

Inhalable microorganisms in Beijing's PM_{2.5} and PM₁₀ pollutants during a severe smog event

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Supporting Information

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Detailed Methods

PM and meteorological data

To address the increasing public health concerns, China revised its Ambient Air Quality Standard in 2012, including PM_{2.5} as an air quality index in addition to PM₁₀¹. The 24 h average concentrations (9:00 AM- 9:00 AM the next day) of PM_{2.5} and PM₁₀ from the 3 nearest official monitoring sites in Haidian District and the averaged PM_{2.5} concentration of Beijing from 35 monitoring sites, operated by the Beijing Municipal Environmental Monitoring Center, were recorded from the official website (<http://zx.bjmemc.com.cn/>) as a comparison (Figure 1A and Supporting Information, Figure S7, Table S4). Air Quality Index (AQI) data (Supporting Information, Table S4) of the same 3 monitoring sites were provided by Beijing Municipal Environmental Protection Bureau website (<http://www.bjepb.gov.cn/>). Meteorological data of Haidian District, including temperature (°C), relative humidity (%), atmospheric pressure (Hpa), and wind velocity (m/s) were downloaded from the China Meteorological Data Sharing Service System (<http://cdc.bjmb.gov.cn/>) hourly (Supporting Information, Table S4 and Figure S8). The meteorological data of Beijing were recorded from the National Meteorological Center of China Meteorological Administration (<http://www.nmc.gov.cn/>). Daily PM_{2.5} concentrations of the 74 key cities in the national PM_{2.5} monitoring network were provided by China National Environmental Monitoring Center (<http://113.108.142.147:20035/emcpublish/>).

Scanning Electron Microscopy

Scanning Electron Microscopy (SEM) was performed using a QUANTA 200 SEM (FEI, Czech). Small pieces (0.5 cm×0.5 cm) of Tissuquartz filter samples were mounted

on aluminum SEM stubs using double-sided adhesive carbon tapes, and gold-coated using an Ion Sputtering Instrument E-1010 (Hitachi, Japan) prior to imaging. The beam voltage was set to 15.0 kV.

OC and EC analysis

We analyzed the organic carbon (OC) and elemental carbon (EC) concentrations of the collected filter samples using a DRI 2001A thermal optical reflectance (TOR) carbon analyzer (Atmoslytic Inc Corp., CA, US). Its detection limits for OC and EC were 0.82 and 0.20 $\mu\text{g C}/\text{cm}^2$, respectively. The IMPROVE_A temperature program was used in our analysis^{2,3}. For OC, a small sample (0.522 cm^2) from the quartz filter was heated under four temperatures sequentially (140, 280, 480, and 580 °C) under a helium environment. In this procedure, carbon compounds in the PM samples were transformed to carbon dioxide, which was then reduced to methane over a manganese dioxide catalyst. A flame ionization detector (FID) was used to detect the peak area of methane for estimating the concentration of organic carbon. For EC, the filter was heated at the temperatures of 580, 740, and 840 °C, respectively, and the carrier gas used was a mixture of oxygen (2%) and helium (98%). Elemental carbon was oxidized to carbon dioxide first, followed by reduction to methane and detection using FID. The reflectance of the filter was monitored with a laser to determine the cut-point of OC and EC. Both helium and oxygen were purchased from Beijing AP BAIF Gases Industry Co., Ltd (Beijing, China).

Elemental analysis

Energy dispersive X-ray fluorescence (PANalytical B. V., the Netherlands Corp., NED) was used to measure the concentrations of elements in the PM samples⁴. Nine elements (Ti, Mn, Ni, Cr, Zn, As, Pb, Fe, and Br) were analyzed. Filter papers from the

MicroMatter company film (Micromatter Corp., Vancouver, Canada) and the NIST SRM 2783 standard substance were used to calibrate the instrument. In addition, for every ten samples, we randomly selected one of them to check for repeatability. A blank filter was analyzed as a negative control. The margin of error was <5% for Fe and Zn, <10% for Mn and Br, and 15% for Ti, Cr, and Pb, respectively. Since Ni and As were low in abundance in samples, the error was ~20%.

Water soluble ions

Ion chromatography (IC) was used to measure the water soluble ions in the PM samples⁴, including five anions (SO_4^{2-} , NO_3^- , Cl^- , F^- , and PO_4^{3-}) and five cations (Na^+ , K^+ , Mg^{2+} , Ca^{2+} , and NH_4^+). Dionex ICS-2000 with CS12 columns (Dionex Corp., CA, US) were used for the cation analysis, and Dionex ICS-90A with AS11-HC columns (Dionex Corp., CA, US) were used for the anion analysis. A blank filter was analyzed as a negative control. The method detection limits were 0.01 mg/L for K^+ , Na^+ , and Mg^{2+} , 0.02 mg/L for F^- , Cl^- , NH_4^+ , and Ca^{2+} , and 0.05 mg/L for SO_4^{2-} , PO_4^{3-} , and NO_3^- .

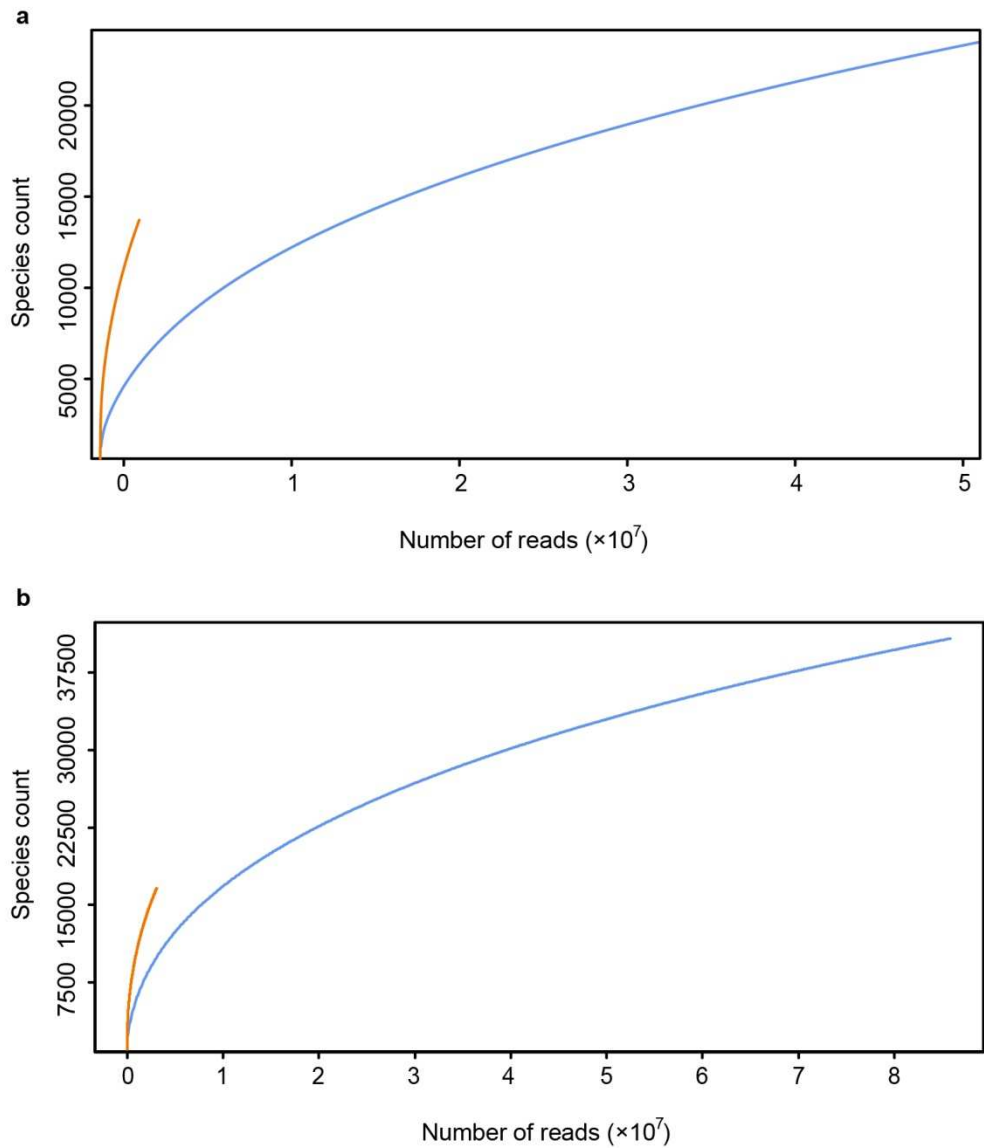
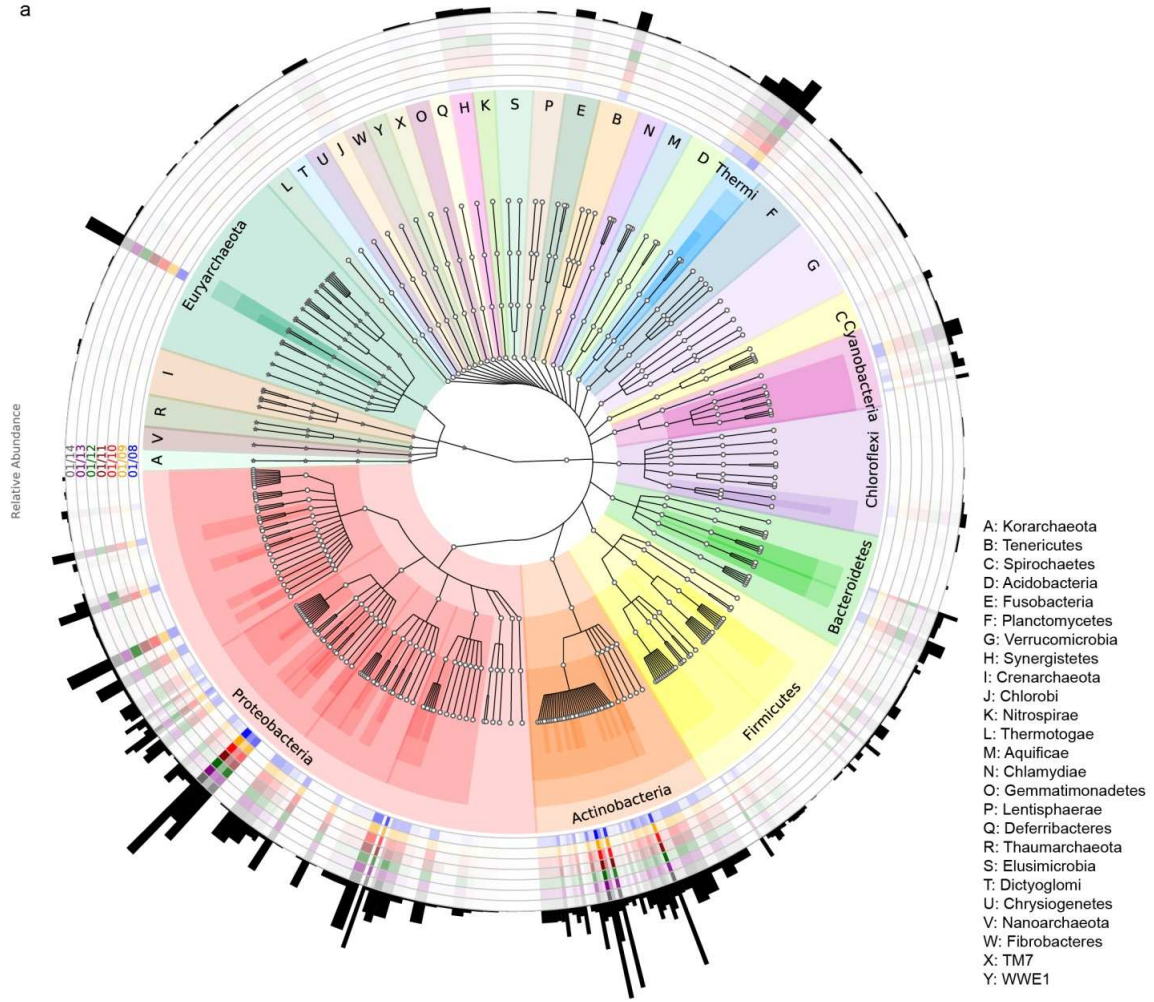


Figure S1. Rarefaction curves of PM_{2.5} and PM₁₀ samples sequenced by MiSeq and HiSeq platforms. Rarefaction curves of **a**) PM_{2.5} and **b**) PM₁₀ samples collected on January 14, 2013, analyzed by MG-RAST. Orange lines correspond to the MiSeq data, while the blue lines correspond to the HiSeq data of the same day.

a



b

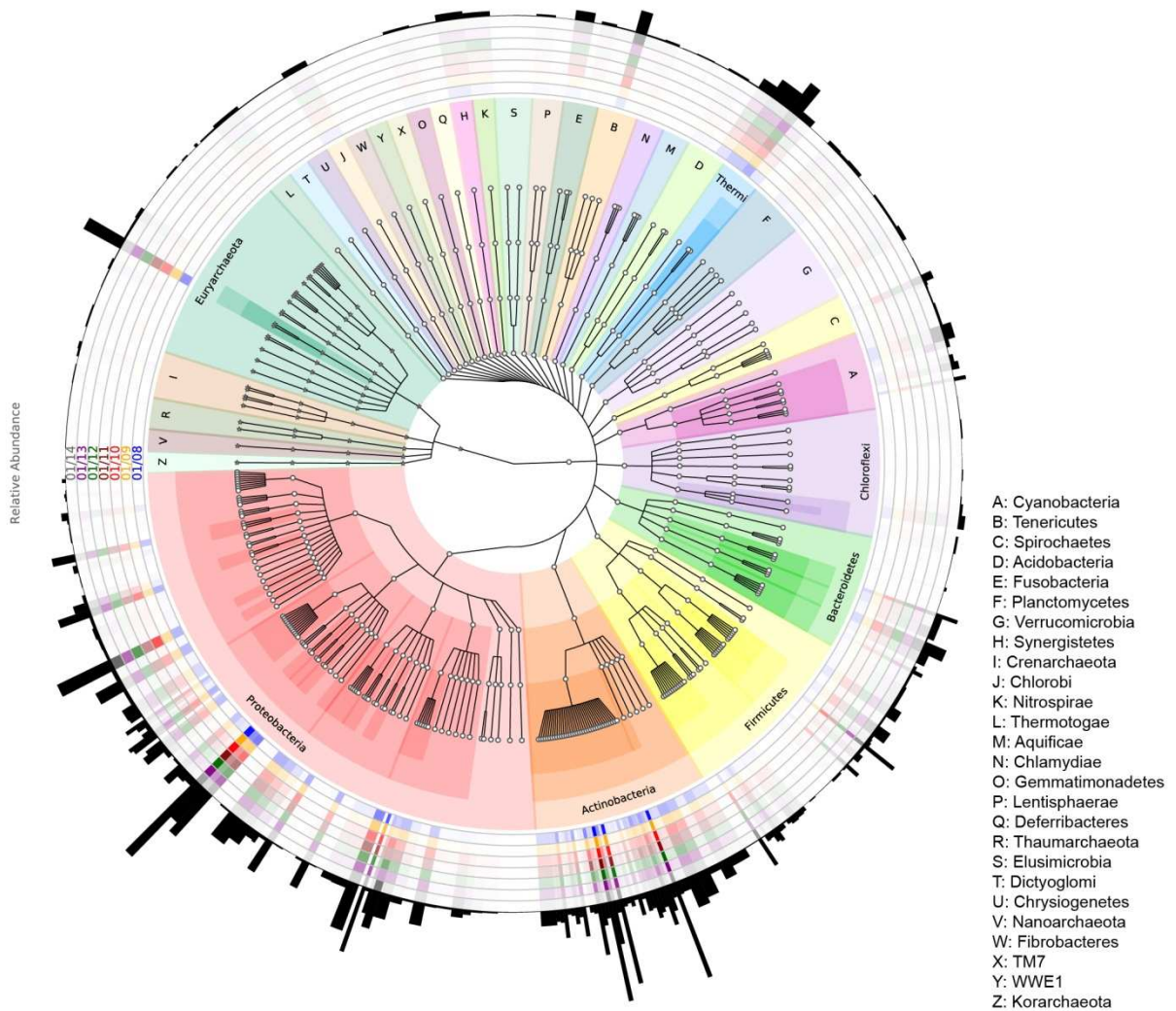
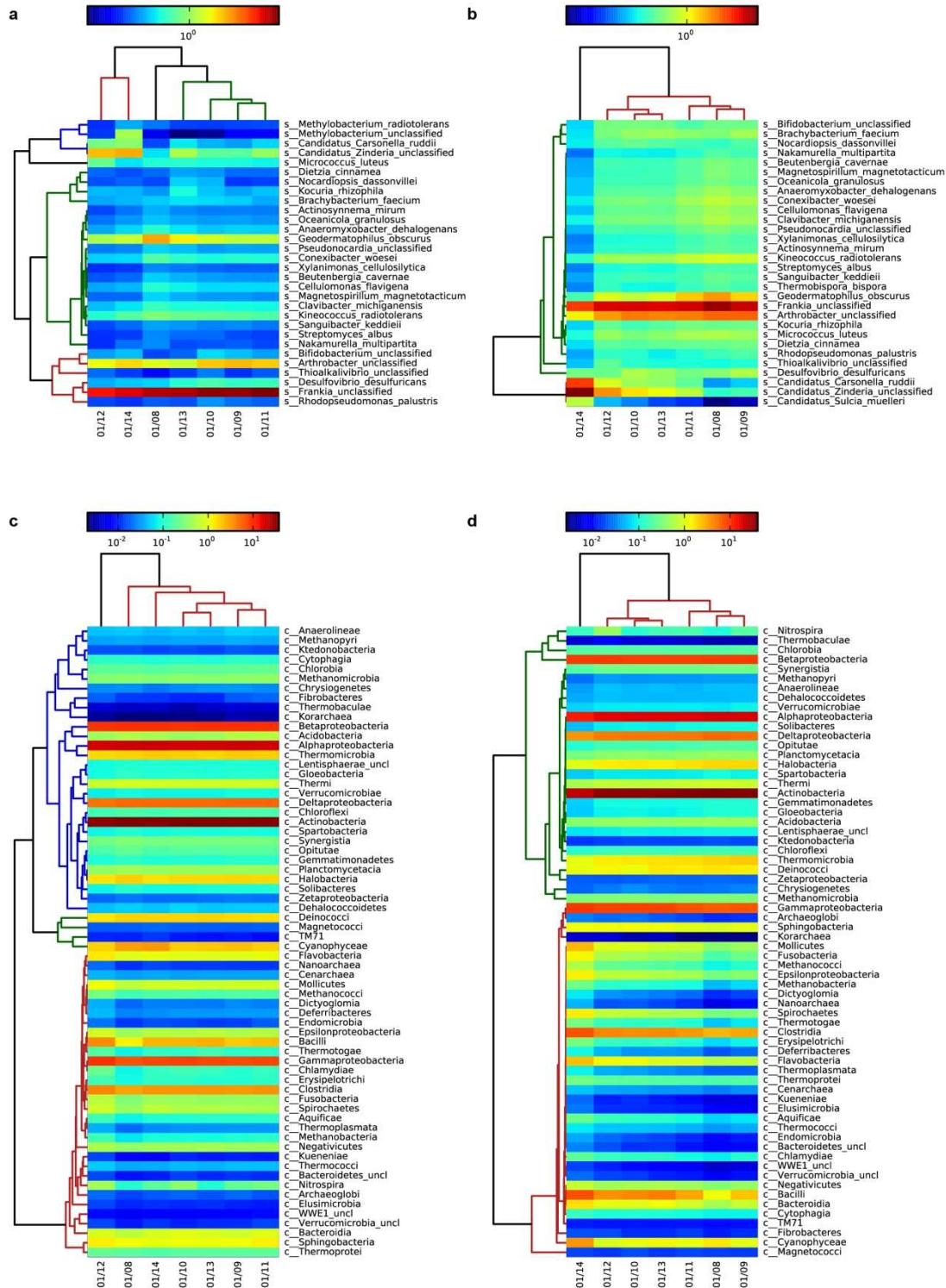


