

Supporting Information

I.	General Procedures	SI-2
II.	Preparation of Organomagnesium Reagents	SI-3
III.	Demonstration of Cross-Coupling Stereochemical Course	SI-4
IV.	Synthesis and Characterization of Substrates	SI-5
V.	General Procedure for Cross-Coupling Reactions	SI-12
VI.	Characterization Data for Products	SI-12
VII.	Mechanistic Studies	SI-22
VIII.	Biological Experiments	SI-26
IX.	References for Supporting Information	SI-29
X.	Crystallographic Data	SI-30
XI.	^1H and ^{13}C NMR spectra	SI-44
XII.	SFC traces	SI-108

I. General Procedures

All reactions were carried out under an atmosphere of N₂ using glassware that was either oven- or flame-dried prior to use. Hexanes, tetrahydrofuran (THF), diethyl ether (Et₂O), and toluene (PhMe), were degassed with argon and then passed through two 4 x 36 inch columns of anhydrous neutral A-2 alumina (8 x 14 mesh; LaRoche Chemicals; activated under a flow of argon at 350 °C for 12 h) to remove H₂O. All other solvents utilized were purchased “anhydrous” commercially, or purified as described (vide infra). ¹H NMR spectra were recorded on Bruker GN-500 (500 MHz ¹H, 125.7 MHz ¹³C), CRYO-500 (500 MHz ¹H, 125.7 MHz ¹³C) or DRX-400 (400 MHz ¹H, 100 MHz ¹³C) spectrometers. Proton chemical shifts are reported in ppm (δ) relative to internal tetramethylsilane (TMS, δ 0.00). Data are reported as follows: chemical shift (multiplicity [singlet (s), broad singlet (br s), doublet (d), doublet of doublets (dd), triplet (t), doublet of triplets (dt), doublet of doublet of triplets (ddt), triplet of triplets (tt), quartet (q), quintet (quint), multiplet (m), apparent singlet (ap s), apparent doublet (ap d), apparent quartet (ap q), broad doublet (br d) and broad multiplet (br m)], coupling constants [Hz], integration). Carbon chemical shifts are reported in ppm (δ) relative to TMS with the respective solvent resonance as the internal standard (CDCl₃, δ 77.16 ppm). Unless otherwise indicated, NMR data were collected at 25 °C. Analytical thin-layer chromatography (TLC) was performed using Silica Gel 60 F₂₅₄ pre-coated plates (0.25 mm thickness). Visualization was accomplished by irradiation with a UV lamp and/or staining with KMnO₄ solution. Flash chromatography was performed using Silica Gel 60Å (170-400 mesh) from Fisher Scientific. Preparatory HPLC was performed on an Agilent 1100 using an Alltima Silica 5 μm, 250 x 22 mm column. Melting points (m.p.) were obtained using a Mel-Temp melting point apparatus and are uncorrected. Infrared spectra were obtained on a Mattson Instruments *Galaxy 5000* (thin film) and Perkin-Elmer Spectrum 1000 FT-IR Systems and are reported in terms of frequency of absorption (cm⁻¹). High resolution mass spectrometry was performed by the University of California, Irvine Mass Spectrometry Center. Optical rotations were measured with a Rudolph Research Analytical Autopol IV Automatic Polarimeter or a Jasco P-1010 digital polarimeter. SFC determinations of enantiopurity were performed on a Berger Analytical instrument using a Daicel™ Chiralpak® column (OD-H, OJ-H, or AD-H; 100 bar, 50 °C, 215 nm).

Ni(cod)₂ was purchased from Strem, stored in a glovebox freezer (-20 °C) under an atmosphere of N₂, and used as received. Ni(PPh₃)₄ was purchased from Aldrich and used as received. Ni(acac)₂ was purchased from Strem, stored in a glovebox under an atmosphere of N₂, and used as received. 1,2-Bis(diphenylphosphino)ethane (dppe) was purchased from Alfa Aesar, stored in a glovebox under an atmosphere of N₂, and used as received. 1,2-Bis(diphenylphosphino)ethane nickel (II) chloride (Ni(dppe)Cl₂) was purchased from Strem and used as received.

Organomagnesium reagents for substrate synthesis were freshly prepared from the halide precursor in THF and molarities were determined by titration with I₂.¹ Activated MnO₂ was prepared according to a procedure reported by Attenburrow and co-workers.² All other chemicals were purchased commercially and used as received.

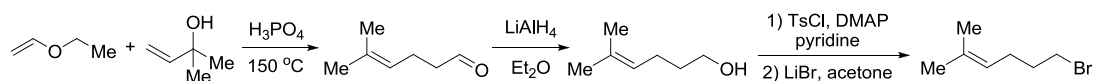
II. Preparation of Organomagnesium Reagents for Cross-Coupling Reactions

For satisfactory yields and enantiospecificities in the cross-coupling reactions, the Grignard reagent must be prepared from the alkyl bromide in diethyl ether.

General Procedure: Magnesium turnings (1.08 g, 45.0 mmol) were added to a round-bottom flask equipped with a stir bar and condenser. The reaction apparatus was flame-dried under vacuum and cooled under N₂. Et₂O (5.0 mL) was added to the reaction apparatus, followed by a single crystal of I₂ (ca. 2 mg). The organohalide³ (15.0 mmol) was added portion-wise over 30 min. The reaction was stirred at ambient temperature for an additional two hours. The resulting Grignard reagents was typically between 2.0 and 3.0 M as titrated using Knochel's method,¹ and could be stored (sealed, under nitrogen) for at least 4 weeks without detrimental effects.

***p*-(*N,N*-Dimethylamino)phenylmagnesium bromide** was prepared according to a procedure reported by Jarvo and co-workers.⁴

(5-methylhex-4-enyl)magnesium bromide was prepared from 5-methylhex-4-enylbromide. The latter has been prepared via the following synthetic route:

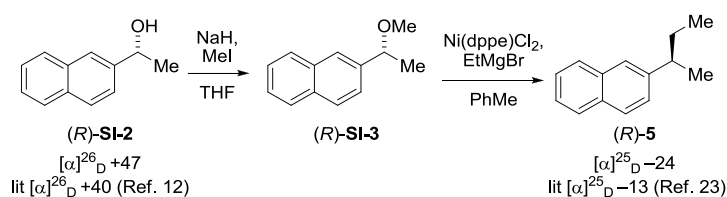


5-methylhex-4-enal was prepared according to a modified procedure reported by Saucy and co-workers.⁵ A sealed tube was charged with ethoxyethane (11.6 mL, 121 mmol), 2-methylbut-3-en-2-ol (6.40 mL, 60 mmol), and 85% phosphoric acid (59 μ L, 0.60 mmol). The tube was sealed, heated to 150 °C, and allowed to stir for 2.5 h. The reaction mixture was cooled to room temp, opened, and neutralized with triethylamine. Purification by fractional distillation (120 °C, 50 torr) afforded the title compound as a clear, colorless oil (2.41 g, 21.5 mmol, 36%). Analytical data is consistent with literature values.⁶

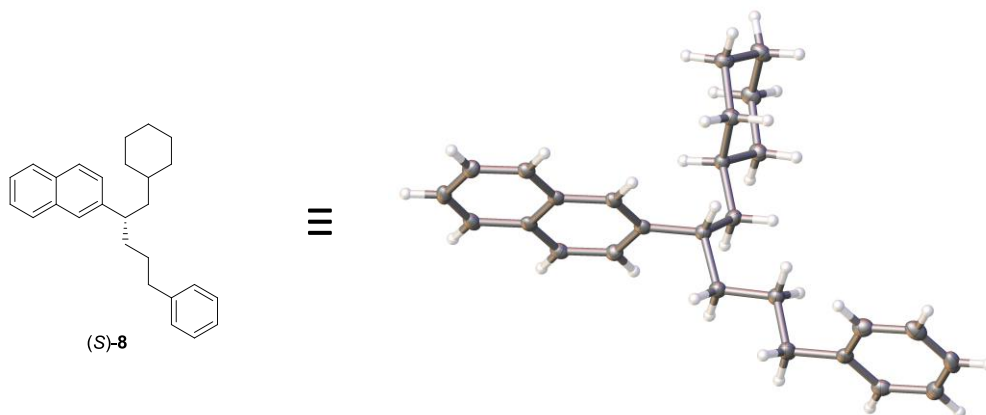
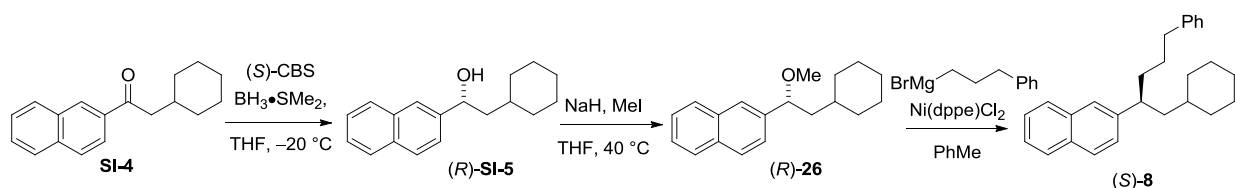
5-methylhex-4-ene-1-ol was prepared according to the procedure reported by Heathcock and co-workers.⁷ Analytical data was consistent with literature values.⁸

6-Bromo-2-methylhex-2-ene was prepared according to the procedure reported by Boyer.⁹ Analytical data was consistent with literature values.⁹

III. Demonstration of Cross-Coupling Stereochemical Course



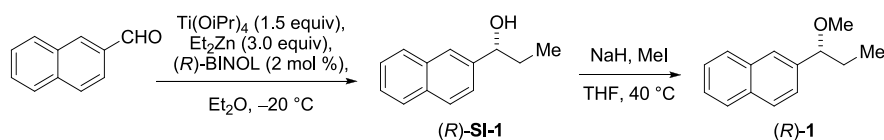
Enantioenriched alcohol **(R)-SI-2** was prepared by CBS reduction using *(S)*-2-methyl-CBS-oxazaborolidine (vide infra), and the stereochemistry was verified by comparison of the optical rotation to the literature value. Conversion to ether **(R)-SI-3** followed by stereospecific cross-coupling produced **(R)-5**, the stereochemistry of which was determined by comparison of the optical rotation to the literature value. This product corresponds to net inversion in the cross-coupling reaction.



Enantioenriched alcohol **(R)-SI-5** was prepared by CBS reduction of **SI-4** using *(S)*-2-methyl-CBS-oxazaborolidine. Absolute configuration was assigned as *R* based on the accepted model for selectivity in CBS reduction¹⁰ and confirmed by Competing Enantioselective Conversion (CEC).¹¹ Conversion to ether **(R)-26** followed by stereospecific cross-coupling produced **(S)-8**, the stereochemistry of which was determined by X-ray crystallographic analysis (vide infra). This product corresponds to net inversion in the cross-coupling reaction. See SI section X for crystallographic data.

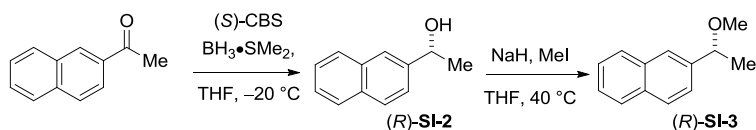
The absolute configurations of all other products were assigned based on the assumption that the cross-coupling reaction proceeds with inversion.

IV. Synthesis and Characterization of Substrates



(R)-1-(naphthalen-2-yl)propan-1-ol ((R)-SI-1). Prepared according to the procedure reported by Jarvo and co-workers.¹² Analytical data is consistent with literature values:¹² $^1\text{H NMR}$ (500 MHz, CDCl_3) δ 7.80 (m, 3H), 7.73 (s, 1H), 7.45 (m, 3H), 4.72 (t, $J = 6.8$ Hz, 1 H), 2.17 (s, 1H), 1.79–1.90 (m, 2H), 0.91 (t, $J = 7.5$ Hz, 3H); $^{13}\text{C NMR}$ (125 MHz, CDCl_3) δ 142.0, 133.3, 133.1, 128.3, 128.0, 127.8, 126.2, 125.9, 124.8, 124.2, 76.2, 31.8, 10.3; $[\alpha]_D^{24} +39.5$ (c 1.01, CHCl_3 , 98% ee), lit. $[\alpha]_D^{23} +41.9$ (c 1.02, CHCl_3 , 96% ee);¹² **SFC** analysis (OD-H, 10% IPA, 2.5 mL/min) indicated 95% ee: t_R (minor) = 9.1 minutes, t_R (major) = 9.9 minutes.

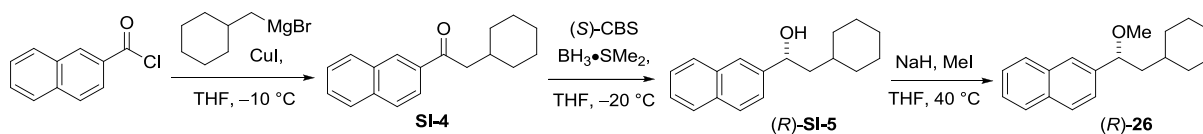
(R)-2-(1-methoxypropyl)naphthalene ((R)-1). Prepared according to the procedure reported by Jarvo and co-workers.¹² Analytical data is consistent with literature values:¹² $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 7.83 (m, 3H), 7.70 (s, 1H), 7.45 (m, 3H), 4.17 (t, $J = 6.6$ Hz, 1 H), 3.24 (s, 3H), 1.92 (m, 1H), 1.75 (m, 1H), 0.89 (t, $J = 7.4$ Hz, 3H); $^{13}\text{C NMR}$ (125 MHz, CDCl_3) δ 139.8, 133.3, 133.2, 128.4, 128.0, 127.8, 126.17, 126.16, 125.8, 124.7, 85.8, 56.9, 30.9, 10.4; $[\alpha]_D^{24} +110.5$ (c 1.07, CHCl_3), lit. $[\alpha]_D^{23} +92.2$ (c 1.82, CHCl_3 , 96% ee);¹² **SFC** analysis (OD-H, 1% IPA, 2.5 mL/min) indicated 95% ee: t_R (minor) = 5.8 minutes, t_R (major) = 6.2 minutes.



(R)-1-(naphthalen-2-yl)ethanol ((R)-SI-2). Prepared according to a modified procedure by Panek and co-workers.¹³ To a cooled ($-30\text{ }^\circ\text{C}$) solution (S) -2-methyl-CBS-oxazaborolidine (0.083 g, 0.30 mmol) in THF (10 mL) was added borane dimethyl sulfide (0.60 mL, 6.0 mmol) and the reaction mixture was stirred for 45 min. Naphthylmethylketone (0.510 g, 3.0 mmol) in THF (5 mL) was added drop-wise and the reaction was further stirred at $-30\text{ }^\circ\text{C}$ for 12 h. The reaction was quenched by slow addition of MeOH (5 mL), warmed to room temperature and diluted with water. The product was extracted with Et_2O (2 x 20 mL) and the combined organics were washed with brine, dried over NaSO_4 , and concentrated in vacuo. Purification by flash chromatography (30% EtOAc in hexanes) afforded the title compound as a white solid (0.48 g, 2.8 mmol, 92%). The product was recrystallized from hexanes to yield higher enantiopurity. Analytical data is consistent with literature values:¹⁴ $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 7.76 (m, 4H), 7.42 (m, 3H), 4.95 (q, $J = 6.0$ Hz, 1H), 2.59 (s, 1H), 1.51 (d, $J = 6.4$ Hz, 3H); $^{13}\text{C NMR}$ (100 MHz, CDCl_3) δ 143.3, 133.4, 133.0, 128.4, 128.0, 127.8, 126.2, 125.9, 123.93, 123.93; $[\alpha]_D^{26} +46.6$ (c 0.86, CHCl_3), lit. $[\alpha]_D^{25} +40.0$ (c 1.00, CHCl_3 , 92% ee);¹⁴ **SFC** analysis (OD-H, 10% IPA, 2.5 mL/min) indicated 92% ee: t_R (minor) = 8.8 minutes, t_R (major) = 9.5 minutes.

(R)-2-(1-methoxyethyl)naphthalene ((R)-SI-3). To a solution of NaH (0.081 g, 3.4 mmol) in THF (10 mL) was added a solution of **(R)-SI-2** (0.291 g, 1.68 mmol) in THF (2 mL) and the mixture was stirred at $40\text{ }^\circ\text{C}$ for 30 min. MeI (0.31 mL, 5.1 mmol) was added and stirring was continued for 8 h. Reaction was quenched with MeOH and the solvent was removed in vacuo. The crude mixture was taken up in CH_2Cl_2 (100 mL), filtered through Celite, and concentrated in vacuo. Purification by flash chromatography (5% EtOAc in hexanes) afforded the title compound as a colorless oil (0.305 g, 1.64 mmol, 97%). Analytical data is consistent with literature values:¹⁵ $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 7.80 (m, 3H), 7.71 (s, 1H), 7.44

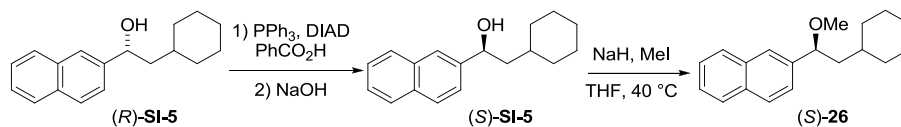
(m, 3H), 4.42 (q, $J = 6.4$ Hz, 1H), 3.23 (s, 3H), 1.50 (d, $J = 6.4$ Hz, 3H); $^{13}\text{C NMR}$ (100 MHz, CDCl_3) δ 141.0, 133.4, 133.1, 128.5, 127.9, 127.8, 126.2, 125.8, 125.2, 124.2, 79.8, 56.6, 24.0; $[\alpha]_{\text{D}}^{26} +105.0$ (c 0.85, CHCl_3); **SFC analysis** (OD-H, 1% IPA, 2.5 mL/min) indicated 92% ee: t_{R} (minor) = 7.6 minutes, t_{R} (major) = 8.3 minutes.



2-cyclohexyl-1-(naphthalen-2-yl)ethanone (SI-4). Prepared according to the procedure reported by Jarvo and co-workers.¹⁶ Analytical data is consistent with literature values:¹⁶ $^1\text{H NMR}$ (500 MHz, CDCl_3) δ 8.45 (s, 1H), 8.03 (d, $J = 9.0$ Hz, 1H), 7.97 (d, $J = 8.0$ Hz, 1H), 7.88 (t, $J = 9.0$ Hz, 2H), 7.46–7.61 (m, 2H), 2.95 (d, $J = 6.5$ Hz, 2H), 2.10–1.97 (m, 1H), 1.85–1.75 (m, 2H), 1.70–1.65 (m, 3H), 1.31–1.26 (m, 2H), 1.22–1.17 (m, 1H), 1.15–1.02 (m, 2H); $^{13}\text{C NMR}$ (125 MHz, CDCl_3) δ 200.4, 135.7, 135.0, 132.7, 129.9, 129.7, 128.52, 128.47, 127.9, 126.8, 124.2, 46.4, 34.9, 33.7, 26.4, 26.3.

(R)-2-cyclohexyl-1-(naphthalen-2-yl)ethanol ((R)-SI-5). Prepared according to the procedure reported by Jarvo and co-workers.¹⁶ Analytical data is consistent with literature values:¹⁶ $^1\text{H NMR}$ (500 MHz, CDCl_3) δ 7.84 (m, 3H), 7.78 (s, 1H), 7.49–7.45 (m, 3H), 4.96 (m, 1H), 1.85–1.77 (m, 4H), 1.71–1.60 (m, 4H), 1.51–1.40 (m, 1H), 1.29–1.13 (m, 3H), 1.04–0.92 (m, 2H); $^{13}\text{C NMR}$ (125 MHz, CDCl_3) δ 142.8, 133.5, 133.1, 128.4, 128.1, 127.8, 126.3, 125.9, 124.6, 124.3, 72.4, 47.1, 34.4, 34.1, 33.1, 26.7, 26.4, 26.3; $[\alpha]_{\text{D}}^{25} +33.3$ (c 0.17, CHCl_3), lit. $[\alpha]_{\text{D}}^{28} +23.8$ (c 1.0, CHCl_3 , 87% ee);¹⁶ **SFC analysis** (AS-H, 3% IPA, 3 mL/min) indicated 96% ee; t_{R} (minor) = 12.2 minutes, t_{R} (major) = 12.8 minutes. Absolute configuration was assigned as *R* based on the accepted model for selectivity in CBS reduction¹⁰ and confirmed by Competing Enantioselective Conversion (CEC).¹¹

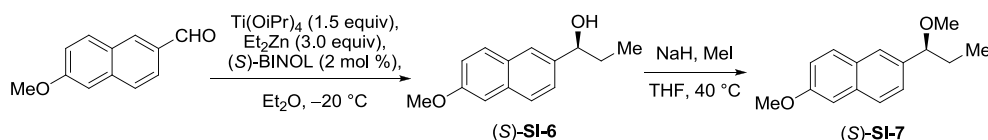
(R)-2-(2-cyclohexyl-1-methoxyethyl)naphthalene ((R)-26). To a solution of NaH (0.514 g, 21.4 mmol) in THF (100 mL) was added (R)-SI-5 (2.72 g, 10.7 mmol) and mixture was stirred at 40 °C for 30 min. MeI (2.00 mL, 32.1 mmol) was added and stirring was continued for 8 h. The reaction was quenched with MeOH and the solvent was removed in vacuo. The crude mixture was taken up in CH_2Cl_2 (100 mL), washed with sat. NaCO_3 , brine, dried over NaSO_4 , and concentrated in vacuo. Purification by flash chromatography (5% EtOAc in hexanes) afforded the title compound as a white solid (2.85 g, 10.6 mmol, 99%): **TLC** $R_{\text{f}} = 0.4$ (5% EtOAc in hexanes); $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 7.83 (m, 3H), 7.70 (s, 1H), 7.49–7.43 (m, 3H), 4.36 (dd, $J = 8.6, 5.7$ Hz, 1H), 3.21 (s, 3H), 1.84–1.76 (m, 3H), 1.69–1.62 (m, 3H), 1.54–1.49 (m, 1H), 1.42–1.37 (m, 1H), 1.25–1.10 (m, 3H), 0.99–0.90 (m, 2H). $^{13}\text{C NMR}$ (100 MHz, CDCl_3) δ 140.5, 133.4, 133.2, 128.4, 127.9, 127.8, 126.2, 125.9, 125.8, 124.6, 81.9, 56.8, 46.2, 34.3, 34.0, 33.3, 26.7, 26.4, 26.3; $[\alpha]_{\text{D}}^{25} +63.2$ (c 1.06, CHCl_3); **SFC analysis** (OJ-H, 3% IPA, 2.5 mL/min) indicated 97% ee t_{R} (minor) = 7.1 minutes, t_{R} (major) = 8.8 minutes.



(S)-2-cyclohexyl-1-(naphthalen-2-yl)ethanol ((S)-SI-5). Prepared according to a modified procedure reported by Presnell and co-workers.¹⁷ To a cooled (brine ice bath) solution of (R)-SI-5 (0.15 g, 0.59 mmol), benzoic acid (0.29 g, 2.4 mmol), and PPh_3 (0.62 g, 2.4 mmol) in THF (5 mL) was added DIAD (0.46 mL, 2.4 mmol) drop-wise over 30 min. After the addition was complete the reaction was allowed to

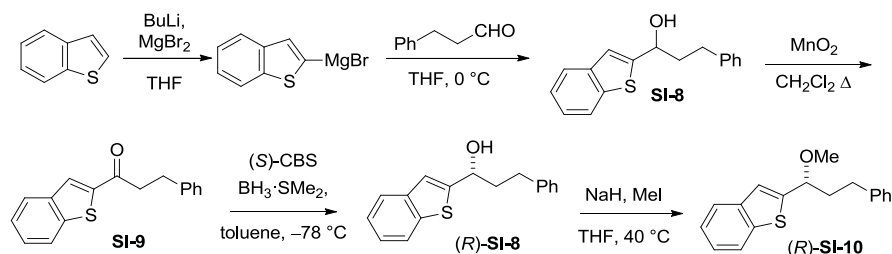
warm to room temperature and stir for 12 h then heated up to 40 °C and stirred for an additional 3 h. The reaction mixture was diluted with Et₂O (10 mL), washed with sat. NaHCO₃, dried over NaSO₄, and concentrated in vacuo. Trituration with hexanes removed the PPh₃ by-products and purification by flash chromatography (10% EtOAc in hexanes) afforded the title compound as a white solid (0.112 g, 0.43 mmol, 73%). The product was recrystallized from hexanes to yield higher enantiopurity. Analytical data is consistent with literature values:¹⁶ ¹H NMR (500 MHz, CDCl₃) δ 7.84 (m, 3H), 7.78 (s, 1H), 7.49–7.45 (m, 3H), 4.96 (m, 1H), 1.85–1.79 (m, 4H), 1.71–1.60 (m, 4H), 1.51–1.40 (m, 1H), 1.29–1.13 (m, 3H), 1.04–0.92 (m, 2H); ¹³C NMR (125 MHz, CDCl₃) δ 142.8, 133.5, 133.1, 128.4, 128.1, 127.8, 126.3, 125.9, 124.6, 124.3, 72.4, 47.1, 34.4, 34.1, 33.1, 26.7, 26.4, 26.3; [α]_D²⁶ –27.4 (c 1.02, CHCl₃); **SFC analysis** (AS-H, 3% IPA, 3 mL/min) indicated 94% ee; t_R (major) = 11.8 minutes, t_R (minor) = 12.7 minutes.

(S)-2-(2-cyclohexyl-1-methoxyethyl)naphthalene ((S)-26). Prepared according to the procedure reported above for (*R*)-26. ¹H NMR (400 MHz, CDCl₃) δ 7.83 (m, 3H), 7.70 (s, 1H), 7.49–7.43 (m, 3H), 4.36 (dd, *J* = 8.6, 5.7 Hz, 1H), 3.21 (s, 3H), 1.84–1.76 (m, 3H), 1.69–1.62 (m, 3H), 1.54–1.49 (m, 1H), 1.42–1.37 (m, 1H), 1.25–1.10 (m, 3H), 0.99–0.90 (m, 2H). ¹³C NMR (100 MHz, CDCl₃) δ 140.5, 133.4, 133.2, 128.4, 127.9, 127.8, 126.2, 125.9, 125.8, 124.6, 81.9, 56.8, 46.2, 34.3, 34.0, 33.3, 26.7, 26.4, 26.3; [α]_D²⁵ –58.1 (c 1.30, CHCl₃); **SFC analysis** (OJ-H, 3% IPA, 2.5 mL/min) indicated 92% ee t_R (major) = 7.0 minutes, t_R (minor) = 8.9 minutes.



(S)-1-(6-methoxynaphthalen-2-yl)propan-1-ol ((S)-SI-6) Prepared according to the procedure reported by Jarvo and co-workers.¹² Analytical data is consistent with literature values:¹² ¹H NMR (500 MHz, CDCl₃) δ 7.72 (m, 3H), 7.44 (d, *J* = 8.5 Hz, 1H), 7.14 (d, *J* = 11.5 Hz, 2H), 4.72 (t, *J* = 6.5 Hz, 1 H), 3.92 (s, 3H), 1.92–1.82 (m, 3H), 0.93 (t, *J* = 7.3 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 157.8, 139.8, 134.2, 129.5, 128.8, 127.2, 124.83, 124.80, 119.1, 105.8, 76.3, 55.5, 31.9, 10.3; [α]_D²⁸ –37.3 (c 0.44, CHCl₃), lit. [α]_D²³ –28.9 (c 0.97, CHCl₃, 90% ee);¹² **SFC analysis** (OD-H, 12.5% IPA, 3 mL/min) indicated 94% ee: t_R (major) = 5.3 minutes, t_R (minor) = 6.5 minutes. Absolute configuration was assigned as *S* based on analogy with the titanium-catalyzed ethylation of aldehydes with Et₂Zn¹⁸ and confirmed by Competing Enantioselective Conversion (CEC).¹¹

(S)-2-methoxy-6-(1-methoxypropyl)naphthalene ((S)-SI-7). Prepared according to the procedure reported by Jarvo and co-workers.¹² Analytical data is consistent with literature values:¹² ¹H NMR (500 MHz, CDCl₃) δ 7.73 (dd, *J* = 5.0, 8.0 Hz, 2H), 7.63 (s, 1H), 7.39 (d, *J* = 8.5 Hz, 1H), 7.14 (m, 2H), 4.14 (t, *J* = 6.8 Hz, 1 H), 3.92 (s, 3H), 3.23 (s, 3H), 1.91 (p, *J* = 7.0 Hz, 1H), 1.74 (quint, *J* = 7.0 Hz, 1H), 0.88 (t, *J* = 7.5 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 157.7, 137.4, 134.3, 129.4, 128.8, 127.2, 126.1, 125.2, 119.0, 105.8, 85.8, 56.8, 55.5, 30.9, 10.4; [α]_D²⁴ –82.3 (c 0.86, CHCl₃), lit. [α]_D²³ –81.9 (c 1.09, CHCl₃, 90% ee);¹² **SFC analysis** (OD-H, 4% IPA, 2.5 mL/min) indicated 93% ee: t_R (major) = 5.4 minutes, t_R (minor) = 5.9 minutes.



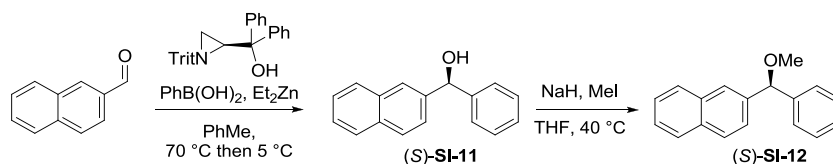
(rac)-1-(benzo[*b*]thiophen-2-yl)-3-phenylpropan-1-ol ((rac)-SI-8). Prepared according to a modified procedure reported by Guinchard and co-workers.¹⁹ To a cooled (ice bath) solution of benzothiophene (3.35 g, 25.0 mmol) in Et₂O (15 mL) was added *n*-BuLi (21 mL, 28 mmol, 1.3 M in hexane) and the reaction was stirred for two hours allowing it to warm to room temperature. The solution was then cooled to -78 °C and hydrocinnamaldehyde (2.57 mL, 20.0 mmol) was slowly added. The reaction was stirred overnight allowing it to warm up to room temperature. The reaction was quenched with MeOH (10 mL) and diluted with Et₂O (100 mL). The solution was washed with water, brine, dried over NaSO₄, and concentrated in vacuo. Purification by flash chromatography (10–15% EtOAc in hexanes) afforded the title compound as a white solid (3.42 g, 12.7 mmol, 64%). **m.p.** 95–97 °C; **TLC** R_f = 0.5 (20% EtOAc in hexanes); **¹H NMR** (400 MHz, CDCl₃) δ 7.82 (d, J = 7.2 Hz, 1H), 7.71 (d, J = 7.2 Hz, 1H), 7.36–7.19 (m, 8H), 5.00 (dd, J = 10.8, 5.6 Hz, 1H), 2.84–2.71 (m, 2H), 2.31–2.15 (m, 2H), 2.13 (d, J = 4.0 Hz, 1H); **¹³C NMR** (125 MHz, CDCl₃) δ 149.2, 141.4, 139.6, 139.5, 128.63, 128.62, 126.2, 124.5, 124.4, 123.6, 122.7, 120.5, 70.2, 40.5, 32.0; **I.R.** (neat) 2929, 1181, 1085, 1054, 873 cm⁻¹; **HRMS** (TOF MS EI+) m/z calcd for C₁₇H₁₆OS [M]⁺ 268.0922, found 268.0912.

1-(benzo[*b*]thiophen-2-yl)-3-phenylpropan-1-one (SI-9). To a solution of *rac*-SI-8 (2.83 g, 10.5 mmol) in CH₂Cl₂ (10 mL) was added MnO₂ (4.58 g, 52.7 mmol) and the reaction mixture was stirred overnight. Upon completion as judged by TLC, the reaction mixture was cooled to room temperature, passed through a plug of Celite, and the solvent was removed in vacuo. The resulting solid was purified by flash chromatography (10–30% EtOAc in hexanes) followed by recrystallization from EtOAc/hexanes to afford the title compound as a colorless crystalline solid (2.03 g, 7.62 mmol, 73% yield): **m.p.** 118–120 °C; **TLC** R_f = 0.6 (20% EtOAc in hexanes); **¹H NMR** (400 MHz, CDCl₃) δ 7.91 (s, 1H), 7.85 (d, J = 9.2 Hz, 2H), 7.45 (m, 1H), 7.39 (m, 1H), 7.32–7.19 (m, 5H), 3.33 (t, J = 4.0 Hz, 2H), 3.10 (t, J = 7.8 Hz, 2H); **¹³C NMR** (125 MHz, CDCl₃) δ 193.8, 143.6, 142.6, 141.0, 139.2, 129.1, 128.7, 128.6, 127.5, 126.4, 126.0, 125.1, 123.1, 41.2, 30.5; **I.R.** (neat) 3052, 3024, 2920, 1665, 1162, 752, 698 cm⁻¹; **HRMS** (TOF MS ES+) m/z calcd for C₁₇H₁₄OS [M+Na]⁺ 289.0663, found 289.0663.

(R)-1-(benzo[*b*]thiophen-2-yl)-3-phenylpropan-1-ol ((R)-SI-8). Prepared according to a modified procedure by Panek and co-workers.¹³ To a cooled (-78 °C) solution (*S*)-2-methyl-CBS-oxazaborolidine (0.211 g, 0.762 mmol) and SI-9 (2.03 g, 7.62 mmol) in toluene (50 mL) was added dropwise BH₃·Me₂S (1.56 mL, 15.2 mmol) and the reaction mixture was stirred for 20 h. The reaction was quenched by slow addition of water (5 mL), warmed to room temperature and extracted with Et₂O (3 x 50 mL). The combined organics were washed with brine, dried over NaSO₄, and concentrated in vacuo. Purification by flash chromatography (3% EtOAc in hexanes) afforded the title compound as a white solid (1.92 g, 7.15 mmol, 94%). The product was recrystallized from EtOAc and hexanes to yield higher enantiopurity. **[α]_D²⁷** +99.2 (c 0.89, CHCl₃); **SFC analysis** (AD-H, 15% MeOH, 3 mL/min) indicated 99% ee t_R (major) = 10.8 minutes, t_R (minor) = 13.6 minutes. Absolute configuration was assigned as *R* based on the accepted model for selectivity in CBS reductions.¹⁰

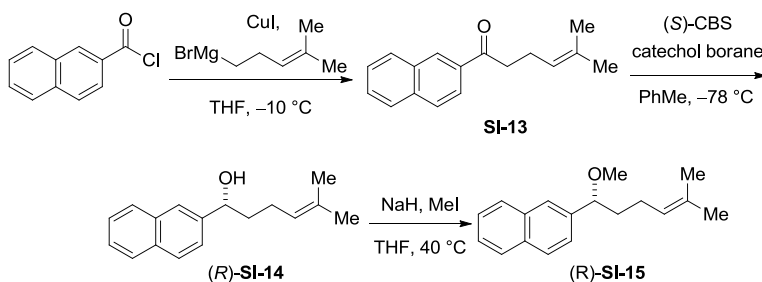
(R)-2-(1-methoxy-3-phenylpropyl)benzo[*b*]thiophene ((R)-SI-10). To a solution of NaH (0.291 g, 12.1 mmol) in THF (30 mL) was added a solution of (*R*)-SI-8 (1.63 g, 6.07 mmol) in THF (10 mL) and the mixture was stirred at 40 °C for 30 min. MeI (1.13 mL, 18.22 mmol) was added and stirring was

continued for 8 h. Reaction was quenched with MeOH and the solvent was removed in vacuo. The crude mixture was taken up in CH₂Cl₂ (100 mL), filtered, and concentrated in vacuo. Purification by flash chromatography (5% EtOAc in hexanes) afforded the title compound as a white solid (1.63 g, 5.77 mmol, 95%). **m.p.** 52–54 °C; **TLC** R_f = 0.7 (15% EtOAc in hexanes); ¹**H NMR** (400 MHz, CDCl₃) δ 7.81 (d, *J* = 7.6 Hz, 1H), 7.71 (d, *J* = 7.2 Hz, 1H), 7.35–7.26 (m, 4H), 7.21–7.17 (m, 4H), 4.40 (t, *J* = 6.8 Hz, 1H), 3.30 (s, 3H), 2.72 (m, 2H), 2.29 (m, 1H), 2.10 (m, 1H); ¹³**C NMR** (125 MHz, CDCl₃) δ 147.1, 141.6, 139.8, 139.5, 128.6, 128.5, 126.0, 124.3 (2C), 123.5, 122.7, 122.3, 79.1, 56.8, 39.6, 32.0; **IR** (neat) 3022, 2930, 2860, 1456, 1099, 839, 750 cm⁻¹; **HRMS** (TOF MS EI⁺) *m/z* calcd for C₁₈H₁₈OS [M]⁺ 282.1078 found 282.1074; [α]_D²⁷ +16.6 (c 1.20, CHCl₃); **SFC analysis** (AD-H, 10% IPA, 3 mL/min) indicated 99% ee t_R (major) = 6.2 minutes, t_R (minor) = 7.4 minutes.



(S)-naphthalen-2-yl(phenyl)methanol ((S)-SI-11). Prepared according to the procedure reported by Jarvo and co-workers.¹² Analytical data is consistent with literature values:¹² ¹**H NMR** (500 MHz, CDCl₃) δ 7.89 (s, 1H), 7.83–7.78 (m, 3H), 7.49–7.41 (m, 5H), 7.35 (m, 2H), 7.27 (m, 1H), 6.00 (d, *J* = 3.5 Hz, 1H), 2.33 (d, *J* = 3.5 Hz, 1H); ¹³**C NMR** (125 MHz, CDCl₃) δ 143.8, 141.2, 133.4, 133.0, 128.7, 128.5, 128.2, 127.83, 127.81, 126.8, 126.3, 126.1, 125.1, 124.9, 76.5; [α]_D²⁹ –4.1 (c 1.22, benzene), lit. [α]_D¹⁹ –3.8 (c 1.70, benzene, >98% ee);²⁰ **SFC analysis** (OD-H, 20% IPA, 3 mL/min) indicated 99% ee: t_R (major) = 6.6 minutes, t_R (minor) = 7.6 minutes.

(S)-2-(methoxy(phenyl)methyl)naphthalene ((S)-SI-12). Prepared according to the procedure reported by Jarvo and co-workers.¹² Analytical data is consistent with literature values:¹² ¹**H NMR** (500 MHz, CDCl₃) δ 7.84–7.78 (m, 4H), 7.48–7.39 (m, 5H), 7.32 (t, *J* = 7.5 Hz, 2H), 7.25 (m, 1H), 5.40 (s, 1H), 3.43 (s, 3H); ¹³**C NMR** (125 MHz, CDCl₃) δ 142.1, 139.6, 133.4, 133.0, 128.6, 128.4, 128.1, 127.8, 127.7, 127.1, 126.2, 126.0, 125.9, 125.1, 85.6, 57.2; [α]_D²⁵ –29.1 (c 1.20, CHCl₃), lit. [α]_D²³ –32.0 (c 1.11, CHCl₃, >99% ee);¹² **SFC analysis** (OD-H, 5% IPA, 3 mL/min, 110 psi) indicated >99% ee: t_R (major) = 5.3 minutes, t_R (minor) = 6.4 minutes.



5-methyl-1-(naphthalen-2-yl)hex-4-en-1-one (SI-13). Prepared according to a modified procedure reported by Hultzsich and co-workers.²¹ To a cooled (brine/ice bath) solution of CuI (0.059 g, 0.31 mmol) and 2-naphthoyl chloride (1.19 g, 6.25 mmol) in THF (7 mL) was slowly added (4-methylpent-3-enyl)magnesium bromide (10.9 mL, 6.25 mmol, 0.574 M in THF) and the reaction mixture was stirred for 1 h. The reaction was quenched by the addition of MeOH and the solvent was removed in vacuo. The crude reaction mixture was taken up in EtOAc (30 mL), washed with 1 N HCl (10 mL), sat. NaHCO₃ (10 mL), brine (10 mL), dried with Na₂SO₄, and concentrated in vacuo. Purification by flash chromatography (4% EtOAc in hexanes) afforded the title compound as a colorless oil (1.01 g, 4.22 mmol, 68% yield).

This compound has been previously reported.²² Our spectral data is not fully consistent with the reported literature values, therefore we provide full characterization data to support our structural assignment: **TLC** R_f = 0.4 (10% EtOAc in hexanes); **¹H NMR** (400 MHz, CDCl₃) δ 8.46 (s, 1H), 8.03 (d, J = 8.8 Hz, 1H), 7.94 (d, J = 8.0 Hz, 1H), 7.86 (t, J = 7.4 Hz, 2H), 7.56 (m, 2H), 5.22 (t, J = 7.2 Hz, 1H), 3.12 (t, J = 7.4 Hz, 2H), 2.48 (q, J = 7.4 Hz, 2H), 1.70 (s, 3H), 1.65 (s, 3H); **¹³C NMR** (100 MHz, CDCl₃) δ 200.0, 135.6, 134.4, 132.8, 132.6, 129.7, 129.6, 128.43, 128.39, 127.8, 126.8, 124.0, 123.1, 38.9, 25.8, 23.2, 17.8; **IR** (neat) 3058, 2967, 2913, 1678, 1277, 1180, 1123 cm⁻¹; **HRMS** (TOF MS Cl⁺) m/z calcd for C₁₇H₁₈O [M+H]⁺ 239.1436, found 239.1429.

(R)-5-methyl-1-(naphthalen-2-yl)hex-4-en-1-ol ((R)-SI-14). Prepared according to a modified procedure by Okamura and co-workers.²³ To a cooled (-78 °C) solution (*S*)-2-methyl-CBS-oxazaborolidine (0.12 g, 0.42 mmol) and ketone **SI-13** (1.01 g, 4.22 mmol) in toluene (20 mL) was added dropwise catecholborane (0.90 mL, 8.4 mmol) and the reaction mixture was stirred for 20 h. The reaction was quenched by slow addition of MeOH (2 mL) followed by sat. NH₄Cl (10 mL), warmed to room temperature and extracted with Et₂O (3 x 20 mL). The combined organics were washed with sat. NaCO₃, brine, dried over NaSO₄, and concentrated in vacuo. Purification by flash chromatography (10–20% EtOAc in hexanes) afforded the title compound as a white solid (0.703 g, 2.91 mmol, 69%). The product was recrystallized from hexanes to yield higher enantiopurity. **m.p.** 53–55 °C; **TLC** R_f = 0.5 (20% EtOAc in hexanes); **¹H NMR** (400 MHz, CDCl₃) δ 7.82 (d, J = 8.0 Hz, 3H), 7.76 (s, 1H), 7.46 (m, 3H), 5.16 (t, J = 7.0 Hz, 1H), 4.83 (t, J = 5.8 Hz, 1H), 2.12–2.04 (m, 3H), 1.94–1.82 (m, 2H), 1.69 (s, 3H), 1.58 (s, 3H); **¹³C NMR** (150 MHz, CDCl₃) δ 142.3, 133.4, 133.1, 132.5, 128.4, 128.1, 127.8, 126.2, 125.9, 124.7, 124.3, 123.9, 74.5, 39.0, 25.9, 24.6, 17.9; **IR** (neat) 3263, 2924, 2856, 1059, 1016 cm⁻¹; **HRMS** (TOF MS Cl⁺) m/z calcd for C₁₇H₂₀O [M+NH₄]⁺ 258.1858, found 258.1847. $[\alpha]_D^{26}$ +4.9 (*c* 1.16, CHCl₃); **SFC analysis** (OD-H, 15% IPA, 3 mL/min) indicated 93% ee t_R (minor) = 4.9 minutes, t_R (major) = 5.3 minutes. Absolute configuration was assigned as *R* based on the accepted model for selectivity in CBS reductions.¹⁰

(R)-2-(1-methoxy-5-methylhex-4-enyl)naphthalene ((R)-SI-15). To a solution of NaH (0.032 g, 1.3 mmol) in THF (3 mL) was added a solution of (*R*)-**SI-14** (0.160 g, 0.67 mmol) in THF (3 mL) and the mixture was stirred at 40 °C for 30 min. MeI (0.124 mL, 2.00 mmol) was added and stirring was continued for 5 h. Reaction was quenched with MeOH and the solvent was removed in vacuo. The crude mixture was taken up in CH₂Cl₂ (10 mL), filtered, and concentrated in vacuo. Purification by flash chromatography (5% EtOAc in hexanes) afforded the title compound as a colorless oil (0.164 g, 0.64 mmol, 96%). **TLC** R_f = 0.5 (10% EtOAc in hexanes); **¹H NMR** (400 MHz, CDCl₃) δ 7.84 (app d, J = 8.0 Hz, 3H), 7.70 (s, 1H), 7.46 (m, 3H), 5.12 (t, J = 6.8 Hz, 1H), 4.24 (t, J = 6.6 Hz, 1H), 3.23 (s, 3H), 2.04 (m, 2H), 1.93 (m, 1H), 1.73 (m, 4H), 1.56 (s, 3H); **¹³C NMR** (125 MHz, CDCl₃) δ 140.0, 133.4, 133.2, 132.2, 128.4, 128.0, 127.9, 126.2, 126.1, 125.8, 124.6, 124.0, 83.6, 56.8, 38.1, 25.9, 24.5, 17.9; **IR** (neat) 3055, 2966, 2926, 1445, 1099, 745 cm⁻¹; **HRMS** (TOF MS Cl⁺) m/z calcd for C₁₈H₂₂O [M]⁺ 254.1671, found 254.1652; $[\alpha]_D^{27}$ +30.2 (*c* 1.11, CHCl₃); **SFC analysis** (OD-H, 5% IPA, 2.5 mL/min) indicated 94% ee t_R (minor) = 4.2 minutes, t_R (major) = 4.6 minutes.



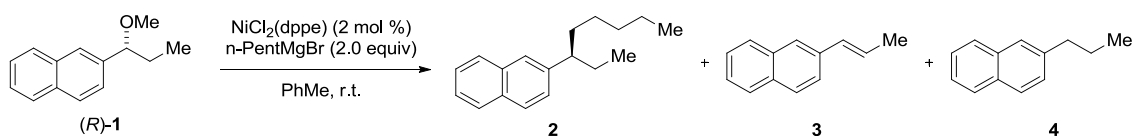
1-(benzofuran-2-yl)-3-methylbutan-1-one (SI-16). Prepared according to a modified procedure reported by Hultsch and co-workers.²¹ To a cooled (brine/ice bath) solution of CuI (0.050 g, 0.260 mmol) and benzofuran-2-carbonyl chloride (0.944 g, 5.23 mmol) in THF (15 mL) was slowly added isobutylmagnesium bromide (7.47 mL, 5.23 mmol, 0.700 M in THF) and the reaction mixture was stirred for 1 h. The reaction was quenched by the addition of MeOH and the solvent was removed in vacuo. The

crude reaction mixture was taken up in EtOAc (30 mL), washed with 1 N HCl (10 mL), sat. NaHCO₃ (10 mL), brine (10 mL), dried with NaSO₄, and concentrated in vacuo. Purification by flash chromatography (4% EtOAc in hexanes) afforded the title compound as a colorless oil (0.833 g, 4.12 mmol, 79% yield): **TLC** R_f = 0.4 (10% EtOAc in hexanes); **¹H NMR** (400 MHz, CDCl₃) δ 7.71 (d, J = 7.6 Hz, 1H), 7.59 (d, J = 8.0 Hz, 1H), 7.50–7.46 (m, 2H), 7.31 (t, J = 7.4 Hz, 1H), 2.83 (d, J = 6.8 Hz, 2H), 2.35 (sept, J = 6.6 Hz, 1H), 1.03 (d, J = 6.4 Hz, 6H); **¹³C NMR** (125 MHz, CDCl₃) δ 191.5, 155.8, 153.1, 128.3, 127.2, 124.0, 123.4, 112.9, 112.6, 47.9, 25.6, 22.9; **IR** (neat) 2957, 2871, 1678, 1556, 1298, 1161, 1143 cm⁻¹; **HRMS** (TOF MS Cl⁺) m/z caclcd for C₁₃H₁₄O₂ [M+H]⁺ 203.1072, found 203.1069.

(R)-1-(benzofuran-2-yl)-3-methylbutan-1-ol ((R)-SI-17). Prepared according to a modified procedure by Panek and co-workers.¹³ To a cooled (–20 °C) solution (*S*)-2-methyl-CBS-oxazaborolidine (0.114 g, 0.41 mmol) in THF (30 mL) was added BH₃·Me₂S (0.800 mL, 8.43 mmol) and the reaction mixture was stirred for 30 min. A solution of **SI-16** (0.700 g, 3.46 mmol) in THF (5 mL) was added and the reaction mixture was stirred for 17 h. at –20 °C. The reaction was quenched by slow addition of MeOH (5 mL) followed by sat. NH₄Cl (30 mL), warmed to room temperature and extracted with EtOAc (3 x 30 mL). The combined organics were washed with sat. NaCO₃, brine, dried over NaSO₄, and concentrated in vacuo. Purification by flash chromatography (5–10% EtOAc in hexanes) followed by recrystallization from hexanes afforded the title compound as white needles (0.461 g, 2.25 mmol, 65%). **m.p.** 72–73 °C; **TLC** R_f = 0.5 (20% EtOAc in hexanes); **¹H NMR** (500 MHz, CDCl₃) δ 7.53 (d, J = 7.5 Hz, 1H), 7.45 (d, J = 8.0 Hz, 1H), 7.26 (m, 1H), 7.21 (t, J = 7.5 Hz, 1H), 6.60 (s, 1H), 4.88 (d, J = 4.0 Hz, 1H), 2.05 (d, J = 4.0 Hz, 1H), 1.87–1.75 (m, 3H), 0.98 (t, J = 5.5 Hz, 6H); **¹³C NMR** (125 MHz, CDCl₃) δ 159.8, 154.9, 128.3, 124.2, 122.9, 121.1, 111.4, 102.5, 66.7, 44.6, 24.7, 23.2, 22.3; **IR** (neat) 3339, 2955, 2869, 1454, 1253 cm⁻¹; **HRMS** (TOF MS Cl⁺) m/z caclcd for C₁₃H₁₆O₂ [M]⁺ 204.1150, found 204.1155. [α]_D²⁴ +25.3 (*c* 1.17, CHCl₃); **SFC analysis** (AD-H, 5% MeOH, 3 mL/min) indicated 95% ee t_R (minor) = 4.9 minutes, t_R (major) = 5.2 minutes. Absolute configuration was assigned as *R* based on the accepted model for selectivity in CBS reductions.¹⁰

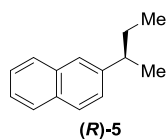
(R)-2-(1-methoxy-3-methylbutyl)benzofuran ((R)-SI-18). To a solution of NaH (0.103 g, 4.30 mmol) in THF (5 mL) was added a solution of (*R*)-**SI-17** (0.439 g, 2.15 mmol) in THF (3 mL) and the mixture was stirred at 40 °C for 30 min. MeI (0.402 mL, 6.44 mmol) was added and stirring was continued for 8 h. Reaction was quenched with MeOH and the solvent was removed in vacuo. The crude mixture was taken up in CH₂Cl₂ (100 mL), filtered, and concentrated in vacuo. Purification by flash chromatography (5% EtOAc in hexanes) afforded the title compound as a colorless oil (0.467 g, 2.14 mmol, 99%): **TLC** R_f = 0.5 (10% EtOAc in hexanes); **¹H NMR** (400 MHz, CDCl₃) δ 7.54 (d, J = 7.6 Hz, 1H), 7.48 (d, J = 8.0 Hz, 1H), 7.28–7.19 (m, 2H), 6.64 (s, 1H), 4.35 (t, J = 7.2 Hz, 1H), 3.32 (s, 3H), 1.90 (m, 1H), 1.75–1.68 (m, 2H), 0.94 (t, J = 6.8 Hz, 6H); **¹³C NMR** (125 MHz, CDCl₃) δ 157.5, 155.1, 128.2, 124.2, 122.8, 121.0, 111.5, 104.6, 75.7, 56.9, 43.3, 24.8, 23.0, 22.5; **IR** (neat) 2955, 2931, 2869, 1454, 1252, 1093 cm⁻¹; **HRMS** (TOF MS Cl⁺) m/z caclcd for C₁₄H₁₈O₂ [M]⁺ 218.1307, found 218.1302; [α]_D²⁴ +100.3 (*c* 1.05, CHCl₃); **SFC analysis** (AD-H, 3% IPA, 3 mL/min) indicated 97% ee t_R (major) = 2.0 minutes, t_R (minor) = 2.2 minutes.

V. Representative Procedure for Cross-Coupling Reactions

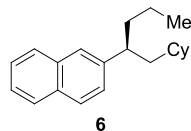


(*R*)-2-(octan-3-yl)naphthalene (**2**, Table 1, entry 16): A vial was charged with (*R*)-1 (0.100 g, 0.500 mmol) and NiCl₂(dppe) (5.3 mg, 0.010 mmol), flushed with nitrogen and capped. Toluene (3.0 mL) was added, followed by *n*-pentylmagnesium bromide (0.60 mL, 1.0 mmol, 1.7 M in Et₂O) and the reaction was allowed to stir at room temperature for 24 h. The reaction was quenched by the addition of EtOAc (1 mL) and the entire reaction mixture was adsorbed onto silica gel (1 g). The solvents were removed in vacuo and the crude was purified by flash column chromatography (100% hexanes). The resulting colorless oil (0.118 g) was a mixture of the title compound (96% calculated yield), the product of reduction (2% calculated yield), and the product of elimination (1% calculated yield). Further purification (100% heptanes) afforded a pure sample of (*R*)-2: **TLC** R_f = 0.6 (100% hexanes); **¹H NMR** (400 MHz, CDCl₃) δ 7.80–7.76 (m, 3H), 7.56 (s, 1H), 7.46–7.38 (m, 2H), 7.30 (dd, J = 8.4, 0.8 Hz, 1H), 2.56 (m, 1H), 1.78–1.60 (m, 4H), 1.26–1.10 (m, 6H), 0.82 (m, 6H); **¹³C NMR** (125 MHz, CDCl₃) δ 143.7, 133.7, 132.2, 127.9, 127.7, 127.6, 126.5, 126.2, 125.8, 125.1, 48.2, 36.6, 32.2, 29.8, 27.5, 22.7, 14.2, 12.4; **IR** (neat) 3056, 2981, 2918, 2850, 1107, 1067 cm⁻¹; **HRMS** (TOF MS CI⁺) m/z calcd for C₁₈H₂₄ [M]⁺ 240.1878, found 240.1871; **$[\alpha]_D^{27}$** -1.6 (c 1.83, CHCl₃); **SFC** analysis (OJ-H, 2% hexanes, 2 mL/min) indicated 96% ee: t_R (minor) = 10.0 minutes, t_R (major) = 10.6 minutes.

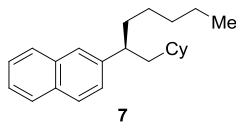
VI. Characterization Data for Products



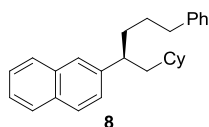
(*R*)-2-sec-butyl-1-naphthalene ((*R*)-5, table 2, entry 1): Prepared according to the representative procedure outlined above using the following amounts of reagents: (*R*)-SI-3 (0.093 g, 0.50 mmol) and NiCl₂(dppe) (5.3 mg, 0.010 mmol), ethylmagnesium bromide (0.36 mL, 1.0 mmol, 2.8 M in Et₂O) and toluene (3.0 mL). Purification by flash column chromatography (100% hexanes) afforded the title compound as a colorless oil (0.73 g, 80%). Analytical data is consistent with literature values:¹² **¹H NMR** (400 MHz, CDCl₃) δ 7.81–7.76 (m, 3H), 7.60 (s, 1H), 7.46–7.34 (m, 2H), 7.35 (d, J = 8.4 Hz, 1H), 2.76 (sextet, J = 7.2 Hz, 1H), 1.73–1.65 (m, 2H), 1.32 (d, J = 6.8 Hz, 3H), 0.85 (t, J = 7.0 Hz, 3H); **¹³C NMR** (100 MHz, CDCl₃) δ 145.3, 133.8, 132.3, 128.0, 127.71, 127.68, 126.0, 125.9, 125.3, 125.1, 42.0, 31.2, 22.0, 12.4; **$[\alpha]_D^{27}$** -26.5 (c 1.17, CHCl₃), **$[\alpha]_D^{25}$** -24.4 (c 1.00, heptane), lit. **$[\alpha]_D^{25}$** -12.7 (c 2.4, heptane, 42% ee);²⁴ **SFC** analysis (OD-H, 1% hexanes, 2.0 mL/min) indicated 91% ee t_R (minor) = 9.9 minutes, t_R (major) = 10.4 minutes.



(+)-2-(1-cyclohexylpentan-2-yl)naphthalene (6, table 2, entry 2): Prepared according to the representative procedure outlined above using the following amounts of reagents: (*R*)-**26** (0.134 g, 0.500 mmol) and NiCl₂(dppe) (5.3 mg, 0.010 mmol), *n*-propylmagnesium bromide (0.44 mL, 1.0 mmol, 2.3 M in Et₂O) and toluene (3.0 mL). Purification by flash column chromatography (100% hexanes) afforded a colorless oil (0.137 g) as a mixture of the title compound (calculated yield 93%) and the product of elimination (5% calculated yield). Further purification (100% heptanes) afforded a sample of analytically pure material: **R_f** = 0.7 (100% hexanes); **¹H NMR** (500 MHz, CDCl₃) δ 7.80–7.76 (m, 3H), 7.55 (s, 1H), 7.45–7.39 (dq, *J* = 5.5, 1.2 Hz, 2H), 7.32 (dd, *J* = 8.5, 1.5 Hz, 1H), 2.84–2.79 (dq, *J* = 9.3, 5.5 Hz, 1H), 1.84 (d, *J* = 12.8 Hz, 1H), 1.85–1.47 (m, 8H), 1.26–1.05 (m, 6H), 0.91–0.86 (m, 5H); **¹³C NMR** (500 MHz, CDCl₃) δ 144.1, 133.7, 132.3, 128.0, 127.73, 127.65, 126.4, 126.2, 125.8, 125.1, 44.9, 42.7, 39.9, 35.0, 34.4, 33.0, 26.8, 26.4, 26.3, 20.9, 14.3; **IR** (thin film) 3052, 2661, 1915, 1633, 1600 cm⁻¹; **HRMS** (TOF MS Cl⁺) *m/z* calcd for C₂₁H₂₈ [M]⁺ 280.2191, found 280.2187; [α]_D²⁹ +17.8 (*c* 1.11, CHCl₃); **SFC** analysis (OJ-H, 2% hexanes, 2.0 mL/min) indicated 97% ee *t_R* (major) = 11.4 minutes, *t_R* (minor) = 12.3 minutes.

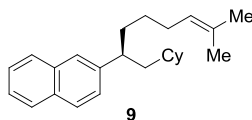


(+)-2-(1-cyclohexylheptan-2-yl)naphthalene (7, table 2, entry 3): Prepared according to the representative procedure outlined above using the following amounts of reagents: (*R*)-**26** (0.134 g, 0.500 mmol) and NiCl₂(dppe) (5.3 mg, 0.010 mmol), *n*-pentylmagnesium bromide (0.50 mL, 1.0 mmol, 2.0 M in Et₂O), and toluene (3.0 mL). Purification by flash column chromatography (100% hexanes) afforded a colorless oil (0.147 g) as a mixture of the title compound (91% calculated yield) and the product of elimination (6% calculated yield). Further purification (100% heptanes) afforded a sample of analytically pure material: **R_f** = 0.3 (100% hexanes); **¹H NMR** (400 MHz, CDCl₃) δ 7.80–7.75 (m, 3H), 7.55 (s, 1H), 7.44–7.37 (m, 2H), 7.31 (dd, *J* = 8.4, 1.2 Hz, 1H), 2.79 (m, 1H), 1.84 (d, *J* = 12.8 Hz, 1H), 1.63–1.44 (m, 8H), 1.21–1.04 (m, 10H), 0.89–0.81 (m, 5H); **¹³C NMR** (500 MHz, CDCl₃) δ 144.2, 133.7, 132.3, 128.0, 127.74, 127.67, 126.4, 126.1, 125.8, 125.1, 45.0, 43.0, 37.6, 35.0, 34.4, 33.0, 32.2, 27.5, 26.8, 26.4, 26.3, 22.7, 14.3; **IR** (neat) 3020, 2920, 2850, 815, 744, 697 cm⁻¹; **HRMS** (TOF MS Cl⁺) *m/z* calcd for C₂₃H₃₂ [M]⁺ 308.2504, found 308.2507; [α]_D²⁷ +9.4 (*c* 1.20, CHCl₃); **SFC** analysis (OJ-H, 2% hexanes, 2.0 mL/min) indicated 97% ee *t_R* (minor) = 11.4 minutes, *t_R* (major) = 12.3 minutes.

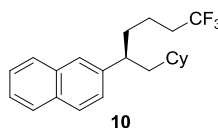


(S)-2-(1-cyclohexyl-5-phenylpentan-2-yl)naphthalene (8, table 2, entry 4): Prepared according to the representative procedure outlined above using the following amounts of reagents: (*R*)-**26** (0.401 g, 1.50 mmol), NiCl₂(dppe) (15.8 mg, 0.030 mmol), (3-phenylpropyl)magnesium bromide (1.59 mL, 1.89 mmol, 1.9 M in Et₂O), and toluene (22 mL). Purification by flash column chromatography (0–5% EtOAc in hexanes) afforded a colorless oil (0.522 g) as a mixture of the title compound (88% calculated yield) and the product of Wurtz coupling of the organomagnesium reagent (1,6-diphenylhexane). Further purification by flash chromatography (100% pentane) afforded a sample of analytically pure material: **TLC R_f** = 0.3 (100% hexanes); **m.p.** 81–81 °C; **¹H NMR** (400 MHz, CDCl₃) δ 7.81 (d, *J* = 8.0 Hz, 1H), 7.77 (d, *J* = 8.5 Hz, 2H), 7.54 (s, 1H), 7.46–7.39 (m, 2H), 7.29 (d, *J* = 8.0 Hz, 1H), 7.22 (m, 2H), 7.13 (t, *J*

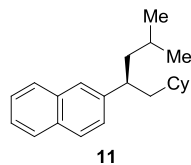
= 7.5 Hz, 1H), 7.08 (d, $J = 7.0$ Hz, 2H), 2.82 (p, $J = 5.0$ Hz, 1H), 2.54 (m, 2H), 1.82 (d, $J = 12.5$ Hz, 1H), 1.69–1.43 (br m, 10H), 1.11–1.00 (m, 4H), 0.85 (m, 2H); ^{13}C NMR (125 MHz, CDCl_3) δ 143.7, 142.7, 133.7, 132.3, 128.5, 128.3, 128.0, 127.73, 127.68, 126.4, 126.1, 125.9, 125.7, 125.1, 44.9, 42.9, 37.2, 36.1, 34.9, 34.4, 32.9, 29.6, 26.8, 26.4, 26.3; **IR** (neat) 2918, 2850, 1442, 822, 745, 696 cm^{-1} ; **HRMS** (TOF MS CI^+) m/z caclcd for $\text{C}_{27}\text{H}_{32}[\text{M}]^+$ 356.2504, found 356.2509; $[\alpha]_{\text{D}}^{20} +1.0$ (c 1.03, CHCl_3); **SFC** analysis (OJ-H, 20% IPA, 2.5 mL/min) indicated 97% ee t_{R} (minor) = 4.2 minutes, t_{R} (major) = 5.2 minutes. Crystals suitable for X-ray diffraction (vide infra) were grown by slow evaporation of solvent from a solution of the title compound in a mixture of methanol and pentane.



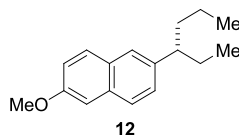
(+)-2-(1-cyclohexyl-7-methyloct-6-en-2-yl)naphthalene (9, table 2, entry 5): Prepared according to the representative procedure outlined above using the following amounts of reagents: (*R*)-**26** (0.0537 g, 0.200 mmol) and $\text{NiCl}_2(\text{dppe})$ (5.3 mg, 0.010 mmol), (5-methylhex-4-enyl)magnesium bromide (0.48 mL, 0.40 mmol, 0.83 M in Et_2O), and toluene (3.0 mL) for 48 h. Purification by flash column chromatography (100% pentane) afforded a colorless oil (0.054 g) as a mixture of the title compound (81% calculated yield) and the product of elimination (12% calculated yield). Further purification by flash chromatography on silver-impregnated silica (0–3% Et_2O in pentane) afforded a sample of analytically pure material: **TLC** R_{f} = 0.4 (100% pentane); ^1H NMR (400 MHz, CDCl_3) δ 7.81–7.76 (m, 3H), 7.55 (s, 1H), 7.46–7.39 (dp, $J = 8.4, 1.2$ Hz, 2H), 7.31 (dd, $J = 8.5, 1.5$ Hz, 1H), 5.01 (t, $J = 7.1$ Hz, 1H), 2.80 (dq, $J = 14.8, 5.3$ Hz, 1H), 1.96–1.82 (m, 3H), 1.68–1.47 (m, 14H), 1.36–1.18 (m, 2H), 1.16–1.01 (m, 4H), 0.94–0.80 (m, 2H); ^{13}C NMR (500 MHz, CDCl_3) δ 144.0, 133.7, 132.3, 131.3, 128.0, 127.7, 127.6, 126.4, 126.1, 125.8, 125.1, 124.9, 44.9, 42.9, 37.2, 35.0, 34.4, 32.9, 28.2, 28.0, 26.8, 26.4, 26.3, 25.8, 17.8; **IR** (neat) 2919, 2850, 1447, 853, 815, 743 cm^{-1} ; **HRMS** (TOF MS EI^+) m/z caclcd for $\text{C}_{25}\text{H}_{34}[\text{M}]^+$ 334.2661, found 334.2660; $[\alpha]_{\text{D}}^{24} +11.8$ (c 5.33, CHCl_3); **SFC** analysis (OJ-H, 3% IPA, 2.5 mL/min) indicated 97% ee t_{R} (minor) = 6.5 minutes, t_{R} (major) = 6.9 minutes.



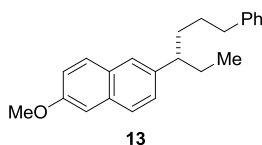
(+)-2-(1-cyclohexyl-6,6,6-trifluorohexan-2-yl)naphthalene (10, table 2, entry 6): Prepared according to the representative procedure outlined above using the following amounts of reagents: (*R*)-**26** (0.0537 g, 0.200 mmol) and $\text{NiCl}_2(\text{dppe})$ (10.6 mg, 0.0200 mmol), (4,4,4-trifluorobutyl)magnesium bromide (0.20 mL, 0.40 mmol, 2.0 M in Et_2O), and toluene (3.0 mL) for a period of 48 h. Purification by flash column chromatography (100% pentane) afforded a colorless oil (0.054 g) as a mixture of the title compound (67% calculated yield) and the product of elimination (15% calculated yield). Further purification by flash chromatography on silver-impregnated silica (100% pentane) afforded a sample of analytically pure material: **TLC** R_{f} = 0.4 (100% pentane); ^1H NMR (400 MHz, CDCl_3) δ 7.82–7.78 (m, 3H), 7.55 (s, 1H), 7.48–7.40 (dp, $J = 6.9, 1.3$ Hz, 2H), 7.29 (dd, $J = 8.5, 1.5$ Hz, 1H), 2.81 (dq, $J = 14.8, 4.9$ Hz, 1H), 2.11–1.90 (m, 2H), 1.83 (d, $J = 12.9$ Hz, 1H), 1.75–1.25 (m, 10H), 1.15–1.01 (m, 4H), 0.93–0.81 (m, 2H); ^{13}C NMR (500 MHz, CDCl_3) δ 142.9, 133.7, 132.4, 128.3, 127.8, 127.7, 126.4, 126.0, 125.7, 125.3, 44.8, 42.8, 36.5, 34.9, 34.3, 33.9 (q, $J = 28.3$ Hz, 1C), 32.9, 26.7, 26.7, 26.3, 26.2, 20.3 (q, $J = 2.8$ Hz, 1C); **IR** (neat) 2920, 2850, 1448, 1253, 1131, 816, 744 cm^{-1} ; **HRMS** (TOF MS EI^+) m/z caclcd for $\text{C}_{22}\text{H}_{27}\text{F}_3[\text{M}]^+$ 348.2065, found 348.2065; $[\alpha]_{\text{D}}^{26} +19.7$ (c 1.2, CHCl_3); **SFC** analysis (OJ-H, 10% hexanes, 2.5 mL/min) indicated 97% ee t_{R} (major) = 4.7 minutes, t_{R} (minor) = 5.3 minutes.



(+)-2-(1-cyclohexyl-4-methylpentan-2-yl)naphthalene (11, table 2, entry 7): Prepared according to the representative procedure outlined above using the following amounts of reagents: (*S*)-**26** (0.134 g, 0.500 mmol) and NiCl₂(dppe) (5.3 mg, 0.010 mmol), *i*-butylmagnesium bromide (0.40 mL, 1.0 mmol, 2.5 M in Et₂O), and toluene (3.0 mL). Purification by flash column chromatography (100% hexanes) afforded a colorless oil (0.067 g) as a mixture of the title compound (40% calculated yield), the product of reduction (8% calculated yield), and the product of elimination (7% calculated yield). Further purification (100% heptanes) afforded a sample of analytically pure material: **TLC** R_f = 0.7 (100% hexanes); **¹H NMR** (500 MHz, CDCl₃) δ 7.79 (q, *J* = 7.5 Hz, 3H), 7.56 (s, 1H), 7.43 (m, 2H), 7.32 (d, *J* = 9.0 Hz, 1H), 2.92 (hept, *J* = 5.0 Hz, 1H), 1.85 (d, *J* = 12.5 Hz, 1H), 1.66–1.54 (m, 6H), 1.49–1.39 (m, 2H), 1.33 (m, 1H), 1.14–1.03 (m, 4H), 0.92–0.84 (m, 5H), 0.80 (d, *J* = 6.5 Hz, 3H); **¹³C NMR** (125 MHz, CDCl₃) δ 144.2, 133.7, 132.3, 128.0, 127.7, 127.6, 126.3, 126.1, 125.8, 125.1, 46.9, 45.4, 40.5, 34.9, 34.3, 33.0, 26.8, 26.4, 26.3, 25.5, 23.7, 22.0; **IR** (neat) 2919, 2849, 1447, 853, 813, 743 cm⁻¹; **HRMS** (TOF MS EI⁺) *m/z* calcd for C₂₂H₃₀ [M]⁺ 294.2347, found 294.2350; **[α]_D²⁷** +14.9 (*c* 0.80, CHCl₃); **SFC** analysis (OJ-H, 2% hexanes, 2.0 mL/min) indicated 90% ee: t_R (major) = 7.0 minutes, t_R (minor) = 8.4 minutes.

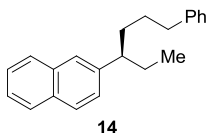


(-)-2-(hexan-3-yl)-6-methoxynaphthalene (12, table 2, entry 8): Prepared according to the representative procedure outlined above using the following amounts of reagents: (*S*)-**SI-7** (0.115 g, 0.500 mmol) and NiCl₂(dppe) (5.3 mg, 0.010 mmol), *n*-propylmagnesium bromide (0.44 mL, 1.0 mmol, 2.3 M in Et₂O), and toluene (3.0 mL). Purification by flash column chromatography (3% EtOAc in hexanes) afforded a white solid (0.108 g) as a mixture of the title compound (80% calculated yield), the product of reduction (6% calculated yield), and the product of elimination (5% calculated yield). Further purification (100% hexanes) afforded a sample of analytically pure material: **TLC** R_f = 0.4 (5% EtOAc in hexanes); **m.p.** 57–58 °C; **¹H NMR** (400 MHz, CDCl₃) δ 7.68 (m, 2H), 7.49 (s, 1H), 7.28 (m, 1H), 7.12 (dd, *J* = 6.0, 2.8 Hz, 2H), 3.91 (s, 3H), 2.54 (m, 1H), 1.73–1.60 (m, 4H), 1.21–1.14 (m, 2H), 0.84 (t, *J* = 7.2, 3H), 0.78 (t, *J* = 7.4, 3H); **¹³C NMR** (125 MHz, CDCl₃) δ 157.2, 141.3, 133.2, 129.1 (2C), 126.8, 126.7, 126.4, 118.6, 105.8, 55.5, 47.7, 39.0, 29.8, 20.9, 14.3, 12.4; **IR** (neat) 2953, 2920, 2856, 816, 744, 697 cm⁻¹; **HRMS** (TOF MS CI⁺) *m/z* calcd for C₁₇H₂₂O [M+H]⁺ 243.1749, found 243.1741; **[α]_D²⁸** -8.2 (*c* 0.97, CHCl₃); **SFC** analysis (OD-H, 10% hexanes, 3.5 mL/min) indicated 92% ee: t_R (minor) = 4.4 minutes, t_R (major) = 4.6 minutes.

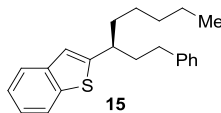


(+)-2-methoxy-6-(6-phenylhexan-3-yl)naphthalene (13, table 2, entry 9): Prepared according to the representative procedure outlined above using the following amounts of reagents: (*S*)-**SI-7** (0.115 g, 0.500 mmol) and NiCl₂(dppe) (5.3 mg, 0.010 mmol), (3-phenylpropyl)magnesium bromide (0.50 mL, 1.0 mmol, 2.0 M in Et₂O), and toluene (3.0 mL). Purification by flash column chromatography (3% Et₂O in heptanes) afforded a colorless oil (0.145 g) as a mixture of the title compound (88% calculated yield), the

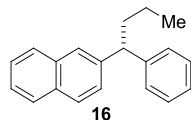
product of reduction (4% calculated yield), and the product of elimination (1% calculated yield). Further purification (preparatory TLC, 2% Et₂O in heptanes) afforded a sample of analytically pure material: **TLC** R_f = 0.4 (5% EtOAc in hexanes); **¹H NMR** (500 MHz, CDCl₃) δ 7.67 (d, J = 8.5 Hz, 2H), 7.47 (s, 1H), 7.22 (m, 3H), 7.14–7.07 (m, 5H), 3.90 (s, 3H), 2.61–2.49 (m, 3H), 1.74–1.57 (m, 4H), 1.52–1.46 (m, 2H), 0.76 (t, J = 7.3 Hz, 3H); **¹³C NMR** (125 MHz, CDCl₃) δ 157.2, 142.8, 140.9, 133.3, 129.1 (2C), 128.5, 128.3, 126.9, 126.6, 126.4, 125.7, 118.6, 105.7, 55.4, 47.9, 36.3, 36.1, 29.9, 29.6, 12.4; **IR** (neat) 3025, 2930, 2856, 1605, 1264, 1032, 850 cm⁻¹; **HRMS** (TOF MS CI⁺) m/z cacl'd for C₂₃H₂₆O [M]⁺ 318.1984, found 318.1994; $[\alpha]_D^{26}$ +12.1 (c 0.50, CHCl₃); **SFC** analysis (OJ-H, 15% IPA, 2.0 mL/min) indicated 93% ee: t_R (minor) = 6.9 minutes, t_R (major) = 8.5 minutes.



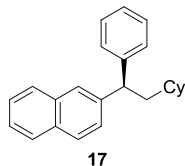
(-)-2-(6-phenylhexan-3-yl)naphthalene (14, table 2, entry 10): Prepared according to the representative procedure outlined above using the following amounts of reagents: (*R*)-**1** (0.040 g, 0.20 mmol) and NiCl₂(dppe) (5.3 mg, 0.010 mmol), (3-phenylpropyl)magnesium bromide (0.24 mL, 0.40 mmol, 1.7 M in Et₂O), and toluene (3.0 mL). Purification by flash column chromatography (100% heptanes) afforded a colorless oil (0.054 g) as a mixture of the title compound (93% calculated yield) and the product of Wurtz coupling of the organomagnesium reagent (1,6-diphenylhexane). Further purification (flash column chromatography in 100% pentanes) afforded a sample of analytically pure material: **TLC** R_f = 0.6 (100% hexanes); **¹H NMR** (500 MHz, CDCl₃) δ 7.79 (d, J = 8.1 Hz, 1H), 7.77 (d, J = 8.4 Hz, 2H), 7.54 (s, 1H), 7.42 (dt, J = 17.4, 6.7 Hz, 2H), 7.28 (d, J = 8.3 Hz, 1H), 7.22 (m, 2H), 7.12 (m, 1H), 7.08 (d, J = 7.3 Hz, 2H), 2.61–2.49 (m, 3H), 1.79–1.59 (m, 4H), 1.57–1.41 (m, 2H), 0.77 (t, J = 7.3 Hz, 3H) **¹³C NMR** (125 MHz, CDCl₃) δ 143.3, 142.7, 133.7, 132.4, 128.5, 128.5, 128.3, 128.3, 128.0, 127.7, 127.7, 126.6, 126.1, 125.9, 125.7, 125.2, 48.1, 36.2, 36.2, 29.8, 29.6, 12.4; **IR** (neat) 3024, 2928, 2856, 815, 743 cm⁻¹; **HRMS** (TOF MS CI⁺) m/z cacl'd for C₂₂H₂₄ [M]⁺ 288.1878, found 288.1880; $[\alpha]_D^{24}$ -15.1 (c 1.08, CHCl₃); **SFC** analysis (OJ-H, 15% IPA, 2.0 mL/min) indicated 97% ee: t_R (minor) = 6.3 minutes, t_R (major) = 7.8 minutes.



