

SUPPLEMENTARY DATA

Assessing the subcellular distribution of oncogenic phosphoinositide 3-kinase using microinjection into live cells

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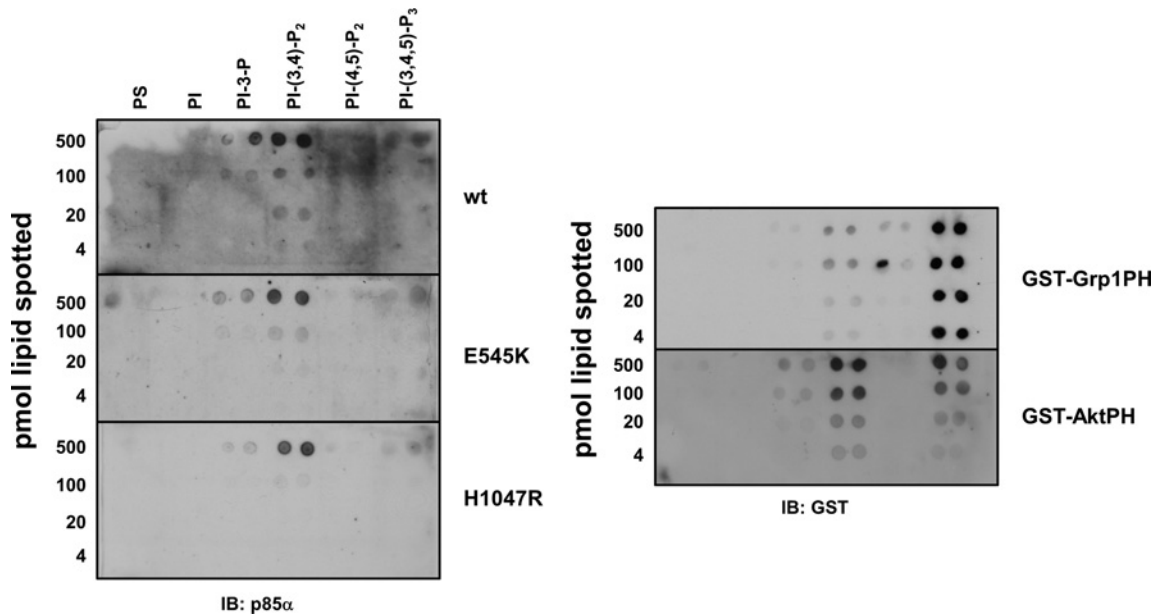


Figure S1 A longer exposure (30 min) of the lipid overlay assay does not provide evidence for increased binding of mutant PI3K α to phosphoinositides compared to wild-type PI3K α

The nitrocellulose membrane from Figure 2(B) (spotted with 4–500 pmol purified PS or a phosphoinositide) that had been incubated with 10 nM wild-type or mutant (E545K or H1047R) PI3K α (upper panel) or 10 nM GST-Grp1PH or GST-Akt PH (lower panel) followed by antibodies that recognize p85 α or GST was exposed to film for 30 min.

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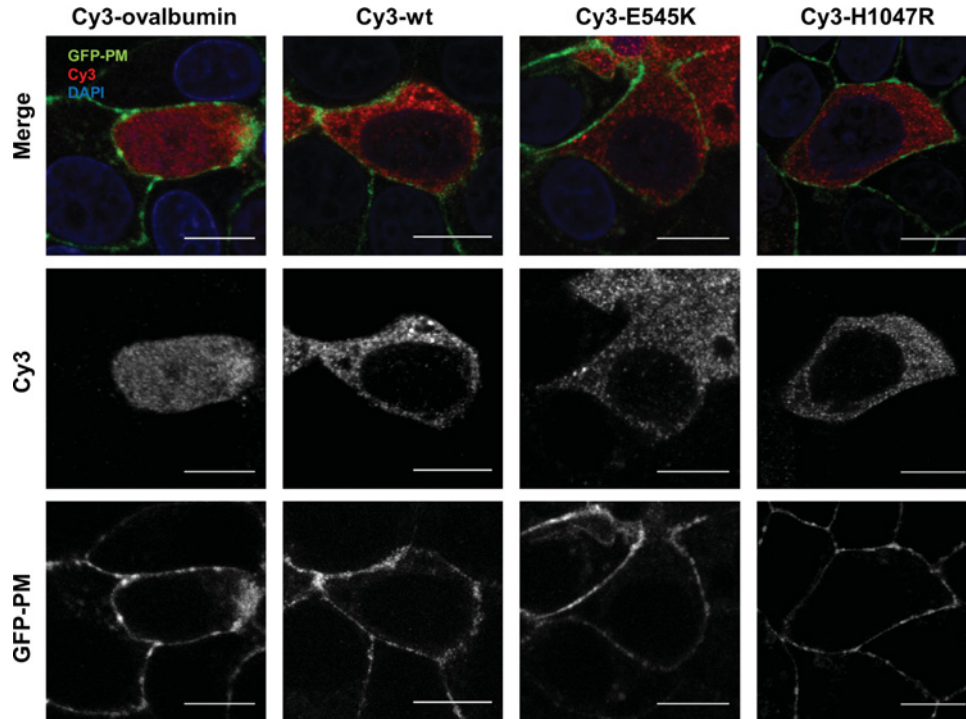


Figure S2 Single-channel images from Figure 2(C)

0.5–2 μ M Cy3-ovalbumin, Cy3-p110 α EE^{WT}/p85 α , Cy3-p110 α EE^{E545K}/p85 α or Cy3-p110 α EE^{H1047R}/p85 α was microinjected into adherent MDCK cells expressing membrane-localized GFP in DME containing 10% (v/v) FBS. Cells were allowed to recover for 2 h post-microinjection, then fixed and washed in PBS and stained with DAPI to detect nuclear DNA. Fluorescence associated with Cy3 (red), GFP (green) or DAPI (blue) was visualized using confocal microscopy (upper panel). Single channel images are shown for Cy3 (middle panel) and the GFP-PM (lower panel). Scale bar = 10 μ m.