Figure S2. Phylogenetic tree of bacteria and archaea identified from PM_{2.5} and PM₁₀ samples.

Each phylum and species with >1% relative abundance were labeled by different colors at the class, order, and family levels. Black bars in the outmost ring represent the overall relative abundance of each family by height. The 7 transparent rings show the relative abundance of each family during the 7 sampling days, represented by the shade of the color blocks. **a)** PM_{2.5} samples. **b)** PM₁₀ samples.



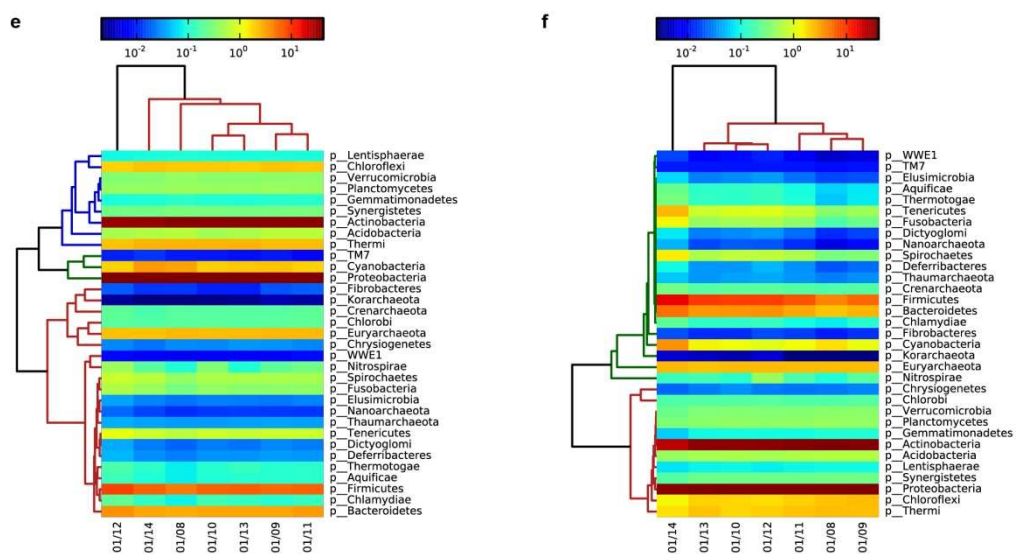
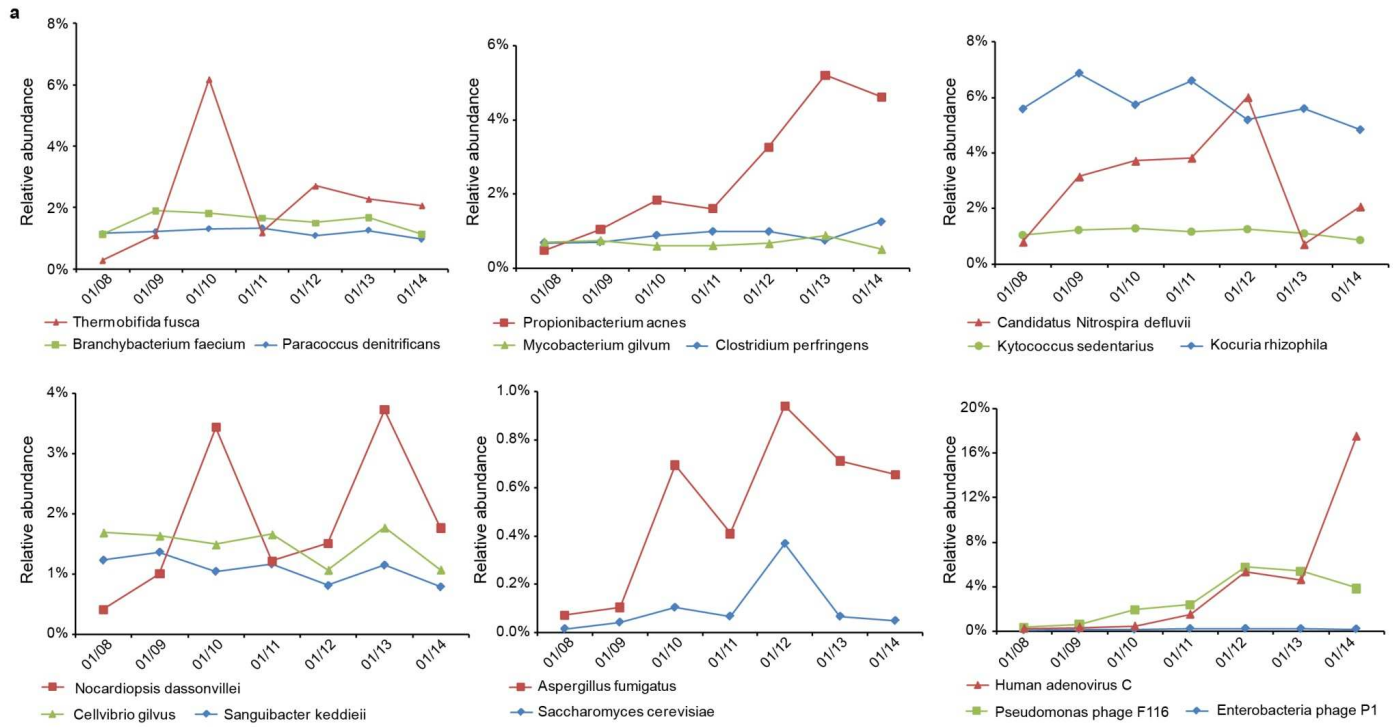


Figure S3. Heatmaps of the relative abundance of bacteria and archaea identified by MetaPhlAn.

The relative abundance (\log_{10} scale) of bacteria and archaea identified by MetaPhlAn at the species (**a**, **b**), class (**c**, **d**) and phylum (**e**, **f**) levels from $PM_{2.5}$ (**a**, **c**, **e**) and PM_{10} (**b**, **d**, **f**) samples during the 7 sampling days (at species level, only the most abundant 30 species were plotted).



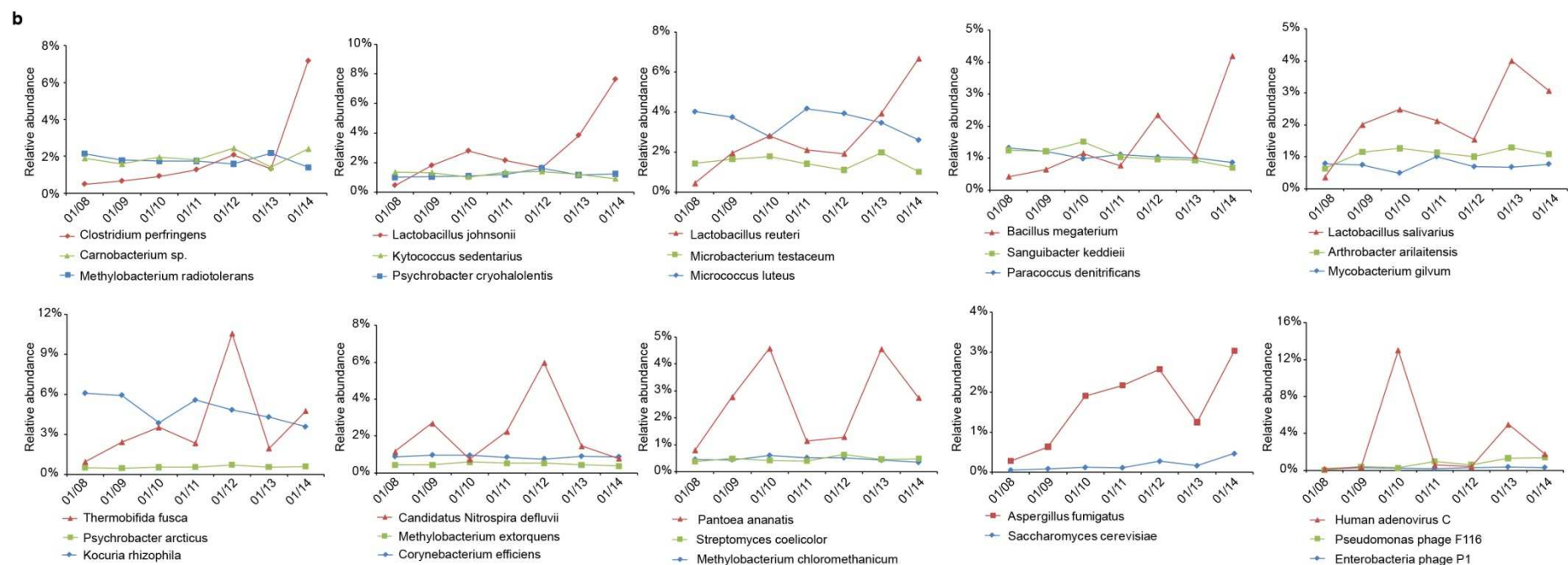


Figure S4. Variations of the genome-size-normalized relative abundance of the most common bacterial, fungal, and viral species. **a)** PM_{2.5} samples. **b)** PM₁₀ samples. Species with S.D. value of their relative abundance (\log_{10} scale) in seven days greater than 0.3 were selected as high variation species (plotted in red), and plotted with one or two other species in the same kingdom for comparison.

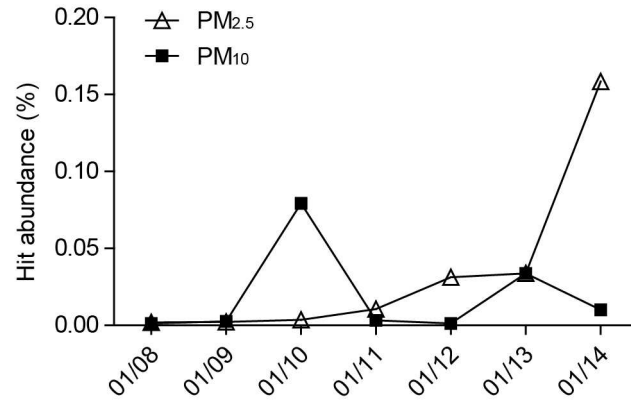


Figure S5. Daily variations of the normalized hit abundance of human adenovirus C in the collected PM_{2.5} and PM₁₀ samples.

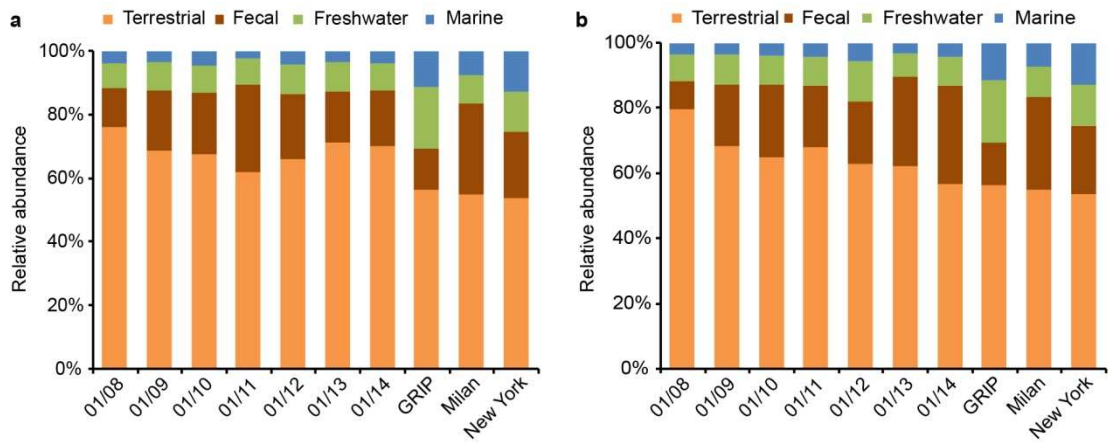


Figure S6. Original habitats of the bacteria identified from the Tsinghua PM_{2.5} and PM₁₀ samples compared with three previous studies.

