

# SUPPLEMENTARY INFORMATION for

## Chromosomal replication dynamics and interaction with the $\beta$ sliding clamp determine orientation of bacterial mobile elements

by Manuel J. Gómez, Héctor Díaz-Maldonado, Enrique González-Tortuero

and Francisco J. López de Saro

### - Supplementary Methods

- **Table S1.** List of Pfam domains used to identify bacterial transposases analyzed in this study.
- **Table S2.** Pfam-based architecture description of IS structures and classification into IS families.
- **Table S3 (see attached PDF file).** List of chromosomes analyzed, IS orientation data, and statistical significance for the non-random orientation of IS elements.
- **Table S4 (see attached PDF file).** List of chromosomes with 6 or more copies of a given IS family.
- **Table S5.** Alignments of regions containing  $\beta$  motifs in transposases.

### SUPPLEMENTARY METHODS

**Purification of *E. coli*  $\beta$ .** The DnaN gene of *E. coli* MG1655 was amplified by PCR from genomic DNA with oligonucleotides #1 (5'-CGGCTAGCCATGGAATTACCGTAGAACGTGAGC-3') and #2 (5'-GGCCTAGGA TCCTTACAGTCTCATTGGCATGACAACATAAGCCGC-3'), cloned (Ncol-BamHI) in vector pET16b (Novagen Inc.), and sequenced. This plasmid was used to overexpress  $\beta$  in *E. coli* BL21(DE3). 15 g of cells (dry weight) were resuspended in Buffer A (100 mM TrisCl, 100 mM NaCl, 2 mM EDTA, 5% glycerine, 1 mM  $\beta$ -mercaptoethanol, 1 mM PMSF, pH 8) and processed four times with a French press. All purification procedures took place at 4°C. The lysed cells were diluted to 150 mL and centrifuged (17,200 g, 30 m, 4°C). The supernatant was treated with 0,2% polyethyleneimine and the precipitate removed by centrifugation. The supernatant was then brought up to 50% saturation with  $[\text{NH}_4]_2\text{SO}_4$ . After centrifugation, the supernatant was further treated with  $[\text{NH}_4]_2\text{SO}_4$  to 70% saturation and centrifuged. The precipitate, containing  $\beta$ , was dissolved in 27 mL of Buffer B (100 mM TrisCl, 100 mM NaCl, 2 mM EDTA, 5% glycerine, 1 mM  $\beta$ -mercaptoethanol, pH 7.2) and dialyzed against 2 L of this buffer for 12 h. This fraction (27 mL) was then applied on a 30 mL Q sepharose FF (GE Healthcare) ion-exchange chromatography column equilibrated in Buffer B. Protein was eluted with Buffer B and a NaCl gradient (320 mL, 0.1-1.0 M) and fractions (3 mL) collected. Fractions containing  $\beta$  (30 mL) were pooled and dialyzed against 1 L of Buffer C (50 mM TrisCl pH 7.2) + 1 M  $[\text{NH}_4]_2\text{SO}_4$ . This fraction (250 mg) was then applied on a 20 mL Phenyl Sepharose 6 FF (GE

Healthcare) chromatography column equilibrated in Buffer C. Hydrophobic interaction chromatography was performed in Buffer C with a double gradient (100 mL) of  $[\text{NH}_4]_2\text{SO}_4$  (1.0-0 M) and ethylene glycol (0-60%). Fractions containing  $\beta$  were pooled and dialyzed against 2 L of Buffer D (50 mM TrisCl, 50 mM NaCl, 1 mM EDTA, 10% glycerine, 1 mM DTT, pH 8). This fraction ( $2.7 \text{ mg mL}^{-1}$ ) was aliquoted and stored at -80°C.

**Purification of GST-Pol IV<sup>LF</sup>.** The C-terminal domain of *E. coli* DNA polymerase IV (residues Val243-Leu351) was amplified by PCR using genomic DNA from strain *E. coli* MG1655 and oligonucleotides #3 (5'-GAATGACGGATCCGTGGCGTGGAACGCACGATGGCGGAAG-3') and #4 (5'-GAATGACCTCGAGTCATAATCCCAGCACCAGTTGTCTTCCATTGCGGG-3'), and cloned (BamHI-Xhol) in pGEX-4T-3 (GE Healthcare). The resulting soluble GST fusion protein (335 amino acids, 39 kDa) was purified by affinity chromatography with Glutathione Sepharose 4B following the standard procedures recommended by the manufacturer (Amersham).

**Purification of *Methanosarcina barkeri* PCNA.** Genomic DNA from *Methanosarcina barkeri* Fusaro (DSM 804) was obtained from DSMZ (Braunschweig, Germany) and used to amplify the gene encoding PCNA by PCR using the oligonucleotides #5 (5'-GGATGACCATGGTTAAGGCAGCAATTAAATGCAGAGCTTC-3') and #6 (5'-GGATGAGGATCCTCAGTCCGACTCAATTCTGGAGCCAGG-3'). The PCNA gene was cloned (Ncol-BamHI) into vector pET16b and the protein overexpressed in *E. coli* BL21(DE3). Purification of PCNA followed the same protocol as for *E. coli*  $\beta$  with the following differences: a)  $[\text{NH}_4]_2\text{SO}_4$  was raised to 40% saturation to remove contaminants and then to 60% to precipitate PCNA; b) the NaCl gradient on the Q sepharose FF chromatography column was from 0.2 to 1.0 M.

**TABLE S1.** List of Pfam domains used to identify the bacterial transposases analyzed in this study.

DDE_2	DUF4158	IstB_IS21_ATP
DDE_3	DUF4277	LZ_Tnp_IS481
DDE_4	DUF4338	LZ_Tnp_IS66
DDE_4_2	DUF4351	MULE
DDE_5	DUF4372	Mu-transpos_C
DDE_Tnp_1	DUF772	Nterm_IS4
DDE_Tnp_1_2	HTH_17	OrfB_IS605
DDE_Tnp_1_3	HTH_21	OrfB_Zn_ribbon
DDE_Tnp_1_4	HTH_23	Phage-MuB_C
DDE_Tnp_1_5	HTH_24	Resolvase
DDE_Tnp_1_6	HTH_28	rve
DDE_Tnp_1_assoc	HTH_29	rve_2
DDE_Tnp_2	HTH_32	rve_3
DDE_Tnp_IS1	HTH_33	Tn7_Tnp_TnsA_C
DDE_Tnp_IS1595	HTH_38	Tn7_Tnp_TnsA_N
DDE_Tnp_IS240	HTH_7	Tn7_TnsC_Int
DDE_Tnp_IS66	HTH_OrfB_IS605	TnPB_IS66
DDE_Tnp_IS66_C	HTH_Tnp_1	Transposase_20
DDE_Tnp_ISAZ013	HTH_Tnp_IS1	Transposase_31
DDE_Tnp_ISL3	HTH_Tnp_IS630	Transposase_mut
DDE_Tnp_Tn3	HTH_Tnp_IS66	Y1_Tnp
DEDD_Tnp_IS110	HTH_Tnp_ISL3	Y2_Tnp
Dimer_Tnp_Tn5	HTH_Tnp_Mu_1	Zn_Tnp_IS1
DUF2080	HTH_Tnp_Mu_2	Zn_Tnp_IS1595
DUF4096	IstB_IS21	Zn_Tnp_IS91

**TABLE S2. Pfam-based architecture description of IS structures and classification into IS families.** The table lists 209 gene clusters corresponding to IS elements. Gene symbols are enclosed by characters that represent an arrow (--gene->), to describe their relative orientation, and are connected by double colons (::). The gene symbols represent the domain organization of encoded proteins. Domain names follow Pfam nomenclature and are connected by equal signs (=). IS family nomenclature follows that of IS finder ([www-is.bioutl.fr//is.html](http://www-is.bioutl.fr//is.html)), except for IS4, IS5, IS66, and ISNCY, as described in accompanying Notes. Importantly, our classification aims to group the domain architectures and proteins of each IS family according to the transposase domains detected and does not take into account other aspects of IS structure, such as the flanking inverted repeats present in many ISs.

