

Table S1 Known SD binding site sequences used to generate position weight matrix to scan the SAM and ORE genomes for predicted SD binding sites.

Gene	Coordinates	Sequence	Source
ct	X:7424390..7424403	TTTGTGAATGAAGT	REDfly TF000140
ct	X:7424422..7424433	TAACATTTAATT	REDfly TF000139
ct	X:7424473..7424487	GATAAACAGCAGTGT	REDfly TF000138
ct	X:7424494..7424520	GCTGTTTTTTTAAATGAATTTCTCTA	REDfly TF000137
ct	X:7424594..7424610	AAAATTATTGAAATTAC	REDfly TF000136
ct	X:7424692..7424701	GGAATGGGAT	REDfly TF000135
ct	X:7424821..7424843	AATGTAATTCGAAAAATGTCGTC	REDfly TF000134
vg	2R:8782964..8783003	GCTAGTTGGAATGTGCTATGAAATGTCGCCGGAATGCGAT	REDfly TF001715
vg	2R:8783977..8783987	GGAAATATCTT	REDfly TF000458
vg	2R:8784014..8784027	TGGGAATCCACGG	REDfly TF000459
vg	2R:8784101..8784114	CACGCGGCATGGCA	REDfly TF000460
vg	2R:8784375..8784386	GTTTGGGAATGTT	REDfly TF000465
bs	2R:20229832..20229857	TAAGAAATTCCTGGCATAGTTAAGT	REDfly TF000512
salm	2L:11454576..11454582	TATGCGA	REDfly TF000032
salm	2L:11454656..11454679	AATGGACATTCGTGGGATTCCAGA	REDfly TF001599
salm	2L:11454657..11454680	ATGGACATTCGTGGGATTCCAGAA	REDfly TF000029
salm	2L:11454657..11454680	ATGGACATTCGTGGGATTCCAGAA	REDfly TF000030
kni	3L:20700051..20700068	TACATTTGTCGCATAGTT	REDfly TF000811
kni	3L:20700243..20700251	ATACATACA	REDfly TF000812
kni	3L:20700303..20700311	AAAATGTCG	REDfly TF000813
kni	3L:20700350..20700358	GAAATGCGT	REDfly TF000814
kni	3L:20700420..20700428	AGAAATAGT	REDfly TF000815
diap1		GCATTCCATT	Wu et al. 2008

Table S2 Position weight matrix used to scan the SAM and ORE genomes for SD binding sites, derived from a MEME (Bailey et al. 2009) scan of 23 known SD binding sites.

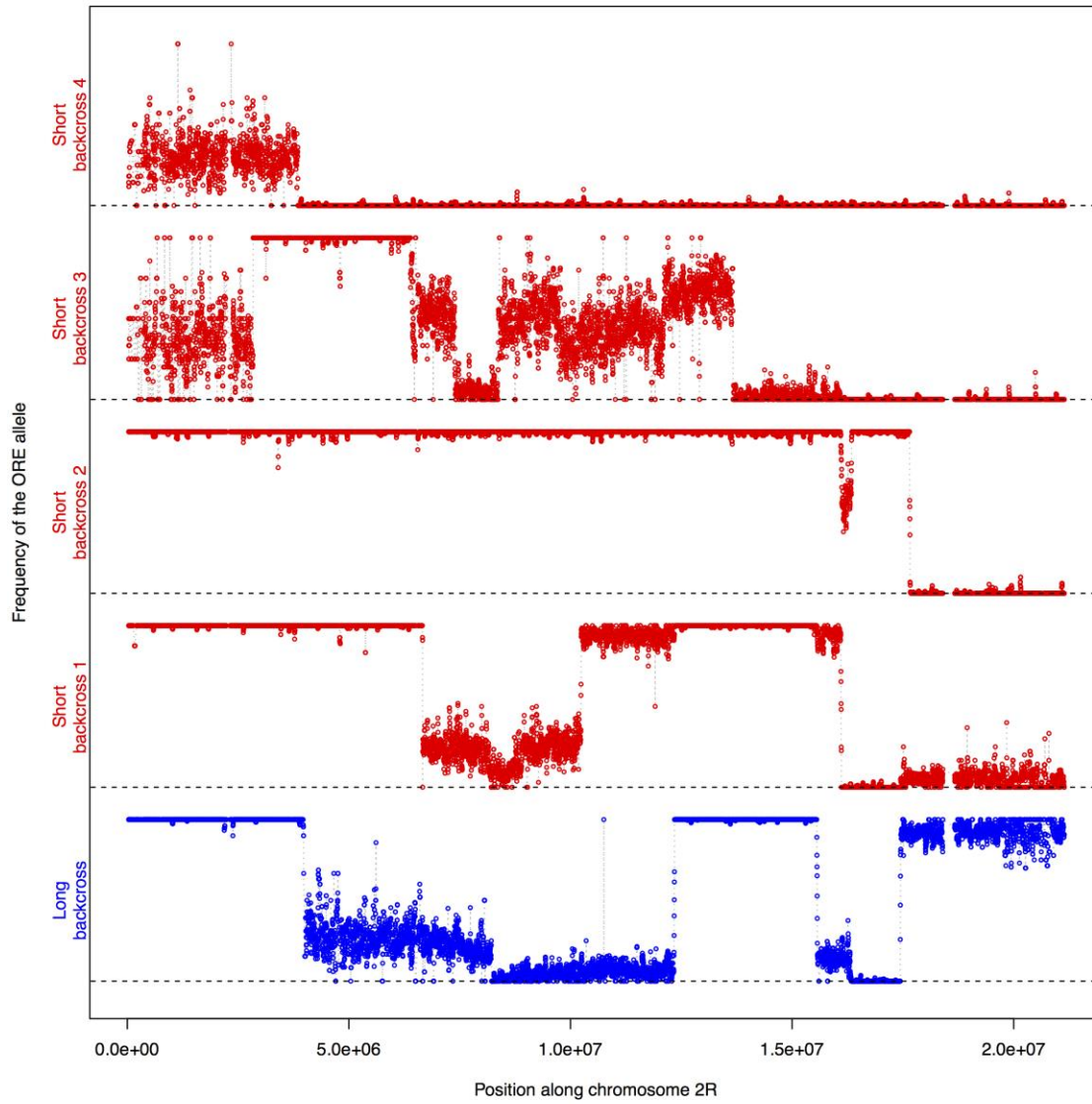
	A	C	G	T
1	0	5	14	4
2	8	1	14	0
3	18	4	0	1
4	22	0	1	0
5	0	0	0	23
6	3	0	14	6
7	3	3	3	14

