## **Selective sweeps at autosomal loci with gene conversion**

The expected reduction in diversity at a neutral site immediately following a sweep by a selectively favorable allele with intermediate dominance, measured relative to the initial diversity,  $\pi_0$ , is given by

$$
\frac{\Delta \pi}{\pi_0} \approx (2N_e s)^{-\frac{4r}{s}} \tag{A1}
$$

where *s* is the selection coefficient on homozygotes for the favored allele, and *r* is the frequency of recombination between this allele and the neutral site (Charlesworth and Charlesworth 2010, p. 409).

If only gene conversion is acting, *r* is approximately twice the product of the initiation rate and the tract length of the events involved, provided that the two sites are sufficiently far apart that they are not included in the same gene conversion tract. This represents two possible types of event; a gene conversion event switching the selected allele onto an ancestral haplotype, and a gene conversion event switching a variant at the neutral site onto a haplotype carrying the selected allele. If conversion is random in direction, each of these occurs at one half of the rate of the relevant gene conversion events themselves, so that we can equate the net rate of events for an autosome to one-half the product of mean tract length and the probability of initiation of an event in female meiosis, thus correcting for the absence of events in males.

For purposes of calculation, it is convenient to rewrite equation (A1) as:

$$
-\left(\frac{s}{4r}\right) \ln\left(\frac{\Delta \pi}{\pi_0}\right) \approx \ln(2N_e) + \ln(s) \tag{A2}
$$

Comeron et al. (2012) estimated the mean gene conversion tract length as 518bp and the rate of initiation of events as  $1.25 \times 10^{-7}$  per female meiosis, giving a value of *r* for an autosome of approximately  $3.2 \times 10^{-5}$  From the mean synonymous site diversity for autosomal recombining regions reported here (0.014) and the estimate of  $5 \times 10^{-9}$  for the per basepair mutation rate per generation in *D. melanogaster* obtained as the mean over the values for the three different strains studied by Keightley et al. (2009) and Schrider et

al. (2013), we have  $N_e = 7.0 \times 10^5$ . Given that the estimated reduction in diversity in autosomal NC regions of *Drosophila* species in the various studies cited in the main text is 85% or more of the value for autosomal C regions, we can equate  $-\Delta \pi / \pi_0$  to 0.85, so that equation (A2) becomes

$$
1270s = 14.15 + \ln(s) \tag{A3}
$$

This implies that *s* is approximately 0.0075. Since the observed diversity includes a contribution for mutations that arose after the sweep, this represents the minimal value of *s* required to explain the data (even larger values would be required for soft rather than hard sweeps).

These results also assume that only a single sweep is in progress at a given time in a given NC, which is not necessarily the case (see the next section). This assumption is probably conservative, as can be seen as follows. If a second favorable mutation initiates a sweep while the first one is in progress to fixation, it will affect the final outcome with respect to level of variability only if it arises on a haplotype that lacks the first mutation. In that case, by driving this haplotype to a higher frequency than it would reach in its absence, the second favorable mutation will increase the opportunity for gene conversion events to exchange variants between haplotypes carrying the two favorable mutations, thereby increasing the final level of variability after fixation of one of the two favorable mutations over that given by equation (A3)

## **Can there be multiple sweeps in the autosomal NC regions?**

The maximal value of  $\omega_a$  for autosomal regions is approximately 0.08 (Table S4 of Supplementary Material 1); given a mutation rate per synonymous site of  $5 \times 10^{-9}$  (see above), this implies a substitution rate for positively selected nonsynonymous mutations of  $4 \times 10^{-10}$  per nonsynonymous site. The average number of codons per gene is 454 for the autosomal NC region (Campos et al. 2012); a generous estimate of the number of genes in a given NC region is 100 (Table 2). With 70% of coding sequence mutations causing an amino-acid change (Misra et al. 2002), this gives an estimate of  $1.27 \times 10^{-5}$ adaptive nonsynonymous substitutions per generation in an NC region, in the absence of

any HRI effects. Adaptive non-coding sequence mutations may be equally important (Sella et al. 2009), so the net rate of adaptive fixations in an NC region could be as high as 2.54  $\times$  10<sup>-5</sup>.

