

ONLINE SUPPLEMENT MATERIAL

Regulation of *CARD8* Expression by *ANRIL* and Association of *CARD8* SNP rs2043211 (p.C10X) with Ischemic Stroke

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Supplemental Materials and Methods

Cell Transfection and Quantitative Real-Time PCR (qRT-PCR) Analysis

HepG2 cells were purchased from ATCC (American Type Culture Collection, USA) and maintained in the Dulbecco's Modified Eagle's medium (DMEM) supplemented with 10% fetal bovine serum (FBS, Gibco Life Technologies, USA). Human umbilical vein endothelial cells (HUVECs) were purchased from Pricells (Wuhan, China) and maintained in the human endothelial basal growth medium supplemented with 10% FBS, 10 ng/ml of EGF and 1 µg/ml of hydrocortisone. Cells were cultured at 37°C in a humidified incubator with 5% CO₂.

Specific small interfering RNA (siRNA) for human *ANRIL* (*ANRIL* siRNA) and negative control siRNA (NC siRNA) were purchased from RioboBio (Guangzhou, China). The sequence of *ANRIL* siRNA was as follows: 5'-GGAATGAGGAGCACAGTGA-3'. HepG2 cells and HUVECs were transfected with siRNA and incubated for 48 hours. Transfection of siRNA was performed using Lipofectamine 2000 with a final concentration of 100 nM according to the manufacturer's protocol.

Plasmid pcDNA3.1-*ANRIL* (NR_003529.3) was purchased from GENEWIZ (Beijing, China). For transfection of pcDNA3.1-*ANRIL* into HepG2 cells, we used a DNA (µg) to Lipofectamine 2000 (µl) ratio of 1:2. Cells were incubated for 6 h at 37°C in the presence of the transfection mixture and grown in fresh media for 48 hrs. Treated and untreated cells were used for preparation of RNA samples for qRT-PCR analysis.

Cells were harvested 48 hrs after transfection and total RNA was isolated using TRIzol reagent (Invitrogen, USA) according to the manufacturer's instructions. Total RNA was converted into cDNA by reverse transcription using the First-Strand cDNA Synthesis kit and random primers (Promega, USA). The qRT-PCR analysis was performed with a FastStart Universal SYBR Green Master kit (Roche Applied Science, Germany) and analyzed on an ABI 7900-HT Genetic Analyzer. The PCR profile included 94 °C for 5 min and 40 amplification cycles of 94 °C for 20 sec and 60 °C for 15 sec. Melting curve analysis was performed at the end of each PCR reaction to verify the specificity of PCR products. *β-actin* (*ACTB*) was used as internal control. The sequences of primers used are listed in Supplement Table I. Results were representative of three independent experiments and each PCR reaction was run in triplicate. Data analysis was performed using the $2^{-\Delta\Delta C_t}$ method as described.¹

Study Subjects

All participants in our study were selected from the GeneID database.² We performed case-control association studies for ischemic stroke in two independent cohorts involving a total of 1,719 ischemic stroke cases and 1,752 controls. The first discovery population of 903 cases and 873 controls was enrolled from Hubei Province in Central China, whereas the second replication population of 816 cases and 879 controls was enrolled from hospitals in Northern China.

Diagnosis of ischemic stroke was made by following the World Health Organization criteria.³ We classified subjects in the patient case group based on a medical history of ischemic stroke or a stroke diagnosis by magnetic resonance imaging (MRI)/computed tomography (CT). Subjects were excluded if they had a known single-gene stroke disorder, central nervous system vasculitis, intracerebral hemorrhage, subarachnoid hemorrhage, brain tumors, embolic brain infarction,

transient ischemic attack, cardioembolic stroke and a relevant brain stem or subcortical hemispheric lesion with a diameter of <1.5 cm.

The study subjects for an association study for CAD included 772 cases and 873 controls (the same controls used for ischemic stroke studies). CAD was diagnosed based on the American College of Cardiology/American Heart Association criteria. Subjects were classified as CAD cases if more than 70% of luminal stenosis was detected by coronary angiography in at least one main branch of the coronary artery, a procedure of either percutaneous coronary intervention or coronary artery bypass graft was performed, and/or myocardial infarction (MI) was diagnosed. Subjects were excluded if they had a myocardial bridge, congenital heart disease, or childhood hypertension.

Individuals without ischemic stroke and CAD were used as control subjects in the present study.