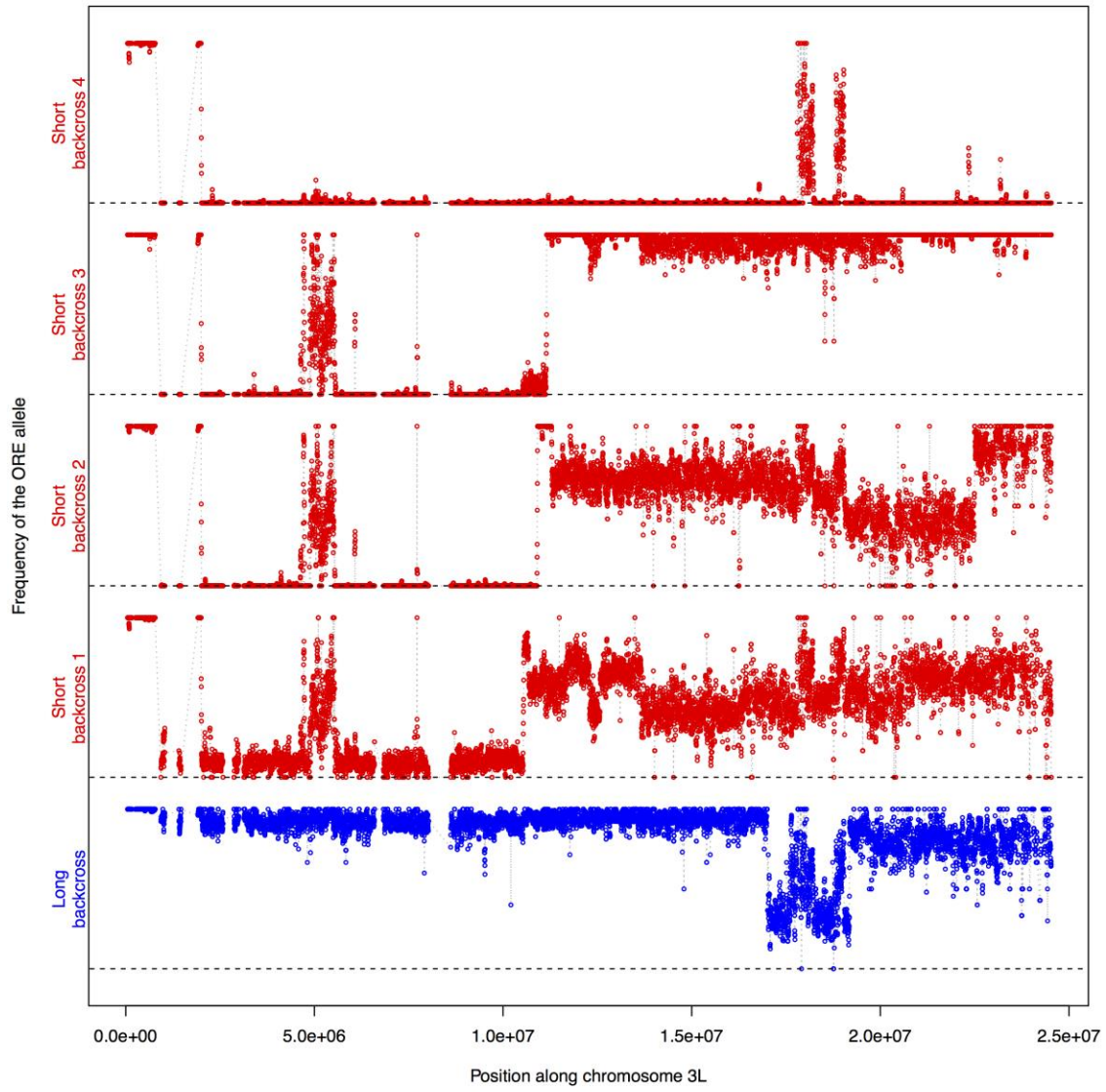
Table S3 Detailed sequence pile-up (Li et al. 2009) for *scalloped* binding site regulating *cut* (Halder & Carrol 2001, Guss et al. 2001).

Chrom.	Pos.	Ref.	# reads (SAM)	Read bases (SAM)	Base quality (SAM)	# reads (ORE)	Read bases (ORE)	Base quality (ORE)
X	7424817	T	15	HFIHIDIBI<GIIII	15	.\$.....	DIIFIIIIIIHII
X	7424818	G	15	HIHGIEIGIGGIIIG	15^]	GIIIIHGIIIIHIE
X	7424819	T	16	,\$.....^].	EHGGICHGIGDIIHGE	15	,\$.....	EIIIIIIIIII
X	7424820	C	15	IIII4IBIBGIIIGI	16^],^].	IGGIIIIIIIBIIEE
X	7424821	A	15G.	IHIICIBIGGII*I	18^],^].	IIIIIIIBIIIIIDBB
X	7424822	A	15	FHHIEIGIDGIIGGI	18	IIIIIIIIIIHIIIEE
X	7424823	T	15	HIIDIGICGIIIGI	18	IIIIIIHHI@IIIGII
X	7424824	G	15	GIIIDI@IEGIGIGI	18	IIIIIIIDIIIIHIIIG
X	7424825	T	15	,\$.....	EGGG=HDCGIIIGI	18	IIIIIIIIIGIIEHII
X	7424826	A	14	IHIBIGIDGGIIGI	18	IIIIIIIIIGIIII
X	7424827	A	14	HII:I4IDGIIIGI	19^].	IIIIIIIIIIHIG>
X	7424828	T	14	III=IEIEGIIII	22^],^],^].	IHIIIIIIIIIIHIEBBB
X	7424829	T	14	III?IBICDIIII	23^].	IIHIIIFIIHIIIFIHICEEED
X	7424830	C	14	IIH?IDIDDIIII	23	IIIIIIIIHIGIIHBFHF
X	7424831	G	14	,\$.....	EDIDIGICGIIII	24^].	IIIIIIIIIIIIIGBIHIE
X	7424832	A	14^].	>>DFGI>EIIII7	25^].	IIHIIIIIGIIIIHGHIGI>
X	7424833	A	14	,\$.....	;<IGIGHIIII:	25	IIIIIIIIHIIIIIDI>II?
X	7424834	A	13	,\$.....	9?IBIGHIIII5	25	IIIIIIIIIIIEIH>IIHIA
X	7424835	A	12	BIGIDHIIII5	25	IIIIIIIIIIIIIGIHHIB
X	7424836	A	12	DIGIDHIIII2	25	IIIGIIIIIIHIIIEIIGIHHIE
X	7424837	T	12	@IBGBEFBIII-	26^].	IIIIIIIIII8IHHHI@ADHDEDIE
X	7424838	G	12	GIGHDDIIII:	26	HIIIIIIIIIIIIIGHHIIII
X	7424839	T	12	GH@IGGHIFHI4	26	HIIIIIIHIIIGIIII3IHHIHI
X	7424840	C	12	GIGI<GIIII5	26	IIIIIIIIIIHIG8IHDHII
X	7424841	G	13^].	:IGIGGIIGII7E	26	IIIIIIIIIIHIIIGIHHII

Table S5 Top 200 genes identified as being differentially expressed between wild-type and *sd^{E3}* mutant flies by DGE analysis. (See SuppTable5.csv in Dryad package, doi:10.5061/dryad.1375s)

