

# Supporting Information

Gonzalez-Perez et al. 10.1073/pnas.1322123111

## SI Materials and Methods

**Measurement of Run-Down and Analysis of Conductance–Voltage Curves.**  $\text{Ca}^{2+}$ - and voltage-activated  $\text{K}^+$  (BK) channels were expressed in *Xenopus* oocytes. To define the loss of  $\gamma 1$ -mediated shifts after patch excision, patches were initially excised into a  $10\text{-}\mu\text{M}$   $\text{Ca}^{2+}$  solution, thus allowing determination of the conductance-voltage (GV) characteristics before any ostensible run-down. The patch was then exposed to one of three different run-down conditions that were tested (condition 1:  $0\text{ Ca}^{2+}$ ,  $0\text{ mV}$ ; condition 2:  $10\text{ }\mu\text{M}$   $\text{Ca}^{2+}$ ,  $0\text{ mV}$ ; condition 3:  $0\text{ Ca}^{2+}$ ,  $+80\text{ mV}$ ) for a 2-min interval before return to  $10\text{ }\mu\text{M}$   $\text{Ca}^{2+}$  to monitor the status of the GV curve. Elapsed times during run-down experiments reflect the cumulative time in the run-down condition. To define the shift, we monitored the time course of appearance of a component of channels with a  $V_h$  (the voltage at which BK channels are half activated) for activation indistinguishable from native Slo1 channels, which we term the unshifted component. Thus, irrespective of the behavior of the shifted portions of channels, the appearance of the unshifted component reflects the fraction of channels for which there is no residual  $\gamma 1$  effect. For analysis of the run-down time course, we only used patches that lasted long enough that the unshifted component reached a fractional value of at least 0.72. Such patches provided a minimum of four points for evaluation of the run-down time course, while also ensuring that we had sufficient patches for analysis.

In experiments in which  $\alpha$ -subunits were titrated with various amounts of  $\gamma 1$ -subunits, we evaluated a variety of fitting procedures. As shown in our examples in Figs. 1 and 2, a two-component Boltzmann function 
$$G(V) = G_s \left( \frac{1}{1 + e^{\frac{z_1 F(V-V_s)}{RT}}} \right) + G_u \left( \frac{1}{1 + e^{\frac{z_2 F(V-V_u)}{RT}}} \right),$$
 where  $G_s$  represents the shifted fraction of conductance of  $V_s$  as the shifted voltage of half activation, and  $G_u$  and  $V_u$  reflect the unshifted fraction.  $z_1$  and  $z_2$  are the terms for voltage-dependence.

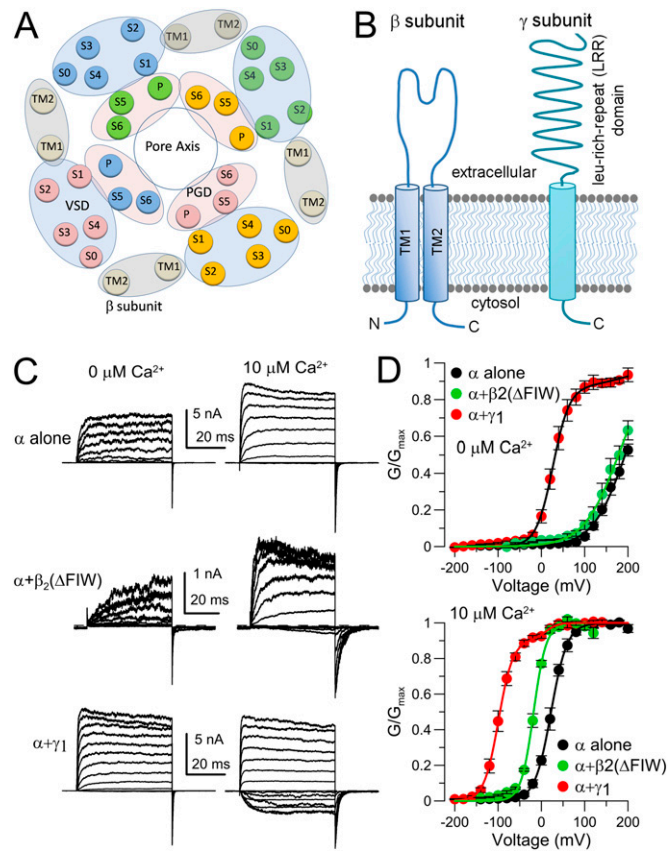
1. Horrigan FT, Aldrich RW (2002) Coupling between voltage sensor activation,  $\text{Ca}^{2+}$  binding and channel opening in large conductance (BK) potassium channels. *J Gen Physiol* 120(3):267–305.

