

Table S1.
Strains, plasmids and oligonucleotides used in this study.

STRAINS

Strains	description and genotype	source
<u>Enteroaggregative <i>E. coli</i></u>		
17-2	WT enteroaggregative <i>Escherichia coli</i>	Arlette Darfeuille-Michaud
17-2 Δ <i>hcp</i>	17-2 deleted of the <i>sci-1 hcp</i> gene	[10]
17-2 Δ <i>sci-1</i>	17-2 deleted of the whole <i>sci-1</i> gene cluster	This study
17-2 Δ <i>tssB1</i>	17-2 deleted of the <i>sci-1 tssB</i> gene	This study
17-2 Δ <i>tssC1</i>	17-2 deleted of the <i>sci-1 tssC</i> gene	This study
17-2 Δ <i>tssBC1</i>	17-2 deleted of the <i>sci-1 tssB</i> and <i>tssC</i> genes	This study
17-2 Δ <i>vgrG1</i>	17-2 deleted of the <i>sci-1 vgrG</i> gene	This study
<u><i>E. coli</i> K12</u>		
DH5 α	F-, Δ (<i>argF-lac</i>)U169, <i>phoA</i> , <i>supE44</i> , Δ (<i>lacZ</i>)M15, <i>relA</i> , <i>endA</i> , <i>thi</i> , <i>hsdR</i>	Laboratory collection
W3110	F-, lambda- IN(<i>rrnD-rrnE</i>)1 <i>rph-1</i>	Laboratory collection

PLASMIDS

pUC12	empty vector, pColE1 origin, <i>Plac</i> , Amp ^R	[22]
pUC-Hcp1-FLAG	<i>sci-1 hcp (hcp1)</i> cloned into pUC12, C-terminal FLAG epitope	[10]
pUC-Hcp2-FLAG	<i>sci-2 hcp (hcp2)</i> cloned into pUC12, C-terminal FLAG epitope	This study
pUC-Hcp1-FLAG N115C	code for Hcp1 harbouring the N115C mutation	This study
pUC-Hcp1-FLAG C38S	code for Hcp1 harbouring the C38S mutation	This study
pUC-Hcp1-FLAG C38S Q24C	code for Hcp1 harbouring the C38S and Q24C mutations	This study
pUC-Hcp1-FLAG C38S G48C	code for Hcp1 harbouring the C38S and G48C mutations	This study
pUC-Hcp1-FLAG C38S A95C	code for Hcp1 harbouring the C38S and A95C mutations	This study
pUC-Hcp1-FLAG C38S G96C	code for Hcp1 harbouring the C38S and G96C mutations	This study
pUC-Hcp1-FLAG C38S N115C	code for Hcp1 harbouring the C38S and N115C mutations	This study
pUC-Hcp1-FLAG C38S S158C	code for Hcp1 harbouring the C38S and S158C mutations	This study
pUC-Hcp1-FLAG C38S Q24C A95C	code for Hcp1 harbouring the C38S, Q24C and A95C mutations	This study
pUC-Hcp1-FLAG C38S G96C S158C	code for Hcp1 harbouring the C38S, G96C and S158C mutations	This study
pUC-Hcp1-FLAG S158W	code for Hcp1 harbouring the S158W mutation	This study
pT18-FLAG	Bacterial Two Hybrid (BACTH) vector	[23]
pT18-Hcp1	<i>hcp1</i> cloned downstream the T18 coding sequence in p-T18-FLAG	This study
pHcp1-T18	<i>hcp1</i> cloned upstream the T18 coding sequence in p-T18-FLAG	This study
pT18-TssB1	<i>tssB1</i> cloned downstream the T18 coding sequence in p-T18-FLAG	This study
pTssB1-T18	<i>tssB1</i> cloned upstream the T18 coding sequence in p-T18-FLAG	This study
pT18-TssC1	<i>tssC1</i> cloned downstream the T18 coding sequence in p-T18-FLAG	This study
pTssC1-T18	<i>tssC1</i> cloned upstream the T18 coding sequence in p-T18-FLAG	This study
pT18-Pal	<i>pal</i> cloned downstream the T18 coding sequence in p-T18-FLAG	[23]
pT25-FLAG	Bacterial Two Hybrid (BACTH) vector	[23]
pT25-Hcp1	<i>hcp1</i> cloned downstream the T25 coding sequence in p-T25-FLAG	This study
pHcp1-T25	<i>hcp1</i> cloned upstream the T25 coding sequence in p-T25-FLAG	This study
pT25-TssB1	<i>tssB1</i> cloned downstream the T25 coding sequence in p-T25-FLAG	This study
pTssB1-T25	<i>tssB1</i> cloned upstream the T25 coding sequence in p-T25-FLAG	This study

