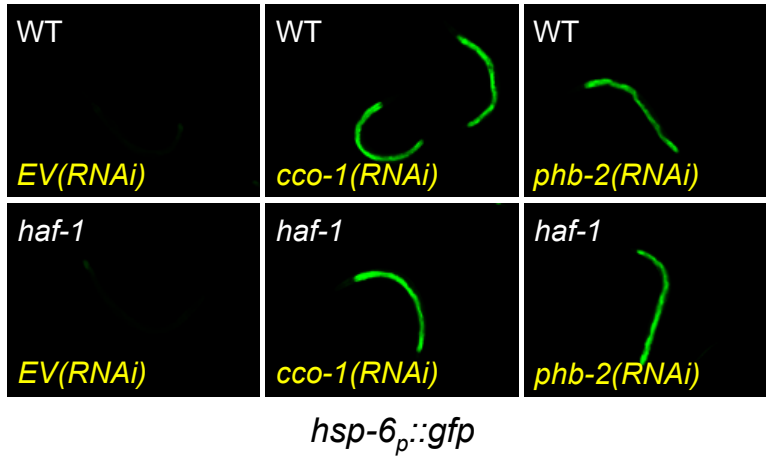
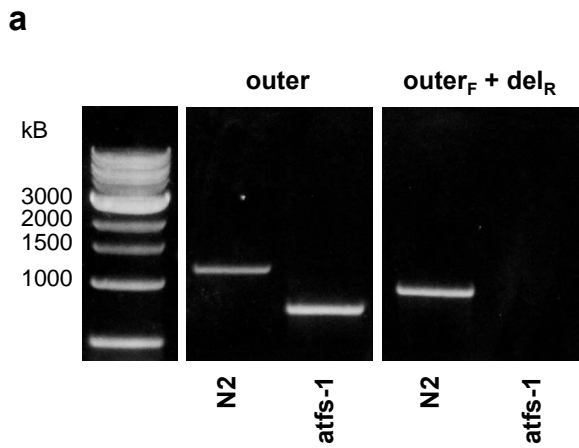


**a**

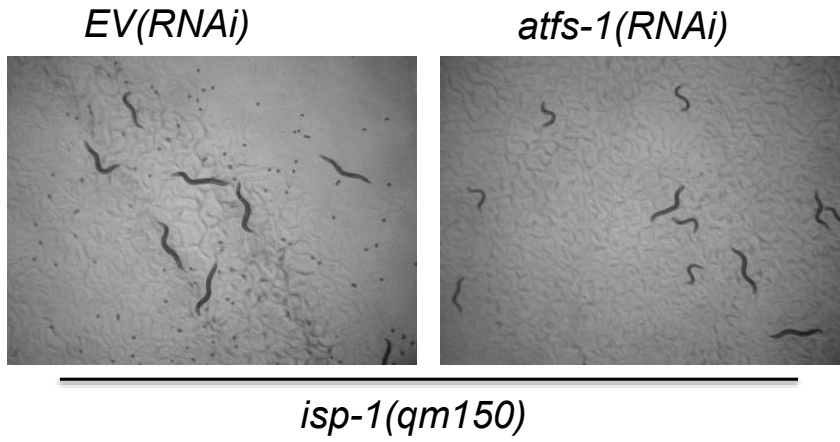


**Supplementary Figure 1. HAF-1 is not required for UPR<sup>mt</sup> induction caused by *cco-1* and *phb-2* RNAi.** *hsp-6<sub>p</sub>::gfp* induction was measured 3 days from hatch at 20°C in wild-type and *haf-1(ok705)* animals.



**Supplementary Figure 2. Genotyping of *atfs-1(tm4525)* mutant.** Outer primers, which bind outside the deletion, give a WT band of ~1200 bp and a *tm4525* band of ~800. The del<sub>R</sub> primer is inside the *tm4525* deletion. The outer<sub>F</sub> and del<sub>R</sub> give a WT band of ~1000 bp and no *tm4525* band.