Original habitats of the bacteria identified from the Tsinghua **a)** PM_{2.5} and **b)** PM₁₀ samples compared with those from the GRIP high-altitude, Milan urban, and New York subway studies

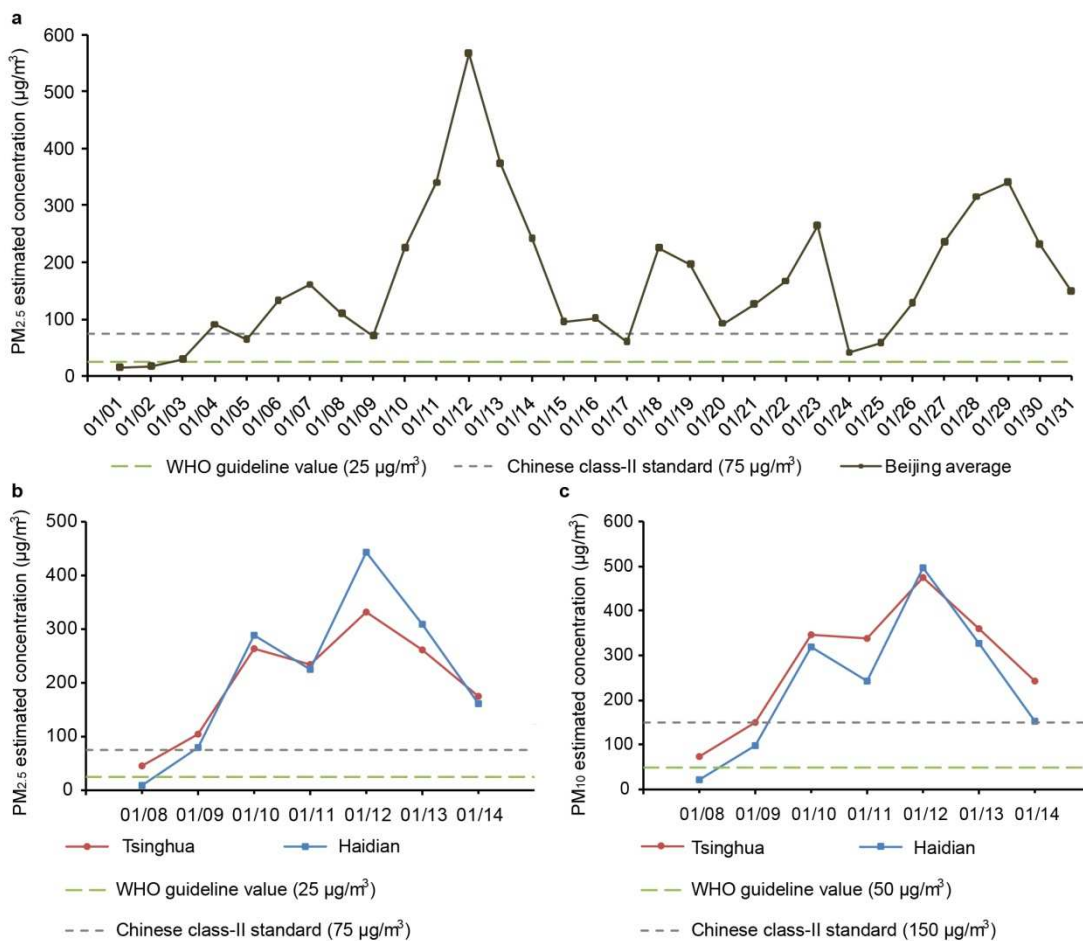


Figure S7. PM_{2.5} and PM₁₀ concentration of Beijing, Haidian district and Tsinghua monitoring site.
 a) Average PM_{2.5} concentration in Beijing in January 2013. b) PM_{2.5} and c) PM₁₀ concentrations of Tsinghua monitoring site and Haidian district during January 8-14, 2013.

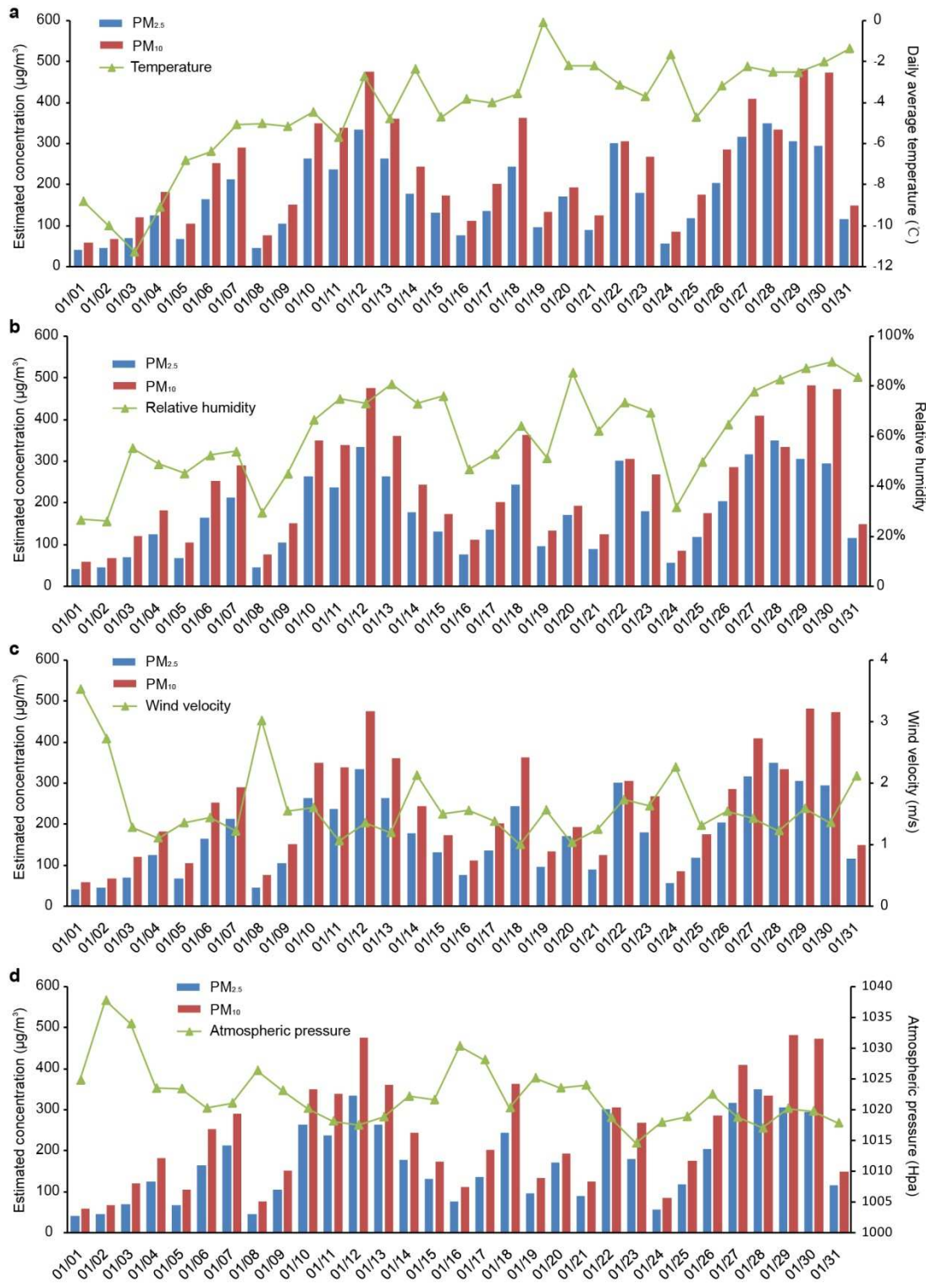


Figure S8. Meteorological data plotted with PM_{2.5} and PM₁₀ concentration in January 2013.

a) Daily average temperature, b) Relative humidity, c) Wind velocity and d) Atmospheric pressure were plotted in green lines, PM_{2.5} concentration was plotted in blue bars and PM₁₀ concentration was plotted in red bars.

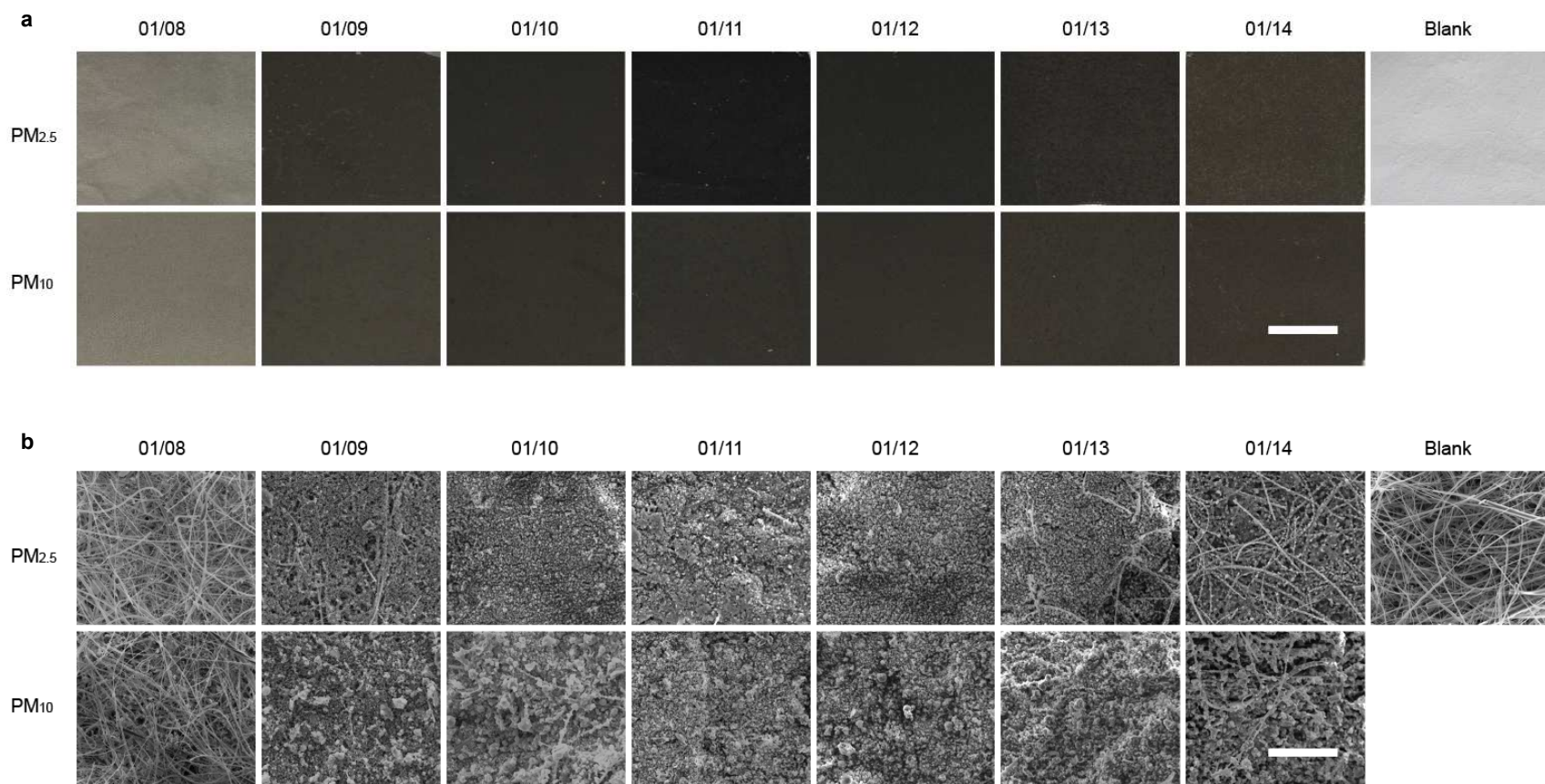


Figure S9. Filter coloration and SEM images of PM_{2.5} and PM₁₀ samples.

a) Filter coloration (Scale bar, 2 cm) and **b)** SEM images (Scale bar, 50 μ m) of PM_{2.5} and PM₁₀ samples collected on the Tissuquartz filters during January 8-14, 2013.

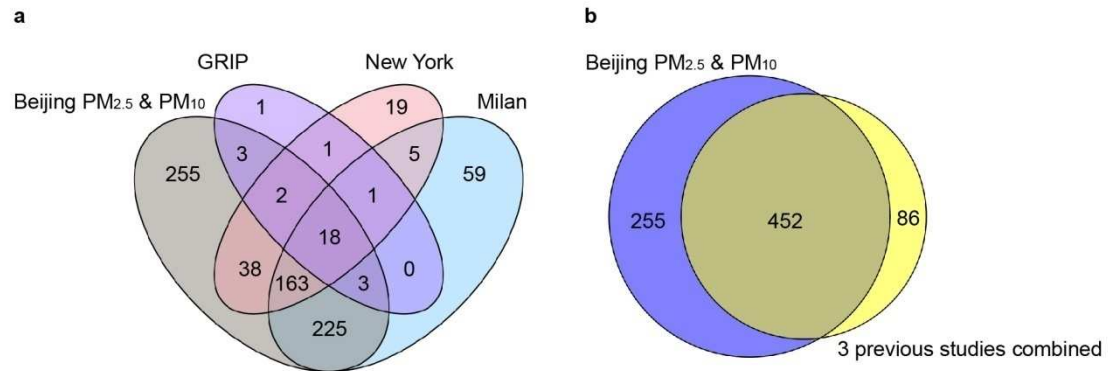


Figure S10. Number of overlapping airborne microbes at the genus level between Tsinghua PM study and three previous studies.
a) Venn diagram of overlapping airborne microbes at the genus level between 4 studies. **b)** Venn diagram of overlapping airborne microbes at the genus level between Tsinghua PM study and three previous studies⁵⁻⁷.

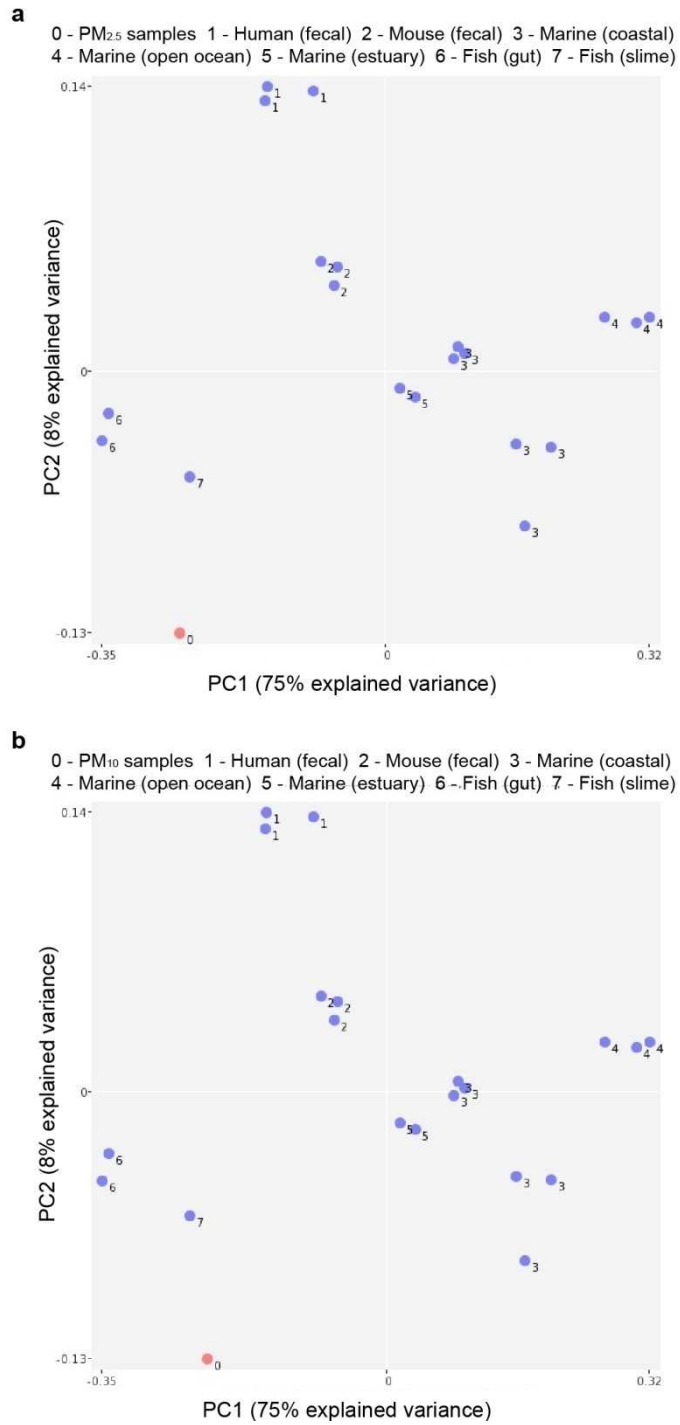


Figure S11. Principle component analysis of dinucleotide frequency of pooled PM_{2.5} samples and PM₁₀ samples. Principle component analysis of dinucleotide frequency of **a**) PM_{2.5} samples and **b**) PM₁₀ samples (represented by a red dot, respectively) compared with other metagenomic studies (calculated by PRINSEQ⁸).