pT25-TssC1	<i>tssC1</i> cloned downstream the T25 coding sequence in p-T25-FLAG	This study
pTssC1-T25	<i>tssC1</i> cloned upstream the T25 coding sequence in p-T25-FLAG	This study
pTolB-T25	<i>tolB</i> cloned upstream the T25 coding sequence in p-T25-FLAG	[23]
pBAD33	empty vector, pACYC184 origin, <i>Para</i> , <i>araC</i> , Cm ^R	[24]
pBAD33-TssB1-VSV-G	<i>tssB1</i> cloned into pBAD33, C-terminal VSV-G epitope	This study
pBAD33-TssB2-VSV-G	<i>tssB2</i> cloned into pBAD33, C-terminal VSV-G epitope	This study
pBAD33-TssB1-sfGFP	<i>sfgfp</i> fused in-frame to <i>tssB1</i> (separated by a 3×Ala 3×Gly linker) in pBAD33-TssB1-VSV-G	This study
pBAD33-VgrG-VSV-G	<i>sci-1 vgrG (vgrG1)</i> cloned into pBAD33, C-terminal VSV-G epitope	This study
pOK12	empty vector, P15A origin, <i>Plac</i> , Kan ^R	[22]
pOK12-TssB1-HA	<i>sci-1 tssB (tssB1)</i> cloned into pOK12, C-terminal HA epitope	M.-S. Aschtgen

OLIGONUCLEOTIDES

For strain construction^a

5-*Δhcp1* TCCCCTGCGCCGGAAGAGGGCGCATCAGAAAACATAACGGAGTAATTTTTGTGTAGGCTGGAGCTGCTTCG
3-*Δhcp1* TGAAGAAAAAATAAAAATGACGGACAGGATGCCCTGTCCGTCCGGCAGAACATATGAATATCCTCCTTAGTTCC
5-*Δsci-1* GGAGGCATCTGCGGTGATGGAACCCCTGAGATGCAGGTTTCACAGGAGAGAGCCTGTGTAGGCTGGAGCTGCTTCG
3-*Δsci-1* CATATATCCATCGCTGTTTATTTATTTCTGTTCTGCCCTCCCTGCTCCGGCCATATGAATATCCTCCTTAGTTC
5-*ΔtssB1* GCATCTGCGGTGATGGAACCCCTGAGATGCAGGTTTCACAGGAGAGAGCCGTGTAGGCTGGAGCTGCTTCG
3-*ΔtssB1* TTCTTTTCTTTCTGTACAGACATCAGCATTTTCTCTCGTAATCCGTTAAACATATGAATATCCTCCTTAGTTCC
5-*ΔtssC1* TGCGTGCACTGGTGCCGGAAAAGGCGTGATTTAACGGATTACGAGAGAAAAGTGTAGGCTGGAGCTGCTTCG
3-*ΔtssC1* GCCCCGTCTTCCCATAATGGGCGATAAATCTTCATTTCCCGACACCTGCCGCATATGAATATCCTCCTTAGTTCC
5-*ΔvgrG* TACCCTTTCAGAGCTGTTTCGTGGAGTGATAAATCATGAATCTCACTGAGTGTAGGCTGGAGCTGCTTCG
3-*ΔvgrG* GGTTTTATGGGTGGCATCTGTTACGTCATATCCCTGATATTTTGTTCATTCATATGAATATCCTCCTTAGTTCC