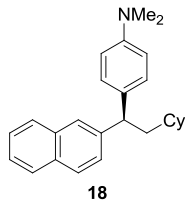
(+)-2-(1-phenyloctan-3-yl)benzo[*b*]thiophene (15, table 2, entry 11): A 7 mL vial was charged with (*R*)-**SI-10** (0.056 g, 0.200 mmol) and NiCl₂(dppe) (5.3 mg, 0.010 mmol), flushed with nitrogen and capped. Toluene (3 mL) was added, followed by *n*-pentylmagnesium bromide (0.18 mL, 0.40 mmol, 2.3 M in Et₂O) and the reaction was allowed to stir at room temperature for 10 h. The vial was cracked open and another batch of NiCl₂(dppe) (5.3 mg, 0.010 mmol) was added. After further stirring for 14 h, the reaction was quenched by the addition of EtOAc (1 mL) and the entire reaction mixture was adsorbed onto silica gel (1 g). The solvents were removed in vacuo and the crude was purified by flash column chromatography (100% hexanes) to afford **15** as a colorless oil (0.035 g, 0.11 mmol, 54%): **TLC** R_f = 0.4 (100% hexanes); **¹H NMR** (500 MHz, CDCl₃) δ 7.79 (d, J = 8.0 Hz, 1H), 7.69 (d, J = 7.5 Hz, 1H), 7.32 (t, J = 7.3 Hz, 1H), 7.26 (m, 3H), 7.18 (d, J = 7.5 Hz, 1H), 7.14 (d, J = 8.0 Hz, 2H), 7.03 (s, 1H), 2.93 (hept, J = 5.0 Hz, 1H), 2.60 (m, 1H), 2.54 (m, 1H), 2.03 (m, 1H), 1.94 (m, 1H), 1.55–1.70 (m, 2H), 1.26–1.13 (m, 6H), 0.82 (t, J = 6.5 Hz, 3H); **¹³C NMR** (125 MHz, CDCl₃) δ 151.2, 142.3, 140.1, 139.2, 128.6, 128.5, 125.9, 124.1, 123.5, 122.9, 122.5, 120.9, 42.0, 39.4, 37.8, 33.8, 31.9, 27.2, 22.7, 14.2; **IR** (neat) 3060, 2925, 2854, 1455, 1436, 821 cm⁻¹; **HRMS** (TOF MS CI⁺) m/z cacl'd for C₂₂H₂₆S [M+H]⁺ 323.1833, found 323.1833; $[\alpha]_D^{26}$ +13.4 (c 0.95, CHCl₃); **SFC** analysis (OD-H, 3% IPA, 2.5 mL/min) indicated 96% ee: t_R (major) = 20.0 minutes, t_R (minor) = 21.4 minutes.



(–)-2-(1-phenylbutyl)naphthalene (**16**, table 2, entry 12): A 7 mL vial was charged with (*S*)-**SI-12** (0.124 g, 0.500 mmol) and NiCl₂(dppe) (5.3 mg, 0.010 mmol), flushed with nitrogen and capped. Toluene (3 mL) was added and the reaction was cooled to 0 °C. *n*-Propylmagnesium bromide (0.18 mL, 0.40 mmol, 2.3 M in Et₂O) was added and the reaction was allowed to stir at 0 °C for 48 h. The reaction was quenched by the addition of EtOAc (1 mL) and the entire reaction mixture was adsorbed onto silica gel (1 g). The solvents were removed in vacuo and the crude was purified by flash column chromatography (100% hexanes). The resulting colorless oil (0.104 g) was an inseparable mixture of the title compound (67% calculated yield) and the product of reduction²⁵ (15% calculated yield). Purification by preparatory HPLC (100 hexanes) afforded a small sample of analytically pure **16**: TLC R_f = 0.5 (100% hexanes); ¹H NMR (400 MHz, CDCl₃) δ 7.78 (t, *J* = 8.0 Hz, 2H), 7.73 (d, *J* = 8.4 Hz, 1H), 7.70 (s, 1H), 7.42 (m, 2H), 7.34 (d, *J* = 8.4 Hz, 1H), 7.28 (m, 4H), 7.17 (m, 1H), 4.02 (t, *J* = 8.2 Hz, 1H), 2.13 (hept, *J* = 5.6 Hz, 2H), 1.32 (sextet, *J* = 7.6 Hz, 2H), 0.95 (t, *J* = 7.2 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 145.3, 142.9, 133.7, 132.2, 128.5, 128.13, 128.12, 127.8, 127.7, 127.0, 126.2, 126.02, 125.99, 125.4, 51.2, 37.8, 21.3, 14.3; IR (neat) 3055, 3024, 2954, 2928, 1599, 1506, 1493 cm⁻¹; HRMS (TOF MS CI⁺) *m/z* calcd for C₂₀H₂₀ [M]⁺ 260.1565, found 260.1567; [α]_D²⁴ –25.2 (*c* 0.54, CHCl₃); SFC analysis (OD-H, 2% IPA, 2.5 mL/min) indicated 91% ee: t_R (minor) = 5.9 minutes, t_R (major) = 7.5 minutes.

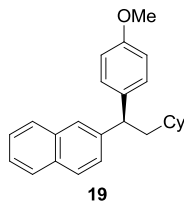


(–)-2-(2-cyclohexyl-1-phenylethyl)naphthalene (**17**, table 3, entry 1): Prepared according to the representative procedure outlined above using the following amounts of reagents: (*R*)-**26** (0.134 g, 0.500 mmol) and NiCl₂(dppe) (5.3 mg, 0.010 mmol), phenylmagnesium bromide (0.37 mL, 1.0 mmol, 2.7 M in Et₂O), and toluene (7.5 mL) for a period of 24 h. Purification by flash column chromatography (100% pentane) afforded the title compound as a colorless oil (0.105 g, 0.33 mmol, 67%): TLC R_f = 0.2 (100% pentane); ¹H NMR (400 MHz, CDCl₃) δ 7.77 (t, *J* = 8.1 Hz, 2H), 7.72 (d, *J* = 8.5 Hz, 1H), 7.69 (s, 1H), 7.41 (quintd, *J* = 6.9, 1.3 Hz, 2H), 7.34 (dd, *J* = 8.5, 1.6 Hz, 1H), 7.29–7.24 (m, 4H), 7.15 (m, 1H), 4.23 (t, *J* = 7.9 Hz, 1H), 2.08–1.95 (m, 2H), 1.80 (t, *J* = 11.7 Hz, 2H), 1.66–1.58 (m, 3H), 1.26–1.05 (m, 4H), 1.01–0.93 (m, 2H); ¹³C NMR (100 MHz, CDCl₃) δ 145.5, 143.0, 133.7, 132.3, 128.5, 128.1, 127.9, 127.7, 127.0, 126.2, 126.0, 125.4, 48.2, 43.5, 35.0, 33.7, 33.5, 26.8, 26.3 IR (neat) 2919, 2849, 1448, 3055, 1490, 743, 698 cm⁻¹; HRMS (TOF MS ES⁺) *m/z* calcd for C₂₄H₂₆ [M]⁺ 314.2035, found 314.2030; [α]_D²⁵ –10.8 (*c* 1.99, CHCl₃); SFC analysis (OJ-H, 15% IPA, 2.5 mL/min) indicated 92% ee: t_R (major) = 6.1 minutes, t_R (minor) = 6.9 minutes.

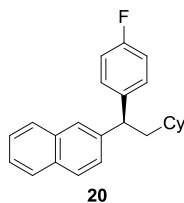


(–)-4-(2-cyclohexyl-1-(naphthalen-2-yl)ethyl)-*N,N*-dimethylaniline (**18**, table 3, entry 2): Prepared according to the representative procedure outlined above using the following amounts of reagents: (*R*)-**26**

(0.054 g, 0.20 mmol) and NiCl₂(dppe) (5.3 mg, 0.010 mmol), 4-(*N,N*-dimethylamino) phenylmagnesium bromide (0.133 g, 0.40 mmol), and toluene (3.0 mL) for a period of 48 h. Purification by flash column chromatography (1% Et₃N, 5% Et₂O in pentane) afforded the title compound as a yellow oil (0.057 g, 0.16 mmol, 79%): **TLC** *R_f* = 0.3 (1% Et₃N, 5% Et₂O in pentane); **¹H NMR** (400 MHz, CDCl₃) δ 7.67 (t, *J* = 8.5 Hz, 2H), 7.71 (d, *J* = 8.4 Hz, 1H), 7.67 (s, 1H), 7.44–7.32 (m, 3H), 7.13 (d, *J* = 8.6 Hz, 2H), 6.66 (d, *J* = 8.7 Hz), 4.14 (t, *J* = 7.9 Hz), 2.88 (s, 6H), 2.04–1.91 (m, 2H), 1.80 (t, *J* = 12.6, 2H), 1.66 (m, 3H), 1.25–1.05 (m, 4H), 1.01–0.92 (m, 2H); **¹³C NMR** (125 MHz, CDCl₃) δ 149.1, 143.9, 133.7, 133.5, 132.1, 128.7, 128.0, 127.8, 127.7, 127.1, 125.8, 125.7, 125.2, 112.9, 47.1, 43.7, 40.9, 35.0, 33.7, 33.6, 26.8, 26.3; **IR** (neat) 2918, 2848, 1614, 1519, 1446, 1345, 813, 731 cm⁻¹; **HRMS** (TOF MS Cl⁺) *m/z* calcd for C₂₆H₃₁N [M+H]⁺ 358.2535, found 358.2529; [α]_D²⁶ -1.55 (*c* 2.59, CHCl₃).

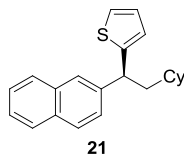


(-)-2-(2-cyclohexyl-1-(4-methoxyphenyl)ethyl)naphthalene (**19**, table 3, entry 3): Prepared according to the representative procedure outlined above using the following amounts of reagents: (*R*)-**26** (0.134 g, 0.500 mmol) and NiCl₂(dppe) (5.3 mg, 0.010 mmol), 4-methoxyphenylmagnesium bromide (0.53 mL, 1.0 mmol, 1.9 M in Et₂O), and toluene (7.5 mL) for a period of 24 h. Purification by flash column chromatography (5% Et₂O in pentane) afforded a colorless oil (0.157 g) as a mixture of the title compound (86% calculated yield) and the starting material (15% calculated yield). Further purification by column chromatography (1–15% benzene in hexanes) afforded a pure sample of **17**: **TLC** *R_f* = 0.7 (5% EtOAc in hexanes); **¹H NMR** (500 MHz, CDCl₃) δ 7.77 (t, *J* = 9.3 Hz, 2H), 7.72 (d, *J* = 8.5 Hz, 1H), 7.66 (s, 1H), 7.41 (m, 2H), 7.32 (d, *J* = 8.5 Hz, 1H), 7.18 (d, *J* = 8.5 Hz, 2H), 6.83 (d, *J* = 9.0 Hz, 2H), 4.18 (t, *J* = 7.3 Hz, 1H), 3.74 (s, 3H), 1.98 (m, 2H), 1.79 (m, 2H), 1.70–1.55 (m, 3H), 1.23–1.04 (m, 4H), 1.02–0.91 (m, 2H); **¹³C NMR** (125 MHz, CDCl₃) δ 157.9, 143.4, 137.6, 133.7, 132.2, 129.0, 128.5, 128.1, 127.8, 127.7, 127.0, 126.0, 125.8, 125.4, 113.9, 55.3, 47.2, 43.7, 35.0, 33.62, 33.57, 26.8, 26.3; **IR** (neat) 2919, 2848, 1509, 1447, 1245, 1177, 1037 cm⁻¹; **HRMS** (TOF MS Cl⁺) *m/z* calcd for C₂₅H₂₈O [M]⁺ 344.2140, found 344.2131; [α]_D²⁴ -6.3 (*c* 1.17, CHCl₃); **SFC** analysis (AD-H, 15% IPA, 3.0 mL/min) indicated 97% ee: *t_R* (major) = 9.0 minutes, *t_R* (minor) = 9.6 minutes.

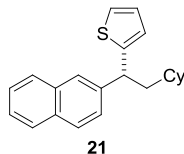


(-)-2-(2-cyclohexyl-1-(4-fluorophenyl)ethyl)naphthalene (**20**, table 3, entry 4): Prepared according to the representative procedure outlined above using the following amounts of reagents: (*R*)-**26** (0.134 g, 0.500 mmol) and NiCl₂(dppe) (5.3 mg, 0.010 mmol), 4-fluorophenylmagnesium bromide (0.46 mL, 1.0 mmol, 2.2 M in Et₂O), and toluene (7.5 mL) for a period of 48 h. Purification by flash column chromatography (100% hexanes) afforded a colorless oil (0.147 g) as a mixture of the title compound (82% calculated yield) and the product of Wurtz coupling of the organomagnesium reagent. Further purification by column chromatography (100% pentane) afforded a pure sample of **18**: **TLC** *R_f* = 0.8 (5% EtOAc in hexanes); **¹H NMR** (500 MHz, CDCl₃) δ 7.78 (t, *J* = 7.5 Hz, 2H), 7.74 (d, *J* = 8.5 Hz, 1H), 7.66 (s, 1H), 7.43 (m, 2H), 7.31 (d, *J* = 8.5 Hz, 1H), 7.22 (dd, *J* = 8.5, 6.0 Hz, 2H), 6.95 (app t, *J* = 8.5 Hz, 2H), 4.21 (t, *J* = 8.0 Hz, 1H), 2.02 (quint, *J* = 7.1 Hz, 1H), 1.94 (quint, *J* = 7.0 Hz, 1H), 1.79 (t, *J* = 12.3

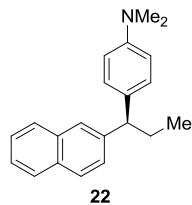
Hz, 2H), 1.70–1.56 (m, 3H), 1.22–1.04 (m, 4H), 0.99 (m, 2H); ^{13}C NMR (150 MHz, CDCl_3) δ 161.4 (d, J = 242.5 Hz, 1C), 142.8, 141.1 (d, J = 3.3 Hz, 1C), 133.7, 132.2, 129.5 (d, J = 7.8 Hz, 2C), 128.2, 127.8, 127.7, 126.8, 126.1, 125.9, 125.5, 115.3 (d, J = 84.0 Hz, 2C), 47.3, 43.6, 35.0, 33.6, 33.5, 26.7, 26.2 (2C); IR (neat) 3053, 2920, 2850, 1506, 1222, 793 cm^{-1} ; HRMS (TOF MS Cl^+) m/z calcd for $\text{C}_{24}\text{H}_{25}\text{F}$ $[\text{M}]^+$ 332.1940, found 332.1930; $[\alpha]_D^{20}$ -11.1 (c 0.91, CHCl_3); SFC analysis (OJ-H, 15% IPA, 2.5 mL/min) indicated 87% ee: t_R (major) = 4.3 minutes, t_R (minor) = 5.6 minutes.



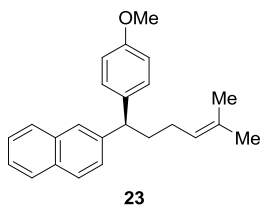
(+)-2-(2-cyclohexyl-1-(naphthalen-2-yl)ethyl)thiophene ((+)-21, table 3, entry 5): Prepared according to the representative procedure outlined above using the following amounts of reagents: (*R*)-**26** (0.054 g, 0.20 mmol) and $\text{NiCl}_2(\text{dppe})$ (10.6 mg, 0.0200 mmol), 2-thienylmagnesium bromide (0.15 mL, 0.40 mmol, 2.6 M in Et_2O), and toluene (3.0 mL) for a period of 48 h. Purification by flash column chromatography (2% Et_2O in pentane) afforded the title compound as a yellow oil (0.049 g, 0.15 mmol, 76%): TLC R_f = 0.5 (100% hexanes); ^1H NMR (400 MHz, CDCl_3) δ 7.78 (t, J = 9.0 Hz, 3H), 7.70 (s, 1H), 7.42 (m, 3H), 7.12 (d, J = 4.4 Hz, 1H), 6.91 (dd, J = 5.2, 3.2 Hz, 1H), 6.83 (d, J = 3.2 Hz, 1H), 4.45 (t, J = 8.0 Hz, 1H), 2.05 (t, J = 7.4 Hz, 2H), 1.85 (d, J = 12.4 Hz, 1H), 1.73 (d, J = 12.4 Hz, 1H), 1.64–1.50 (m, 3H), 1.30–0.96 (m, 6H); ^{13}C NMR (150 MHz, CDCl_3) δ 150.2, 142.5, 133.7, 132.5, 128.4, 127.9, 127.8, 126.7, 126.3, 126.2, 126.1, 125.6, 123.9, 123.6, 45.1, 43.9, 35.1, 33.8, 33.1, 26.7, 26.3, 26.2; IR (neat) 3026, 2922, 2853, 1453, 743, 697 cm^{-1} ; HRMS (TOF MS Cl^+) m/z calcd for $\text{C}_{22}\text{H}_{24}\text{S}$ $[\text{M}+\text{H}]^+$ 321.1677, found 321.1672; $[\alpha]_D^{24}$ $+42.4$ (c 1.20, CHCl_3); SFC analysis (OD-H, 20% hexanes, 3.0 mL/min) indicated 93% ee: t_R (minor) = 14.9 minutes, t_R (major) = 17.5 minutes.



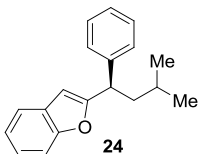
(-)-2-(2-cyclohexyl-1-(naphthalen-2-yl)ethyl)thiophene ((-)-21). Prepared according to the representative procedure outlined above using the following amounts of reagents: (*S*)-**26** (0.039 g, 0.15 mmol) and $\text{NiCl}_2(\text{dppe})$ (7.7 mg, 0.015 mmol), 2-thienylmagnesium bromide (0.17 mL, 0.29 mmol, 2.0 M in Et_2O), and toluene (2.2 mL) for a period of 48 h. Purification by flash column chromatography (2% Et_2O in pentane) afforded the title compound as a yellow oil (0.033 g, 0.10 mmol, 70%): ^1H NMR (500 MHz, CDCl_3) δ 7.78 (t, J = 8.6 Hz, 3H), 7.70 (s, 1H), 7.42 (m, 3H), 7.13 (d, J = 5.2 Hz, 1H), 6.91 (dd, J = 5.2, 3.6 Hz, 1H), 6.84 (d, J = 3.6 Hz, 1H), 4.45 (t, J = 8.0 Hz, 1H), 2.05 (t, J = 7.4 Hz, 2H), 1.86 (d, J = 12.8 Hz, 1H), 1.73 (d, J = 12.4 Hz, 1H), 1.64–1.54 (m, 3H), 1.26–0.96 (m, 6H); ^{13}C NMR (125 MHz, CDCl_3) δ 150.2, 142.5, 133.7, 132.5, 128.4, 127.9, 127.8, 126.7, 126.3, 126.14, 126.09, 125.6, 123.9, 123.6, 45.1, 43.8, 35.1, 33.8, 33.1, 26.7, 26.25, 26.21; $[\alpha]_D^{24}$ -42.9 (c 0.98, CHCl_3); SFC analysis (OD-H, 20% hexanes, 3.0 mL/min) indicated 92% ee: t_R (major) = 14.3 minutes, t_R (minor) = 17.5 minutes.



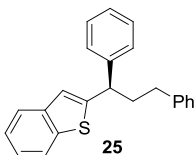
(–)-*N,N*-dimethyl-4-(1-(naphthalen-2-yl)propyl)aniline (**22**, table 3, entry 7): A 7 mL vial was equipped with a stir bar, flame dried, and pumped into a glove box while still warm. (*R*)-**1** (0.040 g, 0.20 mmol), NiCl₂(dppe) (5.3 mg, 0.010 mmol), and 4-(*N,N*-dimethylamino) phenylmagnesium bromide (0.133 g, 0.400 mmol) were added. The vial was capped, removed from the glove box and put under a nitrogen atmosphere. Toluene (3 mL) was added and the reaction mixture was stirred for 24 h at which point the reaction was quenched with methanol. The crude mixture was eluted through a silica plug (100% Et₂O) and concentrated in vacuo. Purification by flash column chromatography (1% Et₃N, 5% Et₂O in pentane) afforded the title compound as a yellow oil (0.047 g, 0.16 mmol, 80% yield). Trace amounts (<5%) of Wurtz coupling product of the organomagnesium reagent could not be separated from the product: **TLC** R_f = 0.2 (1% Et₃N, 5% EtOAc in hexanes) **¹H NMR** (400 MHz, CDCl₃) δ 7.77 (t, *J* = 8.0 Hz, 2H), 7.71 (d, *J* = 8.4 Hz, 1H), 7.68 (s, 1H), 7.40 (quint, *J* = 8.0 Hz, 2H), 7.33 (d, *J* = 8.4 Hz, 1H), 7.14 (d, *J* = 8.4 Hz, 2H), 6.75 (d, *J* = 8.4 Hz, 2H), 3.87 (t, *J* = 7.6 Hz, 1H), 2.89 (s, 6H), 2.17–2.08 (quint, *J* = 7.2 Hz, 2H), 0.93 (t, *J* = 7.4 Hz, 3H); **¹³C NMR** (125 MHz, CDCl₃) δ 149.2, 143.6, 133.7, 133.3, 132.2, 128.7, 128.0, 127.8, 127.7, 127.1, 125.9, 125.8, 125.2, 112.9, 52.4, 40.9, 28.6, 13.0; [α]_D²⁶ –3.4 (*c* 1.17, CHCl₃); **IR** (neat) 2958, 2928, 2871, 2797, 1613, 1564, 1818, 1345, 811, 780, 747; **HRMS** (TOF MS ES⁺) *m/z* calcd for C₂₁H₂₃N [M+H]⁺ 290.1909, found 290.1903; [α]_D²⁵ –3.4 (*c* 1.16, CHCl₃) **SFC** analysis (AD-H, 20% IPA, 2.5 mL/min) indicated 95% ee: t_R (major) = 6.1 minutes, t_R (minor) = 7.4 minutes.



(–)-2-(1-(4-methoxyphenyl)-5-methylhex-4-enyl)naphthalene (**23**, table 3, entry 8): Prepared according to the representative procedure outlined above using the following amounts of reagents: (*R*)-**SI-15** (0.071 g, 0.28 mmol) and NiCl₂(dppe) (5.3 mg, 0.010 mmol), (4-methoxy)phenylmagnesium bromide (0.32 mL, 0.56 mmol, 1.8 M in Et₂O), and toluene (4.2 mL) for a period of 48 h. Purification by flash column chromatography (2% Et₂O in pentane) afforded an oil (0.091 g) as a mixture of the title compound (94% calculated yield) and the product of Wurtz coupling of the organomagnesium reagent. Further purification by column chromatography (1% Et₂O in pentane) afforded a pure sample of **21**: **TLC** R_f = 0.7 (2% EtOAc in hexanes); **¹H NMR** (500 MHz, CDCl₃) δ 7.78 (t, *J* = 8.3 Hz, 2H), 7.72 (d, *J* = 8.0 Hz, 1H), 7.68 (s, 1H), 7.41 (m, 2H), 7.31 (d, *J* = 8.5 Hz, 1H), 7.19 (d, *J* = 9.0 Hz, 2H), 6.81 (d, *J* = 9.0 Hz, 2H), 5.17 (t, *J* = 7.0 Hz, 1H), 4.02 (t, *J* = 7.8 Hz, 1H), 3.75 (s, 3H), 2.14 (m, 2H), 1.97 (app q, *J* = 7.5 Hz, 2H), 1.69 (s, 3H), 1.48 (s, 3H); **¹³C NMR** (125 MHz, CDCl₃) δ 158.0, 143.1, 137.3, 133.7, 132.22, 132.16, 129.1, 128.1, 127.8, 127.7, 126.9, 126.0, 125.9, 125.4, 124.3, 113.9, 55.3, 49.9, 35.8, 26.5, 25.9, 17.9; **IR** (neat) 2925, 1608, 1509, 1440, 1245 cm⁻¹; **HRMS** (TOF MS Cl⁺) *m/z* calcd for C₂₄H₂₆O [M]⁺ 330.1984, found 330.1986; [α]_D²⁰ –2.7 (*c* 1.06, CHCl₃); **SFC** analysis (AD-H, 10% IPA, 3.0 mL/min) indicated 78% ee: t_R (major) = 8.5 minutes, t_R (minor) = 9.2 minutes.



(–)-2-(3-methyl-1-phenylbutyl)benzofuran (**24**, table 3, entry 9): Prepared according to the representative procedure outlined above using the following amounts of reagents: (*R*)-**SI-18** (0.044 g, 0.20 mmol) and NiCl₂(dppe) (10.6 mg, 0.020 mmol), phenylmagnesium bromide (0.13 mL, 0.40 mmol, 3.1 M in Et₂O), and toluene (3.0 mL) for a period of 48 h. Purification by flash column chromatography (100% hexanes) afforded the title compound as a colorless oil (0.037 g, 0.14 mmol, 71%): **TLC** *R*_f = 0.6 (5% EtOAc in hexanes); **¹H NMR** (500 MHz, CDCl₃) δ 7.47 (d, *J* = 7.0 Hz, 1H), 7.39 (d, *J* = 7.5 Hz, 1H), 7.30 (m, 4H), 7.23–7.14 (m, 3H), 6.43 (s, 1H), 4.16 (t, *J* = 8.0 Hz, 1H) 2.08 (m, 1H), 1.89 (m, 1H), 1.51 (m, 1H), 0.95 (d, *J* = 6.5 Hz, 3H), 0.93 (d, *J* = 6.5 Hz, 3H); **¹³C NMR** (125 MHz, CDCl₃) δ 161.6, 154.9, 142.3, 128.8, 128.7, 128.1, 126.8, 123.4, 122.6, 120.5, 111.1, 102.4, 43.7, 43.6, 25.6, 23.0, 22.4; **IR** (neat) 3028, 2954, 2926, 2867, 1453, 1253 cm⁻¹; **HRMS** (TOF MS CI⁺) *m/z* cacl'd for C₁₉H₂₀O [M+H]⁺ 265.1592, found 265.1594; [α]_D²⁴ –45.5 (*c* 1.05, CHCl₃); **SFC** analysis (OD-H, 2% IPA, 3.0 mL/min) indicated 95% ee: *t*_R (minor) = 5.2 minutes, *t*_R (major) = 5.7 minutes.

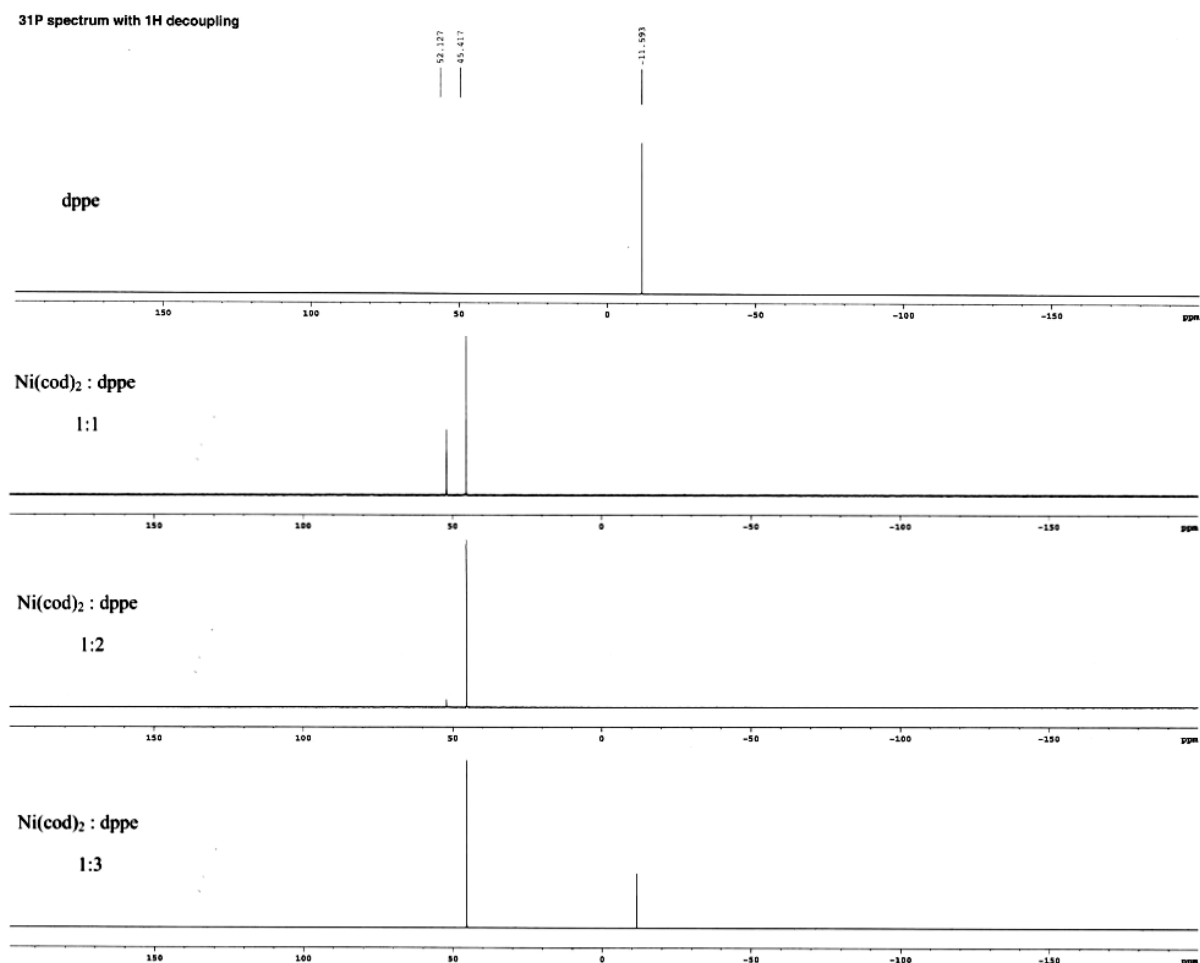
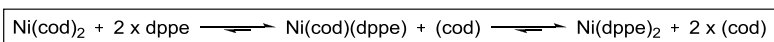


(–)-2-(1,3-diphenylpropyl)benzo[*b*]thiophene (**25**, table 3, entry 10): A 7 mL vial was charged with (*R*)-**SI-10** (0.056 g, 0.20 mmol) and NiCl₂(dppe) (10.6 mg, 0.0200 mmol), flushed with nitrogen and capped. Toluene (3 mL) was added, followed by phenylmagnesium bromide (0.13 mL, 0.40 mmol, 3.1 M in Et₂O) and the reaction was allowed to stir at room temperature for 12 h. The vial was cracked open and another batch of NiCl₂(dppe) (10.6 mg, 0.0200 mmol) was added. After further stirring for 14 h, the reaction was quenched by the addition of EtOAc (1 mL) and the entire reaction mixture was adsorbed onto silica gel (1 g). The solvents were removed in vacuo and the crude was purified by flash column chromatography (2% EtOAc in hexanes) to afford the title compound as a colorless oil (0.050 g, 0.15 mmol, 76%): **TLC** *R*_f = 0.4 (5% EtOAc in hexanes); **¹H NMR** (400 MHz, CDCl₃) δ 7.72 (d, *J* = 8.0 Hz, 1H), 7.66 (d, *J* = 7.6 Hz, 1H), 7.35–7.16 (m, 12H), 7.08 (s, 1H), 4.20 (t, *J* = 7.6 Hz, 1H), 2.66 (m, 2H), 2.56–2.40 (m, 2H); **¹³C NMR** (125 MHz, CDCl₃) δ 150.3, 143.7, 141.8, 140.0, 139.5, 128.8, 128.63, 128.56, 128.0, 127.0, 126.1, 124.3, 123.8, 123.2, 122.3, 120.5, 47.0, 38.4, 34.0; **IR** (neat) 3026, 2922, 2853, 1453 cm⁻¹; **HRMS** (TOF MS CI⁺) *m/z* cacl'd for C₂₃H₂₀S [M+H]⁺ 329.1364, found 329.1362; [α]_D²⁴ –8.4 (*c* 1.00, CHCl₃); **SFC** analysis (OD-H, 5% IPA, 3.0 mL/min) indicated 93% ee: *t*_R (minor) = 16.6 minutes, *t*_R (major) = 17.7 minutes.

VII. Mechanistic Studies

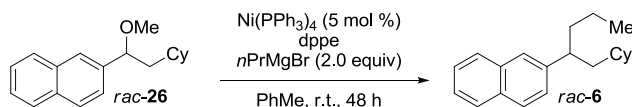
Activity of Ni⁰ Complexes

Procedure for NMR studies: In a glovebox, a vial was charged with Ni(cod)₂ (11 mg, 0.040 mmol), dppe (16 mg, 0.040 mmol) and toluene (3.0 mL). The solution was transferred to a NMR tube and a ³¹P NMR spectrum was collected. Experiment was repeated with 2 and 3 equiv of dppe, (32 mg, 0.080 mmol) and (48 mg, 0.012 mmol) respectively, as well as a control experiment without Ni(cod)₂.



³¹P NMR analysis revealed that the thermodynamically favorable Ni(dppe)₂ complex (45.4 ppm) is the major observed product. At equimolar ratio of Ni and dppe, we observed a peak at 51.1 ppm, which is attributed to a Ni(dppe)(cod) complex. Over time (20 min) this peak disappears and only a signal due to Ni(dppe)₂ is observed.

Procedure for Ni(PPh₃)₄ studies: In a glovebox, a vial was charged with Ni(PPh₃)₄ (11 mg, 0.010 mmol), dppe (4.0 mg, 0.010 mmol), *rac*-**26** (0.054 g, 0.20 mmol) and toluene (3.0 mL). The vial was capped, taken out of the glovebox and placed under N₂ atmosphere. *n*-PrMgBr (0.17 mL, 0.40 mmol, 2.3 M in Et₂O) was added and the mixture was stirred for 48 h. The reaction was quenched by the addition of EtOAc (1 mL), passed through a silica plug (100% Et₂O), the organics were removed under reduced pressure, and the crude was analyzed by ¹H NMR spectroscopy with PhSiMe₃ as internal standard. Experiment was repeated with 12 mol % dppe (9.6 mg), as well as in the absence of dppe.



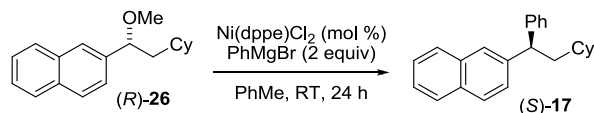
Entry	dppe (mol %)	Recovered 26 (%) ^a	Yield 6 (%) ^a
1	0	91	0
2	5	0	73
3	11	101	0

^a Determined by ¹H NMR analysis using internal standard (PhSiMe₃).

We postulated that the ligand exchange rate between dppe and PPh₃ will be slower than in the case with cod. This difference in rates could preclude the swift formation of the Ni(dppe)₂ complex, thus restoring reactivity in the system. The cross-coupling reaction of *n*-propylmagnesium bromide employing Ni(PPh₃)₄ as the nickel source afforded **6** in 73% yield. Notably, in the absence of dppe as well as when there is an excess of dppe (>2:1 dppe:Ni) no desired reactivity is observed.

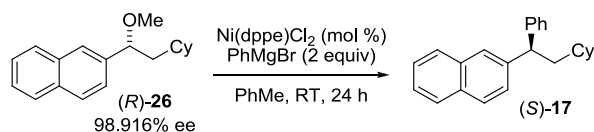
Effect of Catalyst Loading on Enantiospecificity

Procedure: Experiments were performed following the general procedure outlined in Section V. For consistency purposes, only two amounts of NiCl₂(dppe) were weighed out, while the amounts of other reagents were varied to achieve the desired catalyst loading. Benzylic ether (*R*)-**26** was dispensed as a 0.100 M solution in toluene. Phenylmagnesium bromide was dispensed as a 2.8 M solution in Et₂O. For precise amounts of reagents used see Table SI-1 below.



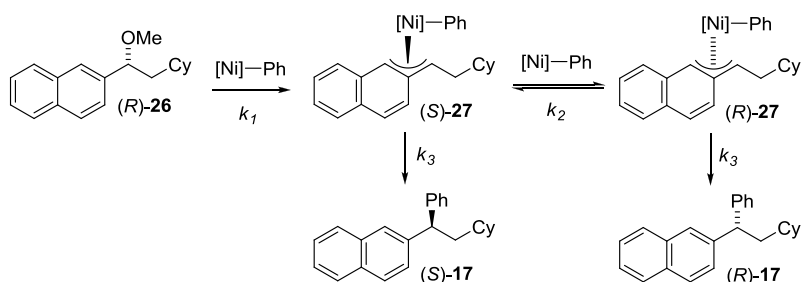
Entry	Ni(dppe)Cl ₂ [mol %]	Ni(dppe)Cl ₂ [mmol]	Ni(dppe)Cl ₂ [mg]	(<i>S</i>)- 26 [mmol]	(<i>R</i>)- 26 [mL]	PhMe [mL]	Total volume [mL]	PhMgBr [mmol]	PhMgBr [μL]
1	4.0	0.010	5.3	0.25	2.50	1.25	3.75	0.50	185
2	6.0	0.012	6.3	0.20	2.00	1.00	3.00	0.40	148
3	8.0	0.012	6.3	0.15	1.50	0.75	2.25	0.30	111
4	10.0	0.010	5.3	0.10	1.00	0.50	1.50	0.20	74
5	12.0	0.012	6.3	0.10	1.00	0.50	1.50	0.20	74

Results:



Entry	Ni(dppe)Cl ₂ [mol %]	ee [%]	es [%]
1	4.0	92.918	93.936
2	6.0	85.178	86.111
3	8.0	81.158	82.047
4	10.0	77.298	78.145
5	12.0	74.358	75.173

Derivation:



We reasoned that one could use a steady state approximation to describe intermediate $(R)\text{-27}$ and express it in terms of concentration of Ni (Eq. 1 and 2). There are a few key assumptions that have to be made: (1) the complex that undergoes oxidative addition and the nucleophilic nickel species responsible for racemization are one and the same or in a fast equilibrium with each other and (2) the above complexes also need to be in fast equilibrium with the precatalyst. Thus, the ratio of the two enantiomers of product formed in the reaction should be inversely proportional to the concentration of nickel (Eq. 3).

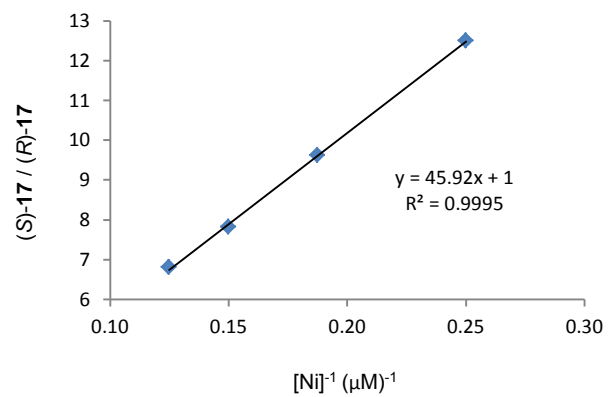
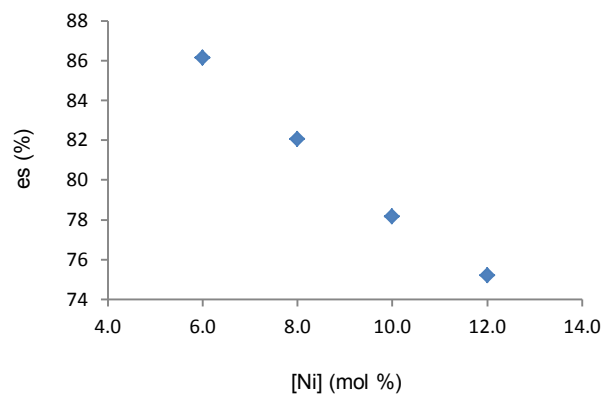
$$\frac{d[(R)\text{-27}]}{dt} = k_2[(S)\text{-27}][\text{Ni}] - k_3[(R)\text{-17}] - k_2^*[(R)\text{-27}][\text{Ni}] = 0 \quad (1)$$

$$[(R)\text{-27}] = \frac{k_2^*[(S)\text{-27}][\text{Ni}]}{k_3 + k_2^*[\text{Ni}]} \quad (2)$$

$$\frac{(S)\text{-17}}{(R)\text{-17}} = \frac{k_3^*[(S)\text{-27}]}{k_3^*[(R)\text{-27}]} = \frac{[(S)\text{-27}]}{\frac{k_2^*[(S)\text{-27}][\text{Ni}]}{k_3 + k_2^*[\text{Ni}]}} = \frac{k_3 + k_2^*[\text{Ni}]}{k_2^*[\text{Ni}]} = \frac{k_3}{k_2^*[\text{Ni}]} + 1 = \frac{k_{obs}}{[\text{Ni}]} + 1 \quad (3)$$

Plots:

Entry	Ni(dppe)Cl ₂ [mol %]	[Ni] [μM]	[Ni] ⁻¹ [μM^{-1}]	(S)-17 [%]	(R)-17 [%]	(S)-17 / (R)-17
1	6.0	4.00	0.250	92.589	7.411	12.493
2	8.0	5.33	0.188	90.579	9.421	9.615
3	10.0	6.67	0.150	88.649	11.351	7.810
4	12.0	8.00	0.125	87.179	12.821	6.800



VIII. Biological Experiments

General Information

Biological experiments were performed according to a modified procedure by Sigman et al.²⁶

Materials: The following reagents were obtained from commercial sources as indicated: Dulbecco's Modified Eagle's Medium (DMEM)/high glucose containing 4.5 g/L glucose and 4.0 mM L-glutamine (HyClone); fetal bovine serum (FBS), heat-inactivated (Omega Scientific); L-glutamine, 200 mM (Gibco); penicillin/streptomycin solution 50X (Mediatech); DMEM/Ham's Nutrient Mixture F12 containing 2.5 mM L-glutamine, 3151 mg/L dextrose, and 55 mg/L sodium pyruvate (Sigma-Aldrich); horse serum (Sigma-Aldrich); 50 μ M hydrocortisone solution (Sigma-Aldrich); human insulin solution (Sigma-Aldrich); cholera toxin (Sigma-Aldrich); human Epidermal Growth Factor (EGF), recombinant (Sigma-Aldrich); 0.25% Trypsin-EDTA (Gibco); nuclease-free sterile water (Fisher Scientific); molecular biology grade DMSO (Sigma-Aldrich); ICI 182,780 (faslodex) (Tocris Bioscience).

Cell Lines and Culture Conditions: MCF-7 cells were maintained in DMEM/high glucose supplemented with 10% FBS, L-glutamine, and penicillin/streptomycin. Experiments with MCF-7 cells were performed in DMEM/high glucose supplemented with 2% FBS, L-glutamine, and penicillin/streptomycin.

MCF-10A cells were maintained in standard medium according to a modified recipe by Brugge et al.²⁷: DMEM/F12 supplemented with 5% horse serum, 10 μ g/mL human insulin, 0.5 μ g/mL hydrocortisone, 10 ng/mL EGF, 100 ng/mL cholera toxin, and penicillin/streptomycin. Experiments with MCF-10A cells were performed in the same medium.

Evaluation of Compounds Against MCF-7 Cells

Procedure: MCF-7 cells were centrifuged in 1X PBS for 20 min, then the pellet was resuspended in DMEM supplemented with 10% FBS and filtered through a 40 μ m nylon cell strainer (Fisher Scientific) to prevent clumping. The cells were seeded at 1,500 cells per well in 96-well flat bottom plates suitable for fluorimetry, using 175 μ L per well DMEM supplemented with 10% FBS, and incubated with 5% CO₂ at 37 °C for 24 h. The compounds (including the faslodex positive control) were dissolved in molecular biology grade DMSO to achieve a 3.5 mM stock, then sterile filtered through a 0.45 μ m PVDF syringe filter unit (Fisher Scientific). The 3.5 mM stock solutions were subsequently diluted to a final concentration of 10 μ M in DMEM supplemented with 2% FBS. Additionally, the corresponding DMSO vehicle control was diluted using the same medium.

The medium was aspirated from the 96-well plates containing the MCF-7 cells using a multichannel pipettor and the medium containing the compounds and controls was added (day 0). The outer row of wells contained medium only to negate edge effects due to the evaporation of the medium. The cells were incubated with compound for 48 h then treated again by aspirating the medium and adding fresh medium containing the compounds and controls (day 2). This procedure was repeated after an additional 48 h (day 4). After incubating a final 24 h, the 96-well plates were flicked dry of medium, rinsed with 1X PBS, blotted dry, and then frozen at -78 °C overnight (day 5). On day 6, cell proliferation was measured using the fluorescence-based CyQUANT Cell Proliferation Assay Kit (Invitrogen).

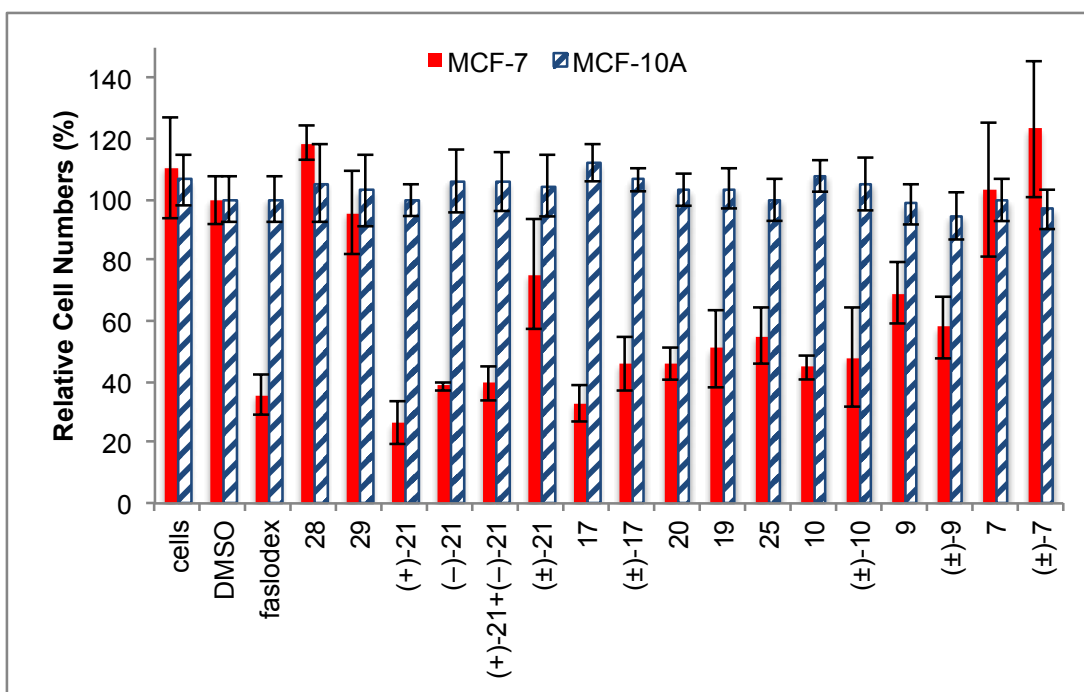
Fluorimetry analysis was performed according to a modified procedure by McGowan et al.²⁸ Cells were stained with 200 μ L/well of 1X CyQUANT GR dye in cell lysis buffer for 10 min in the dark at RT and quantified by fluorimetry at 535 nm with 485 nm excitation, measured using a PerkinElmer VICTOR³ 1420 Multilabel Counter. The fluorescence values were normalized to the DMSO vehicle control. The normalized values were plotted as an average \pm standard deviation of 4 wells per compound.

Evaluation of Compounds Against MCF-10A Cells

Procedure: MCF-10A cells were centrifuged in 1X PBS for 20 min, then the pellet was resuspended in DMEM/F12 and filtered through a 40 μm nylon cell strainer (Fisher Scientific) to prevent clumping. The cells were seeded at 9,000 cells per well in 96-well flat bottom plates suitable for fluorimetry, using 175 μL per well DMEM/F12, and incubated with 5% CO_2 at 37 $^\circ\text{C}$ for 24 h. The 3.5 mM stock solutions of compound in DMSO were subsequently diluted to a final concentration of 10 μM in DMEM/F12. Additionally, the corresponding DMSO vehicle control was diluted using the same medium.

Addition of compounds was performed as specified above for days 0 through 6. Fluorimetry analysis was performed as specified above for MCF-7 cells, with the exception of staining MCF-10A cells with 200 μL /well of 5X CyQUANT GR dye in cell lysis buffer for 10 min in the dark at RT before quantification by fluorimetry. The fluorescence values were normalized to the DMSO vehicle control. The normalized values were plotted as an average \pm standard deviation of 6 wells per compound.

Results:

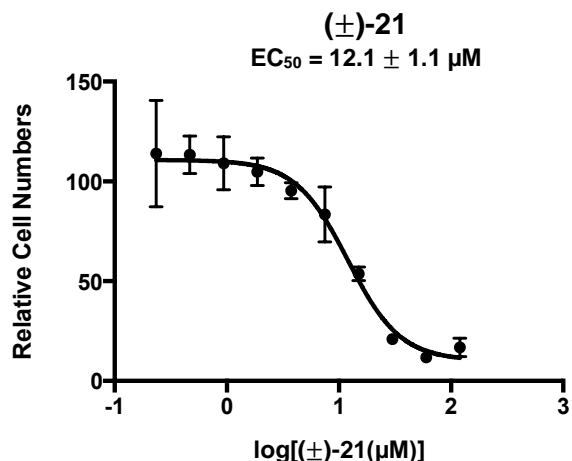
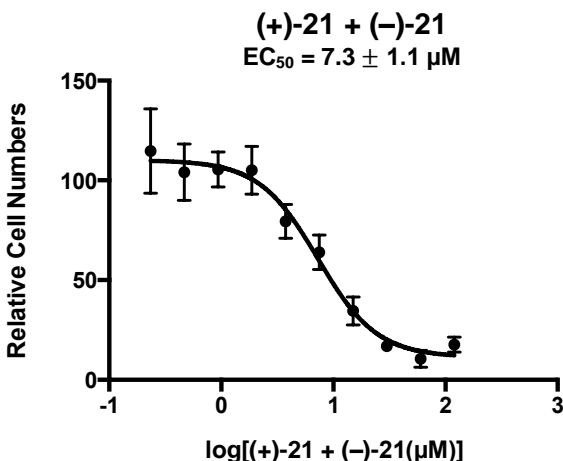
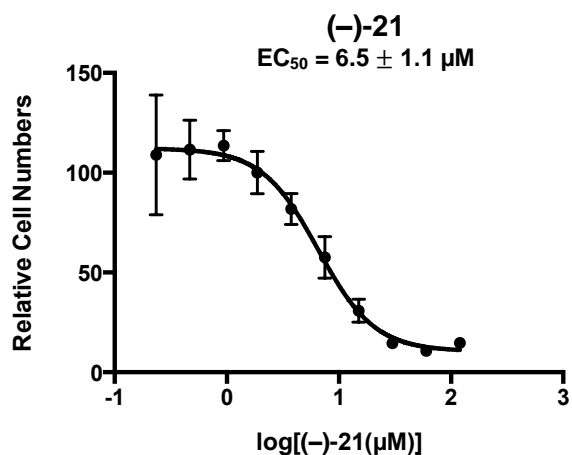
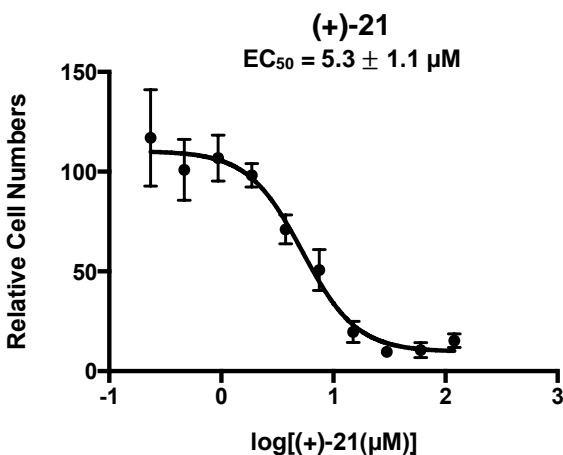


Dose Response of Compounds (+)-21, (-)-21, (+)-21 + (-)-21, and (±)-21

Procedure: MCF-7 cells were centrifuged in 1X PBS for 20 min, then the pellet was resuspended in DMEM supplemented with 10% FBS and filtered through a 40 μm nylon cell strainer (Fisher Scientific) to prevent clumping. The cells were seeded at 1,500 cells per well in 96-well flat bottom plates suitable for fluorimetry, using 175 μL per well DMEM supplemented with 10% FBS, and incubated with 5% CO_2 at 37 $^\circ\text{C}$ for 24 h. The compounds (+)-21, (-)-21, (+)-21 + (-)-21, and (±)-21²⁹ were dissolved in molecular biology grade DMSO to achieve a 42 mM stock, then sterile filtered through a 0.45 μm PVDF syringe filter unit (Fisher Scientific). The 42 mM stock solutions in DMSO were subsequently diluted to 120 μM in DMEM supplemented with 2% FBS, and then serially diluted to achieve 10 different concentrations. Additionally, the corresponding DMSO vehicle controls for each concentration were serially diluted using the same medium.

Addition of compounds was performed as specified above for days 0 through 6. Fluorimetry analysis was performed as specified above for the evaluation of compounds against MCF-7 cells. The fluorescence values were normalized to the DMSO vehicle controls corresponding to each concentration. The normalized values were plotted as an average \pm standard deviation of 4 wells per concentration and these data were analyzed using the dose response nonlinear regression fitting function (log[inhibitor] vs. response with variable slope (four parameters)) with GraphPad Prism 6.

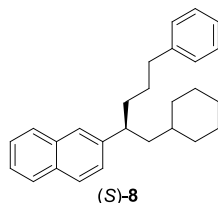
Dose Response Curves:



IX. References

- ¹ Krasovskiy, A.; Knochel, P. *Synthesis* **2006**, *5*, 890–891.
- ² Attenburrow, J.; Cameron, A. F. B.; Chapman, J. H.; Evans, R. M.; Hems, B. A.; Jansen, A. B. A.; Walker, T. *J. Chem. Soc.* **1952**, 1094–1111.
- ³ All alkyl and arylhalides were commercially available and used as received without any purification unless otherwise noted.
- ⁴ Taylor, B. L. H.; Harris, M. R.; Jarvo, E. R. *Angew. Chem. Int. Ed.* **2012**, *51*, 7790–7793.
- ⁵ (a) Marbet, R.; Saucy, G. *Helv. Chim. Acta* **1967**, *50*, 2095–2100. (b) Liu, C.; Kudo, K.; Hashimoto, Y.; Saigo, K. *J. Org. Chem.* **1996**, *61*, 494–502.
- ⁶ Wei, X.; Lorenz, J. C.; Kapdia, S.; Saha, A.; Haddad, N.; Busacca, C. A.; Senanayake, C. H. *J. Org. Chem.* **2007**, *72*, 4250–4253.
- ⁷ Wallace, A. G.; Heathcock, C. H. *J. Org. Chem.* **2001**, *66*, 450–454.
- ⁸ Corey, E. J.; Cheng, H.; Backer, C. H.; Matsuda, S. P. T.; Li, D.; Song, X. *J. Am. Chem. Soc.* **1997**, *119*, 1277–1288.
- ⁹ Boyer, F. D.; Hanna, I. *Org. Lett.* **2007**, *9*, 2293–2295.
- ¹⁰ Corey, E. J.; Helal, C. J. *Angew. Chem. Int. Ed.* **1998**, *37*, 1986–2012.
- ¹¹ (a) Experiment performed by Alexander J. Wagner using the method described in Wagner, A. J.; David, J. G.; Rychnovsky, S. D. *Org. Lett.* **2011**, *13*, 4470–4473. (b) Wagner, A. J.; Rychnovsky, S. D. *J. Org. Chem.* **2013**, *78*, 4594–4598.
- ¹² Taylor, B. L. H.; Swift, E. C.; Waetzig, J. D.; Jarvo, E. R. *J. Am. Chem. Soc.* **2011**, *133*, 389–391.
- ¹³ (a) Li, J. J.; Limberakis, C.; Pflum, D. A. Reductions. *Modern Organic Synthesis in the Laboratory: A Collection of Standard Experimental Procedures*; Oxford University Press: New York, 2007; pp 96–97. (b) Dakin, L. A.; Panek, J. S. *Org. Lett.* **2003**, *5*, 3995–3998.
- ¹⁴ Inagaki, T.; Phong, L. T.; Furuta, A.; Ito, J.-i.; Nishiyama, H. *Chem. Eur. J.* **2010**, *16*, 3090–3096.
- ¹⁵ Guan, B.-T.; Xiang, S.-K.; Wang, B.-Q.; Sun, Z.-P.; Wang, Y.; Zhao, K.-Q.; Shi, Z.-J. *J. Am. Chem. Soc.* **2008**, *130*, 3268–3269.
- ¹⁶ Wisniewska, H. M.; Swift, E. C.; Jarvo, E. R. *J. Am. Chem. Soc.* **2013**, *135*, 9083–9090.
- ¹⁷ Dodge, J. A.; Nissen, J. S.; Presnell, M. *Org. Synth.* **1996**, *73*, 110–112.
- ¹⁸ Zhang, F.-Y.; Yip, C.-W.; Rong, C.; Chan, A. S. C. *Tetrahedron Asymmetry* **1997**, *8*, 585–588.
- ¹⁹ Denis, J.; Guinchard, X. *J. Org. Chem.* **2008**, *73*, 2028–2031.
- ²⁰ Suzuki, K.; Kondo, K.; Aoyama, T. *Synthesis* **2006**, 1360–1364.
- ²¹ Gribkov, D. V.; Hultsch, K. C.; Hampel, F. *J. Am. Chem. Soc.* **2006**, *128*, 3748–3759.
- ²² Narender, T.; Sarkar, S.; Rajendar, K.; Tiwari, S. *Org. Lett.* **2011**, *13*, 6140–6143.
- ²³ Lee, A. S.; Norman, A. W.; Okamura, W. H. *J. Org. Chem.* **1992**, *57*, 3846–3854.
- ²⁴ Caporusso, A. M.; Zampieri, A.; Aronica, L. A.; Banti, D. *J. Org. Chem.* **2006**, *71*, 1902–1910.
- ²⁵ Cho, C.-H.; Sun, M.; Seo, Y.-S.; Kim, C.-B.; Park, K. *J. Org. Chem.* **2005**, *70*, 1482–1485.
- ²⁶ Pathak, T. P.; Gligorich, K. M.; Welm, B. E.; Sigman, M. S. *J. Am. Chem. Soc.* **2010**, *132*, 7870–7871.
- ²⁷ Debnath, J.; Muthuswamy, S. K.; Brugge, J. S. *Methods* **2003**, *30*, 256–268.
- ²⁸ McGowan, E. M.; Alling, N.; Jackson, E. A.; Yagoub, D.; Haass, N. K.; Allen, J. D.; Martinello-Wilks, R. *PLoS ONE* **2011**, *6*, e20623.
- ²⁹ (+)-**21** and (–)-**21** are samples of each single enantiomer with 93% ee and 92% ee, respectively; (±)-**21** is a sample of the racemic standard; (+)-**21** + (–)-**21** is a sample containing an equimolar ratio of the two enantiomers, combined in DMSO prior to biological testing.

X. Crystallographic Data



X-ray Data Collection, Structure Solution and Refinement for (S)-8 (Table 2, entry 4).

The single crystal X-ray diffraction studies were carried out on a Bruker Kappa APEX CCD diffractometer equipped with Cu K_{α} radiation ($\lambda = 1.5478$). Crystals of the subject compound were grown by slow evaporation of solvent from a solution of the title compound in a mixture of methanol and pentane. A 0.217 x 0.095 x 0.053 mm colorless needle was mounted on a Cryoloop with Paratone oil. Data were collected in a nitrogen gas stream at 90(2) K using ϕ and ω scans. Data was collected at two crystal-to-detector distances, 45mm or 60mm, using variable exposure time (2s-10s) depending on θ with a scan width of 1.0°. Data collection was 98.7% complete to 68.00° in θ . A total of 57312 reflections were collected covering the indices, $-7 \leq h \leq 7$, $-16 \leq k \leq 16$, $-17 \leq l \leq 17$. 7355 reflections were found to be symmetry independent, with a R_{int} of 0.0524. Indexing and unit cell refinement indicated a primitive, triclinic lattice. The space group was found to be $P1$. The data were integrated using the Bruker SAINT software program and scaled using the SADABS software program. Solution by direct methods (SHELXT) produced a complete phasing model consistent with the proposed structure.

All nonhydrogen atoms were refined anisotropically by full-matrix least-squares (SHELXL-2013). All hydrogen atoms were placed using a riding model. Their positions were constrained relative to their parent atom using the appropriate HFIX command in SHELXL-2013. Crystallographic data are summarized in Table 1.

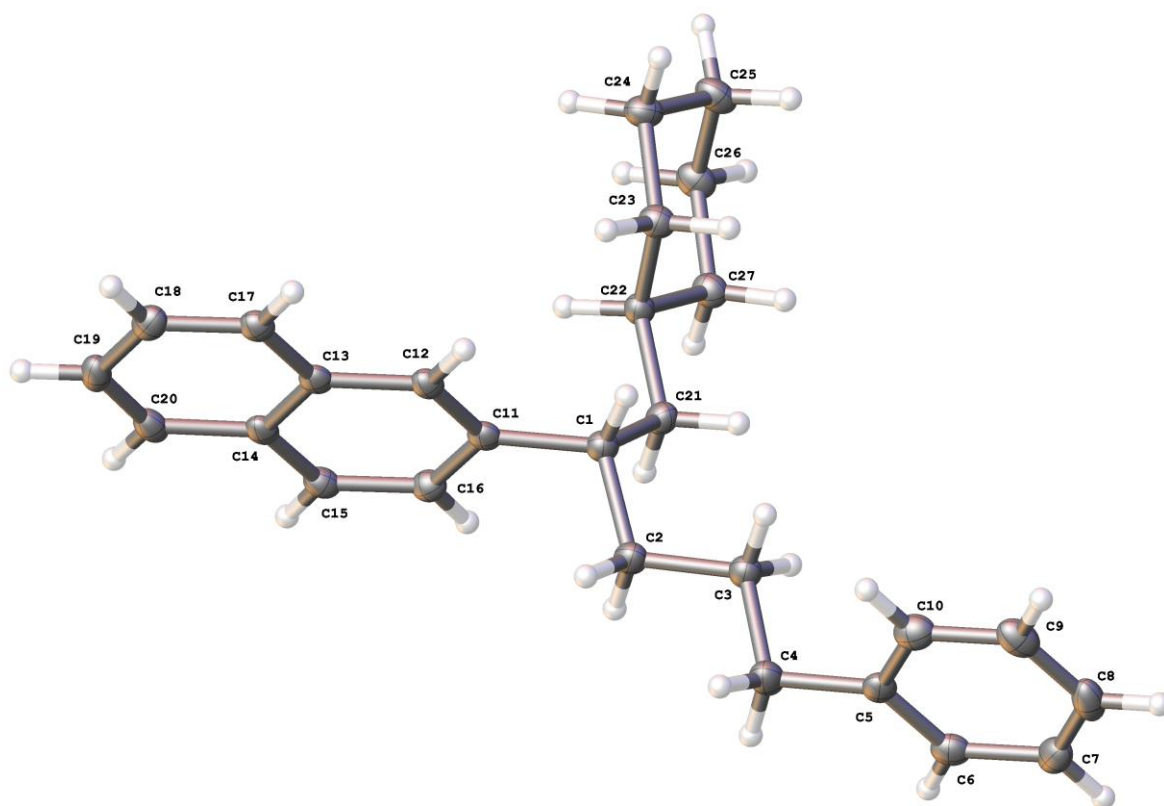
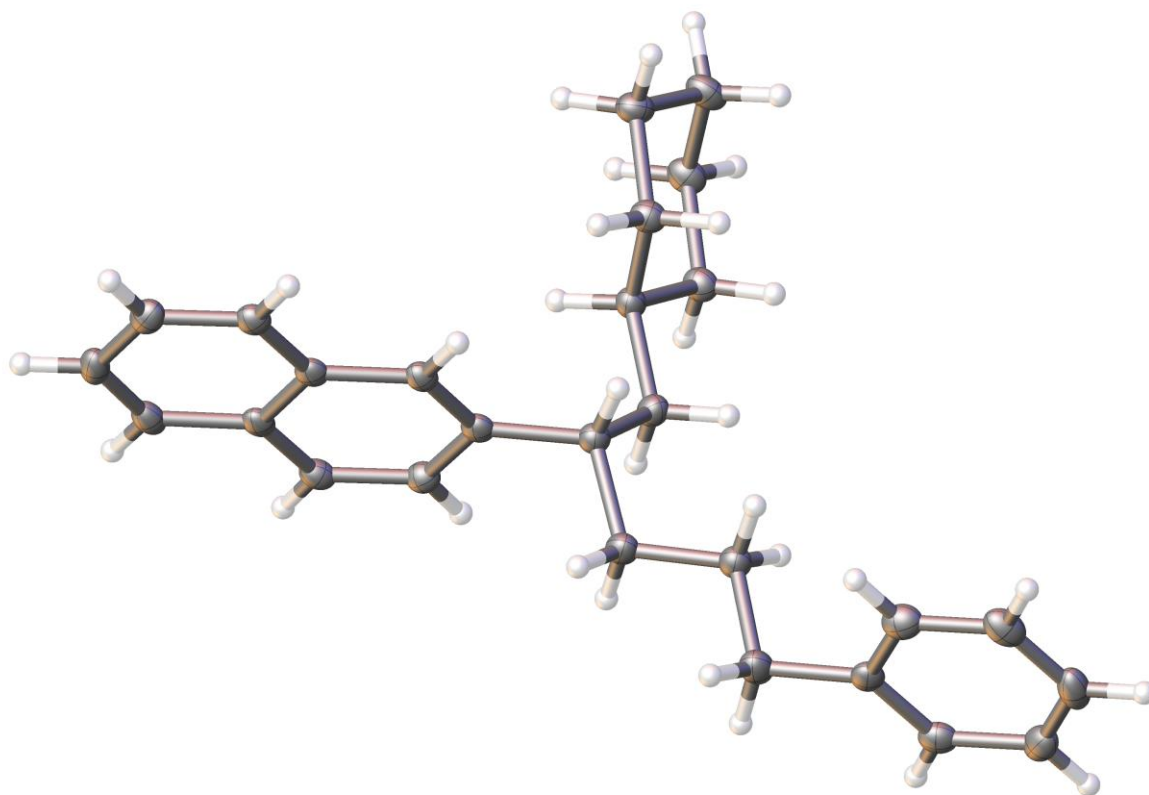


Table 1. Crystal data and structure refinement for Jarvo02.

Identification code	ERJ-14	
Empirical formula	C ₂₇ H ₃₂	
Molecular formula	C ₂₇ H ₃₂	
Formula weight	356.52	
Temperature	90 K	
Wavelength	1.54178 Å	
Crystal system	Triclinic	
Space group	P 1	
Unit cell dimensions	a = 5.9031(2) Å	α = 106.3499(15)°.
	b = 13.8552(5) Å	β = 98.3273(14)°.
	c = 14.0936(5) Å	γ = 101.5540(15)°.
Volume	1058.56(7) Å ³	
Z	2	
Density (calculated)	1.119 Mg/m ³	
Absorption coefficient	0.463 mm ⁻¹	
F(000)	388	
Crystal size	0.217 x 0.095 x 0.053 mm ³	
Crystal color, habit	Colorless Needle	
Theta range for data collection	3.345 to 69.358°.	
Index ranges	-7 ≤ h ≤ 7, -16 ≤ k ≤ 16, -17 ≤ l ≤ 17	
Reflections collected	57312	
Independent reflections	7355 [R(int) = 0.0524]	
Completeness to theta = 68.000°	98.7 %	
Absorption correction	Semi-empirical from equivalents	
Max. and min. transmission	0.7532 and 0.7017	
Refinement method	Full-matrix least-squares on F ²	
Data / restraints / parameters	7355 / 3 / 487	
Goodness-of-fit on F ²	1.029	
Final R indices [I > 2σ(I)]	R1 = 0.0312, wR2 = 0.0787	
R indices (all data)	R1 = 0.0324, wR2 = 0.0799	
Absolute structure parameter	0.065(347)	
Extinction coefficient	n/a	
Largest diff. peak and hole	0.120 and -0.172 e.Å ⁻³	

Table 2. Atomic coordinates ($\times 10^4$) and equivalent isotropic displacement parameters ($\text{\AA}^2 \times 10^3$) for Jarvo02. $U(\text{eq})$ is defined as one third of the trace of the orthogonalized U^{ij} tensor.

	x	y	z	$U(\text{eq})$
C(1)	8627(3)	2712(2)	9759(1)	20(1)
C(2)	8982(4)	1839(2)	10204(2)	23(1)
C(3)	9335(3)	883(2)	9445(1)	21(1)
C(4)	10054(4)	108(2)	9944(2)	27(1)
C(5)	10507(3)	-808(2)	9186(1)	21(1)
C(6)	8896(3)	-1776(2)	8845(2)	23(1)
C(7)	9312(4)	-2603(2)	8126(2)	27(1)
C(8)	11349(4)	-2473(2)	7748(2)	29(1)
C(9)	12964(4)	-1513(2)	8083(2)	28(1)
C(10)	12546(4)	-689(2)	8797(2)	26(1)
C(11)	8776(3)	3692(2)	10609(1)	18(1)
C(12)	10775(3)	4502(2)	10932(1)	18(1)
C(13)	11023(3)	5399(2)	11773(1)	18(1)
C(14)	9133(3)	5458(2)	12292(1)	19(1)
C(15)	7078(3)	4622(2)	11945(1)	21(1)
C(16)	6901(3)	3768(2)	11136(1)	20(1)
C(17)	13086(3)	6234(2)	12111(1)	20(1)
C(18)	13267(3)	7090(2)	12923(2)	23(1)
C(19)	11385(4)	7148(2)	13438(2)	24(1)
C(20)	9373(4)	6355(2)	13131(2)	22(1)
C(21)	6322(3)	2349(2)	8940(2)	21(1)
C(22)	5748(3)	3152(2)	8446(1)	19(1)
C(23)	7800(3)	3596(2)	8012(2)	22(1)
C(24)	7172(4)	4332(2)	7439(2)	24(1)
C(25)	4963(4)	3801(2)	6602(2)	27(1)
C(26)	2893(3)	3389(2)	7036(2)	25(1)
C(27)	3503(3)	2646(2)	7602(2)	22(1)
C(1')	9460(3)	671(2)	3372(1)	21(1)
C(2')	10172(4)	1568(2)	2946(2)	24(1)
C(3')	11922(4)	2533(2)	3706(2)	23(1)
C(4')	12998(4)	3309(2)	3205(2)	28(1)

C(5')	14500(4)	4307(2)	3977(2)	23(1)
C(6')	13548(4)	5157(2)	4278(2)	25(1)
C(7')	14867(4)	6067(2)	5020(2)	28(1)
C(8')	17162(4)	6145(2)	5474(2)	29(1)
C(9')	18138(4)	5304(2)	5183(2)	28(1)
C(10')	16815(4)	4395(2)	4438(2)	26(1)
C(11')	7920(3)	-281(2)	2526(1)	19(1)
C(12')	8759(3)	-1134(2)	2159(1)	19(1)
C(13')	7410(3)	-2009(2)	1326(1)	18(1)
C(14')	5111(3)	-1999(2)	863(1)	19(1)
C(15')	4255(3)	-1114(2)	1263(2)	21(1)
C(16')	5600(3)	-288(2)	2066(2)	21(1)
C(17')	8279(3)	-2891(2)	942(1)	20(1)
C(18')	6937(4)	-3720(2)	136(2)	23(1)
C(19')	4663(3)	-3706(2)	-327(2)	23(1)
C(20')	3773(3)	-2862(2)	29(2)	22(1)
C(21')	8305(3)	988(2)	4284(2)	22(1)
C(22')	7554(3)	139(2)	4758(1)	19(1)
C(23')	9615(3)	-298(2)	5085(2)	23(1)
C(24')	8856(4)	-1122(2)	5585(2)	26(1)
C(25')	7787(4)	-692(2)	6486(2)	28(1)
C(26')	5719(4)	-262(2)	6172(2)	27(1)
C(27')	6475(3)	559(2)	5667(2)	24(1)

Table 3. Bond lengths [Å] and angles [°] for Jarvo02.