IS FAMILY	IS STRUCTURE	NOTES
IS1	--DDE_Tnp_IS1-> --Zn_Tnp_IS1=HTH_Tnp_IS1->::--DDE_Tnp_IS1-> --Zn_Tnp_IS1=HTH_Tnp_IS1-> --Zn_Tnp_IS1=DDE_Tnp_IS1-> --HTH_23=DDE_Tnp_IS1-> --Zn_Tnp_IS1=HTH_Tnp_IS1=DDE_Tnp_IS1-> --HTH_Tnp_IS1->::--DDE_Tnp_IS1-> --Zn_Tnp_IS1->::--DDE_Tnp_IS1->	
IS3	--HTH_Tnp_1->::--HTH_21=rve-> --HTH_21=rve-> --HTH_Tnp_1->::--HTH_21=rve_3-> --HTH_29=rve-> --rve_3-> --HTH_28=HTH_28->::--HTH_21=rve-> --rve_2-> --HTH_Tnp_1=HTH_21=rve-> --HTH_28>::--HTH_21=rve-> --HTH_Tnp_1->::--rve_3-> --HTH_21=rve_3-> --HTH_Tnp_1->::--HTH_21-> --HTH_28=HTH_21=rve-> --HTH_28=rve-> --HTH_28->::--HTH_21=rve_2-> --HTH_21=rve_2-> --HTH_32=rve-> --HTH_28=rve_3-> --HTH_Tnp_1=HTH_28->::--HTH_21=rve-> --HTH_Tnp_1=HTH_21=rve_3-> --HTH_Tnp_1->::--HTH_29=rve-> --HTH_29=rve_3-> --HTH_28=HTH_Tnp_1=HTH_28->::--HTH_21=rve-> --HTH_21->::--rve_2-> --HTH_29=rve=rve_2-> --HTH_Tnp_1->::--HTH_21=rve_2-> --HTH_Tnp_1=rve-> --HTH_32=rve_3-> --HTH_Tnp_1->::--HTH_21->::--rve_3-> --HTH_28=HTH_21=rve_2-> --HTH_38=HTH_Tnp_1->::--HTH_21=rve_2-> --HTH_28->::--rve_2-> --HTH_28=HTH_28=HTH_28->::--HTH_21=rve-> --HTH_21->::--rve_3->	

	--HTH_Tnp_1->::--HTH_32=rve-> <HTH_Tnp_1->::rve_3-> --HTH_28=HTH_28=HTH_21=rve-> --HTH_28->::--HTH_21->::rve_2-> --HTH_28->::rve_3-> --HTH_Tnp_1->::--HTH_38=rve-> --HTH_Tnp_1->::<HTH_38=rve-- --HTH_29=rve=HTH_29-> --HTH_Tnp_1=rve_3-> --HTH_23=HTH_38=rve-> --HTH_28=HTH_28->::--HTH_21->::rve_2-> --HTH_Tnp_1->::rve_2-> --HTH_23=HTH_21=rve->	
<b>IS4a</b>	--Nterm_IS4=DDE_Tnp_1-> --Nterm_IS4-> --Nterm_IS4->::--DDE_Tnp_1->	Includes members of subgroup IS4 of family IS4
<b>IS4b</b>	--DUF4372=DDE_Tnp_1-> --DUF4372-> --DUF4372->::--DDE_Tnp_1->	Includes members of subgroup IS4Sa of family IS4
<b>IS4c</b>	--Dimer_Tnp_Tn5-> --DDE_Tnp_1=Dimer_Tnp_Tn5->	Includes members of the IS50 subgroup of family IS4
<b>IS5a</b>	--DUF772=DDE_Tnp_1-> --DUF772-> --DUF772=DDE_Tnp_1_2-> --DUF772->::--DDE_Tnp_1-> --DUF772->::--DDE_Tnp_1_2->	Includes members of the IS5 subgroup of family IS5
<b>IS5b</b>	--DUF4096->::--DDE_Tnp_1_2-> --DUF4096-> --DUF4096=DDE_Tnp_1-> --DUF4096=DDE_Tnp_1_2-> --DUF4096->::--DDE_Tnp_1->	Includes members of subgroups IS427 and IS1031 of family IS5
<b>IS5c</b>	--DDE_Tnp_1_5->	Includes members of subgroup ISL2 of family IS5
<b>IS5d</b>	--DDE_4-> --DDE_4_2-> --DDE_4_2->::--DDE_4-> --DDE_4_2=DDE_4->	Includes members of subgroup IS903 of family IS5
<b>IS6</b>	--DDE_Tnp_IS240-> --Zn_Tnp_IS1=DDE_Tnp_IS240->	
<b>IS21</b>	--IstB_IS21-> --HTH_38=rve->::--IstB_IS21-> --IstB_IS21_ATP=IstB_IS21-> --HTH_23=rve->::--IstB_IS21-> --HTH_7=rve->::--IstB_IS21-> --HTH_23->::--IstB_IS21-> --HTH_23=rve->::--IstB_IS21_ATP=IstB_IS21-> --HTH_29=rve->::--IstB_IS21-> --IstB_IS21->::--HTH_21=rve->	
<b>IS30</b>	--HTH_38=rve-> --HTH_38->	
<b>IS66a</b>	--HTH_Tnp_1->::--TnpB_IS66->::--LZ_Tnp_IS66=HTH_Tnp_IS66=DDE_Tnp_IS66=DDE_Tnp_IS66_C-> --TnpB_IS66->::--LZ_Tnp_IS66=HTH_Tnp_IS66=DDE_Tnp_IS66=DDE_Tnp_IS66_C->	The IS66 family was divided in two groups according to Gourbeyre <i>et al.</i> (2010) Route 66: investigations into the organisation and