For plasmid construction^{b, c, d}

5pUC-Hcp2-FLAG GGAAACAGCTATGACCATGATTACGAATATGGTCCGGAGTTATGAGTA
3pUC-Hcp2-FLAG AAGAGGATCCCCGGGCGAGCTCGATTATTTATCATACTCATCTTTATAATCTACAAGAGCCT
CTTTATATAAG
5pBAD-TssB1-VSV-G CTCTCTACTGTTTCTCCATACCCGTTTTTTTTGGGCTAGCAGGAGGTATTACACCATGAGCAGTT
CGTTTCAGAATGAAATCCCG
3pBAD-TssB1-VSV-G GGTCGACTCTAGAGGATCCCCGGGTACCTTATTTTCCTAATCTATTCATTTCAATATCTGTA
TACGCCTTTTCCGGCACCAGTGAC
5pBAD-TssB2-VSV-G CTCTCTACTGTTTCTCCATACCCGTTTTTTTTGGGCTAGCAGGAGGTATTACACCATGAGCAATA
CACAACATAAACTTGATAAAAC
3pBAD-TssB2-VSV-G GGTCGACTCTAGAGGATCCCCGGGTACCTTATTTTCCTAATCTATTCATTTCAATATCTGTA
TATTCCTTACTTTCATCGTTTTTCAGGATTG

5pBAD-TssB1-sfGFP	<u>CACTGGTGCCGGAAAAGGCGGCAGCGGCCGGCGGAGGGTCTAAAGGTGAAGAACTG TTCACCG</u>
3pBAD-TssB1-sfGFP	<u>CGGGTACCTTATTTTCTAATCTATTCATTTCAATATCTGTATAGGAAACAGCTATGACCATGATTACG</u>
5pBAD-VgrG1-VSV-G	<u>CTCTCTACTGTTTCTCCATACCCGTTTTTTTTGGGCTAGCAGGAGGTATTACACCATGAATCTCACTGAC</u> TCCCTGCAAAAATGTTTTATCC
3pBAD-VgrG1-VSV-G	<u>GGTCGACTCTAGAGGATCCCCGGGTACCTTATTTTCTAATCTATTCATTTCAATATCTGTATATTC</u> TGTTTCTCCATGAATTTTTACCTTCCCAAACCTC
T25T18C-5-Hcp1	<u>CGGATAACAATTTACACACAGGAAACAGCTATGACCATGGCAATTCAGTTTATCTGTGGCTGAAAG</u> ATG
T18C-3-Hcp1	<u>CCTCGCTGGCGGCTAAGCTTGCGTAATCGCGGTGGTACGCTCACTCCATGCG</u>
T25C-3-Hcp1	<u>GTTTGCGTAACCAGCCTGATGCGATTGCTGCGCGGTGGTACGCTCACTCCATGCG</u>
T25T18N-3-Hcp1	<u>CGAGGTCGACGGTATCGATAAGCTTGATATCGAATTCTAGTTACGCGGTGGTACGCTCACTCCATGCG</u>
T18N-5-Hcp1	<u>CGCCACTGCAGGGATTATAAAGATGACGATGACAAGGCAATTCAGTTTATCTGTGGCTGAAAGATG</u>
T25N-5-Hcp1	<u>GGCGGGCTGCAGATTATAAAGATGACGATGACAAGGCAATTCAGTTTATCTGTGGCTGAAAGATG</u>
T25T18C-5-TssB1	<u>CGGATAACAATTTACACACAGGAAACAGCTATGACCATGAGCAGTTCGTTTCAGAATGAAATCCCGA</u> AAGCAAG
T18C-3-TssB1	<u>CCTCGCTGGCGGCTAAGCTTGCGTAATCGCCTTTTCCGGCACCAAGTGCACGC</u>
T25C-3-TssB1	<u>GTTTGCGTAACCAGCCTGATGCGATTGCTGCGCCTTTTCCGGCACCAAGTGCACGC</u>
T25T18N-3-TssB1	<u>CGAGGTCGACGGTATCGATAAGCTTGATATCGAATTCTAGTTACGCCTTTTCCGGCACCAAGTGCACGC</u>
T18N-5-TssB1	<u>CGCCACTGCAGGGATTATAAAGATGACGATGACAAGAGCAGTTCGTTTCAGAATGAAATCCCGAAAG</u> CAAG
T25N-5-TssB1	<u>GGCGGGCTGCAGATTATAAAGATGACGATGACAAGAGCAGTTCGTTTCAGAATGAAATCCCGAAAGC</u> AAG
T25T18C-5-TssC1	<u>CGGATAACAATTTACACACAGGAAACAGCTATGACCATGCTGATGTCTGTACAGAAAGAAAAGAACGT</u> TGCAGAGAGCG
T18C-3-TssC1	<u>CCTCGCTGGCGGCTAAGCTTGCGTAATCGCCTTTTGCCTTCGGCATCTGCGAAACCAAGTG</u>
T25C-3-TssC1	<u>GTTTGCGTAACCAGCCTGATGCGATTGCTGCGCCTTTTGCCTTCGGCATCTGCGAAACCAAGTG</u>
T25T18N-3-TssC1	<u>CGAGGTCGACGGTATCGATAAGCTTGATATCGAATTCTAGTTACGCTTTTGCCTTCGGCATCTGCGAA</u> ACCAAGTG
T18N-5-TssC1	<u>CGCCACTGCAGGGATTATAAAGATGACGATGACAAGCTGATGTCTGTACAGAAAGAAAAGAACGTT</u> GCAGAGAGCG
T25N-5-TssC1	<u>GGCGGGCTGCAGATTATAAAGATGACGATGACAAGCTGATGTCTGTACAGAAAGAAAAGAACGTTG</u> CAGAGAGCG