C(1)-H(1)	1.0000	C(16)-H(16)	0.9500
C(1)-C(2)	1.546(3)	C(17)-H(17)	0.9500
C(1)-C(11)	1.514(3)	C(17)-C(18)	1.370(3)
C(1)-C(21)	1.542(3)	C(18)-H(18)	0.9500
C(2)-H(2A)	0.9900	C(18)-C(19)	1.415(3)
C(2)-H(2B)	0.9900	C(19)-H(19)	0.9500
C(2)-C(3)	1.523(3)	C(19)-C(20)	1.366(3)
C(3)-H(3A)	0.9900	C(20)-H(20)	0.9500
C(3)-H(3B)	0.9900	C(21)-H(21A)	0.9900
C(3)-C(4)	1.533(3)	C(21)-H(21B)	0.9900
C(4)-H(4A)	0.9900	C(21)-C(22)	1.536(3)
C(4)-H(4B)	0.9900	C(22)-H(22)	1.0000
C(4)-C(5)	1.510(3)	C(22)-C(23)	1.533(3)
C(5)-C(6)	1.388(3)	C(22)-C(27)	1.538(3)
C(5)-C(10)	1.392(3)	C(23)-H(23A)	0.9900
C(6)-H(6)	0.9500	C(23)-H(23B)	0.9900
C(6)-C(7)	1.389(3)	C(23)-C(24)	1.534(3)
C(7)-H(7)	0.9500	C(24)-H(24A)	0.9900
C(7)-C(8)	1.384(3)	C(24)-H(24B)	0.9900
C(8)-H(8)	0.9500	C(24)-C(25)	1.522(3)
C(8)-C(9)	1.382(3)	C(25)-H(25A)	0.9900
C(9)-H(9)	0.9500	C(25)-H(25B)	0.9900
C(9)-C(10)	1.384(3)	C(25)-C(26)	1.526(3)
C(10)-H(10)	0.9500	C(26)-H(26A)	0.9900
C(11)-C(12)	1.373(3)	C(26)-H(26B)	0.9900
C(11)-C(16)	1.424(3)	C(26)-C(27)	1.533(3)
C(12)-H(12)	0.9500	C(27)-H(27A)	0.9900
C(12)-C(13)	1.421(3)	C(27)-H(27B)	0.9900
C(13)-C(14)	1.423(3)	C(1')-H(1')	1.0000
C(13)-C(17)	1.416(3)	C(1')-C(2')	1.539(3)
C(14)-C(15)	1.415(3)	C(1')-C(11')	1.519(3)
C(14)-C(20)	1.420(3)	C(1')-C(21')	1.538(3)
C(15)-H(15)	0.9500	C(2')-H(2'A)	0.9900
C(15)-C(16)	1.366(3)	C(2')-H(2'B)	0.9900

C(2')-C(3')	1.524(3)	C(21')-H(21C)	0.9900
C(3')-H(3'A)	0.9900	C(21')-H(21D)	0.9900
C(3')-H(3'B)	0.9900	C(21')-C(22')	1.531(3)
C(3')-C(4')	1.531(3)	C(22')-H(22')	1.0000
C(4')-H(4'A)	0.9900	C(22')-C(23')	1.533(3)
C(4')-H(4'B)	0.9900	C(22')-C(27')	1.533(3)
C(4')-C(5')	1.505(3)	C(23')-H(23C)	0.9900
C(5')-C(6')	1.391(3)	C(23')-H(23D)	0.9900
C(5')-C(10')	1.392(3)	C(23')-C(24')	1.530(3)
C(6')-H(6')	0.9500	C(24')-H(24C)	0.9900
C(6')-C(7')	1.385(3)	C(24')-H(24D)	0.9900
C(7')-H(7')	0.9500	C(24')-C(25')	1.524(3)
C(7')-C(8')	1.381(3)	C(25')-H(25C)	0.9900
C(8')-H(8')	0.9500	C(25')-H(25D)	0.9900
C(8')-C(9')	1.388(3)	C(25')-C(26')	1.524(3)
C(9')-H(9')	0.9500	C(26')-H(26C)	0.9900
C(9')-C(10')	1.386(3)	C(26')-H(26D)	0.9900
C(10')-H(10')	0.9500	C(26')-C(27')	1.531(3)
C(11')-C(12')	1.369(3)	C(27')-H(27C)	0.9900
C(11')-C(16')	1.425(3)	C(27')-H(27D)	0.9900
C(12')-H(12')	0.9500		
C(12')-C(13')	1.422(3)	C(2)-C(1)-H(1)	107.4
C(13')-C(14')	1.423(2)	C(11)-C(1)-H(1)	107.4
C(13')-C(17')	1.417(3)	C(11)-C(1)-C(2)	109.61(15)
C(14')-C(15')	1.421(3)	C(11)-C(1)-C(21)	113.87(16)
C(14')-C(20')	1.413(3)	C(21)-C(1)-H(1)	107.4
C(15')-H(15')	0.9500	C(21)-C(1)-C(2)	110.96(16)
C(15')-C(16')	1.366(3)	C(1)-C(2)-H(2A)	108.7
C(16')-H(16')	0.9500	C(1)-C(2)-H(2B)	108.7
C(17')-H(17')	0.9500	H(2A)-C(2)-H(2B)	107.6
C(17')-C(18')	1.369(3)	C(3)-C(2)-C(1)	114.19(16)
C(18')-H(18')	0.9500	C(3)-C(2)-H(2A)	108.7
C(18')-C(19')	1.411(3)	C(3)-C(2)-H(2B)	108.7
C(19')-H(19')	0.9500	C(2)-C(3)-H(3A)	109.1
C(19')-C(20')	1.373(3)	C(2)-C(3)-H(3B)	109.1
C(20')-H(20')	0.9500	C(2)-C(3)-C(4)	112.66(16)

H(3A)-C(3)-H(3B)	107.8	C(15)-C(14)-C(13)	118.46(18)
C(4)-C(3)-H(3A)	109.1	C(15)-C(14)-C(20)	122.61(18)
C(4)-C(3)-H(3B)	109.1	C(20)-C(14)-C(13)	118.94(18)
C(3)-C(4)-H(4A)	109.3	C(14)-C(15)-H(15)	119.4
C(3)-C(4)-H(4B)	109.3	C(16)-C(15)-C(14)	121.24(18)
H(4A)-C(4)-H(4B)	107.9	C(16)-C(15)-H(15)	119.4
C(5)-C(4)-C(3)	111.74(16)	C(11)-C(16)-H(16)	119.4
C(5)-C(4)-H(4A)	109.3	C(15)-C(16)-C(11)	121.12(18)
C(5)-C(4)-H(4B)	109.3	C(15)-C(16)-H(16)	119.4
C(6)-C(5)-C(4)	121.28(18)	C(13)-C(17)-H(17)	119.6
C(6)-C(5)-C(10)	118.54(19)	C(18)-C(17)-C(13)	120.76(18)
C(10)-C(5)-C(4)	120.17(19)	C(18)-C(17)-H(17)	119.6
C(5)-C(6)-H(6)	119.8	C(17)-C(18)-H(18)	119.9
C(5)-C(6)-C(7)	120.47(19)	C(17)-C(18)-C(19)	120.16(19)
C(7)-C(6)-H(6)	119.8	C(19)-C(18)-H(18)	119.9
C(6)-C(7)-H(7)	119.8	C(18)-C(19)-H(19)	119.8
C(8)-C(7)-C(6)	120.3(2)	C(20)-C(19)-C(18)	120.5(2)
C(8)-C(7)-H(7)	119.8	C(20)-C(19)-H(19)	119.8
C(7)-C(8)-H(8)	120.2	C(14)-C(20)-H(20)	119.7
C(9)-C(8)-C(7)	119.6(2)	C(19)-C(20)-C(14)	120.66(19)
C(9)-C(8)-H(8)	120.2	C(19)-C(20)-H(20)	119.7
C(8)-C(9)-H(9)	120.0	C(1)-C(21)-H(21A)	108.3
C(8)-C(9)-C(10)	120.0(2)	C(1)-C(21)-H(21B)	108.3
C(10)-C(9)-H(9)	120.0	H(21A)-C(21)-H(21B)	107.4
C(5)-C(10)-H(10)	119.5	C(22)-C(21)-C(1)	116.10(16)
C(9)-C(10)-C(5)	121.0(2)	C(22)-C(21)-H(21A)	108.3
C(9)-C(10)-H(10)	119.5	C(22)-C(21)-H(21B)	108.3
C(12)-C(11)-C(1)	120.95(17)	C(21)-C(22)-H(22)	108.5
C(12)-C(11)-C(16)	118.36(18)	C(21)-C(22)-C(27)	109.54(16)
C(16)-C(11)-C(1)	120.57(18)	C(23)-C(22)-C(21)	111.91(15)
C(11)-C(12)-H(12)	119.0	C(23)-C(22)-H(22)	108.5
C(11)-C(12)-C(13)	121.99(17)	C(23)-C(22)-C(27)	109.80(15)
C(13)-C(12)-H(12)	119.0	C(27)-C(22)-H(22)	108.5
C(12)-C(13)-C(14)	118.84(17)	C(22)-C(23)-H(23A)	109.1
C(17)-C(13)-C(12)	122.18(17)	C(22)-C(23)-H(23B)	109.1
C(17)-C(13)-C(14)	118.98(18)	C(22)-C(23)-C(24)	112.37(15)

H(23A)-C(23)-H(23B)	107.9	C(3')-C(2')-C(1')	114.09(16)
C(24)-C(23)-H(23A)	109.1	C(3')-C(2')-H(2'A)	108.7
C(24)-C(23)-H(23B)	109.1	C(3')-C(2')-H(2'B)	108.7
C(23)-C(24)-H(24A)	109.3	C(2')-C(3')-H(3'A)	109.1
C(23)-C(24)-H(24B)	109.3	C(2')-C(3')-H(3'B)	109.1
H(24A)-C(24)-H(24B)	108.0	C(2')-C(3')-C(4')	112.48(16)
C(25)-C(24)-C(23)	111.43(17)	H(3'A)-C(3')-H(3'B)	107.8
C(25)-C(24)-H(24A)	109.3	C(4')-C(3')-H(3'A)	109.1
C(25)-C(24)-H(24B)	109.3	C(4')-C(3')-H(3'B)	109.1
C(24)-C(25)-H(25A)	109.6	C(3')-C(4')-H(4'A)	109.3
C(24)-C(25)-H(25B)	109.6	C(3')-C(4')-H(4'B)	109.3
C(24)-C(25)-C(26)	110.29(16)	H(4'A)-C(4')-H(4'B)	107.9
H(25A)-C(25)-H(25B)	108.1	C(5')-C(4')-C(3')	111.69(16)
C(26)-C(25)-H(25A)	109.6	C(5')-C(4')-H(4'A)	109.3
C(26)-C(25)-H(25B)	109.6	C(5')-C(4')-H(4'B)	109.3
C(25)-C(26)-H(26A)	109.4	C(6')-C(5')-C(4')	120.02(19)
C(25)-C(26)-H(26B)	109.4	C(6')-C(5')-C(10')	118.34(19)
C(25)-C(26)-C(27)	110.99(16)	C(10')-C(5')-C(4')	121.6(2)
H(26A)-C(26)-H(26B)	108.0	C(5')-C(6')-H(6')	119.5
C(27)-C(26)-H(26A)	109.4	C(7')-C(6')-C(5')	120.91(19)
C(27)-C(26)-H(26B)	109.4	C(7')-C(6')-H(6')	119.5
C(22)-C(27)-H(27A)	109.1	C(6')-C(7')-H(7')	119.9
C(22)-C(27)-H(27B)	109.1	C(8')-C(7')-C(6')	120.3(2)
C(26)-C(27)-C(22)	112.43(17)	C(8')-C(7')-H(7')	119.9
C(26)-C(27)-H(27A)	109.1	C(7')-C(8')-H(8')	120.2
C(26)-C(27)-H(27B)	109.1	C(7')-C(8')-C(9')	119.6(2)
H(27A)-C(27)-H(27B)	107.9	C(9')-C(8')-H(8')	120.2
C(2')-C(1')-H(1')	107.3	C(8')-C(9')-H(9')	120.0
C(11')-C(1')-H(1')	107.3	C(10')-C(9')-C(8')	119.99(19)
C(11')-C(1')-C(2')	109.48(15)	C(10')-C(9')-H(9')	120.0
C(11')-C(1')-C(21')	113.10(16)	C(5')-C(10')-H(10')	119.5
C(21')-C(1')-H(1')	107.3	C(9')-C(10')-C(5')	120.9(2)
C(21')-C(1')-C(2')	112.20(17)	C(9')-C(10')-H(10')	119.5
C(1')-C(2')-H(2'A)	108.7	C(12')-C(11')-C(1')	120.97(17)
C(1')-C(2')-H(2'B)	108.7	C(12')-C(11')-C(16')	118.38(18)
H(2'A)-C(2')-H(2'B)	107.6	C(16')-C(11')-C(1')	120.59(18)

C(11')-C(12')-H(12')	118.9	C(21')-C(22')-C(27')	110.28(16)
C(11')-C(12')-C(13')	122.11(17)	C(23')-C(22')-H(22')	108.1
C(13')-C(12')-H(12')	118.9	C(23')-C(22')-C(27')	109.76(15)
C(12')-C(13')-C(14')	118.90(18)	C(27')-C(22')-H(22')	108.1
C(17')-C(13')-C(12')	122.20(17)	C(22')-C(23')-H(23C)	109.3
C(17')-C(13')-C(14')	118.90(18)	C(22')-C(23')-H(23D)	109.3
C(15')-C(14')-C(13')	118.23(18)	H(23C)-C(23')-H(23D)	107.9
C(20')-C(14')-C(13')	119.09(18)	C(24')-C(23')-C(22')	111.76(15)
C(20')-C(14')-C(15')	122.68(17)	C(24')-C(23')-H(23C)	109.3
C(14')-C(15')-H(15')	119.4	C(24')-C(23')-H(23D)	109.3
C(16')-C(15')-C(14')	121.23(17)	C(23')-C(24')-H(24C)	109.4
C(16')-C(15')-H(15')	119.4	C(23')-C(24')-H(24D)	109.4
C(11')-C(16')-H(16')	119.4	H(24C)-C(24')-H(24D)	108.0
C(15')-C(16')-C(11')	121.13(18)	C(25')-C(24')-C(23')	111.14(18)
C(15')-C(16')-H(16')	119.4	C(25')-C(24')-H(24C)	109.4
C(13')-C(17')-H(17')	119.6	C(25')-C(24')-H(24D)	109.4
C(18')-C(17')-C(13')	120.72(18)	C(24')-C(25')-H(25C)	109.4
C(18')-C(17')-H(17')	119.6	C(24')-C(25')-H(25D)	109.4
C(17')-C(18')-H(18')	119.8	C(24')-C(25')-C(26')	110.98(17)
C(17')-C(18')-C(19')	120.34(19)	H(25C)-C(25')-H(25D)	108.0
C(19')-C(18')-H(18')	119.8	C(26')-C(25')-H(25C)	109.4
C(18')-C(19')-H(19')	119.9	C(26')-C(25')-H(25D)	109.4
C(20')-C(19')-C(18')	120.27(19)	C(25')-C(26')-H(26C)	109.5
C(20')-C(19')-H(19')	119.9	C(25')-C(26')-H(26D)	109.5
C(14')-C(20')-H(20')	119.7	C(25')-C(26')-C(27')	110.94(17)
C(19')-C(20')-C(14')	120.68(18)	H(26C)-C(26')-H(26D)	108.0
C(19')-C(20')-H(20')	119.7	C(27')-C(26')-H(26C)	109.5
C(1')-C(21')-H(21C)	108.4	C(27')-C(26')-H(26D)	109.5
C(1')-C(21')-H(21D)	108.4	C(22')-C(27')-H(27C)	109.2
H(21C)-C(21')-H(21D)	107.5	C(22')-C(27')-H(27D)	109.2
C(22')-C(21')-C(1')	115.48(16)	C(26')-C(27')-C(22')	112.16(17)
C(22')-C(21')-H(21C)	108.4	C(26')-C(27')-H(27C)	109.2
C(22')-C(21')-H(21D)	108.4	C(26')-C(27')-H(27D)	109.2
C(21')-C(22')-H(22')	108.1	H(27C)-C(27')-H(27D)	107.9
C(21')-C(22')-C(23')	112.32(15)		

Table 4. Anisotropic displacement parameters ($\text{\AA}^2 \times 10^3$) for Jarvo02. The anisotropic displacement factor exponent takes the form: $-2\pi^2 [h^2 a^{*2} U^{11} + \dots + 2 h k a^* b^* U^{12}]$

	U ¹¹	U ²²	U ³³	U ²³	U ¹³	U ¹²
C(1)	23(1)	19(1)	18(1)	7(1)	3(1)	6(1)
C(2)	30(1)	20(1)	18(1)	7(1)	2(1)	7(1)
C(3)	24(1)	20(1)	19(1)	7(1)	3(1)	7(1)
C(4)	38(1)	24(1)	21(1)	9(1)	6(1)	13(1)
C(5)	26(1)	20(1)	17(1)	9(1)	1(1)	9(1)
C(6)	22(1)	27(1)	22(1)	13(1)	2(1)	8(1)
C(7)	34(1)	22(1)	20(1)	6(1)	-4(1)	5(1)
C(8)	41(1)	31(1)	18(1)	5(1)	2(1)	20(1)
C(9)	27(1)	39(1)	24(1)	14(1)	7(1)	16(1)
C(10)	24(1)	28(1)	26(1)	12(1)	1(1)	4(1)
C(11)	21(1)	21(1)	16(1)	9(1)	1(1)	8(1)
C(12)	20(1)	21(1)	16(1)	8(1)	3(1)	9(1)
C(13)	22(1)	21(1)	16(1)	10(1)	2(1)	9(1)
C(14)	24(1)	22(1)	15(1)	10(1)	3(1)	11(1)
C(15)	21(1)	26(1)	20(1)	11(1)	6(1)	9(1)
C(16)	20(1)	21(1)	19(1)	8(1)	1(1)	4(1)
C(17)	21(1)	23(1)	18(1)	10(1)	2(1)	8(1)
C(18)	24(1)	21(1)	21(1)	7(1)	-3(1)	5(1)
C(19)	32(1)	21(1)	16(1)	4(1)	-1(1)	12(1)
C(20)	28(1)	28(1)	16(1)	10(1)	6(1)	15(1)
C(21)	25(1)	18(1)	19(1)	6(1)	1(1)	5(1)
C(22)	22(1)	18(1)	16(1)	6(1)	3(1)	7(1)
C(23)	22(1)	25(1)	21(1)	9(1)	4(1)	7(1)
C(24)	28(1)	26(1)	24(1)	13(1)	7(1)	9(1)
C(25)	30(1)	36(1)	19(1)	13(1)	5(1)	12(1)
C(26)	23(1)	31(1)	21(1)	10(1)	1(1)	9(1)
C(27)	22(1)	25(1)	19(1)	7(1)	2(1)	5(1)
C(1')	24(1)	21(1)	18(1)	8(1)	4(1)	5(1)
C(2')	30(1)	22(1)	18(1)	7(1)	3(1)	4(1)
C(3')	28(1)	22(1)	20(1)	8(1)	4(1)	3(1)
C(4')	36(1)	25(1)	21(1)	9(1)	5(1)	2(1)

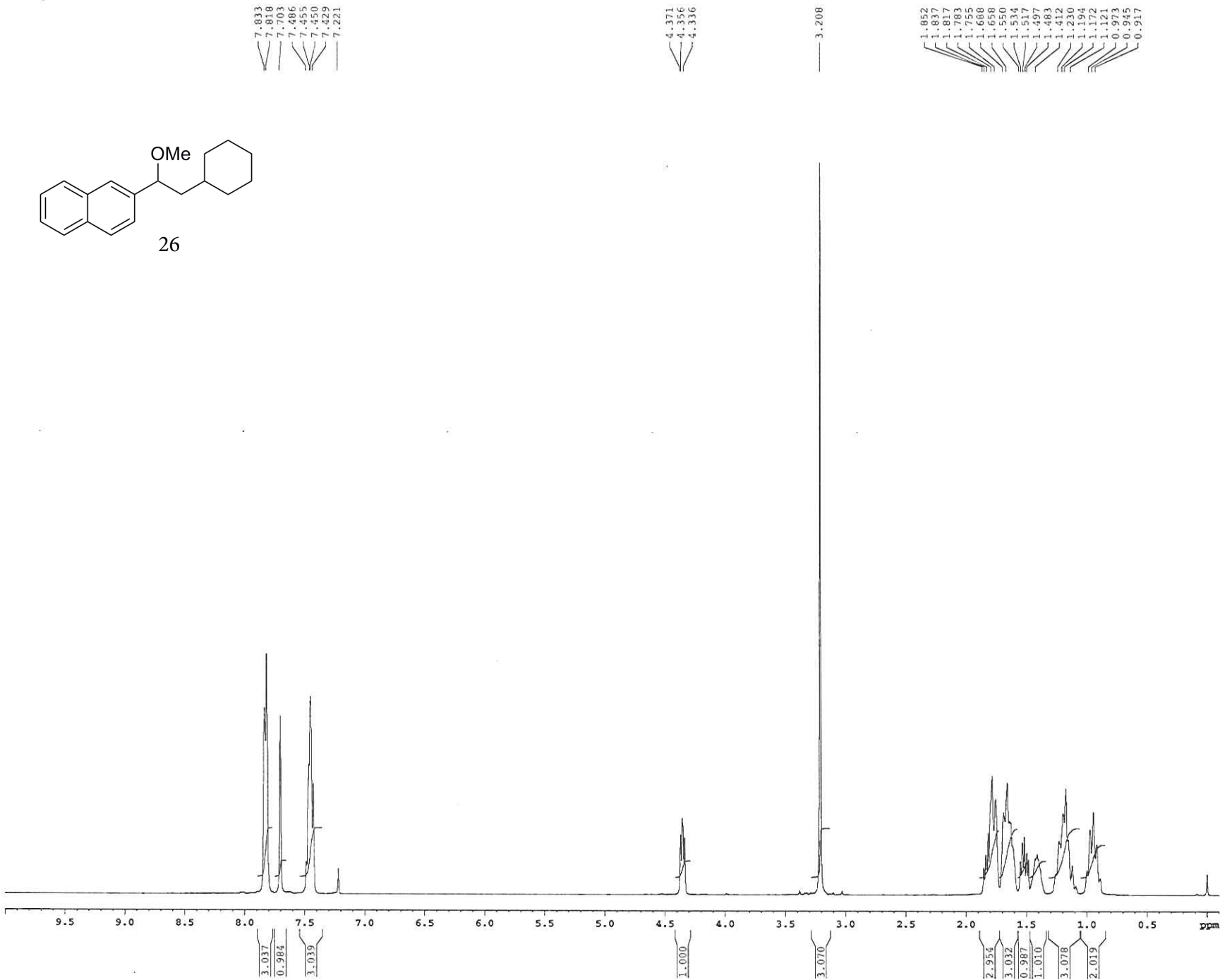
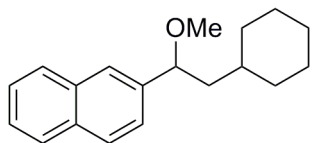
C(5')	28(1)	24(1)	19(1)	12(1)	6(1)	3(1)
C(6')	26(1)	29(1)	21(1)	12(1)	2(1)	6(1)
C(7')	37(1)	25(1)	24(1)	10(1)	5(1)	9(1)
C(8')	36(1)	26(1)	19(1)	9(1)	0(1)	-2(1)
C(9')	22(1)	39(1)	22(1)	14(1)	1(1)	2(1)
C(10')	29(1)	30(1)	24(1)	13(1)	10(1)	9(1)
C(11')	21(1)	21(1)	16(1)	10(1)	4(1)	4(1)
C(12')	18(1)	23(1)	17(1)	10(1)	3(1)	4(1)
C(13')	21(1)	21(1)	15(1)	10(1)	5(1)	5(1)
C(14')	19(1)	24(1)	17(1)	11(1)	4(1)	5(1)
C(15')	18(1)	27(1)	20(1)	12(1)	3(1)	8(1)
C(16')	24(1)	21(1)	22(1)	9(1)	7(1)	10(1)
C(17')	20(1)	23(1)	19(1)	9(1)	3(1)	7(1)
C(18')	28(1)	22(1)	21(1)	8(1)	7(1)	8(1)
C(19')	24(1)	24(1)	16(1)	4(1)	0(1)	2(1)
C(20')	20(1)	27(1)	18(1)	9(1)	2(1)	6(1)
C(21')	28(1)	18(1)	19(1)	6(1)	4(1)	6(1)
C(22')	22(1)	18(1)	17(1)	5(1)	4(1)	4(1)
C(23')	23(1)	27(1)	23(1)	11(1)	7(1)	9(1)
C(24')	27(1)	29(1)	26(1)	15(1)	7(1)	10(1)
C(25')	30(1)	36(1)	23(1)	15(1)	8(1)	11(1)
C(26')	27(1)	33(1)	25(1)	12(1)	11(1)	9(1)
C(27')	27(1)	23(1)	22(1)	6(1)	7(1)	8(1)

Table 5. Hydrogen coordinates ($\times 10^4$) and isotropic displacement parameters ($\text{\AA}^2 \times 10^{-3}$) for Jarvo02.

	x	y	z	U(eq)
H(1)	9972	2865	9422	24
H(2A)	10378	2122	10774	27
H(2B)	7583	1624	10479	27
H(3A)	10576	1106	9092	25
H(3B)	7846	532	8931	25
H(4A)	11504	466	10479	32
H(4B)	8780	-144	10268	32
H(6)	7500	-1873	9104	27
H(7)	8192	-3260	7893	32
H(8)	11636	-3041	7260	35
H(9)	14362	-1419	7824	33
H(10)	13667	-32	9025	31
H(12)	12031	4461	10582	22
H(15)	5797	4654	12280	25
H(16)	5504	3215	10921	24
H(17)	14359	6200	11771	24
H(18)	14659	7645	13142	27
H(19)	11522	7743	14001	28
H(20)	8121	6405	13482	27
H(21A)	6416	1730	8401	25
H(21B)	4988	2127	9248	25
H(22)	5437	3736	8972	23
H(23A)	8240	3016	7546	26
H(23B)	9192	3979	8572	26
H(24A)	6906	4953	7920	29
H(24B)	8515	4567	7142	29
H(25A)	5272	3218	6086	32
H(25B)	4559	4303	6268	32
H(26A)	1488	3019	6480	30
H(26B)	2496	3980	7506	30

H(27A)	3735	2022	7114	27
H(27B)	2157	2418	7901	27
H(1')	10945	490	3617	25
H(2'A)	10880	1318	2357	29
H(2'B)	8727	1763	2700	29
H(3'A)	13209	2320	4063	28
H(3'B)	11102	2878	4216	28
H(4'A)	13979	2994	2755	33
H(4'B)	11710	3462	2783	33
H(6')	11971	5113	3971	30
H(7')	14190	6640	5217	34
H(8')	18067	6770	5981	34
H(9')	19713	5351	5495	34
H(10')	17498	3824	4239	31
H(12')	10291	-1142	2470	22
H(15')	2720	-1095	966	25
H(16')	4978	291	2321	25
H(17')	9809	-2908	1246	24
H(18')	7540	-4306	-113	27
H(19')	3744	-4283	-886	27
H(20')	2243	-2859	-289	26
H(21C)	9433	1592	4813	26
H(21D)	6892	1219	4068	26
H(22')	6310	-444	4239	23
H(23C)	10234	-608	4486	28
H(23D)	10908	277	5567	28
H(24C)	7677	-1729	5083	31
H(24D)	10247	-1361	5815	31
H(25C)	9015	-132	7019	34
H(25D)	7232	-1251	6771	34
H(26C)	5121	53	6777	32
H(26D)	4417	-837	5696	32
H(27C)	7651	1167	6169	28
H(27D)	5081	796	5439	28

1H spectrum



7.833
7.818
7.703
7.486
7.455
7.450
7.329
7.221

4.371
4.356
4.336

3.208

1.852
1.817
1.817
1.783
1.783
1.635
1.638
1.538
1.530
1.514
1.497
1.483
1.412
1.212
1.190
1.174
1.172
0.973
0.943
0.917

```

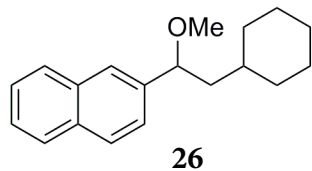
Current Data Parameters
=====
NAME      AGJ_1_190
EXPNO     3
PROCNO    1

F2 - Acquisition Parameters
Date_     20120423
Time      18.59
INSTRUM   dxt400
PROBHD    5 mm QNP 1H/1
PULPROG   zg30
TD         65536
SOLVENT   CDCl3
NS         8
DS         2
SWH        6410.256 Hz
FIDRES     0.097813 Hz
AQ         5.1118579 sec
RG         32
DW         78.000 usec
DE         4.50 usec
TE         298.1 K
D1         0.10000000 sec
MCREST    0.00000000 sec
MCWRX     0.01500000 sec

===== CHANNEL f1 =====
NUC1       1H
PI         12.00 usec
PL1        -0.60 dB
SFO1       400.1328009 MHz

F2 - Processing parameters
SI         65536
SF         400.1300372 MHz
WDW        EM
SCB        0
LB         0.30 Hz
GB         0
PC         2.00
    
```

13C spectrum with 1H decoupling

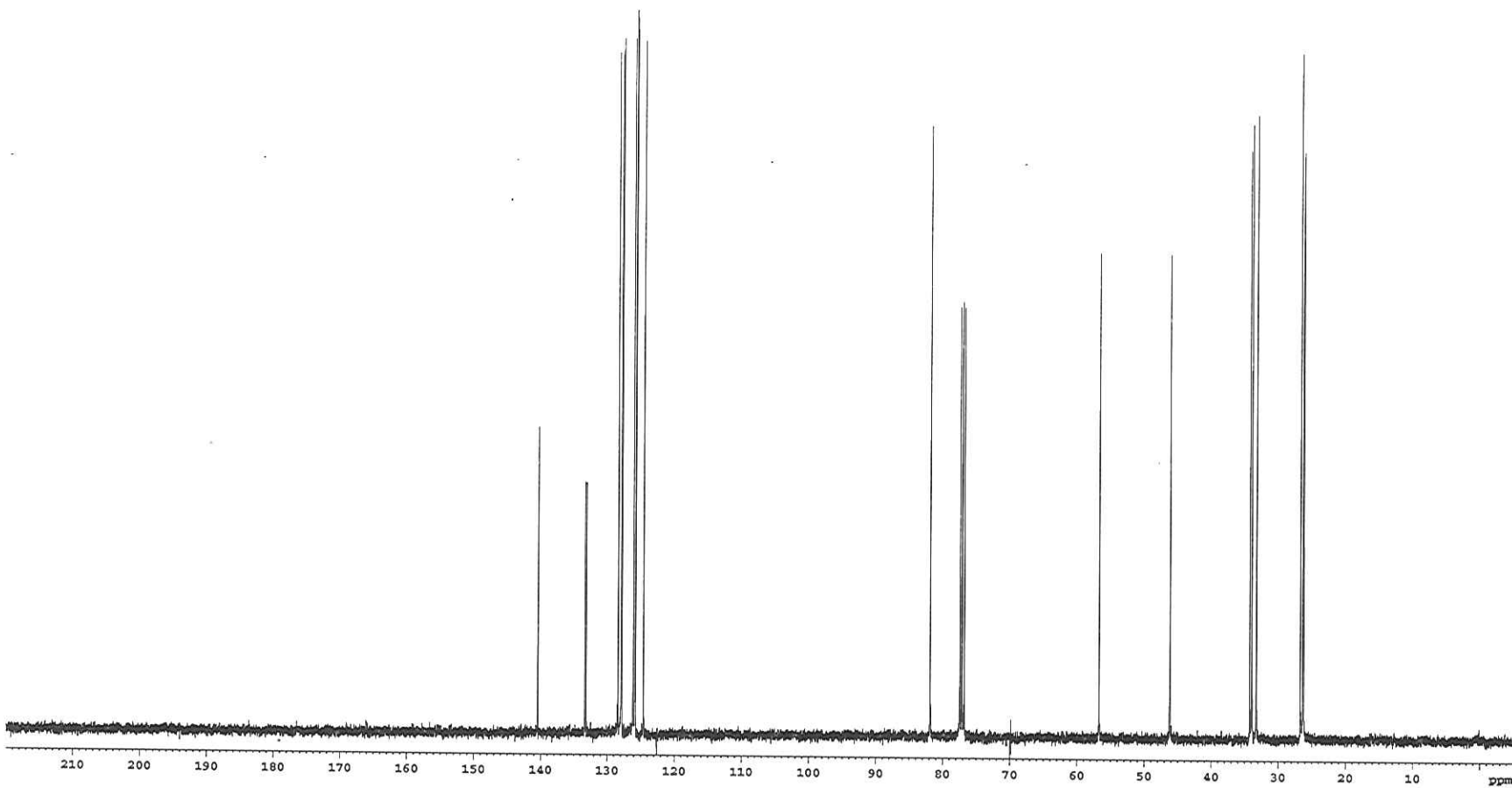


140.453
133.397
133.199
128.441
127.938
127.842
126.155
125.853
125.808
124.636

81.889
77.479
77.161
76.844

56.754
46.210

34.255
33.968
33.287
26.734
26.384
26.299



Current Data Parameters
USER Aaron1
NAME AQ1_1100
EXPRO 2
PROCNO 1

F2 - Acquisition Parameters
Date_ 20120421
Time 17.52
INSTRUM drx400
PROBHD 5 mm QNP H/F/P
PULPROG zgpg30
TD 65536
SOLVENT CDCl3
NS 614
DS 4
SWH 24154.590 Hz
FIDRES 0.368570 Hz
AQ 1.3566452 sec
RG 9195.2
EW 20.700 usec
DE 20.39 usec
TE 298.0 K
D1 0.10000000 sec
d11 0.03000000 sec
MCHRG1 0.00000000 sec
MCHRG2 0.01500000 sec

==== CHANNEL f1 =====
NUC1 13C
P1 11.00 usec
PL1 0.00 dB
SFO1 100.6237964 MHz

==== CHANNEL f2 =====
CPDPRG2 mlev16
NUC2 1H
PCPD2 80.00 usec
PL2 0.00 dB
PL12 16.20 dB
SFO2 400.1328009 MHz

F2 - Processing parameters
SI 65536
SF 100.6127635 MHz
WFW 84
SSB 0
LB 1.00 Hz
GB 0
PC 1.00

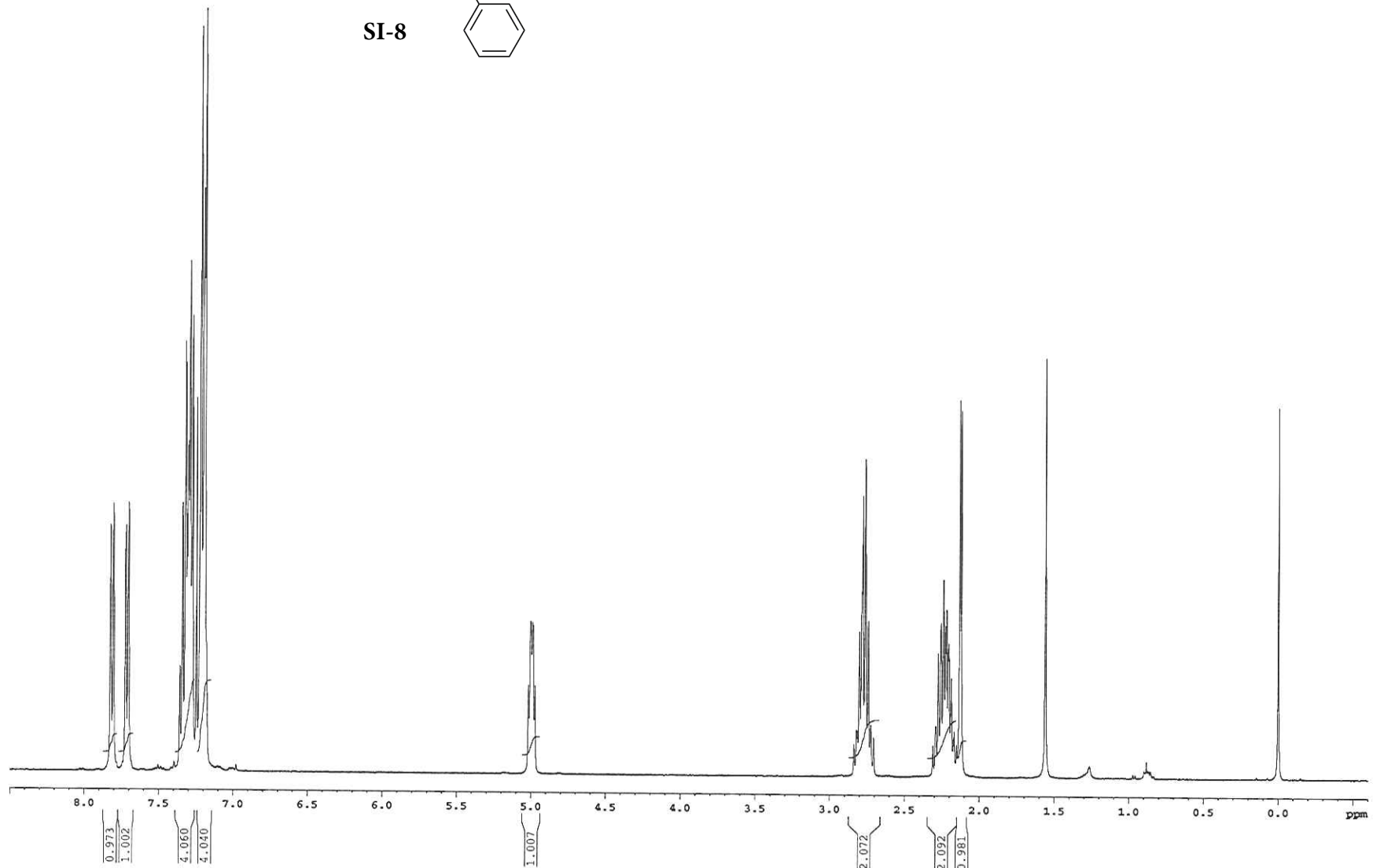
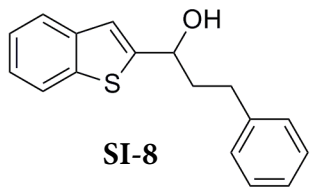
1H spectrum

7.824
7.806
7.724
7.722
7.704
7.357
7.342
7.339
7.324
7.319
7.311
7.304
7.292
7.274
7.247
7.221
7.202
7.191

5.018
5.003
4.991
4.975

2.837
2.821
2.814
2.802
2.786
2.780
2.763
2.741
2.725
2.707
2.310
2.292
2.276
2.257
2.234
2.241
2.225
2.219
2.209
2.204
2.187
2.175
2.169
2.153
2.132
2.122
1.559

-0.000



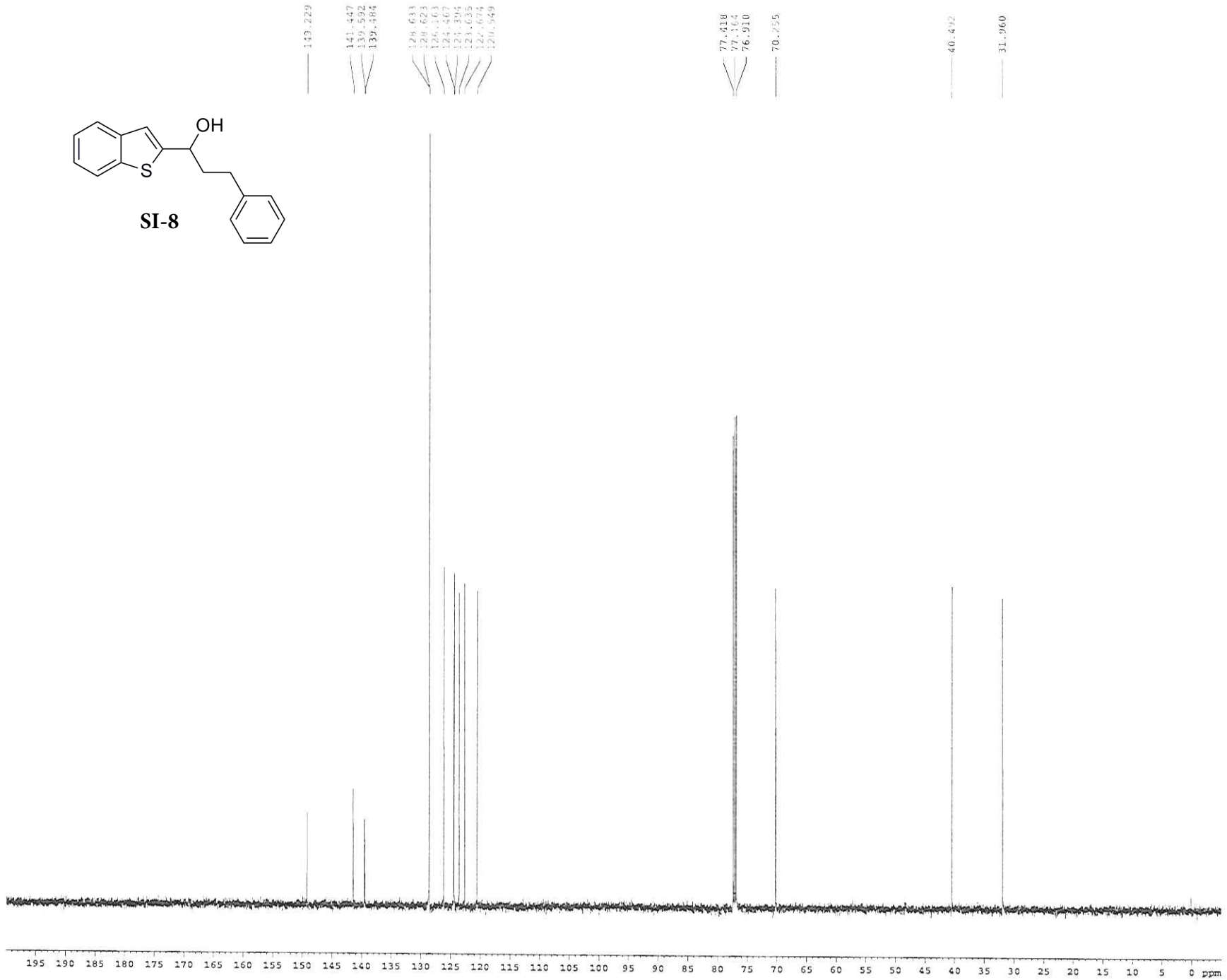
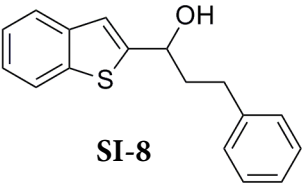
Current Data Parameters
 USER yonova
 NAME IMV5 - 102
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20121201
 Time_ 21.27
 INSTRUM dmx400
 PROBRD 5 mm QNP H/P/P
 PULPROG zg30
 TD 65536
 SOLVENT CDCl3
 NS 8
 DS 2
 SWH 6410.256 Hz
 FIDRES 0.097813 Hz
 AQ 5.1118579 sec
 RG 287.4
 DW 78.000 usec
 DE 4.50 usec
 TE 298.1 K
 D1 0.10000000 sec
 MCHES1 0.00000000 sec
 MCWRR 0.01500000 sec

===== CHANNEL f1 =====
 NUC1 1H
 P1 12.00 usec
 PL1 -0.60 dB
 SFO1 400.1328009 MHz

F2 - Processing parameters
 SI 65536
 SF 400.1300265 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 2.00

Z-restored spin-echo 13C spectrum with 1H decoupling



```

Current Data Parameters
=====
USER          yonova
NAME          1005 - 120
EXPNO        111
PROCNO       1

F2 - Acquisition Parameters
=====
Date_         20121203
Time_        14.29
INSTRUM      cryo500
PULPROG      5 mm CPDPR 1H
SOLVENT      SpinEchop13cp.prd
TD           65536
SOLVENT      CDCl3
NS           270
DS           16
SWH          30303.031 Hz
FIDRES       0.462388 Hz
AQ           1.0814103 sec
RG           2895.3
DS           16.500 usec
DE           6.00 usec
TE           293.2 K
D1           0.25000000 sec
d11          0.03000000 sec
D15         0.00000000 sec
d17         0.00000000 sec
MCARBST     0.00000000 sec
XCFWXR      0.01500000 sec
F2           31.00 usec

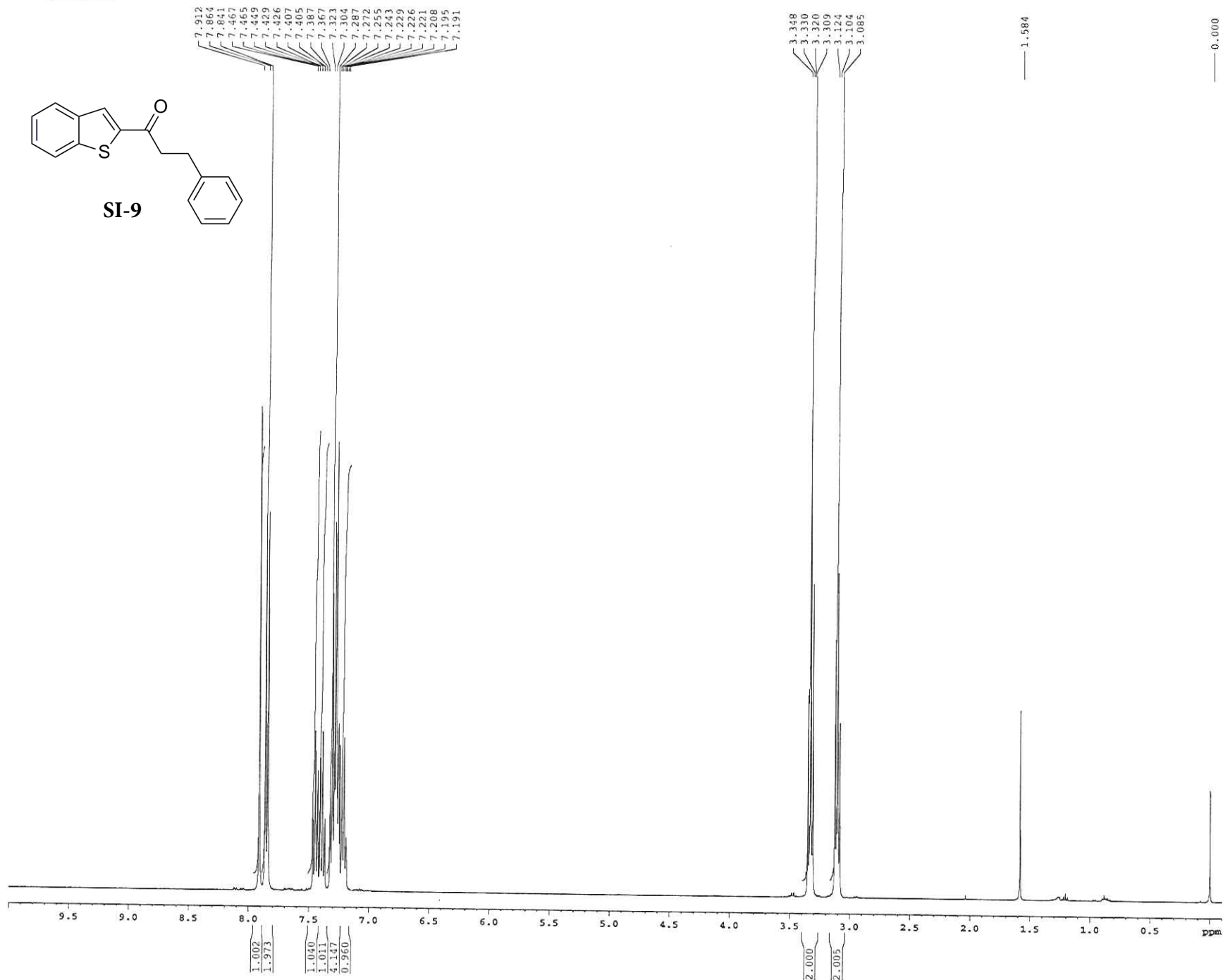
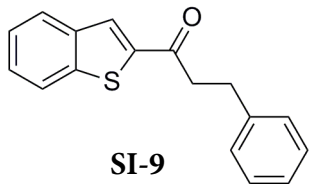
===== CHANNEL f1 =====
NUC1         13C
P1          15.50 usec
PL1         500.00 dB
PL2         2050.00 usec
PL0         120.00 dB
PL1         -1.00 dB
SFO1        125.7942548 MHz
CH1         3.20 dB
SP2         3.20 dB
SFO1        Crp50, 0.5, 20.1
SFO2        Crp60comp, 4
SFO3        0.00 Hz
SFO4        0.00 Hz

===== CHANNEL f2 =====
CPDPRG2     wa1r16
NUC2         1H
PCPD2       100.00 usec
PL2         1.60 dB
SFO2        500.2225011 MHz

===== GRADIENT CHANNEL =====
GPMAX1      SINE.100
GPMAX2      SINE.100
GPE1        0.00 %
GPE2        0.00 %
GPE3        0.00 %
GPE4        0.00 %
GPE5        0.00 %
GPE6        30.00 %
GPE7        50.00 %
GPE8        500.00 usec
GPE9        1000.00 usec

F2 - Processing parameters
=====
SI          65536
SF          125.7804094 MHz
WDW         EM
SSB         0
LB          1.00 Hz
GB          0
PC          2.00
    
```

1H spectrum



```

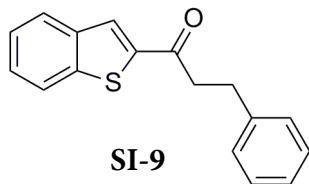
Current Data Parameters
USER          yonova
NAME         IMY5 - 080
EXPNO        1
PROCNO       1

F2 - Acquisition Parameters
Date_        20121121
Time         18.33
INSTRUM      dirx400
PROBHD       5 mm QNP H/P/P
PULPROG      zg30
TD           65536
SOLVENT      CDCl3
NS           8
DS           2
SWH          6410.256 Hz
FIDRES       0.097613 Hz
AQ           5.1118579 sec
RG           181
INW          78.000 usec
DE           4.50 usec
TE           298.1 K
D1           0.10000000 sec
d11          0.00000000 sec
MCREST       0.01500000 sec
MCWRK

===== CHANNEL f1 =====
NUC1         1H
P1           12.00 usec
PL1          -0.60 dB
SFO1         400.1328009 MHz

F2 - Processing parameters
SI           65536
SF           400.1300279 MHz
WDW          EM
SSB          0
LB           0.30 Hz
GB           0
PC           2.00
    
```


Z-restored spin-echo 13C spectrum with 1H decoupling

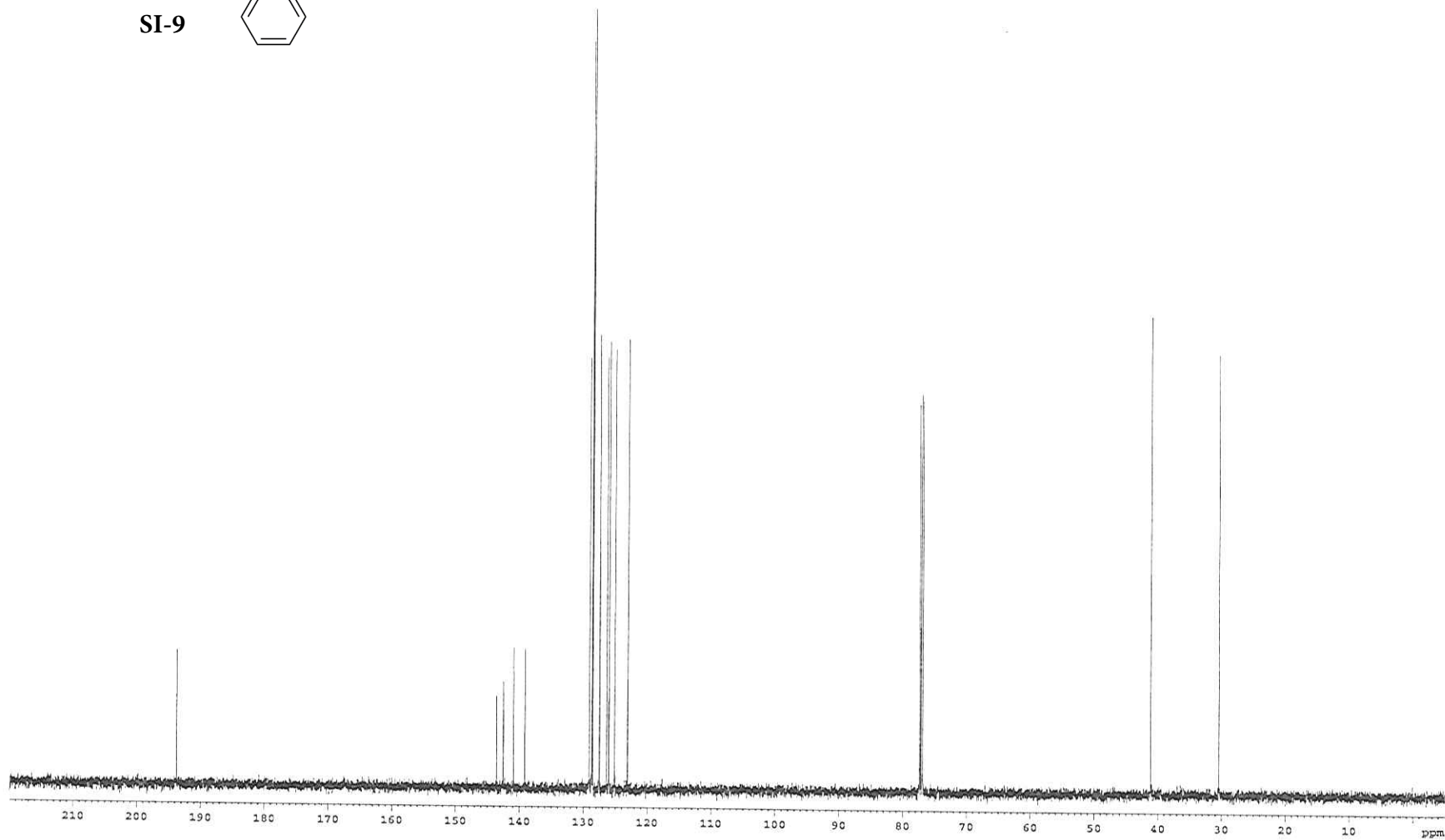


133.805
 143.645
 142.566
 140.990
 139.214
 129.100
 128.708
 128.552
 127.527
 126.396
 126.027
 125.120
 123.115

77.413
 77.159
 76.905

41.156

30.471



```

Current Data Parameters
=====
NAME      yonova
EXPNO     222
PROCNO    1

F2 - Acquisition Parameters
Date_     20121224
Time     15.58
INSTRUM   crys560
PROBHD    5 mm CPY13H-
PULPROG   SpinEcho140pp.prd
TD         65536
SOLVENT   CDCl3
NS         72
DS         16
SWH        30307.511 Hz
FIDRES     0.403389 Hz
AQ         1.0814105 sec
RG         6992
CIV        16.500 usec
DS         5.00 usec
TM         298.0 K
d1         0.25000000 sec
d11        0.33000000 sec
d16        0.20020000 sec
d17        0.00019600 sec
MCHRGFT   0.00000000 sec
MORPH     0.11000000 sec
PC         11.00 usec

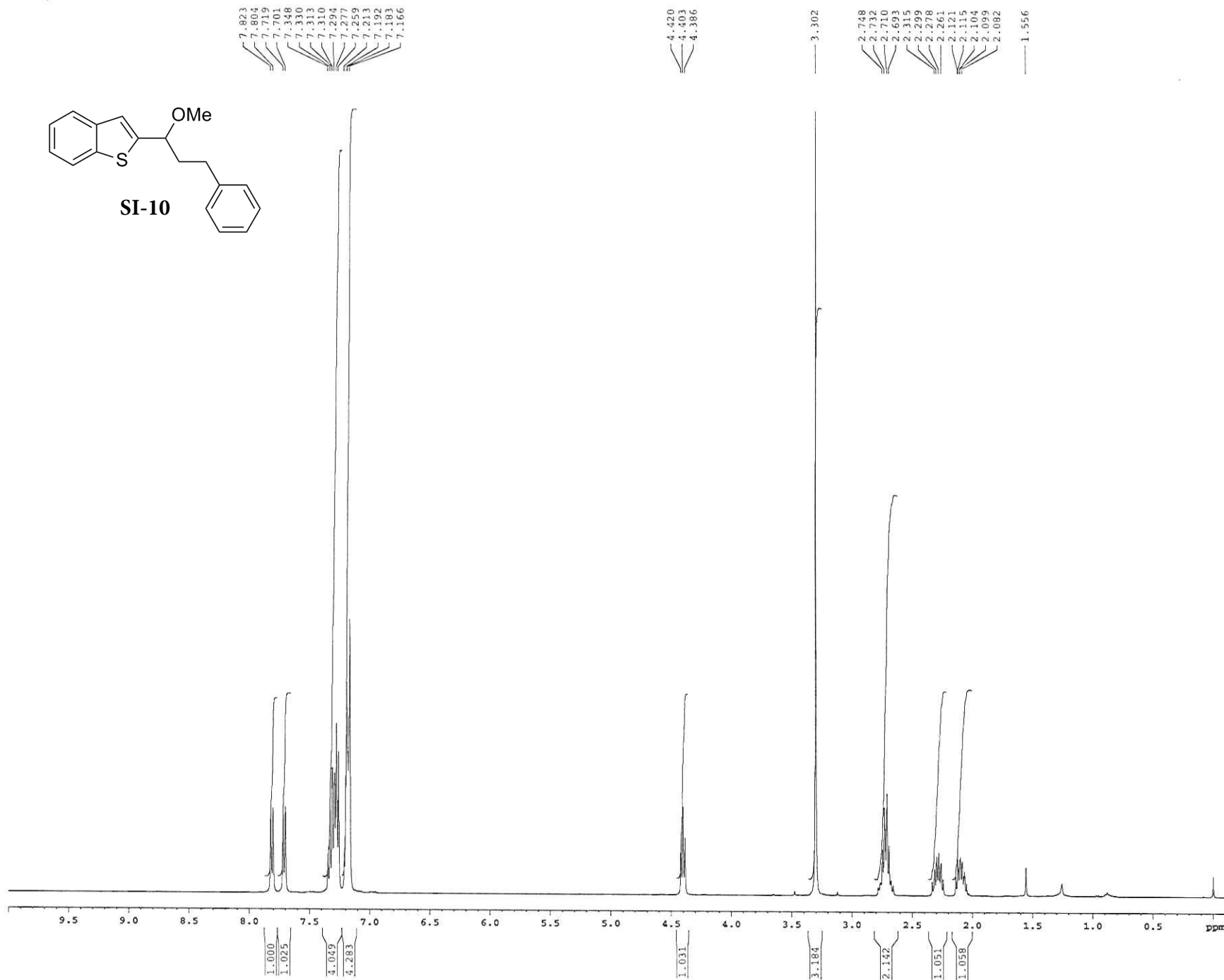
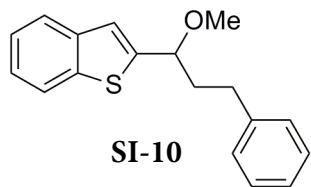
===== CHANNEL f1 =====
NUC1       13C
P1         15.50 usec
P11        500.00 usec
P12        2050.00 usec
PL0        12.00 dB
PL1        -1.00 dB
SFO1       125.7842548 MHz
SF1        3.20 GB
SFX2       3.20 GB
CPDNAME1   Crp60.0.5.70.1
CPDNAME2   Crp60comp.4
SFOFF1     0.00 Hz
SFOFF2     0.00 Hz

===== CHANNEL f2 =====
CPDPRG2    wait16
NUC2       1H
PCPD2     100.00 usec
PL2        1.60 dB
PL12       24.60 dB
SFO2       500.2225011 MHz

===== GRADIENT CHANNEL =====
GPNAM1     SINE.100
GPNAM2     SINE.100
GPX1       0.00 %
GPX2       0.00 %
GPY1       0.00 %
GPY2       0.00 %
GPZ1       30.00 %
GPZ2       50.00 %
D15        500.00 usec
D16        1000.00 usec

F2 - Processing parameters
SI         65536
SF         125.7804131 MHz
WDW        EM
SSB        C
LB         1.00 Hz
GB         C
PC         2.00
    
```

1H spectrum



```

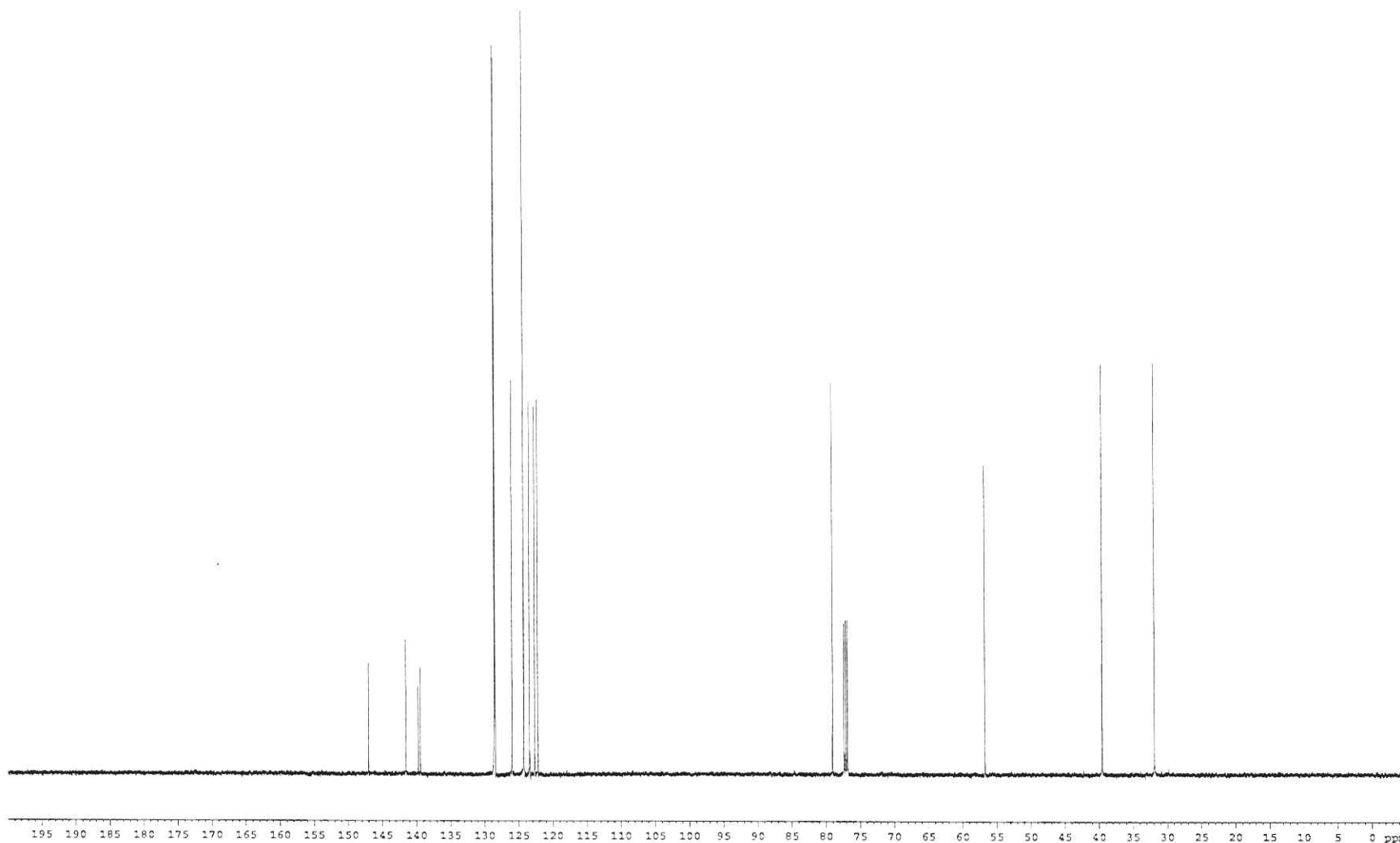
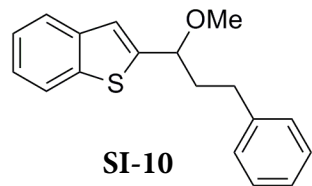
Current Data Parameters
USER yonova
NAME IMV5 - 128
EXPNO 1
PROCNO 1

F2 - Acquisition Parameters
Date_ 20121214
Time 16.21
INSTRUM drx400
PROBHD 5 mm QNP H/P/P
PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 8
DS 2
SWH 6410.256 Hz
FIDRES 0.097813 Hz
AQ 5.1118579 sec
RG 143.7
DM 78.000 usec
DE 4.50 usec
TE 298.1 K
D1 0.10000000 sec
MCHPRT 0.00000000 sec
MCMRK 0.01500000 sec

===== CHANNEL f1 =====
NUC1 1H
P1 12.00 usec
PL1 -0.60 dB
SFO1 400.1328009 MHz

F2 - Processing parameters
SI 65536
SF 400.1300398 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 2.00
    
```

Z-restored spin-echo 13C spectrum with 1H decoupling



Current Data Parameters
 USHR ymbo
 NAME 1MYS - 128
 EXPNO 222
 F2 SNO 1

F2 - Acquisition Parameters
 Date_ 20121214
 Time 17:17
 INSTRUM cryo500
 PROBRD 5 mm CPXI 1H-
 PULPROG SpinEcho90pp.prd
 PC 60.15
 SOLVENT CDCl3
 NS 222
 DS 16
 SWS 30703.011 Hz
 FIDRES 0.462388 Hz
 AQ 1.0814105 sec
 RG 3251
 DS 16.500 usec
 DE 6.00 usec
 TE 295.0 K
 D1 0.2500000 sec
 d11 0.0300000 sec
 D15 0.0002000 sec
 d17 0.0001900 sec
 NDBEST 0.0006000 sec
 NDBRK 0.0150000 sec
 F2 31.00 usec

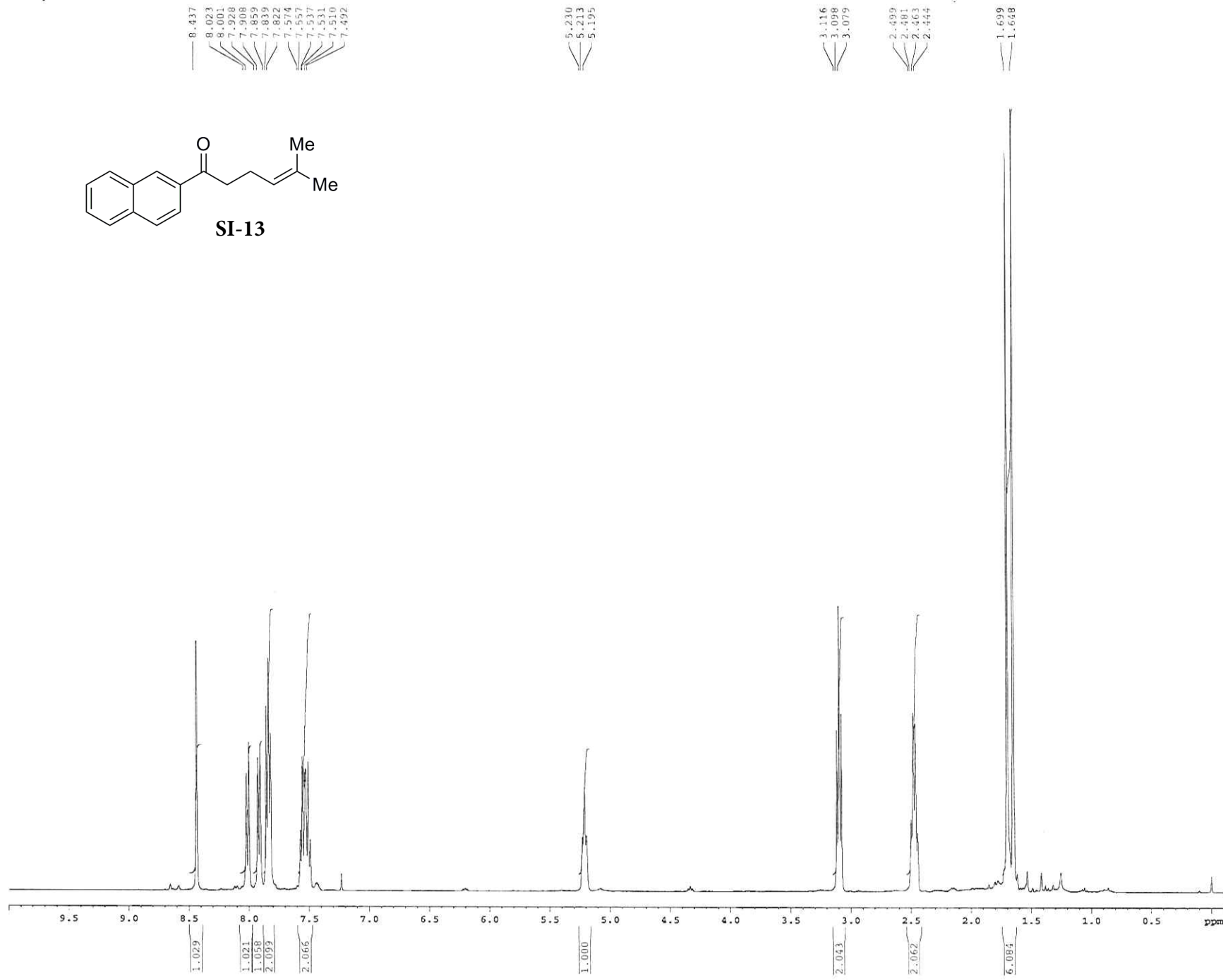
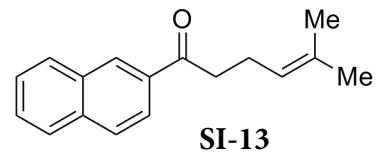
===== CHANNEL f1 =====
 NU1 13C
 P1 15.50 usec
 P11 500.00 usec
 P12 2000.00 usec
 PL0 120.00 dB
 PL1 -1.00 dB
 SFO1 125.764154 MHz
 SF1 3.20 dB
 SF2 3.20 dB
 SFO1 Crp60.0.5.20.1
 SFO2 Crp10comp.4
 SFO1 0.00 Hz
 SFO2 0.00 Hz

===== CHANNEL f2 =====
 CHOPRO2 walz16
 NU2 1H
 PCP2 100.00 usec
 PL0 1.00 dB
 PL1 24.50 dB
 SFO2 500.2225011 MHz

===== GRADIENT CHANNEL =====
 GPNAM1 SINE.100
 GPNAM2 SINE.100
 GPX1 0.00 %
 GPX2 0.00 %
 GPV1 0.00 %
 GPV2 0.00 %
 GPZ1 30.00 %
 GPZ2 50.00 %
 p15 500.00 usec
 p16 1000.00 usec

F2 - Processing parameters
 SI 65536
 SF 125.7604154 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 2.00

1H spectrum



```

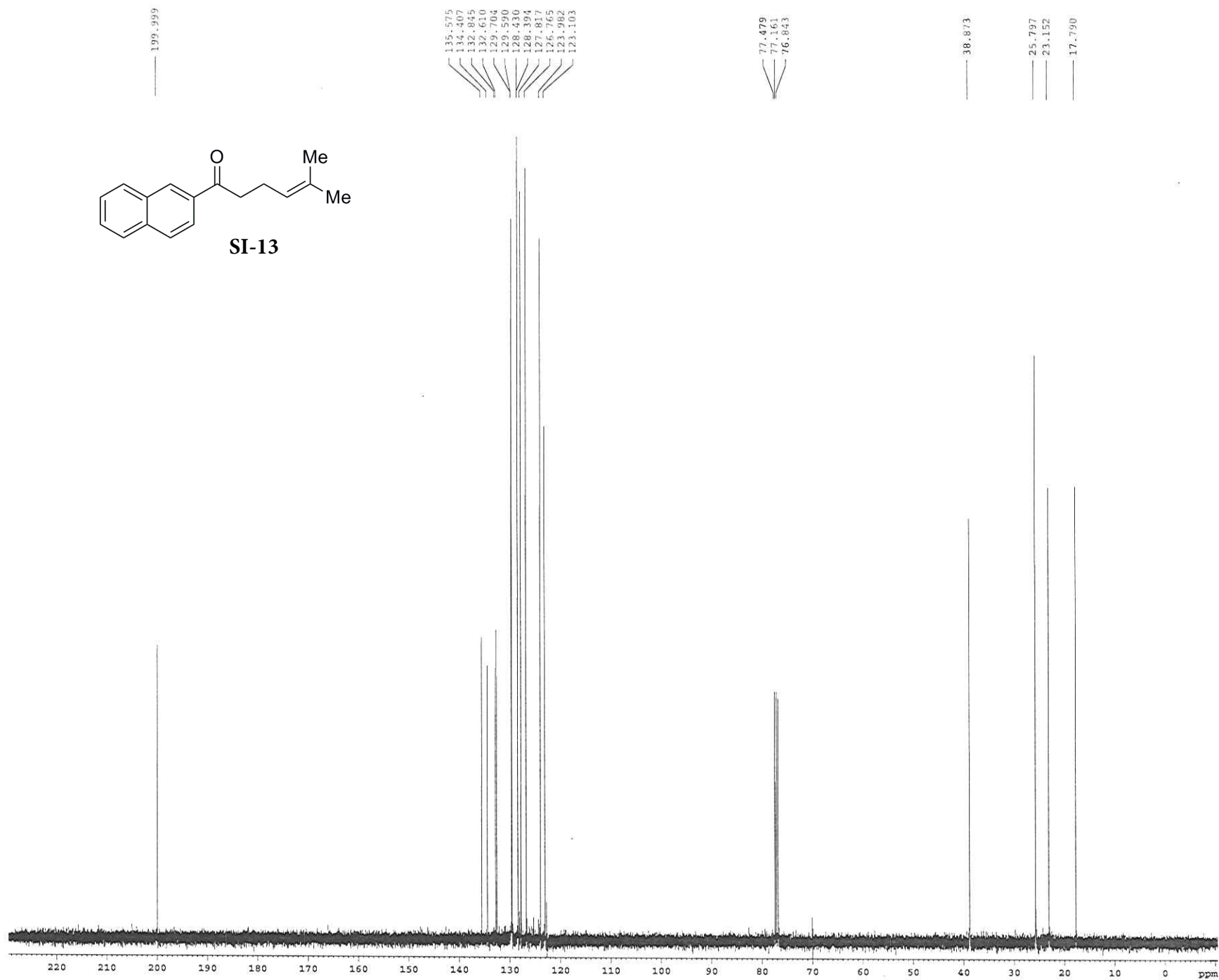
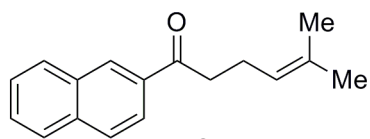
Current Data Parameters
=====
NAME      yonova
EXPNO    1
PROCNO   1

F2 - Acquisition Parameters
=====
Date_    20110615
Time     16.33
INSTRUM  drx400
PROBHD   5 mm QNP 1H/1
PULPROG  zg30
TD        65536
SOLVENT  CDCl3
DS        2
SWH       6410.256 Hz
FIDRES    0.097813 Hz
AQ        5.1118579 sec
RG        32
DW        78.000 usec
DE        4.50 usec
TE        298.1 K
D1        0.10000000 sec
MCREST    0.00000000 sec
MCMRPRK   0.01500000 sec

===== CHANNEL f1 =====
NUC1      1H
P1        12.00 usec
PL1       -0.60 dB
SFO1      400.1328009 MHz

F2 - Processing parameters
=====
SI        65536
SF        400.1300112 MHz
WDW       EM
SSB       0
LB        0.30 Hz
GB        0
PC        2.00
    
```

¹³C spectrum with ¹H decoupling



```

Current Data Parameters
USER      yonova
NAME      imy5 - AG3-3-45
EXPNO     2
PROCNO    1

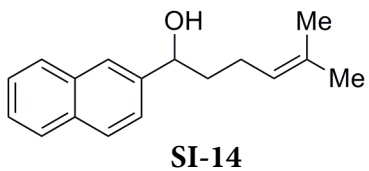
F2 - Acquisition Parameters
Date_     20100615
Time      18.35
INSTRUM   drx400
PROBHD    5 mm QNP H/P/9
PULPROG   zgpg30
TD         65536
SOLVENT   CDCl3
NS         339
DS         4
SWH        24154.590 Hz
FIDRES     0.369570 Hz
AQ         1.3566452 sec
RG         1149.4
EQ         20.700 usec
DE         20.19 usec
TS         298.0 K
DI         0.10000000 sec
d11        0.01000000 sec
MORPH     0.00000000 sec
MORPHK    0.01500000 sec

----- CHANNEL f1 -----
NUC1       13C
P1         11.00 usec
PL1        0.00 dB
SFO1       100.6237964 MHz

----- CHANNEL f2 -----
CPDPRG2   mlev16
NUC2       1H
PCPD2     80.00 usec
PL2        0.00 dB
PL12       16.20 dB
SFO2       400.1328009 MHz

F2 - Processing parameters
SI         65536
SF         100.6127663 MHz
WDW        no
SSB        0
LB         0.00 Hz
GB         0
PC         2.00
    
```

1H spectrum



7.825
7.805
7.760
7.473
7.468
7.458
7.450
7.432
7.239

5.176
5.159
5.141
4.830

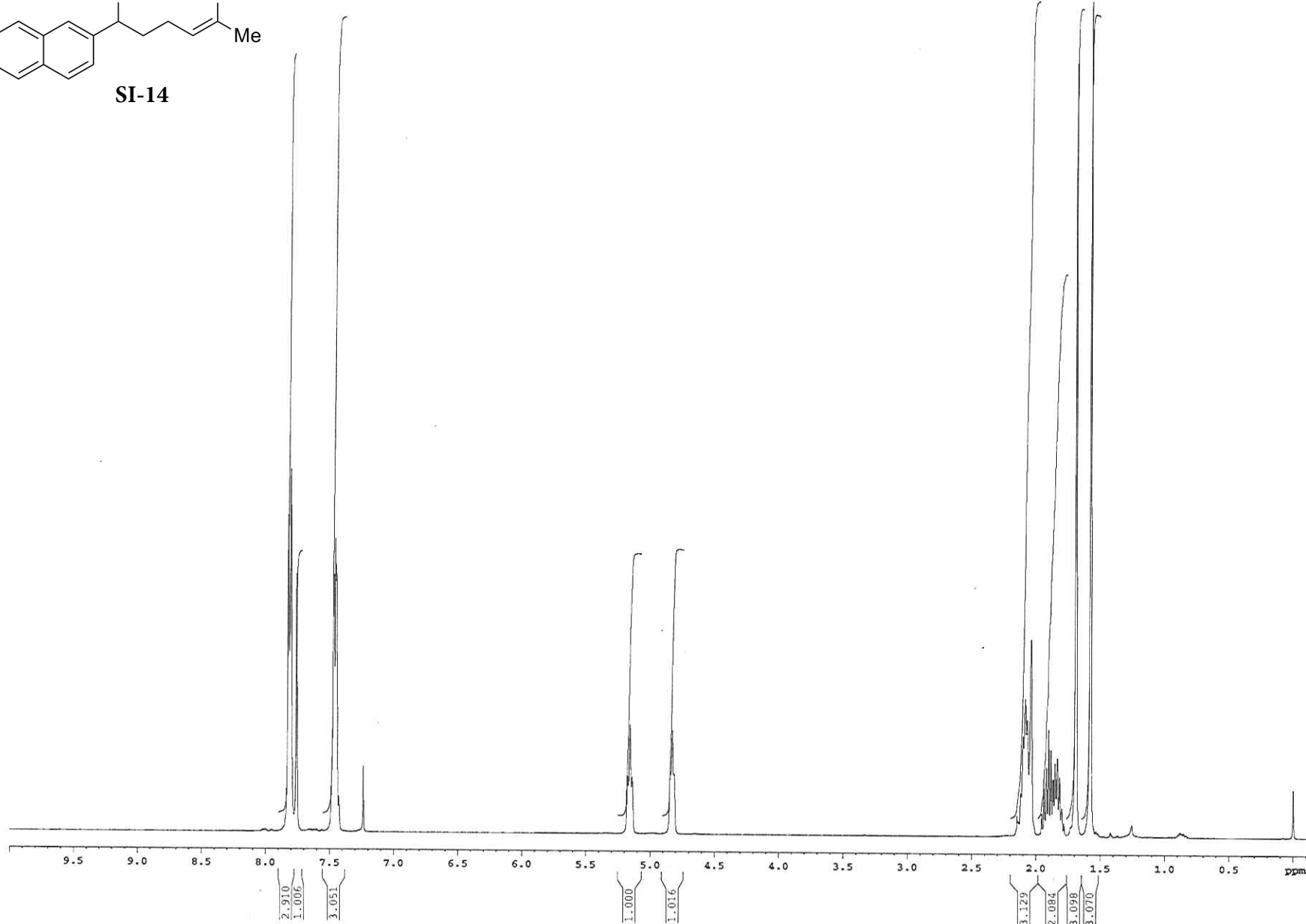
2.118
2.100
2.086
2.083
2.043
1.931
1.920
1.902
1.887
1.867
1.853
1.833
1.691
1.578

Current Data Parameters
 USER aaronj
 NAME AGJ_1_33_c1
 EXPNO 1
 PROCNO 1

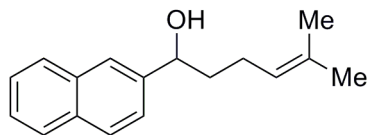
F2 - Acquisition Parameters
 Date_ 20110511
 Time 18.58
 INSTRUM drx400
 PROBHD 5 mm QNP H/P/P
 PULPROG zg30
 TD 65536
 FIDRES 0.097813 Hz
 AQ 5.1118579 sec
 RG 101.6
 DW 4.50 usec
 DE 78.000 usec
 TE 298.2 K
 D1 0.10000000 sec
 MCHRECT 0.00000000 sec
 XCHRX 0.01500000 sec

===== CHANNEL f1 =====
 NUCL 1H
 PL 12.00 usec
 PL1 -0.60 dB
 SFO1 400.1328009 MHz

F2 - Processing parameters
 SI 65536
 SF 400.1300296 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 2.00



11B spectrum with 1H decoupling, using composite pulse for background suppression