Table S6 Genes showing evidence of allelic imbalance in “hybrid” SAM/ORE flies (i.e., in which one of the two alleles is transcribed at significantly higher levels). (See SuppTable6.csv in Dryad package, doi:10.5061/dryad.1375s)





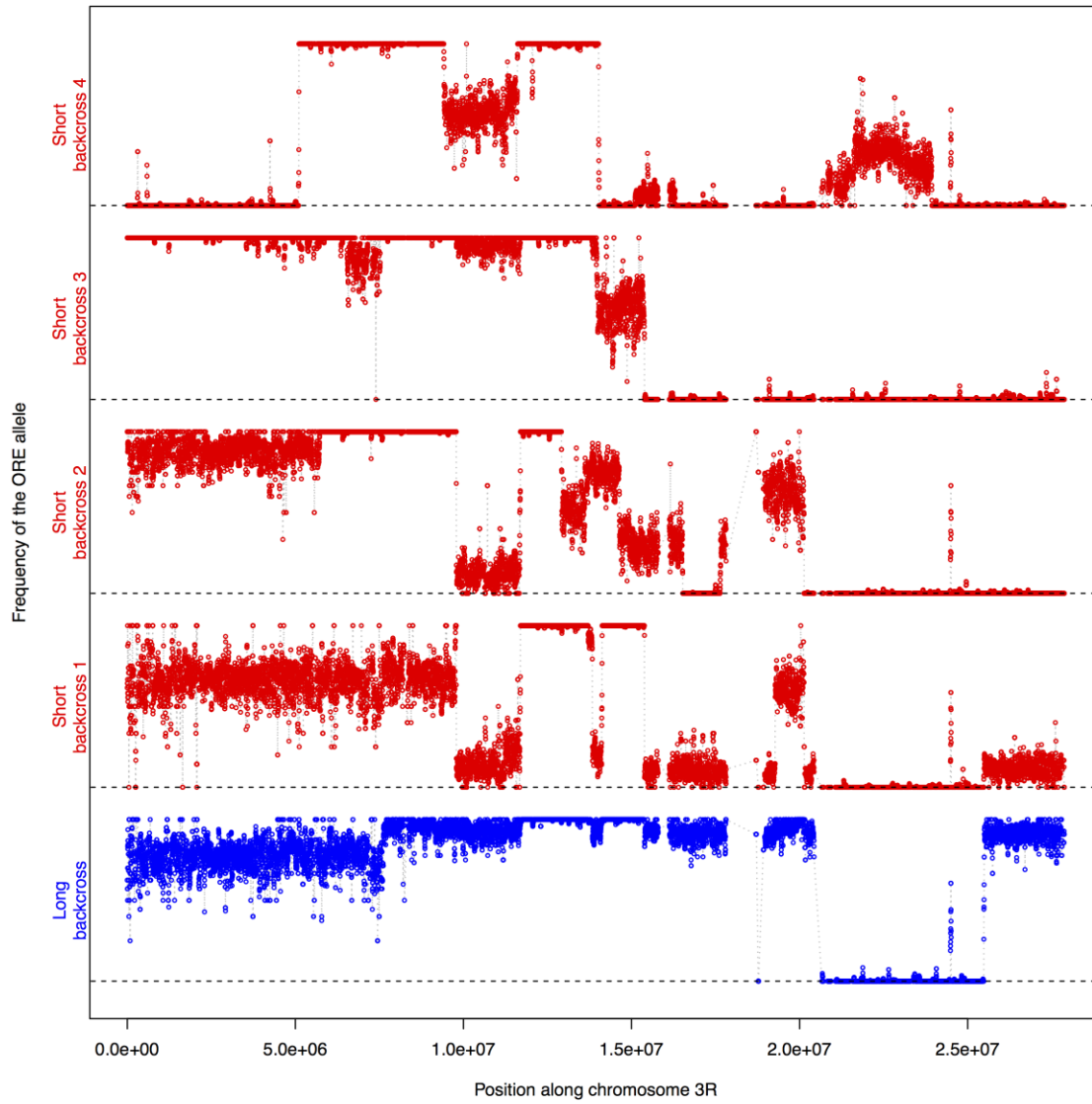


Figure S1 Frequency of the ORE allele along the length of chromosome arms 2R, 3L, and 3R in short- and long-wing sd^{E3} backcross lines.

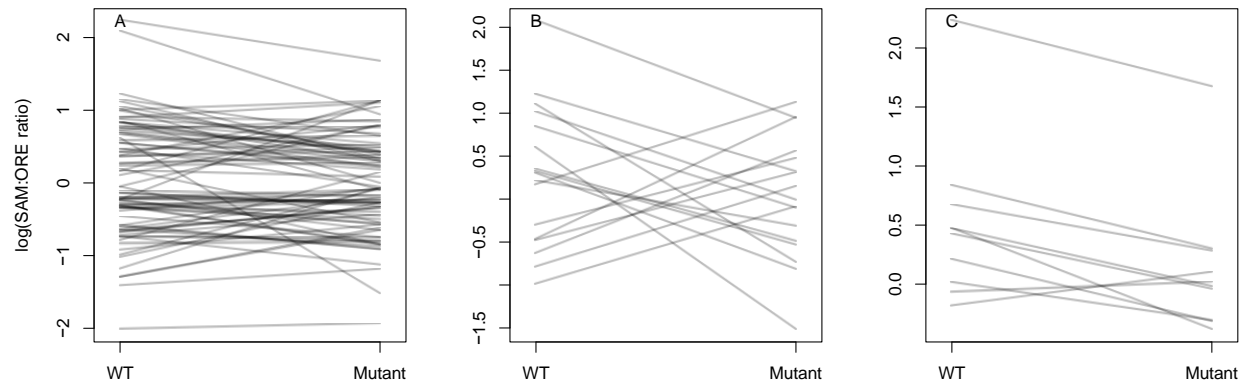


Figure S3 Allelic imbalance in SAM/ORE “hybrid” flies with both wild-type and *sd^{E3}* genotypes. Only sequence tags represented by at least five reads of each allele in all samples are represented in this plot. (A) Genes showing evidence of allelic imbalance ($q < 0.05$). (B) Genes showing evidence of genotype-dependent allelic imbalance ($q < 0.05$). (C) Genes near polymorphic predicted SD binding site ($p < 0.05$).

