

SUPPLEMENTARY FIGURE LEGENDS

Figure S1 (related to Figure 1) *In situ* mito-sypHer pH titration. HeLa cells expressing mito-SypHer were sequentially bathed with a high K^+ solution containing 5 $\mu\text{g/ml}$ Nigericin at pH 5.5, 6.5, 7.0, 7.5 and 8.0. Average of 5 independent experiments (34 cells, means \pm SEM).

Figure S2 (related to Figure 2) Effect of Letm1 levels on pH flash frequency. (A) Letm1 mRNA levels quantified by real time PCR, data normalized by the mRNA averaged levels of 3 housekeeping genes. (B) Frequency of pH flashes in Letm1 siRNA transfected cells compared with control. Data obtained by the analysis of cell frequency in 10 different coverslips per experimental condition. (C) Effect of 2 μM CCCP on oxygen consumption by HeLa cells, measured with a Clark electrode.

Figure S3 (related to Figure 3) *In vitro* superoxide generation and SypHer properties. (A) Superoxide generation by xantine/xanthine oxidase. O_2^- production was measured with MCLA in the buffer used to record SypHer spectra (pH 7.5). Average of 3 independent experiments (means \pm SEM). Addition of 100 mU xanthine oxidase caused a transient increase in luminescence that was completely prevented by superoxide dismutase (SOD, red trace). Representative of 3 independent experiments for each condition. (B-E) Excitation spectra of purified SypHer ($\lambda_{em}=530\text{nm}$) recorded at pH 7.5 in the presence of increasing amounts of SNAP, DTT, CaCl_2 , and ATP. Data are representative of 3 independent experiments for each condition.

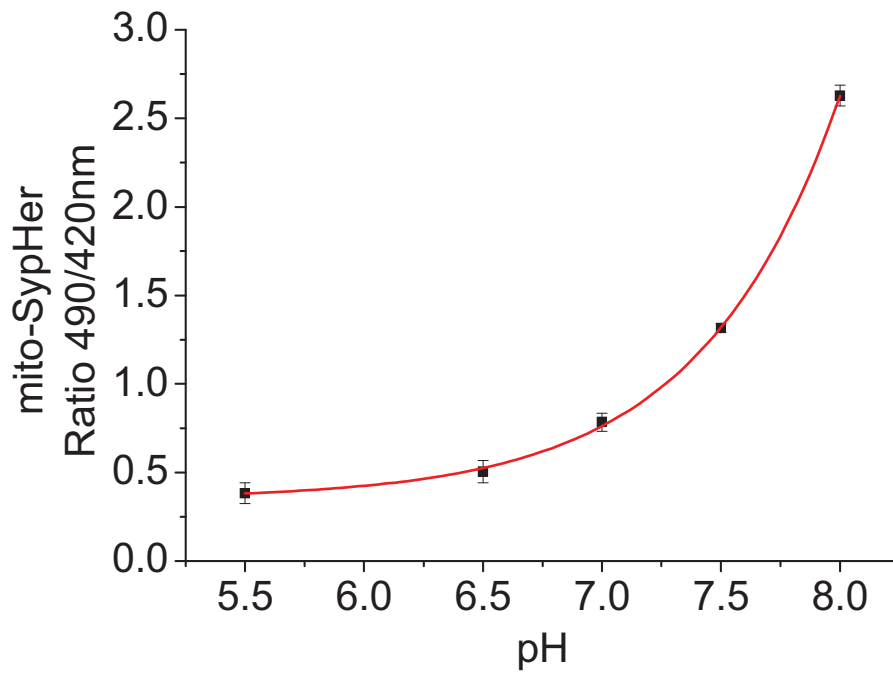
Figure S4 (related to Figure 4). pH_{mito} flashes are driven by concomitant decreases in $\Delta\Psi_m$. (A) HeLa cells expressing mito-SypHer were permeabilized with 100 μM digitonin for 1 min in KCl-based solutions, and the bath pH was then varied from pH 7.5 to 7.0 and 6.5. Traces are representative recordings from 6 different cells. (B) pH_{mito} elevations evoked by short pulses of 30mM NaCl and 30mM LiCl. Permeabilized cells expressing mito-SypHer were equilibrated with 5 μM A23187 and exposed to brief pulses of 30mM NaCl and 30mM LiCl to induce a transient mitochondrial depolarization (n=29 cells). (C) Spatiotemporal coincidence of $\Delta\Psi_m$ and pH_{mito} fluctuations. Simultaneous recordings at 28 frames per second of mito-SypHer and TMRM in HeLa cells. The fluorescence traces correspond to the region of interest zoomed out in the upper panel.