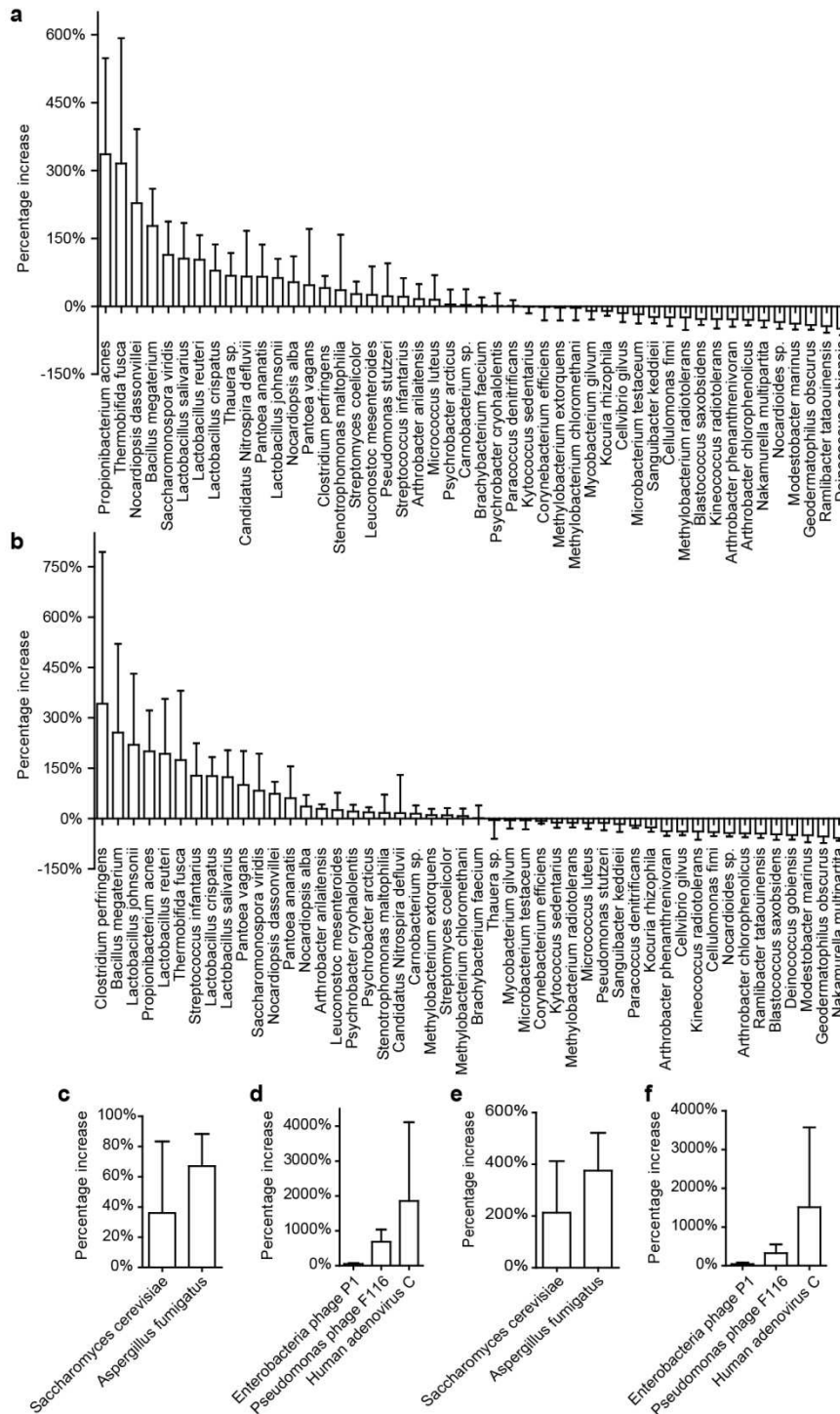


Figure S12. Percentage increase of 53 common species identified from PM samples. Percentage increase of 48 most common bacterial, 2 fungal and 3 viral species from PM_{2.5} (**a, c, d**) and PM₁₀ (**b, e, f**) samples during the last 5 sampling days compared to the first 2 days. Error bars represent S.D. (n=5).

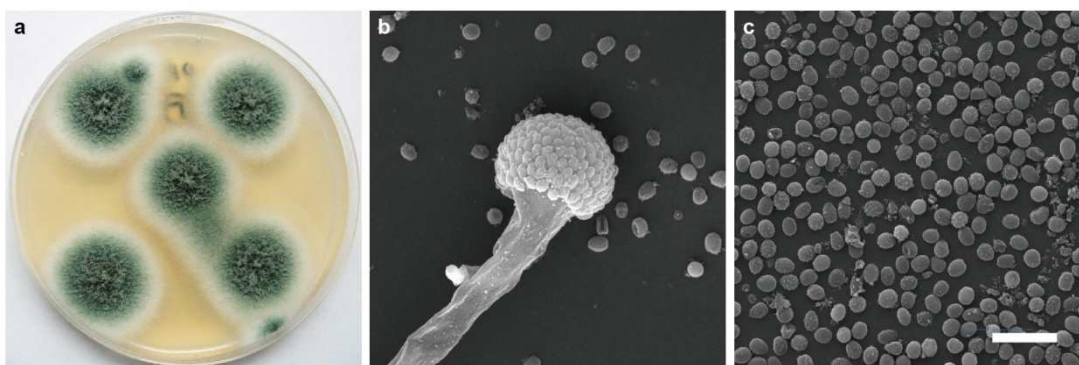


Figure S13. Images of cultured *Aspergillus fumigatus* from PM samples. *Aspergillus fumigatus* colonies on **a)** Potato-Sugar-Agar plate and SEM pictures of the **b)** sporangium and **c,** spores. Scale bar, 10 μm .

Table S1. Summary of sequencing data.

	Sampling date	DNA quantities for library preparation (ng)	Hiseq data				Miseq data		
			Index sequence	Number of reads	Number of bases (bp)	Read length (bp)	Number of reads	Number of bases (bp)	Read length (bp)
PM _{2.5}	01/08	5	CGTGAT	89,678,529	8,071,067,610	90	NA*	NA*	NA*
	01/09	5	GCCTAA	71,393,580	6,425,422,200	90	2,530,054	635,043,554	250
	01/10	5	CACTGT	62,009,579	5,580,862,110	90	3,446,228	865,003,228	250
	01/11	5	GATCTG	77,104,586	6,939,412,740	90	3,256,656	817,420,656	250
	01/12	5	CGTGAT	71,059,996	6,395,399,640	90	3,959,272	993,777,272	250
	01/13	3.84	GTAGCC	79,351,275	7,141,614,750	90	2,718,562	682,359,062	250
	01/14	4.70	CTGATC	49,053,564	4,414,820,760	90	2,885,166	724,176,666	250
PM ₁₀	01/08	5	GATCTG	101,941,930	9,174,773,700	90	NA*	NA*	NA*
	01/09	5	TGGTCA	72,683,317	6,541,498,530	90	2,899,646	727,811,146	250
	01/10	5	ATTGGC	85,462,603	7,691,634,270	90	3,205,264	804,521,264	250
	01/11	5	TCAAGT	78,209,164	7,038,824,760	90	2,115,626	531,022,126	250
	01/12	4.96	TACAAG	78,721,109	7,084,899,810	90	3,198,546	802,835,046	250
	01/13	5	ACATCG	87,108,549	7,839,769,410	90	3,969,610	996,372,110	250
	01/14	5	AAGCTA	85,878,005	7,729,020,450	90	3,499,914	878,478,414	250
Total clean data				1,089,655,786	98,069,020,740		32,646,774	8,194,340,274	
Total raw data				1,192,904,738	107,361,426,420		37,684,544	9,458,820,544	

*Sequencing not performed.

Table S2. Genome coverage of the 48 most common bacterial, 2 fungal, and 3 viral species.

Organism	Species	Genome size (Mb)	Coverage													
			PM _{2.5}							PM ₁₀						
			01/08	01/09	01/10	01/11	01/12	01/13	01/14	01/08	01/09	01/10	01/11	01/12	01/13	01/14
Bacteria	<i>Geodermatophilus obscurus</i>	5.3	54.0%	41.0%	41.0%	43.0%	40.0%	45.0%	35.0%	58.0%	43.0%	34.0%	49.0%	39.0%	39.0%	25.0%
	<i>Modestobacter marinus</i>	5.6	46.0%	39.0%	33.0%	36.0%	28.0%	41.0%	28.0%	42.0%	30.0%	22.0%	38.0%	24.0%	29.0%	17.0%
	<i>Blastococcus saxobsidens</i>	4.9	32.0%	27.0%	28.0%	27.0%	25.0%	33.0%	25.0%	33.0%	29.0%	23.0%	29.0%	24.0%	27.0%	17.0%
	<i>Kocuria rhizophila</i>	2.7	14.0%	17.0%	16.0%	17.0%	17.0%	22.0%	12.0%	17.0%	18.0%	15.0%	17.0%	16.0%	18.0%	14.0%
	<i>Micrococcus luteus</i>	2.5	13.0%	26.0%	18.0%	20.0%	53.0%	25.0%	14.0%	24.0%	19.0%	20.0%	25.0%	26.0%	27.0%	17.0%
	<i>Candidatus Nitrospira defluvii</i>	4.3	11.0%	34.0%	39.0%	40.0%	60.0%	12.0%	23.0%	19.0%	34.0%	12.0%	29.0%	58.0%	26.0%	10.0%
	<i>Methylobacterium radiotolerans</i>	6.1	29.0%	26.0%	20.0%	26.0%	23.0%	22.0%	28.0%	25.0%	21.0%	23.0%	21.0%	19.0%	29.0%	16.0%
	<i>Propionibacterium acnes</i>	2.6	7.1%	13.0%	23.0%	20.0%	42.0%	61.0%	44.0%	10.0%	14.0%	17.0%	27.0%	28.0%	45.0%	37.0%
	<i>Thermobifida fusca</i>	3.6	3.7%	13.0%	57.0%	15.0%	35.0%	35.0%	23.0%	16.0%	31.0%	47.0%	31.0%	80.0%	35.0%	47.0%
	<i>Nocardioides</i> sp.	5.0	12.0%	11.0%	10.0%	10.0%	9.4%	11.0%	7.5%	15.0%	13.0%	12.0%	12.0%	11.0%	12.0%	7.9%
	<i>Nocardiopsis dassonvillei</i>	6.5	4.4%	10.9%	36.0%	13.7%	20.6%	45.7%	18.2%	14.6%	28.4%	46.1%	28.1%	33.6%	39.4%	30.8%
	<i>Brachybacterium faecium</i>	3.6	9.3%	14.0%	16.0%	13.0%	15.0%	18.0%	9.5%	15.0%	31.0%	28.0%	23.0%	19.0%	42.0%	12.0%
	<i>Arthrobacter phenanthrenivorans</i>	4.3	18.0%	15.0%	15.0%	15.0%	12.0%	14.0%	8.1%	26.0%	21.0%	20.0%	20.0%	16.0%	21.0%	11.0%
	<i>Cellvibrio gilvus</i>	3.5	8.6%	7.8%	8.2%	7.9%	6.6%	15.0%	5.8%	9.2%	8.4%	7.5%	7.4%	6.6%	7.4%	4.9%
	<i>Carnobacterium</i> sp.	2.6	18.0%	16.0%	19.0%	16.0%	25.0%	19.0%	8.6%	25.0%	20.0%	26.0%	22.0%	27.0%	23.0%	25.0%
	<i>Microbacterium testaceum</i>	4.0	8.0%	12.0%	10.0%	11.0%	8.9%	11.0%	6.2%	10.0%	12.0%	15.0%	11.0%	8.8%	18.0%	8.6%
	<i>Paracoccus denitrificans</i>	4.6	8.1%	8.0%	8.3%	8.3%	8.3%	9.5%	6.7%	9.7%	8.6%	8.7%	8.4%	8.1%	9.2%	6.7%
	<i>Kytococcus sedentarius</i>	2.8	6.5%	7.8%	9.2%	7.3%	10.0%	9.6%	5.9%	10.0%	10.0%	10.0%	12.0%	12.0%	14.0%	7.7%
	<i>Pantoea vagans</i>	4.0	10.0%	12.0%	32.0%	16.0%	14.0%	9.0%	5.0%	8.8%	20.0%	46.0%	23.0%	17.0%	27.0%	20.0%
	<i>Pseudomonas stutzeri</i>	4.7	12.0%	13.0%	15.0%	9.4%	12.0%	12.0%	25.0%	17.0%	18.0%	19.0%	15.0%	15.0%	21.0%	7.4%
	<i>Kineococcus radiotolerans</i>	4.8	10.0%	8.8%	8.4%	9.9%	6.1%	6.4%	7.9%	11.0%	8.2%	6.1%	11.0%	5.1%	7.2%	3.6%
	<i>Sanguibacter keddieii</i>	4.3	9.5%	10.0%	8.1%	8.1%	7.7%	12.0%	5.9%	9.4%	11.0%	19.0%	8.9%	9.4%	11.0%	6.7%
	<i>Thauera</i> sp.	4.5	6.3%	11.0%	12.0%	17.0%	23.0%	13.0%	11.0%	8.9%	18.0%	10.0%	16.0%	23.0%	13.0%	5.4%
	<i>Pantoea ananatis</i>	4.7	4.2%	15.0%	16.0%	13.0%	26.0%	16.0%	6.6%	13.0%	35.0%	57.0%	16.0%	18.0%	62.0%	31.0%
	<i>Cellulomonas fimi</i>	4.3	8.6%	6.9%	7.1%	7.5%	5.6%	7.5%	5.1%	8.7%	7.7%	6.2%	6.9%	6.0%	6.9%	4.4%
	<i>Lactobacillus johnsonii</i>	2.0	5.2%	13.0%	13.0%	12.0%	22.0%	21.0%	10.0%	8.5%	23.0%	37.0%	28.0%	23.0%	49.0%	58.0%
	<i>Deinococcus gobiensis</i>	3.1	20.0%	14.0%	11.0%	12.0%	9.9%	8.2%	7.3%	19.0%	8.1%	7.6%	9.6%	6.0%	10.0%	4.2%
	<i>Lactobacillus crispatus</i>	2.0	5.2%	12.0%	14.0%	12.0%	23.0%	18.0%	8.1%	8.6%	25.0%	37.0%	30.0%	24.0%	50.0%	32.0%
	<i>Lactobacillus salivarius</i>	1.8	5.0%	9.1%	13.0%	10.0%	22.0%	21.0%	7.0%	6.7%	25.0%	35.0%	28.0%	21.0%	51.0%	33.0%

	<i>Lactobacillus reuteri</i>	2.3	3.2%	11.0%	12.0%	11.0%	20.0%	19.0%	10.0%	8.0%	25.0%	37.0%	28.0%	25.0%	50.0%	51.0%
	<i>Clostridium perfringens</i>	3.3	9.3%	9.1%	12.0%	12.0%	15.0%	12.0%	14.0%	9.1%	9.9%	15.0%	18.0%	26.0%	24.0%	57.0%
	<i>Bacillus megaterium</i>	5.1	3.5%	6.3%	8.5%	9.9%	16.0%	18.0%	16.0%	7.8%	9.7%	19.0%	12.0%	30.0%	20.0%	42.0%
	<i>Psychrobacter cryohalolentis</i>	3.1	9.2%	7.8%	9.1%	9.4%	12.0%	9.6%	5.2%	12.0%	12.0%	14.0%	14.0%	17.0%	17.0%	12.0%
	<i>Arthrobacter arilaitensis</i>	3.9	5.2%	11.0%	9.1%	8.4%	9.7%	12.0%	4.7%	9.2%	15.0%	19.0%	14.0%	13.0%	21.0%	12.0%
	<i>Ramlibacter tataouinensis</i>	4.1	12.0%	9.8%	6.8%	7.8%	5.3%	8.1%	7.5%	11.0%	8.7%	6.8%	7.8%	6.2%	8.1%	3.7%
	<i>Nakamurella multipartita</i>	6.1	12.0%	8.1%	6.0%	6.5%	9.9%	6.0%	8.6%	12.0%	11.0%	5.6%	5.7%	5.3%	5.0%	3.6%
	<i>Streptococcus infantarius</i>	2.0	6.0%	9.8%	8.0%	8.2%	13.0%	17.0%	6.2%	7.9%	12.0%	18.0%	15.0%	13.0%	21.0%	22.0%
	<i>Stenotrophomonas maltophilia</i>	4.9	6.5%	7.2%	23.0%	5.7%	5.4%	6.9%	6.6%	6.0%	8.6%	15.0%	9.9%	6.1%	8.2%	4.2%
	<i>Arthrobacter chlorophenolicus</i>	4.4	8.3%	7.7%	6.4%	7.0%	5.5%	6.5%	6.6%	14.0%	14.0%	11.0%	8.9%	8.7%	10.0%	5.1%
	<i>Mycobacterium gilvum</i>	5.6	8.8%	8.9%	7.4%	6.9%	9.9%	12.0%	5.7%	12.0%	10.0%	8.2%	11.0%	9.6%	11.0%	9.5%
	<i>Methylobacterium extorquens</i>	6.0	9.1%	8.1%	7.3%	9.4%	6.5%	8.7%	9.8%	7.5%	6.6%	9.8%	8.1%	7.9%	8.5%	5.0%
	<i>Methylobacterium chloromethanicum</i>	5.8	8.7%	7.9%	7.7%	9.3%	6.3%	8.4%	9.0%	7.6%	6.6%	9.9%	8.0%	7.8%	8.1%	4.5%
	<i>Corynebacterium efficiens</i>	3.1	6.1%	8.8%	9.8%	9.3%	8.5%	9.2%	3.9%	14.0%	13.0%	15.0%	12.0%	11.0%	16.0%	11.0%
	<i>Saccharomonospora viridis</i>	4.3	2.1%	4.7%	11.0%	5.0%	9.2%	13.0%	5.2%	8.3%	13.0%	16.0%	8.8%	26.0%	13.0%	26.0%
	<i>Nocardiopsis alba</i>	5.8	2.0%	5.8%	9.5%	8.1%	5.3%	8.9%	3.4%	5.3%	11.0%	16.0%	12.0%	10.0%	16.0%	5.9%
	<i>Psychrobacter arcticus</i>	2.7	5.0%	3.9%	4.6%	5.2%	6.1%	5.0%	2.4%	6.6%	5.7%	7.2%	6.2%	7.5%	8.0%	6.1%
	<i>Leuconostoc mesenteroides</i>	2.0	5.5%	3.5%	4.6%	4.5%	11.0%	5.2%	3.6%	7.1%	5.9%	6.1%	6.5%	8.2%	7.1%	11.0%
	<i>Streptomyces coelicolor</i>	8.7	2.3%	3.9%	5.5%	3.6%	4.9%	7.1%	3.3%	5.1%	6.2%	6.4%	5.3%	8.1%	8.0%	5.8%
Fungus	<i>Aspergillus fumigatus</i> Af293	29.4	0.7%	1.0%	8.9%	5.0%	13.4%	11.2%	7.6%	4.7%	9.0%	27.9%	27.8%	31.7%	22.6%	31.7%
	<i>Saccharomyces cerevisiae</i> S288c	12.1	0.2%	0.5%	1.4%	0.9%	5.9%	1.2%	0.6%	1.1%	1.4%	2.2%	1.8%	4.2%	3.5%	6.2%
Virus	Human adenovirus C ₁	0.036	2.4%	3.6%	4.5%	7.7%	9.8%	9.3%	12.0%	2.9%	3.7%	9.6%	4.8%	4.0%	8.8%	8.0%
	<i>Pseudomonas</i> phage F116	0.065	5.4%	8.0%	18.0%	22.0%	34.0%	35.0%	27.0%	0.7%	5.9%	4.7%	13.0%	8.1%	20.0%	16.0%
	Enterobacteria phage P1	0.095	1.1%	1.5%	1.9%	1.9%	2.5%	2.1%	1.9%	1.3%	2.2%	2.8%	2.2%	2.0%	3.2%	2.6%

Table S3. Genome-size-normalized relative abundance of the 48 most common bacterial, 2 fungal, and 3 viral species.