Other clinical information including the age, sex, smoking history, hypertension, diabetes mellitus, and lipid concentrations were collected from the patients' medical records. Hypertension was defined as a systolic blood pressure of higher than 140 mmHg or a diastolic blood pressure of higher than 90 mmHg. Diabetes was diagnosed on ongoing therapy of diabetes or a fasting plasma glucose level of higher than 7.0 mmol/L. Fasting concentrations of the total cholesterol (Tch), triglyceride (TG), LDL cholesterol (LDL-c), and HDL cholesterol (HDL-c) were measured according to standard methods.

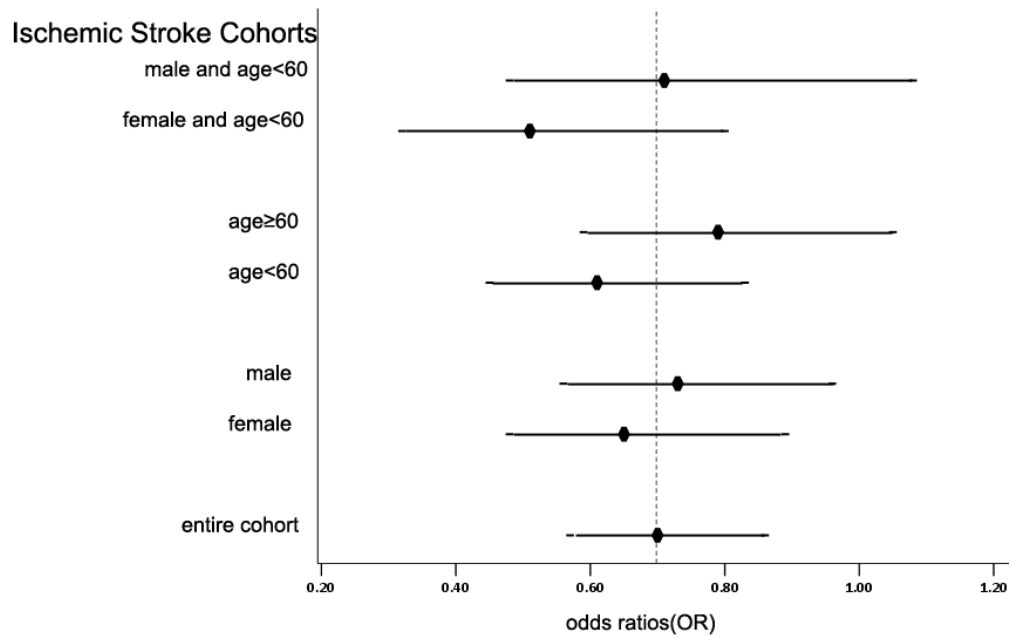
This study followed the principals outlined in the Declaration of Helsinki and has been approved by the local institutional review boards on human subject research. Written informed consents were provided from all participants.

Genotyping and Statistical Analysis

Genomic DNA was isolated from peripheral blood leucocytes with Wizard Genomic DNA Purification Kit (Promega). Genotyping of rs2043211 was carried out using TaqMan SNP Genotyping Assay (c_11708080_1) and analyzed with the ABI 7900-HT Genetic Analyzer according to the manufacturer's instructions (Applied Biosystems). To verify the results by TaqMan assays, we randomly selected 96 DNA samples and genotyped rs2043211 by direct DNA sequencing analysis. The results showed that the accuracy rate of TaqMan assays was 100% in the study. All cluster plots were manually inspected, and ambiguous results were excluded.

The data are reported as mean \pm SEM for qRT-PCR analysis and analyzed using a Student t test (SPSS 17.0). The means were considered significantly different when P was <0.05. A statistical power analysis was carried out using PS software version 3.0.2 for a case-control study.⁴ Hardy-Weinberg disequilibrium tests were carried out in the control group using PLINK version 1.05.⁵ Genotyping data were analyzed for allelic or genotypic association using Pearson's 2×2 and 2×3 contingency table Chi-squared tests, respectively (SPSS version 17.0). Odds ratios (OR), 95% confidence intervals (CIs) and P values were computed by SPSS, version 17.0. Multivariate logistic regression analysis was performed using SPSS version 17.0 by adjusting for traditional risk factors for ischemic stroke or CAD, including the age, sex, body mass index (BMI), smoking history, hypertension, diabetes mellitus and lipid concentrations.