Figure S5 (related to Figure 5). Effects of DRP1^{K38A} and hFIS1 on pH_{mito} flash kinetics and paGFP matrix diffusion. (A) Averaged time to peak, half-life time, and amplitude of the pH_{mito} flashes shown in Figure 5A. (n=90, means ±SEM). (B) paGFP matrix diffusion after photoactivation. Images show merged mito-RFP/paGFP fluorescence at different times after paGFP photoactivation. (C) Kinetics of paGFP spatial propagation following photoactivation in 40 different mitochondrial areas. (D) TMRM and paGFP fluorescence intensities before and immediately after photoactivation in HeLa cells expressing mito-paGFP and loaded with 4nM TMRM. (n=25, means ±SEM). (E) TMRM fluorescence intensity within irradiated area before and after laser illumination in HeLa cells loaded with 4nM TMRM. (n=74, means ±SEM).

Figure S6 (related to Figure 6). Effects of *Opa1* and *Drp1* ablation on pH_{mito} flash properties. (A-D) Averaged frequency, amplitude, and kinetics of the pH_{mito} flashes shown in Figure 6 (n=50 events). (E) Effect of 20 μM Atractyloside on the pH_{mito} flashes recorded in wt and in *Opa1*^{-/-} MEFs cells. (n=50 events; means ± SEM).

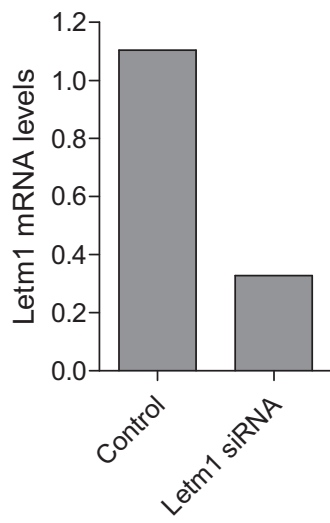
Figure S7 (related to Figure 7). Effect of pH_{mito} flashes on ΔΨ_m and pH_{mito} heterogeneity. (A) Standard deviation of TMRM and SypHer fluorescence intensities measured within flashing regions before and after ΔΨ_m/pH_{mito} flashes (n=32 events) (B) Standard deviations of TMRM/mitoGFP ratio fluorescence measured within flashing regions before and after flashes. (n=20 events; means ± SEM).

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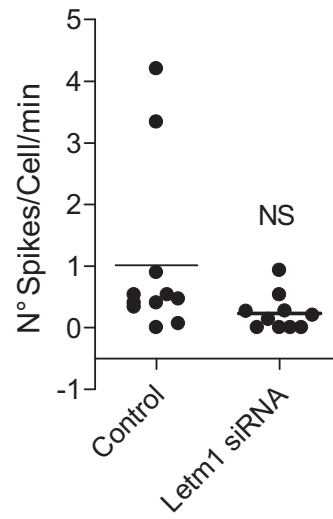