	--TnpB_IS66->::--LZ_Tnp_IS66=HTH_Tnp_IS66=DDE_Tnp_IS66-> --LZ_Tnp_IS66=HTH_Tnp_IS66=DDE_Tnp_IS66=DDE_Tnp_IS66_C-> --LZ_Tnp_IS66=HTH_Tnp_IS66=DDE_Tnp_IS66-> --TnpB_IS66->::--LZ_Tnp_IS66=DDE_Tnp_IS66=DDE_Tnp_IS66_C-> --HTH_Tnp_1->::--TnpB_IS66->::--LZ_Tnp_IS66=HTH_Tnp_IS66=DDE_Tnp_IS66-> --HTH_Tnp_1->::--LZ_Tnp_IS66=HTH_Tnp_IS66=DDE_Tnp_IS66=DDE_Tnp_IS66_C-> --TnpB_IS66->::--LZ_Tnp_IS66->::--HTH_Tnp_IS66=DDE_Tnp_IS66=DDE_Tnp_IS66_C-> --TnpB_IS66->::--LZ_Tnp_IS66=DDE_Tnp_IS66-> --TnpB_IS66->::--HTH_Tnp_IS66=DDE_Tnp_IS66-> --TnpB_IS66->::--LZ_Tnp_IS66=DDE_Tnp_IS66-> --HTH_Tnp_1->::--TnpB_IS66->::--LZ_Tnp_IS66-> --HTH_Tnp_IS66=DDE_Tnp_IS66=DDE_Tnp_IS66_C-> --TnpB_IS66->::--HTH_Tnp_IS66=DDE_Tnp_IS66=DDE_Tnp_IS66_C-> --HTH_23=HTH_Tnp_1->::--TnpB_IS66->::--LZ_Tnp_IS66=HTH_Tnp_IS66=DDE_Tnp_IS66=DDE_Tnp_IS66_C-> --TnpB_IS66->::--LZ_Tnp_IS66=HTH_Tnp_IS66->	distribution of the IS66 family of prokaryotic insertion sequences. Research in Microbiology <b>161</b> : 136-143.  IS66a includes the 'classical' IS66 architecture.
<b>IS66b</b>	--HTH_Tnp_IS66=DDE_Tnp_IS66->	Includes the subgroup ISBst12 of IS66 (Gourbeyre <i>et al.</i> , 2010). See IS66a.
<b>IS91</b>	--Zn_Tnp_IS91=Y2_Tnp-> --Y2_Tnp-> --Zn_Tnp_IS91->::--Y2_Tnp->	
<b>IS110</b>	--DEDD_Tnp_IS110=Transposase_20-> --Transposase_20-> --DEDD_Tnp_IS110-> --DEDD_Tnp_IS110->::--Transposase_20->	
<b>IS200</b>	--Y1_Tnp-> --Y1_Tnp=Y1_Tnp->	
<b>IS200/IS605</b>	<-HTH_OrfB_IS605=OrfB_IS605=OrfB_Zn_ribbon->::--Y1_Tnp-> --Y1_Tnp->::--HTH_OrfB_IS605=OrfB_IS605=OrfB_Zn_ribbon-> --Y1_Tnp->::--OrfB_IS605=OrfB_Zn_ribbon-> <-Y1_Tnp->::--OrfB_IS605=OrfB_Zn_ribbon-> <-OrfB_IS605->::--Y1_Tnp-> <-HTH_OrfB_IS605=OrfB_IS605->::--Y1_Tnp-> <-Y1_Tnp->::--HTH_OrfB_IS605->::--OrfB_IS605=OrfB_Zn_ribbon->	We have chosen TnpA (domain Y1_Tnp) as a proxy to score the orientation of the insertion sequences in this family.
<b>IS256</b>	--Transposase_mut->	
<b>IS481</b>	--LZ_Tnp_IS481=rve-> --LZ_Tnp_IS481->	
<b>IS607</b>	--Resolvase->::--HTH_OrfB_IS605=OrfB_IS605=OrfB_Zn_ribbon-> --HTH_17=Resolvase->::--OrfB_IS605=OrfB_Zn_ribbon-> --MerR=Resolvase-> --Resolvase->::--HTH_OrfB_IS605=OrfB_IS605-> --HTH_17=Resolvase-> --MerR=Resolvase->::--HTH_OrfB_IS605=OrfB_IS605=OrfB_Zn_ribbon-> --MerR_1=Resolvase->::--OrfB_IS605=OrfB_Zn_ribbon-> --HTH_17=Resolvase->::--OrfB_Zn_ribbon-> --MerR=Resolvase->::--OrfB_Zn_ribbon-> --MerR_1=Resolvase->::--OrfB_IS605-> --Resolvase->::--OrfB_IS605=OrfB_Zn_ribbon-> --MerR_1=Resolvase->::--HTH_OrfB_IS605=OrfB_IS605=OrfB_Zn_ribbon-> --MerR=Resolvase->::--OrfB_IS605=OrfB_Zn_ribbon-> --Resolvase->::--OrfB_IS605-> --HTH_17=Resolvase->::--OrfB_IS605->	

<b>IS630</b>	--HTH_29=DDE_3-> --DDE_3-> --HTH_Tnp_IS630=DDE_3-> --HTH_Tnp_IS630->::--DDE_3-> --HTH_29->::--DDE_3-> --HTH_23=HTH_33=DDE_3-> --HTH_Tnp_IS630-> --HTH_23=HTH_33->::--DDE_3-> --HTH_33=DDE_3-> --HTH_28->::--DDE_3-> --HTH_28=HTH_33->::--DDE_3-> --HTH_33->::--DDE_3-> --HTH_23->::--DDE_3-> --HTH_28=DDE_3-> --HTH_23=DDE_3-> --HTH_23->::--HTH_33->::--DDE_3->	
<b>IS701</b>	--DDE_5-> --DDE_5=DDE_Tnp_1-> --DDE_Tnp_1->::--DDE_5-> --DDE_5->::--DDE_Tnp_1-> --DDE_Tnp_1->::<-DDE_5-> <-DDE_Tnp_1->::--DDE_5->	
<b>IS982</b>	--DDE_Tnp_1_3-> --DDE_Tnp_1_3=DDE_Tnp_1_3->	
<b>IS1182</b>	--DUF772=DDE_Tnp_1_6-> --DDE_Tnp_1_6-> --DUF772->::--DDE_Tnp_1_6->	
<b>IS1380</b>	--DDE_Tnp_1_4->	
<b>IS1595</b>	--DDE_Tnp_IS1595-> --Zn_Tnp_IS1595=DDE_Tnp_IS1595-> --Zn_Tnp_IS1595->::--DDE_Tnp_IS1595->	
<b>IS1634</b>	--DUF4277-> --DUF4277=DDE_Tnp_1->	
<b>ISAs1</b>	--DDE_Tnp_1_assoc=DDE_Tnp_1-> --DDE_Tnp_1_assoc-> --DDE_Tnp_1_assoc->::--DDE_Tnp_1-> --DUF4338=DDE_Tnp_1_assoc=DDE_Tnp_1-> --DUF4338->::--DDE_Tnp_1_assoc->	
<b>ISAzo13</b>	--DDE_Tnp_ISAZ013->	
<b>ISL3</b>	--HTH_Tnp_ISL3=DDE_Tnp_ISL3-> --DDE_Tnp_ISL3-> --HTH_Tnp_ISL3->::--DDE_Tnp_ISL3-> --HTH_Tnp_ISL3=DDE_Tnp_ISL3=DDE_Tnp_ISL3-> --DDE_Tnp_ISL3=DDE_Tnp_ISL3-> --HTH_Tnp_ISL3->::--DDE_Tnp_ISL3=DDE_Tnp_ISL3->	
<b>ISNCYa</b>	--Transposase_31-> --DUF4351-> --Transposase_31=DUF4351->	ISPlu15
<b>ISNCYb</b>	--MULE->	ISM1
<b>ISNCYc</b>	--DDE_Tnp_2->	ISC1217
<b>ISNCYd</b>	--DUF2080->	ISA1214