Typically,  $z_1$  and  $z_2$  were constrained to be identical. We also tested whether additional Boltzmann components might improve the quality of the fits to such data but observed no improvement in fit quality by adding additional components.

**Model Predictions.** Predictions for binomially predicted fractional occupancies of channels with  $\gamma 1$ -subunits under different models of assembly were generated using Mathcad 15 (PTC). Predictions for GV curves based on a given fractional occupancy were based on the summation of Horrigan-Aldrich predictions for the GV for each individual stoichiometric contribution, also generated with MathCad 15. Standard values for allosteric constants (1) were used for these calculations:  $L = 10^{-6}$ ,  $z_L = 0.3e$ ,  $J_0 = 0.03$ ,  $z_J = 0.58e$ ;  $D = 25$ ,  $C = 8$ ,  $K = 11\text{ }\mu\text{M}$ ,  $E = 2.4$ . For simulations of incremental and all-or-none gating shifts in which changes in a single allosteric constant were assumed to a given modulatory effect of an auxiliary subunit, a minimal (min) and maximal (max) parameter value were used. For  $D$ ,  $D_{\min} = 25$  and  $D_{\max} = 425$ , consistent with estimates of  $\gamma 1$  induced effect on  $D$  (2); for  $L$ ,  $L_{\min} = 10^{-6}$  and  $L_{\max} = 400 \times 10^{-6}$ ; for  $J_0$ ,  $J_{0\min} = 0.03$  and  $J_{0\max} = 0.83$ ; for  $C$ ,  $C_{\min} = 8$  and  $C_{\max} = 32$ . For  $L$ ,  $J_0$ , and  $C$ , maximum values were simply chosen to produce maximal GV shifts of magnitude comparable to that observed with  $\gamma 1$ -subunits.

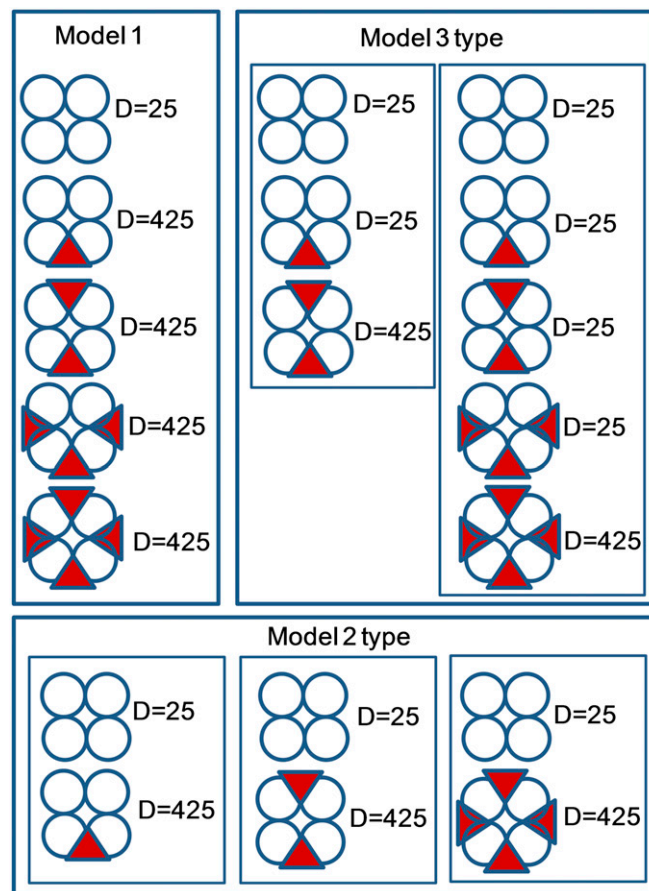
**Chemicals.** Salts and buffers were obtained from Sigma.  $\text{Mg}^{2+}$ -ATP was from Sigma. Solutions of natural phosphatidylinositol 4,5-bisphosphate (PIP<sub>2</sub>) from brain (Avanti Polar Lipids) or synthetic diC16-PIP<sub>2</sub> or diC8-PIP<sub>2</sub> (Echelon Biosciences) were prepared by initially dissolving in water at 1 mM and stored as aliquots. Before the experiments aliquots were dissolved in working solution to make 10 or 25  $\mu\text{M}$  final concentrations.

