Correction (29 October 2013): A label in fig. S2A was corrected.

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Supplementary Materials for

Circadian Clock NAD⁺ Cycle Drives Mitochondrial Oxidative Metabolism in Mice

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Supplementary Text Figs. S1 to S9

Supporting Descriptions:

Supporting Description 1

In order for a biological process to be classified as circadian, it must display an endogenous oscillation of ~24 hrs in the absence of entraining factors. Experimentally, the rhythm must repeat itself for at least two ~24 hr cycles and persist in the absence of external cues (e.g. light or feeding cues). Thus, to determine whether FAO displays circadian oscillations independent of entraining cues caused by either feeding rhythms or the light-dark cycle, we collected tissues for FAO measurements every 4 hrs over the course of 48 hrs in mice that had been both fasted and maintained in constant darkness (Fig. 1A). Prior to sacrifice at each circadian time point, mice were fasted for 18 hrs, a length of time determined to induce maximal FAO (Fig. S2C) and sirtuin expression in liver (*17*). This length of fasting also minimizes potentially confounding effects of behaviorally-generated rhythms of fasting and feeding.

Supporting Description 2

We also compared oxygen consumption rates (OCR) in isolated mitochondria from wild-type mouse liver in response to either palmitoyl-carnitine plus malate or pyruvate plus malate at two different times of day (Fig. S4C). Compared to ZTO, we observed a relative decrease in OCR in response to pyruvate and malate at ZT12 during the transition from the rest to active period in fasted mice (Fig. S4C), suggesting an intrinsic difference in the relative respiration from oxidative versus glycolytic-derived substrate at the two major behavioral and nutrient state transitions (ZT0 and ZT12). Further, since all mice had previously been fasted, this difference does not appear to be the result of differences in the feeding cycle.

Supporting Description 3

While NMN injection led to a complete rescue of uncoupled (FCCP-treated) respiration in isolated mitochondria, ADP-stimulated respiration was not completely increased to WT amounts (Fig. 2B), suggesting that circadian mutants possess both an NAD⁺-dependent defect in oxidative fuel entry into the electron transport chain, as well as an NAD⁺-independent defect in coupled respiration.

Supplementary Figure Legends:

Fig. S1 Circadian clock control of MEF metabolism. Relative amounts of NAD⁺, FAO, mitochondrial ATP (in galactose-containing media), and lactate (in glucose-containing media) in *Bmal1^{-/-}* and *Cry1^{-/-}/Cry2^{-/-}* MEFs compared to controls (n=6). *p<0.05, **p<0.01, ***p<0.001 for Student's two-tailed t-test comparing control and mutant averages. Data are represented as average \pm SEM.

Fig. S2 Cell-autonomous circadian control of FAO in liver. (**A**) Relative NAD⁺ and FAO in liver from fasted liver-specific *Bmal1^{-/-}* mice and littermate controls collected at ZTO (n=4–7). (**B**) Blood glucose in WT and *Bmal1^{-/-}* mice at ZT12 following an 18 hr fast (n=6). (**C**) FAO at ZT12 in WT mice following increasing lengths of fasting. **p<0.01,

***p<0.001 for Student's two-tailed t-test comparing single time points between WT and mutant averages. Data are represented as average ± SEM.

Fig. S3 Circadian regulation of intrinsic mitochondrial function. (**A**) Mitochondrial NAD⁺ in liver of fasted WT mice maintained in constant darkness and collected every 4 hrs for 24 hrs. Shading indicates where light and dark periods would normally occur in 12:12 light-dark conditions (n=3 per time point). (**B-C**) Relative oxygen consumption rates in isolated liver mitochondria from fasted WT and *Bmal1*^{-/-} mice at ZT0 (**B**) or ZT12 (**C**) following FCCP treatment in the presence of indicated substrates (n=3-4). (**D**) Relative liver mitochondrial NAD⁺ in fasted WT and *Bmal1*^{-/-} mice following injection with either saline or NMN 12 hrs prior to ZT0 collection (n=4). *p<0.05, **p<0.01, ***p<0.001 for Student's two-tailed t-test comparing single time points between WT and mutant averages. Data are represented as average ± SEM.