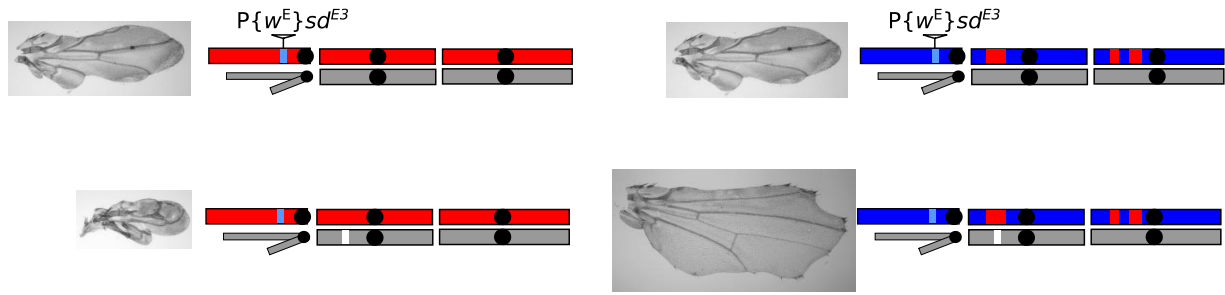
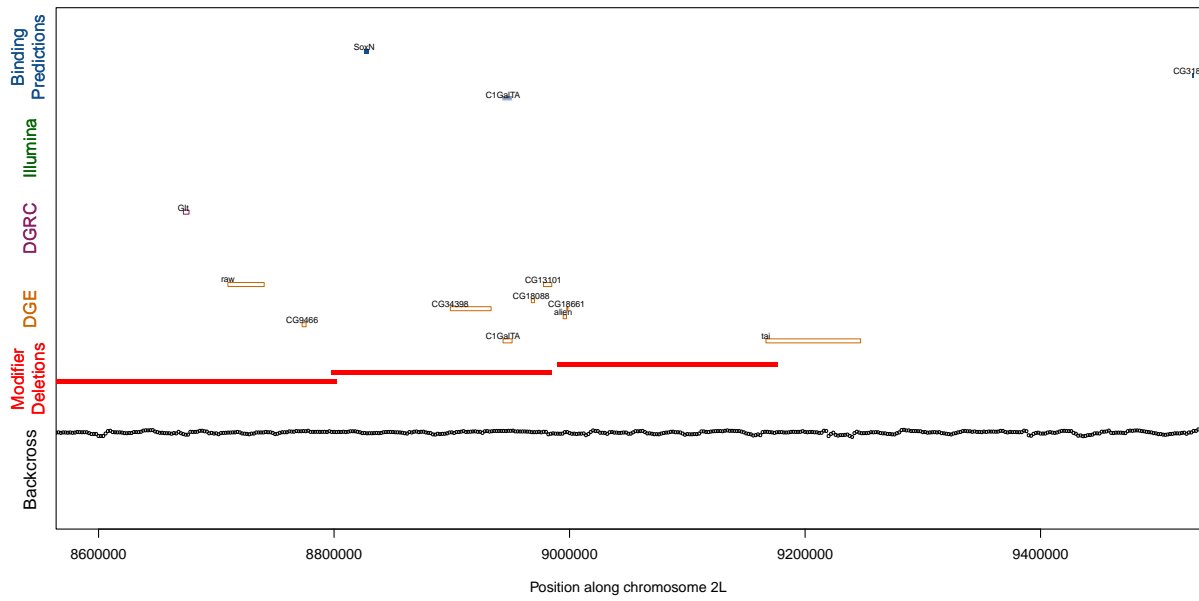
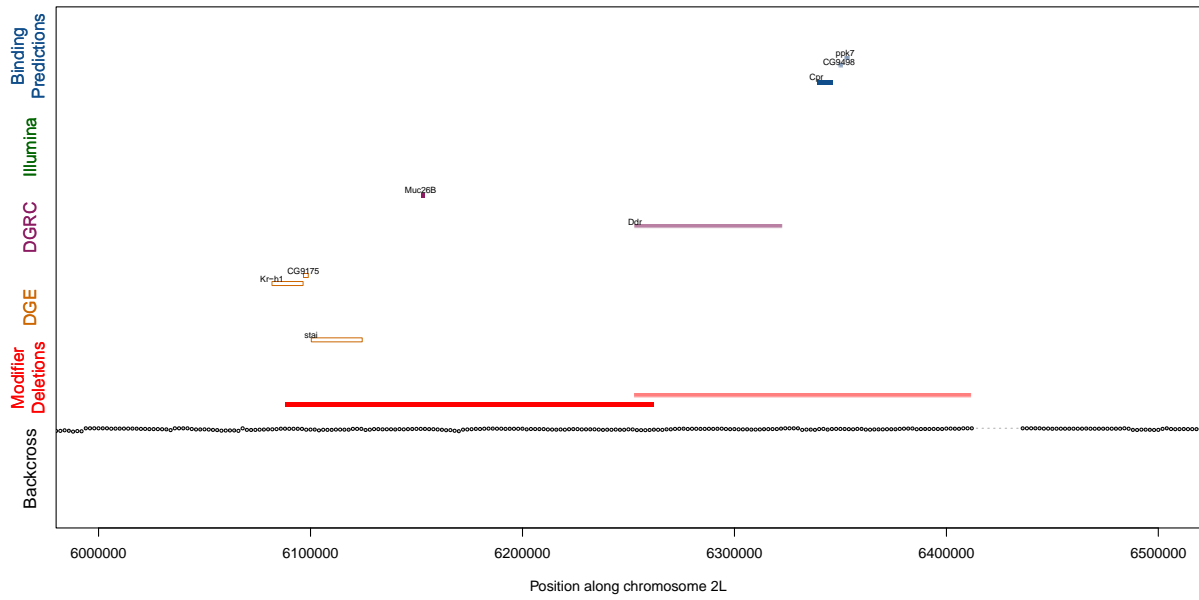
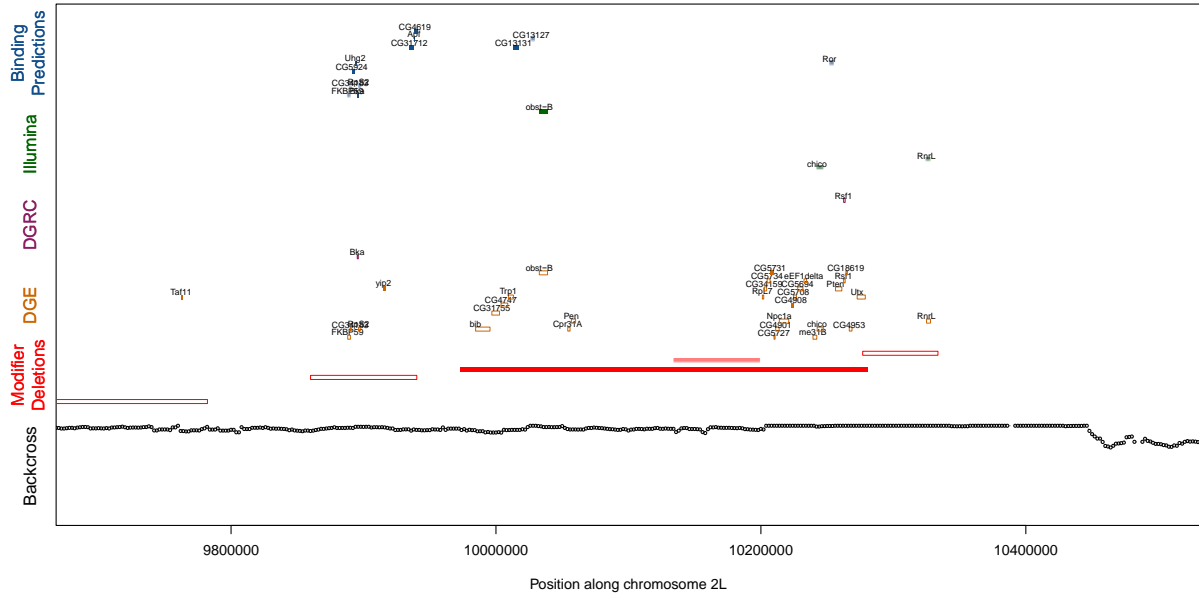