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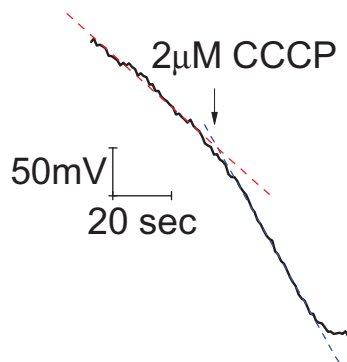
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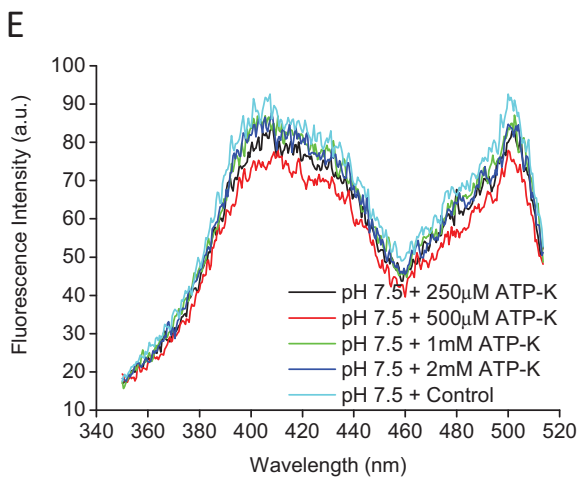
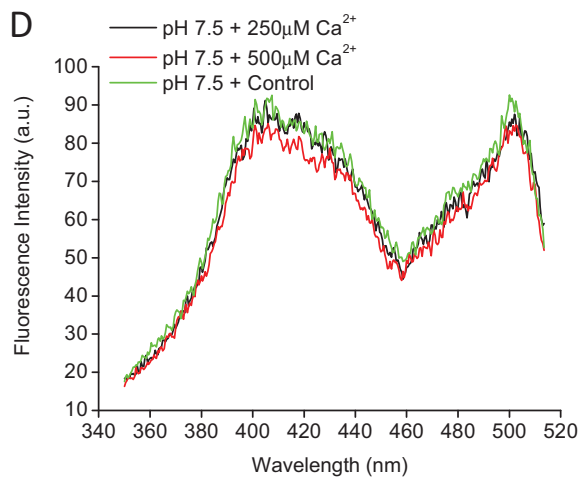
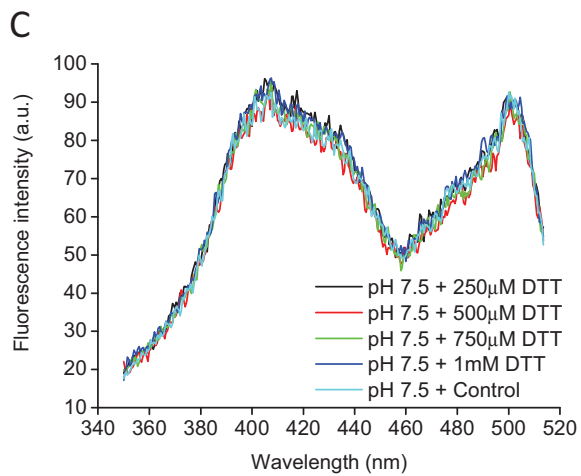
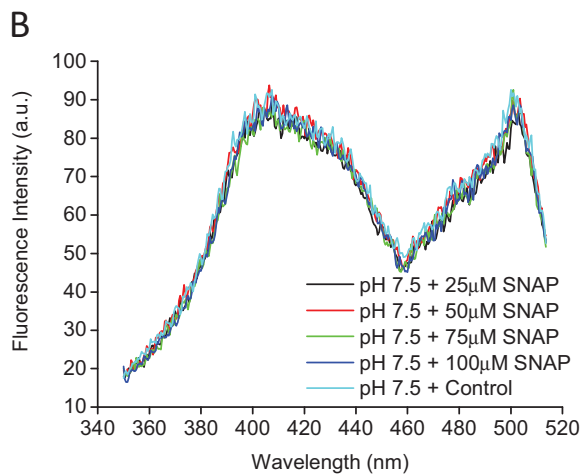
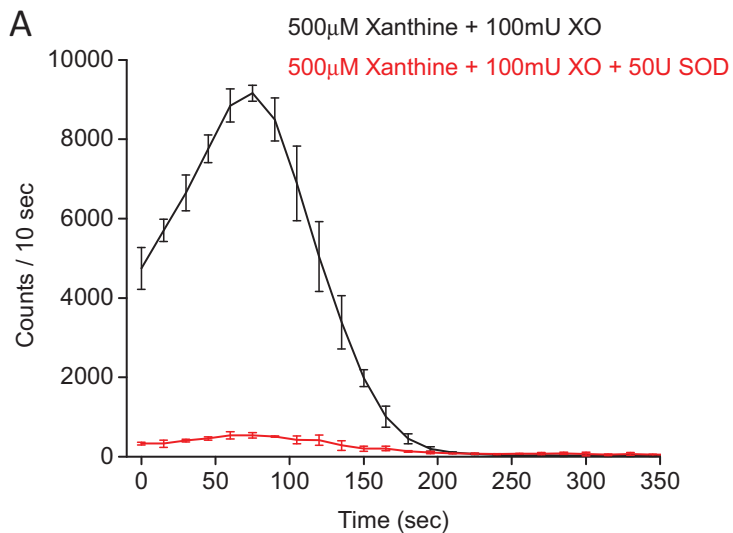