**a**



**Supplementary Figure 3. The developmental rate of *isp-1(qm150)* is delayed by *atfs-1(RNAi)*.** (A) *isp-1(qm150)* were grown from egg on *EV(RNAi)* and *atfs-1(RNAi)* for 6 days.

**Supplementary Table 1. Validated RNAi Inducers of the UPR<sup>mt</sup>.**

Gene Name	NCBI KOG Annotation	PPOD (Human   Yeast Orthologue)	Mito. Targeted	Extended Lifespan	Reported UPR <sup>mt</sup> inducer
<i>E04A4.5</i>	Mitochondrial import inner membrane translocase, subunit TIM17	TIM17A/TIM17 B   TIM17	A, B, D		
<i>F15D3.7</i>	Mitochondrial import inner membrane translocase, subunit TIM23	TIMM23B   TIM23	A, B, D		G, H
<i>T09B4.9</i>	Mitochondrial import inner membrane translocase, subunit TIM44	TIM44   TIM44	A, B, C, D		F
<i>W02F12.5</i>	Dihydrolipoamide succinyltransferase (2-oxoglutarate dehydrogenase, E2 subunit)	DLTS   KGD2	B, C, D		H
<i>dnj-21</i>	Molecular chaperone (DnaJ superfamily)	TIM14;DNAJC1 5; DNAJC19   TIM14/PAM18	A, B, D		F
<i>lpd-9</i>			C, D		
<i>phb-2</i>		PHB2   PHB2	A, B, D		F
<i>C04C3.3</i>	Pyruvate dehydrogenase E1, beta subunit	PDHB   PDB1	A, B, C, D		
<i>wah-1</i>	Programmed cell death 8 (apoptosis-inducing factor)	AIFM1	A, B, C, D		
<i>F45G2.8</i>	Uncharacterized conserved protein	PAM16   PAM16	A, B, D		
<i>Y54G9A.7</i>			C, D		
<i>ech-6</i>	Enoyl-CoA hydratase	ECHS1	A, B, C, D		
<i>Y110A7A.19</i>	Uncharacterized conserved protein	PTCD3	B, C, D		
<i>sco-1</i>	Putative cytochrome C oxidase assembly protein	SCO1   SCO1; SCO2	B, D	E	
<i>cchl-1</i>	Holocytochrome c synthase/heme-lyase	CCHL   CYC3	A, B, D	WBRNAi 00063981	I
<i>ril-1</i>			D	WBRNAi 00063960	I
<i>tomm-22</i>	Translocase of outer mitochondrial membrane complex, subunit TOM22	TOM22	B, D		
<i>Y22D7AL.10</i>	Mitochondrial chaperonin	HSPE1   HSP10	B, C, D		
<i>Y17G9B.5</i>	Intermediate in Toll signal transduction pathway (ECSIT)	ECSIT	B, C, D		
<i>his-5</i>	Histone H4	Histone H4   HHF2	C		
<i>letm-1</i>	Ca <sup>2+</sup> -binding transmembrane protein LETM1/MRS7	LETM1   MDM38	B, C		
<i>C08F8.2</i>	Mitochondrial RNA helicase SUV3, DEAD-box superfamily	SUPV3L1   SUV3	A, B, C		

<i>F02A9.4</i>	3-Methylcrotonyl-CoA carboxylase	MCCC2	A, B, C	
<i>hsp-60</i>	Mitochondrial chaperonin, Cpn60/Hsp60p	HSPD1   HSP60	A, B, C	F, H
<i>Y38E10A.24</i>				
<i>tkt-1</i>	Transketolase	TKTL1; TKTL2   TKL1; TKL2		WBRNAi 00078549 H
<i>ant-1.1</i>	Mitochondrial ADP/ATP carrier proteins	ANT1; ANT2; ANT3   AAC1; AAC2; AAC3	A, B	WBRNAi 00078570
<i>drp-1</i>	Vacuolar sorting protein VPS1, dynamin, and related proteins	DRP1   DNM1	A, B	
<i>Y52B11C.1</i>				
<i>F15D3.6</i>	Predicted member of the intramitochondrial sorting protein family	SLMO2   UPS2; UPS3	B	
<i>Y24D9A.8</i>	Transaldolase	TALDO1   TAL1		
<i>W02B12.9</i>	Mitochondrial carrier protein MRS3/4	MFRN   MRS3; MRS4	A, B	
<i>AC8.6</i>				
<i>F21C3.10</i>				