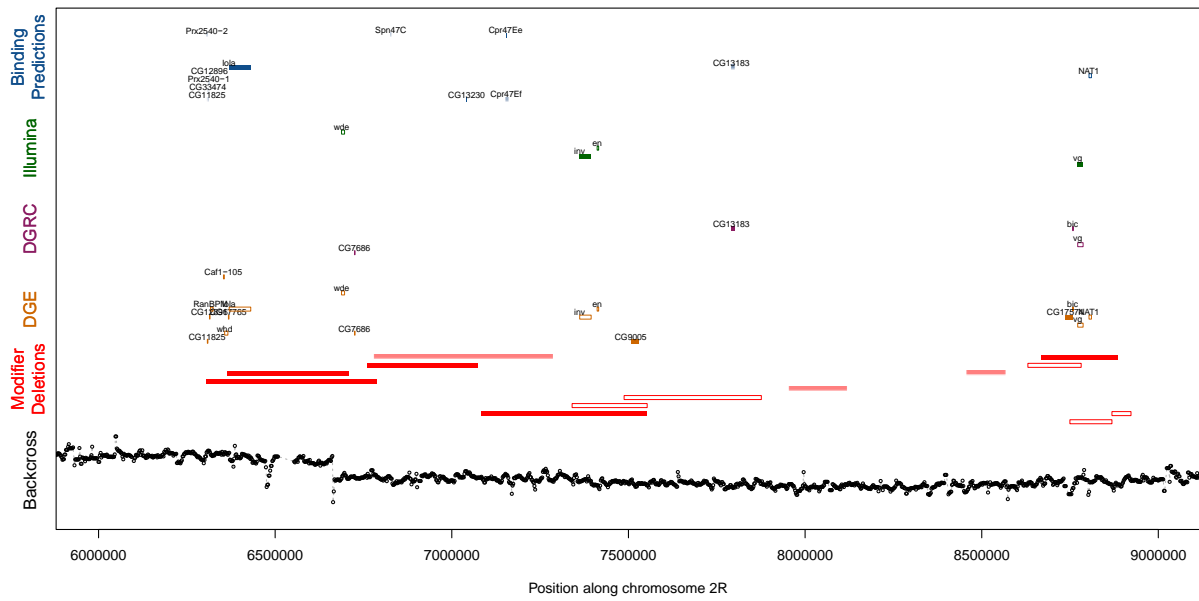
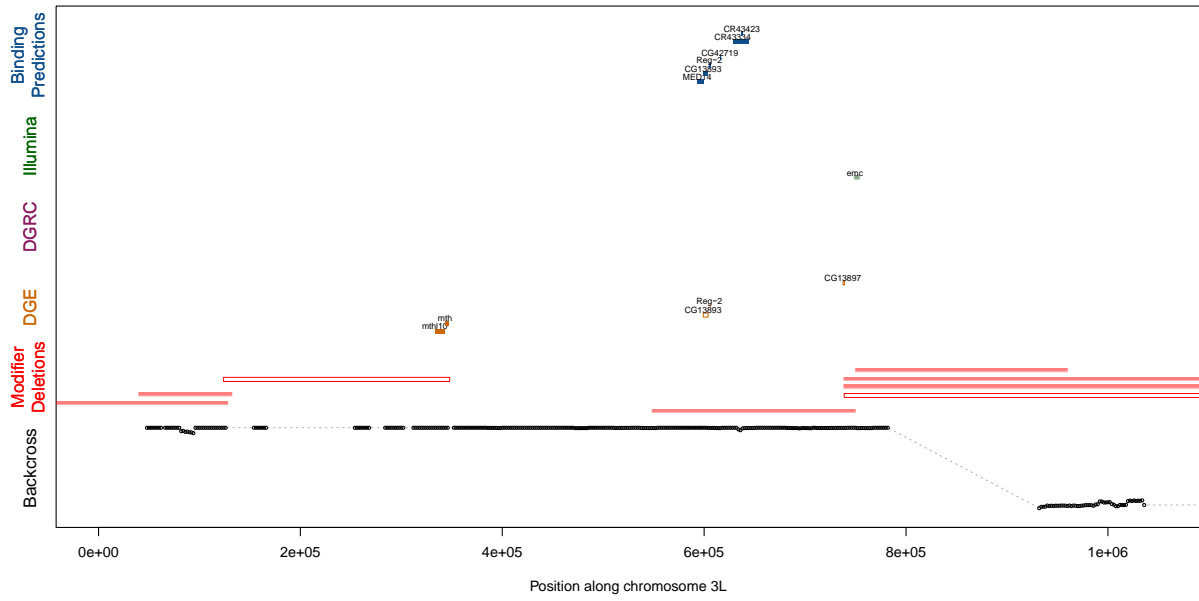
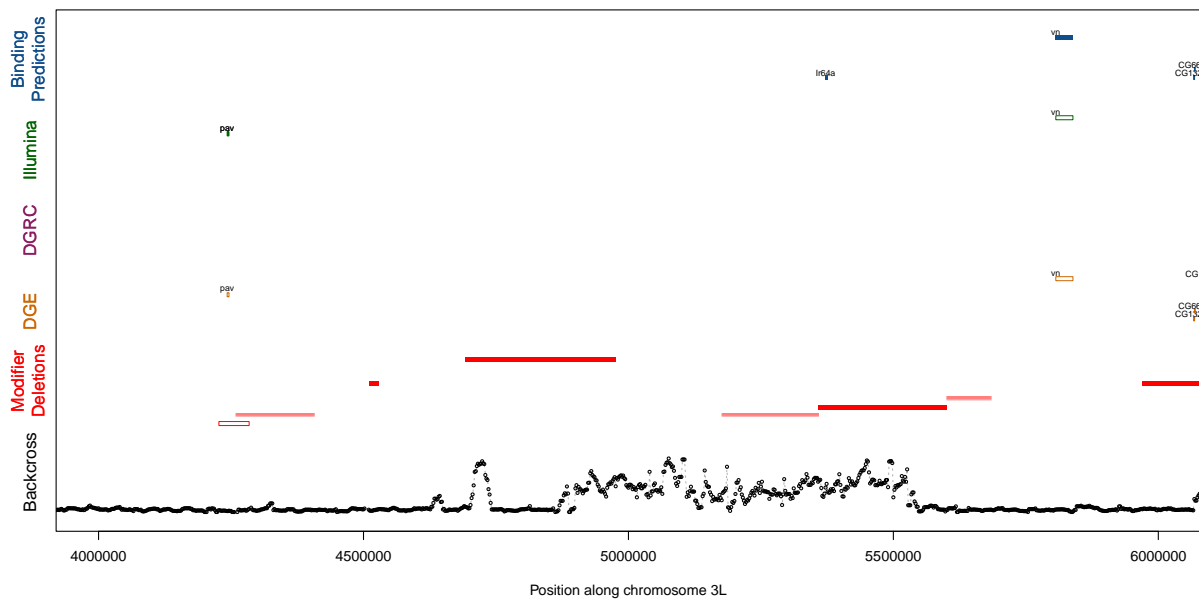
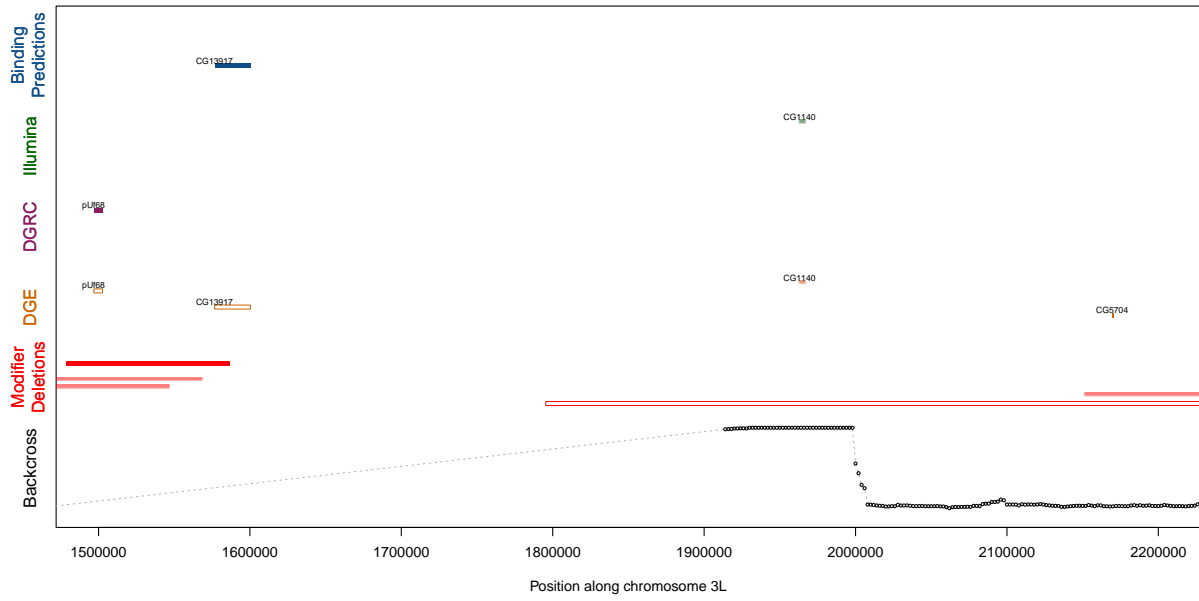