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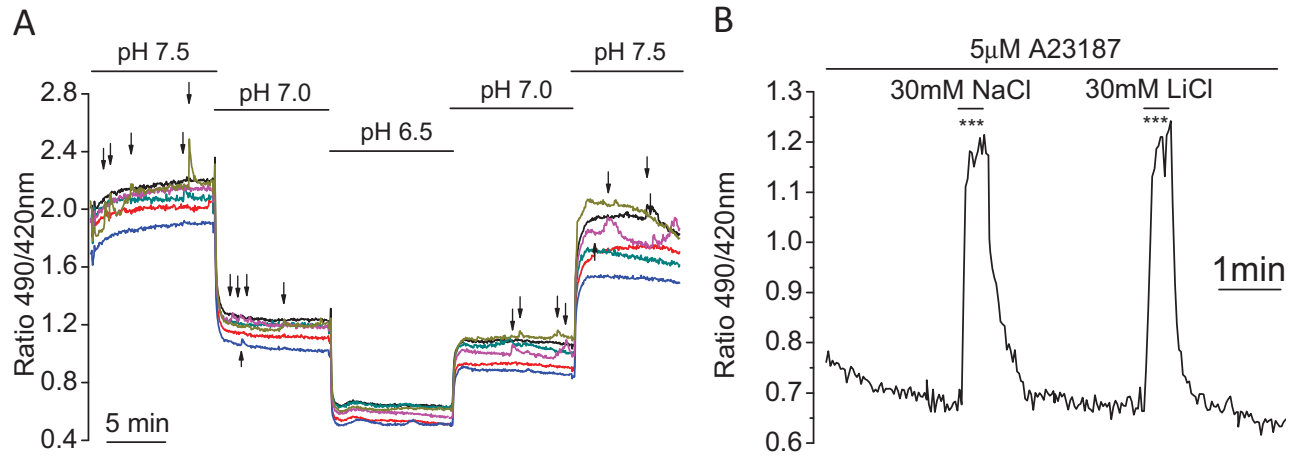
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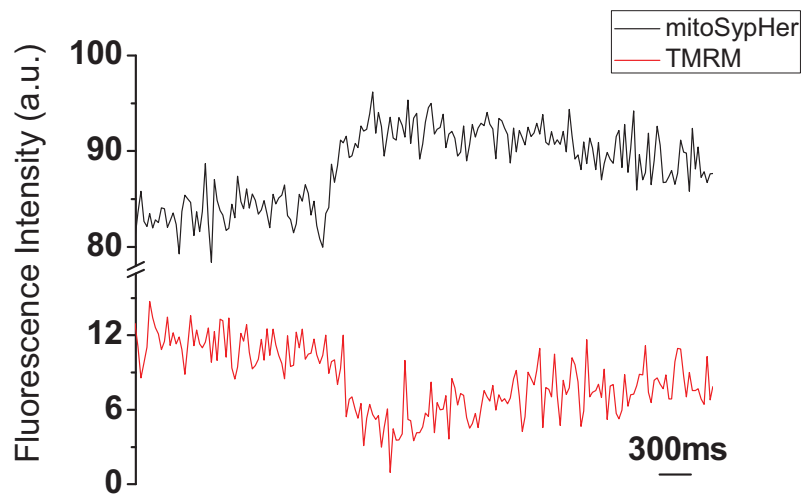
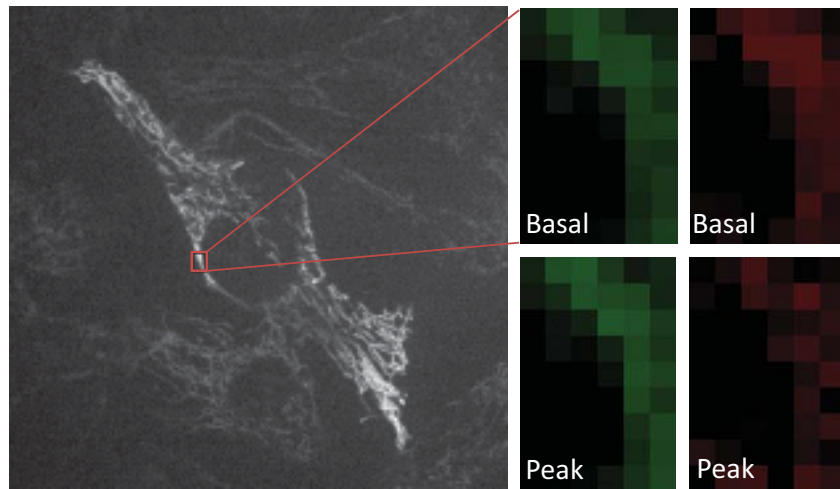
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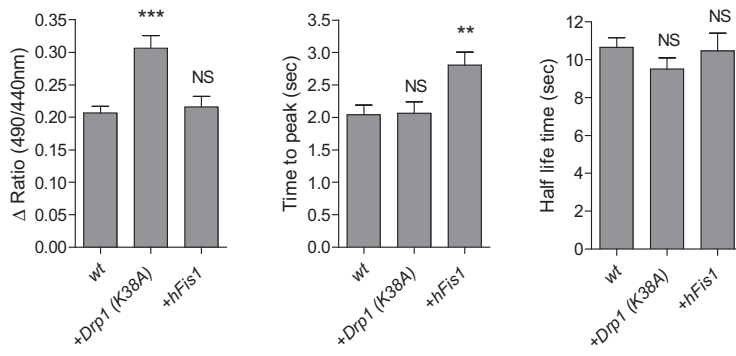