Organism	Species	Genome size (Mb)	Relative abundance													
			PM _{2.5}							PM ₁₀						
			01/08	01/09	01/10	01/11	01/12	01/13	01/14	01/08	01/09	01/10	01/11	01/12	01/13	01/14
Bacteria	<i>Geodermatophilus obscurus</i>	5.3	24.99%	16.34%	12.57%	15.08%	9.49%	12.66%	10.73%	21.81%	12.64%	6.12%	13.51%	8.20%	7.78%	4.47%
	<i>Modestobacter marinus</i>	5.6	16.79%	14.07%	9.64%	12.11%	6.60%	10.40%	8.90%	13.83%	9.45%	4.51%	9.70%	5.46%	5.97%	3.51%
	<i>Blastococcus saxosidens</i>	4.9	11.05%	9.25%	7.49%	8.37%	5.37%	8.50%	6.65%	10.21%	8.15%	4.10%	7.21%	4.97%	5.18%	3.02%
	<i>Kocuria rhizophila</i>	2.7	5.59%	6.87%	5.74%	6.60%	5.20%	5.61%	4.85%	6.09%	5.93%	3.85%	5.57%	4.84%	4.30%	3.58%
	<i>Micrococcus luteus</i>	2.5	2.94%	4.71%	3.48%	4.17%	7.98%	3.44%	2.80%	4.03%	3.75%	2.79%	4.18%	3.94%	3.47%	2.61%
	<i>Candidatus Nitrospira defluvii</i>	4.3	0.80%	3.15%	3.72%	3.83%	6.02%	0.71%	2.07%	1.15%	2.69%	0.75%	2.24%	5.98%	1.46%	0.79%
	<i>Methylobacterium radiotolerans</i>	6.1	3.56%	3.08%	1.94%	2.97%	2.05%	1.66%	3.92%	2.14%	1.79%	1.74%	1.74%	1.60%	2.17%	1.40%
	<i>Propionibacterium acnes</i>	2.6	0.48%	1.04%	1.84%	1.61%	3.27%	5.21%	4.62%	0.61%	0.92%	1.02%	1.92%	2.23%	2.76%	3.50%
	<i>Thermobifida fusca</i>	3.6	0.29%	1.10%	6.18%	1.19%	2.71%	2.29%	2.08%	0.95%	2.43%	3.53%	2.35%	10.56%	1.95%	4.76%
	<i>Nocardioides</i> sp.	5.0	2.78%	2.55%	1.99%	2.22%	1.47%	1.79%	1.23%	3.62%	2.77%	1.93%	2.17%	1.92%	1.81%	1.29%
	<i>Nocardiopsis dassonvillei</i>	6.5	0.41%	1.01%	3.43%	1.21%	1.51%	3.72%	1.76%	1.02%	2.43%	3.97%	2.31%	2.86%	2.74%	3.13%
	<i>Brachybacterium faecium</i>	3.6	1.14%	1.90%	1.81%	1.67%	1.51%	1.68%	1.13%	1.60%	3.33%	2.83%	2.47%	2.01%	3.85%	1.37%
	<i>Arthrobacter phenanthrenivorans</i>	4.3	1.98%	1.79%	1.60%	1.68%	1.24%	1.28%	0.91%	2.99%	2.24%	1.85%	2.09%	1.51%	1.66%	1.12%
	<i>Cellvibrio gilvus</i>	3.5	1.69%	1.64%	1.50%	1.66%	1.07%	1.78%	1.08%	1.78%	1.53%	1.01%	1.31%	1.04%	1.00%	0.77%
	<i>Carnobacterium</i> sp.	2.6	1.40%	1.43%	1.71%	1.43%	2.07%	1.35%	0.77%	1.90%	1.59%	1.95%	1.81%	2.43%	1.41%	2.40%
	<i>Microbacterium testaceum</i>	4.0	1.23%	1.82%	1.39%	1.64%	1.09%	1.33%	0.83%	1.44%	1.65%	1.79%	1.42%	1.11%	1.99%	1.02%
	<i>Paracoccus denitrificans</i>	4.6	1.16%	1.22%	1.31%	1.34%	1.10%	1.26%	0.98%	1.32%	1.21%	0.99%	1.11%	1.04%	1.01%	0.86%
	<i>Kytococcus sedentarius</i>	2.8	1.08%	1.26%	1.31%	1.20%	1.28%	1.12%	0.88%	1.36%	1.33%	1.03%	1.37%	1.40%	1.20%	0.92%
	<i>Pantoea vagans</i>	4.0	0.76%	0.94%	3.04%	1.32%	0.95%	0.54%	0.41%	0.52%	1.44%	3.71%	1.60%	1.20%	1.67%	1.63%
	<i>Pseudomonas stutzeri</i>	4.7	0.85%	1.10%	1.23%	0.77%	0.80%	0.76%	2.42%	0.99%	1.30%	1.13%	1.05%	1.10%	1.16%	0.56%
	<i>Kineococcus radiotolerans</i>	4.8	1.41%	1.31%	1.13%	1.36%	0.68%	0.75%	0.94%	1.37%	1.09%	0.67%	1.27%	0.67%	0.72%	0.45%
	<i>Sanguibacter keddieii</i>	4.3	1.23%	1.36%	1.05%	1.17%	0.81%	1.15%	0.79%	1.26%	1.22%	1.52%	1.05%	0.97%	0.94%	0.71%
	<i>Thauera</i> sp.	4.5	0.51%	0.92%	0.98%	1.46%	1.68%	0.85%	0.99%	0.57%	1.40%	0.62%	1.14%	1.79%	0.75%	0.41%
	<i>Pantoea ananatis</i>	4.7	0.28%	1.13%	1.34%	1.07%	1.91%	0.99%	0.54%	0.79%	2.77%	4.59%	1.14%	1.29%	4.57%	2.75%
	<i>Cellulomonas fimi</i>	4.3	1.32%	1.16%	1.05%	1.28%	0.71%	0.94%	0.72%	1.39%	1.16%	0.75%	0.98%	0.76%	0.75%	0.55%
	<i>Lactobacillus johnsonii</i>	2.0	0.35%	1.06%	1.06%	0.97%	1.56%	1.35%	0.82%	0.46%	1.80%	2.79%	2.15%	1.66%	3.85%	7.65%
	<i>Deinococcus gobiensis</i>	3.1	1.84%	1.28%	1.00%	1.06%	0.66%	0.54%	0.71%	1.39%	0.59%	0.46%	0.68%	0.43%	0.59%	0.34%
	<i>Lactobacillus crispatus</i>	2.0	0.35%	0.90%	1.17%	1.02%	1.67%	1.07%	0.67%	0.47%	1.93%	2.79%	2.27%	1.91%	3.68%	2.97%
<i>Lactobacillus salivarius</i>	1.8	0.34%	0.70%	0.98%	0.83%	1.59%	1.36%	0.57%	0.36%	2.00%	2.49%	2.13%	1.55%	4.01%	3.07%	

	<i>Lactobacillus reuteri</i>	2.3	0.22%	0.82%	0.86%	0.89%	1.44%	1.27%	0.82%	0.44%	1.95%	2.82%	2.10%	1.92%	3.93%	6.68%
	<i>Clostridium perfringens</i>	3.3	0.68%	0.70%	0.89%	0.99%	0.99%	0.75%	1.26%	0.49%	0.66%	0.93%	1.28%	2.08%	1.33%	7.17%
	<i>Bacillus megaterium</i>	5.1	0.24%	0.49%	0.63%	0.81%	1.05%	1.11%	1.40%	0.43%	0.65%	1.16%	0.77%	2.35%	1.07%	4.20%
	<i>Psychrobacter cryohalolentis</i>	3.1	0.84%	0.77%	0.86%	0.96%	1.04%	0.73%	0.46%	1.02%	1.06%	1.09%	1.20%	1.62%	1.16%	1.23%
	<i>Arthrobacter arilaitensis</i>	3.9	0.48%	0.96%	0.80%	0.80%	1.19%	0.88%	0.53%	0.64%	1.15%	1.27%	1.13%	1.01%	1.29%	1.08%
	<i>Ramlibacter tataouinensis</i>	4.1	1.23%	1.10%	0.64%	0.85%	0.41%	0.62%	0.78%	1.02%	0.79%	0.52%	0.64%	0.52%	0.52%	0.30%
	<i>Nakamurella multipartita</i>	6.1	1.17%	0.85%	0.60%	0.71%	0.77%	0.49%	0.90%	1.04%	0.94%	0.42%	0.53%	0.45%	0.35%	0.34%
	<i>Streptococcus infantarius</i>	2.0	0.44%	0.83%	0.59%	0.67%	0.93%	1.14%	0.50%	0.43%	0.87%	1.25%	1.13%	1.05%	1.43%	2.59%
	<i>Stenotrophomonas maltophilia</i>	4.9	0.52%	0.61%	2.00%	0.47%	0.36%	0.42%	0.59%	0.36%	0.62%	0.98%	0.70%	0.44%	0.43%	0.32%
	<i>Arthrobacter chlorophenolicus</i>	4.4	0.87%	0.87%	0.65%	0.76%	0.49%	0.52%	0.64%	1.33%	1.29%	0.86%	0.84%	0.78%	0.71%	0.48%
	<i>Mycobacterium gilvum</i>	5.6	0.71%	0.75%	0.60%	0.61%	0.66%	0.88%	0.51%	0.79%	0.75%	0.50%	1.01%	0.70%	0.68%	0.77%
	<i>Methylobacterium extorquens</i>	6.0	0.67%	0.62%	0.54%	0.77%	0.43%	0.53%	0.87%	0.45%	0.44%	0.59%	0.52%	0.52%	0.44%	0.38%
	<i>Methylobacterium chloromethanicum</i>	5.8	0.64%	0.61%	0.57%	0.76%	0.39%	0.51%	0.80%	0.45%	0.44%	0.60%	0.51%	0.52%	0.42%	0.34%
	<i>Corynebacterium efficiens</i>	3.1	0.49%	0.74%	0.79%	0.76%	0.57%	0.56%	0.35%	0.89%	0.97%	0.96%	0.85%	0.77%	0.90%	0.88%
	<i>Saccharomonospora viridis</i>	4.3	0.19%	0.40%	0.90%	0.41%	0.62%	0.80%	0.42%	0.53%	0.92%	0.97%	0.62%	2.06%	0.68%	2.33%
	<i>Nocardiopsis alba</i>	5.8	0.19%	0.49%	0.77%	0.66%	0.36%	0.54%	0.31%	0.37%	0.77%	0.97%	0.83%	0.76%	0.85%	0.44%
	<i>Psychrobacter arcticus</i>	2.7	0.43%	0.36%	0.43%	0.54%	0.50%	0.35%	0.21%	0.53%	0.48%	0.54%	0.55%	0.72%	0.56%	0.61%
	<i>Leuconostoc mesenteroides</i>	2.0	0.37%	0.27%	0.34%	0.34%	0.76%	0.29%	0.30%	0.39%	0.39%	0.34%	0.42%	0.54%	0.34%	0.82%
	<i>Streptomyces coelicolor</i>	8.7	0.23%	0.38%	0.49%	0.34%	0.35%	0.47%	0.29%	0.38%	0.49%	0.42%	0.40%	0.63%	0.45%	0.48%
Fungus	<i>Aspergillus fumigatus</i> Af293	29.4	0.07%	0.11%	0.70%	0.41%	0.94%	0.71%	0.66%	0.29%	0.64%	1.91%	2.18%	2.58%	1.25%	3.04%
	<i>Saccharomyces cerevisiae</i> S288c	12.1	0.02%	0.04%	0.10%	0.07%	0.37%	0.07%	0.05%	0.06%	0.09%	0.12%	0.12%	0.28%	0.17%	0.47%
Virus	Human adenovirus C	0.036	0.25%	0.36%	0.46%	1.52%	5.33%	4.63%	17.55%	0.16%	0.36%	13.03%	0.56%	0.39%	4.97%	1.75%
	<i>Pseudomonas</i> phage F116	0.065	0.37%	0.62%	1.96%	2.42%	5.78%	5.43%	3.87%	0.03%	0.39%	0.26%	0.98%	0.59%	1.31%	1.38%
	Enterobacteria phage P1	0.095	0.09%	0.16%	0.14%	0.22%	0.22%	0.21%	0.18%	0.09%	0.29%	0.22%	0.18%	0.29%	0.36%	0.29%

Table S4. Summary of PM and meteorological data. The daily temperature, relative humidity, atmospheric pressure, and wind velocity data were 23-hour averages.

Sampling date	Cut-point diameter (μm)	Weather	Temperature ($^{\circ}\text{C}$)	Relative humidity (%)	Atmospheric pressure (Hpa)	Wind velocity (m/s)	Net weight of filters (g)	PM concentration of Tsinghua District ($\mu\text{g}/\text{m}^3$)	PM concentration of Haidian monitoring site ($\mu\text{g}/\text{m}^3$)	AQI of Haidian District		
										BJZWY*	BBXQ*	WL*
01/08	2.5	Clear	-5.03	29.46	1026.36	3.02	0.071	45.21	9.67	83	113	109
	10						0.117	75.03	22.00			
01/09	2.5	Clear	-5.18	44.96	1023.11	1.55	0.163	104.53	80.00	58	NA	77
	10						0.235	150.70	98.33			
01/10	2.5	Overcast	-4.48	66.54	1020.21	1.60	0.411	263.56	288.67	241	242	258
	10						0.543	348.21	321.00			
01/11	2.5	Clear	-5.71	74.88	1018.14	1.06	0.368	235.67	226.67	304	264	413
	10						0.528	338.59	242.67			
01/12	2.5	Misty	-2.75	73.08	1017.49	1.35	0.519	332.82	444.67	360	291	498
	10						0.741	475.18	498.33			
01/13	2.5	Overcast	-4.80	80.75	1018.82	1.20	0.409	261.96	310.33	391	457	401
	10						0.561	359.75	327.67			
01/14	2.5	Clear	-2.38	72.96	1022.15	2.13	0.274	175.39	162.33	280	328	337
	10						0.380	243.68	152.67			

“NA” indicates no available data from the official website of Beijing Municipality Environmental Monitoring Center.

*“BJZWY”, “BBXQ”, “WL” are the abbreviated names of the three monitoring sites in Haidian District.

Table S5. Airborne bacteria and archaea identified at the genus level from Tsinghua PM study and three previous studies combined⁵⁻⁷.