Figure S4 Schematic illustrating tests to distinguish between second- and higher-order epistasis. Red indicates the ORE genetic background; blue indicates the SAM genetic background; grey indicates the genetic background of the deletion strain; white indicates a chromosomal deletion; and the light blue bar on the X chromosome indicates the sd^{E3} allele and the genetic background in which it was originally generated. The genotypes and phenotypes on the left illustrate that this particular deletion enhances the sd^{E3} phenotype in an ORE background, i.e., results in even smaller wings. On the right, however, this deletion suppresses the sd^{E3} phenotype in a short-wing introgression background (i.e., results in larger wings). However, because the short-wing and ORE flies both carry the same genetic background (ORE) opposite the deletion, this background dependence must be due to other loci elsewhere in the genome (in this case, illustrated by the SAM alleles elsewhere in the genome), indicating higher-order epistasis.









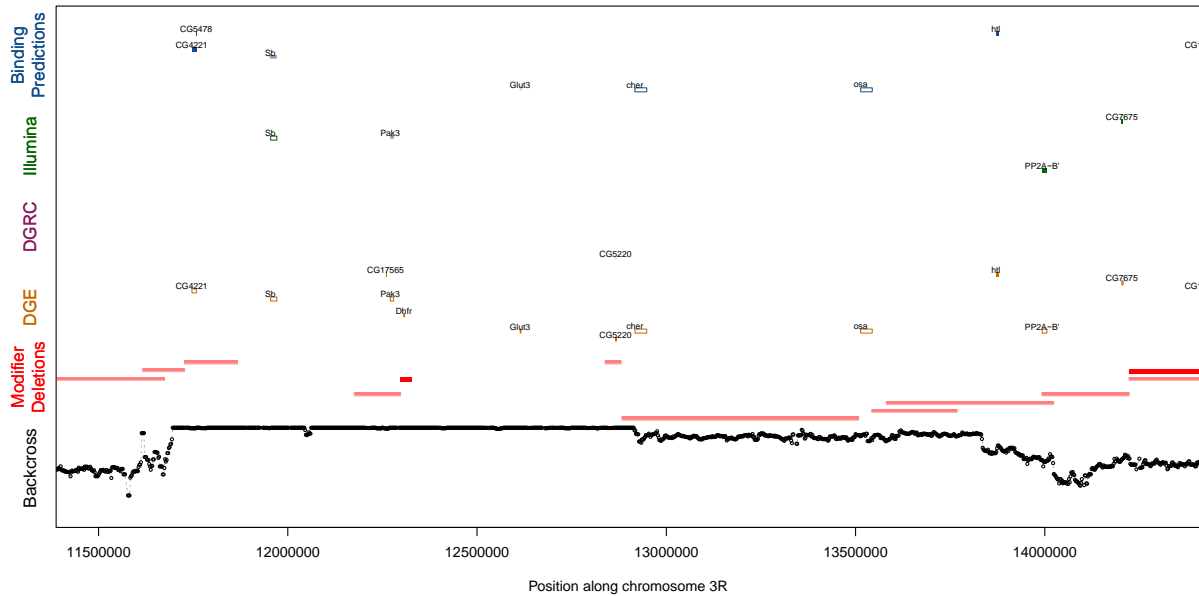
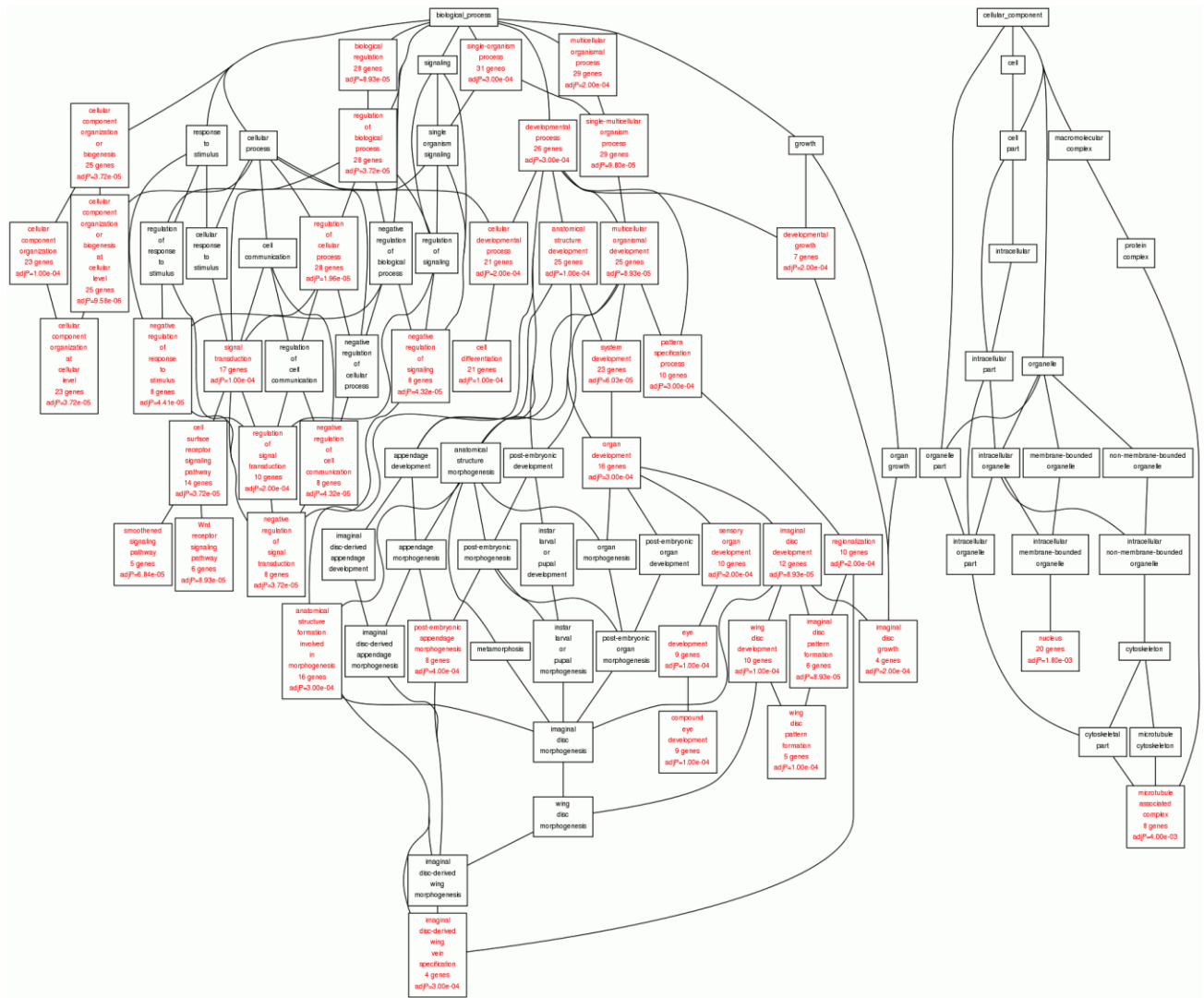