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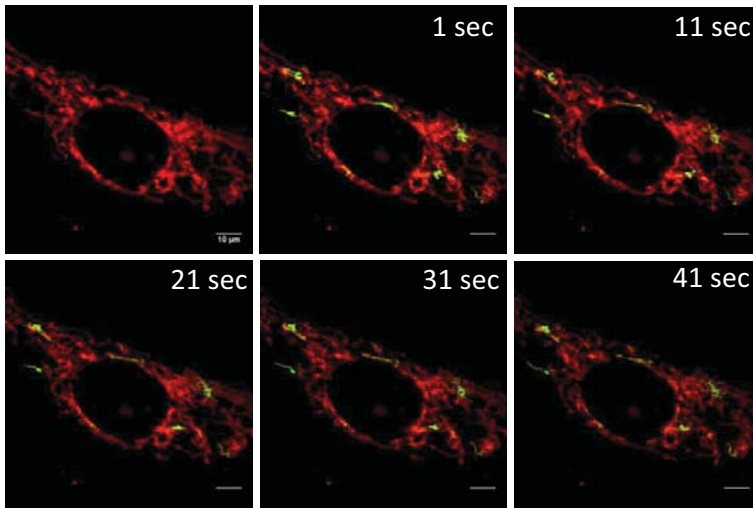