Tn3	--DUF4158=DDE_Tnp_Tn3-> --DDE_Tnp_Tn3-> --Resolvase=HTH_7->:::--DUF4158=DDE_Tnp_Tn3-> <-Resolvase=HTH_7--:::--DUF4158=DDE_Tnp_Tn3-> --Resolvase->:::--DUF4158=DDE_Tnp_Tn3-> <-DUF4158=DDE_Tnp_Tn3--:::--Resolvase-> --DUF4158->:::--DDE_Tnp_Tn3-> --Resolvase=HTH_7->:::--DUF4158->	
Tn7	--Tn7_Tnp_TnsA_N=Tn7_Tnp_TnsA_C-> --Tn7_Tnp_TnsA_N->:::rve=Mu-transpos_C-> --Tn7_Tnp_TnsA_N=Tn7_Tnp_TnsA_C->:::rve=Mu-transpos_C-> --Tn7_Tnp_TnsA_N=Tn7_Tnp_TnsA_C->:::HTH_28=rve=Mu-transpos_C-> --Tn7_Tnp_TnsA_N=Tn7_Tnp_TnsA_C=rve=Mu-transpos_C-> --Tn7_Tnp_TnsA_N=rve=Mu-transpos_C-> --Tn7_Tnp_TnsA_N=Tn7_Tnp_TnsA_C=rve->	

**TABLE S3 (In annexed PDF file). Statistical significance for the non-random distribution of IS elements at the chromosomal level.** Each chromosome is listed with the number of ISs found for each IS family. The number of ISs found in and each of the two possible orientations relative to the local GC skew sign (same/anti) are indicated (See Materials and Methods for details). *p*-values were calculated for each chromosome and IS family, and coloured according to its significance (red for  $p < 10^{-3}$ , orange for  $10^{-3} < p < 10^{-2}$  and yellow for  $10^{-2} < p < 0.05$ ).

Chromosomes used for calculation of statistical significance at the Phylum level (Table 1) are marked red in the first column.

**TABLE S4 (in annexed PDF file). Statistical significance for the non-random distribution of IS elements at the chromosomal level.** As in Table S3, but including only chromosomes with 6 or more copies of a give IS family, and only one chromosome for each species. The table has been ordered by IS family and *p*-value. Significant *p*-values are labelled (red for  $p < 10^{-3}$ , orange for  $10^{-3} < p < 10^{-2}$  and yellow for  $10^{-2} < p < 0.05$ ).

**TABLE S5. Sequences of transposase regions containing the  $\beta$  motif.** *Escherichia coli* peptides used for biochemical analysis (Fig. 2a) are boxed, and residues putatively corresponding to the  $\beta$  binding motif are in bold. Phylum code: Actinobacteria (A), Bacteroidetes/Chlorobi (B), Chlamydiae/Verrucomicrobia (V), Cyanobacteria (C), Deinococcus-Thermus (D), Firmicutes (F), Fusobacteria (U), Nitrospirae (N), Planctomycetes (M), Proteobacteria (P), Spirochaetes (S), Tenericutes (T), Thermotogae (H) and Archaea (Arch.).

### IS5a

P <i>Escherichia coli</i> (AAB53644)	305-RES <b>QIQGVAENDNQLAMLF</b> T LANLFRADQMIRQWERSH*
P <i>Yersinia enterocolitica</i> (CAE46781)	231-TKVRFRGLVRNTA <b>QLVTLF</b> ALS NLWMARRHLLASAGEVRL*
P <i>Vibrio cholerae</i> (EKG68054)	289-IKARYKGLMKNDN <b>QLAMLF</b> T LANLVKVDQLIRRQARSA*
P <i>Pseudomonas putida</i> (CAB99195)	286-VKTRFRGLAKNTA <b>QLVTLF</b> ALS NLWMARRHLLTNAGEVRL*
P <i>Legionella pneumophila</i> (CBX00305)	308-IVICVMVVNQALV <b>QSMDLTAYRY</b> *
P <i>Neisseria meningitidis</i> (CBA09446)	333-ARAAYFGLSKVSA <b>QSHLKAMCLNLLKAANRLSAPAAA</b> *
P <i>Ralstonia solanacearum</i> (AAR23794)	289-RKVRYKGLAKNTA <b>QLFSLF</b> GLANLV LARRQLLASPGSIAS*
F <i>Bacillus</i> sp. (ZP_01722218)	415-RWTTLRGLKKLSM <b>QAMLTFAAMNLKKLATWTWQVA</b> *
F <i>Streptococcus thermophilus</i> (CAE52417)	311-TLTNLLYNIFRFE <b>QIKRLGLKSWA</b> *
F <i>Lactobacillus acidophilus</i> (YP_004292069)	549-LCFYIRAKNRVSS <b>QTTLF</b> KRKIKLKSTSKLNP*
F <i>Thermoanaerobacter ethanolicus</i> (ZP_08211387)	478-KQGLKFYFVNKL <b>VQIKLFAFLYKYYR</b> *
C <i>Crocospshaera watsonii</i> (EAM49691)	446-ITFLVVNL SKLLR <b>QLLSLFLSFTNNRTGNSSNRLSLILIIL</b> *

### IS30

P <i>Escherichia coli</i> (NP_415922)	329-TNGLIRQ <b>YFPKKTCLAQYTQHELDLVA</b> AQLNNRPRKTLKFKT-370
P <i>Yersinia kristensenii</i> (ZP_04623201)	329-TNSLIRQYLPKKTCLAQHS <b>QEVLNQIADELNDRPRKTLKFKT</b> -370
P <i>Pseudoalteromonas citrea</i> (ZP_10273944)	331-TNRLLRQYFPKKTS LHGFD <b>QSYLDKIANKLN</b> NRPRRILNYLT-372
P <i>Burkholderia xenovorans</i> (YP_555985)	331-TNRLLRQYLPHGTQLDHYS <b>QADLNKIAARLN</b> ERPRKTLGFRS-372
P <i>Pseudomonas fluorescens</i> (YP_348496)	332-TNGLLRQYFLKGTDLAEHS <b>QATLNEVARQLNSRPRKTL</b> DYET-373
P <i>Ralstonia eutropha</i> (YP_728612)	316-TNGLLRQYLPKGTDLSVY <b>SQAKLN</b> AIARRLNERPRKTLNFDT-357
P <i>Rickettsia massiliae</i> (YP_001499437)	274-MNSMIHRILPKNTDITTVT <b>QRGLDNVAE</b> I LNMMPRKIFGYKT-315
F <i>Streptococcus mitis</i> (YP_003445190)	312-NHTVLRAILPKGTSFDQLT <b>QKDVLNVISHVN</b> SLKREEFQGKS-353
F <i>Streptococcus salivarius</i> (CAA78948)	289-FNGLLREFIPKGCSLKELN <b>QNLL</b> EDYTKAINERPRRIHGYQS-330
F <i>Streptococcus pneumonia</i> (CCG13893)	313-NHTLIRDILPKGTSFDNL <b>TQEDINLV</b> CSHVNSVKRAALNGKS-354
F <i>Staphylococcus aureus</i> (YP_039551)	261-TNGLLREFFPKKTDLAKVN <b>QEQLNYALDS</b> INYRPRKCLNWKF-302
S <i>Leptospira biflexa</i> (YP_001840433)	286-FFPKGTDFS KLKS <b>QIKKV</b> QTLLNQRPRKTLNWNTPEEEIRA-318
S <i>Treponema succinifaciens</i> (YP_004365962)	319-NGLPRKRLGYKTPEELFNE <b>QLDLIYRL</b> *
A <i>Frankia</i> sp. (YP_001508751)	434-NDRPRKTLDWKTPTEAMNN <b>QLLSLQQPGVARTG</b> *