RNAi knockdown of the following 34 genes significantly induced the *hsp-6p::gfp* reporter. The eukaryotic orthologous groups (KOGs) annotation and PPOD derived human and yeast orthologues for each gene are shown. Annotation for mitochondrial targeting is based off of Go Term “Mitochondrion” annotation (A), orthology to a known mitochondrial genes by PPOD (B), MitoProt II probability estimation >0.8 (C), or presence in a published *C. elegans* mitochondrial proteome data set by Li, J., et al., Proteomics, 2009 <sup>1</sup> (D). Reported effects on longevity are shown as WBRNAi experiments or Chen, D., et al., Aging Cell, 2007 <sup>2</sup> (E). Reported effects on the UPR<sup>mt</sup> are based on the work of Yoneda, T., et al., Journal of Cell Science, 2004 <sup>3</sup> (F), and Nargund, A., et al., Science, 2012 <sup>4</sup> (G). Melo, J., et al., Cell, 2012 <sup>5</sup> (H). Shore, D., et al., PLoS Genet, 2012 <sup>6</sup> (I).

**Supplementary Table 2. Non-Validated RNAi Inducers of the UPR<sup>mt</sup>.**

Gene Name	NCBI KOG Annotation	Putative Mito. Function	Mito. Targeted	Extended Lifespan	Reported UPR <sup>mt</sup> inducer
<i>F44G4.2</i>	Unnamed protein	Complex I	✓		
<i>B0491.5</i>		Complex I	✓	C	
<i>nuo-1</i>	NADH:ubiquinone oxidoreductase, NDUFV1/51kDa subunit	Complex I	✓	WBRNAi 00066177	G
<i>nduf-2.2</i>	NADH:ubiquinone oxidoreductase, NDUFS2/49 kDa subunit	Complex I	✓	WBRNAi 00078565	
<i>ZK809.3</i>	NADH:ubiquinone oxidoreductase, NDUFB6/B17 subunit	Complex I	✓	WBRNAi 00078577	
<i>F31D4.9</i>		Complex I			
<i>gas-1</i>	NADH:ubiquinone oxidoreductase, NDUFS2/49 kDa subunit	Complex I	✓		
<i>nduf-7</i>	NADH-ubiquinone oxidoreductase, NUF57/PSST/20 kDa subunit	Complex I	✓		
<i>C16A3.5</i>	NADH:ubiquinone oxidoreductase, NDUFB9/B22 subunit	Complex I	✓		
<i>nduf-5</i>	NADH:ubiquinone oxidoreductase, NDUFS5/15kDa	Complex I	✓		
<i>D2030.4</i>	NADH:ubiquinone oxidoreductase, NDUFB7/B18 subunit	Complex I	✓	WBRNAi 00065275	D, E, G
<i>F59C6.5</i>	NADH-ubiquinone oxidoreductase, subunit NDUFB10/PDSW	Complex I	✓	WBRNAi 00066189	D, G
<i>T20H4.5</i>	NADH:ubiquinone oxidoreductase, NDUFS8/23 kDa subunit	Complex I	✓	WBRNAi 00065279	E, G
<i>nduf-6</i>	NADH:ubiquinone oxidoreductase, NDUFS6/13 kDa subunit	Complex I	✓		D
<i>Y51H1A.3</i>	NADH:ubiquinone oxidoreductase, NDUFB8/ASHI subunit	Complex I			
<i>F42G8.10</i>	Uncharacterized conserved protein	Complex I			
<i>nuo-4</i>	NADH:ubiquinone oxidoreductase, NDUFA10/42kDa subunit	Complex I		WBRNAi 00065278	E, G
<i>ucr-1</i>	Mitochondrial processing peptidase, beta subunit, and related enzymes (insulinase superfamily)	Complex III	✓	WBRNAi 00078564	