Tsinghua PM study	GRIP, Milan, and New York studies	Overlap
Roseomonas, Chroococcidiopsis, Geodermatophilus, Serinicoccus, Porphyromonas, Streptomyces, Skermanella, Phormidium, Rubellimicrobium, Saccharopolyspora, Friedmanniella, Paracoccus, Alicyclobacillus, Propionicimonas, Aestuariimicrobium, Marmoricola, Deinococcus, Thermoactinomyces, Blastococcus, Nocardioidea, Flavisolibacter, Microlunatus, Kaistobacter, Micromonospora, Cellvibrio, Modestobacter, Paenibacillus, Mesorhizobium, Luteococcus, Archangium, Kocuria, Rhodoplanes, Massilia, Candidatus Microthrix, Actinotalea, UA01, Bacteroides, Pontibacter, Sporosarcina, Arthrobacter, Candidatus Nitrososphaera, Pseudonocardia, Propionibacterium, Planomicrobium, Hymenobacter, Iamia, Gillisia, Arsenicoccus, Tessaracoccus, Sphingomonas, Actinomadura, Bacillus, Rathayibacter, Adhaeribacter, Alkanindiges, Prevotella, Azospirillum, Plesiocystis, Chondromyces, Novosphingobium, Devosia, Mycobacterium, Fusobacterium, Carnobacterium, Actinomycetospira, Turcibacter, Phycococcus, Amaricoccus, Laceyella, Corynebacterium, Planctomyces, Nitrosomonas, Brevundimonas, Psychrobacter, Thiocloava, Myroides, Actinoplanes, Janibacter, Brevibacterium, Alistipes, Clostridium, Desulfovibrio, Epulopiscium, Methylothera, Nannocystis, Herbaspirillum, Azohydromonas, Leptolyngbya, LE30, Cytophaga, Algoriphagus, Oxalobacter, Aeromicrobium, Euzebia, Planococcus, Lysobacter, Methylobacterium, Coprococcus, Anabaena, Myxococcus, Calothrix, Salinicoccus, Ruminococcus, Agrococcus, Promicromonospora, Segetibacter, Lentibacillus, Georgenia, Ramlibacter, Rhodoferrax, Knoellia, Patulibacter, Arcobacter, Allobaculum, Rubrobacter, Treponema, Planifilum, Bifidobacterium, Fibrobacter, Prauserella, Leucobacter, Actinomyces, Microbacterium, Couchioplanes, Flavobacterium, Staphylococcus, Thermobifida, Paludibacter, Bulleidia, Brachyobacterium, Candidatus Solibacter, Jeotgaliococcus, Amycolatopsis, Actinoalloteichus, Methanobrevibacter, Brevibacillus, Hahella, Pedobacter, Nitrososphaera, Virgibacillus, Oerskovia, Gemmata, Oscillatoria, Lactobacillus, Micrococcus, Wautersiella, Microcoleus, Leptospira, Methylobium, Sejongia, Oceanobacillus, Exiguobacterium, Chthoniobacter, Marinilactibacillus, Yaniella, Dietzia, Luteolibacter, Rhodococcus, Asticcacaulis, Megamonas, Dyadobacter, Aerococcus, Rothia, Williamsia, Spirosoma, Enterobacter, Kineococcus, Moraxella, Balneimonas, Weissella, Alkalibacterium, J2-29, Pantoea, Cellulomonas, Caldilinea, Phenylobacterium, Adlercreutzia, Akkermansia, Dehalobacterium, Sphingobium, Parabacteroides, Okibacterium, Pseudomonas, Methylophilus, Shinella, Phascolarctobacterium, Trichococcus, Curtobacterium, Thermomonas, Candidatus Entotheonella, Gemmatimonas, Veillonella, Halomonas, Virgisporangium, Haliangium,	Clostridium, Alkalibacterium, Nocardioidea, Azohydromonas, Georgenia, Ureibacillus, Diaphorobacter, Staphylococcus, Propionibacterium, Ralstonia, Microbacterium, Saccharopolyspora, Rhodococcus, Salinicoccus, Pimelobacter, Stenotrophomonas, Coprococcus, Sphingomonas, Streptococcus, Corynebacterium, Acinetobacter, Pseudomonas, Flavobacterium, Pedobacter, Streptomyces, Mycobacterium, Pantoea, Dyadobacter, Orthomyxococcus, Dietzia, Psychrobacter, Prevotella, Jeotgaliococcus, Acidovorax, Modestobacter, Aquabacterium, Kaistobacter, Friedmanniella, Lactobacillus, Rubellimicrobium, Roseateles, Janthinobacterium, Roseomonas, Deinococcus, Salmonella, Methylobium, Cellulomonas, Aerococcus, Alkanindiges, Bacteroides, Hymenobacter, Geodermatophilus, Bdellovibrio, Curvibacter, Nesterenkonia, Blastococcus, Pontibacter, Kocuria, Planococcus, Aeromonas, Bosea, Massilia, Spirosoma, Fusobacterium, Pseudonocardia, Faecalibacterium, Ruminococcus, Propionicimonas, Chryseobacterium, Bacillus, Lactococcus, Novosphingobium, Microcystis, HTCC2207, Microcoleus, Iamia, Pandoraea, Rhodoferrax, Anaerococcus, Aurantimonas, Skermanella, Rothia, Catenibacterium, Frankia, Treponema, Paracoccus, Salegentibacter, Janibacter, Terracoccus, Agrococcus, Prauserella, Escherichia, Planomicrobium, Dermacoccus, Saccharomonospora, Chroococcidiopsis, Marmoricola, Gordonia, Blautia, Flavisolibacter, Olivibacter, Sphingobacterium, Methylobacterium, Gemella, Amaricoccus, Enterococcus, Chthoniobacter, Paenibacillus, Brevibacterium, Wautersiella, Parabacteroides, Eubacterium, Actinomycetospira, Afipia, Hydrogenophilus, Brachyobacterium, Enhydrobacter, Sphingobium, Cenarchaeum, Burkholderia, Dialister, Micrococcus, Brevundimonas, Bifidobacterium, Roseburia, Oxalobacter, Sejongia, Arthrobacter, Phenylobacterium, Desulfovibrio, Rhizobium, Phormidium, Dokdonella, Alcaligenes, Devosia, Carnobacterium, Glycomyces, Candidatus Xiphinematobacter, Ramlibacter, Cellvibrio, Segetibacter, Pigmentiphaga, Spiroplasma, Micromonospora, Cystobacter, Luteibacter, Marinobacter, Ignatzschineria, Neisseria, Thermobispora, Nocardia, Actinotalea, Halobacterium, Citricoccus, Salinimicrobium, Limnobacter, Alcanivorax, Arsenicoccus, Rubrobacter, Porphyrobacter, Aestuariimicrobium, Mycoplasma, Knoellia, Variovorax, Arcobacter, Tessaracoccus, Helicobacter, Porphyromonas, Yonghaparkia, Peptostreptococcus, Selenomonas, Shinella, Leuconostoc, Gillisia, Finegoldia, Actinomyces, Lamproedia, Paucibacter, Sodalisa, Adhaeribacter, Halomonas, Erythrobacter, Novispirillum, Erwinia, Polaromonas, Lysobacter, PSB-M-3, Nocardioidea, Mesorhizobium, Candidatus Phytoblastia, Planctomyces, Candidatus Rhabdochlamydia, Rhodopirellula, MC18, Candidatus Endobugula, Algoriphagus, Tatlockia, Achromobacter, Balneimonas, Planifilum, Syntrophus,	Roseomonas, Chroococcidiopsis, Geodermatophilus, Serinicoccus, Porphyromonas, Streptomyces, Skermanella, Phormidium, Rubellimicrobium, Saccharopolyspora, Friedmanniella, Paracoccus, Alicyclobacillus, Propionicimonas, Aestuariimicrobium, Marmoricola, Deinococcus, Thermoactinomyces, Blastococcus, Nocardioidea, Flavisolibacter, Microlunatus, Kaistobacter, Micromonospora, Cellvibrio, Modestobacter, Paenibacillus, Mesorhizobium, Luteococcus, Archangium, Kocuria, Rhodoplanes, Massilia, Actinotalea, Bacteroides, Pontibacter, Sporosarcina, Arthrobacter, Candidatus Nitrososphaera, Pseudonocardia, Propionibacterium, Planomicrobium, Hymenobacter, Iamia, Gillisia, Arsenicoccus, Tessaracoccus, Sphingomonas, Actinomadura, Bacillus, Rathayibacter, Adhaeribacter, Alkanindiges, Prevotella, Novosphingobium, Devosia, Mycobacterium, Fusobacterium, Carnobacterium, Actinomycetospira, Turcibacter, Amaricoccus, Corynebacterium, Planctomyces, Nitrosomonas, Brevundimonas, Psychrobacter, Thiocloava, Myroides, Actinoplanes, Janibacter, Brevibacterium, Alistipes, Clostridium, Desulfovibrio, Epulopiscium, Methylothera, Azohydromonas, Leptolyngbya, LE30, Cytophaga, Algoriphagus, Oxalobacter, Algoriphagus, Oxalobacter, Aeromicrobium, Euzebia, Planococcus, Lysobacter, Methylobacterium, Coprococcus, Agrococcus, Myxococcus, Calothrix, Salinicoccus, Ruminococcus, Agrococcus, Promicromonospora, Segetibacter, Lentibacillus, Georgenia, Ramlibacter, Rhodoferrax, Knoellia, Patulibacter, Arcobacter, Allobaculum, Rubrobacter, Treponema, Planifilum, Bifidobacterium, Fibrobacter, Prauserella, Leucobacter, Actinomyces, Microbacterium, Flavobacterium, Staphylococcus, Paludibacter, Bulleidia, Brachyobacterium, Candidatus Solibacter, Jeotgaliococcus, Amycolatopsis, Actinoalloteichus, Methanobrevibacter, Brevibacillus, Pedobacter, Gemmata, Lactobacillus, Micrococcus, Wautersiella, Microcoleus, Methylobium, Sejongia, Oceanobacillus, Exiguobacterium, Chthoniobacter, Marinilactibacillus, Yaniella, Dietzia, Luteolibacter, Rhodococcus, Rhodococcus, Asticcacaulis, Megamonas, Dyadobacter, Aerococcus, Gemmata, Lactobacillus, Micrococcus, Wautersiella, Microcoleus, Kineococcus, Moraxella, Balneimonas, Weissella, Alkalibacterium, J2-29, Pantoea, Cellulomonas, Caldilinea, Phenylobacterium, Adlercreutzia, Akkermansia, Dehalobacterium, Sphingobium, Parabacteroides, Pseudomonas, Shinella, Trichococcus, Curtobacterium, Thermomonas, Candidatus Entotheonella, Gemmatimonas, Veillonella, Halomonas, Haliangium, Glycomyces, Salinimicrobium, Nostoc, Aurantimonas, Rhizobium, Sphingobacterium, Pirellula, Sutterella, Ammoniphilus, Dechloromonas, Eubacterium, Frigoribacterium, Blautia, Terracoccus, Thermobispora, Dokdonella, Aquabacterium, Acinetobacter, Streptacidiphilus, Scytonema, Faecalibacterium, Solirubrobacter, Sharpea, Dermacoccus, Nocardioidea, Caulobacter,

Glycomyces, Salinimicrobium, Nostoc, Aurantimonas, Rhizobium, Sphingobacterium, Pirellula, Sutterella, Ammoniphilus, Dechloromonas, Aeropyrum, Eubacterium, Frigoribacterium, Blautia, Lentzea, Terracoccus, Thermobispora, Dokdonella, Aquabacterium, Jiangella, Acinetobacter, Streptacidiphilus, Gloeobacter, Dermatophilus, Scytonema, Faecalibacterium, Solirubrobacter, Sharpea, Dermacoccus, Shimazuella, Nocardiosis, Caulobacter, Abiotrophia, Mechercharimyces, Actinopolymorpha, Sorangium, Legionella, Caloramator, Roseburia, p-75-a5, Hydrocarboniphaga, Chryseobacterium, Parachlamydia, Rhodocyclus, Pseudoxanthomonas, Simidia, Pimelobacter, Tissierella, Agromyces, Kouleothrix, Waddlia, Thermaerobacter, Cohnella, Candidatus Cloacamonas, Stenotrophomonas, Azonexus, Methanofollis, Methylocaldum, Helcococcus, Frankia, Cellulosimicrobium, Desulfotobacter, ZD0117, Sodalis, Paenisporosarcina, Isoptericola, Shewanella, Hyphomicrobium, Plantibacter, Ectothiorhodospira, Polaromonas, Comamonas, Marinobacter, Achromobacter, Geobacillus, Tsukamurella, Candidatus Rhabdochlamydia, Rhodopirellula, Planktothricoides, Simplicispira, Oceanospirillum, Nocardia, Methylomonas, Clavibacter, Cryobacterium, Marinibacillus, Candidatus Azobacteroides, Jonesia, Azospira, Aquicella, Streptococcus, Bdellovibrio, Saccharomonospora, Acidovorax, Paucibacter, Janthinobacterium, Spirulina, Chloroflexus, Rarobacter, Nitrospira, Lachnospira, Pseudobutyrvibrio, Trablusiella, Arthrobacter, Desulfosporosinus, Capnocytophaga, Oribacterium, Xanthobacter, Thauera, Anaerolinea, Natronorubrum, Hylemonella, Candidatus Odysella, Kaistia, Methylosinus, Solibacillus, Crossiella, Erythromicrobium, Anaeromyxobacter, Rhodopseudomonas, Delftia, Macrococcus, Loktanella, Anaerobiospirillum, Alkaliphilus, Microcella, Pigmentiphaga, MC18, Enterococcus, Oleibacter, Anoxybacillus, u114, Vagococcus, Lactococcus, Candidatus Xiphinematobacter, Lutibacterium, Candidatus Protochlamydia, Chitinophaga, Rubrivivax, Salegentibacter, Methanosarcina, Erythrobacter, Rhodobacter, Acidisoma, Desulfotobacterium, Z4MB24, Aquamonas, Geobacter, Anaerobacillus, Xenococcus, Rheinheimera, Lysinibacillus, Nitrospira, Microbulbifer, Steroidobacter, Niabella, Acetobacter, Gordonia, Edaphobacter, LCP-6, Oscillospira, Verrucomicrobium, Opatutus, Zymomonas, Sporanobacter, Pelotomaculum, Facklamia, Sanguibacter, Planktothrix, WCHB1-05, Bosea, Aeromonas, Erwinia, Kribbella, Bradyrhizobium, Methanospaerula, Thiobacillus, Cryocolla, Methyloversatilis, SHD-231, Roseococcus, Rummelibacillus, Pseudoruegeria, Aminiphilus, Ureibacillus, Thermobacillus, Cryptosporangium, Raoultella, Bartonella, Parapedobacter, Oscillochloris, Desulfobulbus, Inquilinus, Campylobacter, Variovorax, Desulfosporomusa, Labrys, Salinibacterium, Serratia, Luteimonas, Candidatus Koribacter, Mycoplasma, Spiroplasma, Actinokineospora,

Rhodoplanes, Hydrocoleum, Solirubrobacter, Megasphaera, Chloroflexus, Nitrosococcus, Acetobacterium, Oribacterium, Nitrosomonas, Microtetrastora, Fructobacillus, Geobacillus, Cytophaga, Blastochloris, Kineococcus, Desulfosporosinus, Sphingopyxis, Photobacterium, Colwellia, Agrobacterium, Sutterella, Vogesella, LE30, Rubrivivax, Ensifer, SGUS388, Chitinophaga, Proteus, Myroides, Pseudanabaena, Candidatus Solibacter, Demequina, Gemmata, Rhodobacter, Parachlamydia, Isoptericola, Alishewanella, Hyphomicrobium, Butyrvibrio, Rhodocyclus, Rickettsiella, Zymomonas, Fluviicola, Vibrio, Alistipes, Candidatus Liberibacter, Dechloromonas, Thalassobacillus, Candidatus Entotheonella, Methyloversatilis, Aquamonas, Thermomonas, Bulleidia, Sanguibacter, A17, Pseudobutyrvibrio, Agromyces, Methanolinea, Thiobacillus, Williamsia, Thioclava, Cronobacter, SJA-88, Candidatus Odysella, Halobacillus, Shewanella, Parapedobacter, Candidatus Protochlamydia, Caulobacter, Aneurinibacillus, Ideonella, Paludibacter, 4-29, Erythromicrobium, Candidatus Portiera, Exiguobacterium, BHB21, Roseococcus, Methanobrevibacter, Desulfurispirillum, Euzebia, Amycolatopsis, Parvibaculum, Helcococcus, Actinoalloteichus, Pedomicrobium, Singulisphaera, Aeromicrobium, Steroidobacter, Kribbella, Yaniella, Tissierella, A55_D21, Aquicella, Anaeroplasmata, Succinivibrio, RFN20, Facklamia, Azotobacter, Aromatoleum, Cupriavidus, Peptoniphilus, Desemzia, Procabacter, Thermoanaerobacter, Brucella, Luteococcus, Veillonella, Opatutus, Campylobacter, Asticcacaulis, Megamonas, Trichococcus, Candidatus Glomeribacter, Klebsiella, Sharpea, Bradyrhizobium, Luteolibacter, Caldilinea, Runella, Rheinheimera, Lachnobacterium, Zoogloea, Simonsiella, Abiotrophia, Renibacterium, Scytonema, Granulicatella, Leclercia, Mechercharimyces, Xanthomonas, Nitratireductor, Actinopolymorpha, Haemophilus, Hydrogenophaga, Turicibacter, Methylophilus, Weissella, Thauera, Coprobacillus, Collinsella, Slackia, Peptococcus, Eggerthella, Gallionella, Thermoactinomyces, Pseudoclavibacter, Buchnera, Elizabethkingia, Sphaerochaeta, Thiomonas, Leptolyngbya, Alicyclobacillus, Methylothera, Thalassospira, Candidatus Pelagibacter, Adlercreutzia, Epulopiscium, Mycoplasma, Delftia, Maribacter, Rhodobaca, Oceanobacillus, Cyclobacterium, Arenimonas, Pirellula, Oscillospira, Caloramator, Marinimicrobium, Thiothrix, BD2-13, Thermicanus, Legionella, Rhodocista, Hylemonella, Cetobacterium, Sarcina, Akkermansia, Solibacillus, L7A_E11, Actinomadura, Desulfonatronovibrio, Comamonas, Magnetospirillum, Methanocorpusculum, Shigella, Aquiflexum, Simplicispira, Actinokineospora, Limnhabitans, WCHB1-05, Yersinia, Bartonella, Salinibacter, Allobaculum, Verrucomicrobium, Candidatus Koribacter, p-75-a5, Cohnella, Leptotrichia, J2-29, Mucispirillum, Prosthecobacter, Bordetella, Fusibacter, Bacteriovorax, Curtobacterium, Nitrospira, Archangium,