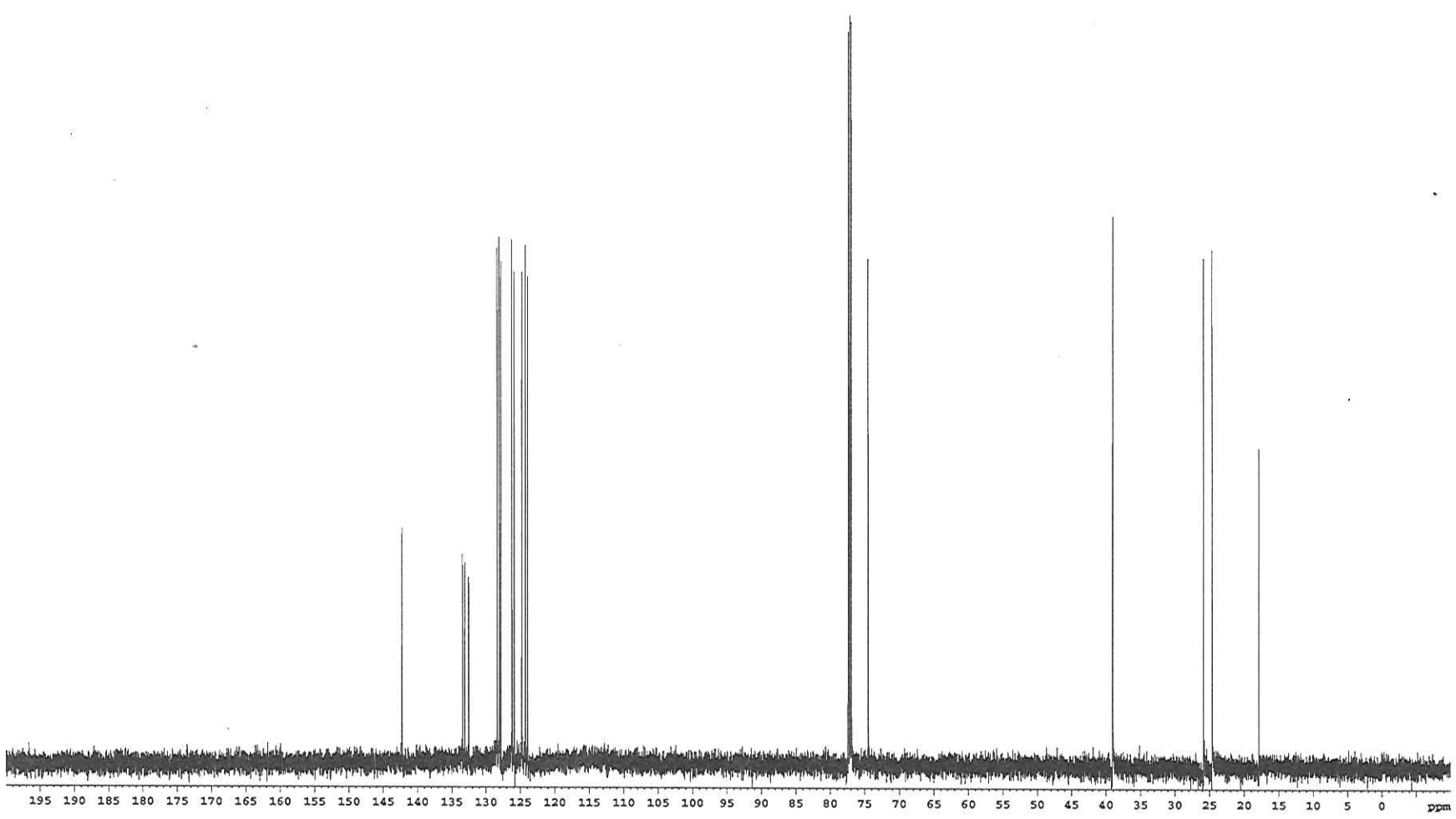
142.264
133.437
133.105
132.524
128.370
128.056
127.809
126.227
125.888
124.741
124.260
123.934

77.370
77.159
76.947
74.502

39.031

25.872
24.639

17.891



```

Current Data Parameters
USER      aaronj
NAME      AGJ_3_13
EXPNO     5
PROCNO    1

F2 - Acquisition Parameters
Date_     20130511
Time      19.24
INSTRUM   av600
PROBHD    5 mm TBI 1H/13
PULPROG   zgpg30
TD         65400
SOLVENT   CDCl3
NS         550
DS         4
SWH        36231.883 Hz
FIDRES     0.554004 Hz
AQ         0.9025700 sec
RG         1030
DW         13.800 usec
DE         6.00 usec
TE         298.2 K
D1         0.40000001 sec
D11        0.03000000 sec
TD0        1

===== CHANNEL f1 =====
NUC1       13C
P1         15.00 usec

F2 - Processing parameters
SI         65536
SP         150.9027869 MHz
WDW        EM
SSB        0
LB         1.00 Hz
GB         0
PC         1.00
    
```

1H spectrum

7.847
7.827
7.704
7.477
7.468
7.453
7.431
7.251

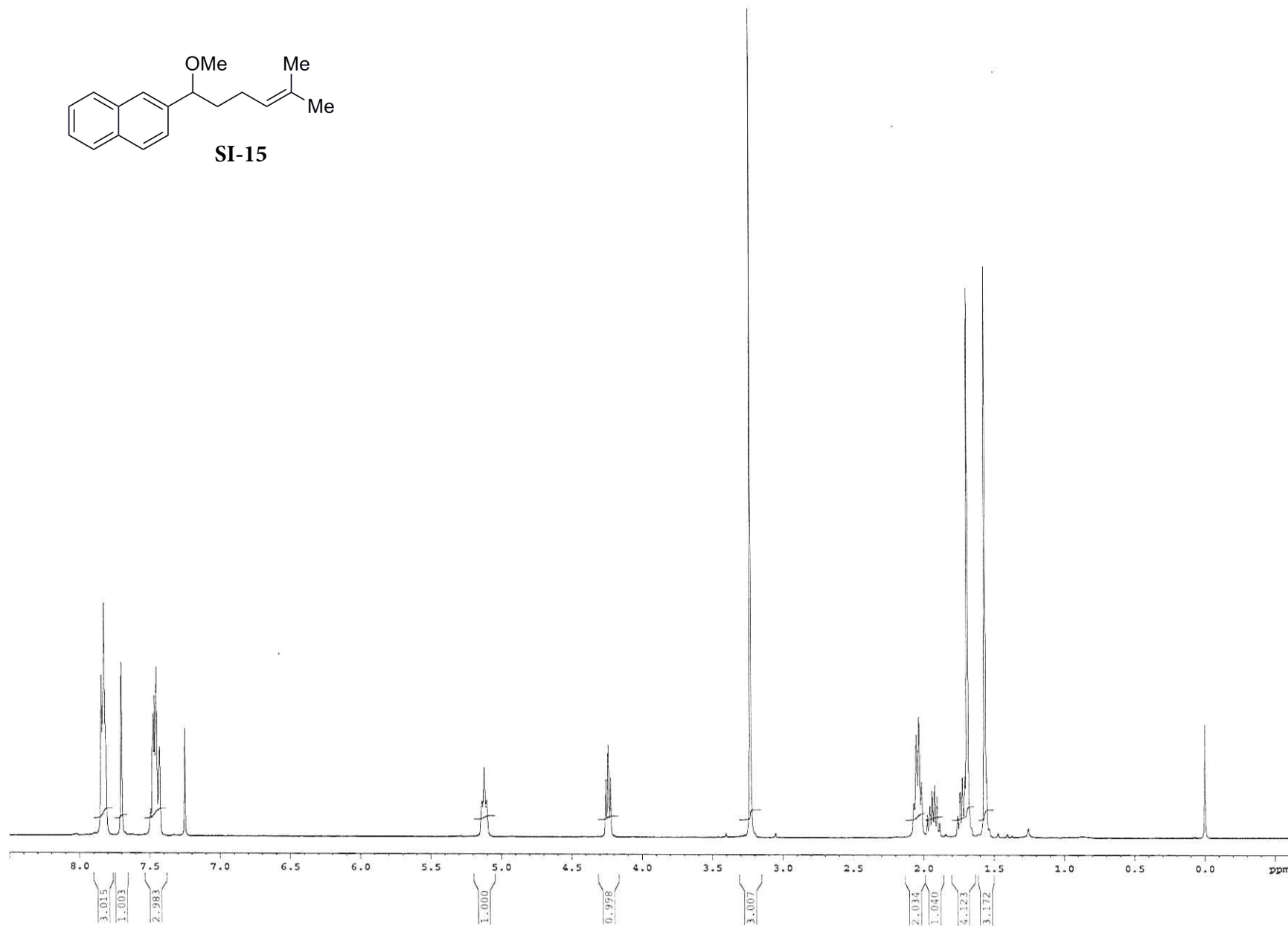
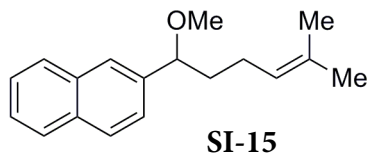
5.140
5.123
5.106

4.258
4.242
4.225

3.231

2.071
2.053
2.034
2.016
1.998
1.942
1.923
1.906
1.742
1.726
1.707
1.690
1.675
1.553

0.000



```

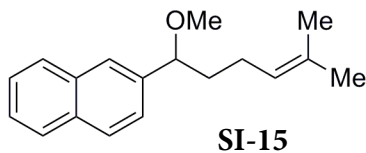
Current Data Parameters
=====
USER          anonj
NAME          AGC_3_19
EXTNO        1
PROCNO       1

F2 - Acquisition Parameters
Date_         20130517
Time          3.10
INSTRUM       dx400
PROBHD        5 mm QNP H/P/P
PULPROG       zgpg
TD            65536
SOLVENT       CDCl3
NS            2
DS            2
SWH           6410.254 Hz
FIDRES        0.097813 Hz
AQ            5.118579 sec
RG            203.1
DW            78.000 usec
DE            4.50 usec
TE            298.0 K
D1            0.10000000 sec
MCHEST        0.00000000 sec
MCVRK         0.01500000 sec

===== CHANNEL f1 =====
NUC1          1H
P1            12.00 usec
PL1           -0.60 dB
SFO1         400.1326009 MHz

F2 - Processing parameters
SI            65536
SF            400.1300246 MHz
WDW           EM
SSB           0
LB            0.30 Hz
GB            0
PC            2.00
    
```


Z-restored spin-echo 13C spectrum with 1H decoupling



139.966
133.360
133.217
132.181
126.411
127.945
127.852
126.175
126.078
125.838
124.648
124.031

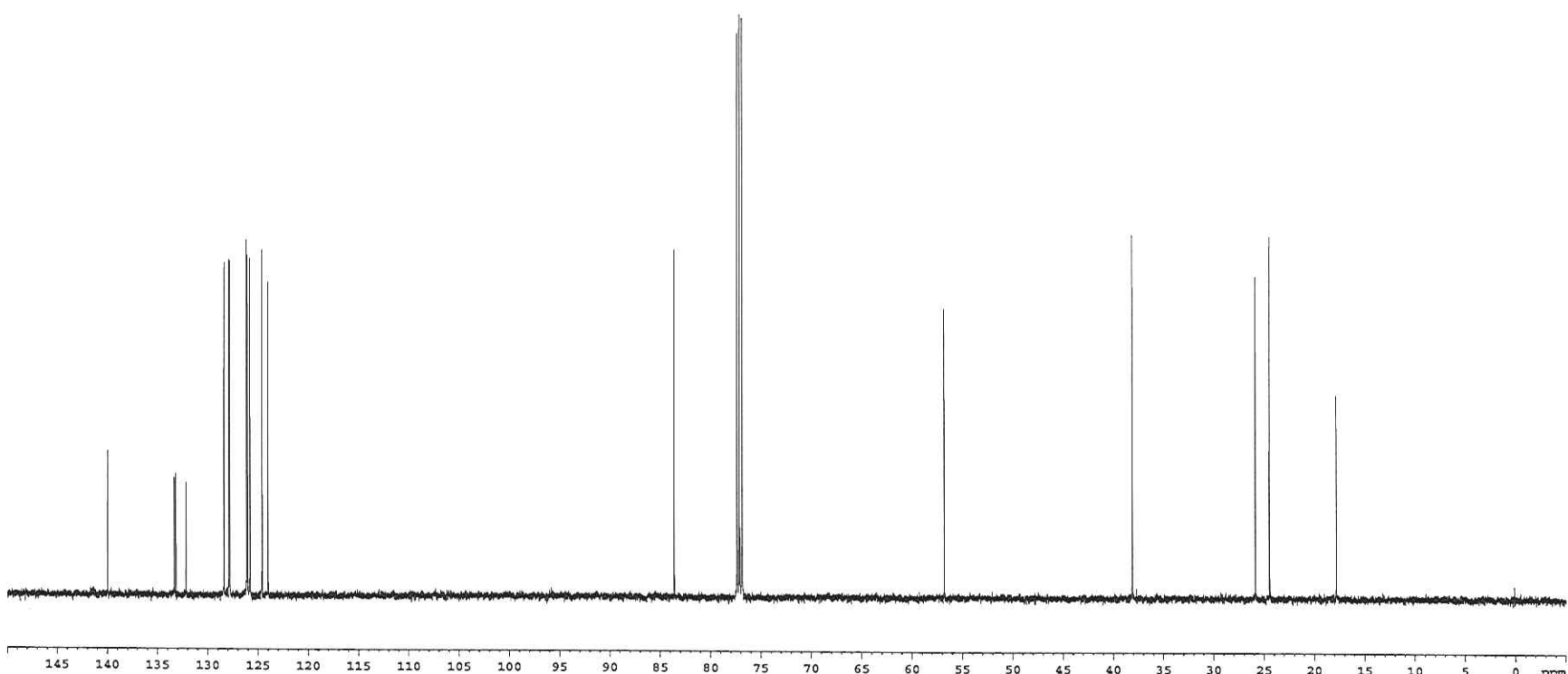
83.634
77.415
77.161
76.907

56.844

38.142

25.897
24.483

17.858



```

Current Data Parameters
USER          axonj
NAME          AGJ_1_39
EXPTNO       2
PROCNO       1

F2 - Acquisition Parameters
Date_        20130527
Time         5.48
INSTRUM      cryo500
PROBHD       5 mm CPTCI 1H-
PULPROG      SpinEchoq10op.prd
TD           65536
SOLVENT      CDCl3
NS           400
DS           16
SWH          30333.031 Hz
FIDRES       0.462388 Hz
AQ           1.0814105 sec
RG           8192
DE           16.500 usec
TE           298.0 K
DL           0.25000000 sec
d11          0.03000000 sec
d16          0.00020000 sec
d17          0.00019600 sec
MCRET       0.00000000 sec
MCWPK       0.01500000 sec
P2           31.00 usec

===== CHANNEL f1 =====
NUC1         13C
P1           15.50 usec
PL1          500.00 usec
P12          2000.00 usec
PL0          120.00 dB
PL1          -1.00 dB
SFO1         125.7942548 MHz
SP1          3.20 dB
SP2          3.20 dB
SFO1         Crp60.0.5.20.1
SFO2         Crp60comp.4
SFOFF1       0.00 Hz
SFOFF2       0.00 Hz

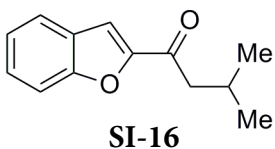
===== CHANNEL f2 =====
CPDPRG2      waltz16
NUC2         1H
P2           100.00 usec
PL2          1.60 dB
PL12         24.60 dB
SFO2         500.2225011 MHz

===== GRADIENT CHANNEL =====
GPNAM1       SINK.100
GPNAM2       SINK.100
GPX1         0.00 %
GPX2         0.00 %
GPY1         0.00 %
GPY2         0.00 %
GPZ1         30.00 %
GPZ2         50.00 %
p15          500.00 usec
p16          1000.00 usec

F2 - Processing parameters
SI           65536
SF           125.7804085 MHz
WDW          EM
GB           0
LB           1.00 Hz
CB           0
PC           2.00
    
```

1H spectrum

7.720
7.700
7.597
7.576
7.498
7.476
7.457
7.332
7.313
7.294
7.261



2.838
2.821
2.387
2.370
2.353
2.336
1.547
1.256
1.040
1.023

-0.000

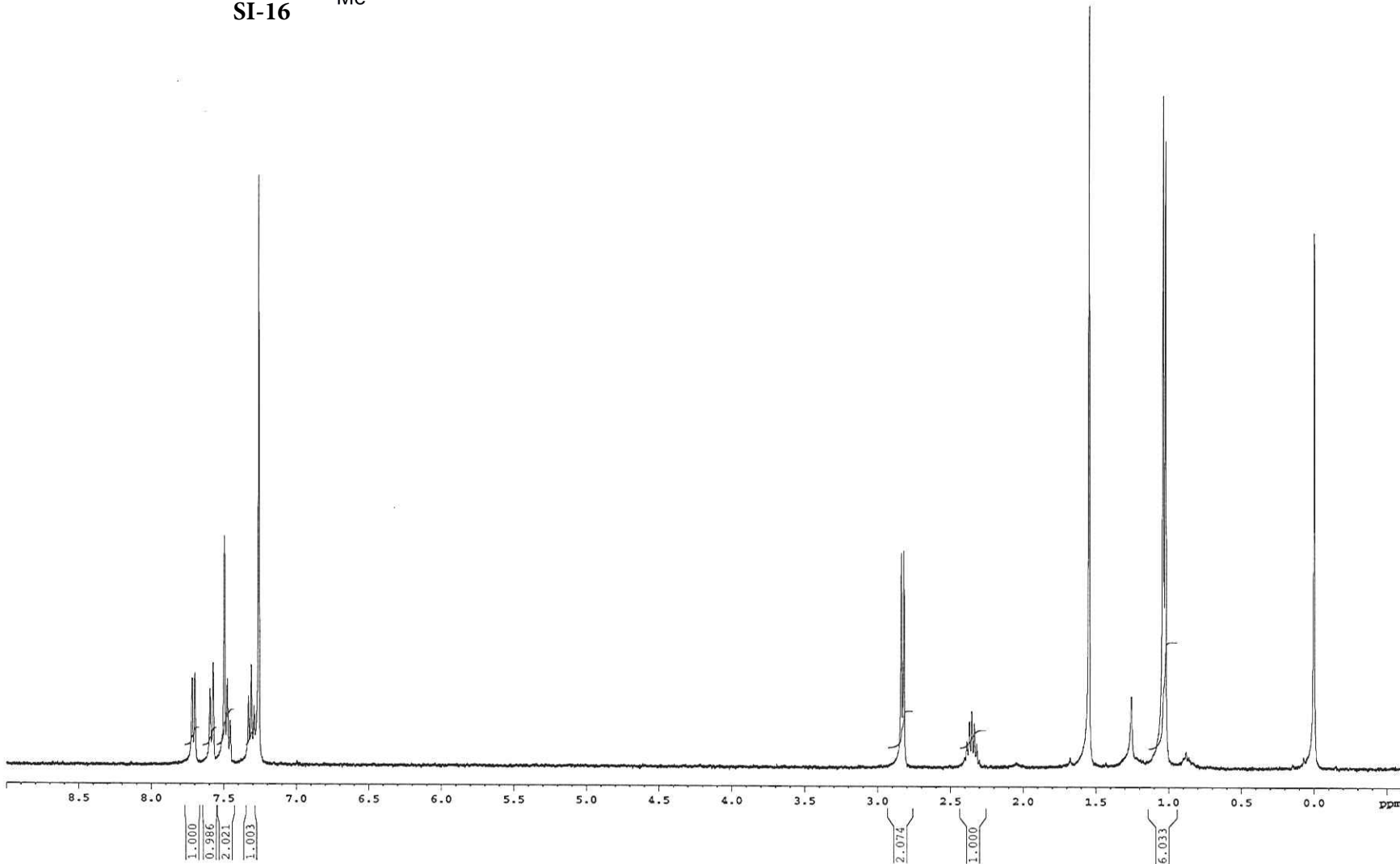
```

Current Data Parameters
USER          yonova
NAME         imy5 - 261
EXPNO       10
PROCNO      1

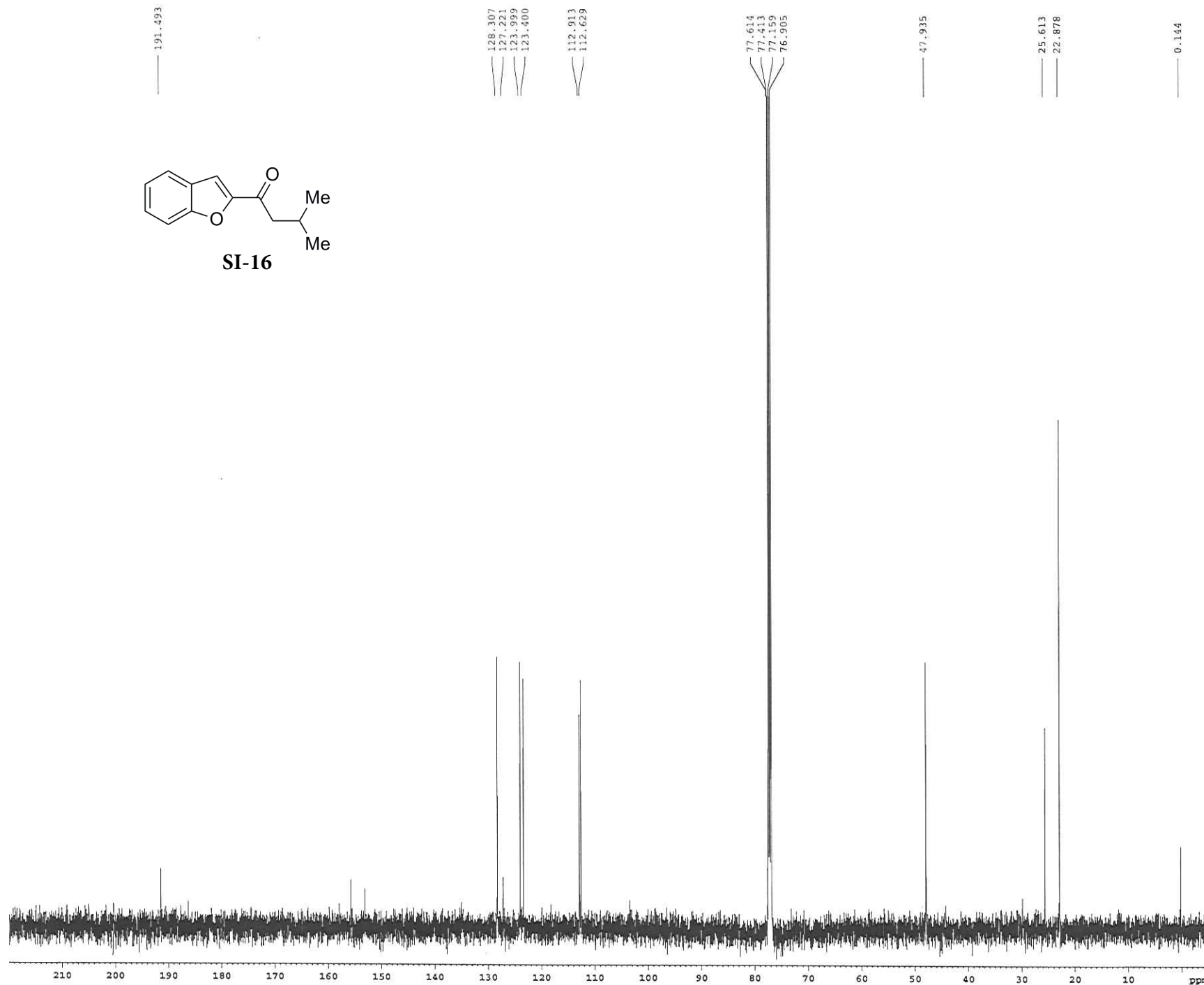
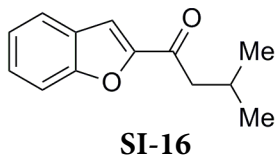
F2 - Acquisition Parameters
Date_       20130616
Time        18.41
INSTRUM     drx400
PROBHD      5 mm QNP 1H/1
PULPROG     zg30
TD          65536
SOLVENT     CDCl3
NS          8
DS          2
SWH         6410.256 Hz
FIDRES      0.097813 Hz
AQ          5.1118579 sec
RG          724.1
EW         78.000 usec
DE         4.50 usec
TE         298.0 K
D1         0.10000000 sec
MCHRG1     0.00000000 sec
MCHRG2     0.01500000 sec

===== CHANNEL f1 =====
NUC1        1H
P1          12.00 usec
PL1         -0.60 dB
SFO1        400.1328009 MHz

F2 - Processing parameters
SI          65536
CF          400.1300207 MHz
WDW         EM
SSB         0
LB          0.30 Hz
GB          0
PC          2.00
    
```



Z-restored spin-echo 13C spectrum with 1H decoupling



```

Current Data Parameters
USER          yonova
NAME         imy5 - 261
EXPNO        222
PROCNO       1

F2 - Acquisition Parameters
Date_        20130716
Time         19.02
INSTRUM      cryo500
PROBHD       5 mm CPCTC 1H-
PULPROG      SpinEcho30up.prd
TD           65536
SOLVENT      CDCl3
NS           3000
DS           16
SWH          30303.031 Hz
FIDRES       0.462388 Hz
AQ           1.0914105 sec
RG           4597.6
LW           16.500 usec
DE           6.00 usec
TE           298.0 K
D1           0.25000000 sec
d11          0.03000000 sec
D16          0.00020000 sec
d17          0.00019600 sec
MCHM2T       0.00000000 sec
MCHM2K       0.01500000 sec
P2           31.00 usec

===== CHANNEL f1 =====
NUC1          13C
P1           15.50 usec
PL1          500.00 usec
PI1          2000.00 usec
PL0          120.00 dB
PL1          -1.00 dB
SFO1         125.7942548 MHz
SF1          3.70 dB
SF2          3.20 dB
SPNAM1       Crp60,0.5,20.1
SPNAM2       Crp60comp.4
SFOFF1       0.00 Hz
SFOFF2       0.00 Hz

===== CHANNEL f2 =====
CPDPRG2      waltz16
NUC2          1H
PCPD2        500.00 usec
PL2          1.60 dB
PL12         24.60 dB
SFO2         500.2225011 MHz

===== GRADIENT CHANNEL =====
GPNAM1       SINE.100
GPNAM2       SINE.100
GPX1         0.00 %
GPX2         0.00 %
GPI1         0.00 %
GPI2         0.00 %
GPZ1         30.00 %
GPZ2         50.00 %
p15          500.00 usec
d15          1000.00 usec

F2 - Processing parameters
SI           65536
SF           125.7804076 MHz
WDW          EM
SSB          0
LB           1.00 Hz
GB           0
PC           2.00
    
```

1H spectrum

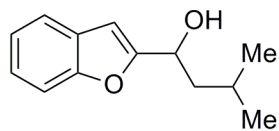
7.537
7.522
7.462
7.446
7.275
7.261
7.247
7.221
7.207
7.192

6.604

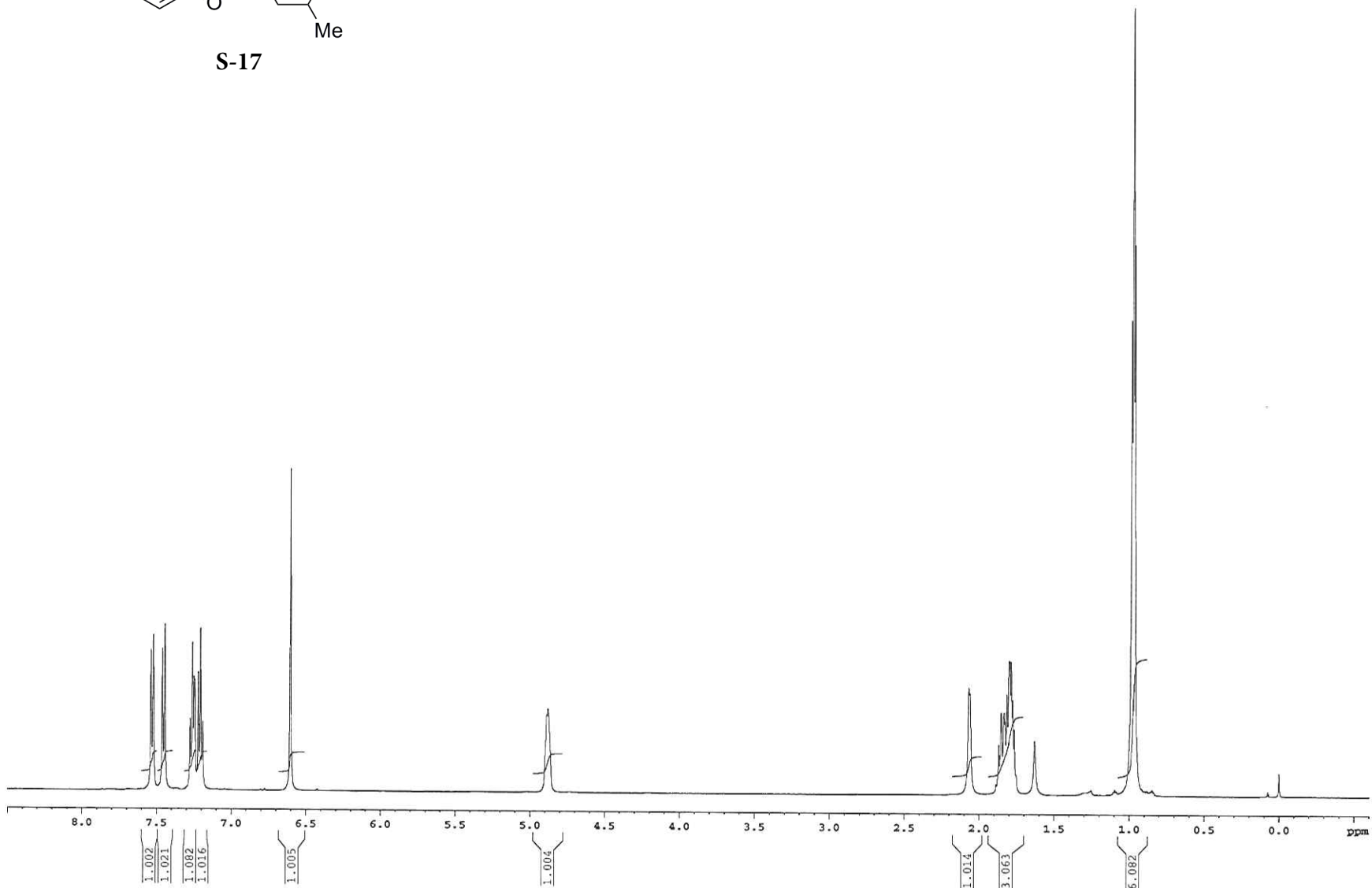
4.868
4.860

2.068
2.061
1.868
1.852
1.836
1.831
1.826
1.813
1.798
1.794
1.786
1.777
1.766
1.629

0.985
0.973
0.963



S-17



```

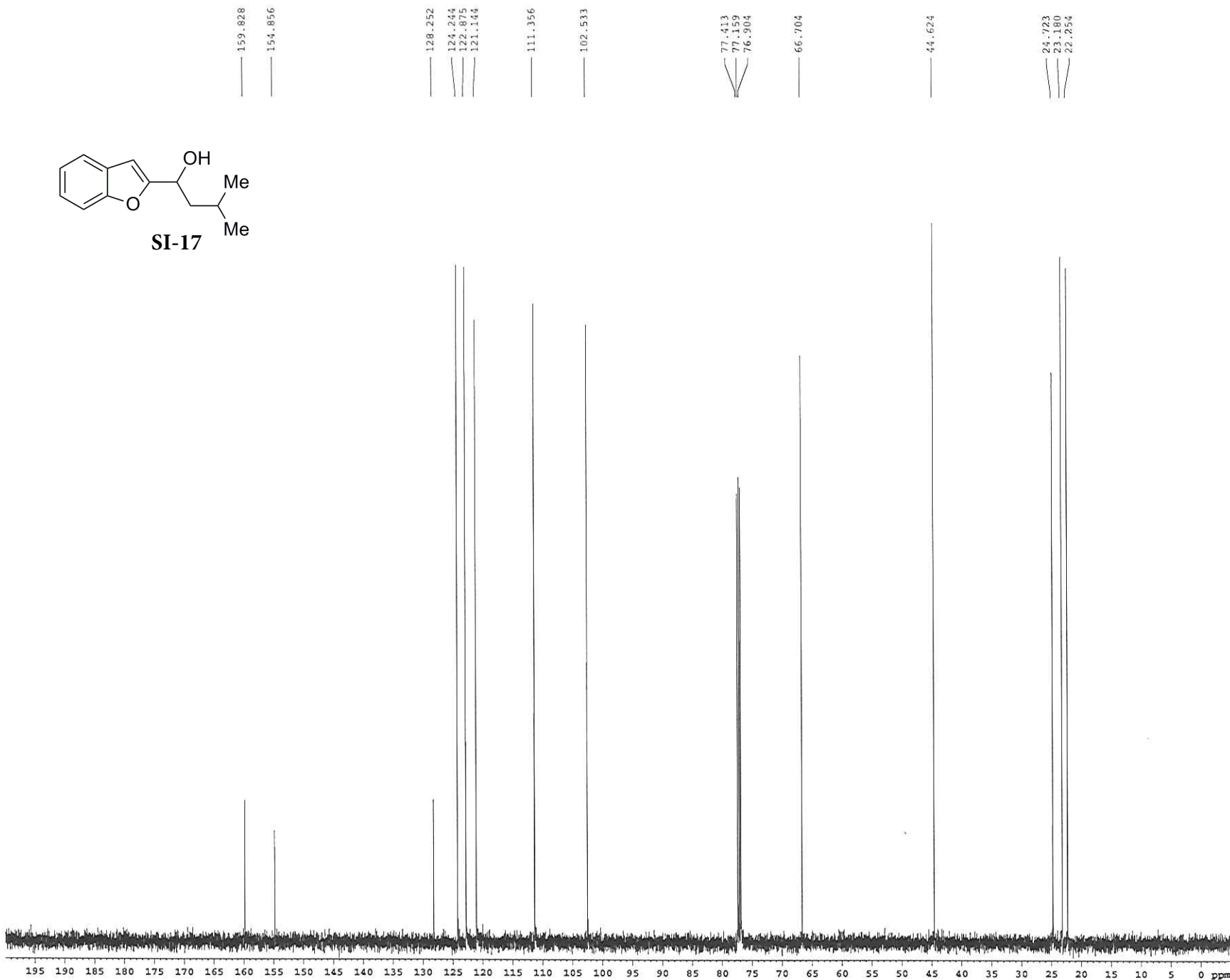
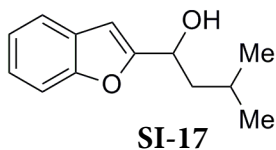
Current Data Parameters
=====
USER          yonova
NAME          agj3 - 32
EXPNO        111
PROCNO       1

F2 - Acquisition Parameters
Date_        20110519
Time         19.37
INSTRUM      cryo500
PROBHD       5 mm CPTCI 1H-
PULPROG      zg30
TD           81728
SOLVENT      CDCl3
NS           8
DS           2
SWH          8012.020 Hz
FIDRES       0.098043 Hz
AQ           5.0998774 sec
RG           9
DW           62.400 usec
DE           6.00 usec
TE           298.0 K
D1           0.10000000 sec
MCHYST       0.00000000 sec
MCHWK       0.01500000 sec

===== CHANNEL f1 =====
NUC1         1H
P1           7.50 usec
PL1          1.60 dB
SFO1         500.2235015 MHz

F2 - Processing parameters
SI           65536
SF           500.2200358 MHz
WDW          EM
SSB          0
LB           0.30 Hz
GB           0
PC           4.00
    
```

Z-restored spin-echo 13C spectrum with 1H decoupling



Current Data Parameters
 USER yonova
 NAME ajj - 32
 EXNO 222
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20130119
 Time 18.39
 INSTRUM cryo500
 PROBHD 5 mm CPTCI 1H-
 PULPROG SpinEchoq10sp.prd
 TD 65536
 SOLVENT CDCl3
 NS 89
 DS 16
 SWH 30303.031 Hz
 FIDRES 0.462388 Hz
 AQ 1.0814105 sec
 RG 8169.6
 DW 16.500 usec
 DE 6.00 usec
 TE 298.0 K
 D1 0.25000000 sec
 d11 0.03000000 sec
 D16 0.00020000 sec
 d17 0.00019600 sec
 MTHRCT 0.00000000 sec
 MCVRK 0.01500000 sec
 P2 31.00 usec

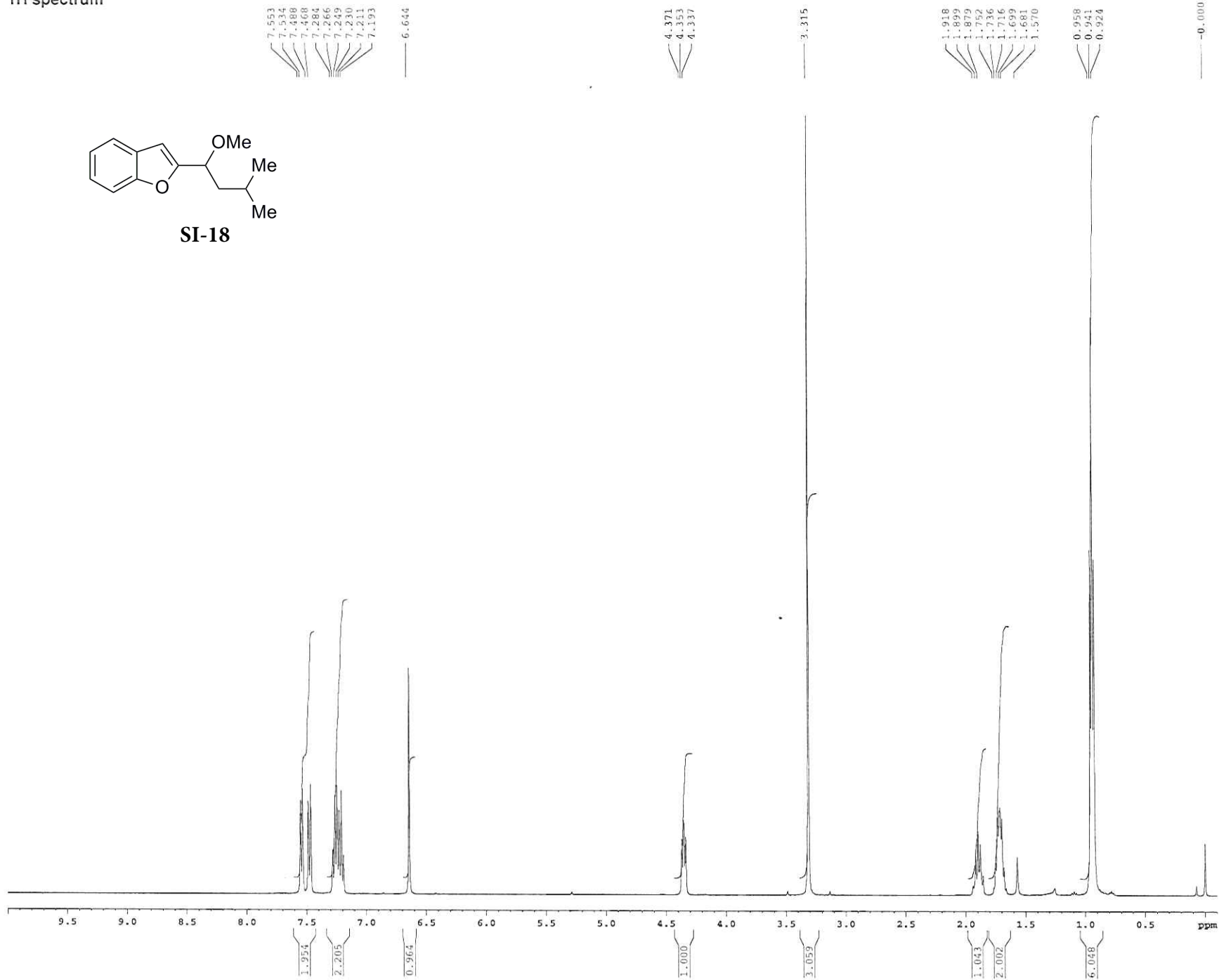
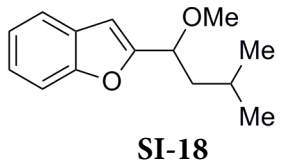
===== CHANNEL f1 =====
 NUC1 13C
 P1 19.50 usec
 PL1 500.00 usec
 P12 2000.00 usec
 PL0 120.00 dB
 PL1 -1.00 dB
 SF01 125.7942548 MHz
 SF1 3.20 dB
 SF2 3.20 dB
 SFO1 Crp60,0.5,20.1
 SFO2 Crp60comp,4
 SFOFF1 0.00 Hz
 SFOFF2 0.00 Hz

===== CHANNEL f2 =====
 CPOPRG2 waltz16
 NUC2 1H
 PCPD2 100.00 usec
 PL2 1.60 dB
 PL22 25.00 dB
 SF02 500.2225011 MHz

===== GRADIENT CHANNEL =====
 G1NAME1 SINE.100
 G1NAME2 SINE.100
 G1X1 0.00 %
 G1X2 0.00 %
 G1Y1 0.00 %
 G1Y2 0.00 %
 G1Z1 30.00 %
 G1Z2 50.00 %
 p15 500.00 usec
 p16 1000.00 usec

F2 - Processing parameters
 SI 65536
 SF 125.7804108 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 2.00

1H spectrum



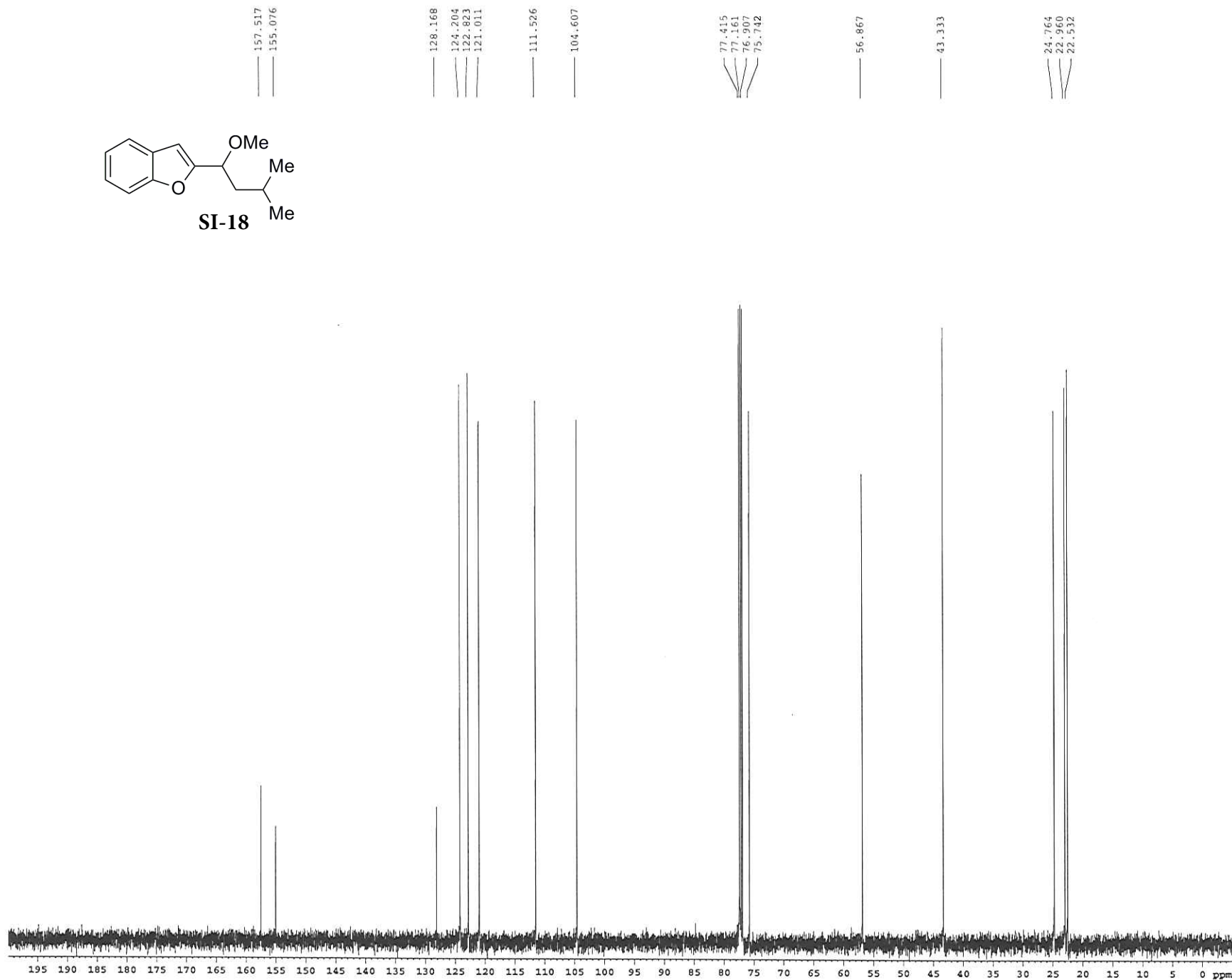
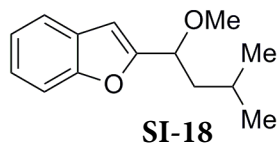
Current Data Parameters
 USEP yonova
 NAME iny5 - 250
 EXNO 1
 PRONO 1

F2 - Acquisition Parameters
 DATE 2010520
 TIME 17:24
 INSTRUM drx400
 PROBHD 5 mm QNP H/P/P
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 8
 DS 2
 SWH 6410.256 Hz
 FIDRES 0.097013 Hz
 AQ 5.111679 sec
 RG 143.7
 DW 78.000 usec
 DE 4.50 usec
 TE 298.2 K
 DI 0.1000000 sec
 MCREST 0.0000000 sec
 MCWRR 0.01500000 sec

----- CHANNEL f1 -----
 NUC1 1H
 P1 12.00 usec
 PL1 -0.50 dB
 SFO1 400.1328009 MHz

F2 - Processing parameters
 SI 65536
 SF 400.130049 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 2.00

Z-restored spin-echo 13C spectrum with 1H decoupling



```

Current Data Parameters
=====
USDR   yonova
NAME    imy5 - 239
EXPNO   22
PROCNO  1

F2 - Acquisition Parameters
Date_   20190519
Time    18.32
INSTRUM cryo500
PROBHD  5 mm CPXI 1H-
PULPROG SpinEchoPsi00p.prd
TD       65536
SOLVENT  CDCl3
NS       134
DS       16
SWH      30303.011 Hz
FIDRES   0.462388 Hz
AQ       1.0814105 sec
RG       7298.2
DW       16.500 usec
DE       6.00 usec
TE       298.0 K
D1       0.25000000 sec
d11      0.03000000 sec
d16      0.00020000 sec
d17      0.00018600 sec
MCHEST  0.00000000 sec
MCWHK   0.01500000 sec
P2       31.00 usec

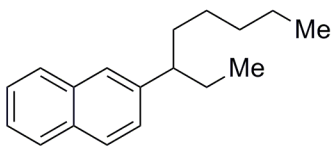
===== CHANNEL f1 =====
NUC1     13C
P1       15.50 usec
PL1      500.00 usec
PI2      2000.00 usec
PL0      120.00 dB
PL1      -1.00 dB
SFO1     125.7942548 MHz
SP1      3.20 dB
SP2      3.20 dB
SFO2
SPNAM1   Crp60,0.5,20.1
SPNAM2   Crp60comp,4
SPOWF1   0.00 Hz
SPOWF2   0.00 Hz

===== CHANNEL f2 =====
CPDPRG2  waltz16
NUC2     1H
PCPD2    100.00 usec
PL2      1.60 dB
PI2      24.60 dB
SFO2     500.2225011 MHz

===== GRADIENT CHANNEL =====
GPNAM1   SINE.100
GPNAM2   SINE.100
GPX1     0.00 %
GPX2     0.00 %
GPT1     0.00 %
GPT2     0.00 %
GPI1     30.00 %
GPI2     50.00 %
p15      500.00 usec
p16      1000.00 usec

F2 - Processing parameters
SI       65536
SF       125.7940094 MHz
WDW      EM
SSB      0
LB       1.00 Hz
GB       0
PC       2.00
    
```

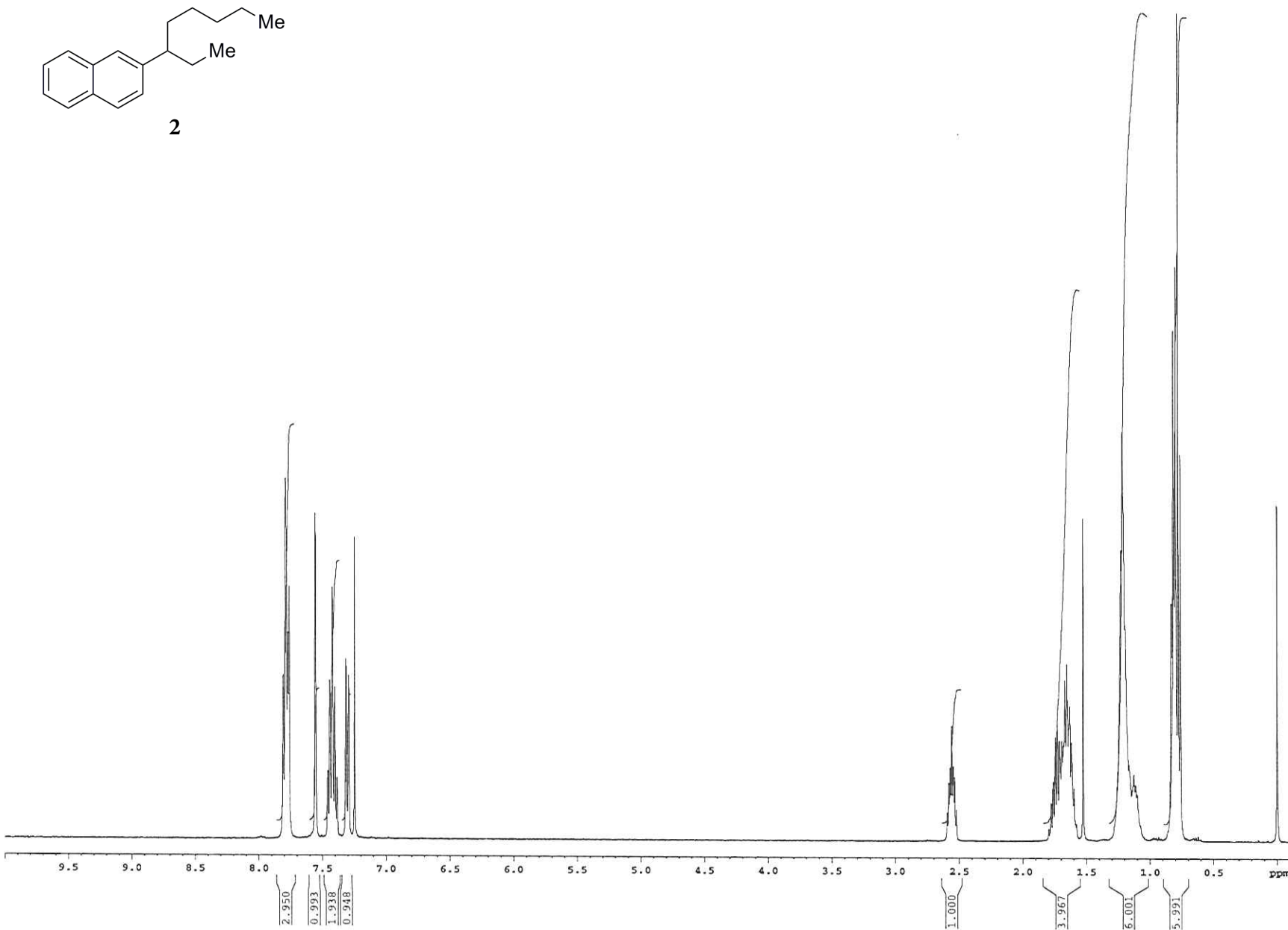
1H spectrum



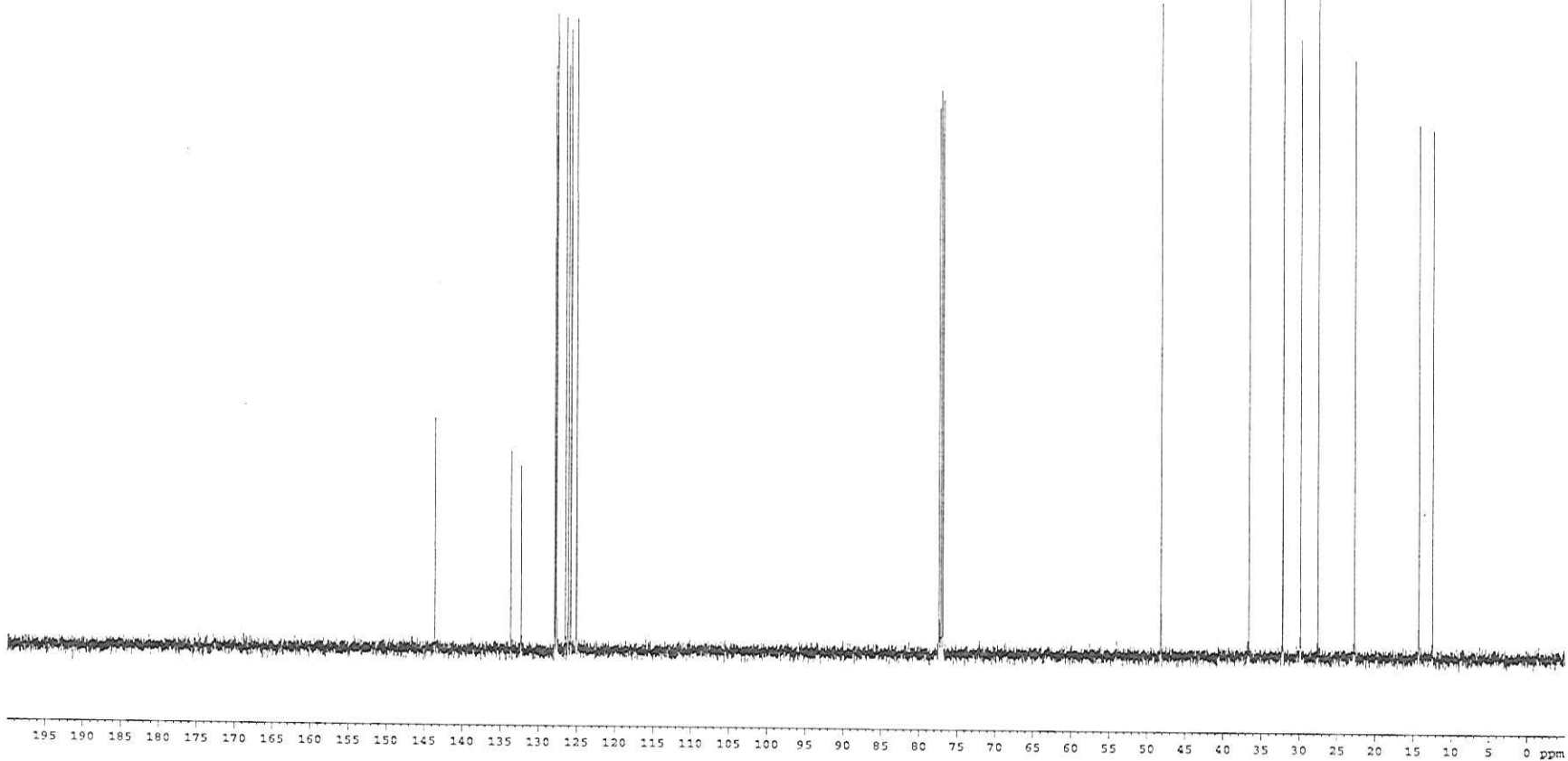
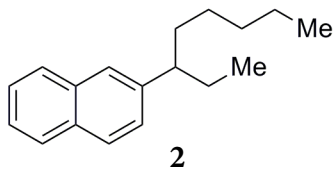
7.811
7.792
7.773
7.764
7.557
7.459
7.444
7.425
7.404
7.387
7.318
7.297
7.250

2.595
2.582
2.572
2.559
2.546
2.536
2.523
1.779
1.764
1.745
1.727
1.713
1.694
1.670
1.653
1.630
1.619
1.596
1.527
1.228
1.216
1.127
1.114
0.832
0.816
0.797
0.779
0.760

Current Data Parameters
 USER yonova
 NAME IMYS - 054
 EXPNO 111
 PROCNO 1
 F2 - Acquisition Parameters
 Date_ 20121204
 Time 0.12
 INSTRUM dx400
 PROBHD 5 mm QNP H/F/P
 PULPROG zg30
 TD 65536
 SOLVENT CDCl3
 NS 8
 DS 2
 SWH 6410.236 Hz
 FIDRES 0.097813 Hz
 AQ 5.1118579 sec
 RG 228.1
 DW 78.000 usec
 DE 4.50 usec
 TE 298.1 K
 D1 0.10000000 sec
 MCHST 0.00000000 sec
 MCMK 0.01500000 sec
 ===== CHANNEL f1 =====
 NUC1 1H
 P1 12.00 usec
 PL1 -0.60 dB
 SFO1 400.1328009 MHz
 F2 - Processing parameters
 SI 65536
 SF 400.1300255 MHz
 WDW EM
 SCA 0
 LB 0.30 Hz
 GB 0
 PC 2.00



Z-restored spin-echo 13C spectrum with 1H decoupling



```

Current Data Parameters
Date_      2011125
MSR       yonova
NAME      IMVS - 035
EXPNO     222
PROCNO    1

F2 - Acquisition Parameters
Date_      2011125
Time       5.35
INSTRUM    cryo500
PROBHD     5 mm CPIC 1H-
PULPROG    SpinEcho3Dm.prd
TD          65536
SOLVENT    CDCl3
NS         193
DS         16
SWH         30103.011 Hz
F2DRIS     0.462188 Hz
AQ          1.082409 sec
RG          729.13
DS          16
DE          6.00 usec
TE          298.0 K
D1          0.25000000 sec
d11         0.01000000 sec
d16         0.00020000 sec
d17         0.00019600 sec
d18         0.00060000 sec
MCHRG1     0.01500000 sec
MCHRG2     0.01500000 sec
P2          31.00 usec

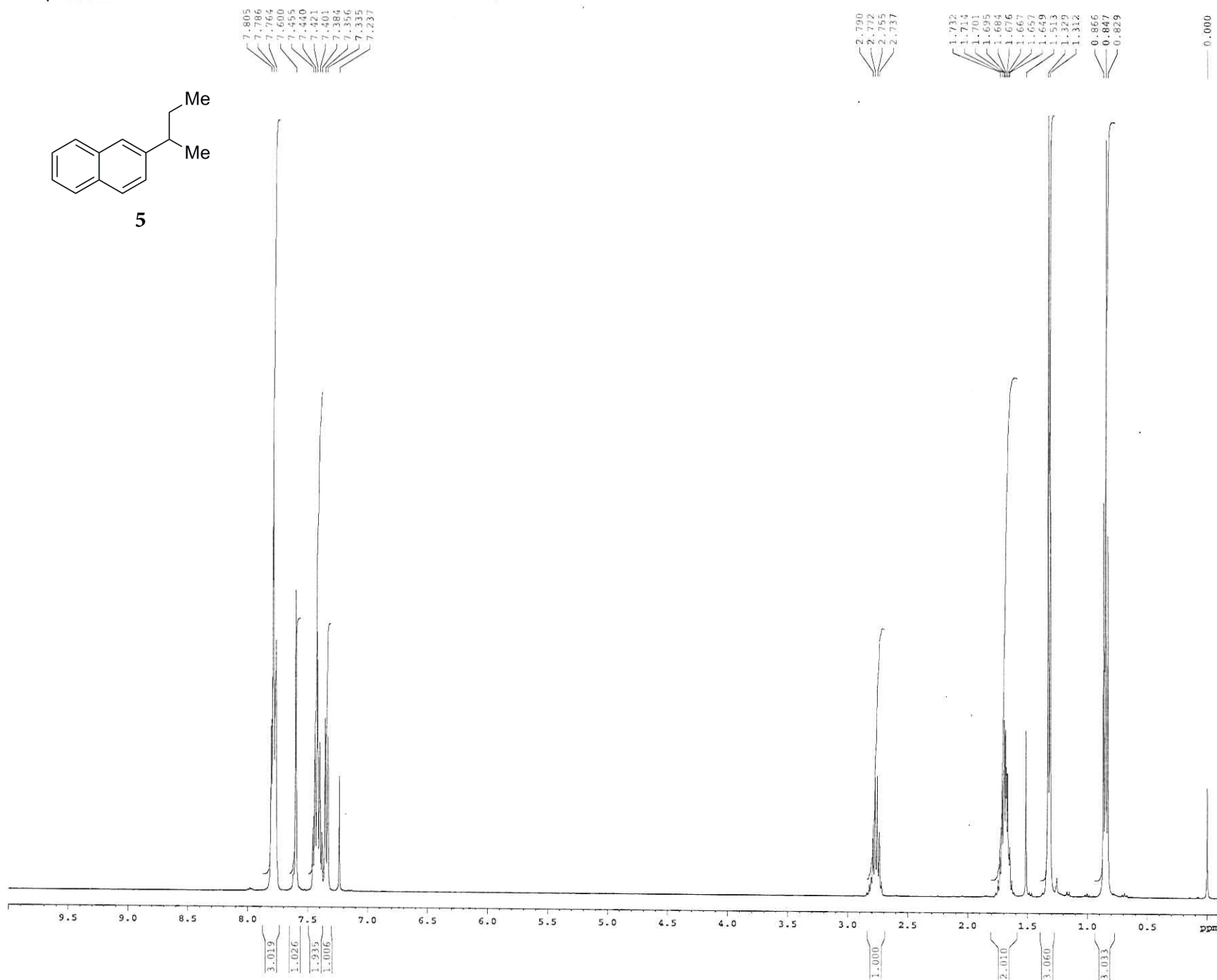
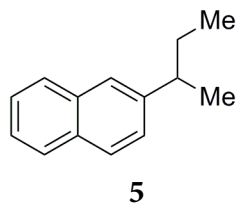
===== CHANNEL f1 =====
NUC1        13C
P1          15.50 usec
PL1         500.00 usec
PL2         2000.00 usec
PL3         120.00 dB
PL4         -1.00 dB
SFO1        125.7942548 MHz
SF1         3.20 dB
SF2         3.20 dB
SFO1        Csp60,0.5,20.1
SFO2        Csp60comp,4
SFO1        0.00 Hz
SFO2        0.00 Hz

===== CHANNEL f2 =====
CPDPRG2     wa1t15
NUC2        1H
PCPD2       100.00 usec
PL2         1.60 dB
PL12        24.60 dB
SFO2        500.222011 MHz

===== GRADIENT CHANNEL =====
GPM1        SINE,100
GPM2        SINE,100
GPR1        0.00 %
GPR2        0.00 %
GPR3        0.30 %
GPR4        0.00 %
GPR5        0.00 %
GPR6        10.00 %
GPR7        00.00 %
p15         500.00 usec
p16         1000.00 usec

F2 - Processing parameters
SI          65536
SF          125.7904090 MHz
WDW         EM
SSB         0
LB          1.00 Hz
GB          0
PC          2.00
    
```

1H spectrum



```

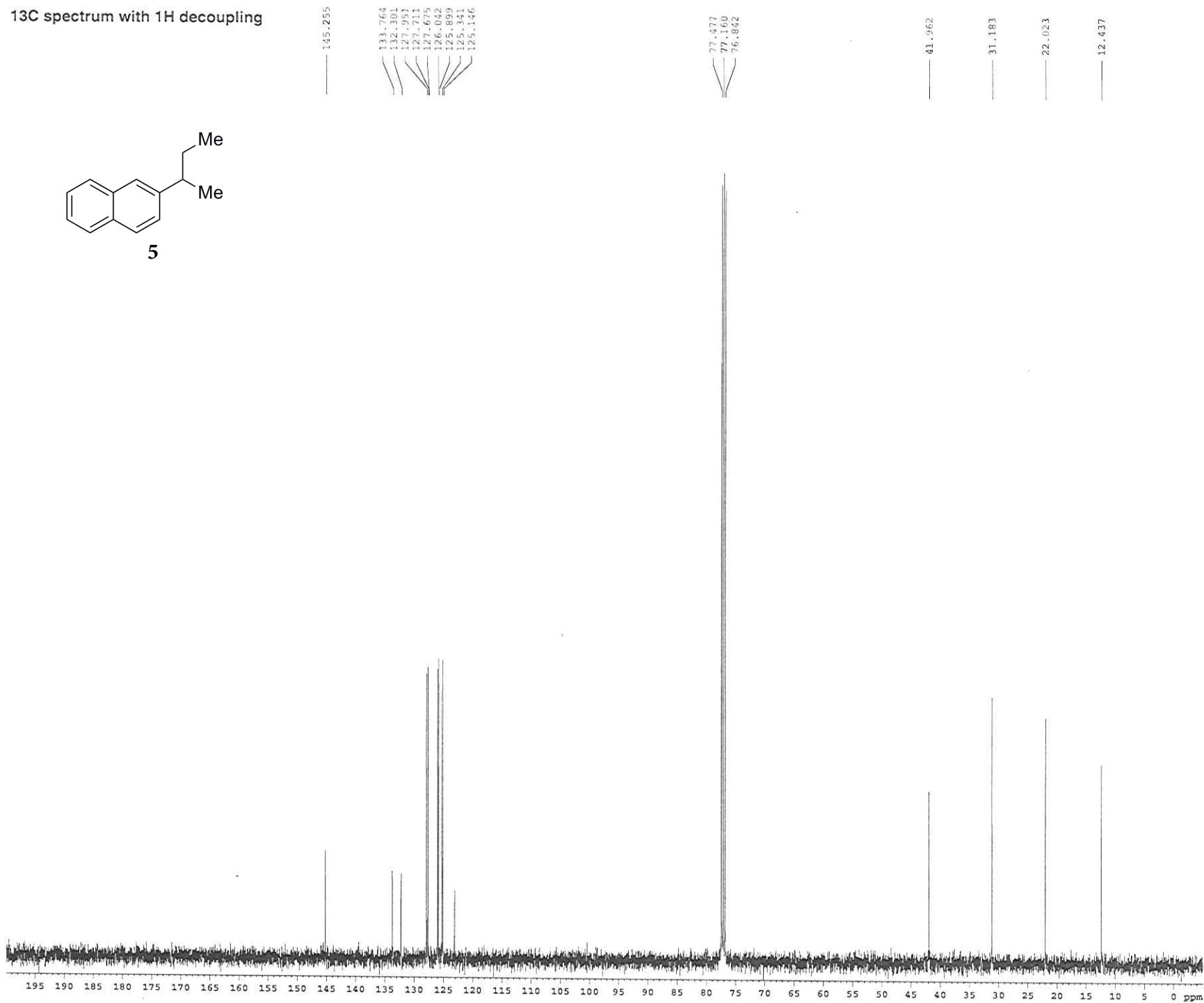
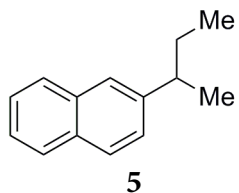
Current Data Parameters
USER          yonova
NAME          (mp) - 712
EXPNO        111
PROCNO        1

P2 - Acquisition Parameters
Date_         20130622
Time          15.14
INSTRUM      spect
PROBHD       5 mm QNP 1H/13
PULPROG      zgpg30
TD           65536
SOLVENT      CDCl3
NS           8
DS           2
SWH          6410.256 Hz
FIDRES       0.097823 Hz
AQ           5.1118579 sec
RG           161.3
DN           78.000 usec
DE           4.50 usec
TE           298.2 K
D1           0.13000000 sec
MCRETOT      0.03000000 sec
MCRMK        0.01500000 sec

===== CHANNEL f1 =====
NUC1          13C
P1            12.00 usec
PL1           -0.60 dB
SFO1          400.1328003 MHz

P2 - Processing parameters
SI           65536
SF           400.1300300 MHz
WDW          EM
SSB          0
GB           0.30 Hz
CB           0
PC           2.00
    
```

¹³C spectrum with ¹H decoupling



```

Current Data Parameters
USER          yonosa
NAME          lmy5 - 232
EXPNO        222
PROCNO       1

F2 - Acquisition Parameters
Date_        20130622
Time         15.18
INSTRUM     dec400
PROBHD      5 mm QNP 3/FP
PULPROG     zgpg30
TD          65536
SOLVENT     CDCl3
NS          307
DS          4
SWH         24154.590 Hz
FIDRES     0.348570 Hz
AQ         1.3566452 sec
RG         5195.2
DW         20.700 usec
DE         20.39 usec
TE         298.1 K
D1         0.10000000 sec
d11        0.03000000 sec
MCHTEST    0.00000000 sec
MCMRK      0.01500000 sec

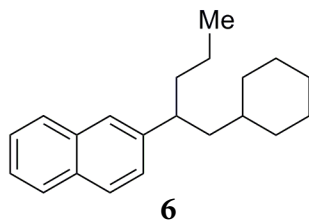
===== CHANNEL f1 =====
NUC1        13C
P1          11.00 usec
PL1         0.00 dB
SFO1        100.6237964 MHz

===== CHANNEL f2 =====
CPDPRG2     mlev16
NUC2        1H
PCPD2       80.00 usec
PL2         0.00 dB
PL12        16.20 dB
SFO2        400.1324009 MHz

F2 - Processing parameters
SI          65536
SF          100.6127595 MHz
WDW         EM
SSB         0
LB          1.00 Hz
GB          0
PC          1.00
    
```

¹H spectrum

7.804
7.786
7.775
7.756
7.754
7.751
7.439
7.438
7.424
7.421
7.416
7.413
7.399
7.386
7.320
7.317
7.303
7.300
7.221



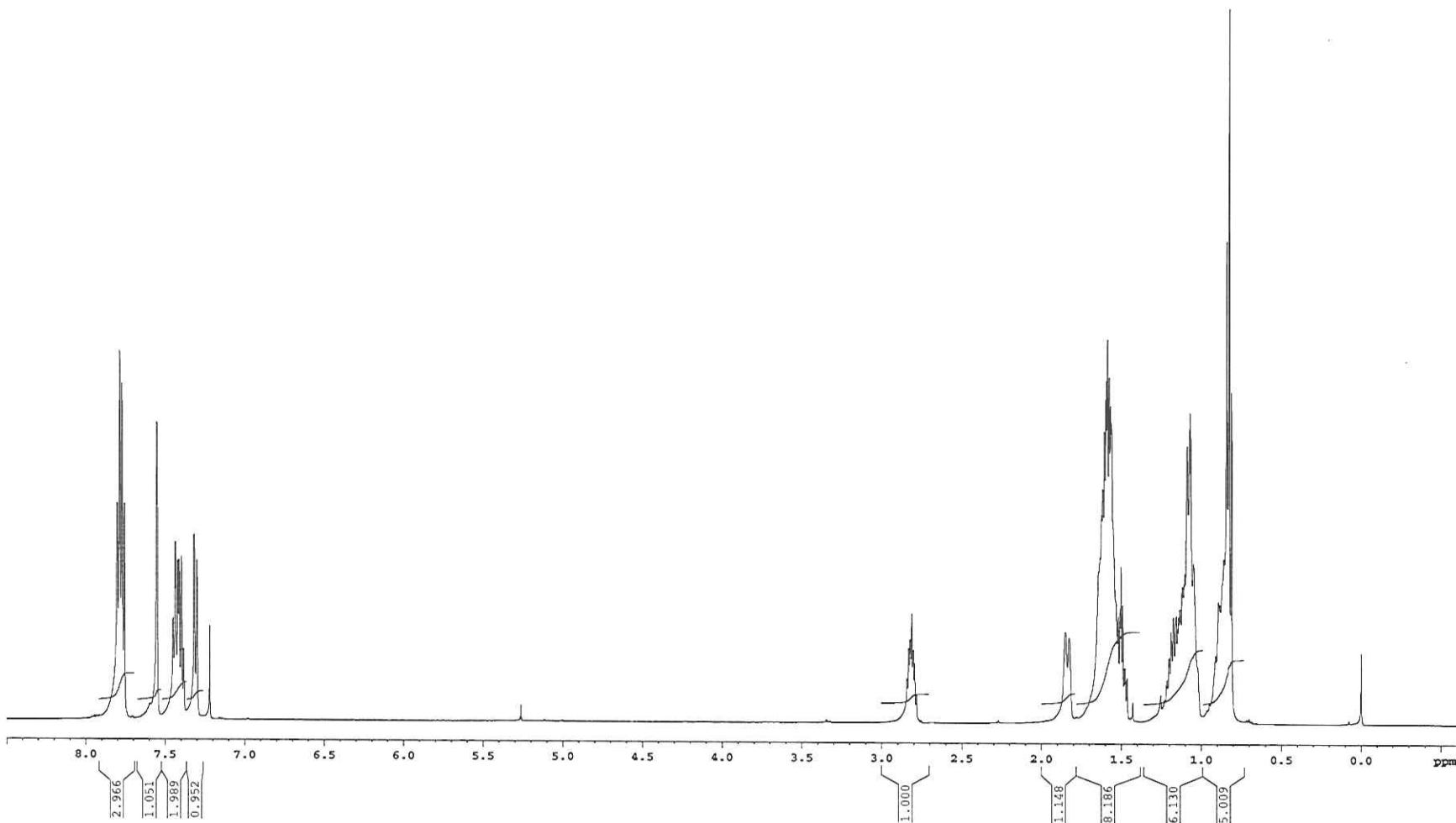
2.844
2.833
2.825
2.814
2.804
2.796
2.785
1.852
1.827
1.645
1.632
1.626
1.620
1.617
1.608
1.597
1.590
1.579
1.570
1.564
1.542
1.510
1.504
1.494
1.483
1.477
1.466
1.295
1.217
1.203
1.190
1.176
1.173
1.158
1.150
1.144
1.138
1.136
1.121
1.114
1.113
1.109
1.105
1.091
1.073
1.052
0.942
0.894
0.884
0.874
0.861

```
Current Data Parameters
=====
USER          aaronj
NAME          AGJ_1_100
EXPNO        3
PROCNO       1

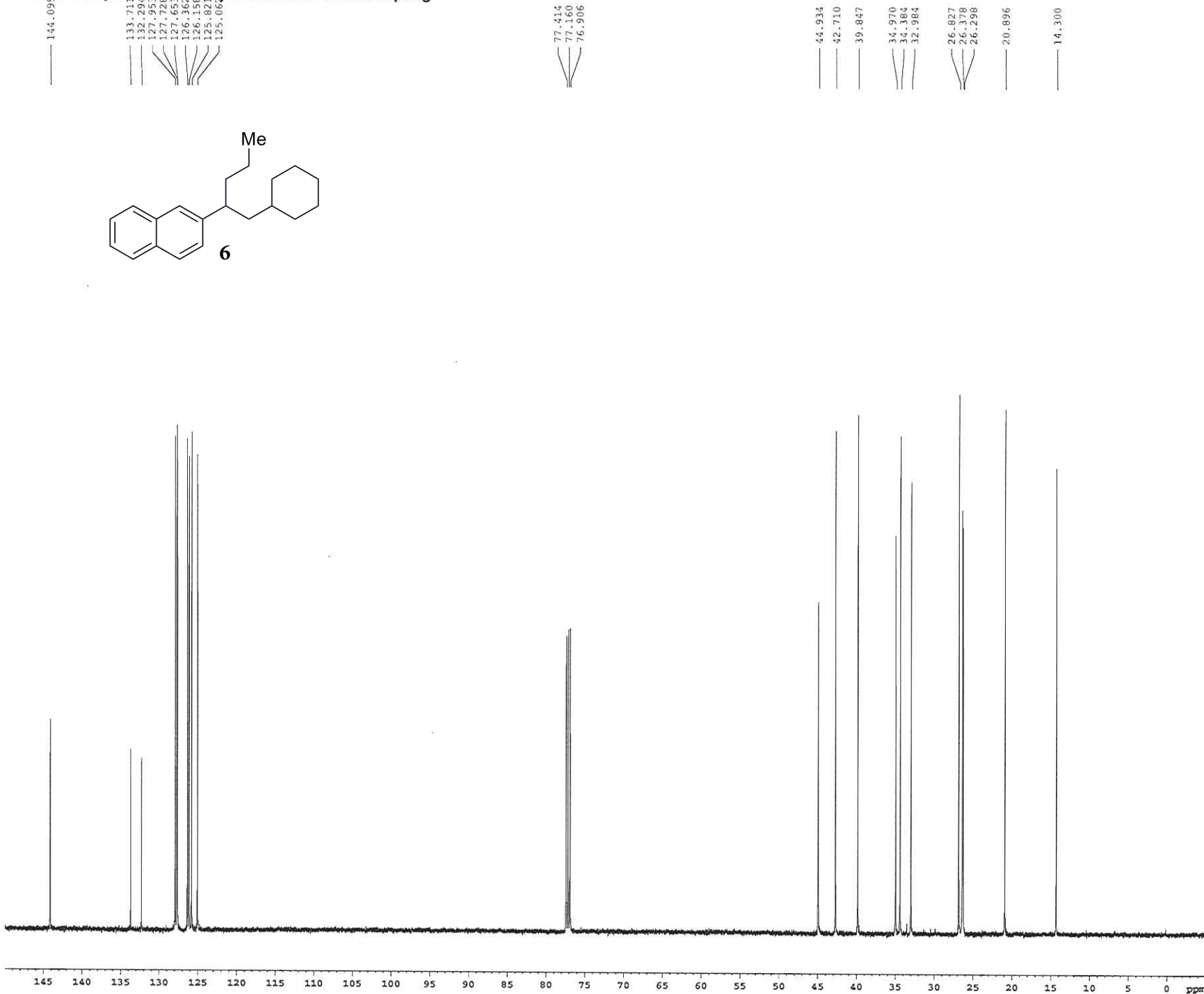
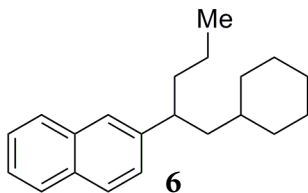
F2 - Acquisition Parameters
=====
Date_        20120502
Time         23.25
INSTRUM      cryo500
PROBHD       5 mm CPTCI 1H-
PULPROG      zg30
TD            81728
SOLVENT      CDCl3
NS            8
DS            2
SWH           8012.820 Hz
FIDRES       0.098041 Hz
AQ            5.0998774 sec
RG            5
DW            62.400 usec
DE            6.00 usec
TE            298.0 K
D1            0.10000000 sec
MCRET        0.00000000 sec
MCWRK        0.01500000 sec

===== CHANNEL f1 =====
NUC1          1H
P1            7.50 usec
PL1           1.50 dB
SFO1          500.2235015 MHz

F2 - Processing parameters
=====
SI            65536
SF            500.2200512 MHz
WDW           EM
SSB           0
LB            0.30 Hz
GB            0
PC            4.00
```