Supplemental Figure



Supplemental Figure I . Comparison of odds ratios (ORs). Horizontal axis shows OR after adjustment for covariates under a recessive model. Vertical axis indicates different groups of patients with ischemic stroke subdivided based on sex and age. Solid rhombus centered on the OR estimate and scaled in proportion to sample size with 95% CI (horizontal bar) are shown for each subgroup.

Supplemental Tables

Supplemental Table I . Primers used in real time PCR analysis for *CARD8*, *ANRIL* and β -*actin*

Primer	Sequence
<i>CARD8</i> -Forward	5' ctg aag gaa atg tgg atg ttg agt 3'
<i>CARD8</i> -reverse	5' cca cag ata cca gcc agc agt 3'
<i>ANRIL</i> -Forward	5' tgg aga cac aca gat gcc taa cg 3'
<i>ANRIL</i> -reverse	5' act cgg gaa gtg cta gca gca atg 3'
β - <i>actin</i> -Forward	5' gga ctt cga gca gga gat gg 3'
β - <i>actin</i> -reverse	5' gca ccg tgt tgg cgt aga gg 3'

**Supplemental Table II . Identification of potential eQTLs for 9p21 SNPs
(rs10116277, rs7865618, rs564398, rs496892, rs7044859)**

Gene	Probe	effect	SE	h2	LOD	pvalue
<i>ABLIM1</i>	200965_s_at	0.275	0.079	3.64	2.652	0.00047
<i>AJAP1</i>	215789_s_at	-0.273	0.078	3.55	2.623	0.00051
<i>ALDOB</i>	211357_s_at	0.284	0.079	4.04	2.772	0.00035
<i>ANKRD50</i>	225731_at	0.263	0.079	3.36	2.444	0.00079
<i>ARL17</i>	229028_s_at	0.283	0.078	3.81	2.873	0.00028
<i>BCAT1</i>	214452_at	0.291	0.077	4.36	3.101	0.00016
<i>BTF3L4</i>	238675_x_at	0.274	0.078	3.58	2.647	0.00048
<i>C10orf88</i>	222852_at	0.271	0.078	3.52	2.627	0.00051
<i>C11orf32</i>	212560_at	0.27	0.079	3.49	2.556	0.0006
<i>C11orf58</i>	200084_at	0.271	0.078	3.67	2.616	0.00052
<i>C18orf30</i>	1563584_at	0.276	0.078	3.83	2.689	0.00043
<i>C1orf108</i>	222459_at	0.265	0.079	3.35	2.458	0.00077
<i>C6orf120</i>	221787_at	0.264	0.079	3.31	2.413	0.00086
<i>CARD8*</i>	<i>1554479_a_at</i>	0.263	0.078	3.45	2.456	0.00077
<i>CD44</i>	<i>212063_at</i>	0.271	0.079	3.67	2.577	0.00057
<i>CHCHD3</i>	217972_at	0.256	0.077	3.42	2.413	0.00086
<i>CLCN3</i>	201733_at	0.259	0.078	3.39	2.413	0.00086
<i>CSF3</i>	207442_at	-0.282	0.079	3.81	2.792	0.00034
<i>CYP2S1</i>	223385_at	0.266	0.078	3.56	2.497	0.0007
<i>DDX58</i>	<i>242961_x_at</i>	0.272	0.079	3.52	2.591	0.00055
<i>DFNA5</i>	203695_s_at	0.272	0.077	3.67	2.698	0.00042
<i>DIP2B</i>	224872_at	0.33	0.08	5.13	3.722	3.50E-05
<i>DKFZP564O0823</i>	225809_at	0.261	0.079	3.4	2.384	0.00092
<i>DMKN</i>	226926_at	0.286	0.079	3.94	2.87	0.00028
<i>DYNC2H1</i>	1565149_at	-0.268	0.078	3.44	2.587	0.00056
<i>EIF2S3</i>	224936_at	0.244	0.071	3.72	2.606	0.00053
<i>FAM129C</i>	230983_at	0.278	0.078	3.7	2.776	0.00035
<i>FKBP5</i>	224856_at	0.26	0.078	3.21	2.38	0.00093
<i>FLJ35348</i>	212547_at	0.263	0.078	3.33	2.482	0.00072
<i>FOXP3</i>	218031_s_at	0.263	0.079	3.28	2.434	0.00081
<i>GLOD4</i>	209092_s_at	0.267	0.08	3.35	2.432	0.00082
<i>GLUL</i>	<i>200648_s_at</i>	0.26	0.078	3.42	2.436	0.00081
<i>GOLM1</i>	217771_at	0.259	0.078	3.44	2.419	0.00084
<i>HECW2</i>	232080_at	0.282	0.078	4.07	2.862	0.00028
<i>HLA-DQA1*</i>	<i>212671_s_at</i>	0.257	0.078	3.32	2.366	0.00096
<i>HOXC4</i>	206194_at	-0.26	0.078	3.48	2.39	0.00091
<i>IAH1</i>	230621_at	0.257	0.078	3.32	2.367	0.00096
<i>IGHV1-69</i>	240915_at	-0.265	0.079	3.36	2.448	0.00079
<i>INADL</i>	223681_s_at	0.298	0.079	4.18	3.06	0.00017
<i>JARID2</i>	203298_s_at	0.261	0.079	3.22	2.377	0.00094
<i>KIF1B</i>	209234_at	0.269	0.079	3.45	2.521	0.00066
<i>KLHL6</i>	1555275_a_at	0.268	0.078	3.59	2.534	0.00064
<i>LARGE</i>	215543_s_at	0.289	0.079	4.16	2.914	0.00025