## IS66a (TnpB)

P <i>Escherichia coli</i> (YP_424826)	66-LFTKRLERGRFWWPVTRDG-KVHLTPAQLSMLLEGINWKHPKRTERAGIRI*
P <i>E. coli</i> (YP_003235003)	64-LFTKRLEEGQFIWPAVRDG-KVSITRSQLAMLLDKLDWRQPKTSSRNSLTM*
P <i>Shigella sonnei</i> (YP_313273)	66-LFTRRLERGRFWWPVTRDG-KVHLTPAQLSMLLEGIDWKHPKRTERAGIRI*
P <i>S. boydii</i> (YP_407150)	66-LFTRRLERGRFWWPVTRDG-KVSI TRSQLAMLLDKLDWRQPKTSRLNALTM*
P <i>Polymorphum gilvum</i> (YP_004305836)	66-LVHKRLECGKFVWPQAQDG-VMRISSAQMAALFEGLDWRLVRPERARRPLVAG*
P <i>Ochrobactrum intermedium</i> (ZP_04679718)	72-LFTKKLERGRFIWPSAADG-TVVIITPAQLGYLLEGIDWRMPQKTWRPTSAG*
P <i>Methylocystis</i> sp. (ZP_08074780)	66-LFAKRLEDGEFRWPKIEDG-TMRLSATQFSALLEGLDWKRVHTKETPAPALPG*
P <i>Agrobacterium vitis</i> (YP_002540116)	72-LFYKVLERGYFPWPRAKEG-VAPLTQAQLSMLVEGIDWRRPAWTSAPARTG*
P <i>Rhizobium leguminosarum</i> (YP_002973065)	66-LAYKRLEEHTFTWPGIKDG-LMTLTHAQFEALFAFDLVVSGHA*
P <i>Pelagibaca bermudensis</i> (ZP_01442077)	66-MAYKRLEESTFTWPAIRDG-AMTLNRAQFEALFAGLDWRRVRSLEVRRPAVAE*
P <i>Desulfovibrio magneticus</i> (YP_002952289)	66-LWHKRLERHVFRWPTR-EAEVLAIDSRLQAWLLDGLDPLAVTGHSRLEYSTLF*
P <i>Magnetococcus marinus</i> (YP_864253)	67-LWQKRLEKDRFHWRQGGAAEIQITGRQLNWLLDGYNLAAMKGHNKLHFSSIV*
P <i>A. ferrooxidans</i> (YP_002218731)	65-LVYRRLDQGRLHWPRADAG-ALELSAAQWAMLVEGRPWTPLPTLEKCTPKLL*
P <i>Burkholderia glumae</i> (YP_002907560)	66-LMLKRLEADHFVWPCHR-EQAVIELTTEQLHWLLDGIDIDAMQRHPARRYRHAS*
P <i>Cupriavidus taiwanensis</i> (YP_001795972)	66-LFLKRLEADRFAWPR--GAAVATLSVEQLHWLLDGIDISAVQRHPPRHYQRAV*
F <i>Streptococcus pneumoniae</i> (YP_002742797)	67-LLYKRFENGRLTWPS-EKDVKALTPEQVDWLMKGFSITPKINPSES RDFY*
F <i>Bacillus pseudofirmus</i> (YP_003426249)	66-LYYRRLEKGTFWPWPDSSSSPQMISHRQFRWLLDGLSIDQKSAHPKVT AQRVI*
F <i>B. selenitireducens</i> (YP_003698933)	66-LYYRRLEKGRFPWPTSGSDEPMIITERQLRWLLDGLPLDQKGKAHRKMNP EKVV*

## IS66a (TnpC)

P *Escherichia coli* (YP\_003235004)  
 P *Shigella dysenteriae* (EFW49566)  
 P *Yersinia pestis* (YP\_003566509)  
 P *Klebsiella pneumoniae* (YP\_001687996)  
 P *Pantoea vagans* (YP\_003933479)

P *Escherichia coli* (ZP\_07592975)  
 P *Shewanella benthica* (ZP\_02157382)  
 P *Fulvimarina pelagi* (ZP\_01440753)  
 P *Magnetococcus marinus* (YP\_866522)  
 P *Aliivibrio salmonicida* (YP\_002262549)  
 P *Brucella ceti* (ZP\_03787083)  
 P *B. cellullosilyticus* (YP\_004096872)  
 P *Methylocystis* sp. (ZP\_08074907)  
 P *Polymorphum gilvum* (YP\_004305837)  
 M *Planctomyces maris* (ZP\_01851762)  
 B *Parabacteroides merdae* (ZP\_02032857)  
 B *Bacteroides salanitronis* (YP\_004258102)  
 V *Chthoniobacter flavus* (ZP\_03131569)  
 F *Lactobacillus casei* (YP\_005858453)

F *Streptococcus pneumoniae* (NP\_358890)  
 F *Lactobacillus parafarraginis* (ZP\_09394150)  
 F *Enterococcus faecium* (EJX82788)