For site-directed mutagenesis ^e

A-Hcp1-N115C	CGTGAAGGTGGTGAAGGTTT <u>G</u> CCCCGGTCATGCACGACATC
B-Hcp1-N115C	GATGTCGTGCATGACCGGG <u>C</u> AAACCTTCACCACCTTCACG
A-Hcp1-S158C	CATTCATTCCGACGCATGGT <u>G</u> TGAGCGTACCACCGCGTAA
B-Hcp1-S158C	TTACGCGGTGGTACGCTC <u>A</u> CACCATGCGTCGGAATGAATG
A-Hcp1-Q24C	CAAAGGGTCCGTGGATGTTT <u>G</u> CGACCGTGAGGGCAGCATTG
B-Hcp1-Q24C	CAATGCTGCCCTCACGGTC <u>G</u> CAAACATCCACGGACCCTTTG
A-Hcp1-G48C	TCCCGACGGATAACAATACCT <u>G</u> TAAGCTGACCGGTACCCG
B-Hcp1-G48C	CGGGTACCGGTCAGCTT <u>A</u> CAGGTATTGTTATCCGTCGGGA
A-Hcp1-A95C	GTGGTACAAAATCAATGAT <u>T</u> GCGGTCAGGAGGTGGAGTATTC
B-Hcp1-A95C	GAAATACTCCACCTCCTGACCG <u>C</u> AATCATTGATTTTGTACCAC
A-Hcp1-G96C	GTACAAAATCAATGATGCCT <u>G</u> TGCAGGAGGTGGAGTATTC
B-Hcp1-G96C	GAAATACTCCACCTCCTG <u>A</u> CAGGCATCATTGATTTTGTAC
A-Hcp1-C38S	GTGGTGGCTCAGGAGCAC <u>A</u> GCCTCTACATCCCGACGG
B-Hcp1-C38S	CCGTCGGGATGTAGAGGCTGTGCTCCTGAGCCACCAC
A-Hcp1-S158W	CATTCATTCCGACGCATGGT <u>G</u> GGAGCGTACCACCGCGTAA
B-Hcp1-S158W	TTACGCGGTGGTACGCTC <u>A</u> CCCATGCGTCGGAATGAATG

^a Sequences corresponding to the downstream and upstream regions of the gene to be deleted underlined.

^b Sequence annealing on the target plasmid underlined.

^c FLAG or VSV-G epitope coding sequence in **Bold**

^d 3×Ala-3×Gly linker in *italics*.

^e Mismatch codon underlined.