Previous theoretical work has shown that beneficial mutations will establish themselves in the population without mutual interference if the mean time between the occurrence of mutations that get fixed by selection is less than the time it takes for a beneficial mutation that has established itself to spread to a high frequency or fixation, which is approximately  $\ln(4N_e s_h)/s_h$  for semidominant mutations (Maynard Smith 1971; Desai and Fisher 2007), i.e. HRI due to competing beneficial mutations requires the reciprocal of the rate of substitution,  $K_a$ , of adaptive mutations in the NC regions to be smaller than the mean of  $\ln(4N_e s_h)/s_h$ . With the above estimate of  $s = 0.0075$  to account for the observed reduction in diversity from a selective sweep with gene conversion, and with  $N_e = 7.0 \times 10^5$ ,  $\ln(4N_e s_h)/s_h$  is approximately 1.3 x 10<sup>3</sup>, which is far smaller than  $1/(2.54 \times 10^{-5}) = 3.9 \times 10^{4}$ . It is therefore very difficult to reconcile the observed rates of gene conversion and reduction in synonymous diversity in NC regions with a model in which clonal interference is solely responsible for the reduced level of adaptive evolution. This does not, of course, rule out possible joint effects of background selection and clonal interference.

## **Effects of weak selection on site frequency spectra**

The model used is similar to that studied numerically by Zeng and Charlesworth (2009). It assumes a randomly mating, diploid, discrete-generation population with effective population size  $N_e$ . Over a long sequence of nucleotide sites, each site has two alternative types,  $A_1$  and  $A_2$ , with mutation rates *u* and *v* from  $A_1$  to  $A_2$  and vice versa. ( $A_1$  and  $A_2$ ) can be taken to correspond to unpreferred versus preferred synonymous codons, respectively.) The mutational bias parameter,  $\kappa$ , is equal to  $v/u$ . If selection is acting, semidominance is assumed, with  $A_2$  having a selective advantage *s* over  $A_1$  when homozygous.

If the population is at statistical equilibrium, the mean numbers of sites in each state are constant over time, despite continual changes in frequencies at individual sites. There is independence among sites, and all evolutionary forces are weak, so that diffusion approximations can be employed.

These assumptions allow the use of Wright's stationary distribution formula (Wright 1931, 1937) to describe the probability density of the frequency  $q$  of  $A_2$  at a given site

$$
\phi(q) = C \exp(\gamma q) p^{\alpha - 1} q^{\beta - 1} \tag{1}
$$

where  $p = 1 - q$ ,  $\alpha = 4N_e v$ ,  $\beta = 4N_e u$ ,  $\gamma = 2N_e s$ , and the constant *C* is such that the integral of  $\phi(q)$  between  $q = 0$  and  $q = 1$  is equal to 1.

When *k* haploid genomes are sampled from a population, the probability of obtaining a frequency x of  $A_2$  at a given site in the sample is given by the integral of the product of φ(*q*) and the binomial probability of obtaining *x*, conditioned on *q* (e.g. Sawyer and Hartl 1992). This can be used to calculate quantities such as the expected value of Tajima's *D* for a sequence of length *m* (Tajima 1989) and the expected proportion of singletons in the sample, by numerical evaluation of the relevant formulae. Analyses of selection on codon usage bias in *D. melanogaster* suggest that γ is typically around 1 and κ is between 2 and 3 (Zeng and Charlesworth 2009), so that the results presented below for these values are probably most relevant for the present study.

$\gamma$	$\pmb{\mathit{K}}$	$\pi$ (%)	$\theta_w$ (%)	Tajima's	Proportion of
				D(%)	Singletons $(\% )$
$\boldsymbol{0}$	$\mathbf{1}$	0.980	0.977	1.14	31.1
$\mathbf{1}$	$\mathbf{1}$	0.908	0.920	$-3.78$	32.3
$\overline{2}$	$\mathbf{1}$	0.751	0.796	$-16.4$	35.4
$\overline{4}$	$\mathbf{1}$	0.479	0.576	$-47.9$	43.7
$\boldsymbol{0}$	$\overline{2}$	1.29	1.29	0.005	30.9
$\mathbf{1}$	$\overline{2}$	1.42	1.43	$-2.88$	32.0
$\overline{2}$	$\overline{2}$	1.33	1.40	$-4.82$	35.1
$\overline{4}$	$\overline{2}$	0.936	1.12	$-48.4$	43.3
$\boldsymbol{0}$	$\overline{3}$	1.44	1.32	0.007	30.8
$\mathbf{1}$	3	1.73	1.74	$-0.007$	31.8
$\overline{2}$	3	1.78	1.87	$-0.593$	34.7
$\overline{4}$	3	1.37	1.63	$-47.8$	42.9

**Table S1-4.** Population statistics for weakly selected sites as a function of the strength of selection γ and the mutational bias κ

A sample size of 17 haploid genomes was assumed. The scaled mutation parameter β was equal to 0.02; the sequence length for calculating Tajima's *D* was 450bp.

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