Z-restored spin-echo ¹³C spectrum with ¹H decoupling



```

Current Data Parameters
USER      aaronj
NAME      AGJ_160
EXPNO     4
PROCNO    1

F2 - Acquisition Parameters
Date_     20120502
Time      23.32
INSTRUM   cryo00
PROBHD    5 mm CPCL 1H-
PULPROG   SpinEcho30op.prd
TD         65536
SOLVHT    CDCl3
NS         412
DS         16
SWH        30303.031 Hz
FIDRES     0.442388 Hz
AQ         1.0813940 sec
RG         5792.6
DW         2.6500 usec
DE         6.00 usec
TE         298.0 K
D1         0.25000000 sec
d11        0.03000000 sec
d16        0.00000000 sec
d17        0.00019600 sec
MCREST     0.00000000 sec
MCWRK     0.01500000 sec
F2         31.00 usec

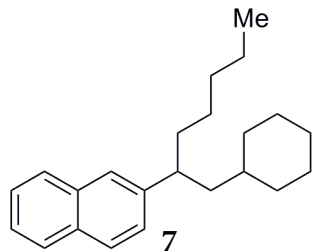
===== CHANNEL f1 =====
NUC1       13C
P1         15.50 usec
P11        500.00 usec
P12        2000.00 usec
PL0        120.00 dB
PL1        -1.00 dB
SFO1      125.7942548 MHz
SP1        3.20 dB
SP2        3.20 dB
SFO2      Crp60.05.20.1
SPNAM1    Crp60.05.20.1
SPNAM2    Crp60comp.4
SPOPF1    0.00 Hz
SPOPF2    0.00 Hz

===== CHANNEL f2 =====
CPDPRG2   waltz16
NUC2       1H
PCPD2     100.00 usec
PL2       1.60 dB
PL12      24.60 dB
SFO2      500.225011 MHz

===== GRADIENT CHANNEL =====
GPNAM1    SINE.100
GPNAM2    SINE.100
GPX1      0.00 %
GPX2      0.00 %
GPY1      0.00 %
GPY2      0.00 %
GPE1      30.00 %
GPE2      50.00 %
p15       500.00 usec
p16       1000.00 usec

F2 - Processing parameters
SI         65536
SF         125.7804122 MHz
WDW        EM
SSB        0
LB         1.00 Hz
GB         0
PC         2.00
    
```

¹H spectrum



7.796
7.779
7.768
7.748
7.549
7.443
7.429
7.407
7.384
7.370
7.315
7.312
7.294
7.291
7.175

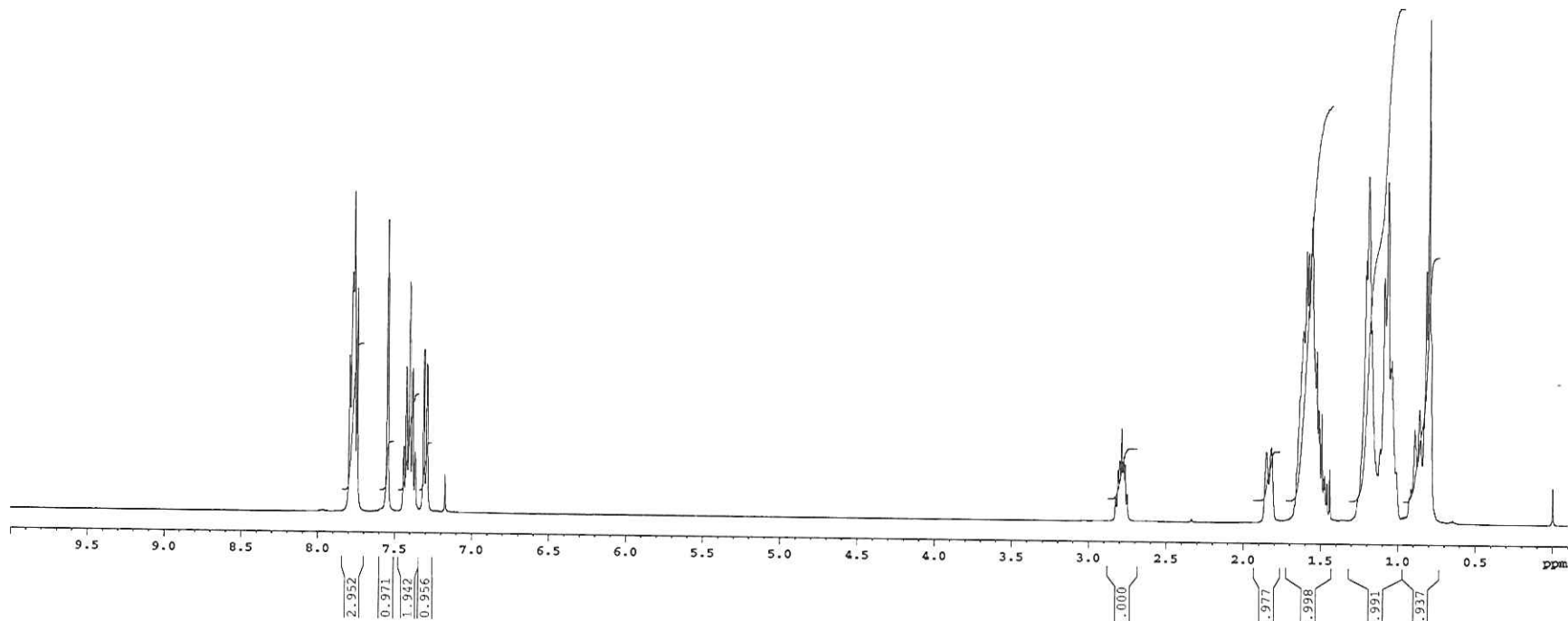
2.826
2.812
2.802
2.789
2.776
2.766
2.752
1.853
1.821
1.617
1.597
1.584
1.560
1.540
1.516
1.509
1.495
1.482
1.474
1.461
1.453
1.411
1.397
1.093
1.071
1.044
1.033
0.893
0.853
0.822
0.805

Current Data Parameters
USER ycnova
NAME IMY5 - 117
EXPNO 11
PROCNO 1

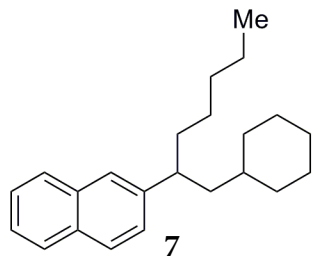
F2 - Acquisition Parameters
Date_ 20121209
Time 15.41
INSTRUM dx400
PROBHD 5 mm QNP H/P/P
PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 8
DS 2
SWH 6410.256 Hz
FIDRES 0.097813 Hz
AQ 5.1118579 sec
RG 32
DW 78.000 usec
DE 4.50 usec
TE 298.1 K
D1 0.1000000 sec
MCRET 0.0000000 sec
MCWREX 0.01500000 sec

===== CHANNEL f1 =====
NUCL 1H
P1 12.00 usec
PL1 -0.00 dB
SFO1 400.1328009 MHz

F2 - Processing parameters
S2 65536
SF 400.1300549 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 2.00



Z-restored spin-echo 13C spectrum with 1H decoupling

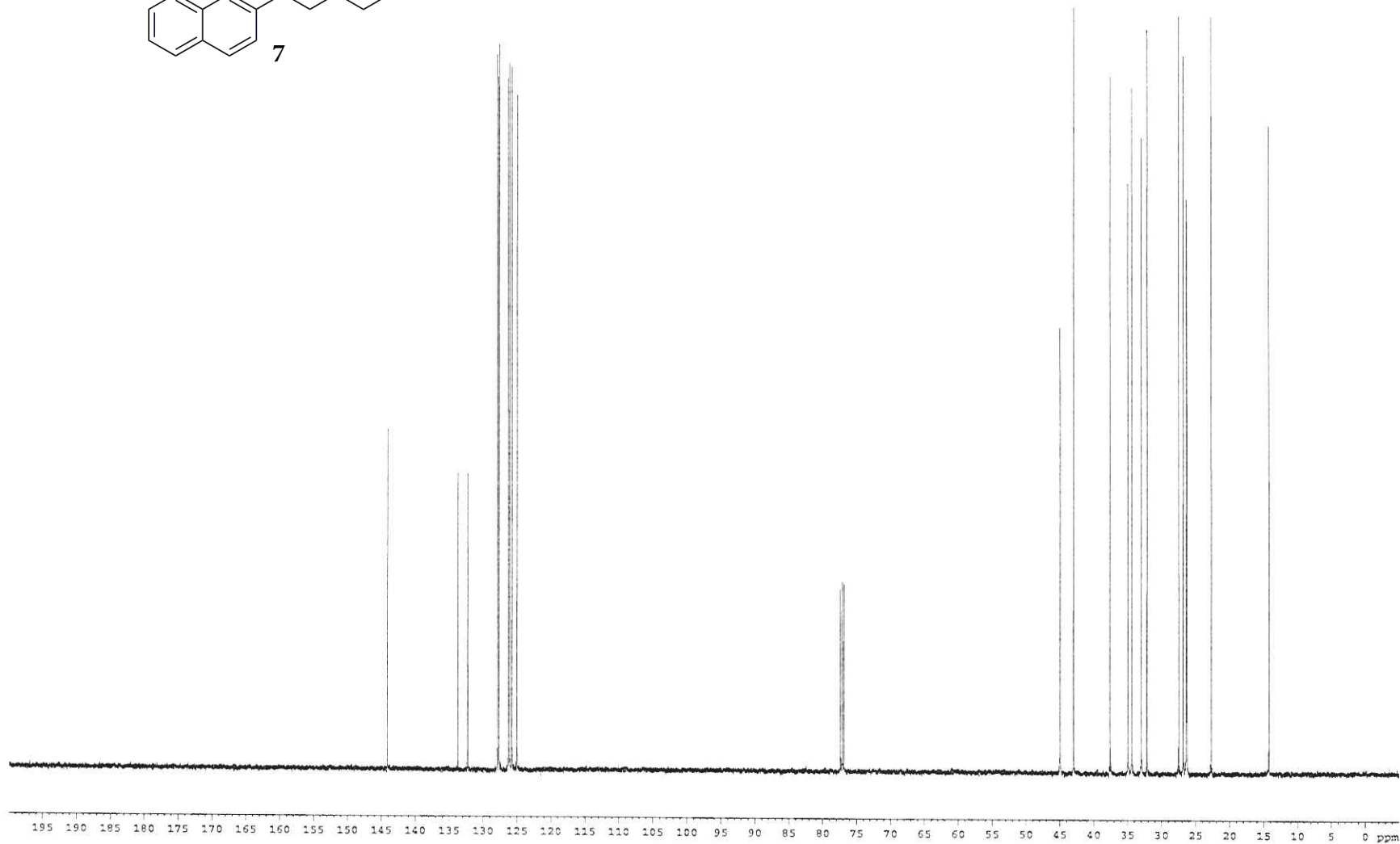


144.151
133.738
132.118
127.967
127.735
127.672
126.956
126.149
126.805
125.051

77.413
77.159
76.905

45.002
43.009
37.610
35.990
34.404
32.991
32.162
27.573
26.879
26.306
26.235

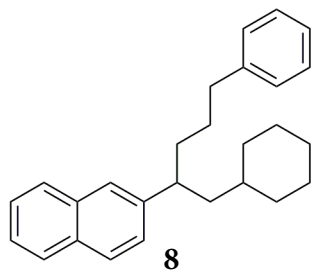
14.255



```

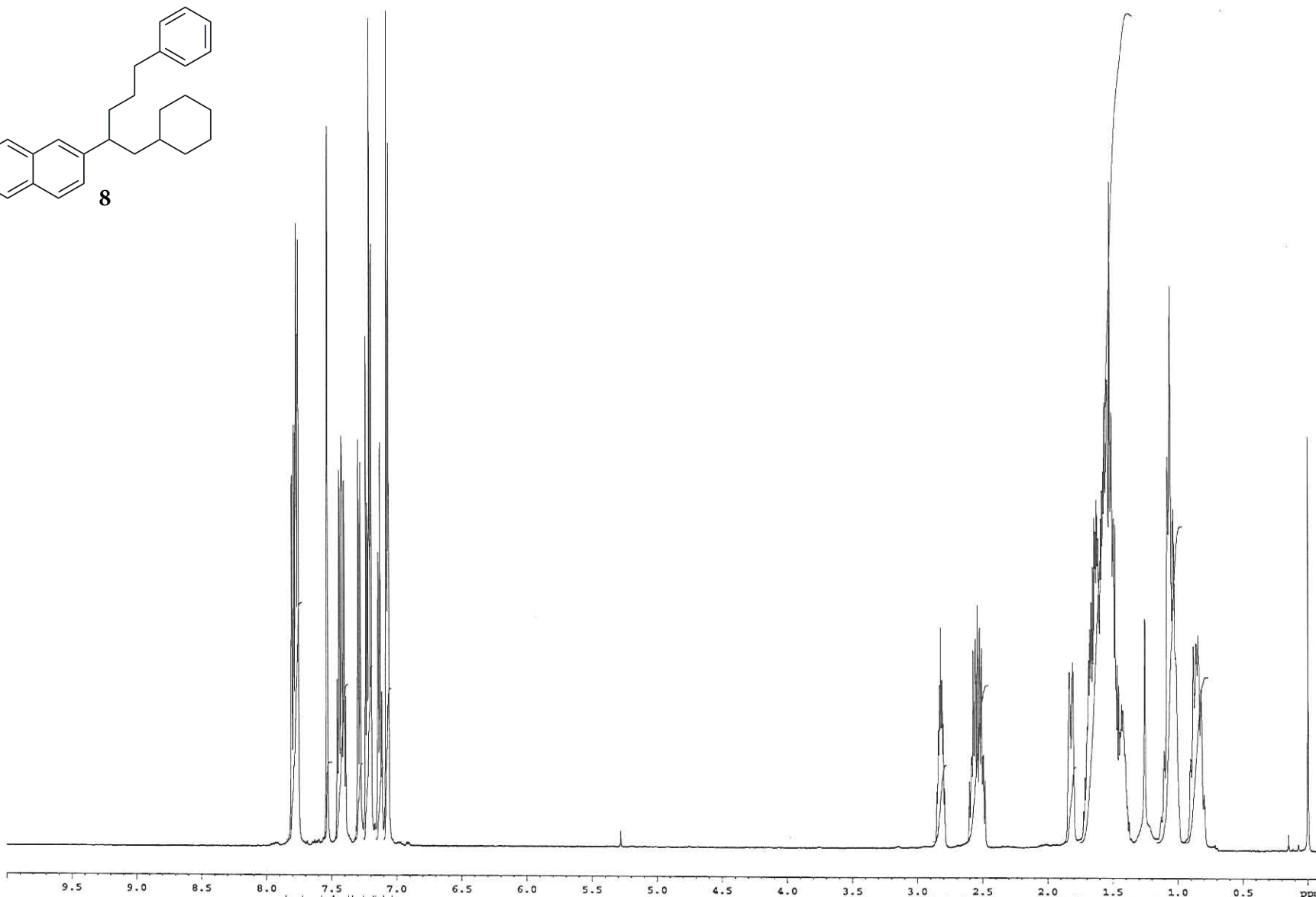
Current Data Parameters
JED0      Y00000
NAME      IMV5   117
EXPNO     222
PROCNO    1
----- Acquisition Parameters
Date_     20121209
Time      15:53
INSTRUM   cryo600
PROBHD    5 mm CPIC1 1H-
PULPROG   Spinschope30app.grd
TD         65536
SOLVENT   CDCl3
NS         32
DS         16
SWH        30302.931 Hz
FIDRES     0.442388 Hz
AQ         1.0814205 sec
RG         8.32
DE         6.00 usec
TE         298.0 K
D1         0.25000000 sec
d11        0.03000000 sec
D15        0.05000000 sec
d17        0.05019600 sec
XCFIRST   0.00000000 sec
XCFWH     0.01500000 sec
P2         31.00 usec
----- CHANNEL f1 -----
NUC1       13C
P1         15.50 usec
P11        500.00 usec
P12        2000.00 usec
PL1        120.00 dB
PL2        -1.00 dB
SFO1       125.7942649 MHz
SD1        3.20 dB
SD2        1.20 dB
SFO2       Crp60.0.5.20.1
SFO3       Crp60comp.4
SFOFF1     0.00 Hz
SFOFF2     0.00 Hz
----- CHANNEL f2 -----
CPDPRG2   waltz16
NUC2       1H
PCPD2     100.00 usec
PL2        1.00 dB
PL12       24.60 dB
SFO2       500.2225011 MHz
----- GRADIENT CHANNEL -----
G1NAME1    SINE.100
G1NAME2    SINE.100
GPM1       0.00 %
GPM2       0.00 %
GPM3       0.00 %
GPM4       0.00 %
GPM5       30.00 %
GPM6       50.00 %
p15        500.00 usec
p16        1000.00 usec
P2 - Processing parameters
SI         65536
SF         125.7804169 MHz
RGW        EX
SSB         0
LB         1.00 Hz
GB         0
PC         2.00
    
```

1H spectrum



7.808
7.792
7.775
7.758
7.536
7.457
7.444
7.425
7.408
7.393
7.297
7.281
7.240
7.229
7.214
7.199
7.145
7.130
7.115
7.080
7.066

2.832
2.822
2.813
2.601
2.573
2.556
2.541
2.522
2.509
2.481
1.835
1.810
1.687
1.677
1.666
1.654
1.643
1.635
1.624
1.594
1.585
1.575
1.565
1.544
1.510
1.494
1.483
1.465
1.454
1.433
1.254
1.080
1.061
1.035
0.881
0.845
0.822



3.058
1.004
2.011
0.992
2.243
0.917
1.954
1.000
2.028
0.980
10.661
4.079
2.144

```

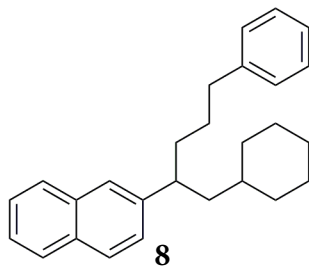
Current Data Parameters
=====
NAME      yonova
EXPNO     11
PROCNO    1

F2 - Acquisition Parameters
Date_     20130620
Time      20.57
INSTRUM   cryo500
PROBHD    5 mm CPTCI 1H-
PULPROG   zg30
TD         81728
SOLVENT   CDCl3
NS         8
DS         2
SWH        8012.820 Hz
FIDRES    0.098043 Hz
AQ         5.0999398 sec
RG         7.2
DE         62.400 usec
TE         298.0 K
D1         0.10000000 sec
MCHRECT   0.00000000 sec
MCWPK     0.01500000 sec

===== CHANNEL f1 =====
NUC1       1H
P1         7.50 usec
PL1        1.60 dB
SFO1       500.2235015 MHz

F2 - Processing parameters
SI         65536
SF         500.2200398 MHz
WDW        EM
SSB        0
LB         0.30 Hz
GB         0
PC         4.00
    
```

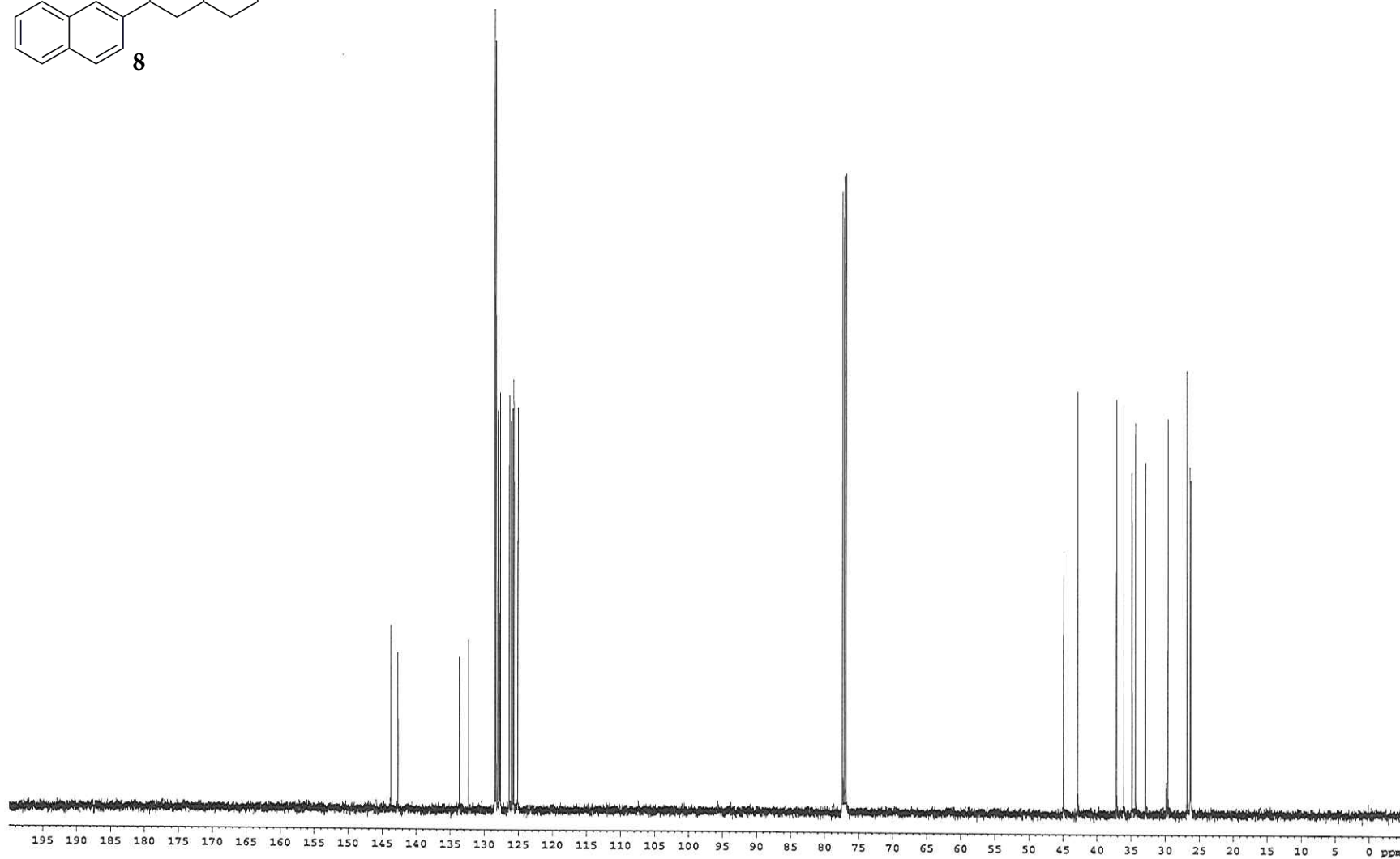

Z-restored spin-echo 13C spectrum with 1H decoupling



143.749
142.719
133.701
132.328
128.516
128.326
128.045
127.726
127.676
126.991
126.090
125.853
125.708
123.122

77.416
77.162
76.908

44.924
42.869
37.159
36.133
36.021
34.379
33.928
32.616
22.987
20.349
20.267



Current Data Parameters
USER ym0008
NAME imy5 - 109
EXPTNO 222
PROCNO 1

F2 - Acquisition Parameters
Date_ 20130620
Time 21.03
INSTRUM crys500
PROBHD 5 mm CPTCI 1H-
PULPROG SpinEcho30pp.prd
TD 65536
SOLVENT CDCl3
NS 520
DS 16
SWH 30303.031 Hz
FIDRES 0.462388 Hz
AQ 1.0814105 sec
RG 10321.3
DM 16.500 usec
DE 8.00 usec
TE 298.0 K
D1 0.25000000 sec
d11 0.03000000 sec
d16 0.00200000 sec
d17 0.00019600 sec
MCRET 0.00000000 sec
MCWRR 0.01500000 sec
P2 31.00 usec

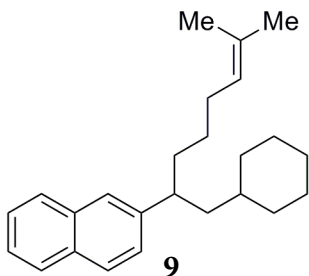
===== CHANNEL f1 =====
NUC1 13C
P1 15.50 usec
P11 500.00 usec
P12 2000.00 usec
PL0 120.00 dB
PL1 -1.00 dB
SFO1 125.7942548 MHz
SP1 3.20 dB
SP2 3.20 dB
SFXAM1 Crp60,0.5,20.1
SFXAM2 Crp60comp.4
SFOFF1 0.00 Hz
SFOFF2 0.00 Hz

===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 100.00 usec
PL0 1.60 dB
PL12 24.60 dB
SFO2 500.225011 MHz

===== GRADIENT CHANNEL =====
GPNAM1 SINE.100
GPNAM2 SINE.100
GPI1 0.00 %
GPI2 0.00 %
GPI3 0.00 %
GPI4 0.00 %
GPI5 30.00 %
GPI6 50.00 %
d15 500.00 usec
d16 1000.00 usec

F2 - Processing parameters
SI 65536
SF 125.7804090 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 2.00

Z-restored spin-echo ¹³C spectrum with ¹H decoupling



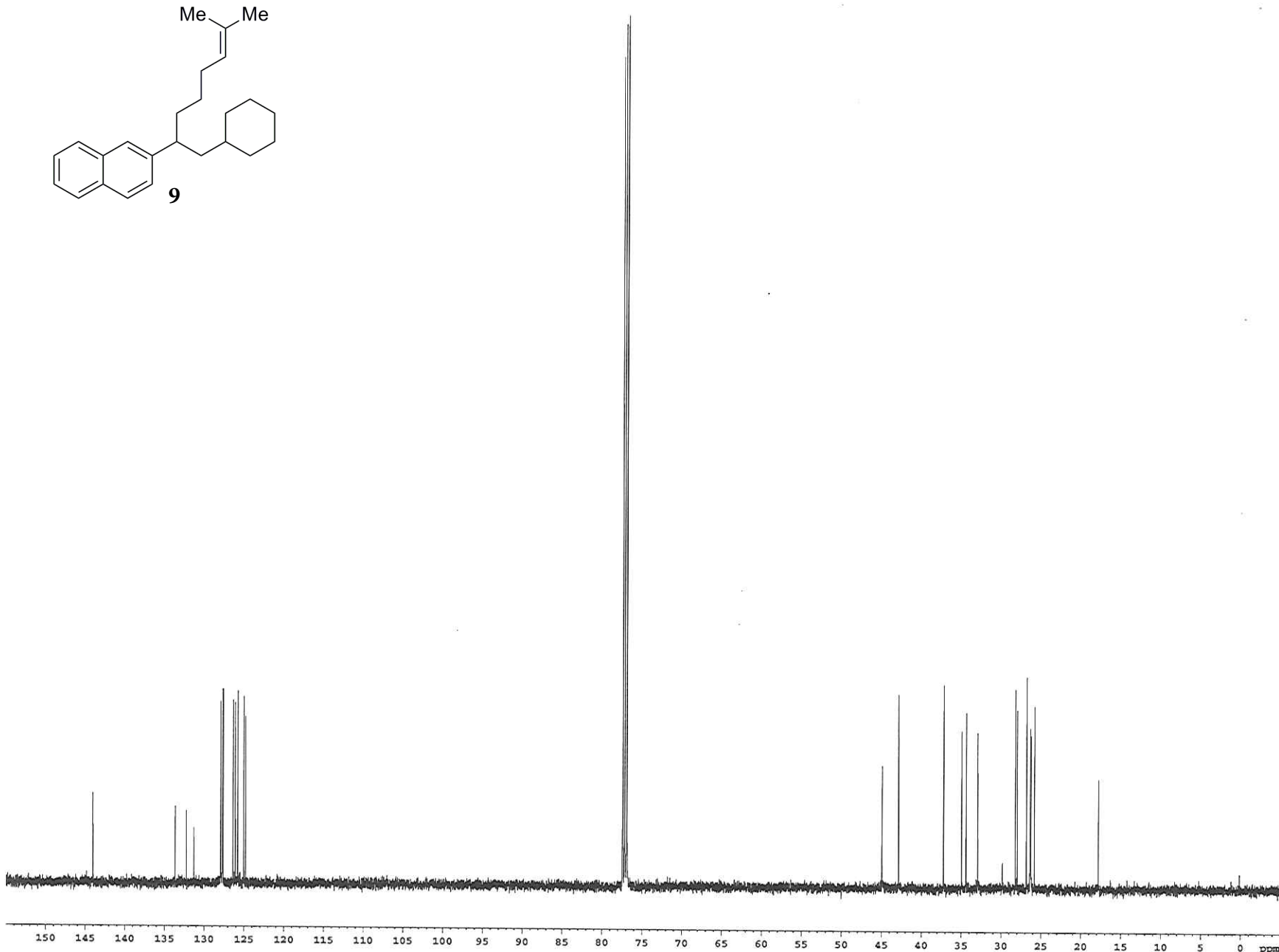
144.025
133.706
132.296
131.349
127.973
127.725
127.662
126.403
126.146
125.818
125.065
124.853

77.415
77.161
76.907

44.943
42.891

37.236
34.957
34.401
32.945
28.209
27.995
26.819
26.370
26.286
25.844

17.812



```
Current Data Parameters
USHR          aar01
NAME          AGI_2_223_c2_1
EXPNO         3
PROCNO        1

F2 - Acquisition Parameters
Date_         20110130
Time          18.31
INSTRUM       cryo100
PROBHD        5 mm CPTCI 1H-
PULPROG       SpinEchoq10pp.prd
TD            65536
SOLVENT       CDCl3
NS            713
DS            16
SWH           30303.031 Hz
FIDRES        0.442388 Hz
AQ            1.0813940 sec
RG            7298.2
DW            16.500 usec
DE            5.00 usec
TE            298.0 K
D1            0.25000000 sec
d11           0.03000000 sec
d16           0.00020000 sec
d17           0.00019600 sec
MCHRECT       0.00000000 sec
MCWRRK        0.01500000 sec
P2            31.00 usec

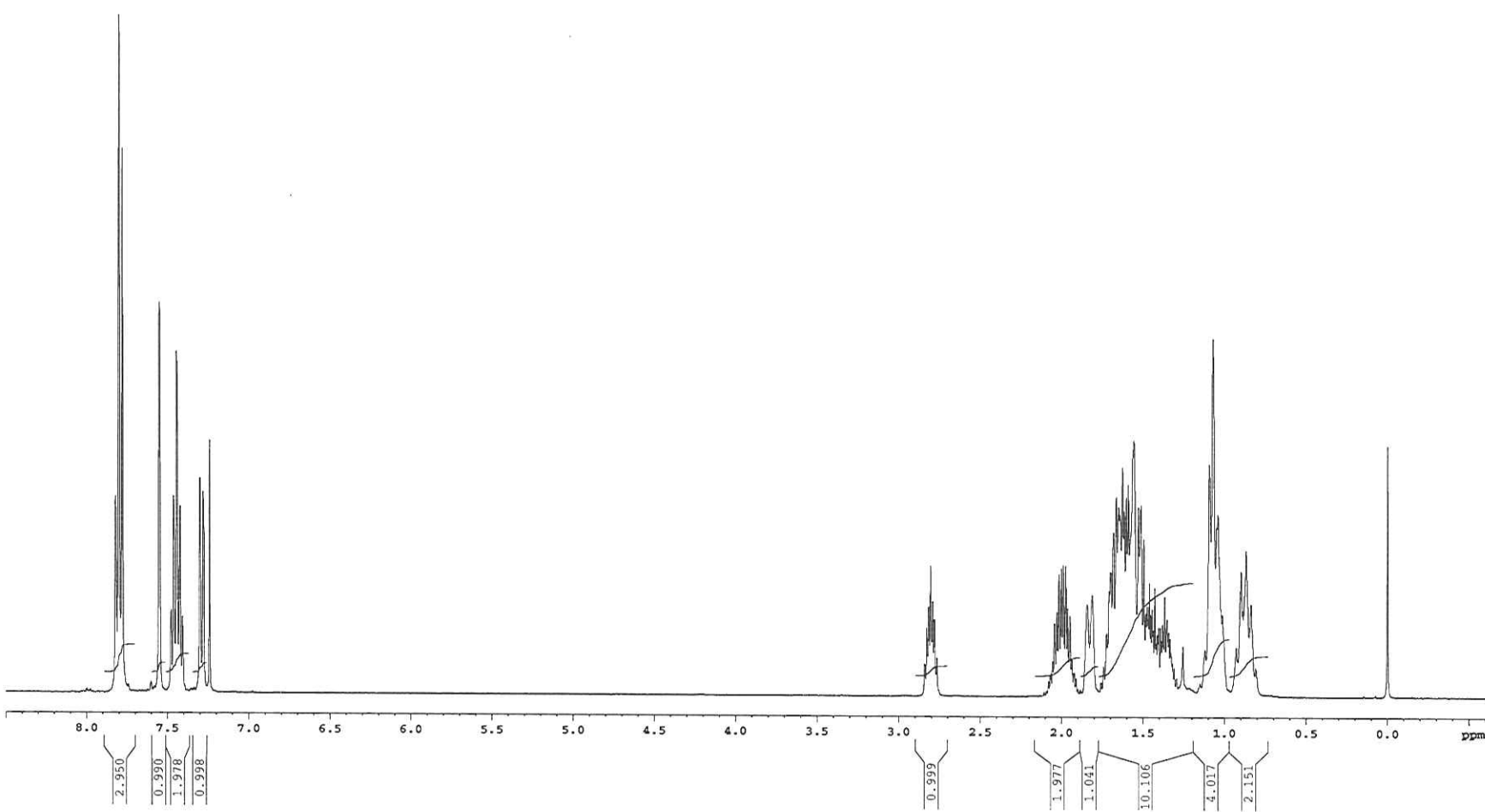
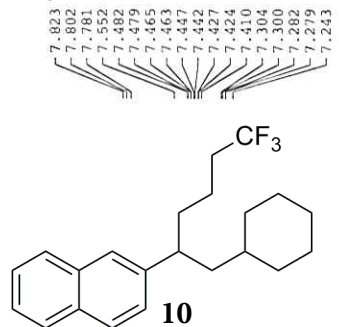
===== CHANNEL f1 =====
NUC1           13C
P1            15.50 usec
PL1           500.00 usec
P12           2000.00 usec
PL0           120.00 dB
PL1           -1.00 dB
SFO1          125.7842548 MHz
SP1           3.20 dB
SP2           3.20 dB
SPNAM1        Crp60.0.5.20.1
SPNAM2        Crp60comp.4
SPOFF1        0.00 Hz
SPOFF2        0.00 Hz

===== CHANNEL f2 =====
CPDPRG2       waltz16
NUC2           1H
PCPD2         100.00 usec
PL2           1.60 dB
PL12          24.60 dB
SFO2          500.2225011 MHz

===== GRADIENT CHANNEL =====
GPNAM1        SINE.100
GPNAM2        SINE.100
GPX1          0.00 %
GPX2          0.00 %
GPY1          0.00 %
GPY2          0.00 %
GPZ1          30.00 %
GPZ2          50.00 %
p15           500.00 usec
p16           1000.00 usec

F2 - Processing parameters
SI            65536
SP            125.7804076 MHz
WDW           EM
SSB           0
LB            1.00 Hz
GB            0
PC            2.00
```

1H spectrum



2.852
2.829
2.818
2.805
2.793
2.782
2.768
2.056
2.044
2.029
2.017
2.002
1.992
1.976
1.964
1.949
1.937
1.928
1.922
1.845
1.814
1.744
1.725
1.709
1.701
1.683
1.665
1.651
1.644
1.628
1.617
1.604
1.592
1.559
1.530
1.517
1.513
1.496
1.483
1.474
1.468
1.462
1.453
1.444
1.438
1.428
1.420
1.413
1.405
1.393
1.382
1.368
1.354
1.343
1.334
1.328
1.319
1.309
1.296
1.120

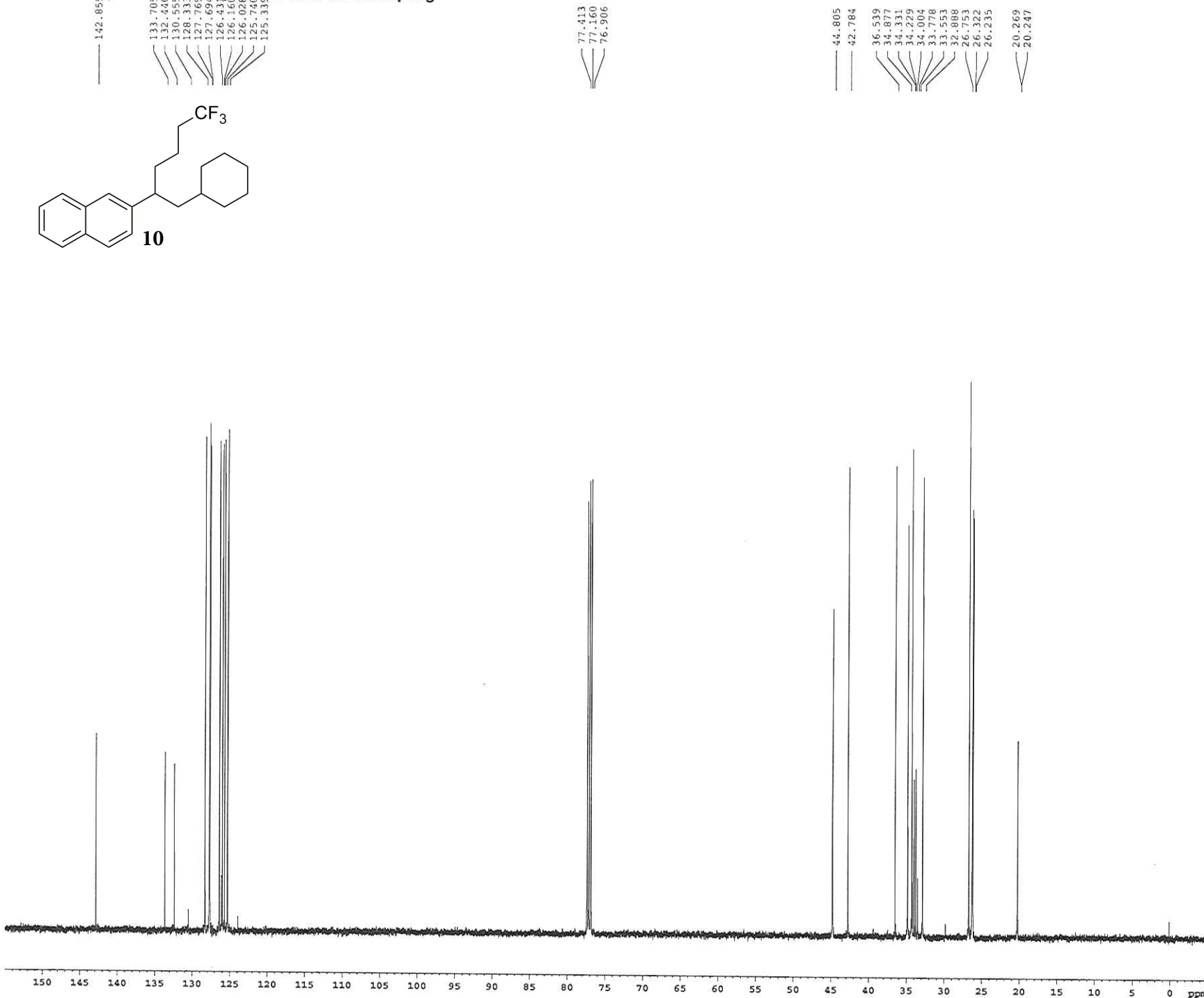
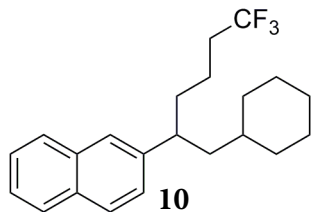
Current Data Parameters
USER aaronj
NAME AGJ_2_235_c1
EXFNO 1
PROCNO 1

F2 - Acquisition Parameters
Date_ 20110128
Time 11.08
INSTRUM drx400
PROBHD 5 mm QNP H/P/P
PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 8
DS 2
SWH 6410.256 Hz
FIDRES 0.097813 Hz
AQ 5.1119579 sec
RG 143.7
EW 78.000 usec
DE 4.50 usec
TE 298.1 K
D1 0.10000000 sec
MCHSST 0.00000000 sec
MCORR 0.01500000 sec

===== CHANNEL F1 =====
NUC1 1H
P1 12.00 usec
PL1 -0.60 dB
SFO1 400.1328009 MHz

F2 - Processing parameters
SI 6536
SF 400.1300280 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 2.00

Z-restored spin-echo 13C spectrum with 1H decoupling



Current Data Parameters
 USER [saron]
 NAME AGJ_2_235_r2
 EXPRNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20130129
 Time 8.29
 INSTRUM cryo500
 PROBHD 5 mm CPYHT 1H
 PULPROG SpinEchoPsi0pp.prd
 TD 65536
 SOLVENT CDCl3
 NS 64
 DS 16
 SWH 10303.031 Hz
 FIDRES 0.462368 Hz
 AQ 1.0813940 sec
 RG 2896.3
 DW 16.500 usec
 DE 6.00 usec
 TE 298.0 K
 D1 0.25000000 sec
 d11 0.03000000 sec
 D16 0.00020000 sec
 E17 0.00019000 sec
 XCHHEST 0.00000000 sec
 MCWRK 0.01500000 sec
 P2 31.00 usec

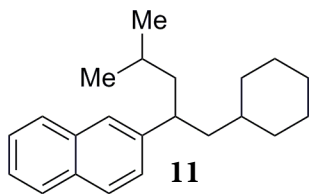
===== CHANNEL f1 =====
 NUC1 13C
 P1 15.50 usec
 P12 500.00 usec
 P12 2000.00 usec
 PL0 120.00 dB
 PLL -1.00 dB
 SFO1 125.7942548 MHz
 SF1 3.20 dB
 SP2 3.20 dB
 SFOAM1 Crp60.0.5.20.1
 SFNAM2 Crp60comp.4
 SFOFF1 0.00 Hz
 SFOFF2 0.00 Hz

===== CHANNEL f2 =====
 CPDPRG2 waltz16
 NUC2 1H
 PCPD2 100.00 usec
 PL2 1.60 dB
 PLL2 24.60 dB
 SFO2 500.2225011 MHz

===== GRADIENT CHANNEL =====
 GPNAM1 SINE.100
 GPNAM2 SINE.100
 GPX1 0.00 %
 GPX2 0.00 %
 GPY1 0.00 %
 GPY2 0.00 %
 GPZ1 30.00 %
 GPZ2 50.00 %
 P15 500.00 usec
 P16 1000.00 usec

F2 - Processing parameters
 SI 65536
 SF 125.7804090 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 2.00

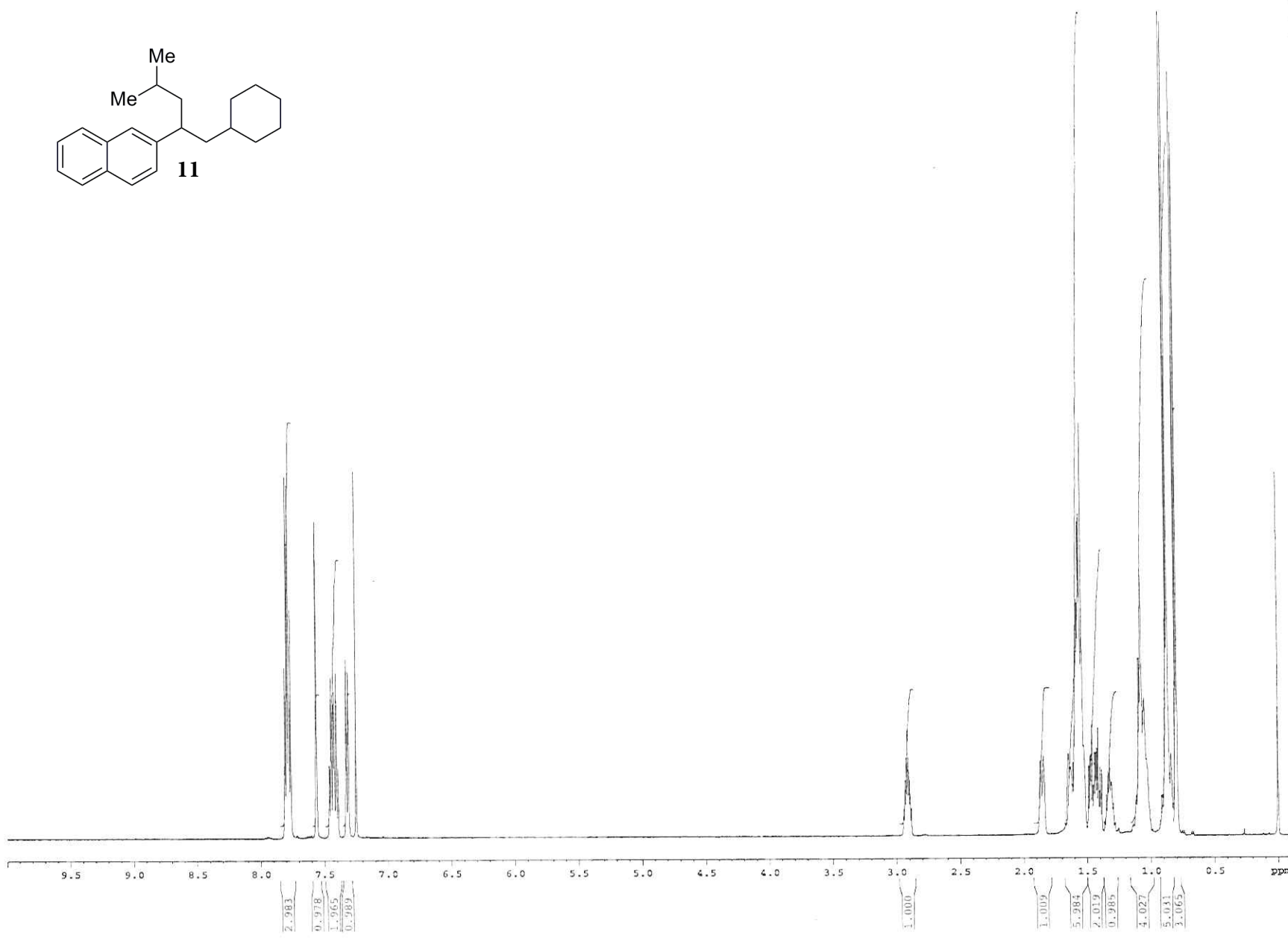
1H spectrum



7.811
7.795
7.782
7.766
7.564
7.462
7.448
7.433
7.423
7.407
7.394
7.340
7.312
7.252

2.935
2.916
2.916
2.916
2.896
2.867
1.852
1.646
1.609
1.599
1.589
1.581
1.572
1.562
1.553
1.548
1.488
1.477
1.471
1.461
1.441
1.423
1.413
1.403
1.396
1.385
1.348
1.335
1.325
1.093
1.074
1.048
0.879
0.866
0.842

Current Data Parameters
NAME yonova
INSTRUM cryo500
PULPROG 5 mm CPTCI 1H-
TD 2930
F2 - Acquisition Parameters
Date_ 20121224
Time 12.55
PROBHD 5 mm CPTCI 1H-
PULPROG 2930
TD 81728
SOLVENT CDCl3
NS 2
DS 2
SWH 8012.820 Hz
FIDRES 0.098043 Hz
AQ 5.0999398 sec
RG 37
DA 62.400 usec
DE 6.00 usec
TE 298.0 K
D1 0.1000000 sec
MREDT 0.2000000 sec
HOWRK 0.0150000 sec
===== CHANNEL f1 =====
NUC1 1H
P1 7.50 usec
PL1 2.00 dB
SFO1 500.2235015 MHz
F2 - Processing parameters
SI 65536
SF 500.2200344 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 4.00



Z-restored spin-echo 13C spectrum with 1H decoupling

Current Data Parameters
 USER yoshida
 NAME DM5 - 125
 EXPNO 222
 PROCNO 1

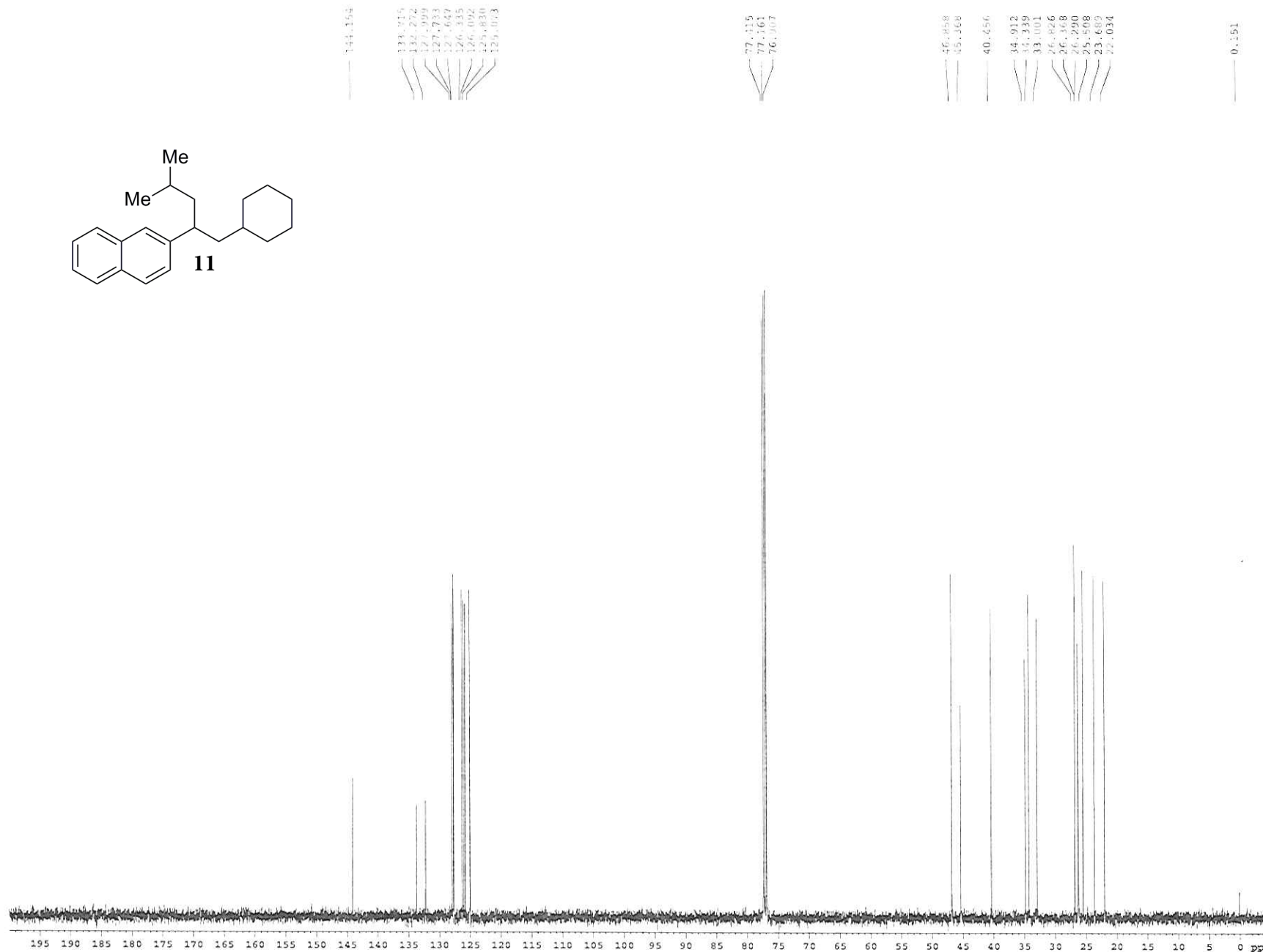
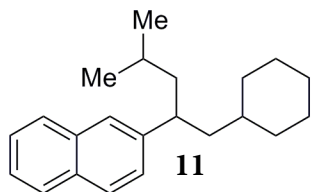
F2 - Acquisition Parameters
 Date_ 20111214
 Time 13:00
 INSTRUM cryo500
 PROBRD 5 mm QNP1H-
 PULPROG SpinEcho30cp
 TD 55336
 SOLVENT cdcl3
 NS 275
 DS 16
 SWH 35383.031 Hz
 HYRES 0.462388 Hz
 AQ 1.0514105 sec
 RG 5150.6
 DM 16.500 usec
 DE 6.00 usec
 TE 298.0 K
 D1 0.3500000 sec
 d11 0.0500000 sec
 d16 0.0000000 sec
 d17 0.0000000 sec
 XCFRST 0.0000000 sec
 XCFR2 0.0150000 sec
 P2 31.00 usec

***** CHANNEL f1 *****
 NUCL 13C
 P1 15.50 usec
 P11 500.00 usec
 P12 2000.00 usec
 PL0 120.00 dB
 PL1 -1.00 dB
 SFO1 125.7642548 MHz
 SF1 3.20 dB
 SFC 3.20 dB
 GPNAM1 Crp50.0.5.120.1
 SPNAM2 Crp60comp.1
 SFOFF1 0.00 Hz
 SFOFF2 0.00 Hz

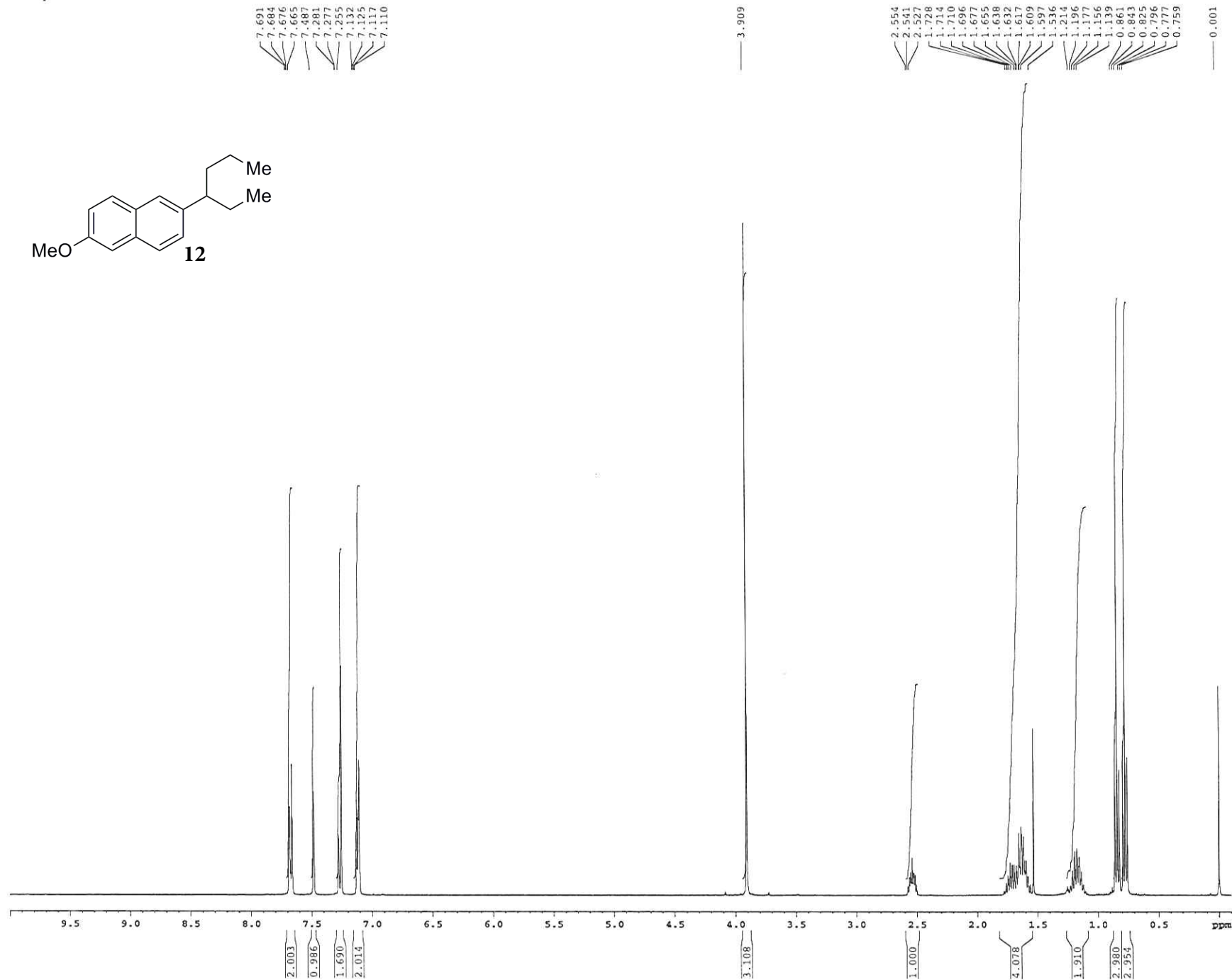
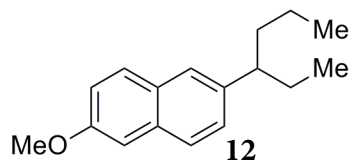
***** CHANNEL f2 *****
 CPDPRG2 waltz16
 NUCL2 1H
 PCPD2 100.00 usec
 PL2 1.60 dB
 PL12 24.60 dB
 SFO2 500.2225011 MHz

***** GRADIENT CHANNEL *****
 GPNAM1 SINE.100
 GPNAM2 SINE.100
 GPX1 0.00 %
 GPX2 0.00 %
 GPY1 0.00 %
 GPY2 0.00 %
 GPZ1 30.00 %
 GPZ2 50.00 %
 p15 500.00 usec
 p16 1000.00 usec

F2 - Processing parameters
 SI 55536
 SF 125.7604076 MHz
 NSM RM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 2.00



1H spectrum



```

Current Data Parameters
USER          yonova
NAME          IMY5 - 606NapRtDP
EXPNO        111
PROCNO       1

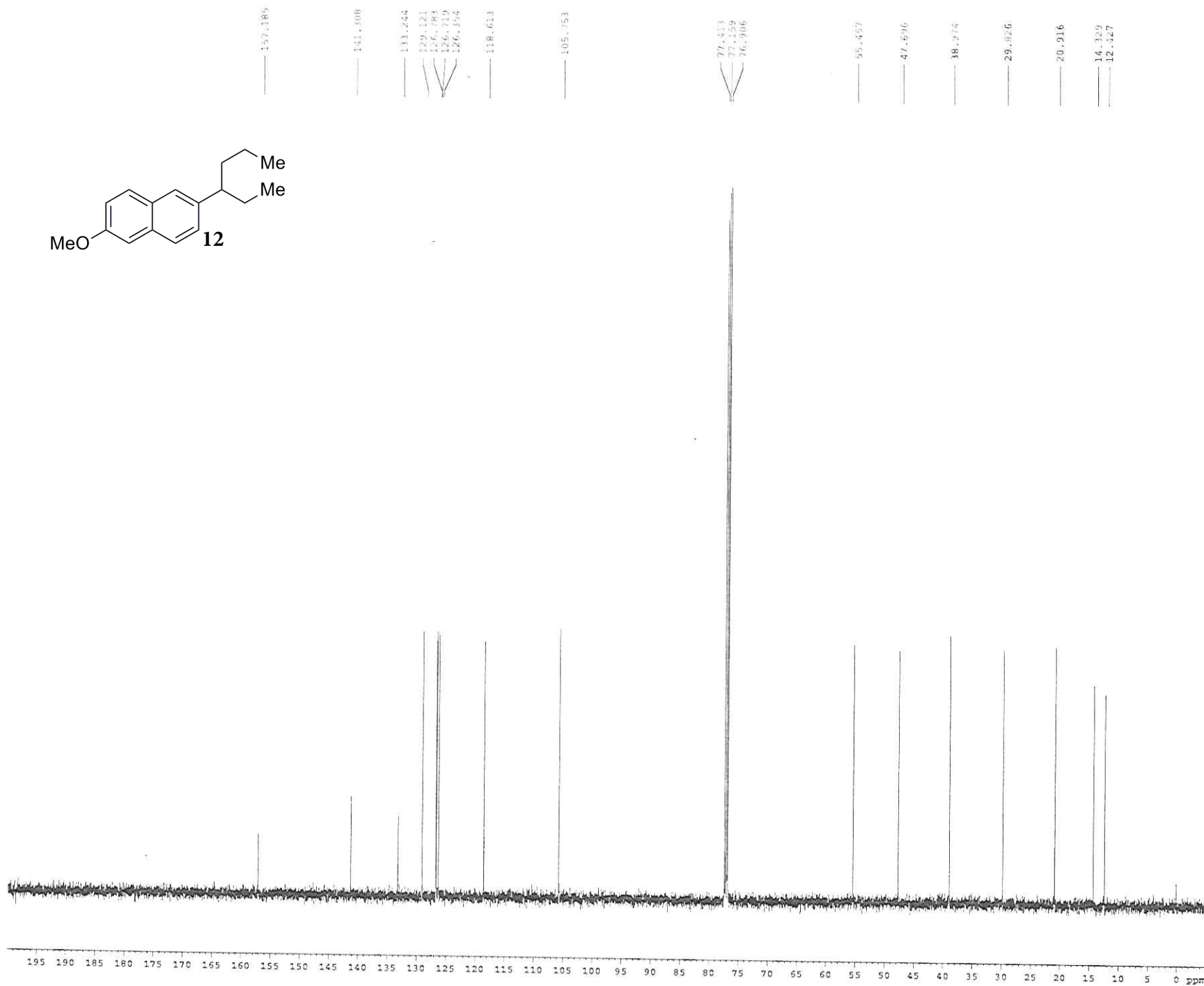
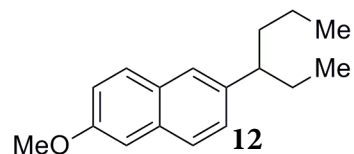
F2 - Acquisition Parameters
Date_        20121128
Time         23.02
INSTRUM      dx400
PROBHD       5 mm QNP 1H/1
PULPROG      zg30
TD           65536
SOLVANT      CDCl3
NS           8
DS           2
SWH          6410.256 Hz
FIDRES       0.097813 Hz
AQ           5.113879 sec
RG           362
DW           78.000 usec
DE           4.50 usec
TE           298.0 K
D1           0.10000000 sec
MCHPST       0.00000000 sec
MCWRK        0.01500000 sec

===== CHANNEL f1 =====
NUC1         1H
P1           12.00 usec
PL1         -0.60 dB
SFO1         400.1326009 MHz

F2 - Processing parameters
SI           65536
SF           400.1300231 MHz
WDW          EM
SSB          0
LB           0.30 Hz
GB           0
CB           2.00

```


Z-restored spin-echo 13C spectrum with 1H decoupling



```

Current Data Parameters
=====
UNIQ          Y
NAME          DM4 - 129
EXPNO        222
PROCNO        1

F2 - Acquisition Parameters
Date_         2012129
Time_         5:07
INSTRUM       cryo500
PROBHD        5 mm CPXI 1H-
PULPROG       SpinEcho10pp.prd
TD            65536
SOLVENT       CDCl3
NS            378
DS            4
SWH           30303.031 Hz
FIDRES        0.462388 Hz
AQ            1.0414105 sec
RG            2580.3
PC            2580.3
DQ            16.500 usec
DE            6.00 usec
TE            284.0 K
D1            0.25000000 sec
d11           0.03000000 sec
d15           0.00250000 sec
d17           0.00019000 sec
MCHEMT        0.00000000 sec
MORPH         0.01500000 sec
P2            31.00 usec

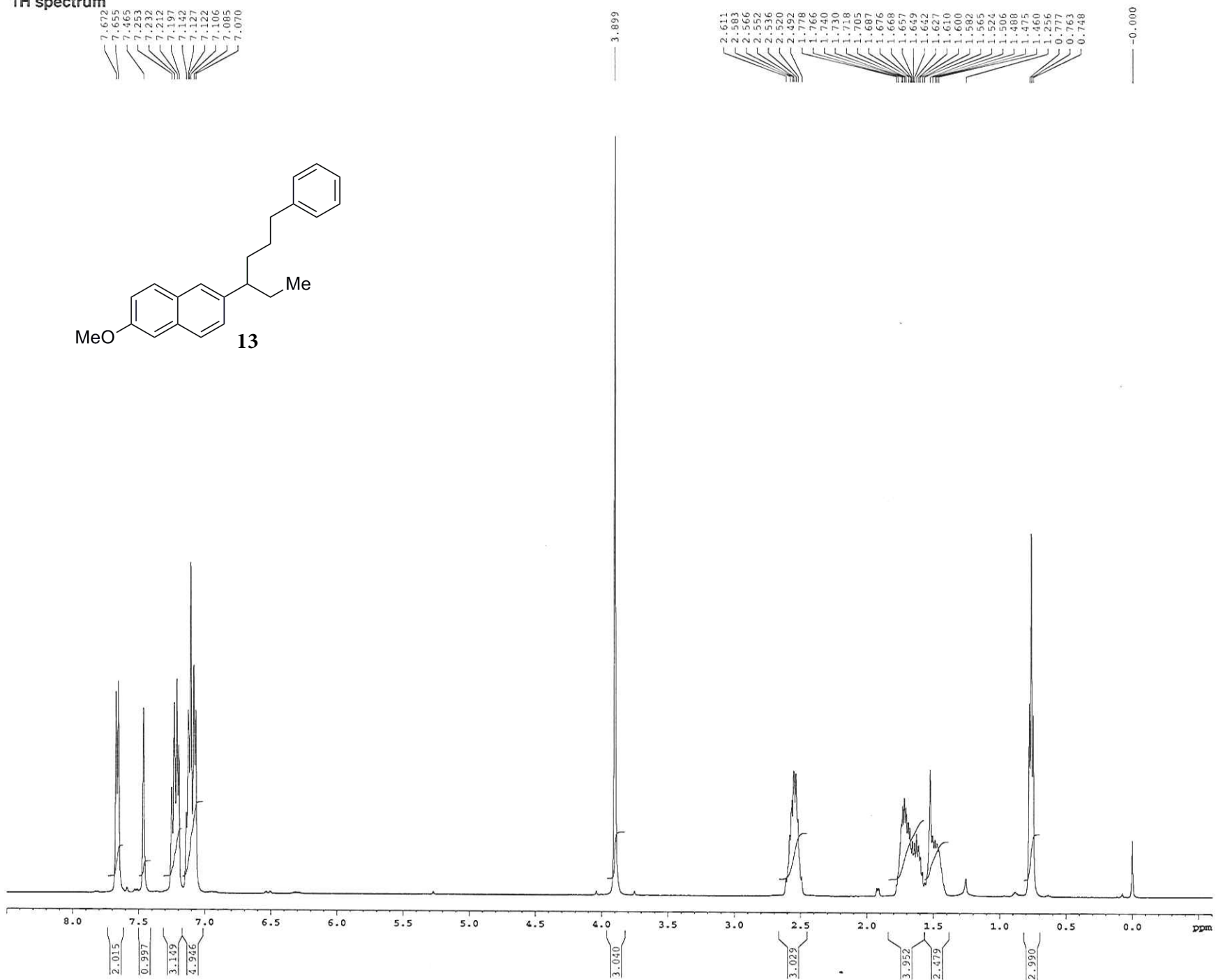
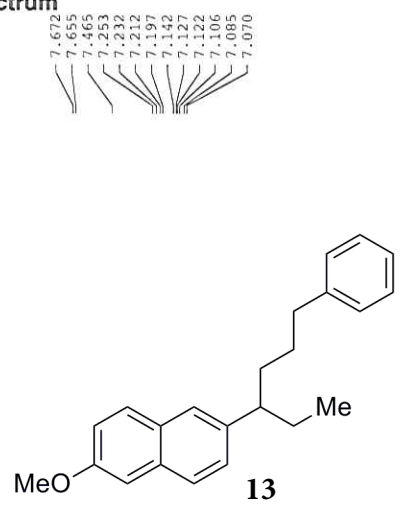
===== CHANNEL f1 =====
NUC1          13C
P1            15.50 usec
P11           500.00 usec
P12           2000.00 usec
PL0           120.00 dB
PL1           -1.00 dB
SFO1         125.7844071 MHz
SF1           3.20 dB
SP2           3.20 dB
SFO1         Crp60.0.5.20.1
SFO2         Crp60comp.4
SFOFF1        0.00 Hz
SFOFF2        0.00 Hz

===== CHANNEL f2 =====
CPDPRG2       walcz16
NUC2          1H
PCPD2         100.00 usec
PL2           1.00 dB
PL12          24.80 dB
SFO2         500.1325011 MHz

===== GRADIENT CHANNEL =====
GPNAM1        SINE.100
GPNAM2        SINE.100
CP1           0.00 %
CP2           0.00 %
CPV1          0.00 %
CPV2          0.00 %
GP11          30.00 %
GP22          50.00 %
PL5           500.00 usec
PL6           1000.00 usec

F2 - Processing parameters
SI            5536
SF            125.7804071 MHz
RG            84
SSB           0
LB            1.00 Hz
GB            0
PC            2.00
    
```

1H spectrum



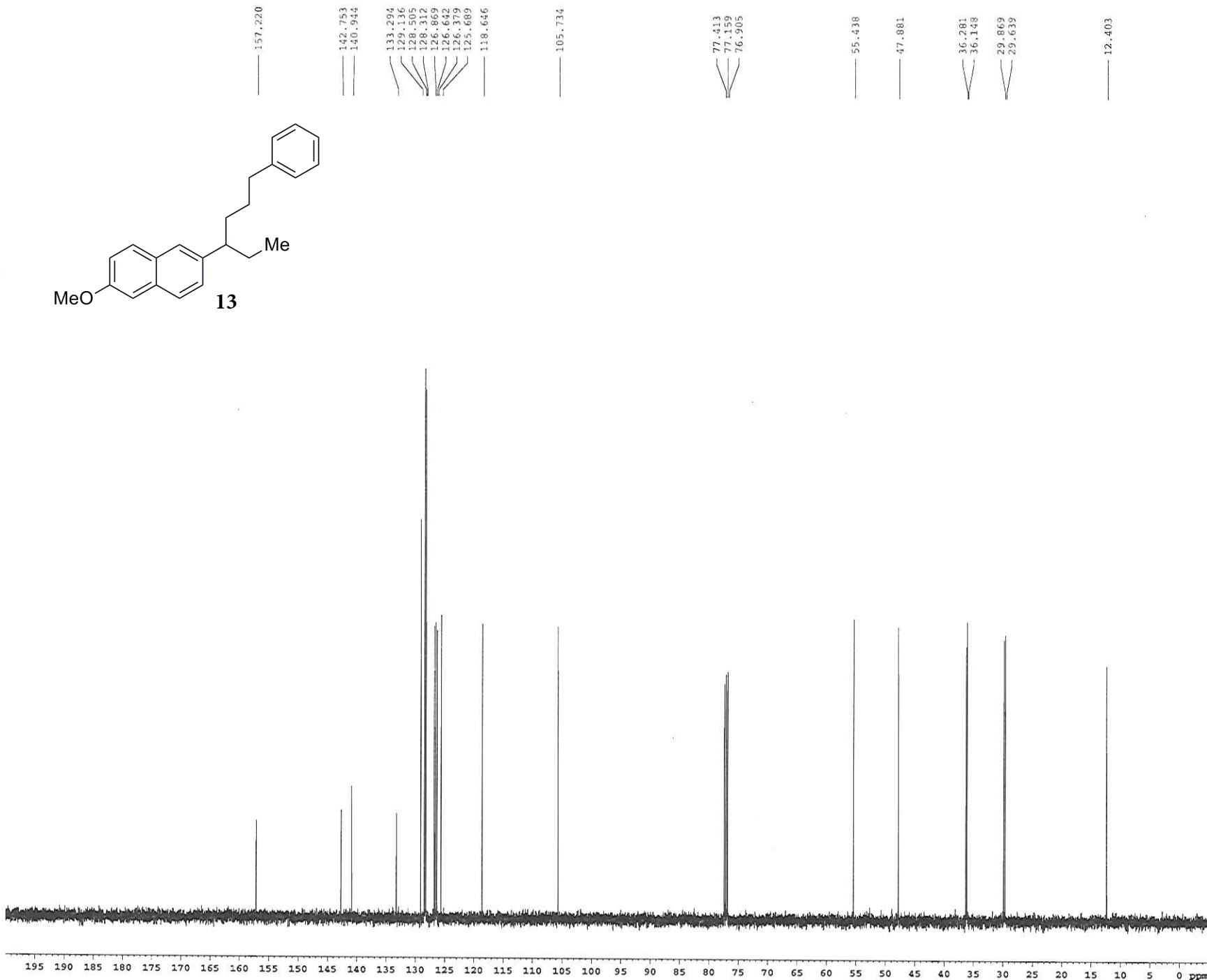
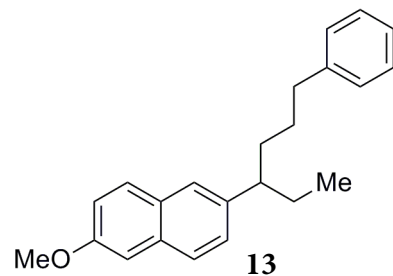
Current Data Parameters
 UMR yonova
 NAME imy5 - 099 - bottom
 ETVNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20121203
 Time_ 20.47
 INSTRUM gn500
 PROBHD 5 mm broadband
 PULPROG zg30
 TD 81728
 SOLVENT CDCl3T
 NS 8
 DS 2
 SWH 8012.820 Hz
 FIDRES 0.098043 Hz
 AQ 5.0998774 sec
 RG 114
 DW 62.400 usec
 DE 6.00 usec
 TE 298.0 K
 DI 0.10000000 sec
 MCHYST 0.80000000 sec
 MCWPK 0.01500000 sec

===== CHANNEL f1 =====
 NUCL 1H
 P1 12.20 usec
 PL1 -5.00 dB
 SFO1 499.4034958 MHz

F2 - Processing parameters
 SI 6536
 SF 499.4000446 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

Z-restored spin-echo ¹³C spectrum with ¹H decoupling



Current Data Parameters
 USER yonova
 NAME IVYS - 099
 EXPNO 222
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20121203
 Time_ 21.04
 INSTRUM cryo000
 PROBHD 5 mm CPXI 1H-
 PULPROG SpinEchopg30op.prd
 TD 65536
 SOLVENT CDCl3
 NS 56
 DS 26
 SWH 30303.031 Hz
 FIDRES 0.462388 Hz
 AQ 1.0813940 sec
 RG 4096
 DW 16.500 usec
 DE 6.00 usec
 TE 298.0 K
 D1 0.25000000 sec
 d11 0.03000000 sec
 d16 0.00200000 sec
 d17 0.00190000 sec
 MCERS2 0.00000000 sec
 MCMK 0.01500000 sec
 P2 31.00 usec

===== CHANNEL f1 =====
 NUC1 ¹³C
 P1 15.50 usec
 P11 500.00 usec
 P12 2000.00 usec
 PL0 120.00 dB
 PL1 -1.00 dB
 SFO1 125.7942548 MHz
 SP1 3.20 dB
 SP2 3.20 dB
 SPNAM1 Crp60.0.5.20.1
 SPNAM2 Crp60comp.4
 SPOFF1 0.00 Hz
 SPOFF2 0.00 Hz

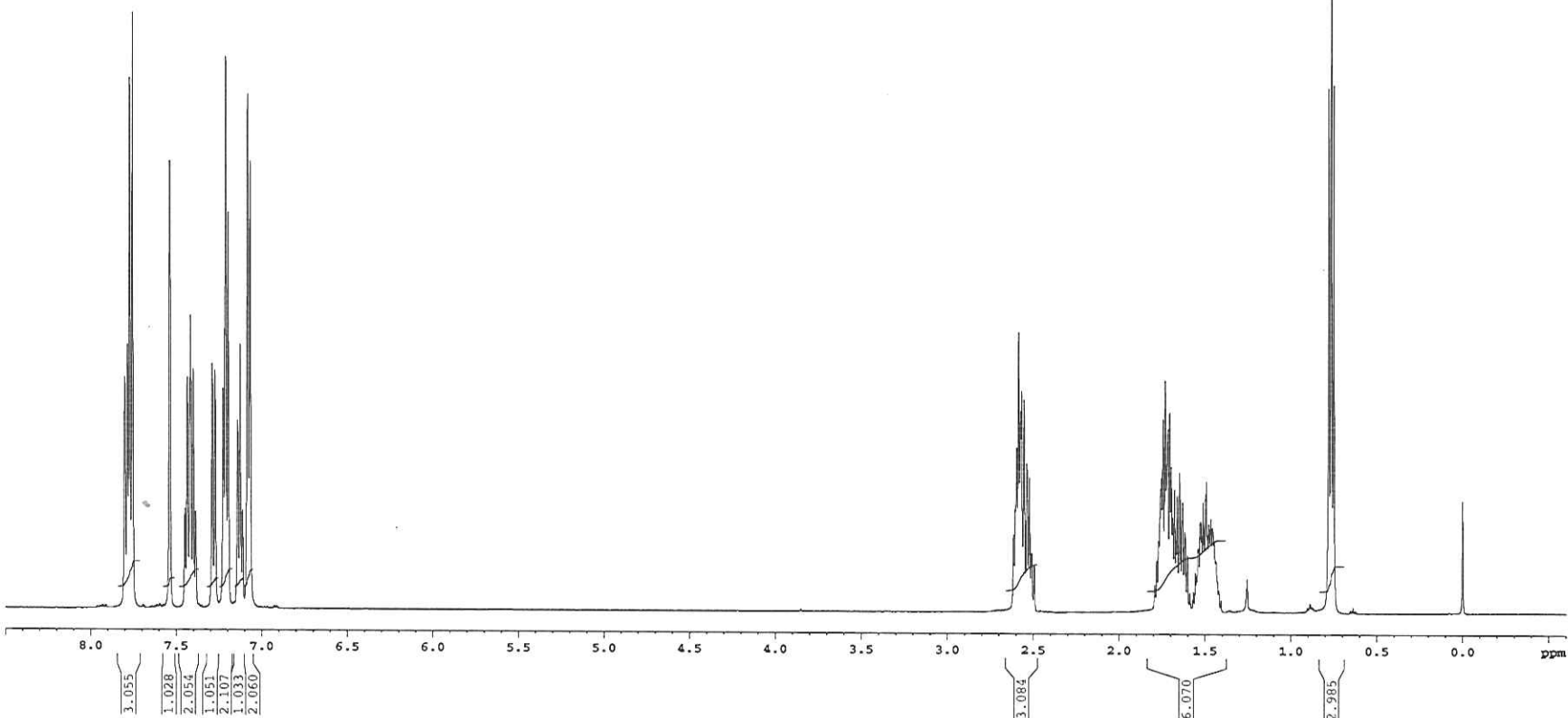
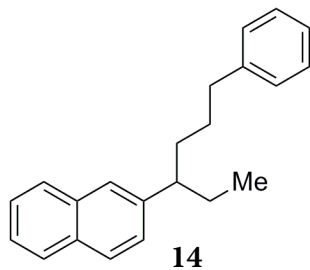
===== CHANNEL f2 =====
 CPDPRG2 waltz16
 NUC2 ¹H
 PCPD2 100.00 usec
 PL2 2.00 dB
 PL12 24.00 dB
 SFO2 500.2225011 MHz