2. Yan J, Aldrich RW (2010) LRRC26 auxiliary protein allows BK channel activation at resting voltage without calcium. *Nature* 466(7305):513–516.



**Fig. S1.** Two distinct types of auxiliary subunits of BK channels,  $\gamma_1$  and  $\beta$ , produce robust shifts in gating. (A) Schematic of deduced positions (1, 2) of  $\alpha$ - and  $\beta$ -subunit transmembrane (TM) segments at the extracellular face. TM segments for a given  $\alpha$ -subunit share an identical color. Pink ellipses highlight pore-gate domain (PGD) elements; blue ellipses highlight voltage-sensor domain (VSD) elements; gray ellipses highlight  $\beta$ -subunits. (B) Cartoons of the topology of  $\beta_2$ - and  $\gamma_1$ -subunits highlighting N termini (NT), cytosolic C termini (CT), and extracellular domains. (C) Current activation over voltages from  $-200$  to  $+200$  mV at  $0$  and  $10 \mu\text{M}$   $\text{Ca}^{2+}$  for  $\alpha$ -subunit alone (Top),  $\alpha+\beta_2(\Delta\text{FIW})$  (Middle), and  $\alpha+\gamma_1$  (Bottom). (D) GV curves from sets of currents as in C at  $0 \text{ Ca}^{2+}$  (Upper) and  $10 \mu\text{M}$   $\text{Ca}^{2+}$  (Lower) for  $\alpha$  alone (black),  $\alpha+\beta_2(\Delta\text{FIW})$  (green), and  $\alpha+\gamma_1$  (red), highlighting the more than  $100\text{-mV}$  leftward shift in  $V_h$  produced by  $\gamma_1$  at both  $0$  and  $10 \mu\text{M}$   $\text{Ca}^{2+}$ , whereas  $\beta_2$  produces no shift at  $0$  mV relative to  $\alpha$  alone, but an  $\sim 50\text{-mV}$  shift at  $10 \mu\text{M}$   $\text{Ca}^{2+}$ .

1. Wu RS, Marx SO (2010) The BK potassium channel in the vascular smooth muscle and kidney:  $\alpha$ - and  $\beta$ -subunits. *Kidney Int* 78(10):963–974.
2. Liu G, et al. (2010) Location of modulatory beta subunits in BK potassium channels. *J Gen Physiol* 135(5):449–459.



**Fig. S2.** Possible stoichiometric combinations of  $\gamma_1:\alpha$ -subunits that could underlie all-or-none gating shifts. Model 1 allows one to four auxiliary subunits per channel, but one subunit is sufficient to produce a full change in a given allosteric constant. Here the value for the allosteric constant  $D$  in relation to a given subunit combination reflects an all-or-none effect on that parameter when a single auxiliary subunit is present. Model 1 would also include a version in which only up to two  $\gamma_1$ -subunits can assemble in a channel, but one is sufficient to produce a full effect on an allosteric constant. Model 2 allows a single  $\gamma_1$ -subunit (or  $\gamma_1$ -subunit complex) to assemble per channel, such that an all-or-none change in an allosteric constant occurs for channels associated with the full  $\gamma_1$  complex. Model 3 represents a case when up to some number of  $\gamma_1$ -subunits can be present in a channel complex, but the gating shift only occurs when all required subunits are present.