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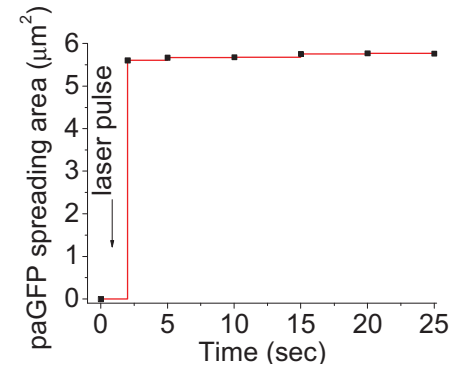
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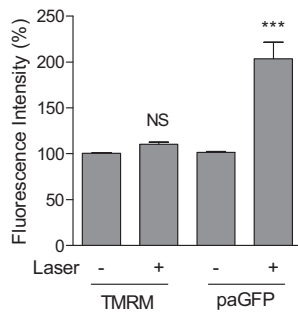
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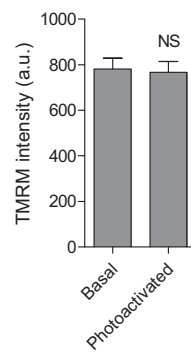
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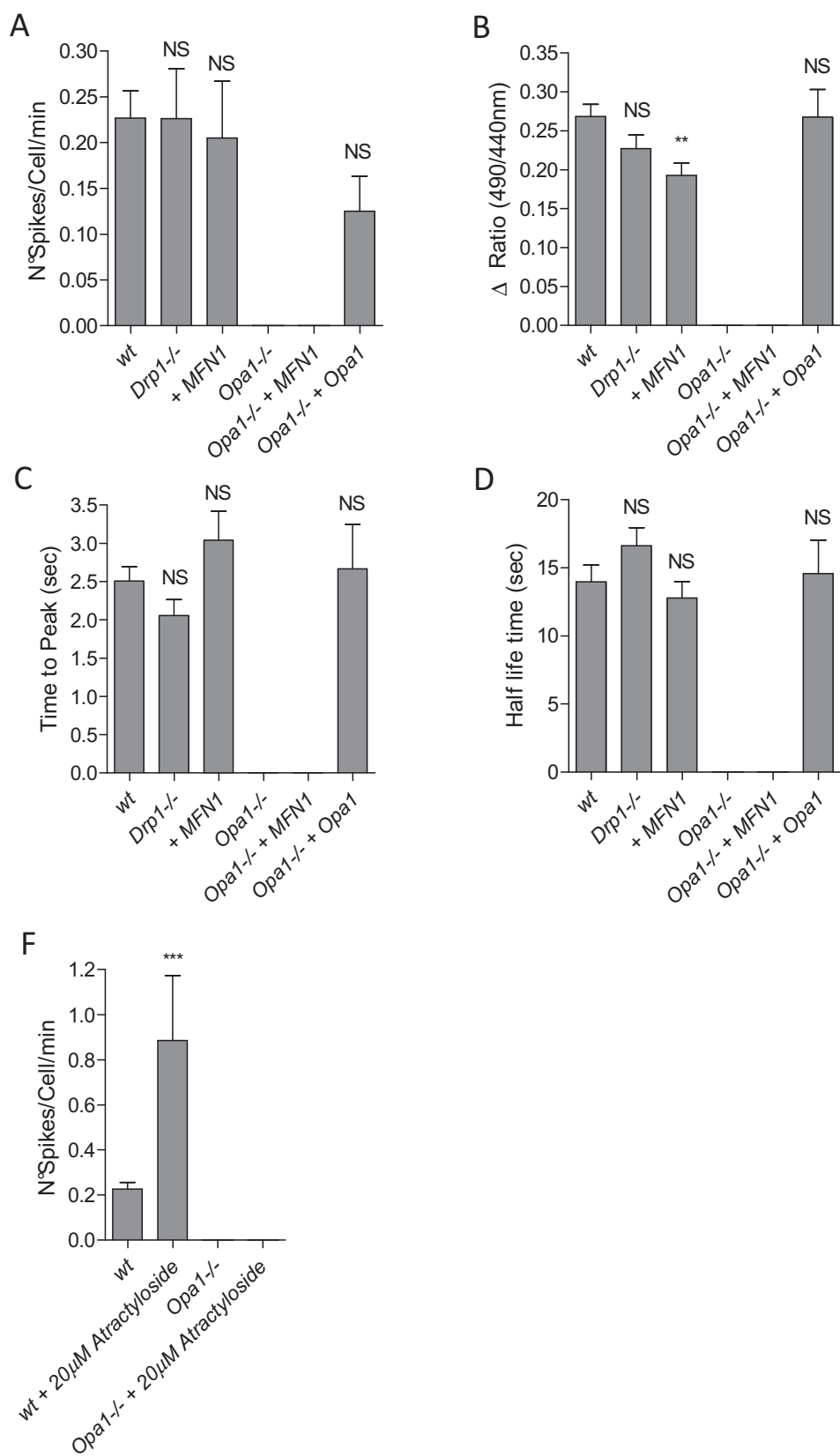
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