===== GRADIENT CHANNEL =====
 GPNAM1 SINE.100
 GPNAM2 SINE.100
 GPX1 0.00 %
 GPX2 0.00 %
 GPY1 0.00 %
 GPY2 0.00 %
 GPZ1 30.00 %
 GPZ2 50.00 %
 p15 500.00 usec
 p16 1000.00 usec

F2 - Processing parameters
 SI 65536
 SF 125.7804113 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 2.00

1H spectrum

7.803
7.787
7.774
7.757
7.540
7.450
7.437
7.419
7.402
7.389
7.292
7.290
7.276
7.273
7.228
7.213
7.198
7.143
7.129
7.114
7.085
7.070



2.615
2.605
2.596
2.587
2.575
2.568
2.555
2.540
2.536
2.523
2.512
2.509
2.495
1.781
1.771
1.765
1.758
1.754
1.744
1.733
1.721
1.716
1.706
1.696
1.687
1.677
1.669
1.662
1.648
1.645
1.630
1.620
1.617
1.602
1.556
1.550
1.542
1.538
1.530
1.524
1.518
1.511
1.497
1.493
1.479
1.466
1.458
1.454
1.448
1.440
1.436
0.781
0.766
0.751
0.000

Current Data Parameters
 USER aaronj
 NAME AGJ_2_142_c2
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20130523
 Time 11.50
 INSTRUM cryo500
 PROBHD 5 mm CPTCI 1H-
 PULPROG zg30
 TD 8176
 TO
 SOLVENT CDCl3
 NS 8
 DS 2
 SWH 8012.820 Hz
 FIDRES 0.098043 Hz
 AQ 5.0998774 sec
 RG 6.3
 DW 62.400 usec
 DE 6.00 usec
 TE 298.0 K
 D1 0.10000000 sec
 MCREST 0.00000000 sec
 MCWRRK 0.01500000 sec

===== CHANNEL f1 =====
 NUC1 1h
 P1 7.50 usec
 PLL 1.60 dB
 SFO1 500.2235015 MHz

F2 - Processing parameters
 SI 6536
 SF 500.2200519 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 4.00

Z-restored spin-echo ¹³C spectrum with ¹H decoupling

143.280
142.710
133.670
132.378
128.511
128.331
128.023
127.720
127.653
126.577
126.090
125.880
125.710
125.15

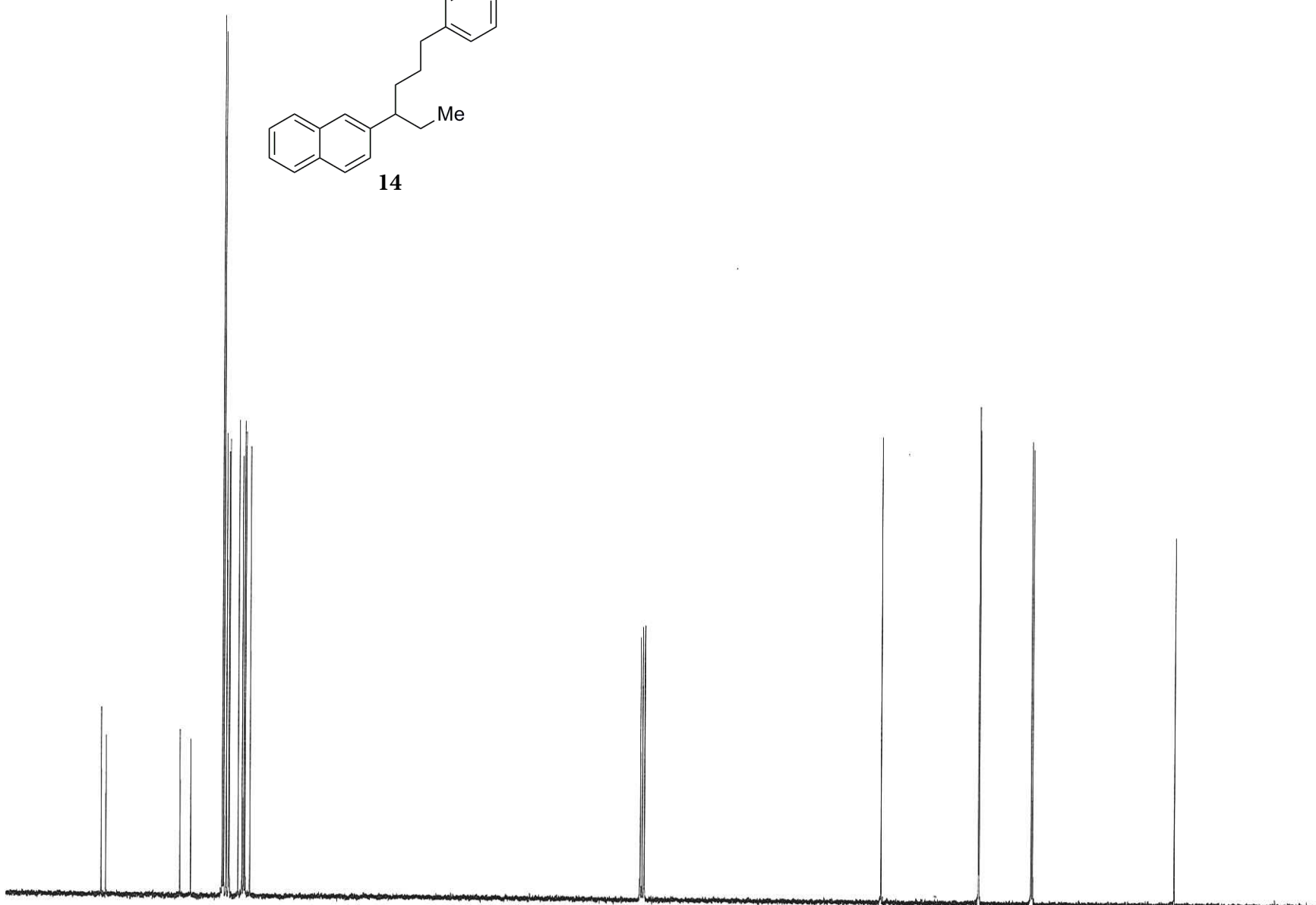
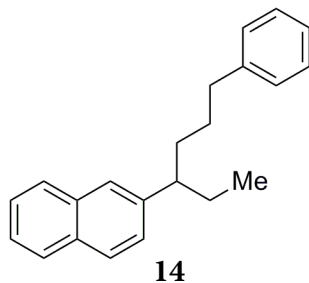
77.416
77.162
76.908

48.101

36.226
36.151

29.826
29.639

12.409



Current Data Parameters
USER Aaron
NAME AG1_2_142.c
EXPNO 2
PROCNO 1

F2 - Acquisition Parameters
Date_ 20130523
Time 11.54
INSTRUM cryo500
PROBHD 5 mm CPXI 1H-
PULPROG SpinEchoq10p.prd
TD 65536
SOLVENT CDCl3
NS 332
DS 16
OS 16
SMH 30303.031 Hz
FIDRES 0.462388 Hz
AQ 1.0813940 sec
RG 4597.6
DW 16.500 usec
DE 6.00 usec
TE 298.0 K
D1 0.25000000 sec
d11 0.03000000 sec
d16 0.00020000 sec
d17 0.00019600 sec
MCHPST 0.00000000 sec
MCHRK 0.01500000 sec
P2 11.00 usec

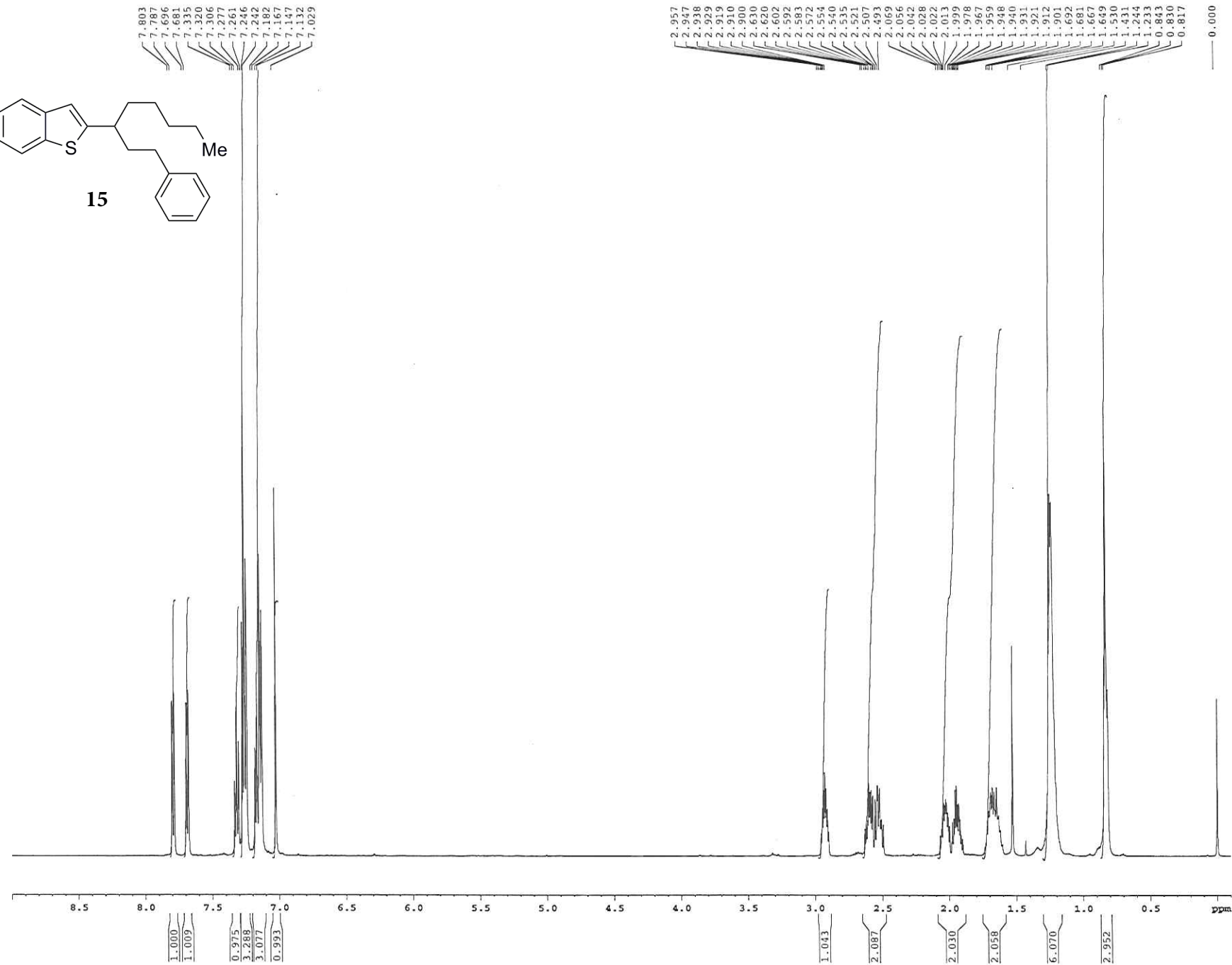
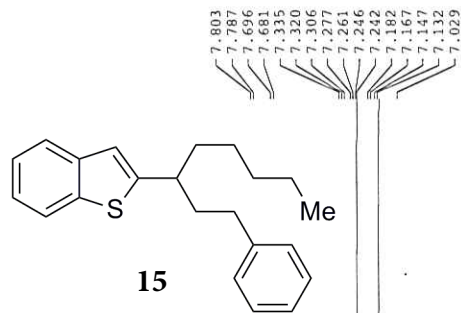
***** CHANNEL f1 *****
NUC1 ¹³C
P1 15.50 usec
PL1 500.00 usec
PL2 2000.00 usec
PL0 120.00 dB
PL1 -1.00 dB
SFO1 125.7942544 MHz
SF1 3.20 dB
SP2 1.20 dB
SPNAM1 Crp60,0.5,20.1
SPNAM2 Crp60comp.4
SFOFF1 0.00 Hz
SPOFF2 0.00 Hz

***** CHANNEL f2 *****
CPDPRG2 waltz16
NUC2 ¹H
PCPD2 100.00 usec
PL2 1.60 dB
PL12 24.60 dB
SFO2 500.225011 MHz

***** GRADIENT CHANNEL *****
GPNAM1 SINE.100
GPNAM2 SINE.100
GPX1 0.00 %
GPX2 0.00 %
GPY1 0.00 %
GPY2 0.00 %
GPZ1 30.00 %
GPZ2 50.00 %
p15 500.00 usec
p16 1000.00 usec

F2 - Processing parameters
SI 65536
SF 125.7804122 MHz
WDW EM
GB 0
CB 1.00 Hz
PC 2.00

1H spectrum



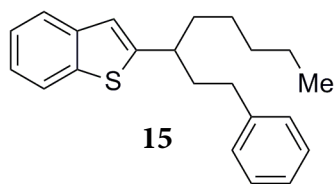
Current Data Parameters
 USER yonova
 NAME imy5 - 210
 EXPNO 111
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20130329
 Time 21.59
 INSTRUM cryo500
 PROBHD 5 mm CPTCI 1H-
 PULPROG zg30
 TD 91728
 SOLVENT CDCl3
 NS 16
 DS 2
 SWH 8012.420 Hz
 FIDRES 0.098043 Hz
 AQ 5.0999398 sec
 RG 6.3
 DM 62.400 usec
 DE 6.00 usec
 TE 298.0 K
 D1 0.10000000 sec
 MCHWST 0.00000000 sec
 MCMRK 0.01500000 sec

===== CHANNEL f1 =====
 NUC1 1H
 P1 7.50 usec
 PL1 1.60 dB
 SFO1 500.2235015 MHz

F2 - Processing parameters
 SI 65536
 SF 500.2200403 MHz
 WDW 0
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 2.00

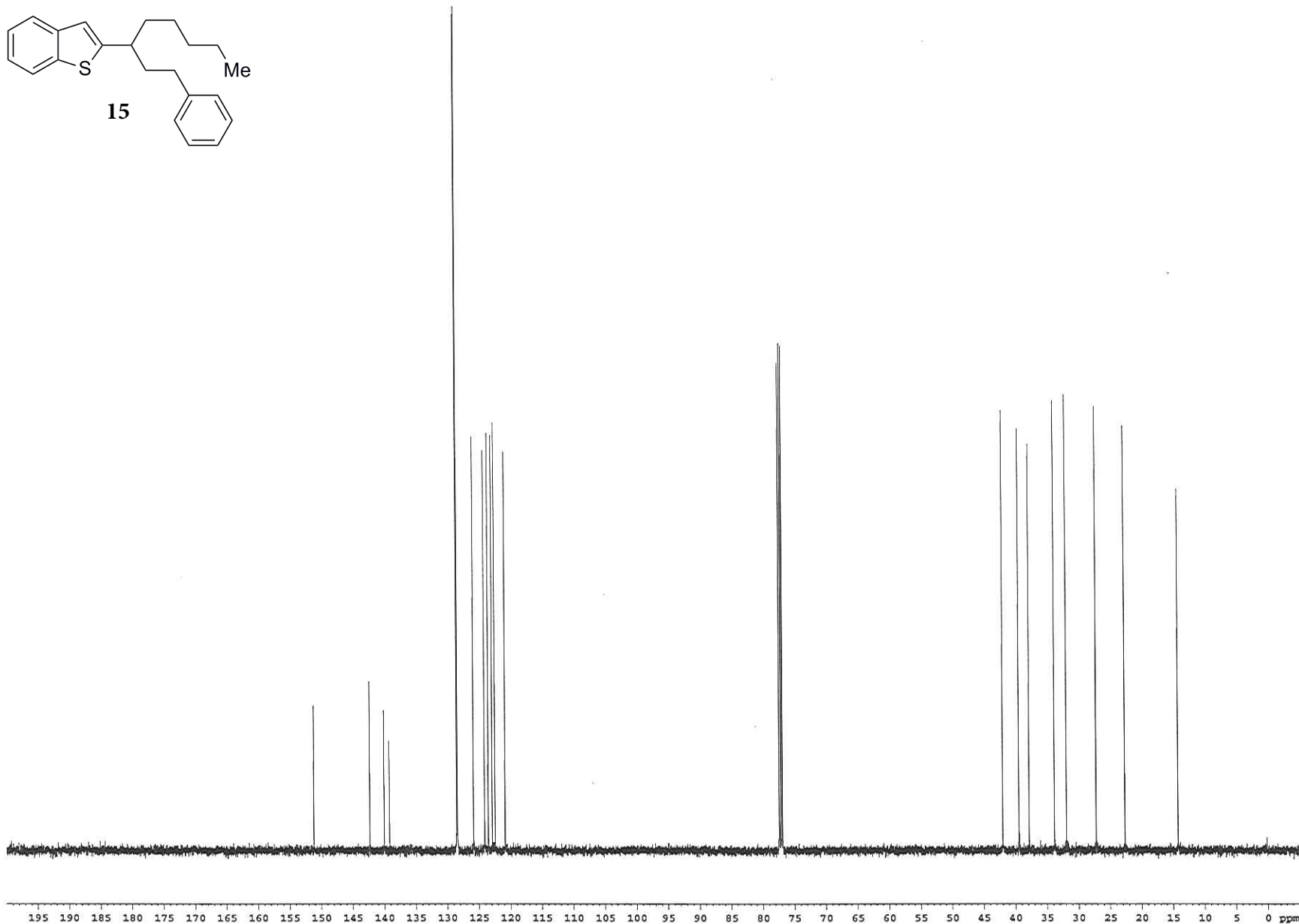
Z-restored spin-echo 13C spectrum with 1H decoupling



151.192
 142.349
 140.062
 139.245
 128.567
 128.450
 125.878
 124.139
 123.517
 122.892
 122.479
 120.860

77.415
 77.161
 76.907

42.000
 39.443
 37.801
 33.807
 31.932
 27.226
 22.673
 14.219



```

Current Data Parameters
USER          yonova
NAME          imy5 - 210
EXPNO        222
PROCNO       1

F2 - Acquisition Parameters
Date_        20130329
Time         22.08
INSTRUM      cryo500
PROBHD       5 mm CPTCI 1H-
PULPROG      SpinEchoq30pp.prd
TD           65536
SOLVENT      CDCl3
NS           304
DS           16
SWH          30303.611 Hz
FIDRES       0.462388 Hz
AQ           1.0814105 sec
RG           7298.2
DK           16.500 usec
DE           6.00 usec
TE           298.0 K
D1           0.25000000 sec
d11          0.03000000 sec
D16          0.00020000 sec
d17          0.00019600 sec
MCRETZ      0.00000000 sec
MORPH       0.01500000 sec
P2           11.00 usec

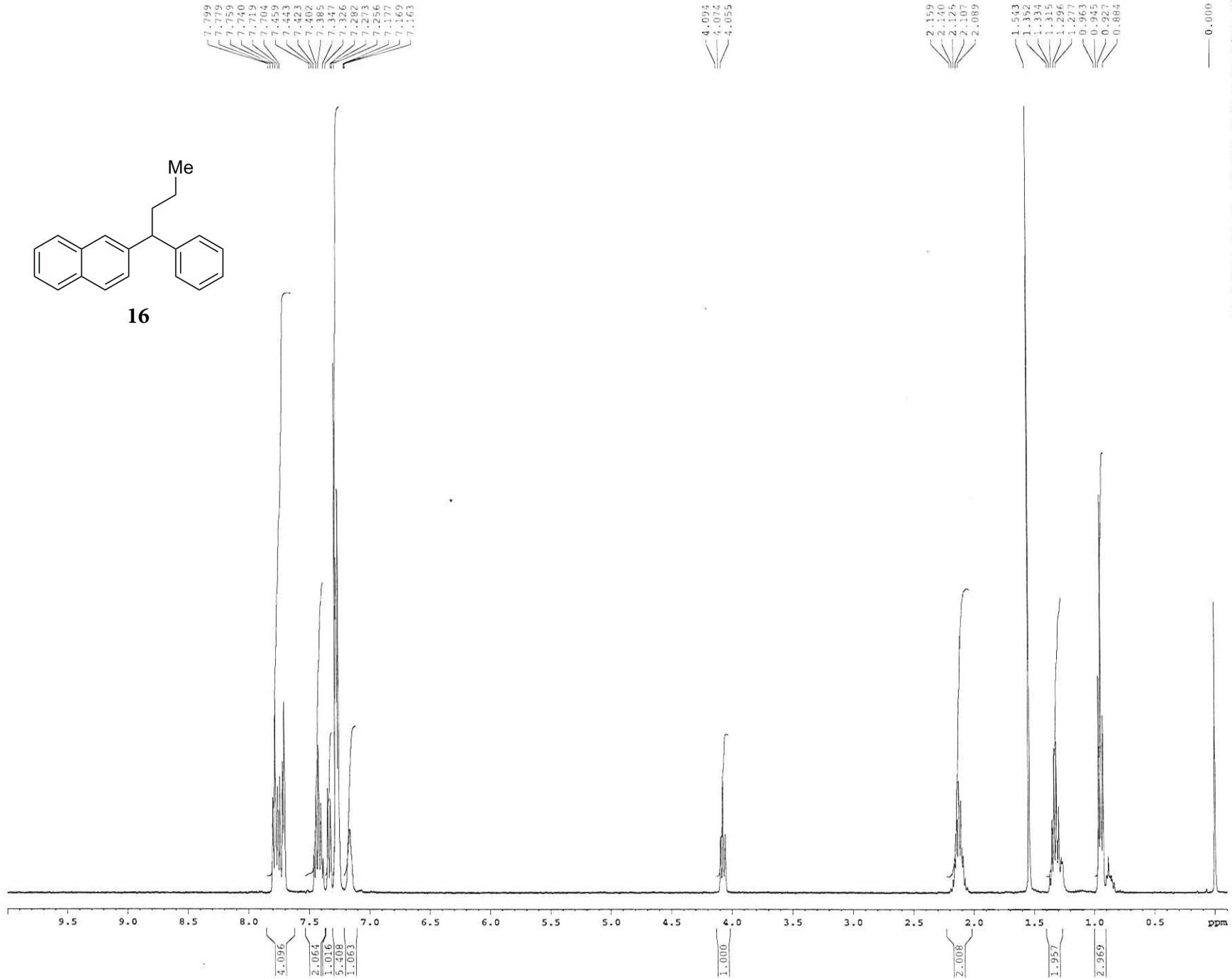
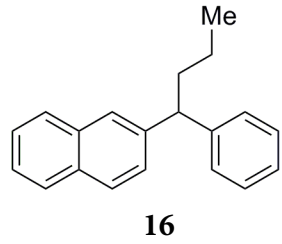
===== CHANNEL f1 =====
NUC1         13C
P1           15.50 usec
P11          500.00 usec
P12          2000.00 usec
PL0          120.00 dB
PL1          -1.00 dB
SFO1         125.7842548 MHz
SP1          3.20 dB
SP2          3.20 dB
SFO2         Crp60.0.5.26.1
SFO3         Crp60comp.4
SPOFF1       0.00 Hz
SPOFF2       0.00 Hz

===== CHANNEL f2 =====
CPDPRG2      waltz16
NUC2         1H
PCPD2       100.00 usec
PL2         1.60 dB
PL12        24.60 dB
SFO2        500.2225011 MHz

===== GRADIENT CHANNEL =====
GPNAM1       SINE.100
GPNAM2       SINE.100
GFX1         0.00 %
GFX2         0.00 %
GPY1         0.00 %
GPY2         0.00 %
GZ1          30.00 %
GZ2          50.00 %
p15          500.00 usec
p16          1000.00 usec

F2 - Processing parameters
SI           65536
SF           125.7804099 MHz
WDW          EM
SSB          0
LB           1.00 Hz
GB           0
PC           2.00
  
```

1H spectrum



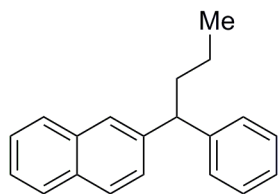
Current Data Parameters
 Date_ 20110807
 User_ yonova
 NAME_ imy5 - 143
 EXPNO_ 31
 PROCNO_ 1

F2 - Acquisition Parameters
 Date_ 10.11
 Time_ 19.11
 INSTRUM_ drx400
 PROBRD_ 5 mm QNP H/P/P
 PULPROG_ zg30
 TD_ 65536
 SOLVENT_ CDCl3
 NS_ 2
 DS_ 2
 SWH_ 6410.256 Hz
 FIDRES_ 0.097813 Hz
 AQ_ 5.118579 sec
 RG_ 406.4
 DW_ 78.000 usec
 DE_ 4.50 usec
 TE_ 299.1 K
 EI_ 0.10000000 sec
 MCREST_ 0.00000000 sec
 MCWRR_ 0.01500000 sec

***** CHANNEL f1 *****
 NUC1_ 1H
 P1_ 12.00 usec
 PL1_ -0.60 db
 SFO1_ 400.1320009 MHz

F2 - Processing parameters
 SI_ 65536
 SF_ 400.1300227 MHz
 WDW_ EM
 SSB_ 0
 LB_ 0.30 Hz
 GB_ 0
 PC_ 2.00

Z-restored spin-echo ¹³C spectrum with ¹H decoupling



16

145.321
142.882
133.659
132.238
128.520
128.134
128.119
127.830
127.690
126.967
126.202
126.022
125.988
125.435

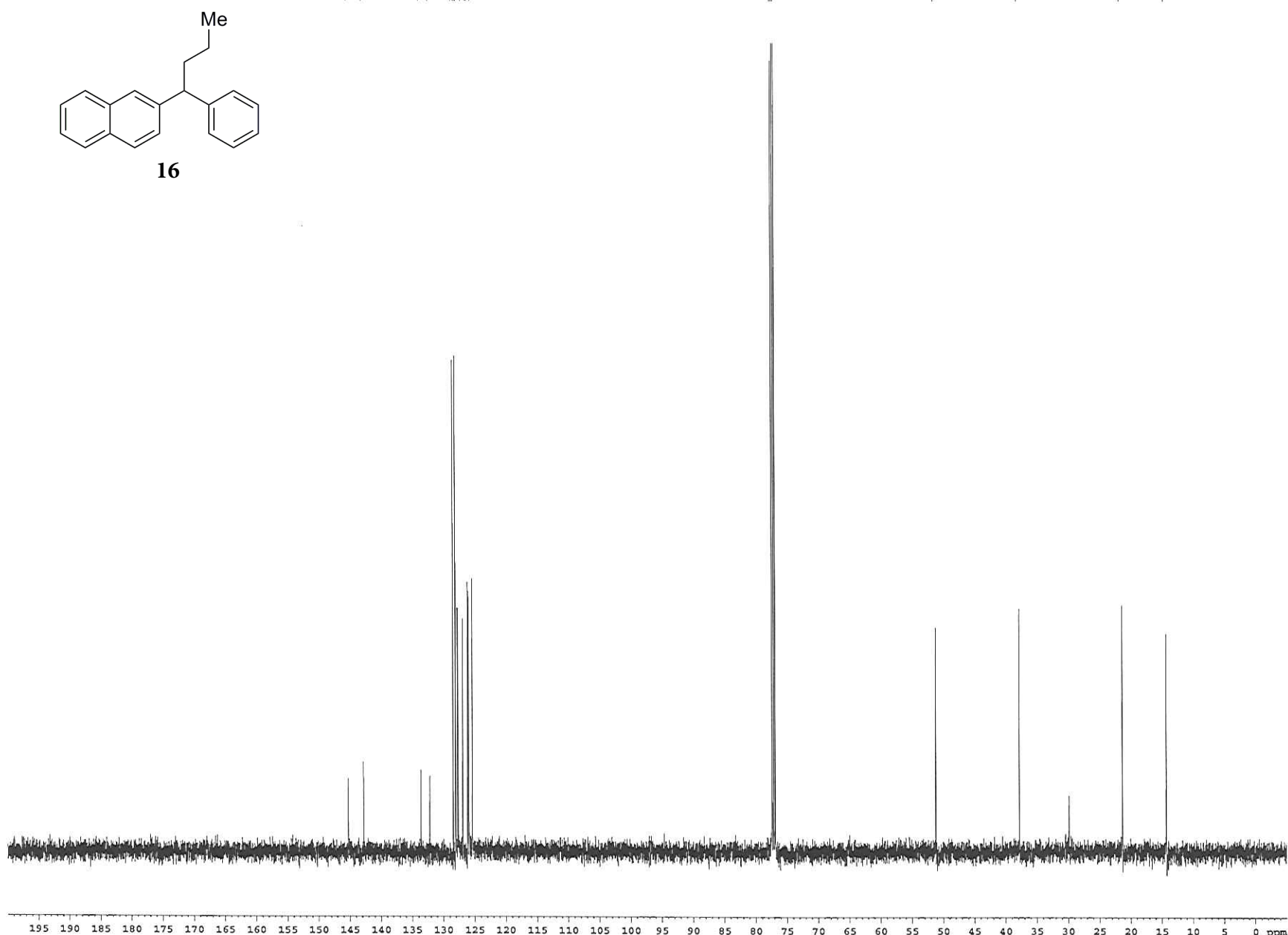
77.414
77.160
76.906

51.209

37.809

21.326

14.256



```

Current Data Parameters
USER          yonova
NAME          imy1 - 143
EXPNO        22222
PROCNO       1

F2 - Acquisition Parameters
Date_        20130809
Time         16.04
INSTRUM      cryo500
PROBHD       5 mm CPTCI 1H-
PULPROG      SpinEchoq30qp.prd
TD           65536
SOLVSMT      CDCl3
NS           218
DS           16
CWX          30303.031 Hz
FIDRES       0.462388 Hz
AQ           1.0814109 sec
PC           9195.2
DM           16.500 usec
DE           6.00 usec
TE           298.0 K
D1           0.25000000 sec
d11          0.03000000 sec
DL6          0.00020000 sec
d17          0.00019000 sec
MCHRGFT      0.00000000 sec
MCMRK        0.01500000 sec
F2           31.00 usec

===== CHANNEL f1 =====
NUC1         13C
P1           15.50 usec
PL1          500.00 usec
P12          2000.00 usec
PL0          120.00 dB
PL1         -1.00 dB
SFO1        125.7942548 MHz
SP1          3.20 dB
SP2          3.20 dB
GPNAM1       Crp60,0.5,20.1
GPNAM2       Crp60comp,4
SPOFF1       0.00 Hz
SPOFF2       0.00 Hz

===== CHANNEL f2 =====
CPDPRG2      waltz16
NUC2         1H
PCPD2        100.00 usec
PL2          1.60 dB
PL12         24.60 dB
SFO2        500.2225011 MHz

===== GRADIENT CHANNEL =====
GPNAM1       SINE.100
GPNAM2       SINE.100
GPX1         0.00 %
GPX2         0.00 %
GPY1         0.00 %
GPY2         0.00 %
GPR1         30.00 %
GPR2         50.00 %
p15         500.00 usec
p16         1000.00 usec

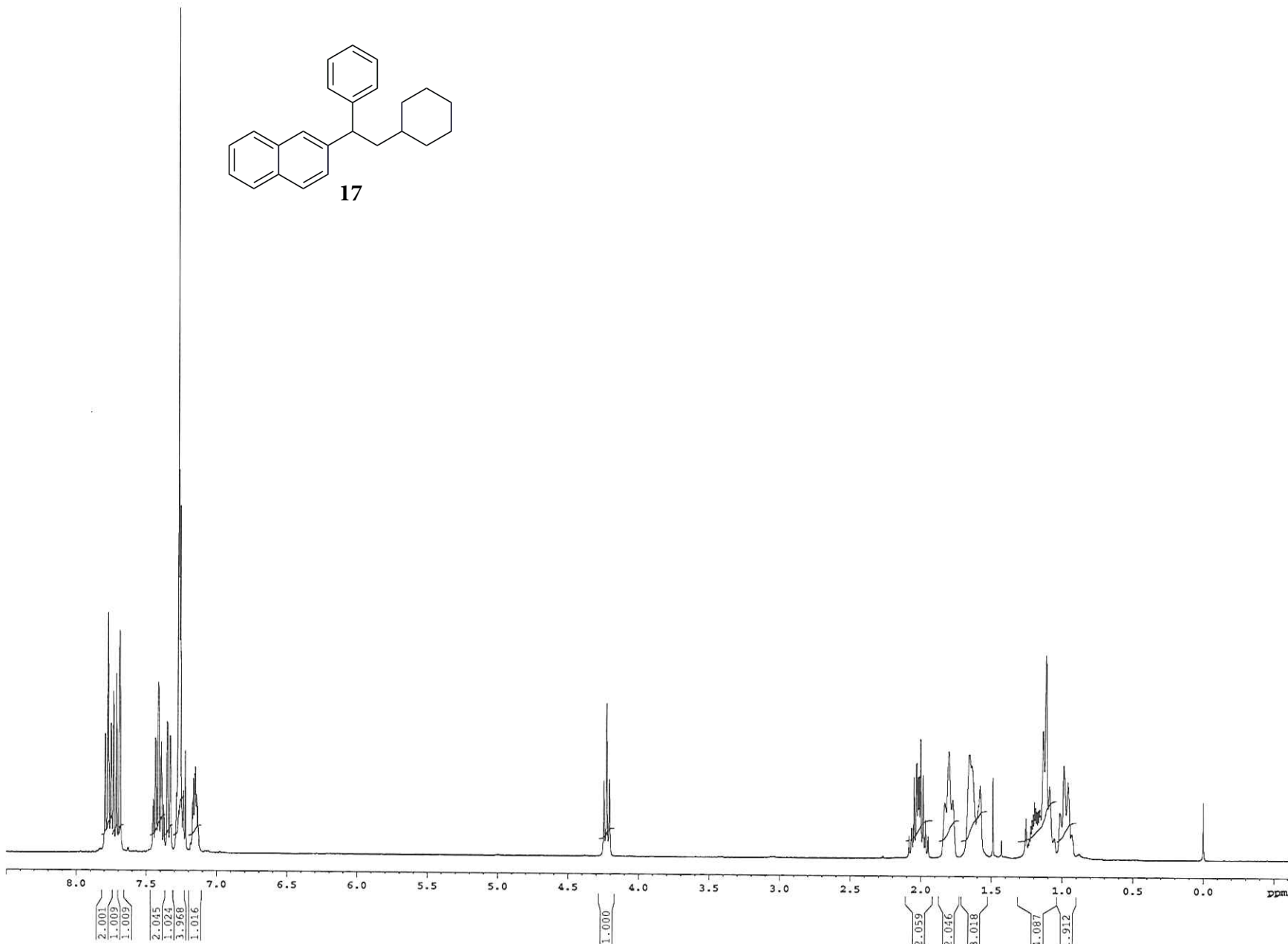
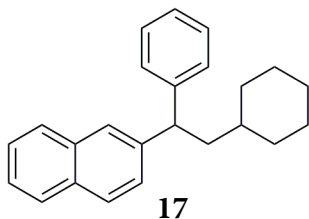
F2 - Processing parameters
SI           65536
SF           125.7804085 MHz
WDW          EM
SSB          0
LB           1.00 Hz
GB           0
PC           2.00
    
```

1H spectrum

7.735
7.714
7.690
7.453
7.450
7.436
7.433
7.415
7.388
7.372
7.352
7.342
7.326
7.321
7.306
7.286
7.272
7.265
7.239
7.223
7.213
7.172
7.166
7.158
7.151
7.144
7.136

4.246
4.226
4.206

2.067
2.064
2.049
2.032
2.030
2.020
2.013
2.002
1.984
1.968
1.831
1.801
1.773
1.655
1.641
1.636
1.592
1.579
1.489
1.255
1.222
1.213
1.204
1.195
1.186
1.177
1.168
1.160
1.152
1.133
1.113
1.088
1.016
0.987
0.959
0.000



Current Data Parameters
 USER aaronj
 NAME AGJ_2_231
 EXPRNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20130108
 Time 11.57
 INSTRUM brx400
 PROBNM 5 mm QNP H/P/P
 PULPROG zg30
 TD 65536
 SOLVENT CDCl3
 NS 8
 DS 2
 SWH 6410.256 Hz
 FIDRES 0.097813 Hz
 AQ 5.118579 sec
 RG 114
 DW 78.000 usec
 DE 4.50 usec
 TE 298.0 K
 D1 0.10000000 sec
 MCRECT 0.00000000 sec
 MCWRK 0.01500000 sec

===== CHANNEL f1 =====
 NUC1 1H
 P1 12.00 usec
 PL1 -0.60 dB
 SF01 400.1328009 MHz

F2 - Processing parameters
 SI 65536
 SF 400.1300361 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 2.00

13C spectrum with 1H decoupling

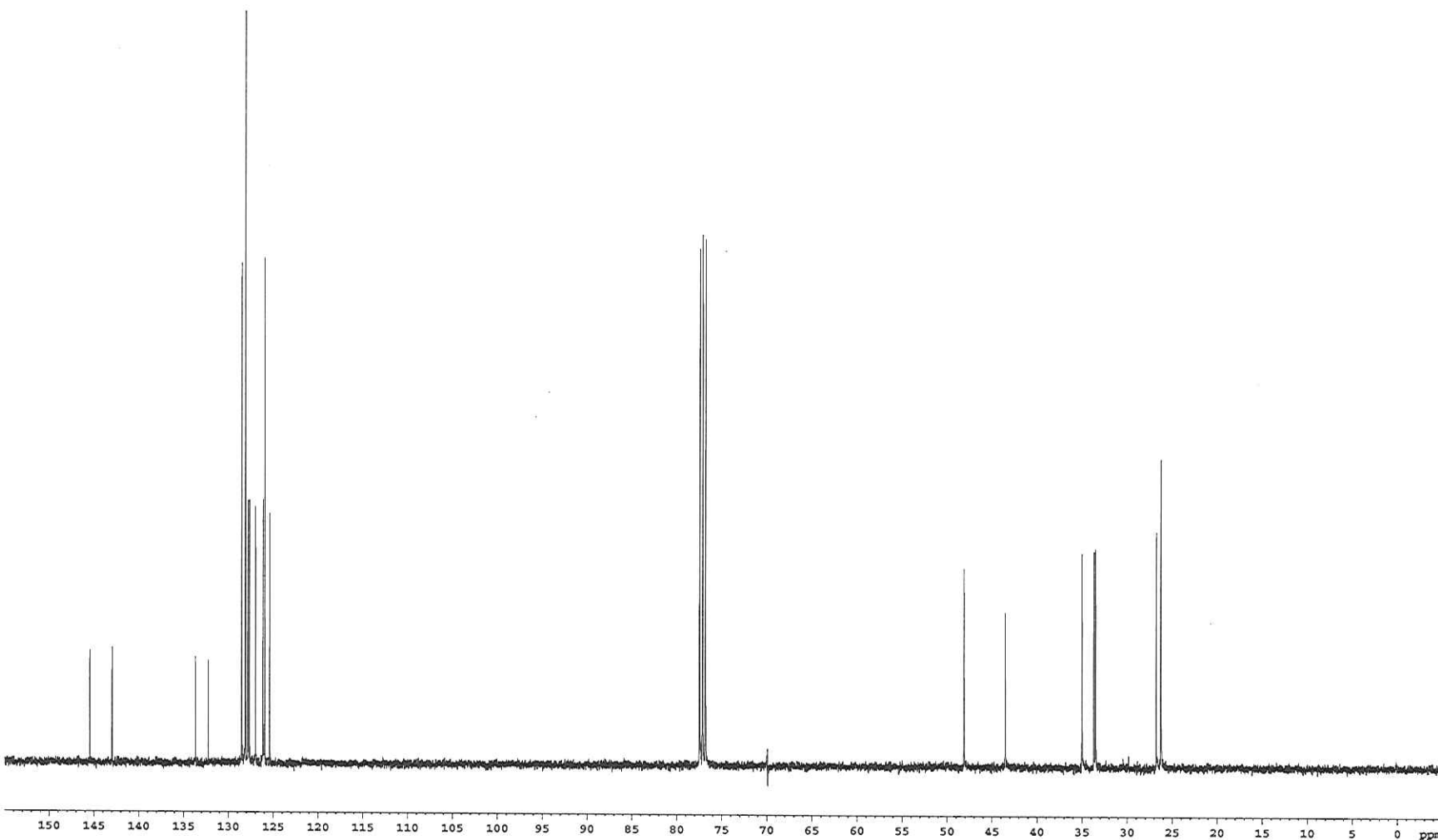
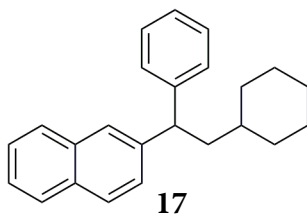
145.495
143.001
133.711
132.249
128.545
128.131
127.855
127.690
127.024
126.163
125.422

77.478
77.160
76.842

48.150
43.543

35.037
33.705
33.529

26.784
26.285



```

Current Data Parameters
USER          AaronJ
NAME          ACQ_2_231
EXPNO        2
PROCNO       1

F2 - Acquisition Parameters
Date_        20130108
Time         12.52
INSTRUM      dxs400
PROBHD       5 mm QNP H/F/P
PULPROG      zgpg30
TD           65536
SOLVNT       CDCl3
NS           613
DS           4
SWH          24154.590 Hz
FIDRES       0.168570 Hz
AQ           1.3566452 sec
RG           10321.3
DW           20.700 usec
DE           20.39 usec
TE           298.0 K
D1           0.10000000 sec
d11          0.01000000 sec
MCHRG1      0.00000000 sec
MCHRG2      0.01500000 sec

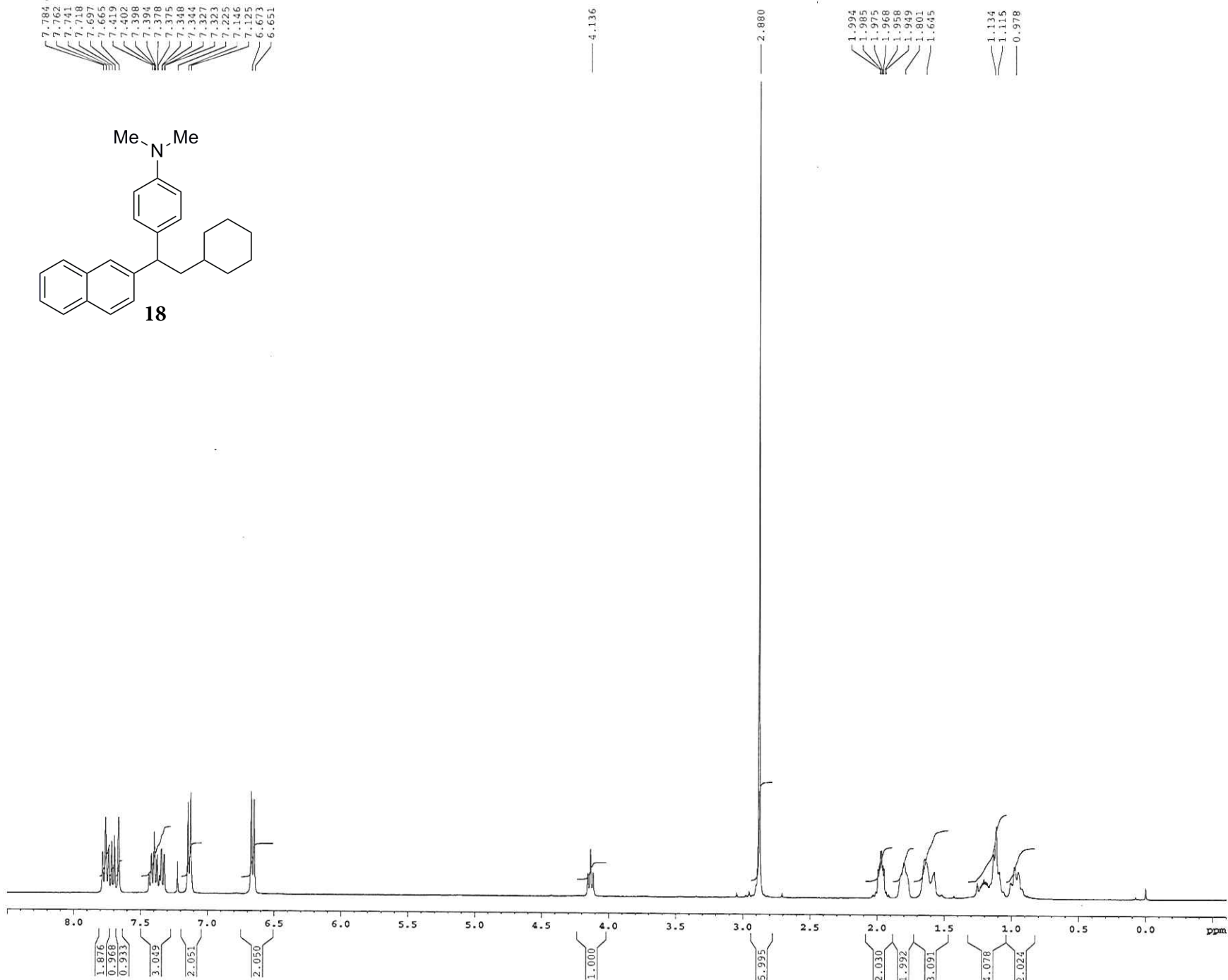
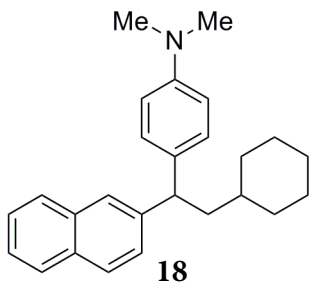
===== CHANNEL f1 =====
NUC1         13C
P1           11.00 usec
PL1          0.00 dB
SFO1         100.6237964 MHz

===== CHANNEL f2 =====
CPDPRG2      mlev16
NUC2         1H
PCPD2        80.00 usec
PL2          0.00 dB
PL12         16.20 dB
SFO2         400.1326009 MHz

F2 - Processing parameters
SI           65536
SF           100.6127610 MHz
WDW          EM
SSB          0
LB           1.00 Hz
GB           0
PC           1.00
    
```

1H spectrum

7.784
7.762
7.741
7.719
7.697
7.665
7.419
7.402
7.398
7.378
7.376
7.374
7.348
7.346
7.327
7.325
7.262
7.242
7.196
7.152
6.613
6.611



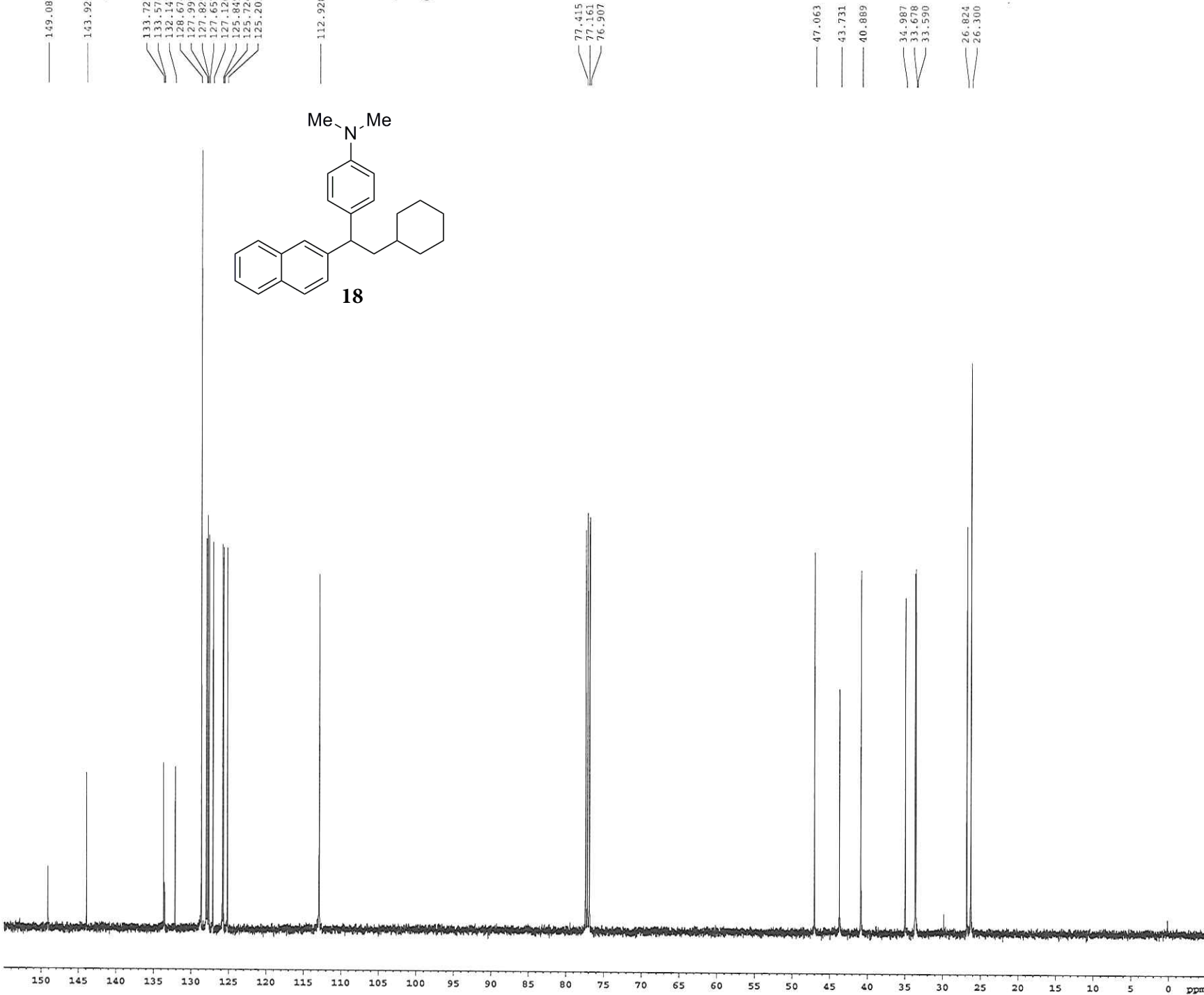
Current Data Parameters
 USER aaronj
 NAME AQJ_2_251_c1
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20130122
 Time 21.02
 INSTRUM dx400
 PROBHD 5 mm QNP H/P/P
 PULPROG zg30
 TD 65536
 SOLVENT CDCl3
 NS 8
 DS 2
 SWH 6410.256 Hz
 FIDRES 0.0978113 Hz
 AQ 5.1118579 sec
 RG 114
 DW 78.000 usec
 DE 4.50 usec
 TE 298.0 K
 D1 0.10000000 sec
 MCHPRT 0.00000000 sec
 MCHW 0.01500000 sec

===== CHANNEL f1 =====
 NUCL 1H
 P1 12.00 usec
 PL1 -0.60 dB
 SFO1 400.1328009 MHz

F2 - Processing parameters
 SI 65536
 SF 400.1300353 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 2.00

Z-restored spin-echo 13C spectrum with 1H decoupling



```

Current Data Parameters
USER      aaronj
NAME      AGJ_2_251_ci
EXPNO     1
PROCNO    1

F2 - Acquisition Parameters
Date_     20130130
Time      13.44
INSTRUM   cryo500
PROBHD    5 mm CPTCT 1H-
PULPROG   SpinEchoq10pp.prd
TD         65535
SOLVENT   CDCl3
NS         200
DS         16
SWH        30303.034 Hz
FIDRES     0.442388 Hz
AQ         1.0813940 sec
RG         7298.2
KW         16.500 usec
DE         6.00 usec
TE         298.0 K
D1         0.25000000 sec
d11        0.03000000 sec
d16        0.00000000 sec
d17        0.00019600 sec
MCHRGST   0.00000000 sec
MCWRK     0.01500000 sec
F2         31.00 usec

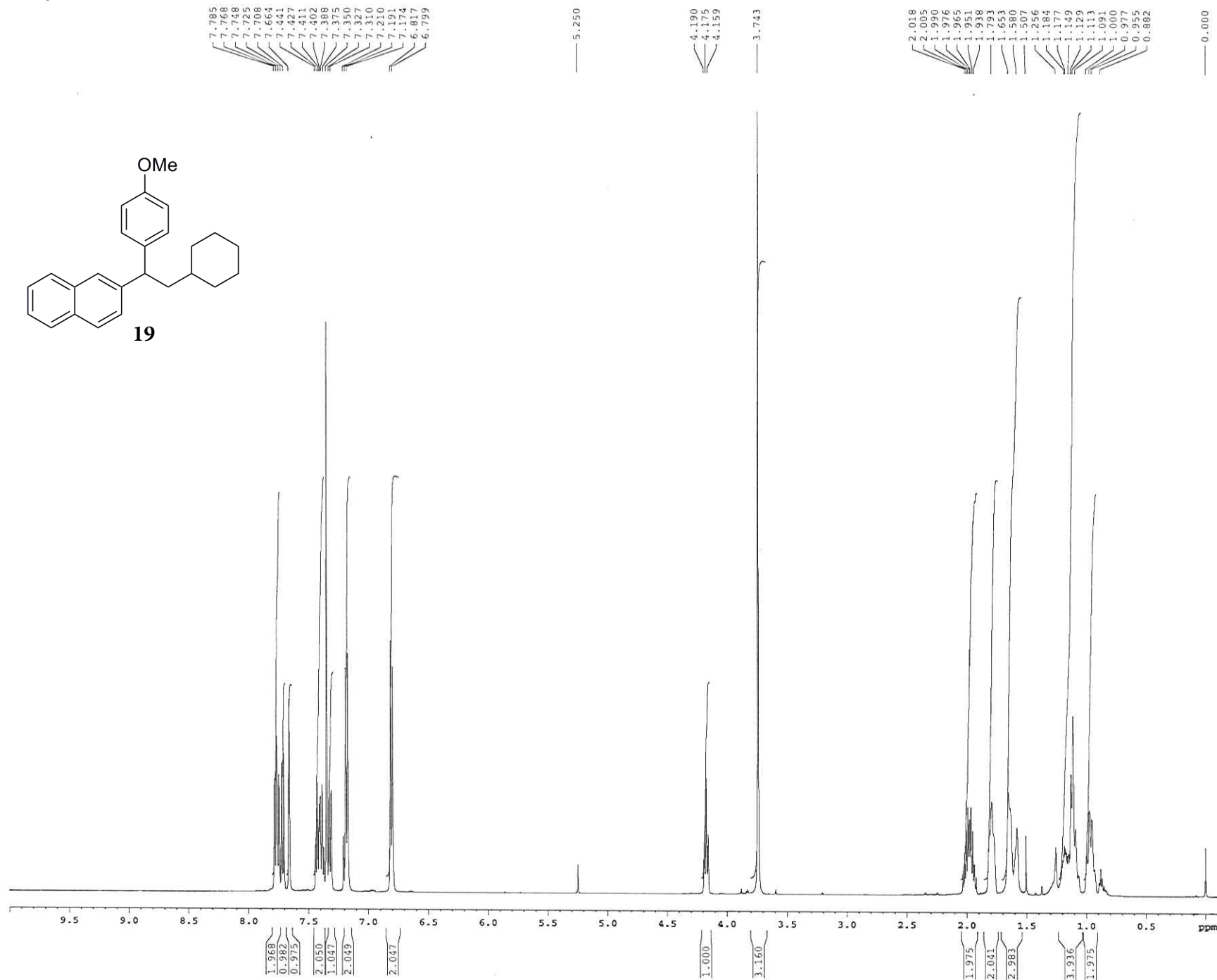
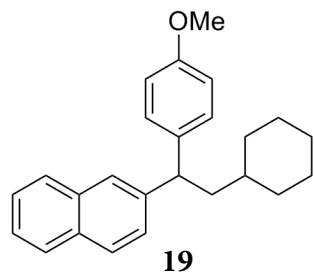
===== CHANNEL f1 =====
NUC1       13C
P1         15.50 usec
P11        500.00 usec
P12        2000.00 usec
PL0        120.00 dB
PL1        -1.00 dB
SFO1      125.7942548 MHz
SP1        3.20 dB
SP2        3.20 dB
SFO2      Crp60,0.5,20.1
SPNAM1    Crp60comp.4
SPOPF1    0.00 Hz
SPOPF2    0.00 Hz

===== CHANNEL f2 =====
CPDPRG2   wait16
NUC2       1H
PCPD2     100.00 usec
PL2       1.60 dB
PL12      24.60 dB
SFO2      500.225011 MHz

===== GRADIENT CHANNEL =====
GPNAM1    SINE.100
GPNAM2    SINE.100
GPX1      0.00 %
GPX2      0.00 %
GPV1      0.00 %
GPV2      0.00 %
GPE1      30.00 %
GPE2      50.00 %
p15       500.00 usec
p16       1000.00 usec

F2 - Processing parameters
SI         65536
SF         125.7804122 MHz
WDW        EM
SSB        0
LB         1.00 Hz
GB         0
PC         2.00
    
```

1H spectrum



7.785
7.768
7.748
7.725
7.708
7.664
7.441
7.427
7.411
7.402
7.388
7.375
7.350
7.327
7.310
7.210
7.191
7.174
6.817
6.799

5.250

4.190
4.175
4.159

3.743

2.018
2.005
1.990
1.976
1.965
1.951
1.938
1.793
1.653
1.580
1.507
1.256
1.184
1.177
1.149
1.129
1.113
1.091
1.000
0.977
0.955
0.882

0.000

```

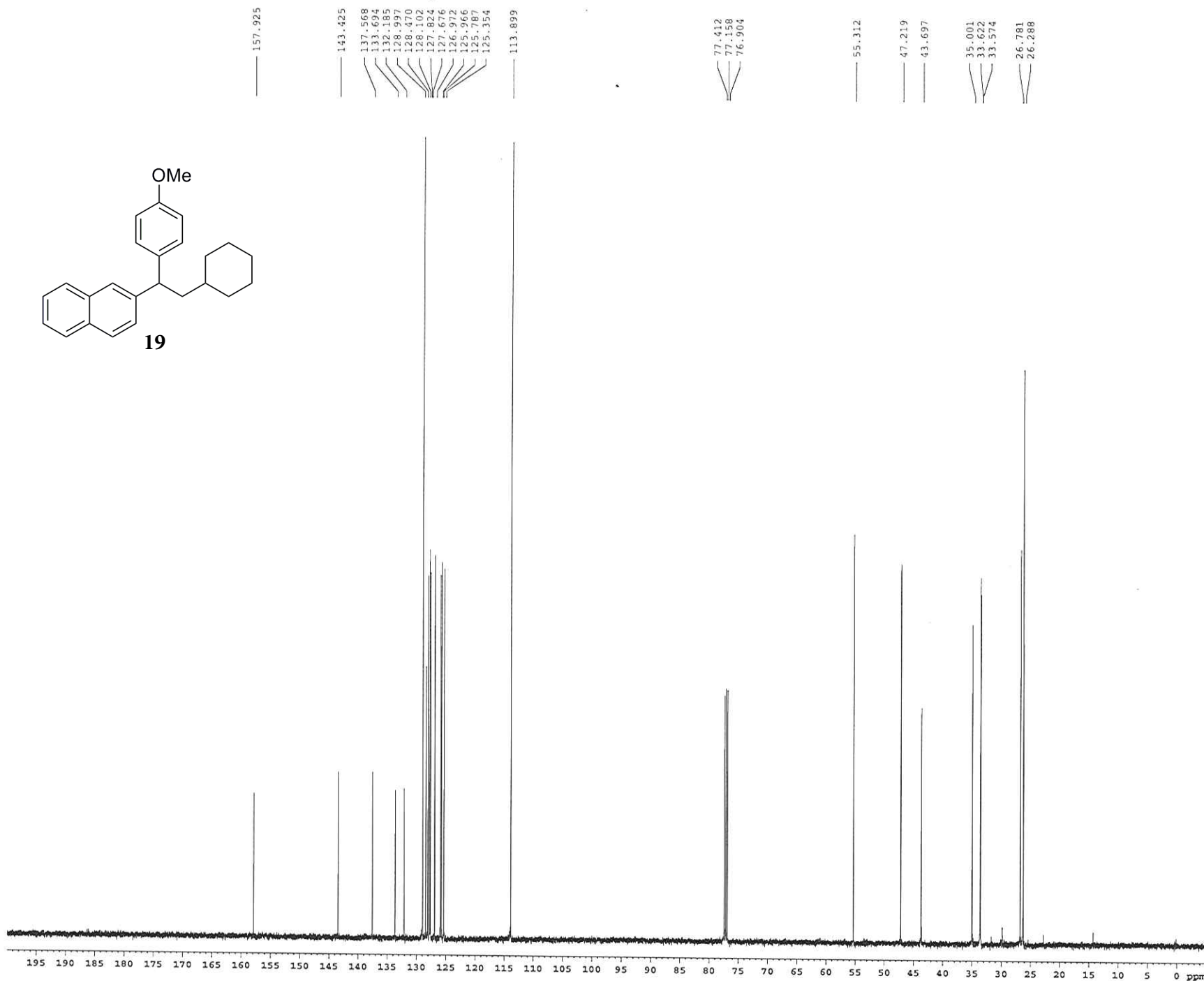
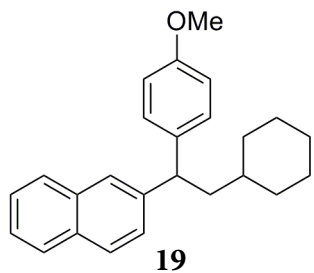
Current Data Parameters
USER          yonova
NAME         imy5 - ag32-272
EXPNO        111
PROCNO       1

F2 - Acquisition Parameters
Date_        20130624
Time         19.13
INSTRUM     cryo500
PROBHD      5 mm CPTCI 1H-
PULPROG     zg30
TD           81728
SOLVENT     CDCl3
NS           16
DS           2
SWH          8012.820 Hz
FIDRES      0.098041 Hz
AQ           5.0998774 sec
RG           5
DW           62.400 usec
DE           6.00 usec
TE           294.0 K
D1           0.10000000 sec
MCHREST     0.00000000 sec
MCWRK       0.01500000 sec

===== CHANNEL f1 =====
NUC1         1H
P1           7.50 usec
PL1          1.60 dB
SFO1         500.2235015 MHz

F2 - Processing parameters
SI           65536
SF           500.220946 MHz
WDW          EM
SSB          0
LB           0.30 Hz
GB           0
PC           2.00
    
```

Z-restored spin-echo 13C spectrum with 1H decoupling



Current Data Parameters
 USER yonova
 NAME imy5 - agj2-272
 EXPRNO 222
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20130624
 Time_ 19.20
 INSTRUM cryo500
 PROBNM 5 mm CPTCI 1H-
 PULPROG SpinEcho30pp.prd
 TD 65536
 F2 125.7604140
 SOLVENT CDCl3
 NS 207
 DS 16
 SFR 30303.031 Hz
 FIDRES 0.462388 Hz
 AQ 1.0813940 sec
 RG 7298.2
 DW 16.500 usec
 DE 6.00 usec
 TE 298.0 K
 D1 0.25000000 sec
 d11 0.03000000 sec
 d16 0.00020000 sec
 d17 0.00019600 sec
 MCRRET 0.00000000 sec
 MCRMK 0.01500000 sec
 P2 31.00 usec

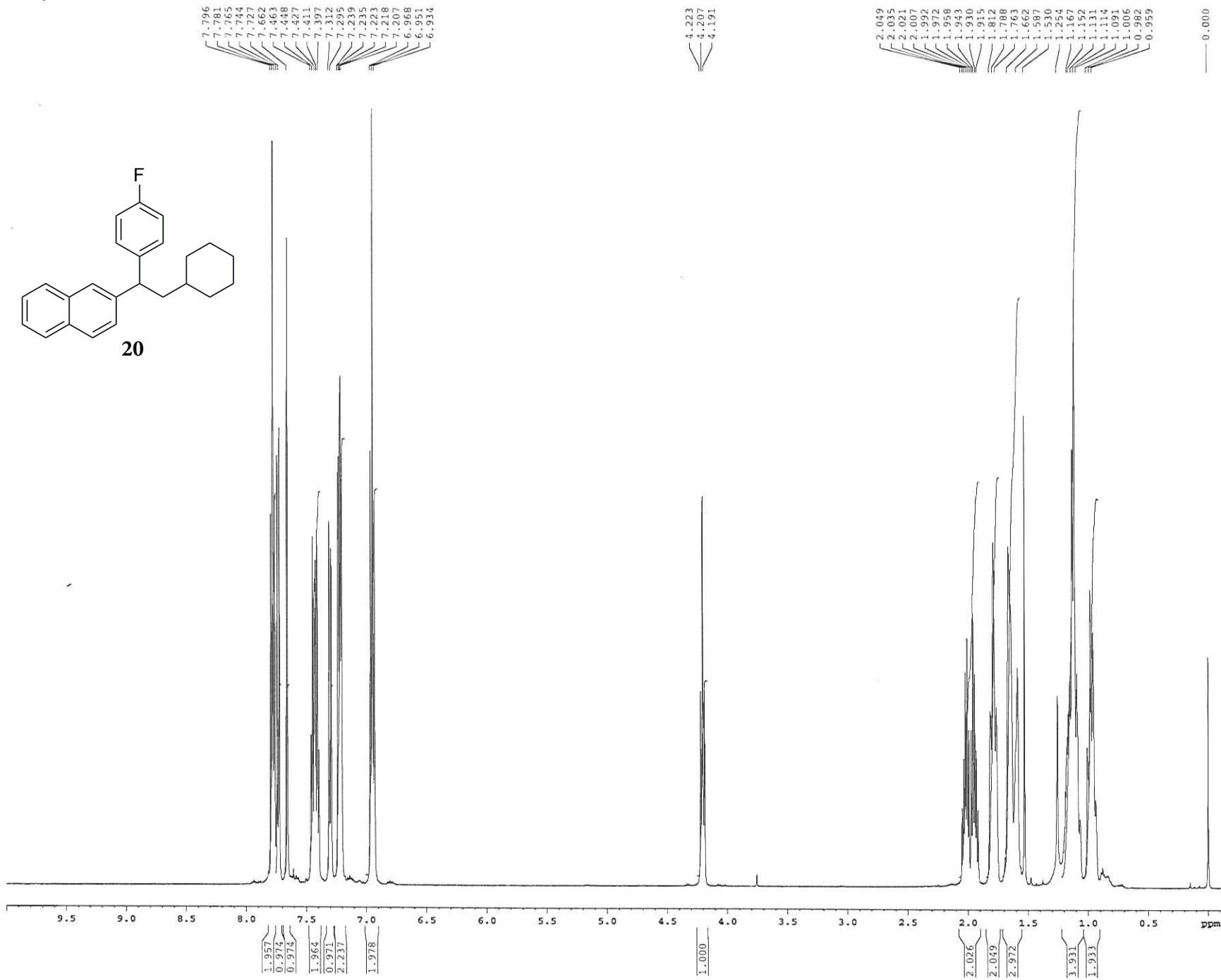
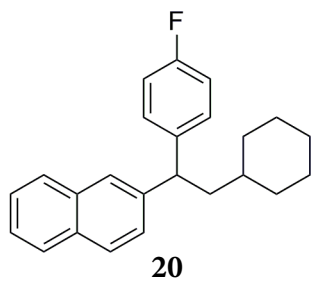
==== CHANNEL f1 =====
 NUC1 13C
 P1 15.50 usec
 P11 500.00 usec
 P12 2000.00 usec
 PL0 120.00 dB
 PL1 -1.00 dB
 SFO1 125.7942548 MHz
 SF1 3.20 dB
 SP2 1.20 GB
 SFRAM1 Crp60.0.5.20.1
 SFRAM2 Crp60comp.4
 SPOFF1 0.00 Hz
 SPOFF2 0.00 Hz

==== CHANNEL f2 =====
 CPDPRG2 waltz16
 NUC2 1H
 P1P2 100.00 usec
 PL2 1.60 dB
 PL12 24.60 dB
 SFO2 500.2225011 MHz

==== GRADIENT CHANNEL =====
 GPNAM1 SINE.100
 GPNAM2 SINE.100
 GPX1 0.00 %
 GPX2 0.00 %
 GPY1 0.00 %
 GPY2 0.00 %
 GPZ1 30.00 %
 GPZ2 50.00 %
 p15 500.00 usec
 p16 1000.00 usec

F2 - Processing parameters
 SI 65536
 SF 125.7604140 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 4.00

1H spectrum



```

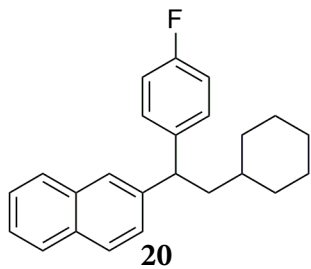
Current Data Parameters
USER      yonova
NAME      imy5 - agj2-201
EXPNO     111
PROCNO    1

F2 - Acquisition Parameters
Date_     20130621
Time      21.01
INSTRUM   cryo500
PROBHD    5 mm CPTCI 1H-
PULPROG   zg30
TD         81728
SOLVENT   CDCl3
NS         8
DS         2
SWH        8012.820 Hz
FIDRES    0.098043 Hz
AQ         5.0999398 sec
RG         5
DS         62.400 usec
DE         6.00 usec
TE         298.0 K
D1         0.10000000 sec
MCHRGST   0.00000000 sec
MCWRR     0.01500000 sec

===== CHANNEL f1 =====
NUC1       1H
P1         7.50 usec
PL1        1.60 dB
SFO1       500.2235015 MHz

F2 - Processing parameters
SI         65536
SF         500.2200399 MHz
WDW        EM
SSB        0
LB         0.30 Hz
GB         0
PC         2.00
    
```


Z-restored spin-echo 13C spectrum with 1H decoupling



162.346
160.406

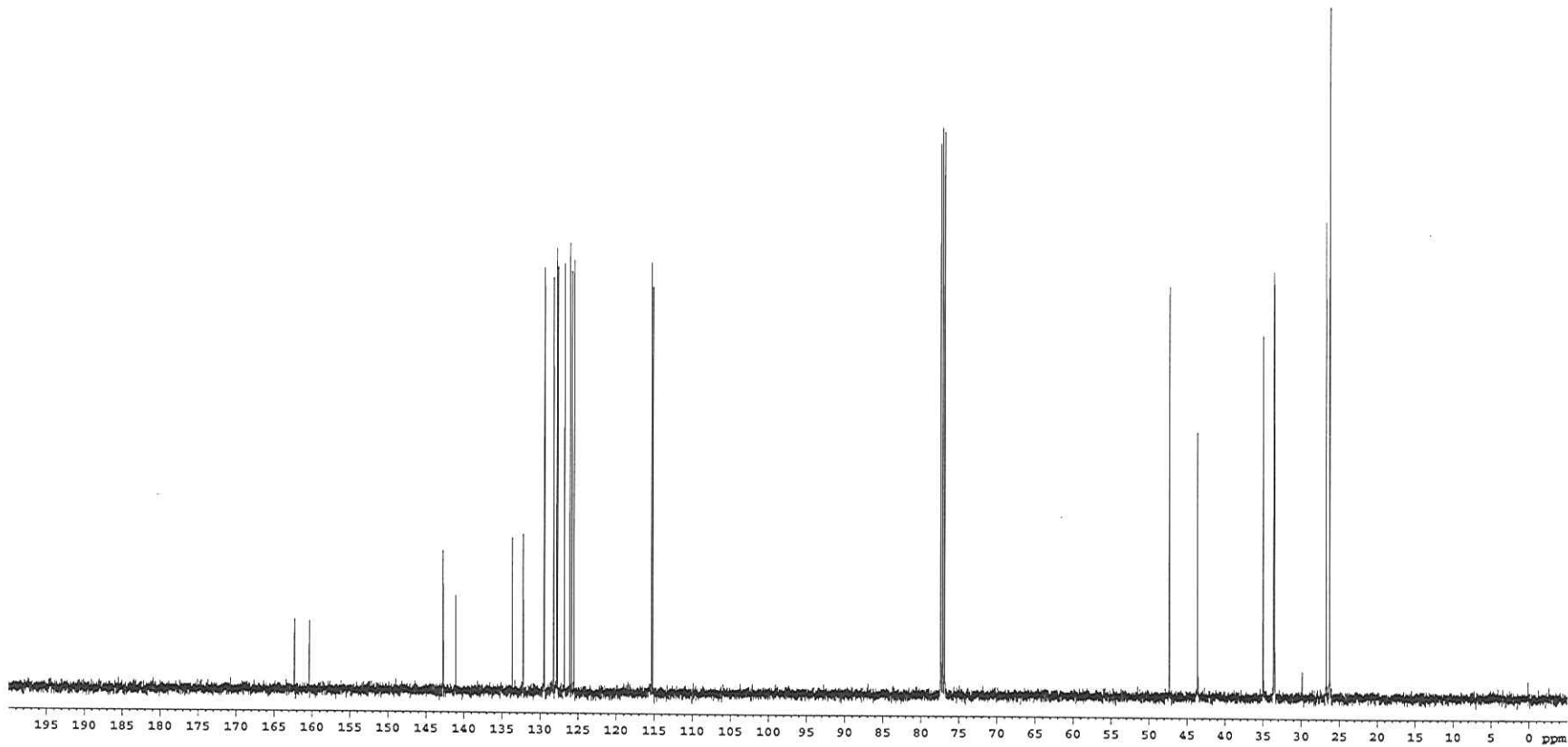
142.781
141.132
141.106
133.669
132.248
129.484
129.422
128.245
127.836
127.713
126.823
126.111
125.893
125.539
115.370
115.202

77.413
77.159
76.905

47.334
43.638

34.998
33.612
33.530

26.741
26.272



Current Data Parameters
 USHR yonova
 NAME imy5 - agj2-281
 EXPMO 222
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20130621
 Time 21.07
 INSTRUM cryo500
 PROBRD 5 mm CPTCI 1H-
 PULPROG Spinecho30op.prd
 DS 65536
 SOLVENT CDCl3
 NS 239
 DS 16
 SWH 30303.031 Hz
 FIDRES 0.462388 Hz
 AQ 1.0814105 sec
 RG 7298.2
 RM 16.500 usec
 RE 6.00 usec
 TE 298.0 K
 D1 0.25000000 sec
 d11 0.03000000 sec
 d16 0.00000000 sec
 d17 0.00019600 sec
 MCHSET 0.00000000 sec
 MCWRR 0.01000000 sec
 PC 31.00 usec

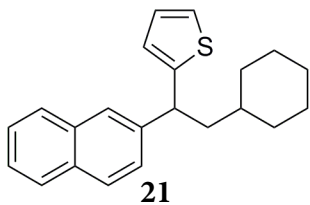
==== CHANNEL f1 =====
 NUC1 13C
 P1 15.50 usec
 P11 500.00 usec
 P12 2000.00 usec
 PL0 120.00 dB
 PL1 -1.00 dB
 SFO1 125.7842548 MHz
 SF1 3.20 dB
 SF2 3.20 dB
 SFOAM1 Crp60.0.5.20.1
 SFOAM2 Crp60comp.4
 SFOFF1 0.00 Hz
 SFOFF2 0.00 Hz

==== CHANNEL f2 =====
 CPDPRG2 waitz16
 NUC2 1H
 P12 100.00 usec
 PL2 1.60 dB
 PL12 24.60 dB
 SFO2 500.2225011 MHz

==== GRADIENT CHANNEL =====
 GPNAM1 SINE.100
 GPNAM2 SINE.100
 GPX1 0.00 %
 GPX2 0.00 %
 GPY1 0.00 %
 GPY2 0.00 %
 GPZ1 30.00 %
 GPZ2 50.00 %
 p15 500.00 usec
 p16 1000.00 usec

F2 - Processing parameters
 SI 65536
 SF 125.7804094 MHz
 MDW 8K
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 2.00

1H spectrum



7.807
7.785
7.762
7.698
7.473
7.459
7.441
7.424
7.403
7.381
7.251
7.135
7.112
6.917
6.908
6.895
6.841
6.833

4.470
4.450
4.430

2.067
2.049
2.030
1.870
1.939
1.743
1.742
1.687
1.287
1.255
1.235
1.223
1.224
1.215
1.206
1.189
1.179
1.171
1.162
1.152
1.132
1.085
1.088
1.076
1.076
0.986
0.939
0.000