<i>LAT2</i>	221581_s_at	0.267	0.078	3.53	2.566	0.00059
<i>LCP2</i>	205270_s_at	0.26	0.077	3.47	2.453	0.00078
<i>LIX1L</i>	225793_at	0.276	0.079	3.65	2.665	0.00046
<i>LOC153546</i>	236124_at	-0.275	0.078	3.83	2.723	0.0004
<i>LOC200830</i>	1556898_at	-0.268	0.079	3.39	2.494	0.0007
<i>LOC389831</i>	225046_at	0.26	0.078	3.24	2.383	0.00092
<i>NPM1</i>	200063_s_at	0.292	0.078	4.35	3.05	0.00018
<i>LRPPRC</i>	211971_s_at	0.325	0.078	5.33	3.784	3.00E-05
<i>MAPK8IP1</i>	213013_at	-0.264	0.079	3.29	2.403	0.00088
<i>MIAT</i>	240607_at	-0.264	0.078	3.32	2.478	0.00073
<i>MPEG1</i>	226818_at	0.271	0.078	3.74	2.625	0.00051
<i>MPZL3</i>	227747_at	0.289	0.078	3.99	2.973	0.00022
<i>NARG2</i>	228960_at	0.331	0.079	5.19	3.836	2.60E-05
<i>NCBP2</i>	201517_at	0.279	0.078	3.91	2.751	0.00037
<i>NEXN</i>	1552309_a_at	-0.265	0.078	3.63	2.48	0.00073
<i>NID1</i>	202007_at	0.267	0.078	3.66	2.579	0.00057
<i>NROB2</i>	206410_at	-0.285	0.079	3.83	2.832	0.0003
<i>PAQR8</i>	227626_at	0.29	0.078	4.2	2.988	0.00021
<i>PCMTD1</i>	235507_at	0.258	0.078	3.34	2.362	0.00097
<i>PIGN</i>	219048_at	0.274	0.077	3.63	2.717	0.0004
<i>PIK3CB</i>	<i>212688_at</i>	0.269	0.079	3.44	2.517	0.00066
<i>PNMA6A</i>	235758_at	-0.262	0.078	3.28	2.445	0.00079
<i>PPP1CB</i>	201409_s_at	0.268	0.079	3.6	2.536	0.00063
<i>R3HDM2</i>	203831_at	0.276	0.078	3.65	2.734	0.00039
<i>RAB6IP1</i>	212561_at	0.27	0.08	3.45	2.508	0.00068
<i>RABEP1</i>	225064_at	0.273	0.079	3.73	2.627	0.00051
<i>RIPK3</i>	228139_at	-0.262	0.077	3.31	2.492	0.0007
<i>RPRM</i>	219370_at	-0.264	0.079	3.3	2.438	0.00081
<i>SCRNI</i>	201462_at	0.259	0.078	3.19	2.387	0.00092
<i>SIX1</i>	205817_at	-0.262	0.079	3.25	2.385	0.00092
<i>SLC16A9</i>	227506_at	0.288	0.078	4.01	2.942	0.00023
<i>SLC34A3</i>	1569926_s_at	-0.286	0.079	3.87	2.85	0.00029
<i>SLC7A11</i>	217678_at	0.263	0.079	3.28	2.42	0.00084
<i>SMU1</i>	222618_at	0.267	0.078	3.56	2.511	0.00067
<i>SPG20</i>	236600_at	-0.267	0.078	3.6	2.515	0.00067
<i>ST3GAL6</i>	213355_at	0.272	0.08	3.49	2.533	0.00064
<i>TGFB111</i>	209651_at	0.256	0.078	3.21	2.352	0.001
<i>TMEM185A</i>	227880_s_at	0.266	0.079	3.37	2.451	0.00078
<i>TMPRSS3</i>	220177_s_at	0.27	0.077	3.82	2.689	0.00043
<i>TNFRSF11A</i>	238846_at	0.267	0.079	3.43	2.514	0.00067
<i>TRIM6</i>	223599_at	0.283	0.076	4.05	2.968	0.00022
<i>UBE1L2</i>	222602_at	0.284	0.077	4.09	2.912	0.00025
<i>USH1C</i>	211184_s_at	-0.268	0.079	3.41	2.508	0.00068
<i>ZMYM5</i>	206744_s_at	-0.263	0.079	3.48	2.434	0.00081