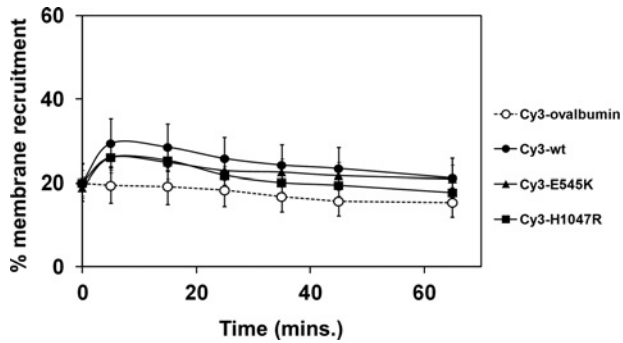


Figure S3 Data from Figure 3 represented as the means \pm S.E.M. of the percentage of Cy3-PI3K α co-incident with the plasma membrane

0.5–2 μ M Cy3-ovalbumin (○), Cy3-p110 α EE^{WT}/p85 α (●), Cy3-p110 α EE^{E545K}/p85 α (▲) or Cy3-p110 α EE^{H1047R}/p85 α (■) was microinjected into adherent MDCK cells expressing membrane-localized GFP in DME containing 10% FBS. Cells were allowed to recover for 2 h post-microinjection, then starved for 4 h and imaged prior to stimulation and at 5, 15, 25, 35, 45 and 65 min post-stimulation with 10% FBS, 50 ng/ml EGF and 5 μ g/ml insulin. The percentage of Cy3-PI3K α fluorescence that was co-incident with GFP-plasma membrane marker fluorescence was quantified in 25 successive focal planes in individual cells at each time point using Imaris v7.4 software. The means \pm S.E.M. of the percentage of Cy3-PI3K α co-incident with the plasma membrane were calculated from measurements of a minimum of five individual cells from three independent experiments at each time point.

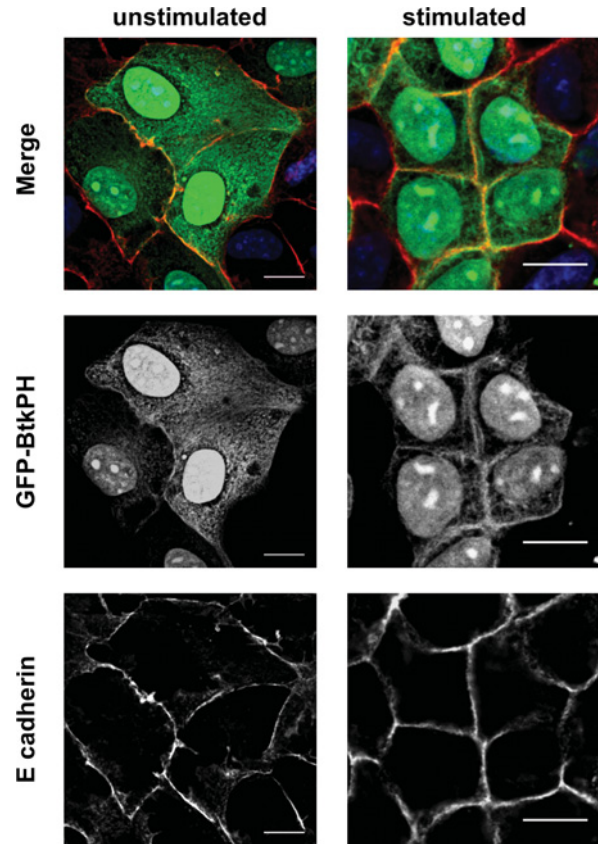


Figure S4 To measure whether of stimulation of MDCK results in activation of endogenous PI3K α resulting in PIP₃ production, cells were transfected with GFP-BtkPH, a PIP₃ reporter [1]

Two days after transfection, cells were starved for 4 h in media containing no FBS at 37 °C then stimulated for 10 min with 10% FBS, 50 ng/ml EGF and 5 μ g/ml insulin to activate RTKs. Cells were fixed and washed in PBS and stained with DAPI to detect nuclear DNA and E cadherin, as a PM marker, followed by an Alexa594-labelled secondary antibody (red). Fluorescence associated with GFP (green), Alexa 594 (red) or DAPI (blue) was visualized using confocal microscopy (upper panel). Single-channel images are shown for GFP-BtkPH (middle panel) and E cadherin/Alexa 594 as a PM marker (lower panel). Scale bar = 10 μ m.

REFERENCE

- 1 Varnai, P, Rother, K. I. and Balla, T. (1999) Phosphatidylinositol 3-kinase-dependent membrane association of the Bruton's tyrosine kinase pleckstrin homology domain visualized in single living cells. *J. Biol. Chem.* **274**, 10983–10989 [CrossRef PubMed](#)

Received 10 December 2013/18 February 2014; accepted 4 March 2014

Published as Immediate Publication 5 March 2014, doi 10.1042/BSR20130133