<i>T02H6.11</i>	Ubiquinol cytochrome c reductase, subunit QCR7	Complex III	✓	B	
<i>F45H10.2</i>	Ubiquinol cytochrome c reductase, subunit QCR8	Complex III	✓		
<i>R07E4.3</i>	Ubiquinol cytochrome c reductase, subunit QCR8	Complex III	✓		
<i>F29B9.11</i>		Complex IV	✓		
<i>cco-2</i>	Cytochrome c oxidase, subunit Va/COX6	Complex IV	✓	WBRNAi 00063991	G
<i>F29C4.2</i>	Unnamed protein	Complex IV	✓	WBRNAi 00078571	
<i>cco-1</i>	Cytochrome c oxidase, subunit Vb/COX4	Complex IV	✓	WBRNAi 00063967	D, E, F
<i>F26E4.6</i>	Cytochrome c oxidase, subunit VIIc/COX8	Complex IV	✓	WBRNAi 00065276	D, E, G
<i>atp-2</i>	F0F1-type ATP synthase, beta subunit	Complex V	✓	WBRNAi 00066144	F, G
<i>atp-4</i>	Mitochondrial F1F0-ATP synthase, subunit Cf6 (coupling factor 6)	Complex V	✓	WBRNAi 00063979	G
<i>R53.4</i>	Mitochondrial F1F0-ATP synthase, subunit f	Complex V	✓	C	
<i>F32D1.2</i>	Mitochondrial F1F0-ATP synthase, subunit epsilon/ATP15	Complex V	✓		
<i>F58F12.1</i>	Mitochondrial F1F0-ATP synthase, subunit delta/ATP16	Complex V	✓		
H28O16.1	F0F1-type ATP synthase, alpha subunit	Complex V		WBRNAi 00066165	D, F, G
<i>atp-3</i>	Mitochondrial F1F0-ATP synthase, subunit OSCP/ATP5	Complex V		WBRNAi 00063970	G
<i>R04F11.2</i>	Mitochondrial F1F0-ATP synthase, subunit e	Complex V	✓		
<i>asg-2</i>	Mitochondrial F1F0-ATP synthase, subunit g/ATP20	Complex V	✓	WBRNAi 00065274	E, G
<i>asg-1</i>	Mitochondrial F1F0-ATP synthase, subunit g/ATP20	Complex V	✓		D
T26E3.7	F0F1-type ATP synthase, alpha subunit	Complex V			
Y82E9BR.3	Mitochondrial F1F0-ATP synthase, subunit c/ATP9/proteolipid	Complex V			
<i>cyc-2.1</i>	Cytochrome c	Cytochrome C	✓	WBRNAi 00078573	
<i>F33D4.5</i>	50S ribosomal protein L1	Mitochondrial Translation	✓	WBRNAi 00078562	A
<i>C30C11.1</i>	Mitochondrial ribosomal protein L32	Mitochondrial Translation	✓		
<i>F09G8.3</i>	Mitochondrial/chloroplast ribosomal protein S9	Mitochondrial Translation	✓	WBRNAi 00078568	
<i>F59A3.3</i>	Mitochondrial/chloroplast ribosomal protein L24	Mitochondrial Translation	✓	C	
<i>K01C8.6</i>	Mitochondrial ribosomal protein L10	Mitochondrial Translation	✓	C	