Figure S5 Integrated plots showing results of independent genomic datasets used to investigate the genetic basis of background dependence of the sd^{E3} phenotype. Backcross: Average frequency of the ORE (short-wing) allele across four short-wing introgression lines. Modifier deletions: open bars represent deletions with a significant main effect on the sd^{E3} phenotype; light shaded bars represent deletions with a significant background-dependent effect on the sd^{E3} phenotype; and dark shaded bars represent deletions in which both the main and interaction effects are significant. DGE: open bars represent genes whose transcript counts are influenced by sd genotype; light shaded bars represent genes showing evidence of genotype-dependent allelic imbalance; and dark shaded bars represent genes showing evidence of both an overall effect of sd genotype and genotype-dependent allelic imbalance. DGRC and Illumina: open bars represent genes showing evidence of an effect of sd genotype on expression; light shaded bars represent genes showing evidence of a genotype-by-background interaction effect; and dark shaded bars represent genes showing evidence of both the main and interaction effects. Binding predictions: open bars represent genes predicted to be overall SD binding targets (in at least one of the two genetic backgrounds); light shaded bars represent genes predicted to show differential affinity for SD between the two backgrounds; and dark shaded bars represent genes showing evidence of both overall SD binding and differential affinity between backgrounds. Only genes showing evidence of at least four significant effects across all datasets are shown.