MSQKYLIRIAELERLI[SEQAEALRQKDQQQLSLVVEETEAFLRSALTRAEEKIEEDEREIEHLRAQIEKLRRLMFGTRSEKLRREVEAALL----KREQQDSDRY  
 MNQKYLIRIAELECQL-----RQKDQQQLSLVVEETEAFLRSALARAEKKIEEDEREIEHLRAQIEKLRRLMFGTRSEKLRREVEAALL----KREQQDSDRY  
 MVMSQDYLARIAALEDAL-----RQKDNLQSLSLVAETESFLRSALARAEKKIEENEREIEHLRAQIEKLRRLMFGTRSEKLRRVVEAEALL----KQQEQQSDRY  
 MNHDYLARIAALEDAL-----RQKDSLQSLSLVAETESFLRSALARAEKKIEENEREIEHLRAQIEKLRRLMFGTRSEKLRRVVERAEALL----KQQEQQSDRY  
 MKRSLSAENDRLRALL-----DTQQRSLQLEQMAEYNRLLS----RRVAAYASEINRLKALVALQRMQFGKSSEKLREKTARQVREAE--RISGLQEEMA  
 MDISLLSTTR-----DPEQLRALAIAMVQKAMAESQNLANVQEKDRNIAELQNRIRILEQMQLARQQ[RGKKCESLAG---MQRSLFEE-DVDADIAEISAH  
 MASQYSEIAELKQS-----VQRLLEPFRLAQQQRFGASSESHN---YQGELFNE---AEVTLDEPE  
 MATALEALPD-----DPGTLKAMLIA--ERVRSE-----RLEQIIKELQRHRFGRRAETLPE---DQLLLGLE-DVEQGVAVEEAE  
 MKIKPQTLPD-----DPAELKALVQSLQEEMLK-----LREQLHILISKRFGRSSKEYDP---NQLGLFDEAELIGATAADVE  
 MIDIKIP-----LPDTIDELKALVLQLENK-----YNRLLLEQFRRLAQHQRFQGSSES---TQFDLFNE---TEEEIIIEND  
 MLNRGQHLP-----DPDILVGMILERDAEIER-----LKVLLKAANAKPFGQRSEQLAHMVERQIRLDLG-DVVHEPEVASAE  
 MRPMBKIKNDKDAQF-----YKERAEKLEKEELEAK-----LKYWEEQFRLSQKRFQGSSEQT---CQLSLFNE---IEDTSNKDV  
 MSRAAADLPE-----DPAELRRFAEALAAEVHAK-----TLLIEKLKMQLAVLRRARFGRSEKLDRDI[QLELLIG-DMEESDAERQA-  
 MDAAVLAREN-----ALLKARLIEVEAALAES-----QEANRRLDEDILRTSQREKFGRSEKLPSP--DQFNLPLE-DAELAQGVLEA-  
 MIHQQLG---ETVGE-----QREVEQLKHFIDRLRQRFGARSEKIA---PNQMSLFDE--PEAAEATDPE  
 MIHTDTMELIIKNQQEQIKGLLETNRTLVESNQKLMEQTGE-----LQQKVQELLSQVAWLNRQLFGRKSEKLASLDPNQQLALFDT---LANPRQEE---  
 MIRQDTMEQIIRSQQEQIAGLLETNRSLVESNGKLLQEQTDA-----LQRKIQELLSQIAWLNRQLFGRSEKLAALDPNQSLFDS--VPATGQDEDIR  
 MPDSTFPN-----AAELLARIAELEKE-----NALLRQKIDALARKIYGVSSKLDP---AQLHLLQ---GLDEPGKAPE  
 MSAEAVVTTEQFEYLKQE-----NALLREQVEFLMRRLYGTKRESLTD--GQVDFDQ--TKTFVAPTV-  
 MKIIQQQSAIIDSLTNELSLCEQVAYLTQKLSGSSEKSVCPCFG[QLNLFEEESPSEKDGDVPS\*  
 MILSLKVKQRTVTQLEIDE[LKENAELRALVAKQAKQIELLQE[QVNLYLMSKLYGKSEQT[PEDGQTS[LFEDDENGVFEPESTGE\*  
 METTDTLLQLLQEAKTNQAQQQT[QNLTTIEQLLNEKVNLYLTKLFGRSKETLFEETNG[QNLFSDEEISVSVPEA-77

## IS91

P *Escherichia coli* (ACO24927)  
 P *Gamma proteobacterium* (ZP\_01617481)  
 P *Gamma proteobacterium* (CBL46704)  
 P *Nitrococcus mobilis* (ZP\_01126307)  
 P *Aliivibrio salmonicida* (CAQ77725)  
 F *Lactobacillus casei* (YP\_806573)  
 F *Clostridium hiranonis* (ZP\_03294328)  
 M *Kuenenia stuttgartiensis* (Q1Q731)

463-IEDPKVIEQILKHLQKTTAKANAAKQRELPP[ERAPPLTPSLFDPSQSRLFD\*  
 453-IEDPSVIKKILEHLDAKSMAL--TSANQLPEPRAPP-[QAEFLD\*  
 464-IEDPAVIEKILQHLMKESLP---LPRVHEARAPPD[QAAFLQL\*  
 435-IEDPEIIEKILAHDHVTEP---EATRRPPCRAPP-[QRGLFDETG\*  
 357-GYVKVDPYECILCESRLVFTNFRVGNSVNDLVTHAIV[QSELRAA\*  
 394-RHYMLEVNQNIAKEAYQT[KYQAEAAAYDRCRFSWER[QRIIYLSEMPQA\*  
 154-IWHYKYGLIYNVLDKSNYKRIIYEEIIKEIISLNTT[QKELF\*  
 249-IYNEIEEIMRGKYEPKEEKVIKPEGDGTTIRPTPRRV[QIPLFSM\*