Abiotrophia, Mechercharimyces, Actinopolymorpha, Legionella, Caloramator, Roseburia, p-75-a5, Hydrocarboniphaga, Chryseobacterium, Parachlamydia, Rhodocyclus, Pimelobacter, Tissierella, Agromyces, Kouleothrix, Waddlia, Cohnella, Stenotrophomonas, Helcococcus, Frankia, Cellulosimicrobium, Sodalis, Paenisporosarcina, Isoptericola, Shewanella, Hyphomicrobium, Polaromonas, Comamonas, Marinobacter, Achromobacter, Geobacillus, Candidatus Rhabdochlamydia, Rhodopirellula, Simplicispira, Nocardia, Methylomonas, Marinibacillus, Aquicella, Streptococcus, Bdellovibrio, Saccharomonospora, Acidovorax, Paucibacter, Janthinobacterium, Chloroflexus, Lachnospira, Pseudobutyrvibrio, Desulfosporosinus, Capnocytophaga, Oribacterium, Thauera, Hylemonella, Candidatus Odysella, Solibacillus, Erythromicrobium, Rhodopseudomonas, Delftia, Macrococcus, Pigmentiphaga, MC18, Enterococcus, Lactococcus, Candidatus Xiphinematobacter, Lutibacterium, Candidatus Protochlamydia, Chitinophaga, Rubrivivax, Salegentibacter, Erythrobacter, Rhodobacter, Acidisoma, Aquamonas, Geobacter, Rheinheimera, Nitrospira, Steroidobacter, Acetobacter, Gordonia, Oscillospira, Verrucomicrobium, Opatutus, Zymomonas, Facklamia, Sanguibacter, WCHB1-05, Bosea, Aeromonas, Erwinia, Kribbella, Bradyrhizobium, Thiobacillus, Methyloversatilis, Roseococcus, Ureibacillus, Raoultella, Bartonella, Parapedobacter, Campylobacter, Variovorax, Serratia, Luteimonas, Candidatus Koribacter, Mycoplasma, Spiroplasma, Actinokineospora, Nitratireductor, Rhodobaca, Candidatus Amoebophilus, Catellatospora, Pseudoclavibacter, Burkholderia, Pedomicrobium, Mycoplasma, Anaeroplasmata, A55_D21, Citricoccus, Anaerofustis, Succinivibrio, Aromatoleum, Salmonella, Frateuria, Butyrvibrio, A17, Zoogloea, Singulisphaera, Pseudaminobacter, Phyllobacterium, Diaphorobacter, Hydrocoleum, Blastochloris, Halobacillus, Granulicatella, Leuconostoc, Runella, Gemella, Agrobacterium, Haererehalobacter, Methylophilus, Anaerococcus, Saccharothrix, Afipia, Gallionella, Pseudidiomarina, Ideonella, RFN20, Neisseria, Ralstonia, SJA-88, Halobacterium, Xanthomonas, Coprobacillus, Azotobacter, Zhihengliuella, BD2-13, L7A_E11, Kineospora, Arcanobacterium, Leptotrichia, Cyclobacterium, Peptococcus, Megasphaera, Demequina, Sphaerochaeta, Alcanivorax, Methanolinea, Aquiflexum, Dolichospermum, Arenimonas, Muricauda, Kushneria, Solimonas, Nesterenkonina, Mitsuoella, Curvibacter, Yonghaparkia, Citrobacter, Dysgonomonas, Ignatzschineria, Dyella, Ochrobactrum, Streptobacillus, Symbiobacterium, Actinoallomurus, Pseudanabaena, Meiothermus, Collinsella, Mucispirillum, Lachnobacterium, Prochlorococcus, Procabacter, Emticicia, Klebsiella, Pseudoalteromonas, Sphingopyxis, Conchiformibius, Syntrophomonas, Teredinibacter, Nitrosococcus, Cupriavidus, Olivibacter, Cystobacter, Alcaligenes, Tatlockia, Sarcina, Haloanella, Caedibacter, Desemzia, Peptoniphilus, Acholeplasma,

Candidatus Methanoregula, Synechococcus, Nitratireductor, Streptobacillus, Wolbachia, Symbiobacterium, Desulfonatronum, Aneurinibacillus, Acetivibrio, Pelomonas, Balneola, Luteibacter, Nonomuraea, Rhodobaca, Candidatus Amoebophilus, Actinoallomurus, Rhodopseudomonas, Pseudoalteromonas, Luedemannella, Desulfotomaculum, Xylanimicrobium, Catellatospora, Pseudoclavibacter, Burkholderia, Pilmelia, Meiothermus, Dehalobacterium, Alteromonas, Waddlia, Actinobaculum, Renibacterium, Marinimicrobium, Syntrophus, Pedomicrobium, Mycoplasma, Atopobium, Anaeroplasm, Nitrosopumilus, Congregibacter, Oceanithermus, Prochlorococcus, Gelidibacter, Vibrio, Ensifer, Thalassobacillus, Desulfobacterium, Myceligenerans, Salinisphaera, A55_D21, Sedimentibacter, Pelagicoccus, Dysgonomonas, Actinoplanes, Phyllobacterium, Eggerthella, Phaeospirillum, Magnetospirillum, Fluviicola, Citricoccus, Anaerofustis, Succinivibrio, Aromatoleum, Crenothrix, Acidobacterium, Acidisoma, nsmpVI18, Lewinella, Candidatus Liberibacter, 4-29, SGUS388, Ornithinococcus, Salmonella, Frateuria, Butyrivibrio, Collimonas, A17, Conchiformibus, Dolichospermum, Saccharothrix, Fusibacter, Microcystis, Slackia, Thermicanus, Thiothrix, Sinorhizobium, Rhodanobacter, Averyella, Zoogloea, Sulcia, Teredinibacter, Cellulosimicrobium, Mitsuokella, Aquitalea, Spirochaeta, Verrucosipora, Selenomonas, Leclercia, Singulisphaera, Pseudaminobacter, Phyllobacterium, Lautropia, Serinicoccus, Polaribacter, Dyella, Pelomonas, Oleispira, Porphyrobacter, Cetobacterium, Congregibacter, Prosthecobacter, Diaphorobacter, Methylophaga, Hydrocoleum, Blastochloris, Plesiomonas, Lutibacterium, Emticicia, Spirochaeta, Acholeplasma, Pandoraea, Rickettsiella, Kurthia, Thiomonas, Flectobacillus, Schlegelella, Halobacillus, Granulicatella, Leuconostoc, Runella, Haloplanus, Caedibacter, Ammoniphilus, Anaerofustis, Candidatus Phytolasma, PSB-M-3, Desulfomicrobium, Candidatus Gemella, Agrobacterium, Haererehalobacter, Catonella, Verminephrobacter, Methylomonas, Brevibacillus, Catellatospora, Mannheimia, Alysia, Methylophaga, Xenophilus, Desulfobacca, Pseudidiomarina, Acetivibrio, Citrobacter, Luedemannella, Anaerococcus, Saccharothrix, Dorea, Methylobacillus, Caldicellulosiruptor, Desulfotomaculum, Dehalobacter, Sporocytophaga, Afipia, Gallionella, Rhodovibrio, HB2-32-21, Sporosarcina, Flectobacillus, Frigoribacterium, Kouleothrix, Pseudidiomarina, Ideonella, RFN20, Neisseria, Ralstonia, SJA-88, Actinobaculum, Arcanobacterium, Leucobacter, Streptacidiphilus, Nordella, Halobacterium, Halorubrum, Xanthomonas, Frateuria, Gemmatimonas, Acidimicrobium, Kineosporia, Syntrophomonas, Sphaerotilus, Serratia, Desulfobacterium, Sphaerospermopsis, Coprobacillus, Kibdelosporangium, Gelidibacter, Nostoc, Luteimonas, Candidatus Amoebophilus, Azotobacter, Zhihengliuella, BD2-13, L7A_E11, Dethiobacter, Geobacter, Haliangium, Myxococcus, Desulfomicrobium, Kineosporia, Arcanobacterium, Candidatus Scalindua, Leptotrichia, Beijerinckia, Cardiobacterium, Fibrobacter, Blastomonas, Nevskia, Cyclobacterium, Belliella, Peptococcus, Megasphaera, Demequina, Methylocystis, Halostagnicola, Enterobacter, Cycloclasticus, Sphaerochaeta, Alcanivorax, Methanolinea, Aequorivita, Fulvivirga, Halococcus, Macrococcus, Phaeospirillum, Pseudaminobacter, Aquiflexum, Dolichospermum, Arenimonas, Muricauda, Solimonas, Salimicrobium, Marinococcus, Lachnospira, Streptomonospora, Kushneria, Solimonas, Methylococcus, Halalkalicoccus, Candidatus Nitrososphaera, Rathayibacter, Thermomonospora, Chelativorans, Olsenella, Nesterenkonia, Natronococcus, Blattabacterium, Idiomarina, Marinibacillus, Thermaetogonium, Mitsuokella, Thermus, Wolinella, Curvibacter, Tetrathiobacter, Marinilactibacillus, Balneola, Marivita, Yonghaparkia, Leucothrix, Citrobacter, Terribacillus, Zhihengliuella, Microlunatus, Azorhizobium, Haererehalobacter, Chthonomonas, Dysgonomonas, Ignatzschineria, Dyella, Verrucosipora, Moraxella, Acetobacter, Raoultella, Ochrobactrum, Halomiconema, Streptosporangium, Hydrocarboniphaga, Haloanella, Xylanimicrobium, Desulfocapsa, Streptobacillus, Symbiobacterium, Actinoallomurus, Muricauda, Ochrobactrum, Capnocytophaga, Paenisporosarcina, Pseudanabaena, Meiothermus, Collinsella, Tenacibaculum, Kurthia, Insolitispirillum, Kushneria, Patulibacter, Mucispirillum, Lachnobacterium, Polynucleobacter, Prochlorococcus, Procabacter, Emticicia, Acidiphilium, Marinomonas, Klebsiella, Erysipelothrix, Pseudoalteromonas, Ruegeria, Sphingopyxis, Conchiformibus, Actinocorallia, Syntrophomonas, Aminobacter, Teredinibacter, Sinomonas, Hydrogenobaculum, Nitrosococcus, Rivularia, Cupriavidus, Azomonas, Olivibacter, Ancylobacter, Sulfuricurvum, Methanosphaera, Cystobacter, Terriglobus, Azorhizophilus, Alcaligenes, Tatlockia, Sarcina, Haloanella, Caldimonas, Asteroleplasma, Caedibacter, Desemzia, Kitasatospora, Peptoniphilus, Acholeplasma, Heliorestis, Aneurinibacillus, Niastella, Acetivibrio, Pelomonas, Balneola, Luteibacter, Methanocella, Luedemannella, Oceanisphaera, Desulfotomaculum, Trichodesmium, Xylanimicrobium, Moorella, Skermania, Anaerofilum, Providencia, Dactylosporangium, Gardnerella, Catenulispora, Actinobaculum, Renibacterium, Gluconacetobacter, Candidatus Aquirestis, Sporichthya, Coriobacterium, Sulfurospirillum, Candidatus Contubernalis, Marinimicrobium,

Syntrophus, Gluconobacter, Gelidibacter, Thermosynechococcus, Vibrio, Stigmatella, Ensifer, Oceanimonas, Thalassobacillus, Martellella, Buttiauxella, Segniliparus, Methyloarcula, Aphanothece, Desulfobacterium, Eggerthella, Aphanizomenon, Phaeospirillum, Marinospirillum, Octadecabacter, Tropheryma, JG37-AG-70, Fischerella, Azoarcus, Sphaerisporangium, Magnetospirillum, Fluviicola, Candidatus Liberibacter, Chroococcus, Actinocatenispora, Mariprofundus, Avibacterium, 4-29, SGUS388, Candidatus Aquiluna, Candidatus Cardinium, Ornithinococcus, Rhodovulum, Fusibacter, Telmatospirillum, Microcystis, Slackia, Symploca, Thermicanus, Dermabacter, Thiothrix, Spirochaeta, Ferrimicrobium, Brachyspira, Salinispora, Acrocarpospora, Psychroserpens, Verrucosipora, Selenomonas, Leclercia, Leptospirillum, Thioalkalivibrio, Thermosinus, Porphyrobacter, Natronincola, Cetobacterium, Sneathia, Ignavibacterium, Coralloccoccus, T78, YNPFP6, Microbispora, Candidatus Tammella, Rhodothermus, Thiohalospira, Arthronema, Brachymonas, Congregibacter, Haloferula, Prostheco bacter, Rhodobium, Halochromatium, Prochloron, Jannaschia, Methanobacterium, Pandoraea, Saccharospirillum, Halanaerobium, Magnetococcus, Umboniibacter, Rickettsiella, Psychromonas, Kurthia, Dichelobacter, Tetrasphaera, Thiomonas, Thermodesulfobivibrio, Thermovenabulum, Thermincola, Flectobacillus, Actinosynnema, Actinopolyspora, Pediococcus, Phaeobacter, Gloeotheca, Candidatus Phytoplasma, PSB-M-3, Rickettsia, Pelobacter, Piscirickettsia, Desulfomicrobium, Candidatus Portiera

Table S6. Public data of metagenomic studies used in PCA analysis (relative abundance as the input variables).

Habitat	Project Name	Project link	Sample ID
Bloom lake	Freshwater Microcystis Bloom Metagenomes	http://metagenomics.anl.gov/linkin.cgi?project=1207	4467058.3, 4467059.3
Agricultural soil	EarlhamMetagenomes2012	http://metagenomics.anl.gov/linkin.cgi?project=2850	4508937.3, 4508938.3, 4508939.3, 4508940.3, 4508941.3, 4508942.3
Open ocean	Global Ocean Sampling Expedition	http://metagenomics.anl.gov/linkin.cgi?project=27	4441126.3, 4441129.3, 4441139.4, 4441615.3, 4441576.3, 4441578.3, 4441592.3, 4441661.3, 4441588.3, 4441594.3
Coastal ocean	Global Ocean Sampling Expedition	http://metagenomics.anl.gov/linkin.cgi?project=27	4441580.3, 4441583.3, 4441148.3, 4441613.3, 4441596.4, 4441601.3, 4441586.3, 4441660.3, 4441591.3, 4441605.3
Anarobic sludges	Beer-to-Caproate	http://metagenomics.anl.gov/linkin.cgi?project=397	4480719.3, 4480764.3, 4480776.3, 4480815.3, 4480859.3, 4480861.3, 4480863.3, 4480864.3, 4480865.3, 4480867.3

Table S7. Annotations of the 48 most common bacterial, 2 fungal, and 3 viral species.