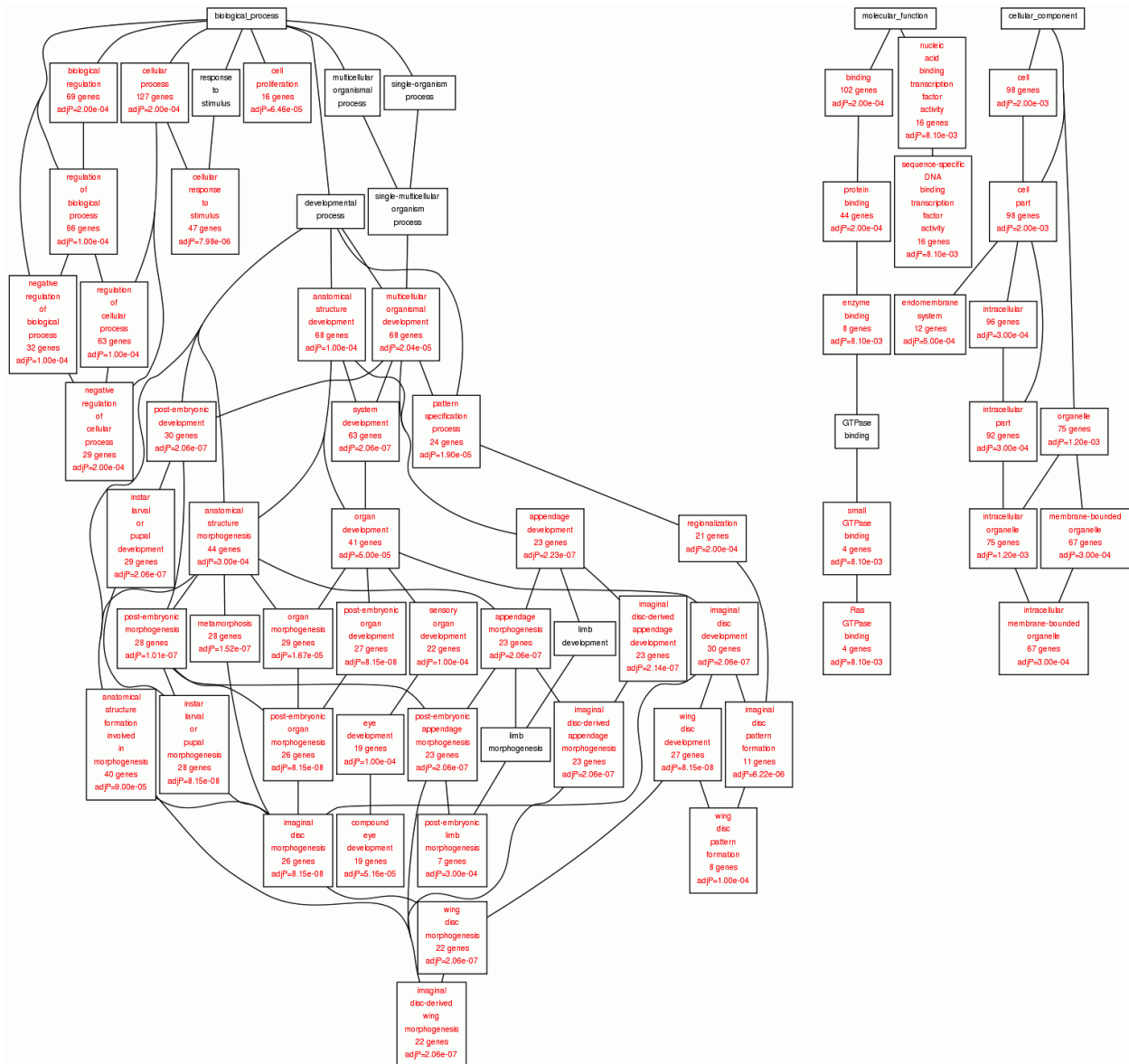


Figure S6 Top scoring GO hits for candidate genes identified by integrated analysis using (A) significant *sd* genotype effects for expression datasets and (B) significant genotype-by-background interaction effects for expression datasets.

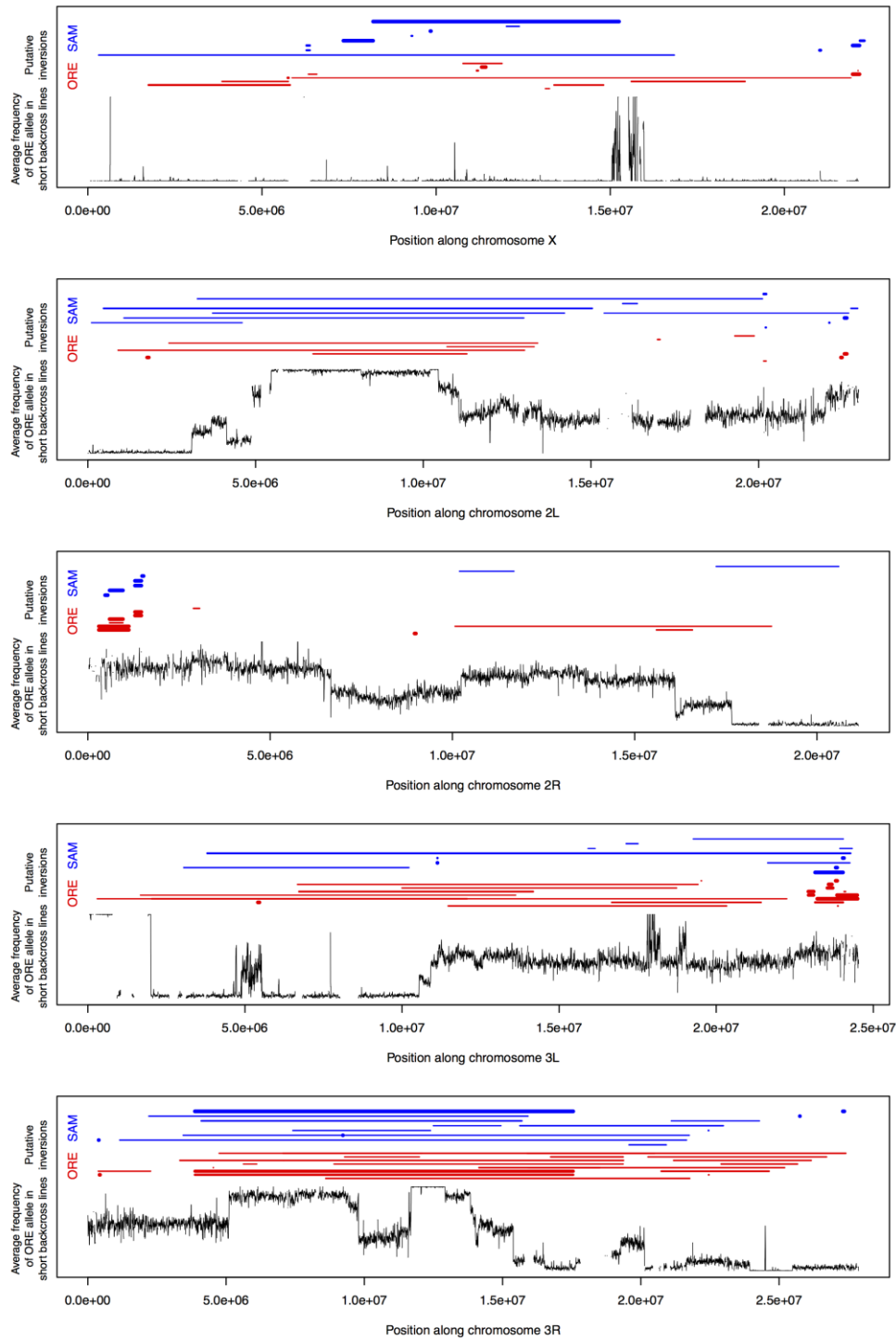


Figure S7 Weak evidence for putative inversions based on paired-end reads mapping to discordant locations in the genome. Thicker bars indicate stronger evidence. Inversions are relative to the *D. melanogaster* reference genome. We searched for inversions using BreakDancer v1.1 (Chen et al. 2009).

References

- Bailey TL, Bodén M, Buske FA, Frith M, Grant CE, Clementi L, Ren J, Li WW, Nole WS: **MEME SUITE: tools for motif discovery and searching.** *Nucleic Acids Res* 2009, **37**:W202-W208.
- Chen K, Wallis JW, McLellan MD, Larson DE, Kalicki JM, Pohl CS, McGrath SD, Wendl MC, Zhang Q, Locke DP, Shi X, Fulton RS, Ley TJ, Wilson RK, Ding L, Mardis ER: **BreakDancer: an algorithm for high-resolution mapping of genomic structural variation.** *Nat Methods* 2009, **6**:677-681.
- Guss KA, Nelson CE, Hudson A, Kraus ME, Carrol SB: **Control of a genetic regulatory network by a selector gene.** *Science* 2001, **292**: 1164-1167.
- Halder G, Carrol SB: **Binding of the Vestigial co-factor switches the DNA-target selectivity of the Scalloped selector protein.** *Development* 2001, **128**: 3295-3305.