Bold and italic: genes with possible involvement in atherosclerosis;

*Reported phenotypic association.

Supplemental Table III. Allelic association analysis between SNP rs2043211 and ischemic stroke.

Cohort (n, case/control)	MAF Case/Control	without adjustment		with adjustment	
		<i>P-obs</i>	OR(95%CI)	<i>P-adj</i>	OR(95%CI)
GeneID-Central					
IS(903/873)	(0.48/0.51)	0.077	0.89(0.78-1.01)	0.092	0.84(0.68-1.03)
GeneID-North					
IS(816/879)	(0.50/0.50)	0.982	1.00(0.88-1.15)	0.697	0.97(0.82-1.15)
Combined IS					
Entire cohort (1719/1752)	(0.49/0.50)	0.210	0.94(0.86-1.03)	0.337	0.94(0.84-1.06)

MAF: Minor allele (T) frequency; OR, odds ratio; *P-obs*, *P* value using 2×2 contingency table χ^2 tests before adjustment for covariates;

P-adj, *P* value adjusted by multivariate logistic regression analysis for traditional risk factors, including age, sex, BMI, smoking history, hypertension, DM, TC, TG, HDL- and LDL-C)

Supplement Table IV. Association analysis between SNP rs2043211 and CAD

Model	Case (MAF or num)	Control (MAF or num)	<i>P-obs</i>	<i>P-adj</i>	OR(95%CI)
allelic association					
entire cohort	0.49(772)	0.51(873)	0.235	0.300	0.90(0.74-1.10)
Male	0.49(538)	0.51(495)	0.430	0.721	0.95(0.75-1.22)
Female	0.48(234)	0.51(378)	0.361	0.372	0.85(0.60-1.22)
Additive association					
entire cohort	174/407/191	230/430/213	0.182	0.296	0.90(0.90-1.11)
Male	124/280/134	125/253/117	0.693	0.718	0.95(0.75-1.22)
Female	50/127/57	105/177/96	0.134	0.359	0.84(0.58-1.22)
Dominant association					
entire cohort	581/191	660/213	0.872	0.408	0.87(0.63-1.20)
Male	404/134	378/117	0.634	0.386	0.83(0.56-1.25)
Female	177/57	282/96	0.773	0.668	1.14(0.63-2.04)
Recessive association					
entire cohort	174/598	230/643	0.073	0.381	0.86(0.62-1.20)
Male	124/414	125/370	0.408	0.782	1.05(0.71-1.59)
Female	50/184	105/273	0.076	0.053	0.55(0.30-1.01)

Model: Additive (TT/AT/AA), Dominant (TT+AT/AA), Recessive (TT/AT+AA);
MAF: Minor allele (T) frequency; OR, odds ratio; *P-obs*, *P* value using 2 × 2 contingency table χ^2 tests before adjustment for covariates; *P-adj*, *P* value adjusted by multivariate logistic regression analysis for traditional risk factors, including age, sex, BMI, smoking history, hypertension, DM, TC, TG, HDL- and LDL-C)

Supplemental References

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