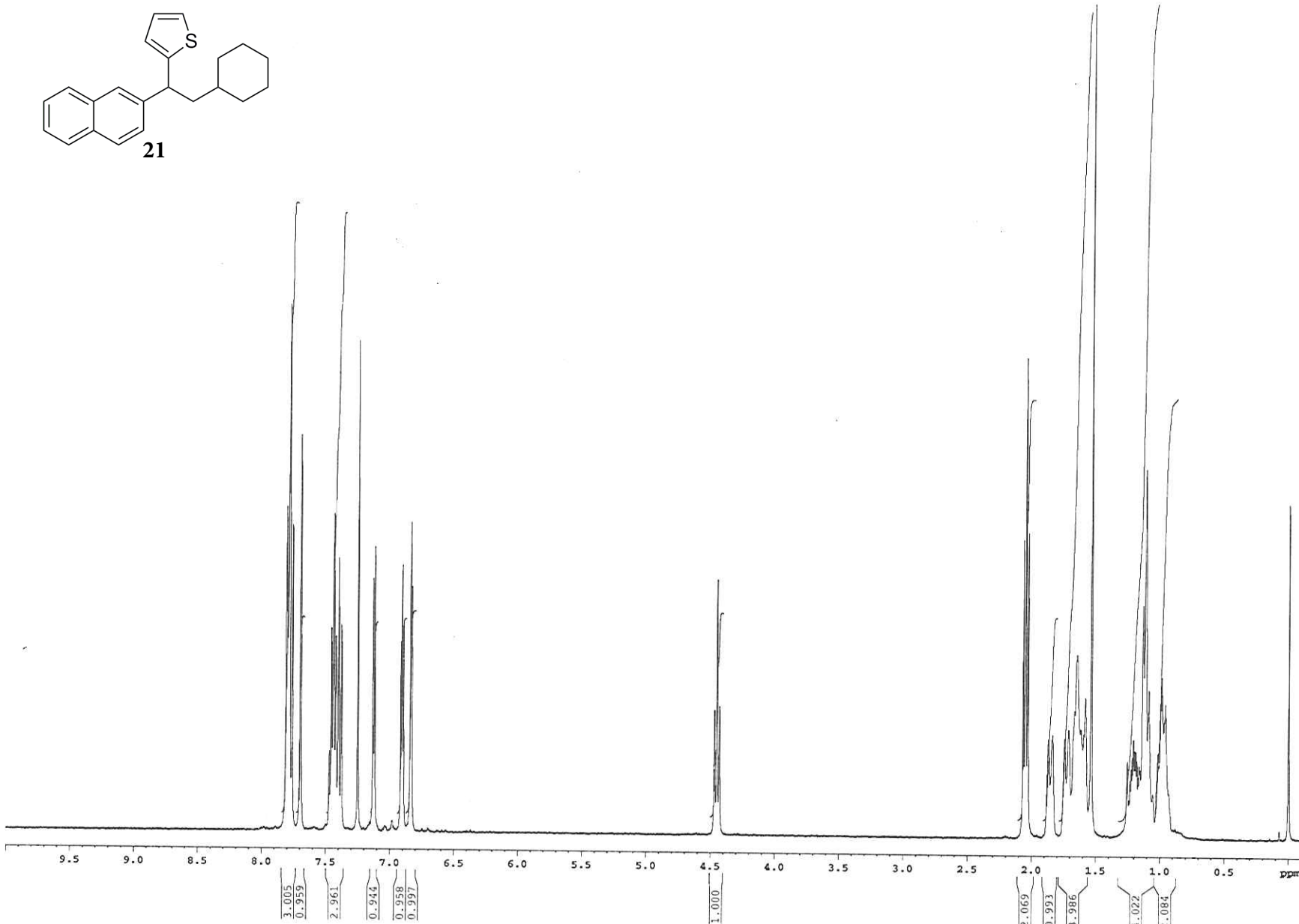
```

Current Data Parameters
USER          yonova
NAME          lmy5 - 243
EXTRNO       17
PROCNO        1

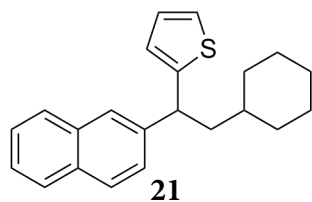
F2 - Acquisition Parameters
Date_         20130509
Time          13.57
INSTRUM       dx400
PROBHD        5 mm QNP H/P/P
PULPROG       zg30
TD            65536
SOLVENT       CDCl3
NS            8
DS            2
SWH           6410.256 Hz
FIDRES        0.097913 Hz
AQ            5.1118579 sec
RG            287.4
EW            78.000 usec
DE            4.50 usec
TE            298.1 K
D1            0.10000000 sec
MCHECT        0.00000000 sec
MCHRR        0.01500000 sec

===== CHANNEL f1 =====
NUC1          1H
P1            12.00 usec
PL1           -0.40 dB
SFO1         400.1328009 MHz

F2 - Processing parameters
SI            65536
SF            400.1300249 MHz
WDW           EM
SSB           0
GB            0.30 Hz
PC            2.00
    
```



Z-restored spin-echo 13C spectrum with 1H decoupling

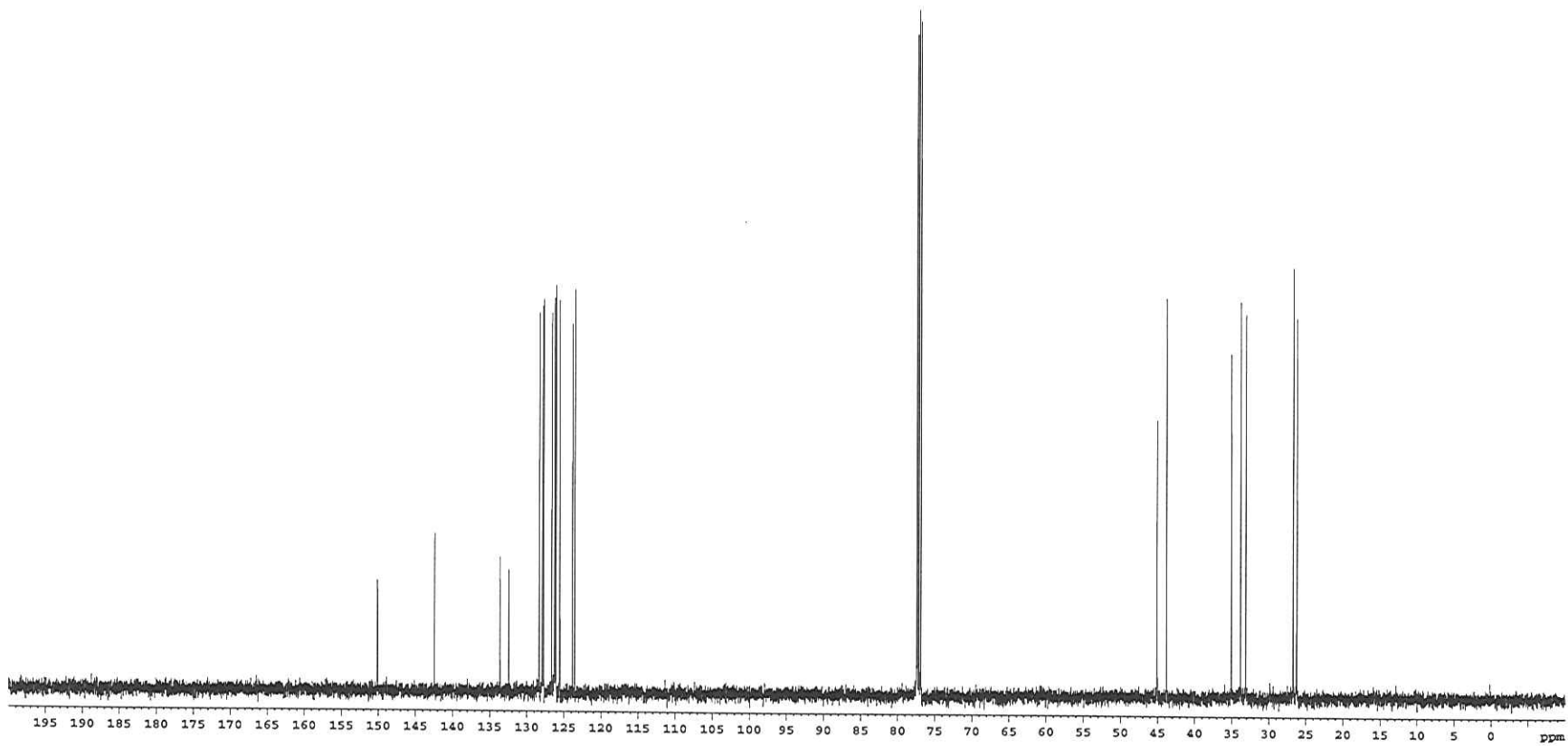


150.154
142.455
133.667
132.466
128.356
127.911
127.754
126.672
126.297
126.149
126.090
125.599
123.875
123.557

77.415
77.163
76.907

45.090
43.821

35.050
33.803
33.089
26.718
26.245
26.214



Current Data Parameters
USER yonova
NAME imys - 243
EXPNO 222
PROCNO 1

F2 - Acquisition Parameters
Date_ 20110510
Time 14.26
INSTRUM crys000
PROBHD 5 mm CP13 1H-
PULPROG SpinEcho30op.prd
TD 65536
SOLVENT CDCl3
NS 325
DS 16
SWH 30303.031 Hz
FIDRES 0.462198 Hz
AQ 1.0814105 sec
RG 5792.6
DW 16.500 usec
DE 6.00 usec
TE 298.0 K
D1 0.25000000 sec
d11 0.03000000 sec
D16 0.00000000 sec
d17 0.00019600 sec
MCHRGPT 0.00000000 sec
MCWRR 0.01500000 sec
P2 31.00 usec

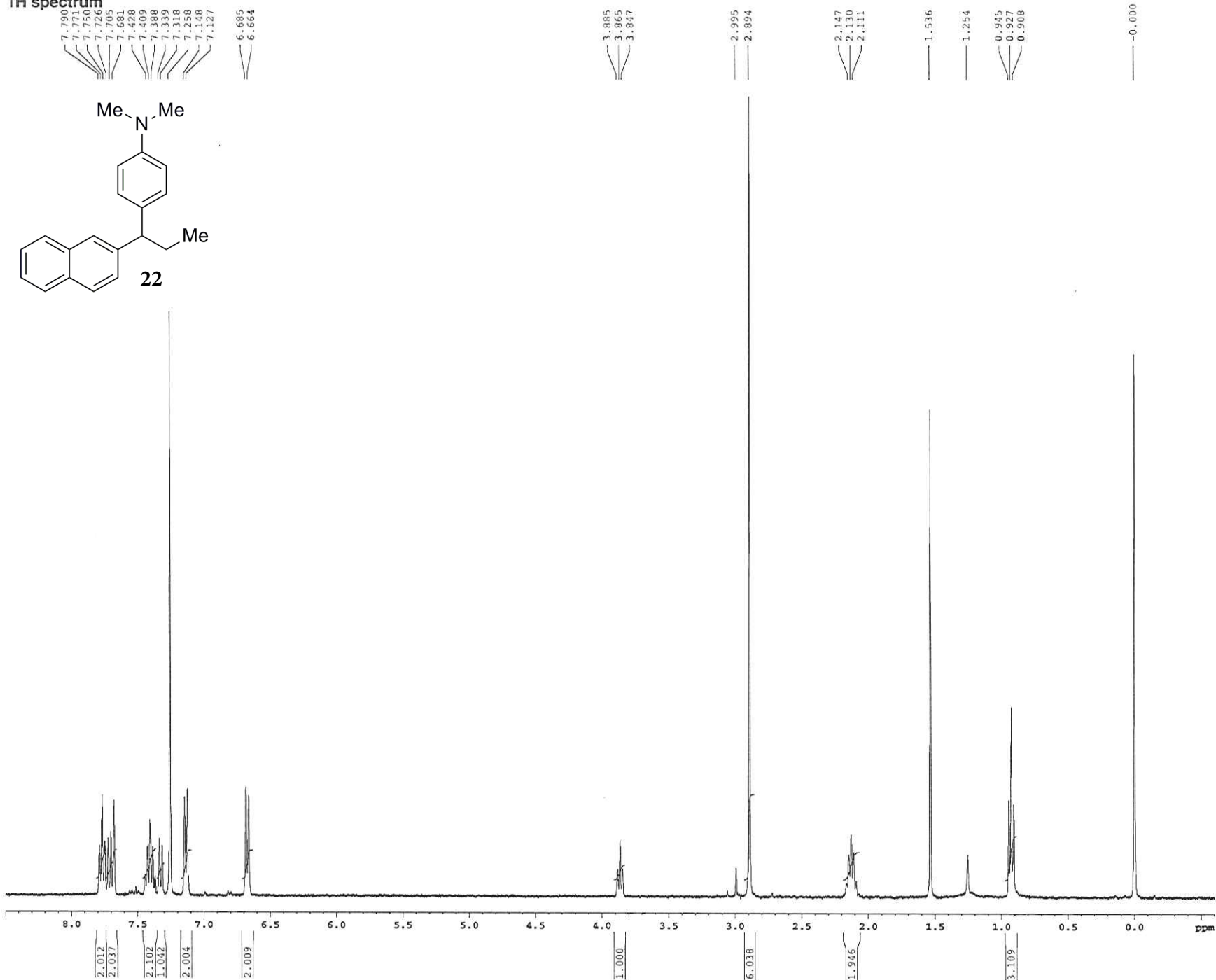
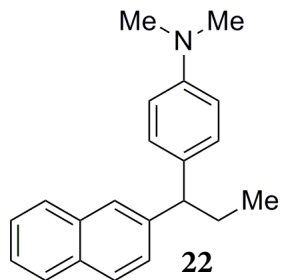
===== CHANNEL f1 =====
NUC1 13C
P1 15.50 usec
P11 500.00 usec
P12 2000.00 usec
PL0 120.00 dB
PL1 -1.00 dB
SFO1 125.7942548 MHz
SF1 3.20 dB
SF2 3.20 dB
SFXAM1 Crp60.0.5.20.1
SFXAM2 Crp60comp.4
SPOFF1 0.00 Hz
SPOFF2 0.00 Hz

===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCP2 100.00 usec
PL2 1.60 dB
PL12 24.60 dB
SFO2 500.2225011 MHz

===== GRADIENT CHANNEL =====
GPNAM1 SINE.100
GPNAM2 SINE.100
GPX1 0.00 %
GPX2 0.00 %
GPY1 0.00 %
GPY2 0.00 %
GPZ1 30.00 %
GPZ2 50.00 %
p15 500.00 usec
p16 1000.00 usec

F2 - Processing parameters
SI 65536
SF 125.7804094 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 2.00

1H spectrum



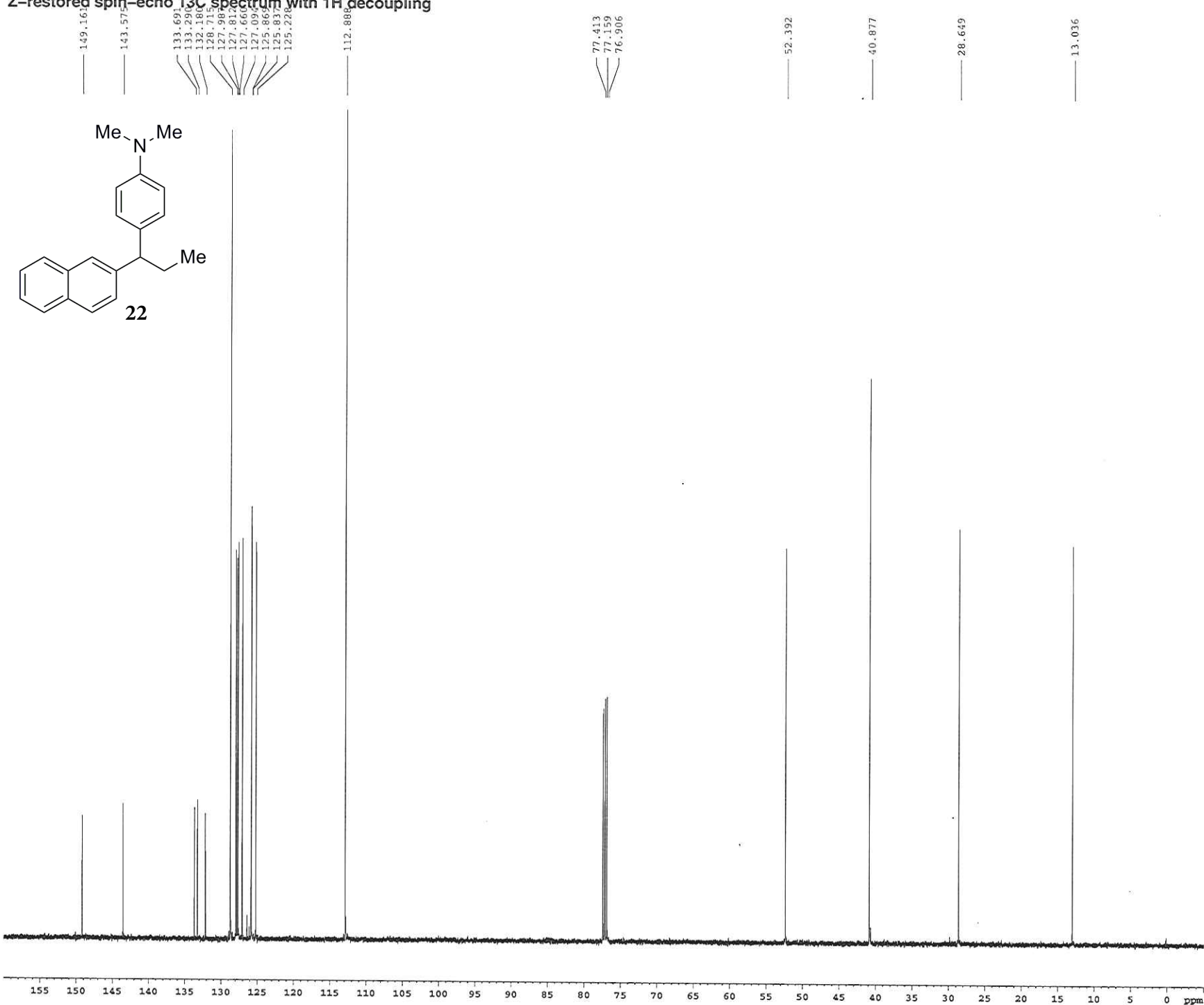
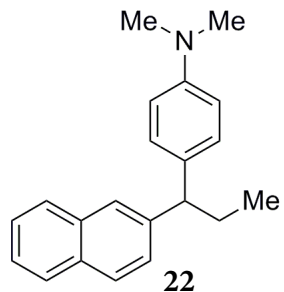
```

Current Data Parameters
=====
Date_      20110625
Time       17.41
INSTRUM    dx400
PROBHD     5 mm QNP H/P/P
PULPROG    zg30
TD          65536
SOLVENT    CDCl3
NS          8
DS          2
SWH         6410.256 Hz
FIDRES     0.097813 Hz
AQ          5.118579 sec
RG          724.1
DM          78.000 usec
DE          4.50 usec
TE          297.9 K
D1          0.10000000 sec
d11         0.00000000 sec
MCRES2     0.00000000 sec
MCWVK      0.01500000 sec

===== CHANNEL f1 =====
NUC1       1H
P1         12.00 usec
PL1        -0.60 dB
SFO1       400.1328009 MHz

F2 - Processing parameters
SI         65536
SP         400.1300221 MHz
WDW        EM
SSB        0
LB         0.30 Hz
GB         0
PC         2.00
    
```

Z-restored spin-echo ¹³C spectrum with ¹H decoupling



```

Current Data Parameters
USER          aaronj
NAME          AGJ_3_11
EXPNO        5
PROCNO       1

F2 - Acquisition Parameters
Date_        20130625
Time         19.40
INSTRUM      cryo500
PROBHD       5 mm CPTCI 1H-
PULPROG      SpinEchopp30pp.prd
TD           65536
SOLVENT      CDCl3
NS           440
DS           16
SWH          30303.031 Hz
FIDRES       0.462188 Hz
AQ           1.0813940 sec
RG           7298.2
DW           16.500 usec
DE           6.00 usec
TE           298.0 K
D1           0.25000000 sec
d11          0.03000000 sec
D16          0.00200000 sec
d17          0.00019600 sec
MCREST       0.00000000 sec
MCWRK        0.01500000 sec
P2           31.00 usec

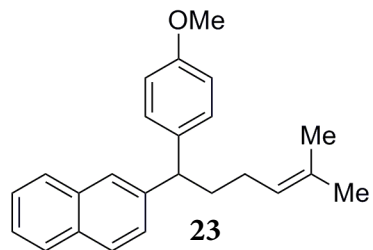
===== CHANNEL f1 =====
NUC1         13C
P1           15.50 usec
P11          500.00 usec
P12          2000.00 usec
PL0          120.00 dB
PL1          -1.00 dB
SFO1         125.7942548 MHz
SP1          3.20 dB
SP2          3.20 dB
SFRAM1       Crp60.0.5.20.1
SFRAM2       Crp60comp.4
SPOPF1       0.00 Hz
SPOPF2       0.00 Hz

===== CHANNEL f2 =====
CPDPRG2      waltz16
NUC2         1H
PCPD2        100.00 usec
PL2          1.60 dB
PL12         24.60 dB
SFO2         500.2225011 MHz

===== GRADIENT CHANNEL =====
GPNAM1       SINM.100
GPNAM2       SINM.100
GPX1         0.00 %
GPX2         0.00 %
GPY1         0.00 %
GPY2         0.00 %
CFZ1         30.00 %
CFZ2         50.00 %
p15          500.00 usec
p16          1000.00 usec

F2 - Processing parameters
SI           65536
SF           125.7804127 MHz
WDW          EM
SSB          0
LB           1.00 Hz
GB           0
PC           2.00
    
```

1H spectrum



7.784
7.768
7.751
7.728
7.710
7.678
7.445
7.431
7.415
7.408
7.394
7.379
7.320
7.303
7.228
7.194
7.176
6.822
6.804

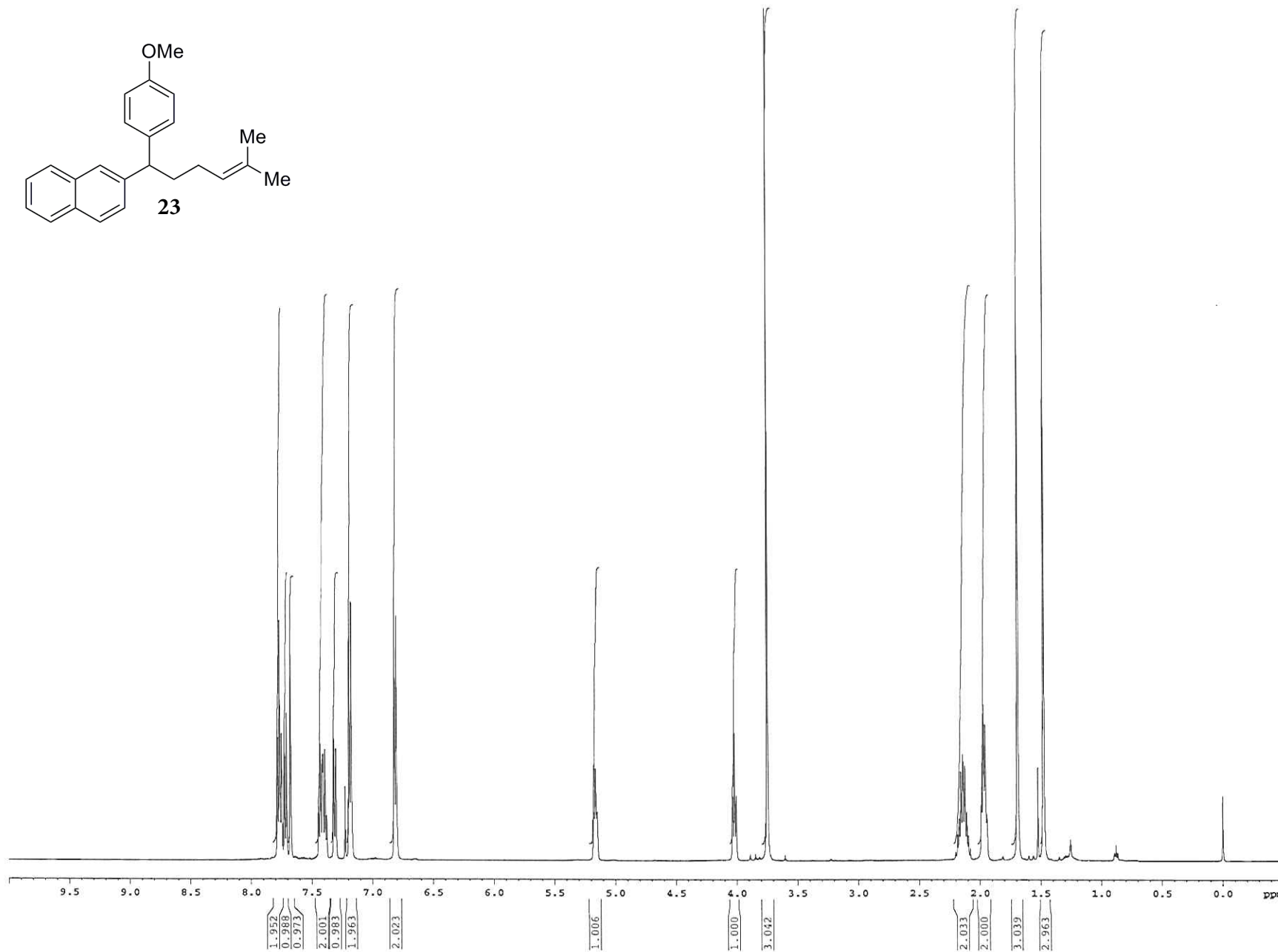
5.179
5.165
5.151

4.037
4.022
4.006

3.751

2.171
2.159
2.143
2.124
2.109
1.987
1.972
1.958
1.943
1.690
1.524
1.476

0.000



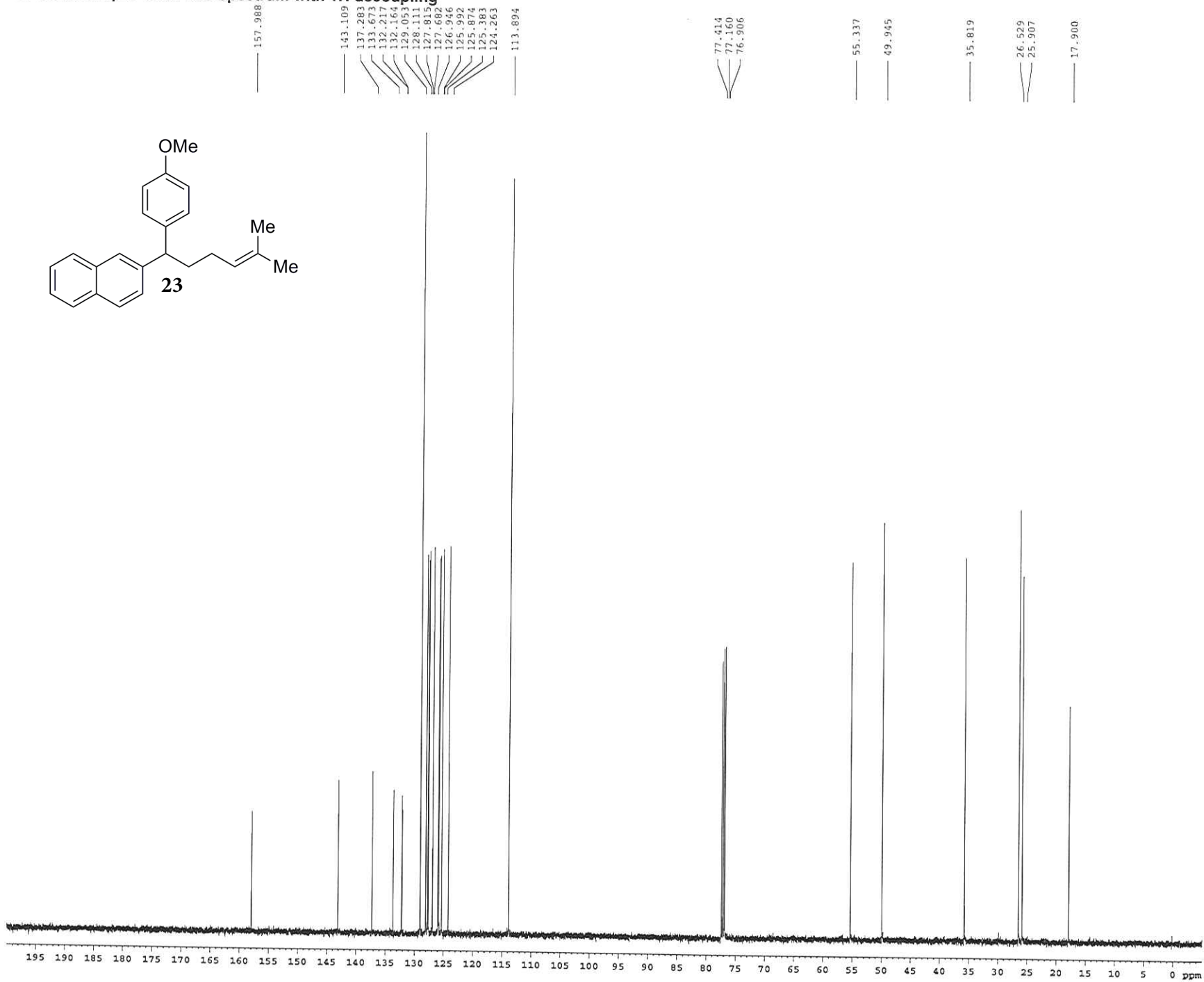
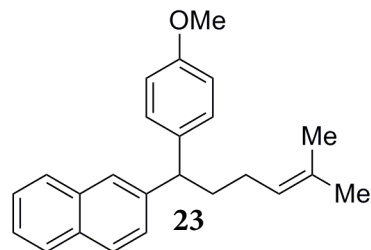
Current Data Parameters
 USER yonova
 NAME imy5 - ag31-49
 EXTRN 111
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20130211
 Time 21.12
 INSTRUM cryo500
 PROBRD 5 mm CPYX 1H-
 PULPROG zg30
 TD 81728
 SOLVENT CDCl3
 NS 8
 DS 2
 SWH 8012.820 Hz
 FIDRES 0.098041 Hz
 AQ 5.0999398 sec
 RG 6.3
 DW 62.400 usec
 DE 6.00 usec
 TE 298.0 K
 D1 0.10000000 sec
 MCREST 0.00000000 sec
 MCWRR 0.01500000 sec

===== CHANNEL f1 =====
 NUC1 1H
 P1 7.50 usec
 PL1 1.60 dB
 SFO1 500.2235015 MHz

F2 - Processing parameters
 SI 65336
 SF 500.2200457 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 2.00

Z-restored spin-echo 13C spectrum with 1H decoupling



Current Data Parameters
 USER yonova
 NAME imy5 - agj1-49
 EXPNO 222
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20110621
 Time 21.20
 INSTRUM cryo500
 PROBRD 5 mm CPCCI 1H-
 PULPROG spinstochop3dpp,prd
 TD 65536
 SOLVENT CDCl3
 NS 16
 DS 276
 SWH 30303.031 Hz
 FIDRES 0.462388 Hz
 AQ 1.0814105 sec
 RG 5792.6
 DW 16.500 usec
 DE 6.00 usec
 TE 298.0 K
 D1 0.25000000 sec
 d11 0.03000000 sec
 DL6 0.00020000 sec
 dL7 0.00019600 sec
 MCREST 0.00000000 sec
 MCWRR 0.01500000 sec
 P2 31.00 usec

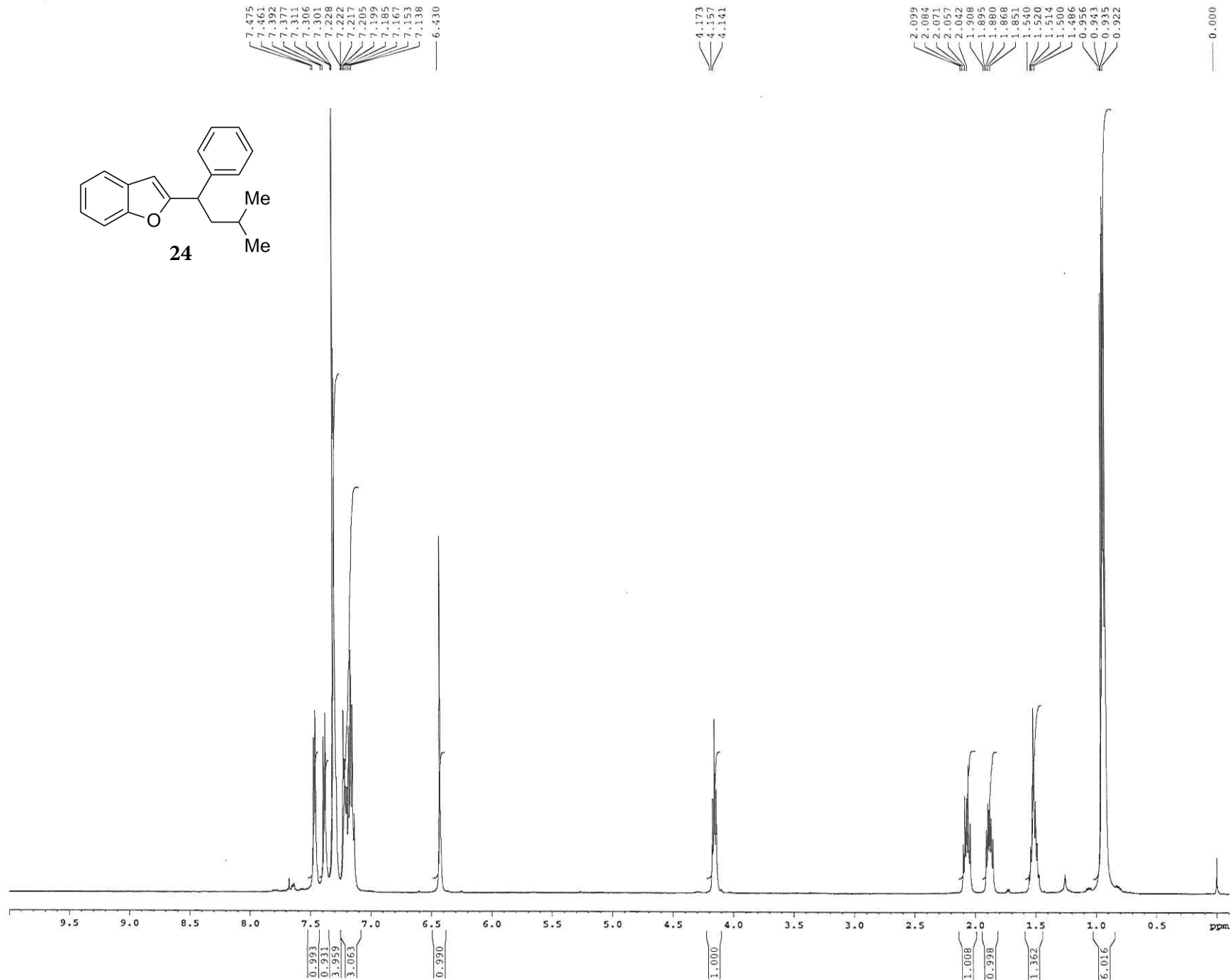
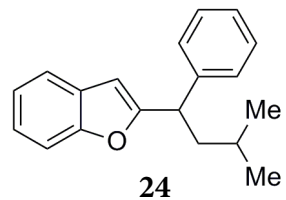
***** CHANNEL f1 *****
 NUC1 13C
 P1 15.50 usec
 PL1 500.00 usec
 PL2 2000.00 usec
 PL0 120.00 dB
 PL1 -1.00 dB
 SFO1 125.7942548 MHz
 SP1 3.20 dB
 SP2 3.20 dB
 SPNAM1 Crp60.0.5.20.1
 SPNAM2 Crp60comp.4
 SPOFF1 0.00 Hz
 SPOFF2 0.00 Hz

***** CHANNEL f2 *****
 CPDPRG2 waltz16
 NUC2 1H
 PCPD2 100.00 usec
 PL2 2.60 dB
 PL12 24.60 dB
 SFO2 500.225011 MHz

***** GRADIENT CHANNEL *****
 GRNAM1 SINE.100
 GRNAM2 SINE.100
 GPX1 0.00 %
 GPX2 0.00 %
 GPY1 0.00 %
 GPY2 0.00 %
 GPZ1 30.00 %
 GPZ2 50.00 %
 p15 500.00 usec
 p16 1000.00 usec

F2 - Processing parameters
 SI 65536
 SF 125.7804113 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 4.00

1H spectrum



```

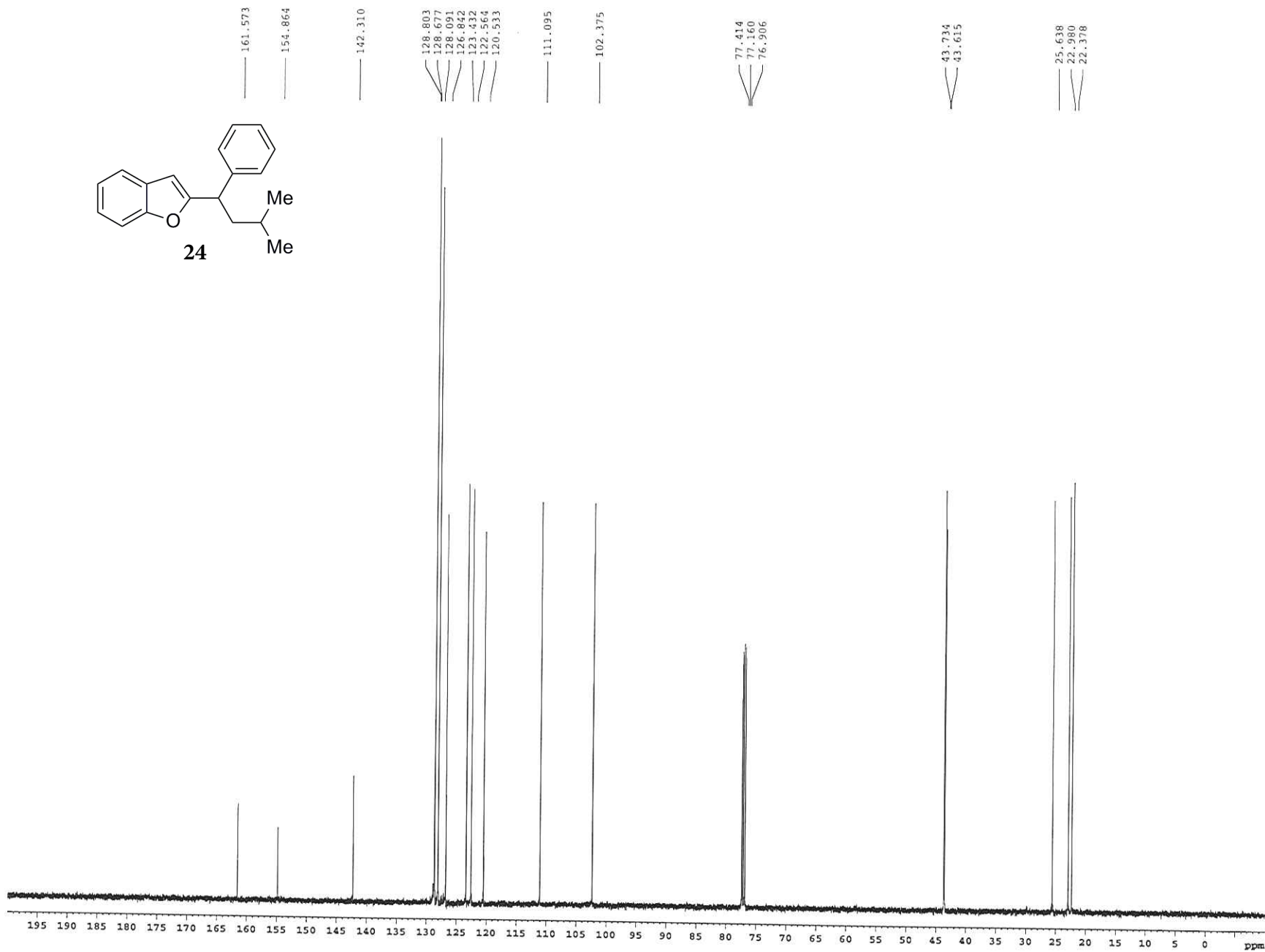
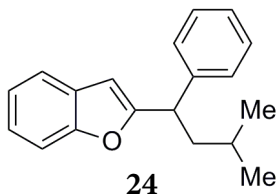
Current Data Parameters
=====
USER      yonova
NAME      imy5 - 240
EXPNO     111
PROCNO    1

F2 - Acquisition Parameters
=====
Date_     20110510
Time      14.16
INSTRUM   cryo500
PROBHD    5 mm CPTCI 1H-
PULPROG   zg30
TD         81728
SOLVENT   CDCl3
NS         8
DS         2
SWH        8012.820 Hz
FIDRES     0.098043 Hz
AQ         5.0999398 sec
RG         4.5
RW         62.400 usec
DE         6.00 usec
TE         298.0 K
D1         0.10000000 sec
MCHRES     0.00000000 sec
MCWRK     0.01500000 sec

===== CHANNEL f1 =====
NUC1      1H
P1        7.50 usec
PL1       1.60 dB
SFO1      500.2235015 MHz

F2 - Processing parameters
=====
SI         65536
SF         500.2200474 MHz
WDW        EM
SSB        0
LB         0.30 Hz
GB         0
PC         2.00
    
```


Z-restored spin-echo 13C spectrum with 1H decoupling



Current Data Parameters
 USER ycnova
 NAME imy5 - 240
 EXPNO 222
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20130510
 Time 14:24
 INSTRUM cryo500
 PROBRD 5 mm CPTCI 1H-
 PULPROG SpinEcho30op.prd
 TD 65536
 SFO1 125.780117
 SOLVENT CDCl3
 NS 274
 DS 16
 SWH 30303.011 Hz
 FIDRES 0.462398 Hz
 AQ 1.0814105 sec
 RG 5792.6
 LW 6.00 usec
 DE 16.500 usec
 TE 298.0 K
 D1 0.25000000 sec
 d11 0.00000000 sec
 D16 0.00020000 sec
 d17 0.00019600 sec
 MCHPST 0.00000000 sec
 MCHWK 0.01500000 sec
 P2 31.00 usec

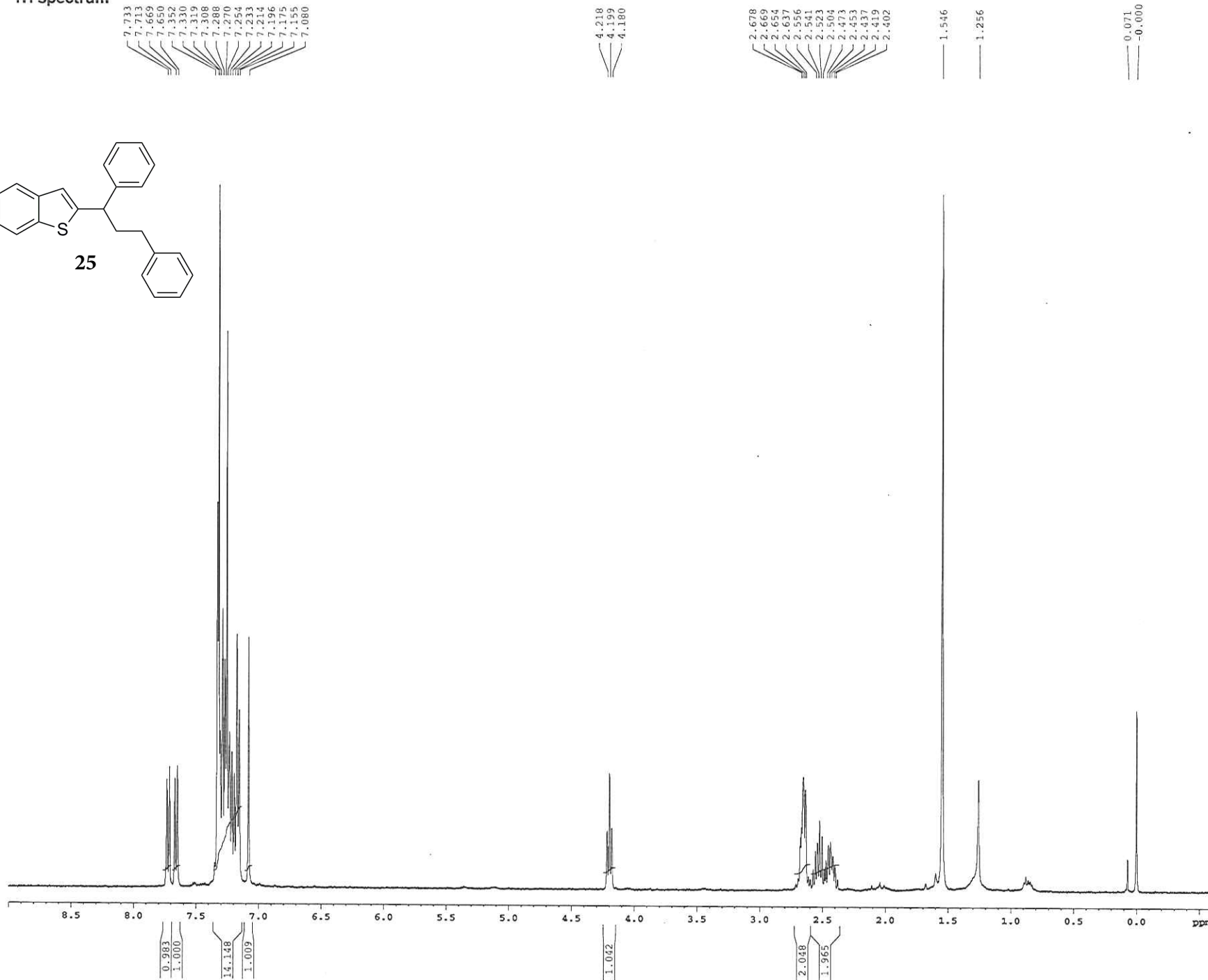
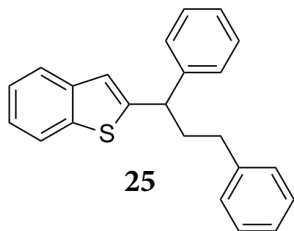
***** CHANNEL f1 *****
 NUC1 13C
 PL 15.50 usec
 P11 500.00 usec
 P12 2000.00 usec
 PL0 120.00 dB
 PL1 -1.00 dB
 SFO1 125.7842548 MHz
 SF1 3.20 dB
 SF2 3.20 dB
 SFO2 Crp60,0.5,20,1
 SFO1 Crp60comp,4
 SPOFF1 0.00 Hz
 SPOFF2 0.00 Hz

***** CHANNEL f2 *****
 CPDPRG2 waltz16
 NUC2 1H
 P12 100.00 usec
 PL2 1.60 dB
 PL1 24.60 dB
 SFO2 500.2225011 MHz

***** GRADIENT CHANNEL *****
 GPNAM1 SINE.100
 GPNAM2 SINE.100
 GPX1 0.00 %
 GPX2 0.00 %
 GPY1 0.00 %
 GPY2 0.00 %
 GPZ1 30.00 %
 GPZ2 50.00 %
 p15 500.00 usec
 p16 1000.00 usec

F2 - Processing parameters
 SI 65536
 SF 125.7804117 MHz
 RM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 4.00

1H spectrum



7.733
7.713
7.669
7.650
7.352
7.330
7.319
7.308
7.288
7.270
7.254
7.233
7.214
7.196
7.175
7.155
7.080

4.218
4.199
4.180

2.678
2.669
2.654
2.637
2.556
2.541
2.521
2.504
2.473
2.453
2.437
2.419
2.402

1.546
1.256

0.071
-0.000

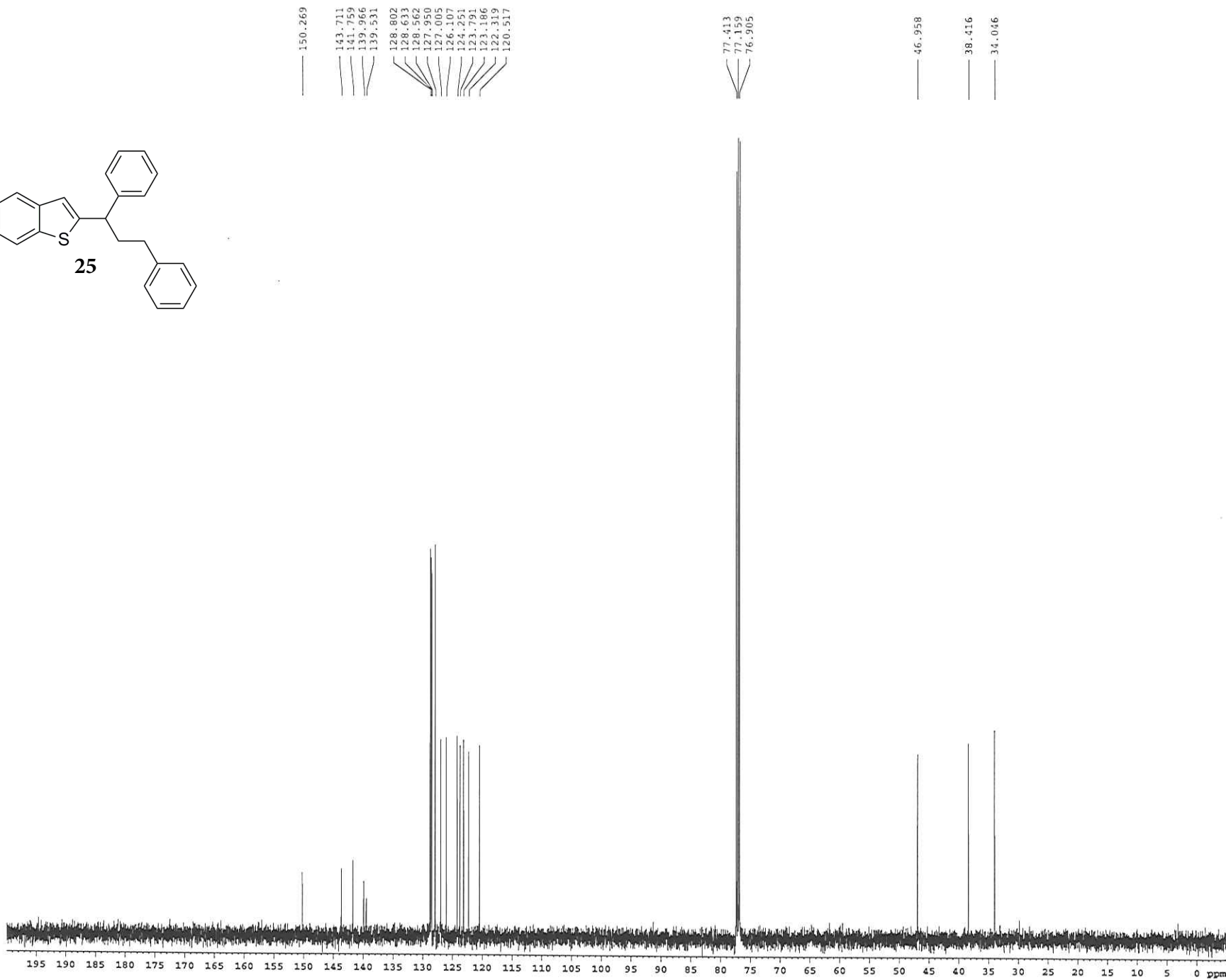
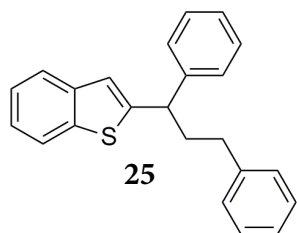
Current Data Parameters
 USER yonova
 NAME imy5 - 233
 EXPNO 20
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20130516
 Time 14.52
 INSTRUM dxs400
 PROBRD 5 mm QNP H/F/P
 PULPROG zg30
 TD 65536
 SOLVENT CDCl3
 NS 8
 DS 2
 SWH 6410.256 Hz
 FIDRES 0.097811 Hz
 AQ 5.1118579 sec
 RG 456.1
 DW 78.000 usec
 DE 4.80 usec
 TE 297.9 K
 D1 0.10000000 sec
 MCREST 0.00000000 sec
 MCMRK 0.01500000 sec

==== CHANNEL f1 =====
 NUC1 1H
 P1 12.00 usec
 PL1 -0.60 dB
 SFO1 400.1328009 MHz

F2 - Processing parameters
 SI 65536
 SF 400.1300212 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 2.00

Z-restored spin-echo 13C spectrum with 1H decoupling



Current Data Parameters
USER ymova
NAME imy5 - 133
EXPNO 222
PROCNO 1

F2 - Acquisition Parameters
Date_ 20130519
Time 18.26
INSTRUM cryso00
PROBHD 5 mm CPTCI 1H-
PULPROG SpinEcho30op.prd
TD 65536
SOLVENT cdcl3
NS 288
DS 16
SWH 30303.031 Hz
FIDRES 0.462388 Hz
AQ 1.0814105 sec
RG 5160.6
DM 16.500 usec
DE 8.00 usec
TE 298.0 K
D1 0.25000000 sec
d11 0.03000000 sec
d16 0.00200000 sec
d17 0.00019600 sec
MCRET 0.00000000 sec
MCRBK 0.01500000 sec
F2 31.00 usec

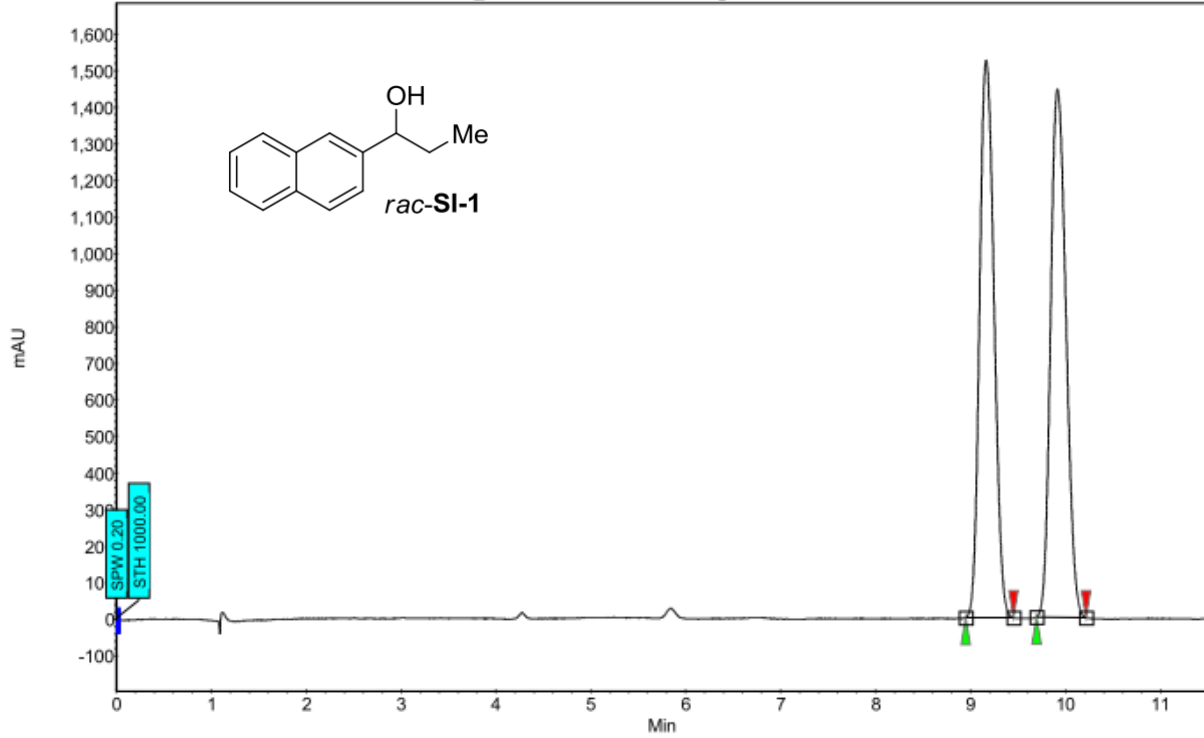
***** CHANNEL F1 *****
NUC1 13C
P1 15.50 usec
P11 500.00 usec
P12 2000.00 usec
PL0 120.00 dB
PL1 -1.00 dB
SFO1 125.7942548 MHz
SP1 3.20 dB
SP2 3.20 dB
GPNAM1 Crp60.0.9.220.1
GPNAM2 Crp60comp.4
SPOFF1 0.00 Hz
SPOFF2 0.00 Hz

***** CHANNEL F2 *****
CPDPRG2 waltz16
NUC2 1H
PCPD2 100.00 usec
PL2 1.60 dB
PL12 24.60 dB
SFO2 500.2225011 MHz

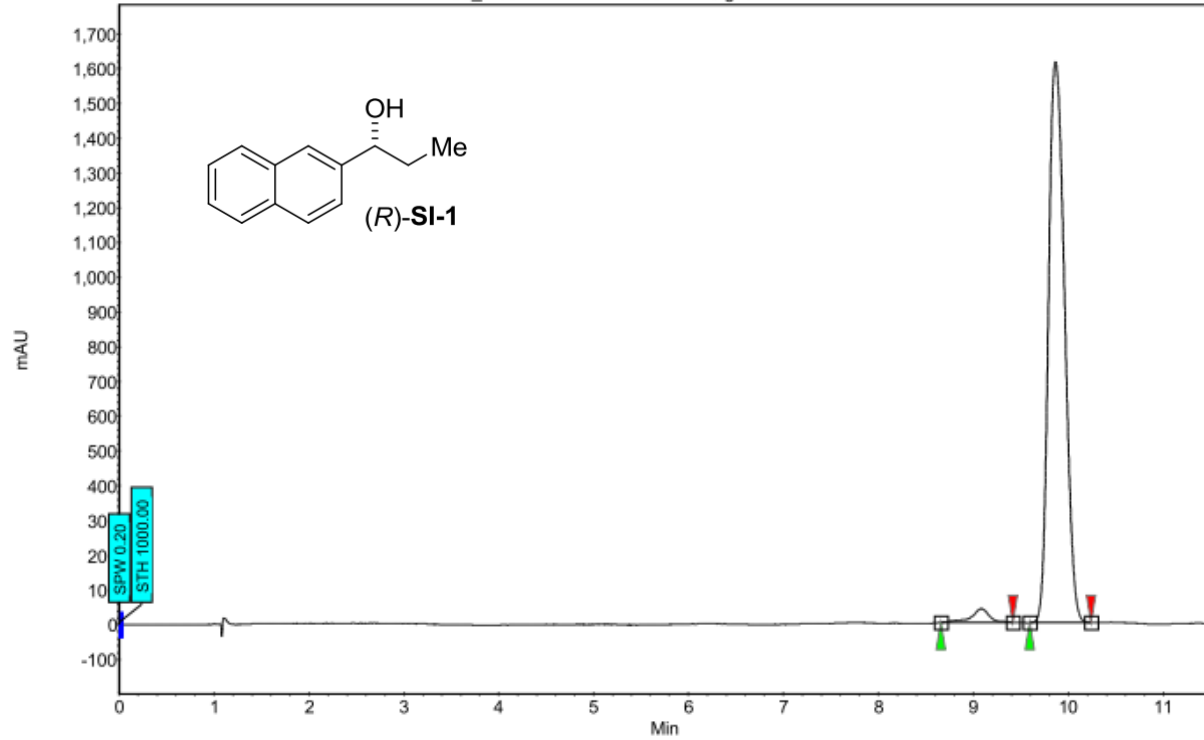
***** GRADIENT CHANNEL *****
GPNAM1 SINE.100
GPNAM2 SINE.100
GPX1 0.00 %
GPX2 0.00 %
GPY1 0.00 %
GPY2 0.00 %
GPI1 30.00 %
GPI2 50.00 %
p15 500.00 usec
p16 1000.00 usec

F2 - Processing parameters
SI 65536
SF 125.7804085 MHz
WDW EM
GB 0
LB 1.00 Hz
GB 0
PC 4.00

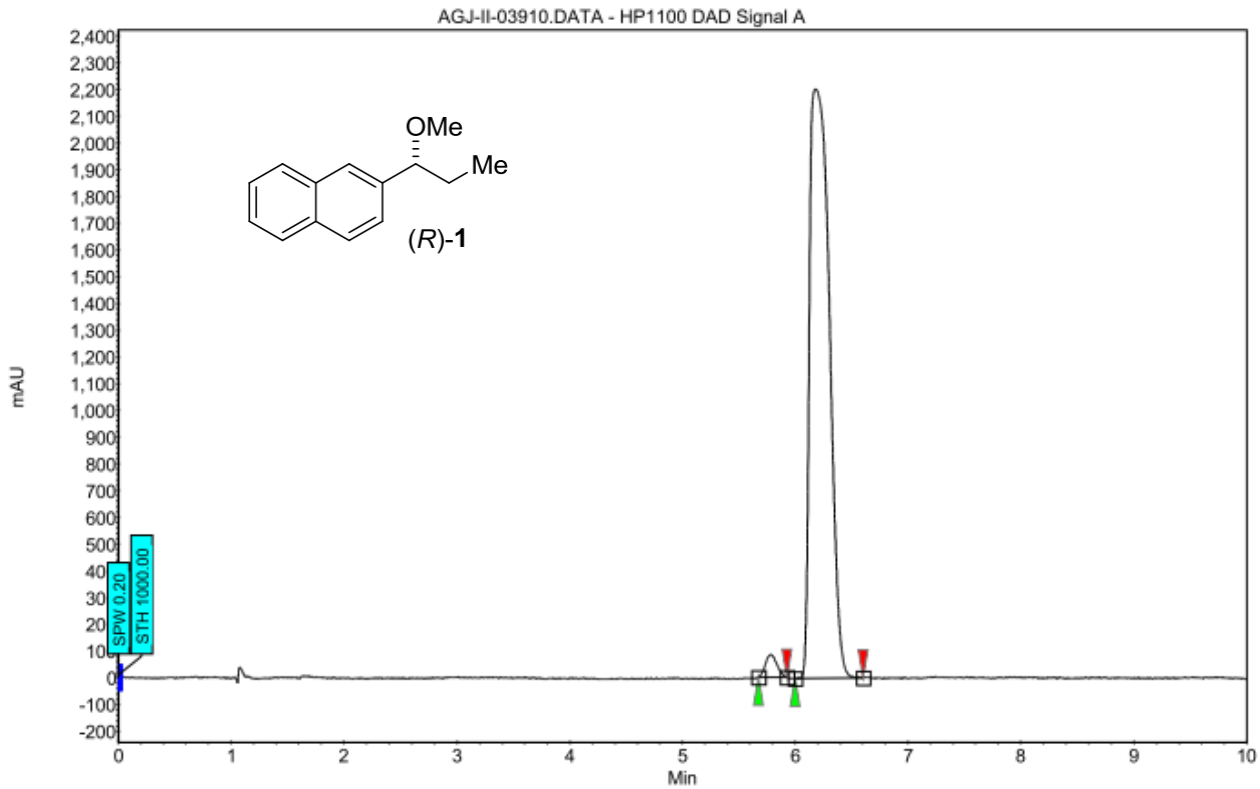
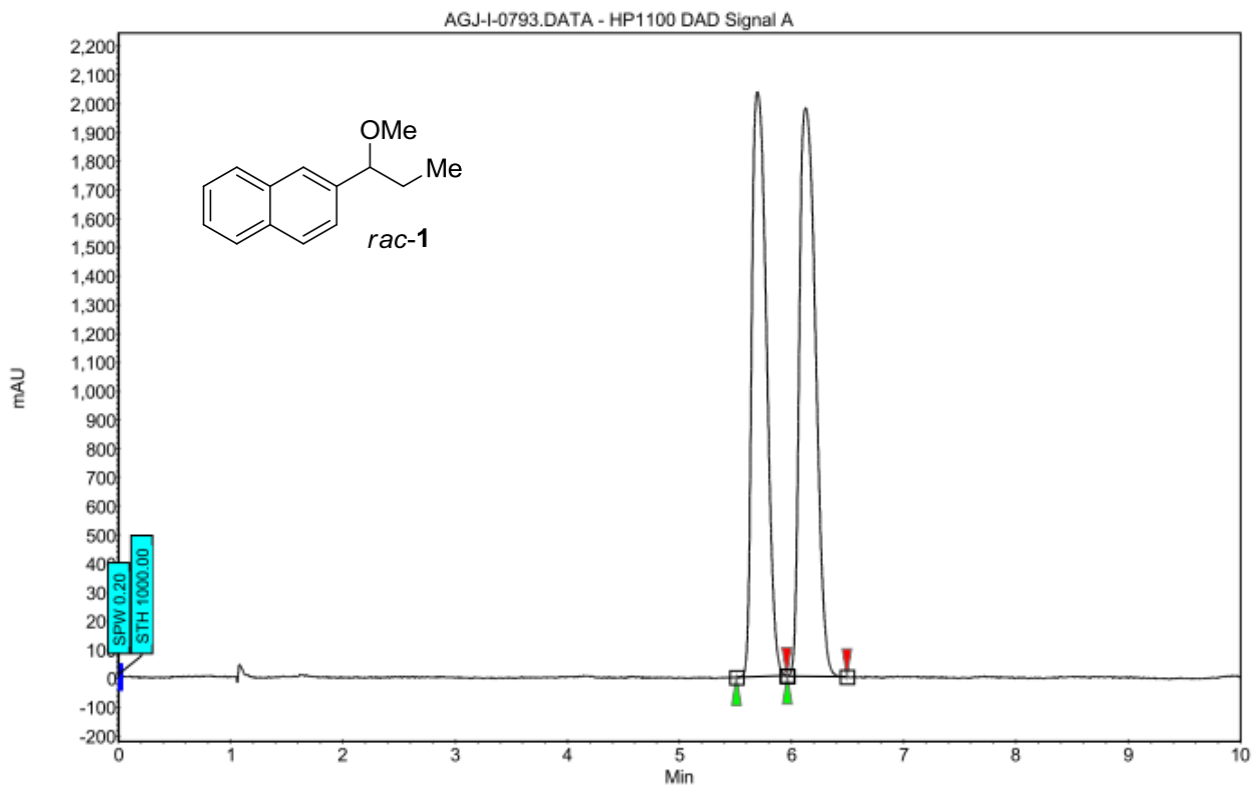
AGJ_2-82.DATA - HP1100 DAD Signal A



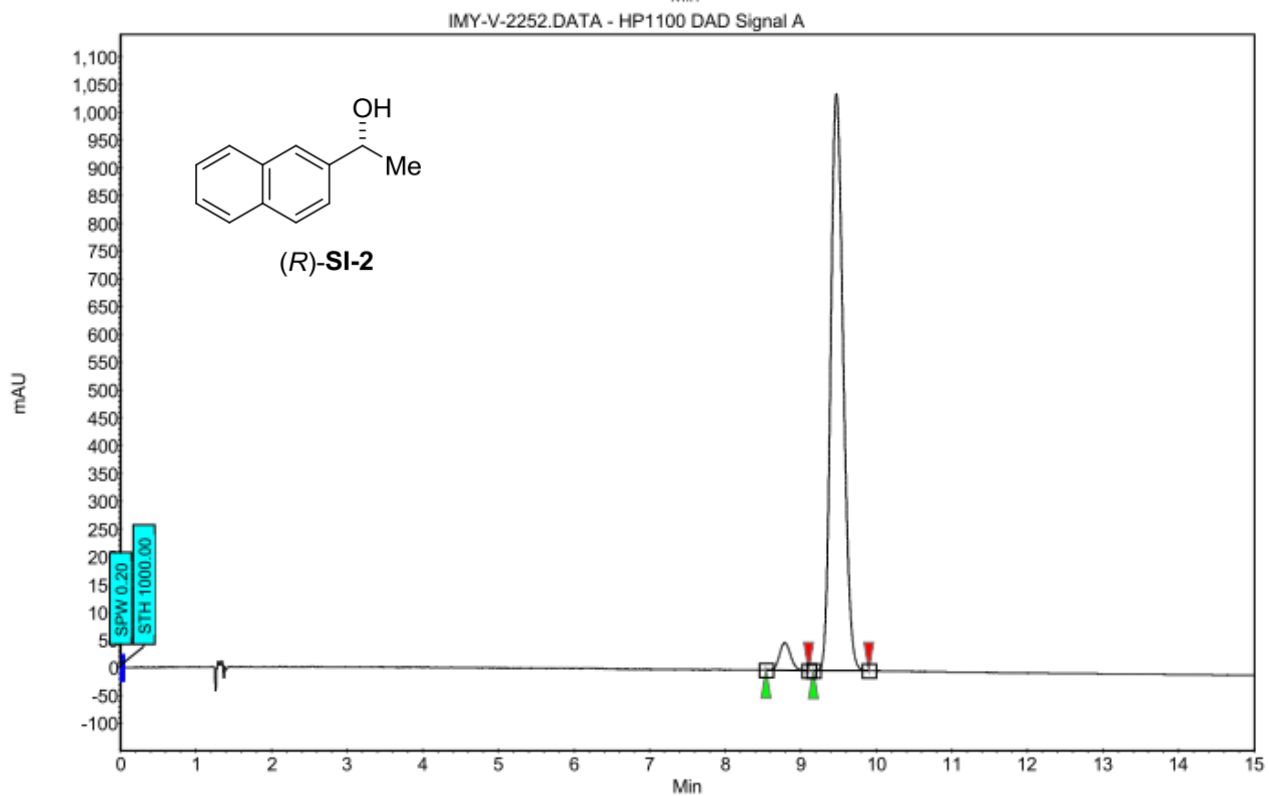
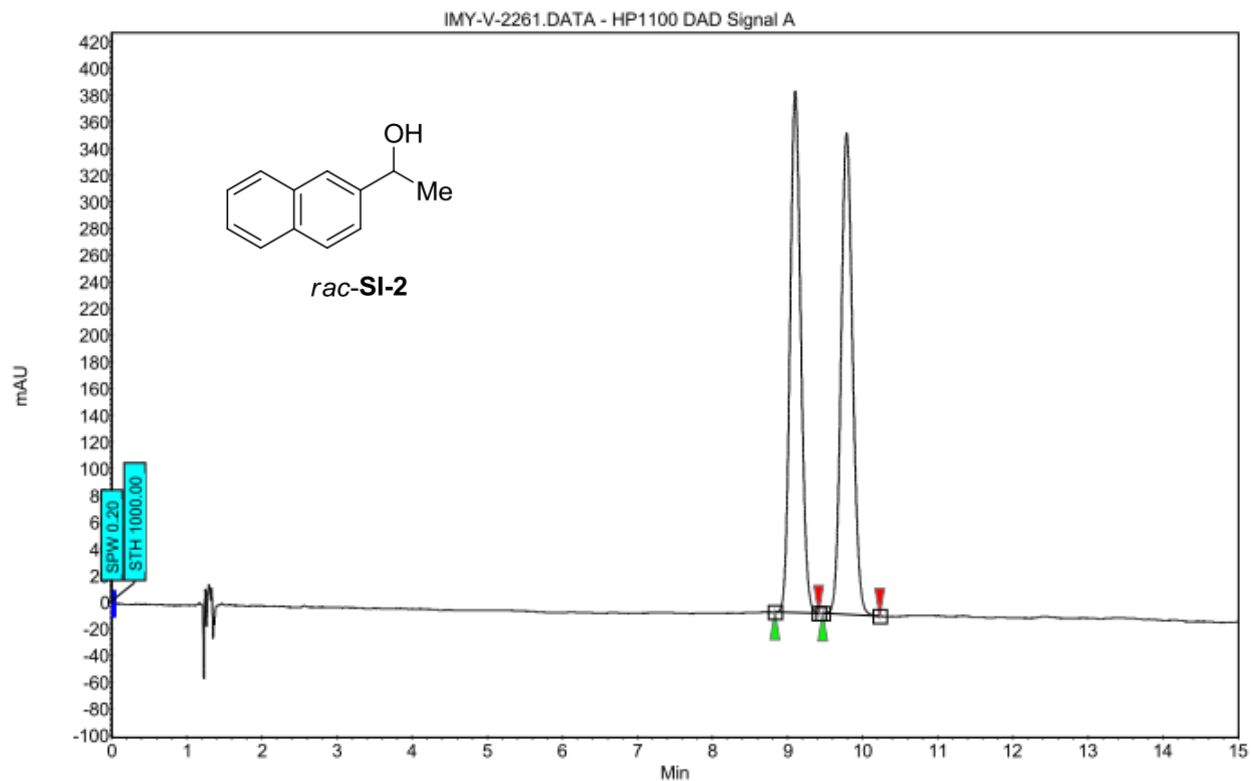
AGJ_2-382.DATA - HP1100 DAD Signal A



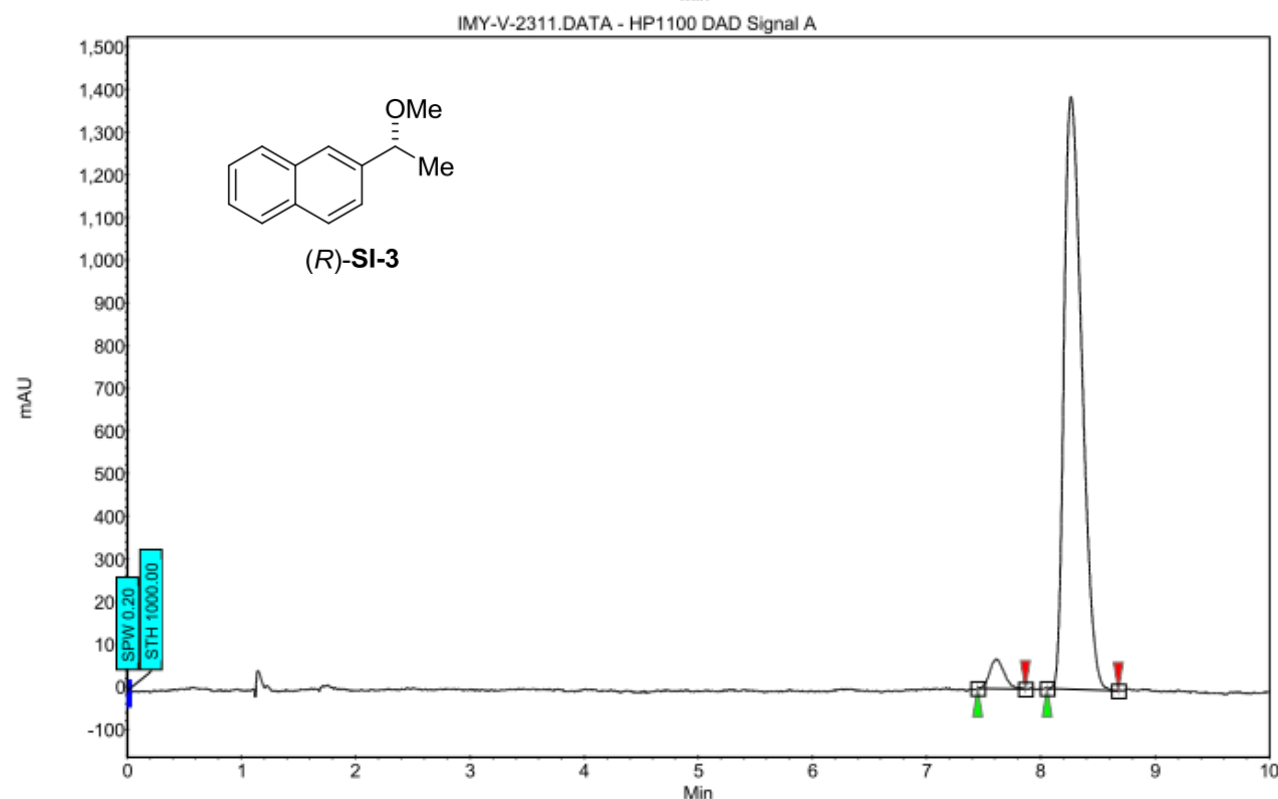
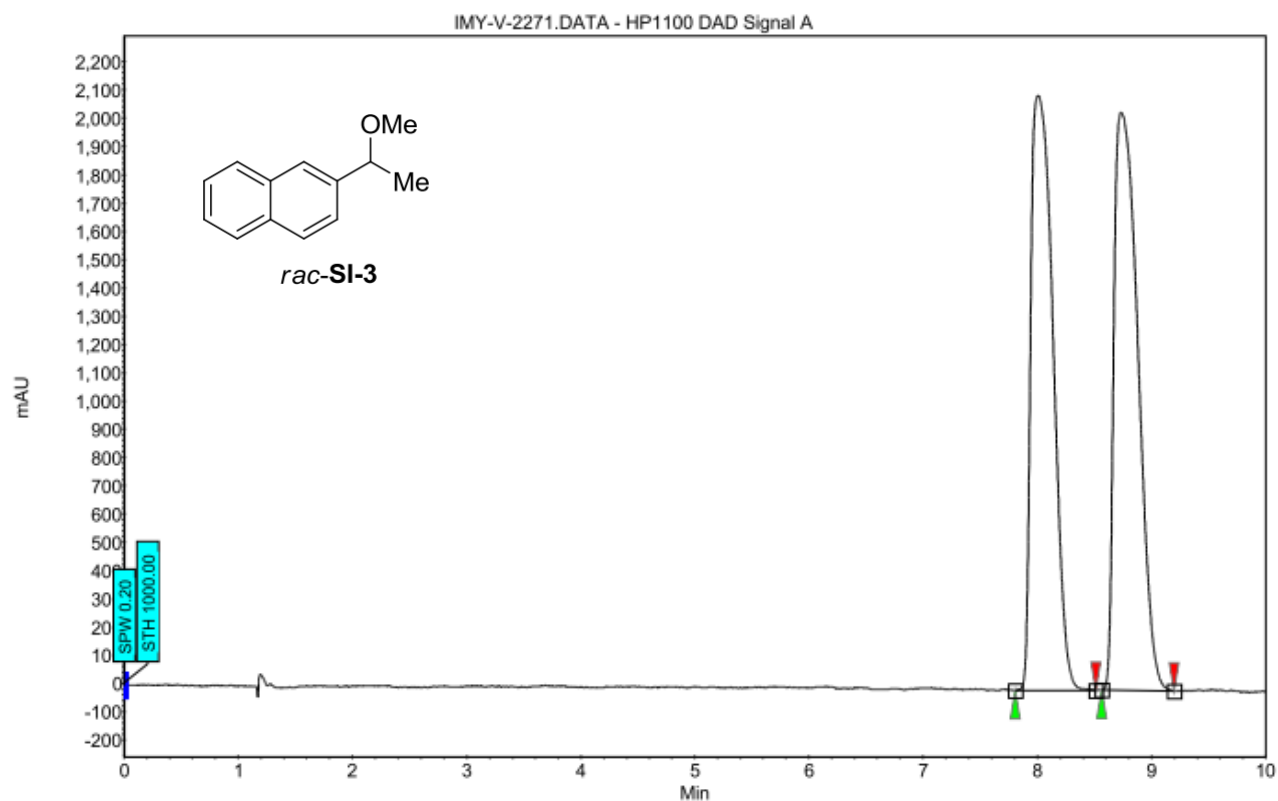
Index	Name	Start	Time	End	RT Offset	Quantity	Height	Area	Area
		[Min]	[Min]	[Min]	[Min]	[% Area]	[μ V]	[μ V.Min]	[%]
2	UNKNOWN	8.66	9.08	9.42	0.00	2.69	39.6	8.8	2.691
1	UNKNOWN	9.60	9.86	10.24	0.00	97.31	1613.2	316.9	97.309
Total						100.00	1652.8	325.7	100.000