Organism	Species	Aerobic /anaerobic	Descriptions	References
Bacteria	<i>Geodermatophilus obscurus</i>	aerobic	soil and rock varnish, lytic capability of yeast cell walls, UV-C resistance, strong production of extracellular functional amyloid and manganese oxidation	⁹
	<i>Modestobacter marinus</i>	aerobic	predominant in the first two millimeters of calcarenite stone surfaces, access to various trace carbon sources, availability highly resistant to desiccation and to gamma- and high-energy UV radiations	¹⁰
	<i>Blastococcus saxosidens</i>	aerobic	thriving in inhospitable and bleak biotopes as the interior of stones, thermophilic esterases, resistance to several metals higher than those of its phylogenetic relatives, low resistance to several oxidizing stresses	¹¹
	<i>Kocuria rhizophila</i>	aerobic	halotolerant, tolerant to a wide variety of organic solvents, ability to grow rapidly and at high cell density, robust at various growth conditions	¹²
	<i>Micrococcus luteus</i>	aerobic	diverse chemotaxonomic features dormancy, able to concentrate heavy metals from low-grade ores, Fleming strain	¹³
	<i>Candidatus Nitrospira defluvii</i>	aerobic	uncultivated nitrite-oxidizing bacteria, resistance to ampicillin	¹⁴
	<i>Methylobacterium radiotolerans</i>	aerobic	multilayered cell wall structure, facultative methylotroph, high resistance to gamma radiation	¹⁵
	<i>Thermobifida fusca</i>	aerobic	moderately thermophilic, filamentous, a major degrader of plant cell walls in heated organic materials, spores can be allergenic, associated with farmers lung	¹⁶
	<i>Nocardioides</i> sp.	aerobic	ability to metabolize unusual substrates, including butane, jet fuel, phenanthrene, p-nitrophenol, trinitrophenol, atrazine, ethene, and vinyl chloride	¹⁷
	<i>Propionibacterium acnes</i>	anaerobic	a commensal organism on human skin, implicated in various infections including acne vulgaris, endocarditis, osteomyelitis, and prostate cancer; the only microorganism that has been isolated from sarcoid lesions by bacterial culture	¹⁸
	<i>Brachybacterium faecium</i>	aerobic	capable of degrading uric acid and fermenting cellobiose, glucose, maltose, and man-nose, but not cellulose, chitin, or gelatin	¹⁹
	<i>Cellvibrio gilvus</i>	aerobic	isolated from bovine feces, cellulolytic, disaccharide preference	²⁰
	<i>Arthrobacter phenanthrenivorans</i>	aerobic	phenanthrene and anthracene as sole carbon sources, isolated from a creosote-contaminated site	²¹
	<i>Nocardiopsis dassonvillei</i>	aerobic	isolated from a large variety of natural habitats such as soil and marine sediments, from different plant and animal materials as well as from human patients, participating actively in biopolymer degradation, associated with actinomycetoma, conjunctivitis, cholangitis	²²
	<i>Carnobacterium</i> sp.	facultative	ubiquitous lactic acid bacteria, tolerant to freezing/thawing and high pressure, able to grow at low temperatures	²³
	<i>Microbacterium testaceum</i>	aerobic	endophytic within plant hosts, N-acylhomoserine lactone-degrading activity	²⁴
	<i>Paracoccus denitrificans</i>	facultative	capable of thriving in soil, ability to single-handedly convert nitrate to dinitrogen via denitrification	²⁵
	<i>Kytococcus sedentarius</i>	aerobic	production of oligoketide antibiotics, opportunistic pathogen causing valve endocarditis, hemorrhagic pneumonia, and pitted keratolysis	²⁶
	<i>Sanguibacter keddiei</i>	facultatively anaerobic	blood isolate, short, irregular rods, mesophilic	²⁷
	<i>Kineococcus radiotolerans</i>	aerobic	isolated from high-level radioactive waste, radiation and desiccation resistance	²⁸

<i>Pantoeaananatis</i>	aerobic	epiphyte or endophyte, an emerging phytopathogen infecting a wide range of important crop and forest plants, an occasional clinical isolate and a presumptive opportunistic human pathogen, associated with septicemia following penetrating trauma with plant material, nosocomial infections due to exposure to contaminated hospital materials, and secondary complications of preexisting illnesses	²⁹
<i>Lactobacillus johnsonii</i>	facultatively anaerobic	acidophilus, attachment to epithelial cells, immunomodulation, competitive exclusion of pathogens	³⁰
<i>Cellulomonas fimi</i>	facultatively anaerobic	cellulolytic, secreting multi-domain glycoside hydrolases	³¹
<i>Lactobacillus crispatus</i>	aerobic	beneficial microbiota present in the vertebrate gastrointestinal and human genitourinary tracts	³²
<i>Deinococcus gobiensis</i>	facultatively anaerobic	higher tolerance to gamma radiation and UV light than all other known microorganisms, isolated from desert	³³
<i>Thauera</i> sp.	facultative	isolated from activated sludge samples from industrial wastewater, ability to produce abundant exopolysaccharide and degrade various aromatic compounds with nitrate	³⁴
<i>Pantoea vagans</i>	facultatively anaerobic	enterobacterial plant epiphyte of a broad range of plants, providing effective disease control of fire blight	³⁵
<i>Clostridium perfringens</i>	anaerobic	a common inhabitant of the avian and mammalian gastrointestinal tracts, commensal or pathogenical	³⁶
<i>Lactobacillus reuteri</i>	facultatively anaerobic	heterofermentative lactic acid bacterium that naturally inhabits the gut of humans and other animals, production of the broad-spectrum antimicrobial compound reuterin during anaerobic metabolism of glycerol	³⁷
<i>Pseudomonas stutzeri</i>	facultative	ability to use maltose and starch as sole carbon and energy sources, degradation of environmental pollutants and high-molecular-weight polyethylene glycols, production of dinitrogen gas from nitrate, associated with human infections	³⁸
<i>Psychrobacter cryohalolentis</i>	aerobic	capable of growth at temperatures from -10°C to 30°C, isolated from permafrost	³⁹
<i>Lactobacillus salivarius</i>	anaerobic	part of the indigenous microbiota of the gastrointestinal tract and oral cavity of humans and hamsters, isolated from human breast milk and from the intestinal tracts of swine and chickens	⁴⁰
<i>Bacillus megaterium</i>	aerobic	with a cell length of up to 4 µm and a diameter of 1.5 µm, nonpathogenic host for the biotechnological production of several substances, including vitamin B12, penicillin acylase, and amylases	⁴¹
<i>Arthrobacter arilaitensis</i>	aerobic	one of the main bacterial species found at the surface of cheeses, especially in smear-ripened cheeses, where it contributes to the typical colour, flavour and texture properties of the final product	⁴²
<i>Ramlibacter tataouinensis</i>	aerobic	chemo-organotrophic, cyst-producing, first isolated from meteorite fragments buried in the sands of a desert	⁴³
<i>Nakamurella multipartita</i>	aerobic	polysaccharide-accumulating, septa-forming, isolated from activated sludge acclimated with sugar-containing synthetic waste water	⁴⁴
<i>Streptococcus infantarius</i>	facultatively anaerobic	ability to grow in the presence of bile, production of β-glucosidase leading to hydrolysis of aesculin	⁴⁵
<i>Mycobacterium gilvum</i>	aerobic	pleomorphic, acid-fast, resistant to isoniazid, sodium aminosalicilate and rifampicin	⁴⁶
<i>Arthrobacter chlorophenolicus</i>	aerobic	growth on and degradation of high concentrations of 4-chlorophenol as the sole carbon and	⁴⁷

		energy source, isolated from soil	
	Methylobacterium extorquens	aerobic	ability to grow on one or several reduced one carbon (C1) compounds other than methane, associated with plants 48
	Methylobacterium chloromethanicum	aerobic	isolated from soil at a petrochemical factory in Tatarstan, Russia, able to grow using chloromethane as the sole carbon and energy source 49
	Corynebacterium efficiens	facultatively anaerobic	thermostability, glutamic-acid-producing, isolated from soil and vegetables 50
	Stenotrophomonas maltophilia	aerobic	commonly found in water and soil in association with plants, beneficial effects on plants including protection from pathogens, promotion of growth, and biodegradation of pollutants; an emerging opportunistic human pathogen involved in infections at hospitals and in patients with cystic fibrosis 51
	Nocardiopsis alba	aerobic	inhibitory bioactivity against several Bacillus isolates, some Gram-positive human pathogens, or honeybee pathogens 52
	Psychrobacter arcticus	facultative	capable of growth at temperatures from -10°C to 28°C, isolated from permafrost 39
	Saccharomonospora viridis	aerobic	thermophile, spores can cause farmer's lung disease, bagassosis and humidifier fever, pentachlorophenol metabolism 53
	Streptomyces coelicolor	aerobic	broad range of metabolic processes and biotransformations, soil-dwelling, production of most natural antibiotics used in human and veterinary medicine 54
	Leuconostoc mesenteroides	facultative	lactic acid bacteria, key players responsible for kimchi fermentation 55
Fungus	Aspergillus fumigatus	aerobic	prolific conidia production, isolated from human habitats and vegetable compost heaps, a primary and opportunistic pathogen, a major allergen, linked to severe asthma and sinusitis 56
	Saccharomyces cerevisiae	facultative	domesticated organism that occasionally establishes synanthropic natural populations, capable of malolactic fermentation, able to form biofilms 57
Virus	Pseudomonas phage F116		temperate, pilus-specific, generalized transducing phage, linear dsDNA 58
	Human adenovirus C		non-enveloped, icosahedral, linear dsDNA, causing a self-limited febrile illness 59
	Enterobacteria phage P1		lysogenizing enteric bacteria as a circular, low-copy-number plasmid 60

Table S8. Primers and Sanger sequencing results.

Primers	Sequence
funga118sF	5'GATACCGTYGTAGTCTTA3'
funga118sR	5'TGTCTGGACCTGGTGAGT3'
gliIRTF	5'TCGTTGCTGGAGAATCTGCTGTATGA3'
gliIRTR	5'ACAAAGGAATCTGGTGGAAAGGCGA3'
18s RNA gene PCR product	GATACCGTCGTAGTCTTAACCATAAACTATGCCGACTAGGGATCGGGCGG TGTTTCTATGATGACCCGCTCGGCACCTTACGAGAAATCAAAGTTTTTGGG TTCTGGGGGAGTATGGTCGCAAGGCTGAAACTTAAAGAAAATTGACGGAA GGCACCACAAGGCGTGGAGCCTGCGGCTTAATTTGACTCAACACGGGGA AACTCACCAGTCCAGACA
<i>gliI</i> gene PCR product	TCGTTGCTGGAGAATCTGCTGTATGACATCTGCGAGCCGGGCAATTGCGAC GGGGAGGCGTGTGATCGAGACGCCGTTCTGGGGTGCCTACCATCGGGGG TTTCGTTGGCACCAGCATGCATGGTCTTAGCTGATGTATACGAATAGGGGGC TTCGAGACGAGCTTCGTCCTGCGCTCCAACGTCACCGCTGTTCATGTGAGG CCTCCGTGTCATGGCAATGGATCCGCAGATCTGGATCGTCTCGTGTGGCG TATATTGAAGCGTACGAACGGGTTCTGCGTCAGGCCCCCTGCCGCATCAAG GCCATCCTCGTGTGTAACCCGCACAATCCATGCGGGCACATCTACCCACCG AGAGTAATCCAGGCCTTGCTTCAATTCGCCAACGGCATGATCTATTCTAC ATCTCCGATGAGATCTACGCCCTCTCCACGCTGGATGAGCAAACCGCCTTC ACCTCGGTCCTGAGCATCGATGTAGCCGCCCTGGGTGTAGACCTGGCGCGC GTATTCACCCTGTACAGTATCAGCAAAGACCTCGGCAGTAGCGGATTGCG ACTGGTATACTGTCTTATCTCCTCCGACTCTCAAGGCCTCCGCAGAACTGA CACTGTGCAGGGCTTCGGCATCACCAAGCACATCCCGACCTGCGTCTCTC CCTCGCCATCTCGAACCATTCCCGCGTCTCGACATTCACCTCGCTGGTCAT CACCGCTCTCCTGCACGACCCGGAAGCCGCCACGGCAATCCTGCACCAGA ATCGAGCCGCTCTGCAGCGCAGCGCCAGCTCATCAGAAAACCTTCTCGCCT CACGACCGTTGGGAGAGAGTGAGAGAGAAATCGCCTTCCACCAGATTCCC TTTGT

Table S9. Estimated concentrations of organic carbon and elemental carbon measured in PM_{2.5} and PM₁₀ samples.

Sampling date	PM _{2.5}		PM ₁₀	
	OC (µg/m ³)	EC (µg/m ³)	OC (µg/m ³)	EC (µg/m ³)
01/08	6.753	2.151	9.93	3.283
01/09	28.537	6.351	36.421	6.904
01/10	71.036	13.514	59.588	8.186
01/11	45.022	12.42	53.829	8.12
01/12	68.733	13.667	68.814	8.774
01/13	49.18	9.452	53.9	7.588
01/14	25.429	6.005	32.816	6.068

Table S10. Estimated concentrations of anions and cations measured in PM_{2.5} and PM₁₀ samples.

Sampling date	01/08		01/09		01/10		01/11		01/12		01/13		01/14	
	PM _{2.5}	PM ₁₀	PM _{2.5}	PM ₁₀	PM _{2.5}	PM ₁₀	PM _{2.5}	PM ₁₀	PM _{2.5}	PM ₁₀	PM _{2.5}	PM ₁₀	PM _{2.5}	PM ₁₀
SO ₄ ²⁻ (µg/m ³)	3.208	4.256	13.193	16.832	90.795	58.364	57.511	81.258	105.016	111.108	70.914	92.471	43.381	57.847
NH ₄ ⁺ (µg/m ³)	5.202	5.698	18.982	21.078	62.365	45.588	44.328	54.280	62.925	63.475	48.271	55.871	36.012	39.618
NO ₃ ⁻ (µg/m ³)	1.219	1.526	11.616	14.084	52.437	37.954	32.293	49.089	48.984	52.278	36.509	50.547	25.074	33.535
K ⁺ (µg/m ³)	0.681	0.928	3.553	4.451	14.174	9.214	9.331	12.663	17.149	16.621	19.077	22.134	8.691	11.241
Cl ⁻ (µg/m ³)	0.960	2.285	4.022	6.930	15.694	12.604	8.489	16.881	14.320	18.409	10.072	16.228	5.571	9.740
Na ⁺ (µg/m ³)	1.318	4.213	2.857	8.326	6.924	10.426	5.275	17.956	8.265	16.884	5.963	13.238	3.945	12.261
Ca ²⁺ (µg/m ³)	2.566	10.394	2.276	14.256	2.913	15.906	1.758	21.257	3.067	16.324	1.613	17.054	1.376	11.268
Mg ²⁺ (µg/m ³)	0.519	0.854	0.524	1.570	0.968	2.471	0.491	3.149	0.828	2.851	0.737	3.444	0.504	2.758
F ⁻ (µg/m ³)	0.060	0.124	0.181	0.442	0.505	0.906	0.270	0.943	0.417	0.741	0.352	1.043	0.148	0.400
PO ₄ ³⁻ (µg/m ³)	0.021	0.031	0.035	0.047	0.070	0.068	0.050	0.080	0.070	0.090	0.051	0.076	0.042	0.061

Table S11. Estimated concentrations of elements measured in PM_{2.5} and PM₁₀ samples.

Sampling date	01/08		01/09		01/10		01/11		01/12		01/13		01/14	
	PM _{2.5}	PM ₁₀	PM _{2.5}	PM ₁₀	PM _{2.5}	PM ₁₀	PM _{2.5}	PM ₁₀	PM _{2.5}	PM ₁₀	PM _{2.5}	PM ₁₀	PM _{2.5}	PM ₁₀
Fe (µg/m ³)	0.249	1.844	0.482	2.479	1.302	3.389	0.824	3.496	2.191	4.543	2.042	8.048	1.016	4.564
Zn (µg/m ³)	0.046	0.068	0.285	0.428	1.227	1.022	0.586	0.842	1.695	1.585	1.104	1.686	0.561	0.870
Pb (µg/m ³)	0.032	0.040	0.164	0.221	0.636	0.452	0.352	0.437	1.114	0.890	1.033	1.271	0.449	0.576
As (ug/m ³)	0.000	0.002	0.031	0.051	0.178	0.168	0.077	0.095	0.237	0.221	0.115	0.139	0.067	0.092
Mn (µg/m ³)	0.000	0.064	0.056	0.110	0.147	0.136	0.085	0.057	0.141	0.181	0.002	0.289	0.080	0.157
Br (µg/m ³)	0.004	0.003	0.025	0.032	0.107	0.067	0.061	0.065	0.139	0.110	0.087	0.115	0.041	0.057
Ti (µg/m ³)	0.020	0.145	0.028	0.190	0.063	0.233	0.029	0.218	0.076	0.240	0.042	0.253	0.025	0.170
Cr (µg/m ³)	0.001	0.006	0.006	0.015	0.018	0.021	0.010	0.020	0.020	0.028	0.020	0.039	0.006	0.020
Ni (µg/m ³)	0.001	0.008	0.003	0.008	0.008	0.005	0.005	0.010	0.007	0.012	0.012	0.021	0.003	0.007

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