**IS200**

P <i>Escherichia coli</i> (ZP_03029803)	125-WSWGYFVDTVGV- -NEEIIRR[YARYQEKM <b>EQTHEQQMELLE</b> *
P <i>Vibrio cholera</i> (AAC01554)	106-WARGYFVDTVGV- -NEEIIRR[YVRHQDKKELEPEQQ <b>QLELLRD</b> *
P <i>Shewanella baltica</i> (YP_001364938)	106-WARGYFVDTVGV--NEEIIRR[YVRHQDK <b>QDQEHEAQLSLQMM</b> *
P <i>Shewanella woodyi</i> (YP_001761123)	107-WQRGYFVDTVGI- -NEEVIRR[YVKHQEK <b>VEKQEQPQLDLK</b> *
P <i>Serratia odorifera</i> (ZP_06639291)	138-WARGYCVDTVGI- -NEEMIRKYVKY <b>QEKEV-ESQLPLKEV</b> *
P <i>Nitrosomonas</i> sp. (YP_004295562)	107-WTDGYYVATVGERADGEVERYVKNQGKPKEE-LR <b>QLEFF</b> *
P <i>Escherichia coli</i> (NP_752024)	123-WCRGYYVDTVGK- -NTAKI <b>QDYIKHQLEEDKM-GEQLSIPYP</b> GSPFTGRK*
P <i>Yersinia pestis</i> (YP_005509676)	123-WCRGYYVDTVGK- -NTARI <b>QEYIKHQLEEDKM-GEQLSIPYP</b> PSPFTGRK*
F <i>Streptococcus pneumoniae</i> (ZP_01829269)	107-WCRGYYVDTVGR- -NQKVIAEYI <b>QNQLQEDRV-ADQLTLFESVDPFT</b> -150
F <i>Clostridium botulinum</i> (YP_002650746)	105-WCRGYYVDTVGR- -NKKAI <b>TEYIKNQQKEDMI-SDQISLKEYMDPFK</b> -148
F <i>Bacillus halodurans</i> (BAD18222)	106-WCRGFYVDTVGR- -NKKQI <b>QEYIRNQLREDYM-GDQLTLFEYDPFT</b> -149
F <i>Enterococcus faecium</i> (ZP_05832064)	105-WCRGYYVDTVGK- -NAKKIEEYIANQL <b>QEDLE-YDQMTLKEYIDPFT</b> -148
F <i>Ruminococcus albus</i> (YP_004103952)	108-WCRGYYVDTVGK- -NKKKIAEYIRNQL <b>QEDIV-CDQISLFETVDPFT</b> -151
F <i>Staphylococcus epidermidis</i> (YP_188749)	107-WCKGFYVDTVGR- -NKKVI <b>ENYIRNQLQEDIV-ADQISMEEYLDPFT</b> -150
F <i>Filifactor alocis</i> (YP_005053889)	110-WCKGYYVDTVGK- -NTKAI <b>QEYISNQLKVDRE-SDQISIFDPRDPFTGSK</b> *
U <i>Fusobacterium nucleatum</i> (ZP_00143858)	105-WCRGYYVDTVGR- -NKERIAQYIK <b>NQIEEDIKI-MDQMTLKEYFDPFN</b> -148
P <i>Xenorhabdus nematophila</i> (YP_003713015)	106-WTDGYFASTVGKH <b>DEQMIGRYVQNQGKKYHK-LHSDHQLALF</b> *
P <i>Psychromonas ingrahamii</i> (YP_944951)	107-WTQSVFVETIGNATEE-VIRKYV <b>QNQLIELDRKEINSDQLDF</b> *
C <i>Microcystis aeruginosa</i> (CAO87910)	106-WSDGYYASTVGKH <b>GDEGMIARYVKEQDKEYLQ-LHQNLQLSLF</b> *
C <i>Acaryochloris marina</i> (YP_001519691)	104-WSRGYFVSTVGR- -DEEVIRR[YRH <b>QEQQEQK-LE---QLNLFR</b> A*
B <i>Pelodictyon phaeoclathratiforme</i> (YP_002018173)	106-WTDGYYAGTVGKH <b>GNEDMIGKYVKQGGTYQK-RYSDYQLSLF</b> *
(Arch.) <i>Methanosaeta concilii</i> (YP_004384718)	106-WSDGGYIGTVGDGTTSDVI <b>KSYIEQNQNQEEKEAYKQMKIIDFQ</b> *
(Arch.) <i>Sulfolobus islandicus</i> (YP_002831273)	106-WSSGKFYRSVGNVTAD-TIKHYI <b>KESQKKPKTEVKSSSAKPDQRIIDDF</b> *
(Arch.) <i>Halobacterium salinarum</i> (YP_001688288)	106-WSSGKFFRSVGNVTADTIQ-HYI <b>KESQGKPKAESKVCRSRESGQRRLLDDF</b> *
(Arch.) <i>Haloarcula marismortui</i> (YP_134246)	108-WSPSYFLATSG <b>QVTLEVLIK-KYVESQGKE</b> *
	106-WQPGYFLATTG <b>QVSIDTLM-DYVDDQ</b> *
	108-WNDSYCLISTG <b>QVSLDVLIK-QYVEDQRE</b> *

## IS200/IS605 (OrfB) Cyanobacteria

<i>Cyanothece</i> sp. (YP_002371738)	367-DLNHGLLAGTAPNFMTQKERIGEYIQLSLFDPTLFGG*
<i>Microcystis aeruginosa</i> (YP_001656728)	371-IENRGKNAVGLTVLENACGGDLTGVVQLNLFDLVKSLSRTKN-411
<i>Arthrosira platensis</i> (BAI93705)	374-NGCGERVRLSVKKAHLNEASTRPAFQQLSIFDLLK*
<i>Synechococcus</i> sp. (ZP_05040300)	360-ASVMGVVASADGVFDNPLNSMNQSAQQQLTLFPMSA*
<i>Synechocystis</i> sp. (NP_441190)	368-KNGRGGKRQTTSVAASGEASTHRKAIQLTLFAS*
<i>Trichodesmium erythraeum</i> (YP_721194)	404-ALSKGKTCRKKPANCDEMLTRFESFKQLNLFD*
<i>Microcoleus chthonoplastes</i> (ZP_05026728)	410-ALVGLSINQPGBTGLSCKLSRTIKYVQLSLFDDFRATKNPDLS*
<i>Crocospaera watsonii</i> (ZP_00513803)	435-TPKPELTGSSHRETSVSLEIEPGNPQQQLSLFEWVNGEVIPC*
<i>Acaryochloris marina</i> (YP_001521276)	266-NGCRRECKSEVNSAVLSDASTRLVDKQLELFAS*

## IS1380

P <i>Escherichia coli</i> (YP_003829282)	271-WEKDRRFVVSRVLIKPEKE---RAQLSLLEGS---EYDYFFFVTN-TTLLSEKV-316
P <i>Gamma proteobacterium</i> (ZP_05061507)	293-WRSRRLFIAVRRLAKVKK-EGPQQQLIIEPV----YDYFCYVTT-ERLTPWQA-339
P <i>Azoarcus</i> sp (YP_195484)	286-WKKAKRTLRLVVRVTERTIDKKGQHLLAPEIEIEGWWTSDL-----VPMADV-332
F <i>Bacillus coagulans</i> (YP_004859304)	280-WEKPRRVAVIRKADKYEE---DQLQLFDF----LWDYEAIVTT-MDWEPMDI-323
F <i>Enterococcus casseliflavus</i> (AACX38177)	270-WEKDRRFVVSRVLKPEKE---RAQISLLEG---SEYEYFFFVTN-TTLLSEKV-315
F <i>Desulfotomaculum</i> sp. (YP_001111807)	316-WSKARRFVFIRETQEPKV--SGEQLNF-DL---KTFDYQVIITSSDEYNPEEV-362
F <i>Geobacillus</i> sp. (YP_002949920)	289-IDGNTTYTYVQVTQVTERTMRNGQLMLVPDYEVES-YWVRLKGY--EHVRMSDV-339
B <i>Tannerella forsythia</i> (YP_005013031)	278-WQKPRRIIVIVRQKIEKRPQAGGKQLSLFPEDEIHRNYRYSAYFTN-QTCSMVDV-330
B <i>Prevotella buccae</i> (ZP_07884135)	291-PRRIVMVRQEVEKRPKAAGKQVRQLELFEDEQDFGKYRYSCFVTN-LALPAKIV-343
A <i>Rhodococcus jostii</i> (YP_707285)	314-HQIPGR-LVVRRIPLRPPKDQGQGTLFDI---WRFHAFFTTDPDDLDVTDA-362
A <i>Mycobacterium gilvum</i> (YP_001133130)	278-RGWPAGMRVIARR--ERPHPGA-QLRLTDDNG--WRITCFATNTP--GWSIADL-323