Fig. S4 Circadian control of fatty acid oxidation in C2C12 cells. (A) Relative NAD⁺ in synchronized C2C12 cells (n=3). (B) Relative FAO and GO in synchronized C2C12 cells (n=1). (C) Ratio of OCR in the presence of palmitoyl-carnitine to pyruvate and malate in WT isolated liver mitochondria collected at ZT0 or ZT12 (n=5). **p<0.01 for Student's two-tailed t-test comparing WT averages at each time point. Data are represented as average \pm SEM.

Fig. S5 Increased glycolytic gene expression in circadian mutants. (A) Relative mRNA expression of *liver pyruvate kinase* (*L-Pk*) in liver of 18-hr fasted WT mice

maintained in constant darkness and collected every 4 hrs for 48 hrs. FAO rhythm is shown for reference (n=4). (**B**) Relative mRNA expression of *pyruvate dehydrogenase kinase 1 (Pdk1), lactate dehydrogenase A (Ldha),* and *phosphofructokinase 1 (Pfk1)* in *Bmal1*^{-/-} and *Cry1*^{-/-}/*Cry2*^{-/-} MEFs compared to controls. (**C**) Relative mRNA expression of *Pdk1, Ldha,* and *L-Pk* in *Bmal1*^{-/-} mutant mouse liver compared to controls at ZTO (n=9–11) and ZT12 (n=3-4). (**D**) Relative mRNA expression of glycolytic genes in liverspecific *Nampt*^{-/-} liver (n=5-6). (**E**) Relative NAD⁺ and lactate production in WT MEFs treated with FK866 (n=2-3). *p<0.05, **p<0.01, ***p<0.001 for Student's two-tailed ttest comparing single time points between WT and mutant averages. Data are represented as average ± SEM.

Fig. S6 Normal mitochondrial biogenesis in circadian mutants. (A) Ratio of mitochondrial to nuclear DNA in liver from fasted WT and *Bmal1*^{-/-} animals at ZT0 and ZT12 (n=9–11). (B) Immunoprecipitated PGC1 α from WT and *Bmal1*^{-/-} fasted liver at ZT0 immunoblotted for acetylated-lysine and PGC1 α . (C-D) Relative mRNA expression of *Ppar* α (C) and genes involved in mitochondrial biogenesis (D) in liver from fasted WT and *Bmal1*^{-/-} animals at ZT0 and ZT12 (n=9–11). *p<0.05, **p<0.01, ***p<0.001 for Student's two-tailed t-test comparing WT and mutant averages at either ZT0 or ZT12. Data are represented at average ± SEM.

Fig. S7 Circadian control of SIRT3 activity. (**A**) Relative LCAD activity in liver mitochondrial extracts from fasted WT and *Bmal1*^{-/-} animals at ZT0 (n=6). (**B**) Relative OTC activity in liver mitochondria from fasted WT mice maintained in constant darkness

and sacrificed every 4 hrs for 48 hrs (n=4). (C) OCR in mitochondria isolated from liver of fasted WT and *Sirt3^{-/-}* mice in the presence of palmitoyl-carnitine following FCCP treatment at ZT0 (n=3). **p<0.01, ***p<0.001 for Student's two-tailed t-test comparing single time points between WT and mutant averages. Data are represented as average \pm SEM.

Fig. S8 Normal SIRT3 protein expression in circadian mutants. (A) SIRT3 protein abundance in liver from fasted WT mice maintained in constant darkness and collected every 4 hrs for 48 hrs. COXIV immunoblotting was performed as a loading control. (B) SIRT3 and MCAD protein in liver mitochondria from fasted WT and *Bmal1*^{-/-} animals at ZT0. COXIV immunoblotting was performed as a loading control (n=3). (C) SIRT3 and β -actin protein in liver from fasted WT and *Bmal1*^{-/-} mice collected at ZT0 following injection with NMN for either 12 hr (top) or 10 consecutive days (bottom) prior to collection.

Fig. S9 Circadian control of SIRT3 activity is independent of redox ratio and nicotinamide levels. (A) NAD⁺/NADH ratio in WT and *Bmal1^{-/-}* MEFs (n=6). (B) *In vitro* deacetylase activity of recombinant SIRT3 on acetylated SIRT3-selective target peptide in the presence of constant NAD⁺ and increasing concentrations of NADH. (C) Nicotinamide (left) and acetyl-CoA (right) measurements in liver from fasted WT and *Bmal1^{-/-}* mice at ZTO (n=3-6). *p<0.05, **p<0.01 for Student's two-tailed t-test comparing single time points between WT and mutant averages. Data are represented as average ± SEM.

