<i>tag-264</i>	Mitochondrial 28S ribosomal protein S30	Mitochondrial Translation	✓	C	D, A
<i>tufm-1</i>	Mitochondrial translation elongation factor Tu	Mitochondrial Translation	✓	WBRNAi 00078595	
<i>T04A8.11</i>		Mitochondrial Translation	✓		
<i>dap-3</i>	Mitochondrial ribosome small subunit component, mediator of apoptosis DAP3	Mitochondrial Translation	✓		
F45E12.5	Mitochondrial ribosomal protein L14	Mitochondrial Translation			
<i>mrpl-37</i>		Mitochondrial Translation	✓	A	A
<i>mrps-5</i>	Ribosomal protein S5	Mitochondrial Translation	✓	A	A
<i>F56B3.8</i>	Mitochondrial/chloroplast ribosomal protein L2	Mitochondrial Translation	✓	A	A
<i>sars-2</i>	Seryl-tRNA synthetase	Mitochondrial Translation	✓		
<i>T13H5.5</i>	Mitochondrial ribosomal protein S18b	Mitochondrial Translation	✓		
<i>T23B12.2</i>	Mitochondrial/chloroplast ribosomal protein L4	Mitochondrial Translation	✓		
<i>T23B12.3</i>	Mitochondrial/chloroplast ribosomal protein S2	Mitochondrial Translation	✓		
<i>W04B5.4</i>	Mitochondrial ribosomal protein L30	Mitochondrial Translation	✓		
<i>Y39A1A.6</i>	Mitochondrial/chloroplast ribosomal protein L22	Mitochondrial Translation	✓		
<i>Y39B6A.39</i>	Mitochondrial ribosomal protein S28	Mitochondrial Translation	✓		
<i>Y48C3A.10</i>	Mitochondrial/chloroplast ribosomal protein L20	Mitochondrial Translation	✓		
<i>Y34D9A.1</i>	Phosphatidylethanolamine binding protein	Mitochondrial Translation	✓		

This table represents 61 positive inducers of *hsp-6p::gfp* identified in our screen that target components of the electron transport chain or mitochondrial translation machinery.

Since similar hits have been reported to induce the UPR<sup>mt</sup>, we focused our validation elsewhere, but this list is expected to represent genuine inducers of the UPR<sup>mt</sup> and in some cases, have been validated by other groups. Putative Mitochondrial Function is based off KOG annotation, PPOD orthology analysis, or blastP homology analysis.

Mitochondrial targeting is based of inclusion in the *C. elegans* mitochondrial proteome data set by Li, J., et al., Proteomics, 2009<sup>1</sup>. Reported effects on longevity or UPR<sup>mt</sup>



induction are shown as WBRNAi experiments or Houtkooper, R., et al., Nature, 2013<sup>7</sup> (A), Lee, S.S., et al., Nature Gen, 2003<sup>8</sup> (B), Chen, D., et al., Aging Cell, 2007<sup>2</sup> (C), Yoneda, T., et al., Journal of Cell Science, 2004<sup>3</sup> (D), Hamilton, B., et al., Genes Dev, 2005<sup>9</sup> (E), Melo, J., et al., Cell, 2012<sup>5</sup> (F), and Shore, D., et al., PLoS Genet, 2012<sup>6</sup> (G).

**Supplementary Table 3. Effects of 34 Validated RNAi inducers of the UPR<sup>mt</sup> on other stress response reporters.**