### Tn7 (TnsC)

P <i>Escherichia coli</i> (EIL57895)	491-VKRTEWGCLSEEDLRRRYAD <b>GPESEAYDRFKQAGLILDDLRLPLRAS*</b>
P <i>Escherichia coli</i> (YP_006903369)	438-LQTADDLFERIGNPDLMQHAAAEEKPLELLR <b>QEGVFCEHPLKVFA*</b>
P <i>Acinetobacter haemolyticus</i> (ZP_06726085)	508-KQWNELADTDLRHIFSMNPTSSESIYTQLDR <b>QGLLILQMSDLLS*</b>
P <i>Acidithiobacillus caldus</i> (YP_004750330)	505-KKEKWHALDSHDLRFLFSQASSEEDVYKSLG <b>QKGLFFDTESWVD SYG*</b>
P <i>Vibrio cholerae</i> (EGS60813)	509-PNSWDELPSSDLRFIKSQCKNSTDVHTALNK <b>QGMLLDLSSV LNL AG*</b>
P <i>Desulfovibrio aespoensis</i> (YP_004122276)	500-IPRKDWHS LGSEDLRFVFSQ TDSDKDMYEHLK <b>QNSMIFDMEDWLRKPG*</b>
F <i>Clostridium thermocellum</i> (YP_001037540)	511-MDENDIRYIVEEGRKNKKSAYESLNEKGLIK <b>QVEKDFFKAV*</b>
F <i>Bacillus cellulosilyticus</i> (YP_004093238)	487-LNNNDKSTEKVISNDLRTIIKKGKEQQVTAY <b>QALLDAGVIKQDYPERVVS*</b>
C <i>Crinalium epipsammum</i> (YP_007140767)	482-RKITNKPQLITRETANNATTLIEIVTIGTAH <b>QISGYEALKLAGLIQPTTEY*</b>

### ISL3

P <i>Escherichia coli</i> (ZP_07122173)	351-IIRGEENYA FRFISLMCEKEPELKIA <b>QQLVLEFYRILKT*</b>
P <i>Methylobacterium extorquens</i> (YP_003065656)	478-GPVEGVNRLKLIKRS MYGRAGFDLLR <b>QRVLHAA*</b>
P <i>Shewanella</i> sp. (YP_733579)	364-NGLAEAINSQRQLRVKAMGYRN SERF <b>QRAILFHFGKLDMGFHQ*</b>
P <i>Syntrophus aciditrophicus</i> (YP_461433)	276-LKPDHKERLDALMQANQPLFVMHSMKE <b>QLRLF*</b>
A <i>Mycobacterium gilvum</i> (YP_001131597)	398-RLINARGYGHHTAETLTSMIYLCLGG <b>LQVKLPTRT*</b>
F <i>Lactobacillus brevis</i> (ZP_03940084)	386-EGVNRKIKAI GRTAYGYRNFDHYRTRI <b>QMELFA*</b>
F <i>Streptococcus mitis</i> (YP_003445409)	408-FRNYYHFRNRIILITKMFGPKQKG <b>IQQQLVV*</b>
F <i>Staphylococcus haemolyticus</i> (YP_252293)	407-YRNYDNLRNRIIITSRLFASTTKKEIK <b>QPKVA*</b>
H <i>Petrotoga mobilis</i> (YP_001567687)	342-GLAEGSVNKLKVVKRIMYGRNK <b>FEMLRQKVLFLENNG*</b>

### ISNCYa

P <i>Escherichia coli</i> (EIL58166)	261-RQEERQEFALRLLSKGMSRE <b>DVAEMANLPLAEIDKVINLI*</b>
P <i>Photorhabdus luminescens</i> (Q7N2E9)	284-EEGRAEGKLETARALLRHGVSDL IIVTSTGLSRD <b>KIEMLK H*</b>
P <i>Proteus mirabilis</i> (YP_002152329)	267-EKGIGLGVEKVARSL LQQGVDLNIIMQCTGLTRE <b>KIESLKH*</b>
F <i>Selenomonas sputigena</i> (ZP_05899258)	253-VTEIVLNMLKKKKWSLQDISEVSQWPLDKIESLG <b>KMHQLL*</b>
F <i>Thermoanaerobacter italicus</i> (YP_003476143)	251-GKKEGKIAVAKLLEKGMDIDEIAEITELSKEE <b>IKKLLN*</b>
F <i>Clostridium botulinum</i> (ZP_04824118)	294-KKAIEDAIGFLRLGVSEEIVSKGTGLPIEK <b>VRELKD KINN*</b>
F <i>Caldicellulosiruptor owensensis</i> (YP_004003521)	256-ATIEIAKRLIQKGFSD DEEV AELTELQIEKV <b>KELRKSMLN*</b>
S <i>Leptospira interrogans</i> (Q8F556)	265-GKVETKIDIARNM LLGASLEYVLK ITRLTEQEL <b>KDHGLL*</b>

**TABLE S6. Statistical significance for the non-random orientation of IS elements in the Phyla Bacteroidetes, Cyanobacteria and Spirochaeta. See legend to Table 1 for details.**

	Bacteroidetes			Cyanobacteria			Spirochaeta		
	Chr.	IS	Orient.	Chr.	IS	Orient.	Chr.	IS	Orient.
<b>IS1</b>	7	29	0.426	7	49	0.283	-	-	-
<b>IS3</b>	28	171	0.333	-	-	-	13	50	0.894
<b>IS4b</b>	11	48	0.206	-	-	-	-	-	-
<b>IS4c</b>	-	-	-	4	26	0.254	-	-	-
<b>IS5a</b>	16	112	0.775	-	-	-	5	28	0.656
<b>IS5b</b>	10	68	0.630	13	142	0.470	-	-	-
<b>IS5c</b>	-	-	-	2	22	0.312	-	-	-
<b>IS5d</b>	5	62	0.199	11	154	0.556	-	-	-
<b>IS21</b>	14	62	0.393	-	-	-	-	-	-
<b>IS30</b>	6	23	0.812	-	-	-	6	24	0.869
<b>IS66a</b>	9	30	0.513	-	-	-	-	-	-
<b>IS66b</b>	2	21	0.263	2	22	0.663	-	-	-
<b>IS110</b>	38	151	0.119	6	26	0.398	8	116	0.577
<b>IS200</b>	36	132	0.546	14	59	0.825	-	-	-
<b>IS200/IS605</b>	-	-	-	11	45	0.459	-	-	-
<b>IS256</b>	17	68	0.265	-	-	-	7	66	0.0603
<b>IS607</b>	-	-	-	8	39	0.833	-	-	-
<b>IS630</b>	7	71	0.533	16	392	0.884	-	-	-
<b>IS701</b>	3	11	0.551	11	85	0.855	-	-	-
<b>IS982</b>	17	104	0.676	-	-	-	-	-	-
<b>IS1182</b>	20	78	0.926	3	23	0.583	-	-	-
<b>IS1380</b>	-	-	-	4	22	0.833	-	-	-
<b>IS1595</b>	16	88	0.263	-	-	-	6	24	0.124
<b>IS1634</b>	-	-	-	5	46	0.518	-	-	-
<b>ISAs1</b>	9	28	0.147	6	34	0.366	-	-	-
<b>ISAzo13</b>	-	-	-	5	29	0.359	-	-	-
<b>ISL3</b>	23	81	0.764	8	73	0.775	-	-	-
<b>ISNCYa</b>	-	-	-	19	102	0.247	8	21	0.572