RNAi/cond.	<i>hsp-6<sub>p</sub>::gfp</i>	<i>hsp-60<sub>p</sub>::gfp</i>	<i>hsp-4<sub>p</sub>::gfp</i>	<i>hsp-16.2<sub>p</sub>::gfp</i>
<i>EV</i>	1 ± 0.02	1 ± 0.02	1 ± 0.04	1 ± 0.03
<i>phb-2</i>	12.29 ± 0.52	1.88 ± 0.07	1.23 ± 0.05	0.82 ± 0.07
<i>EV + Heat Shock</i>	#N/A	#N/A	#N/A	7.13 ± 0.46
<i>EV + Tunicamycin</i>	#N/A	#N/A	7.15 ± 0.63	#N/A
<i>Y17G9B.5</i>	1.4 ± 0.11	1.07 ± 0.05	0.92 ± 0.05	0.84 ± 0.04
<i>his-5</i>	2.27 ± 0.14	0.96 ± 0.03	1.18 ± 0.07	0.09 ± 0.01
<i>tkt-1</i>	2.42 ± 0.28	0.76 ± 0.02	0.98 ± 0.05	0.57 ± 0.02
<i>ant-1.1</i>	2.46 ± 0.33	0.86 ± 0.03	0.72 ± 0.04	0.83 ± 0.03
<i>F21C3.10</i>	2.56 ± 0.2	1.02 ± 0.05	1.09 ± 0.06	0.94 ± 0.06
<i>drp-1</i>	2.96 ± 0.39	0.92 ± 0.04	0.78 ± 0.03	1.02 ± 0.06
<i>C04C3.3</i>	3.19 ± 0.35	1.07 ± 0.05	1.19 ± 0.06	0.87 ± 0.04
<i>Y24D9A.8</i>	3.49 ± 0.25	0.75 ± 0.04	0.91 ± 0.08	0.55 ± 0.02
<i>W02B12.9</i>	3.58 ± 0.34	1.01 ± 0.03	1.16 ± 0.12	0.81 ± 0.05
<i>sco-1</i>	3.87 ± 0.59	1.16 ± 0.04	0.84 ± 0.09	0.92 ± 0.05
<i>C08F8.2</i>	5.08 ± 0.72	1.06 ± 0.05	0.9 ± 0.03	0.87 ± 0.05
<i>Y52B11C.1</i>	5.35 ± 0.39	1.35 ± 0.05	0.95 ± 0.06	1.1 ± 0.05
<i>F15D3.6</i>	6.03 ± 0.35	1.36 ± 0.07	0.82 ± 0.03	0.86 ± 0.05
<i>cchl-1</i>	8.2 ± 1.04	1.42 ± 0.04	0.84 ± 0.04	1.01 ± 0.04
<i>ech-6</i>	8.97 ± 1.37	1.45 ± 0.06	0.93 ± 0.04	0.93 ± 0.04
<i>hsp-60</i>	11.14 ± 0.78	1.01 ± 0.03	1.09 ± 0.04	0.59 ± 0.03
<i>F02A9.4</i>	11.17 ± 0.96	1.33 ± 0.06	0.87 ± 0.07	0.79 ± 0.03
<i>AC8.6</i>	11.62 ± 1	1.28 ± 0.06	0.76 ± 0.04	1 ± 0.04
<i>tomm-22</i>	11.93 ± 0.81	1.14 ± 0.05	1.19 ± 0.11	0.65 ± 0.04
<i>T09B4.9</i>	12.04 ± 0.73	1.65 ± 0.14	1.02 ± 0.04	0.7 ± 0.03
<i>Y38E10A.24</i>	12.16 ± 0.67	1.07 ± 0.05	1.44 ± 0.08	0.72 ± 0.02
<i>Y22D7AL.10</i>	12.85 ± 0.71	0.93 ± 0.03	0.76 ± 0.06	0.84 ± 0.04
<i>wah-1</i>	13.17 ± 1.99	1.36 ± 0.04	1.14 ± 0.04	0.93 ± 0.06
<i>E04A4.5</i>	13.33 ± 1.12	1.4 ± 0.11	0.97 ± 0.09	0.64 ± 0.02
<i>Y54G9A.7</i>	14.04 ± 1.31	1.28 ± 0.05	1.14 ± 0.08	0.92 ± 0.06
<i>Y110A7A.19</i>	14.41 ± 1.3	1.45 ± 0.08	0.84 ± 0.05	0.77 ± 0.03
<i>ril-1</i>	14.71 ± 2.81	1.05 ± 0.06	0.59 ± 0.04	0.35 ± 0.02
<i>letm-1</i>	16.06 ± 2.44	1.22 ± 0.05	0.83 ± 0.03	0.73 ± 0.03
<i>phb-2</i>	16.11 ± 2.36	1.99 ± 0.11	1.2 ± 0.06	0.55 ± 0.03
<i>lpd-9</i>	16.37 ± 1.45	1.46 ± 0.07	1.15 ± 0.08	0.97 ± 0.05
<i>W02F12.5</i>	16.47 ± 1.73	1.15 ± 0.06	1.1 ± 0.07	0.73 ± 0.03
<i>F45G2.8</i>	16.95 ± 1.4	1.37 ± 0.05	1.07 ± 0.09	0.81 ± 0.04
<i>F15D3.7</i>	17.52 ± 1.13	1.67 ± 0.08	0.93 ± 0.06	0.61 ± 0.04
<i>dnj-21</i>	22.12 ± 2.48	1.93 ± 0.1	1.19 ± 0.08	0.87 ± 0.04

RNAi knockdown of the shown genes significantly induced expression of the *hsp-6<sub>p</sub>::gfp* reporter. The effect on mean GFP fluorescence from each RNAi clone is shown relative to empty vector (EV) treated animals (± SEM, N=3). Worms were grown at 20°C and

imaged 3 days from hatching. Gene knockdowns that significantly increased (student's t-test with Bonferroni correction) each reporter are highlighted in gray.

**Supplementary Table 4. Primers used for qRT-PCR.** qRT-PCR was used to measure expression of mitoUPR targets.

<b>Primer</b>	<b>Sequence</b>
hsp-6 <sub>F</sub>	TCGTGAACGTTTCAGCCAGA
hsp-6 <sub>R</sub>	CTCAGCGGCATTCTTTTCGG
hsp-60 <sub>F</sub>	GGGGAAGCCCAAAGATCACA
hsp-60 <sub>R</sub>	TCCAGCCTCCTCATTAGCCT
timm-23 <sub>F</sub>	CTCCGATCGATCTCAGTGCC
timm-23 <sub>R</sub>	ATAGGGTGTCATTTGCCGGG

## SUPPLEMENTARY REFERENCES

- 1) Li, J., Cai, T., Wu, P., Cui, Z., Chen, X., Hou, J., Xie, Z., Xue, P., Shi, L., Liu, P., *et al.* (2009). Proteomic analysis of mitochondria from *Caenorhabditis elegans*. *Proteomics* 9, 4539-4553.
- 2) Chen, D., Pan, K.Z., Palter, J.E., and Kapahi, P. (2007). Longevity determined by developmental arrest genes in *Caenorhabditis elegans*. *Aging Cell* 6, 525-533.
- 3) Yoneda, T. et al. Compartment-specific perturbation of protein handling activates genes encoding mitochondrial chaperones. *J Cell Sci* 117, 4055-4066, doi:10.1242/jcs.01275 (2004).
- 4) Nargund, A. M., Pellegrino, M. W., Fiorese, C. J., Baker, B. M. & Haynes, C. M. Mitochondrial import efficiency of ATFS-1 regulates mitochondrial UPR activation. *Science* 337, 587-590, doi:10.1126/science.1223560 (2012).
- 5) Melo, J.A., and Ruvkun, G. Inactivation of Conserved *C. elegans* Genes Engages Pathogen- and Xenobiotic-Associated Defenses. *Cell* 149, 452–466 (2012).
- 6) Shore, D.E., Carr, C.E., and Ruvkun, G. Induction of Cytoprotective Pathways Is Central to the Extension of Lifespan Conferred by Multiple Longevity Pathways. *PLoS Genet* 8, e1002792 (2012).
- 7) Houtkooper, R. H. et al. Mitonuclear protein imbalance as a conserved longevity mechanism. *Nature* 497, 451-457, doi:10.1038/nature12188 (2013).
- 8) Lee, S. S. et al. A systematic RNAi screen identifies a critical role for mitochondria in *C. elegans* longevity. *Nat Genet* 33, 40-48, doi:10.1038/ng1056 (2003).
- 9) Hamilton, B. et al. A systematic RNAi screen for longevity genes in *C. elegans*. *Genes Dev* 19, 1544-1555 (2005).