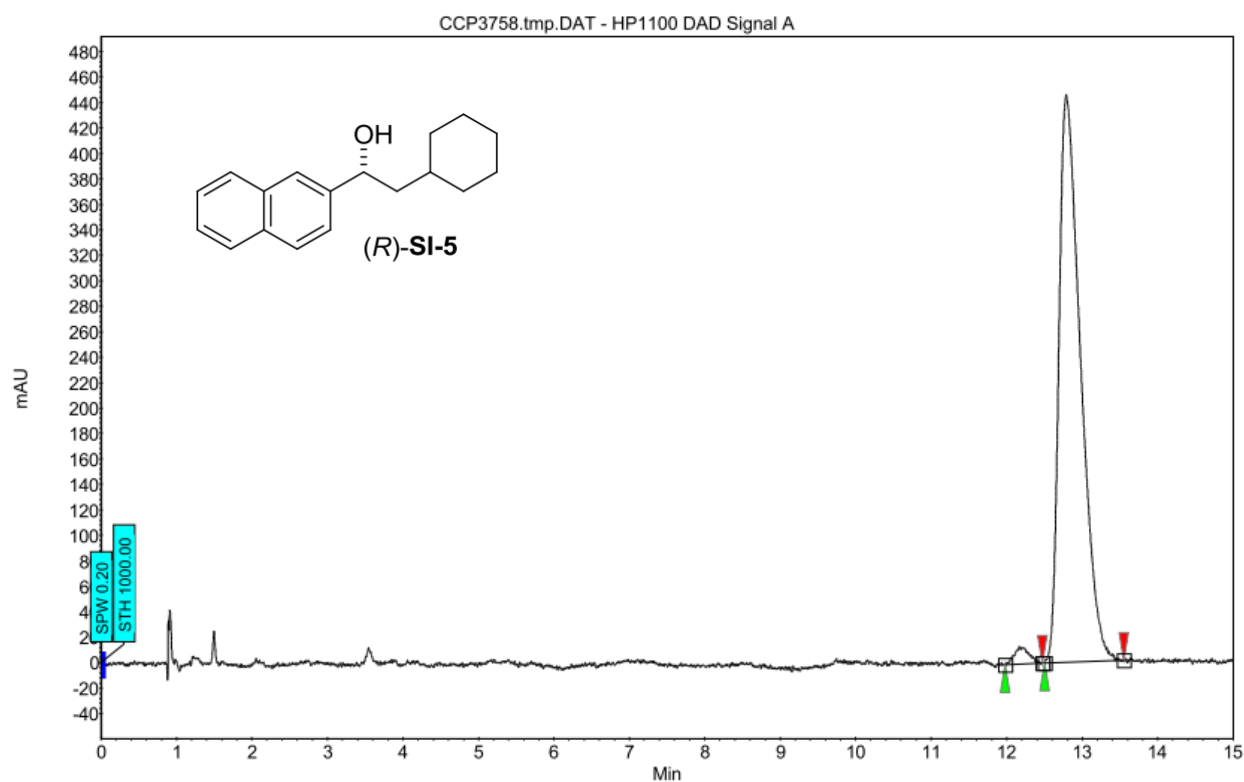
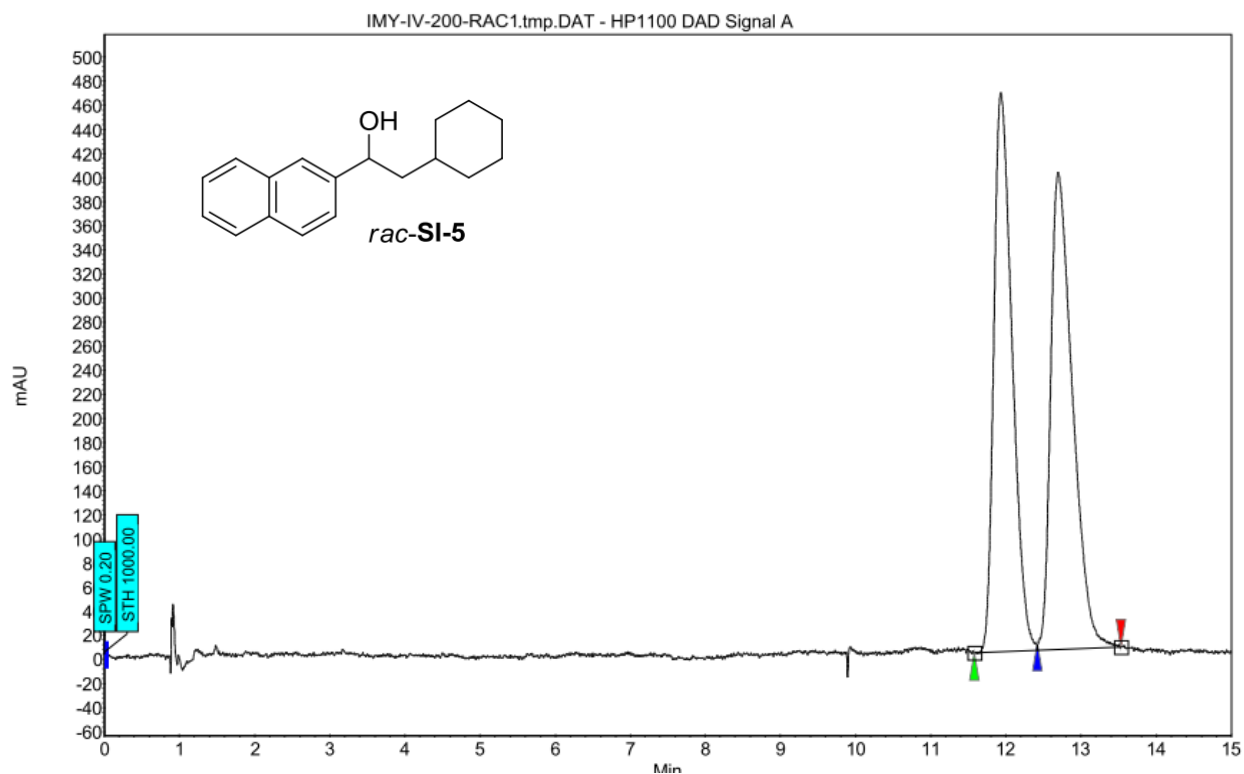
Index	Name	Start	Time	End	RT Offset	Quantity	Height	Area	Area
		[Min]	[Min]	[Min]	[Min]	[% Area]	[μ V]	[μ V.Min]	[%]
2	UNKNOWN	5.67	5.78	5.93	0.00	2.19	84.6	10.0	2.186
1	UNKNOWN	6.00	6.18	6.60	0.00	97.81	2204.3	445.6	97.814
Total						100.00	2288.8	455.6	100.000



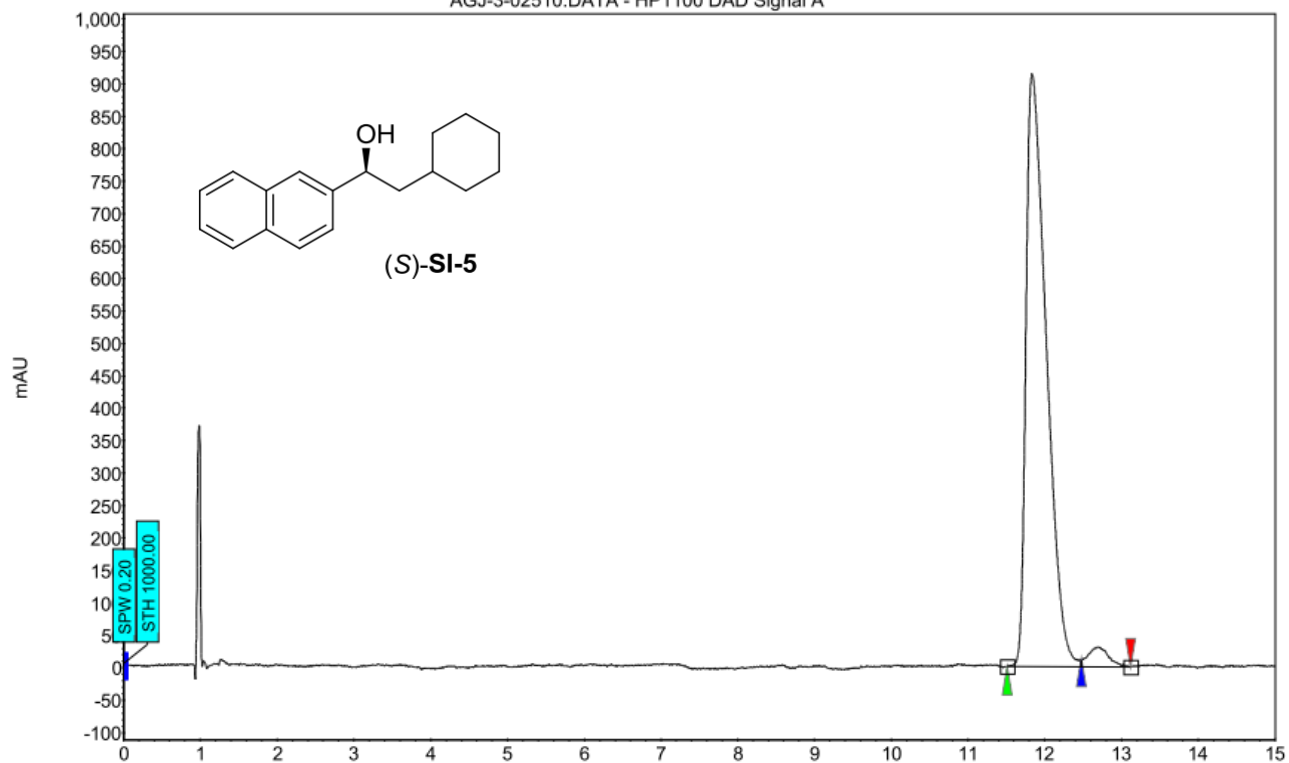
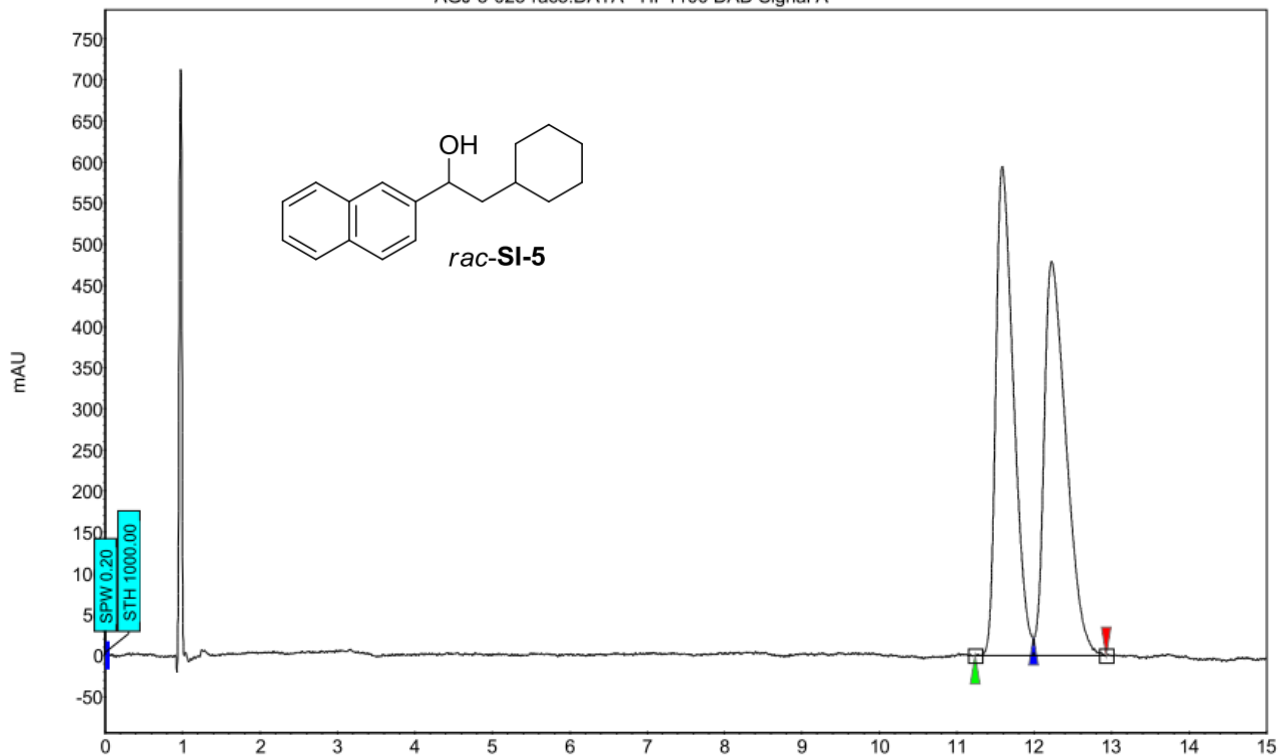
Index	Name	Start	Time	End	RT Offset	Quantity	Height	Area	Area
		[Min]	[Min]	[Min]	[Min]	[% Area]	[μ V]	[μ V.Min]	[%]
1	UNKNOWN	8.54	8.79	9.10	0.00	4.06	49.8	8.4	4.057
2	UNKNOWN	9.17	9.48	9.91	0.00	95.94	1038.5	198.9	95.943
Total						100.00	1088.3	207.3	100.000



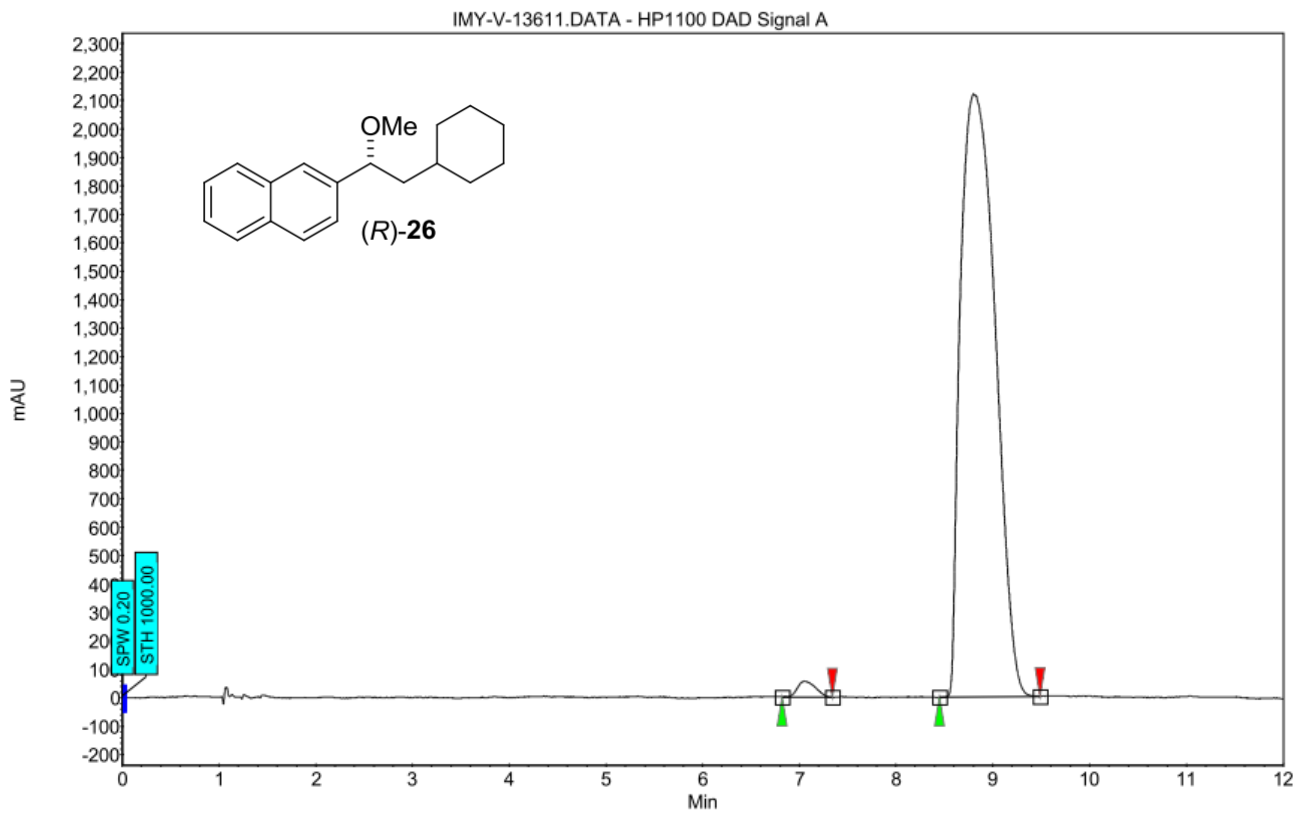
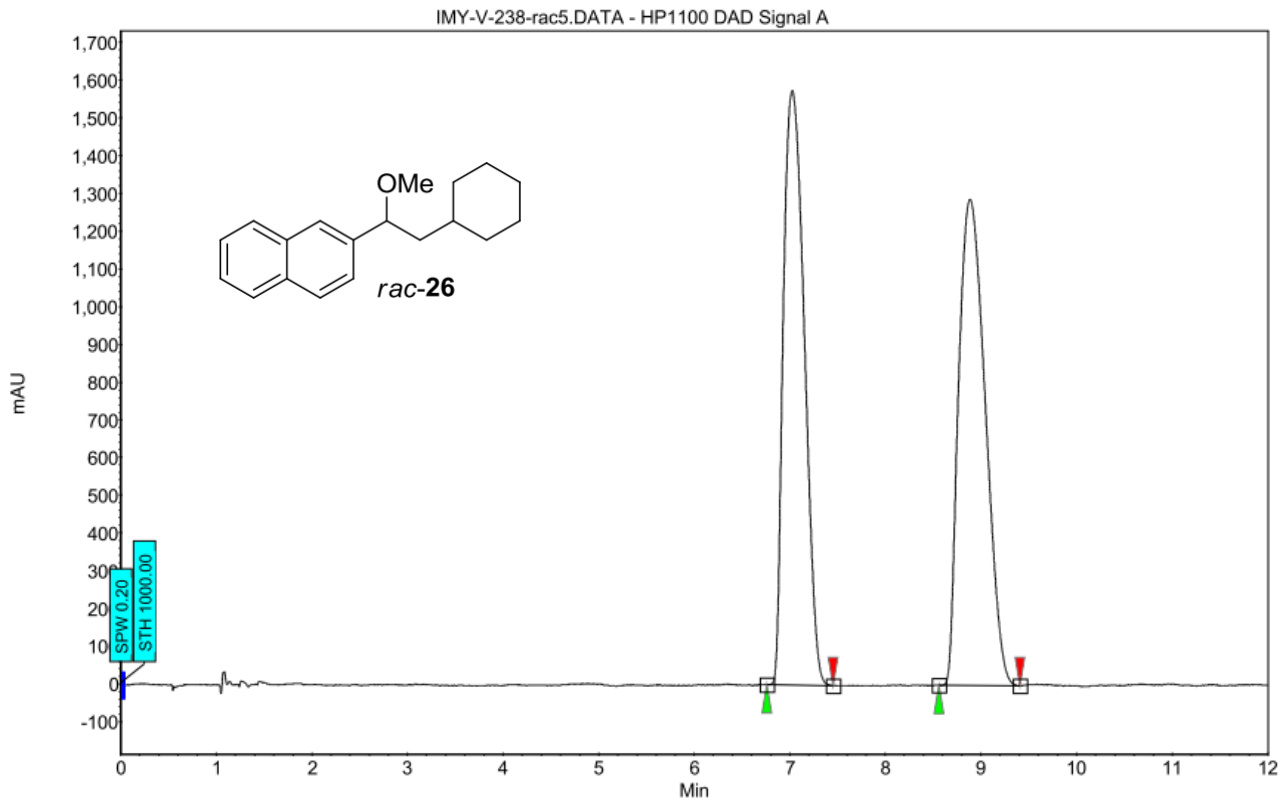
Index	Name	Start	Time	End	RT Offset	Quantity	Height	Area	Area
		[Min]	[Min]	[Min]	[Min]	[% Area]	[μ V]	[μ V.Min]	[%]
1	UNKNOWN	7.45	7.61	7.86	0.00	3.88	69.8	10.1	3.884
2	UNKNOWN	8.06	8.26	8.68	0.00	96.12	1389.0	250.7	96.116
Total						100.00	1458.8	260.8	100.000



Index	Name	Start	Time	End	RT Offset	Quantity	Height	Area	Area
		[Min]	[Min]	[Min]	[Min]	[% Area]	[μ V]	[μ V.Min]	[%]
1	UNKNOWN	11.98	12.17	12.47	0.00	2.01	13.9	3.0	2.010
2	UNKNOWN	12.50	12.79	13.55	0.00	97.99	445.8	147.3	97.990
Total						100.00	459.7	150.3	100.000

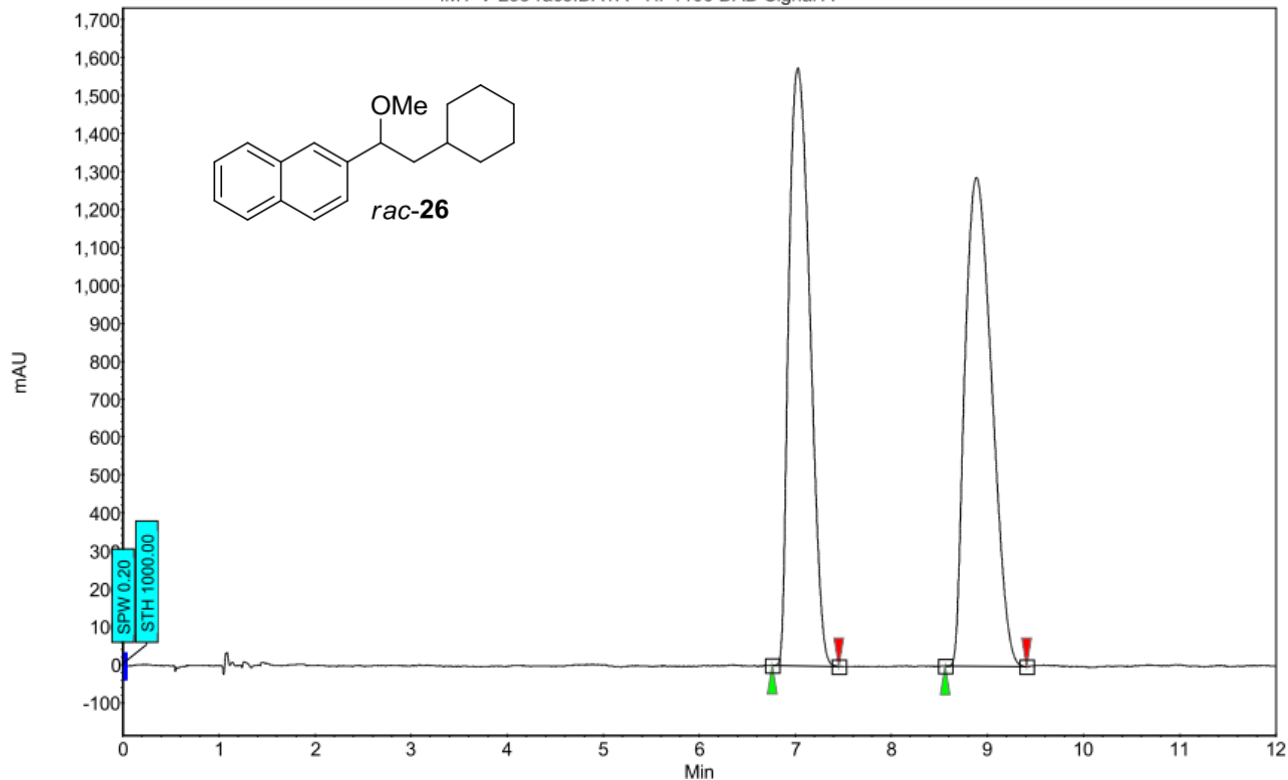


Index	Name	Start	Time	End	RT Offset	Quantity	Height	Area	Area
		[Min]	[Min]	[Min]	[Min]	[% Area]	[μ V]	[μ V.Min]	[%]
1	UNKNOWN	11.51	11.83	12.47	0.00	96.72	913.1	279.8	96.723
2	UNKNOWN	12.47	12.70	13.11	0.00	3.28	30.9	9.5	3.277
Total						100.00	944.0	289.3	100.000

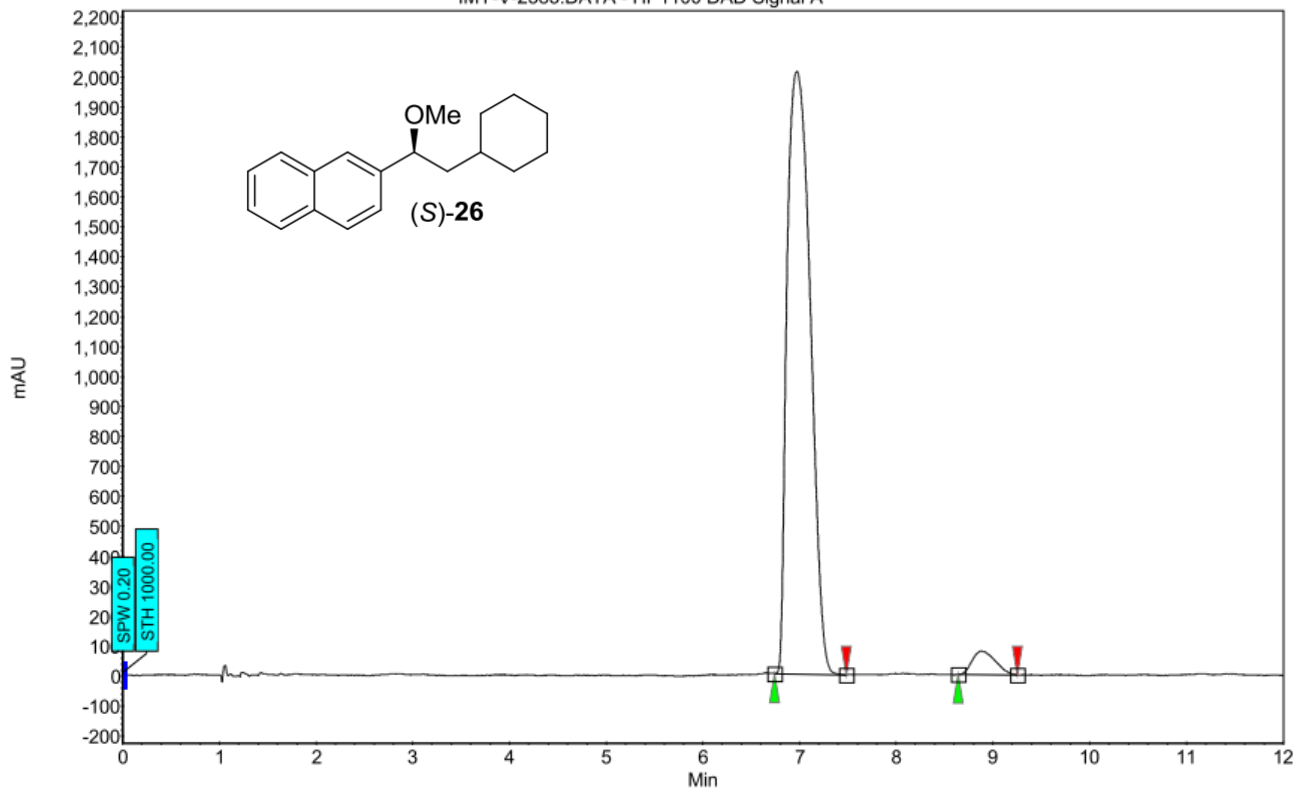


Index	Name	Start	Time	End	RT Offset	Quantity	Height	Area	Area
		[Min]	[Min]	[Min]	[Min]	[% Area]	[μ V]	[μ V.Min]	[%]
2	UNKNOWN	6.82	7.05	7.34	0.00	1.35	56.2	12.1	1.352
1	UNKNOWN	8.45	8.80	9.49	0.00	98.65	2120.6	879.3	98.648
Total						100.00	2176.8	891.4	100.000

IMY-V-238-rac5.DATA - HP1100 DAD Signal A

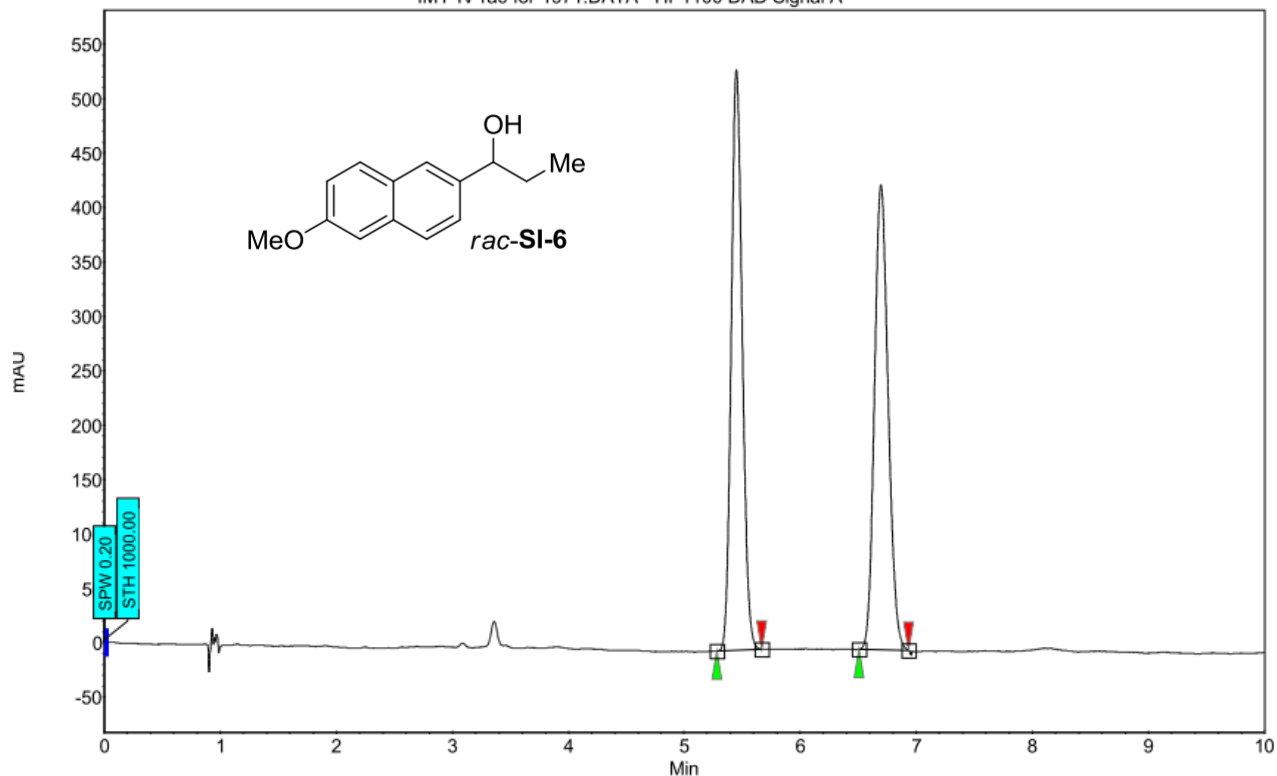


IMY-V-2385.DATA - HP1100 DAD Signal A

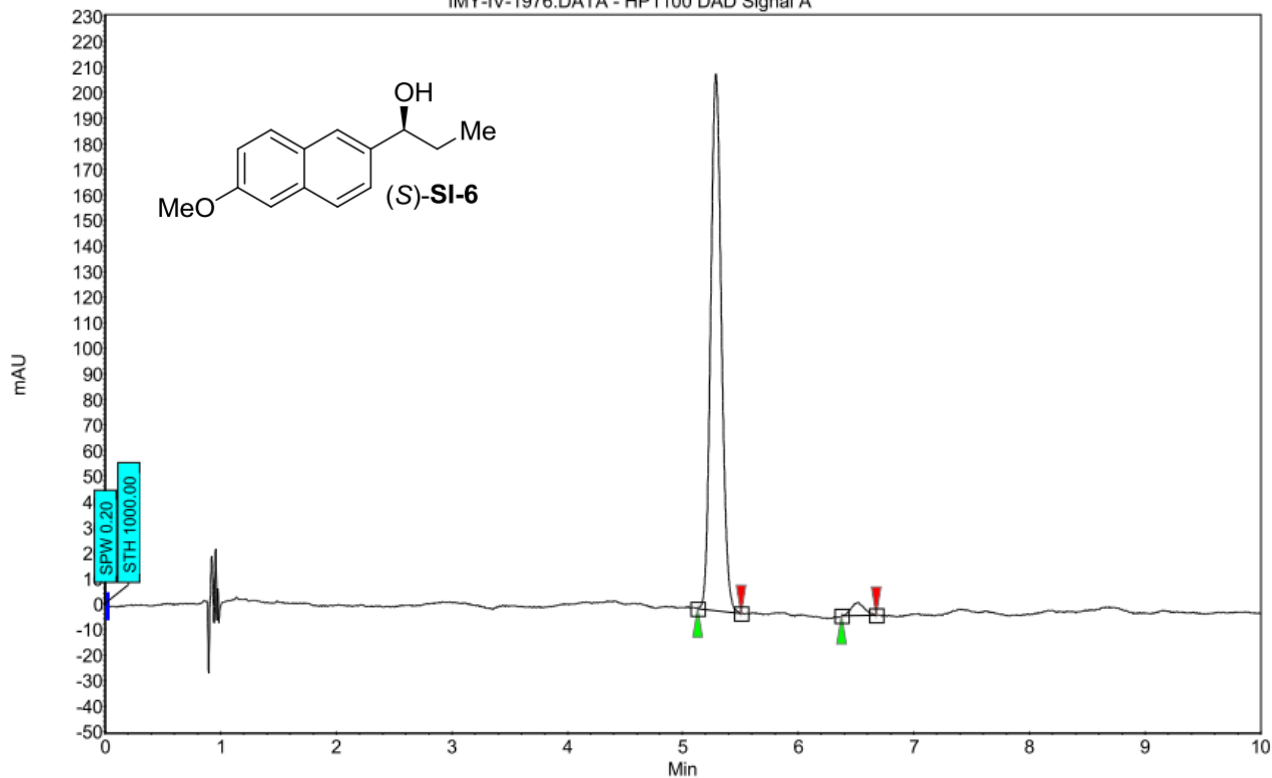


Index	Name	Start	Time	End	RT Offset	Quantity	Height	Area	Area
		[Min]	[Min]	[Min]	[Min]	[% Area]	[μ V]	[μ V.Min]	[%]
1	UNKNOWN	6.75	6.98	7.49	0.00	96.23	2011.3	558.6	96.229
2	UNKNOWN	8.64	8.87	9.25	0.00	3.77	78.5	21.9	3.771
Total						100.00	2089.9	580.5	100.000

IMY-IV-rac-for-1971.DATA - HP1100 DAD Signal A

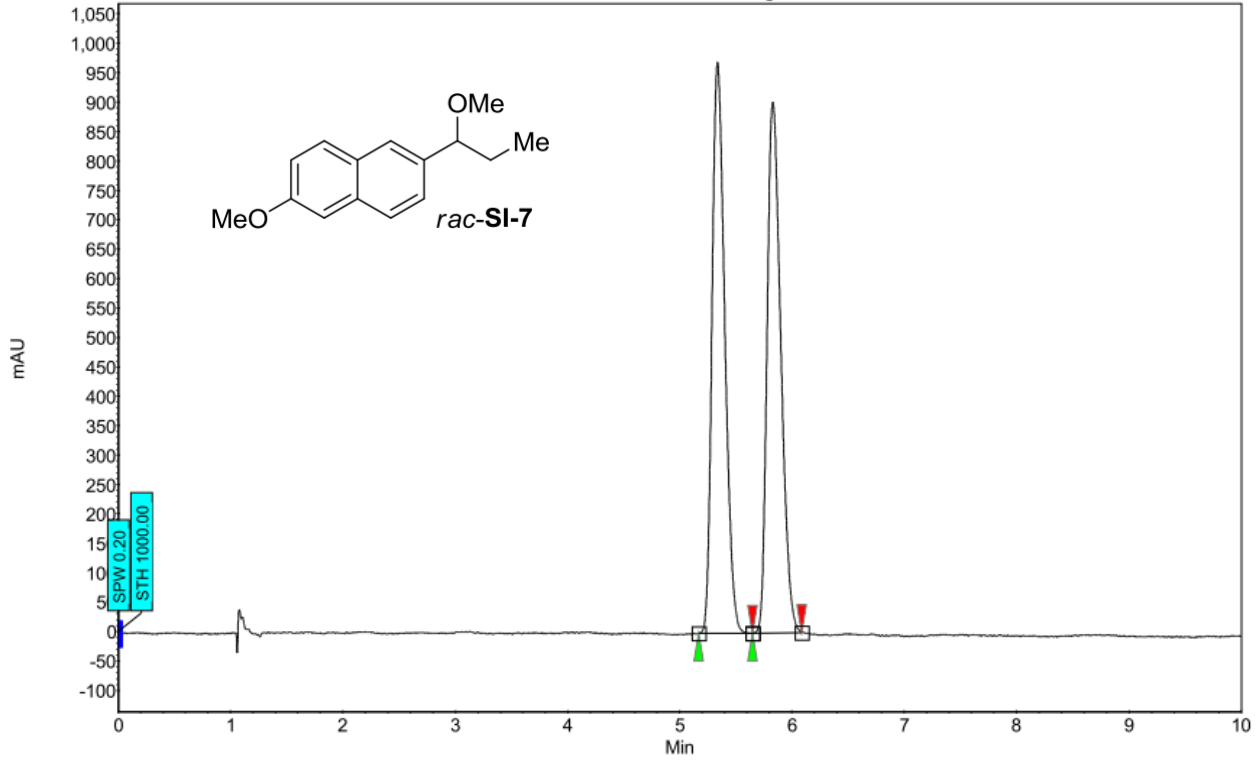


IMY-IV-1976.DATA - HP1100 DAD Signal A

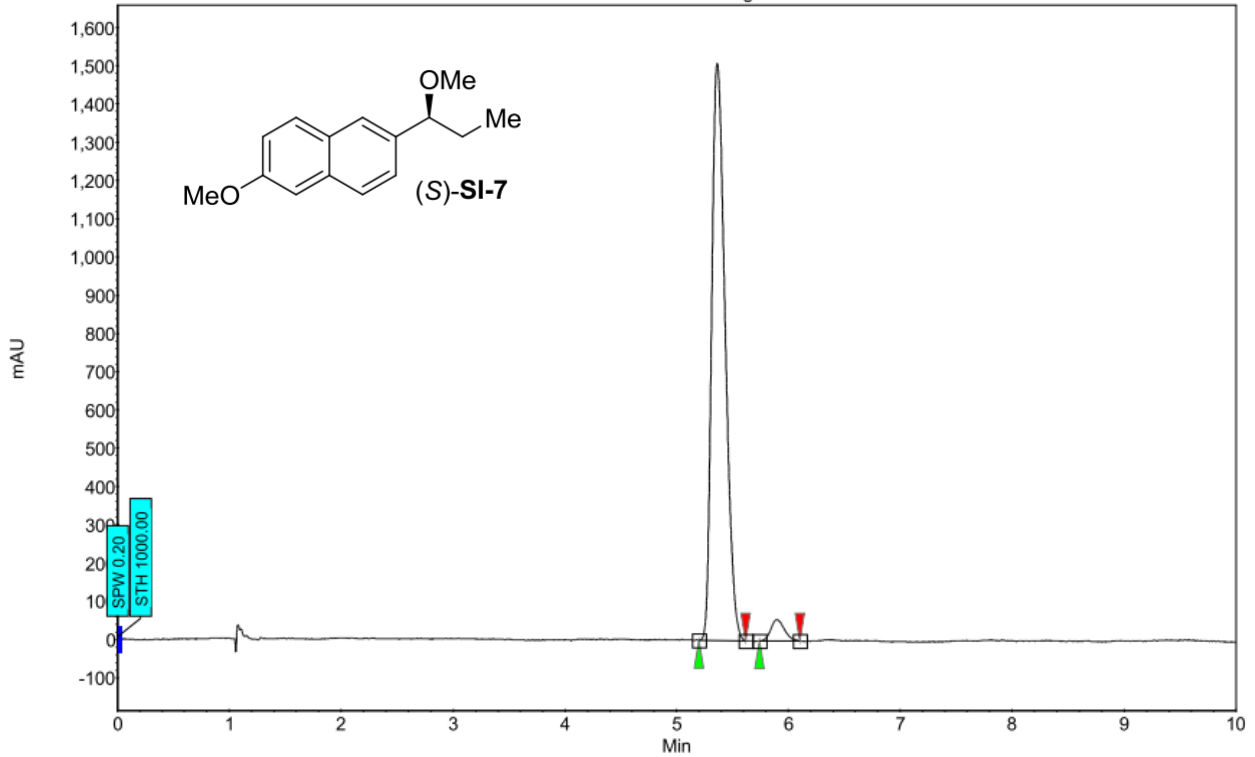


Index	Name	Start	Time	End	RT Offset	Quantity	Height	Area	Area
		[Min]	[Min]	[Min]	[Min]	[% Area]	[μ V]	[μ V.Min]	[%]
1	UNKNOWN	5.13	5.29	5.51	0.00	97.12	209.9	22.7	97.119
2	UNKNOWN	6.38	6.52	6.67	0.00	2.88	5.2	0.7	2.881
Total						100.00	215.0	23.4	100.000

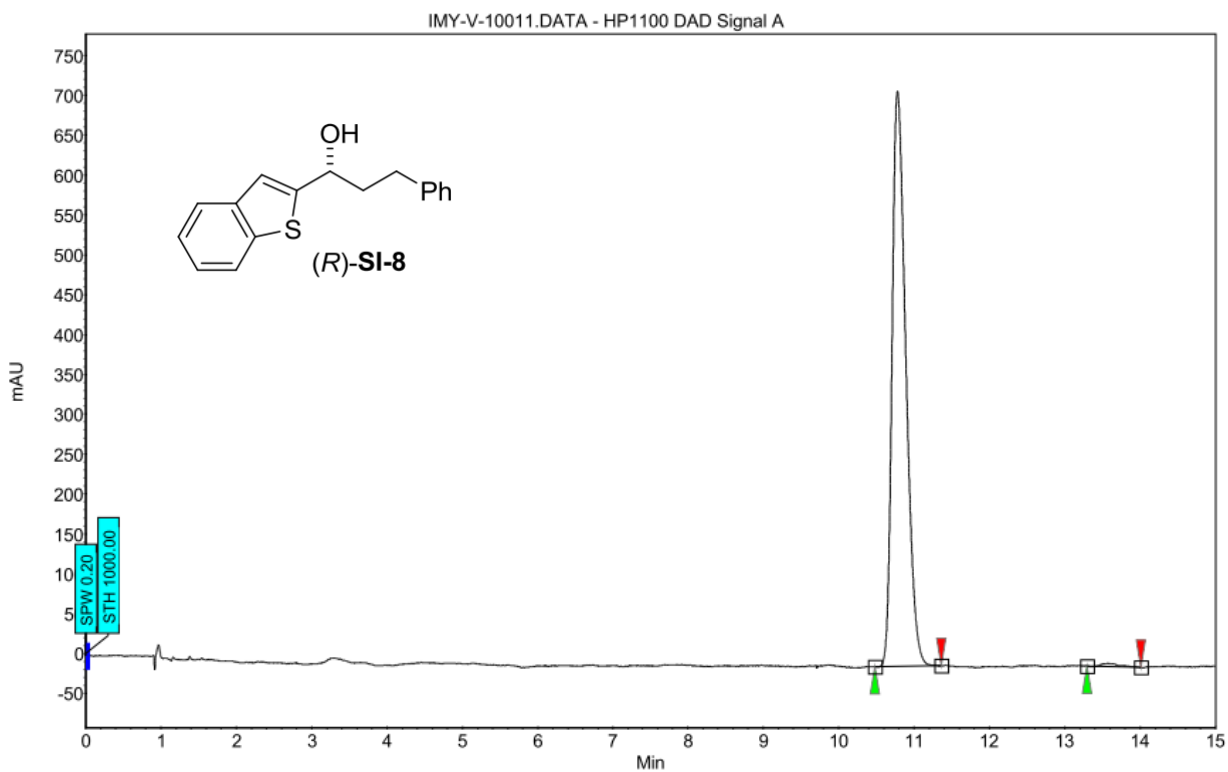
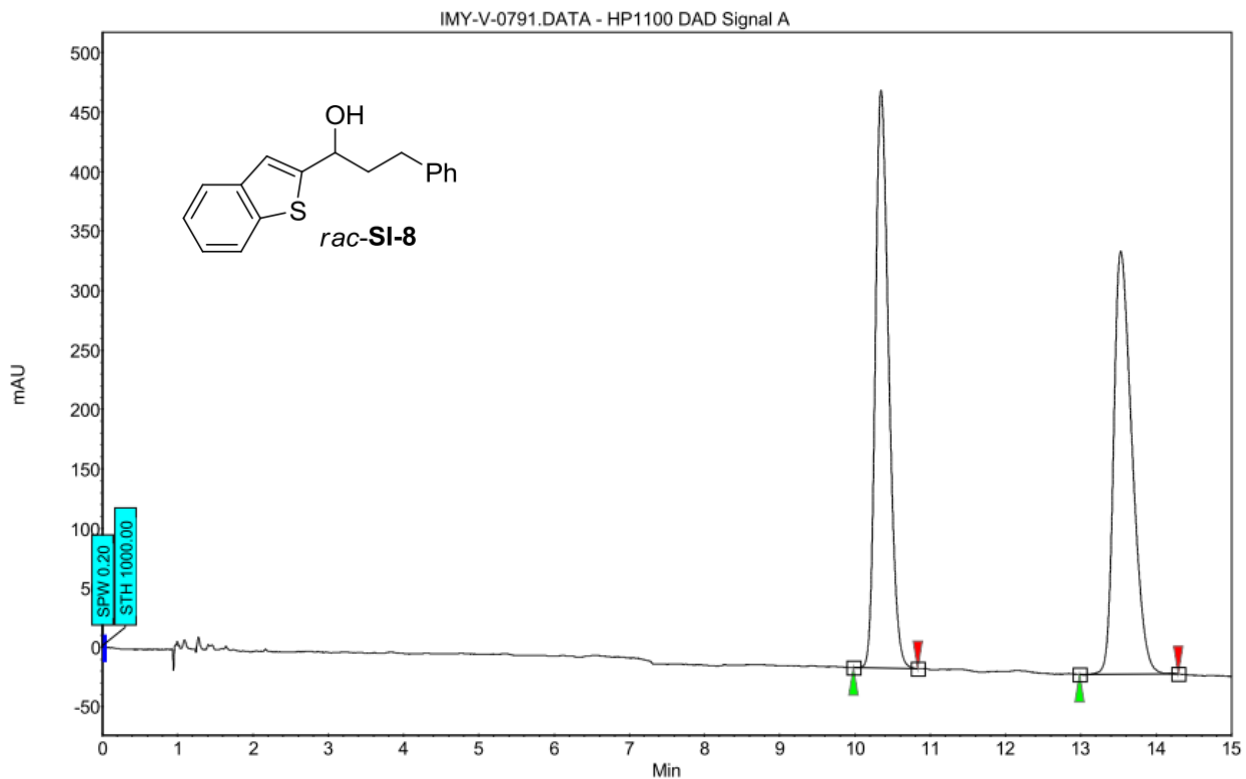
IMY-IV-2012RAC2.DATA - HP1100 DAD Signal A



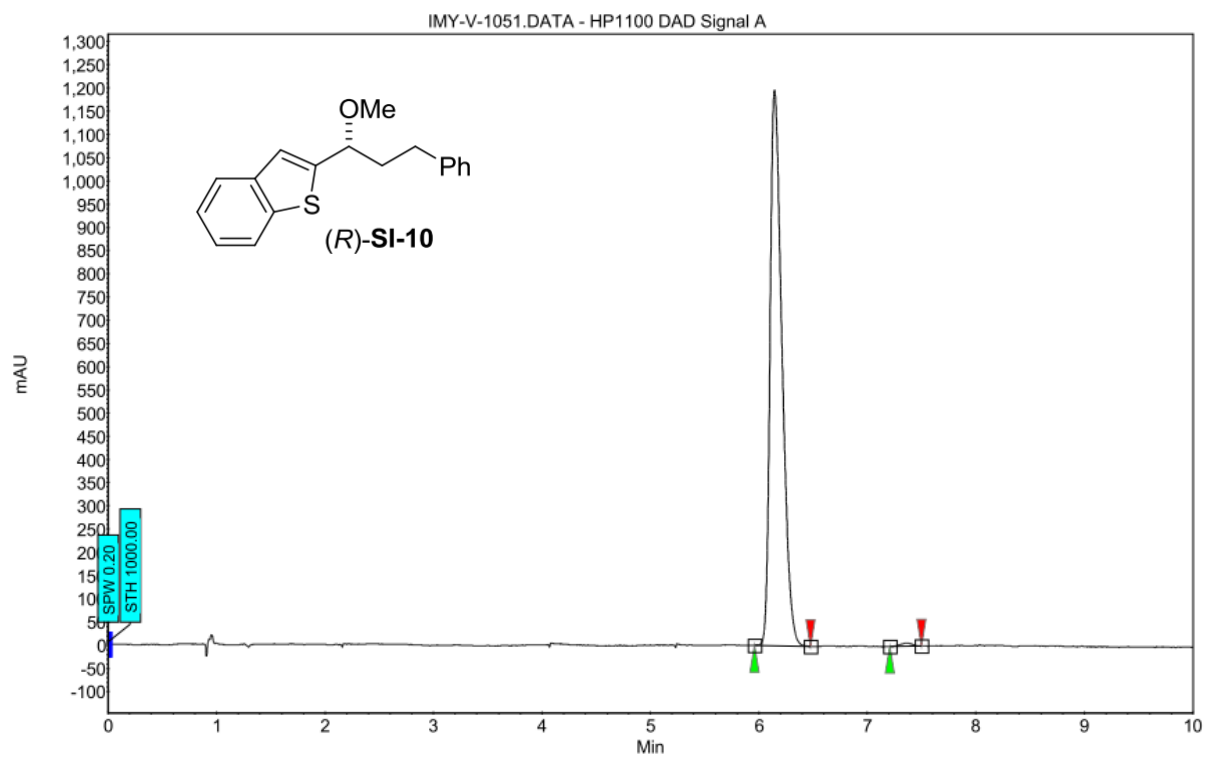
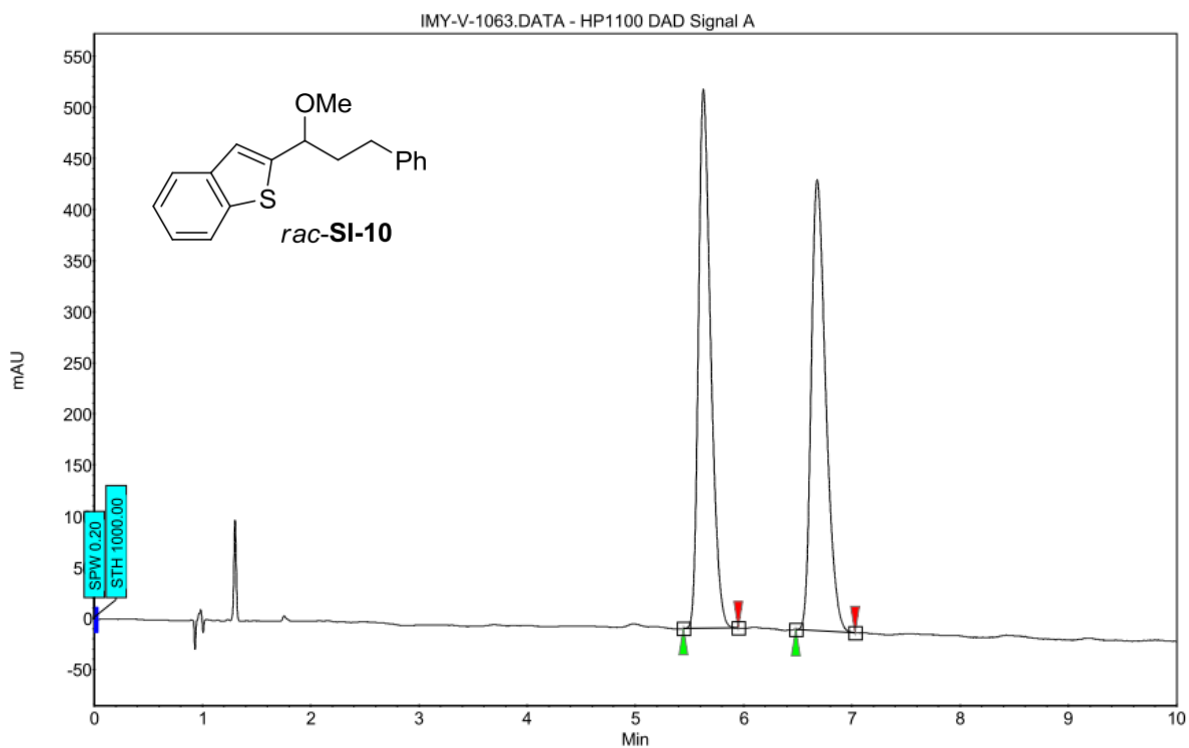
IMY-IV-2012.DATA - HP1100 DAD Signal A



Index	Name	Start	Time	End	RT Offset	Quantity	Height	Area	Area
		[Min]	[Min]	[Min]	[Min]	[% Area]	[μ V]	[μ V.Min]	[%]
1	UNKNOWN	5.20	5.37	5.62	0.00	96.49	1508.6	209.6	96.491
2	UNKNOWN	5.74	5.90	6.10	0.00	3.51	55.5	7.6	3.509
Total						100.00	1564.1	217.3	100.000

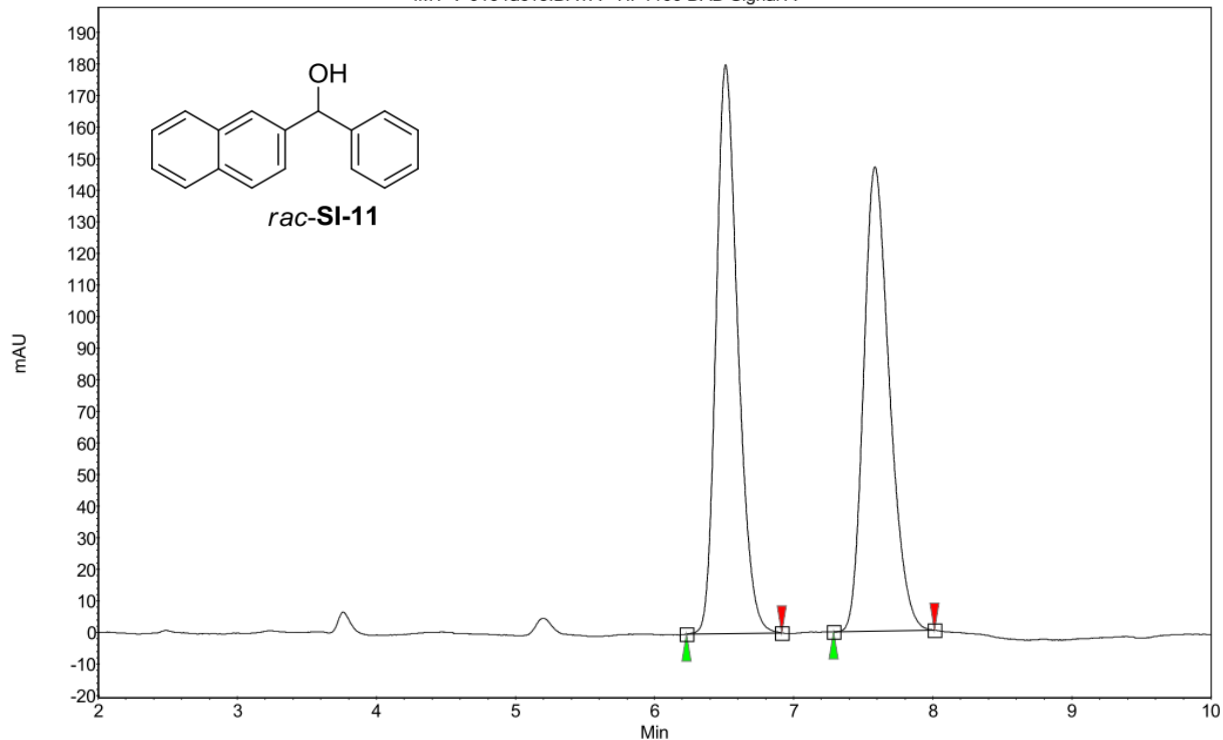


Index	Name	Start	Time	End	RT Offset	Quantity	Height	Area	Area
		[Min]	[Min]	[Min]	[Min]	[% Area]	[μ V]	[μ V.Min]	[%]
1	UNKNOWN	10.48	10.78	11.36	0.00	99.34	720.8	151.9	99.336
2	UNKNOWN	13.30	13.59	14.01	0.00	0.66	4.2	1.0	0.664
Total						100.00	725.0	153.0	100.000

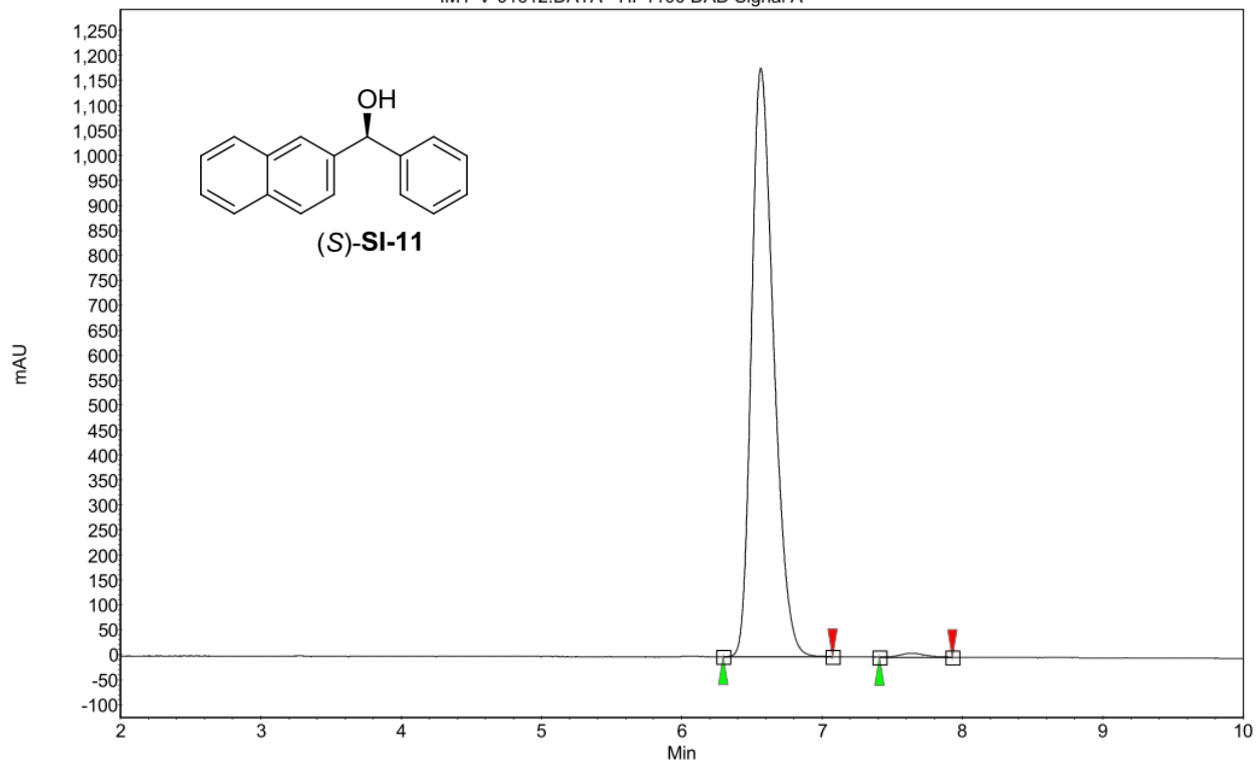


Index	Name	Start	Time	End	RT Offset	Quantity	Height	Area	Area
		[Min]	[Min]	[Min]	[Min]	[% Area]	[μ V]	[μ V.Min]	[%]
1	UNKNOWN	5.96	6.15	6.48	0.00	99.49	1196.1	152.3	99.490
2	UNKNOWN	7.21	7.37	7.50	0.00	0.51	6.6	0.8	0.510
Total						100.00	1202.7	153.0	100.000

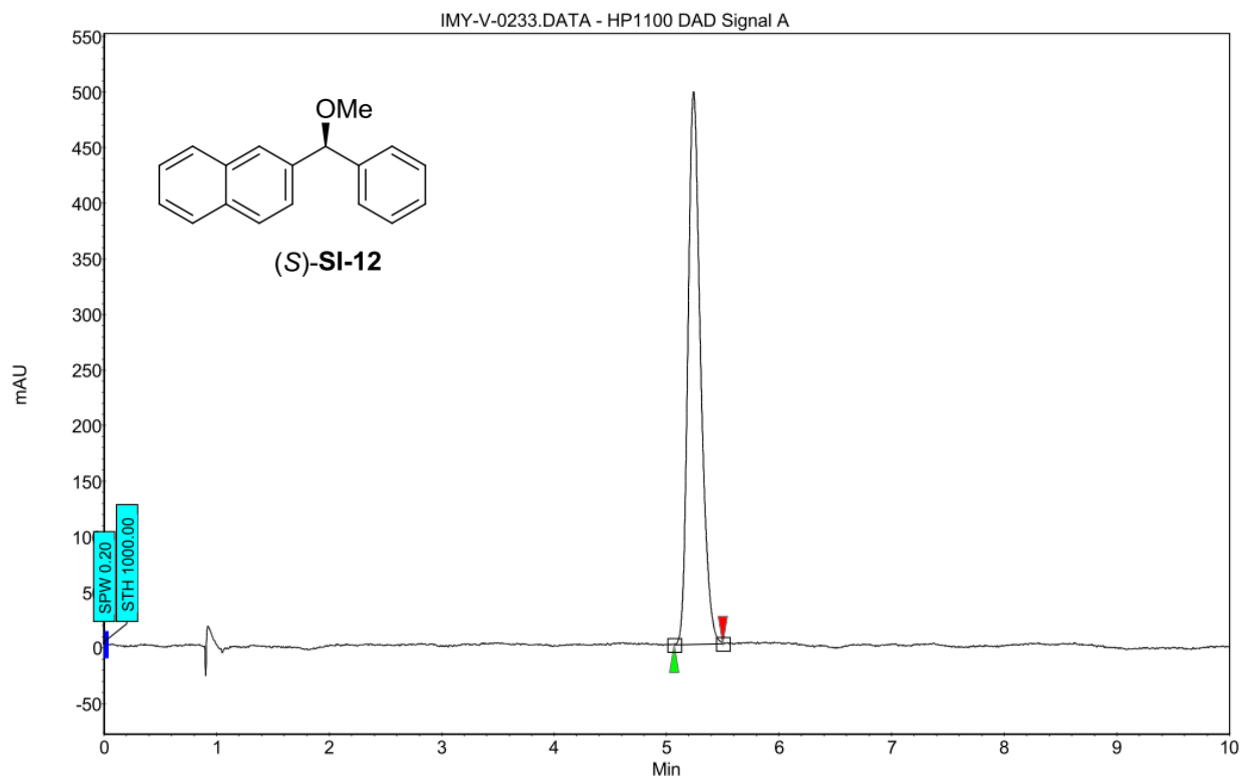
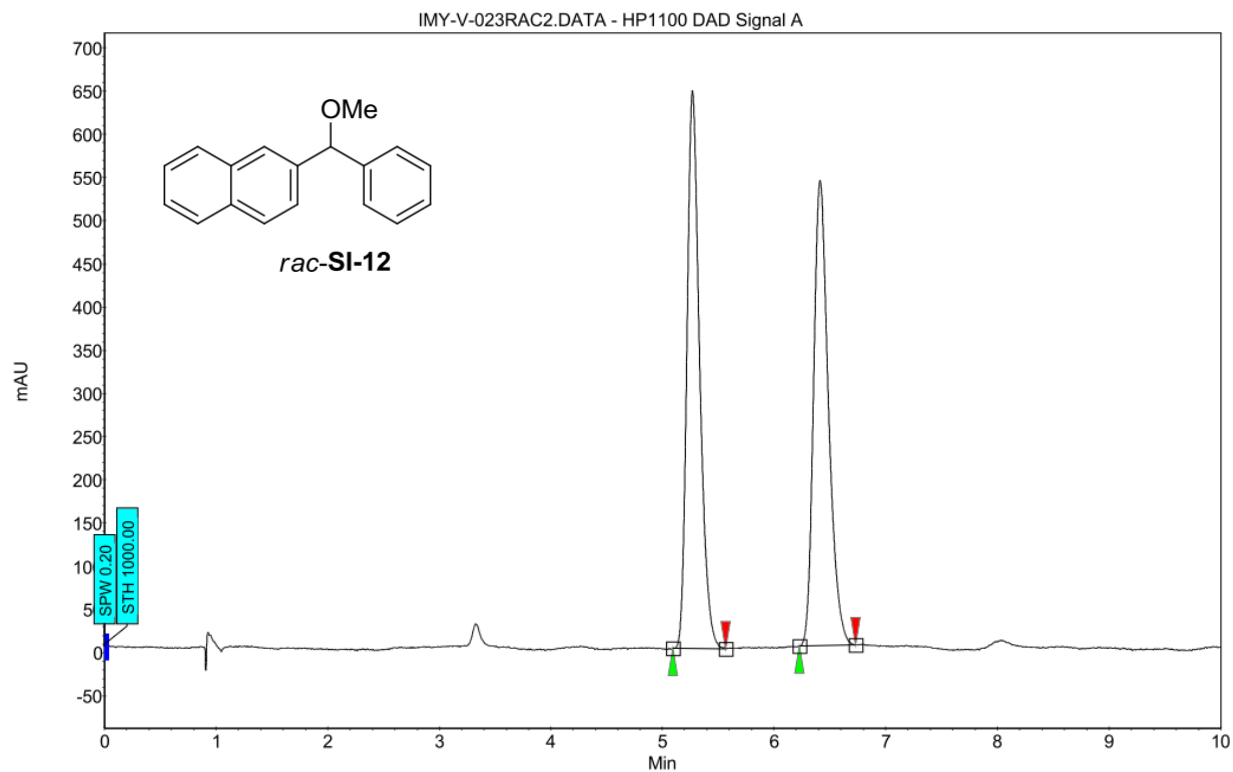
IMY-V-016 rac13.DATA - HP1100 DAD Signal A



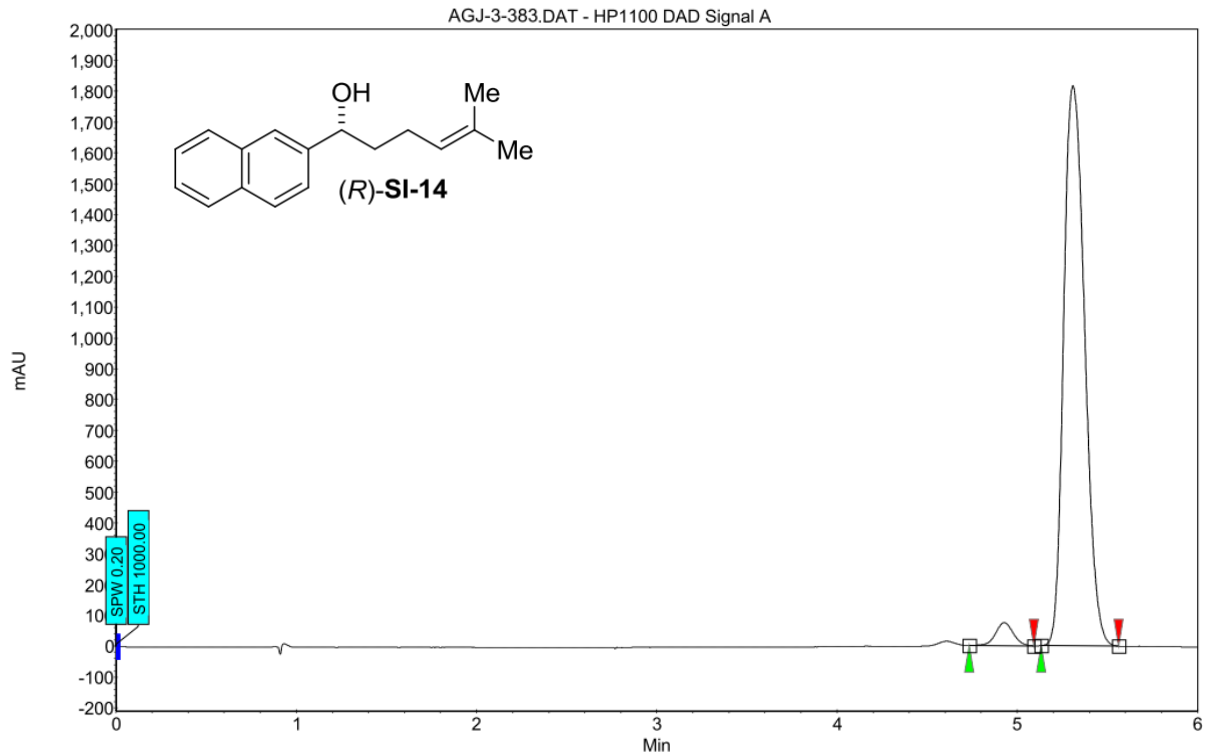
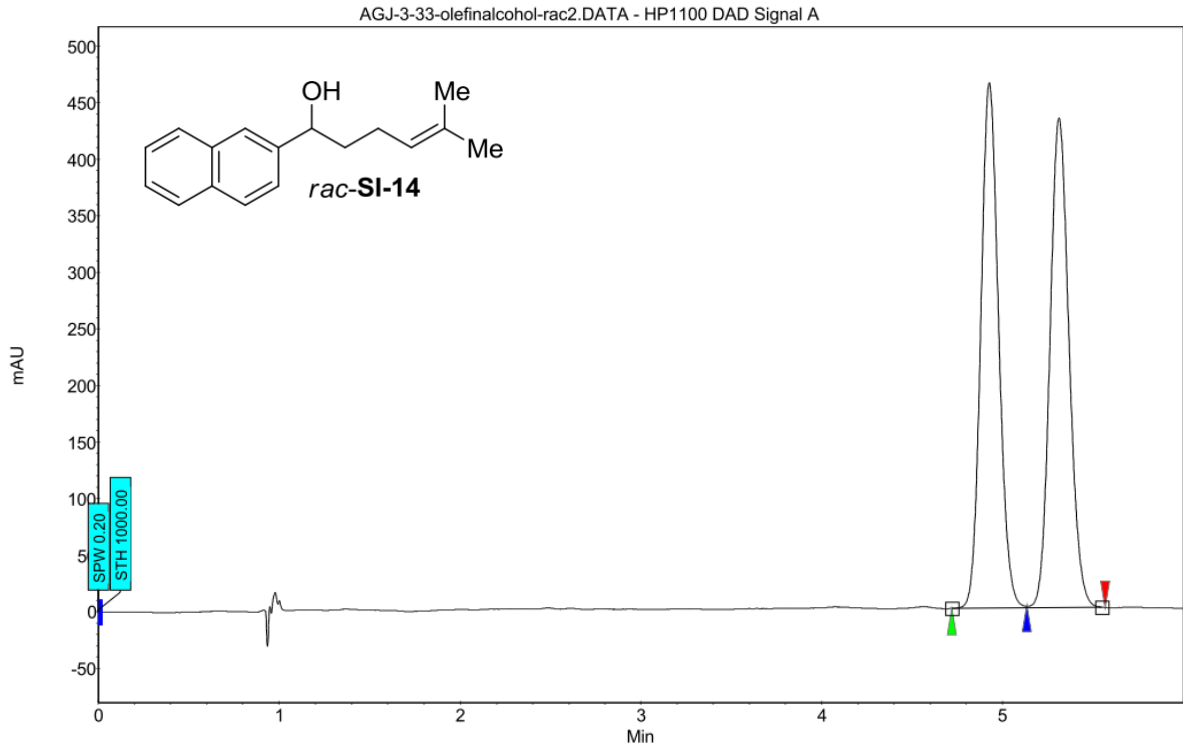
IMY-V-01612.DATA - HP1100 DAD Signal A



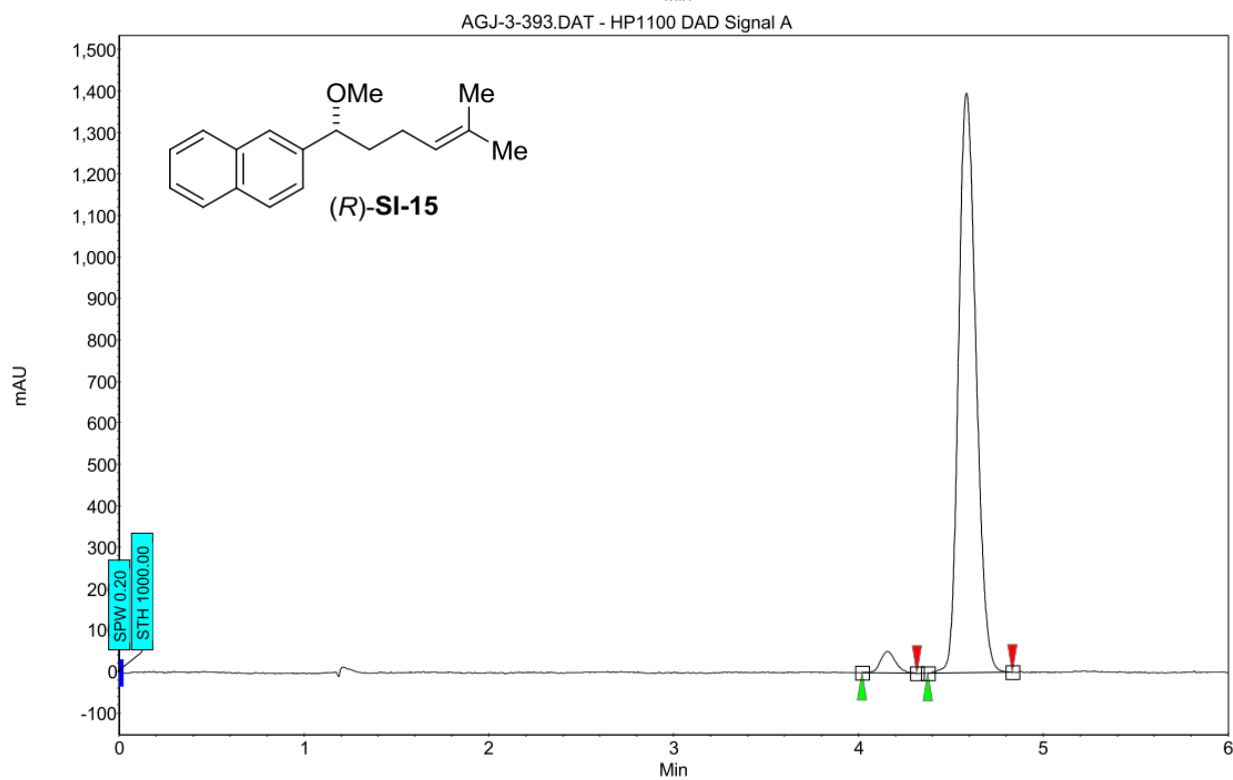
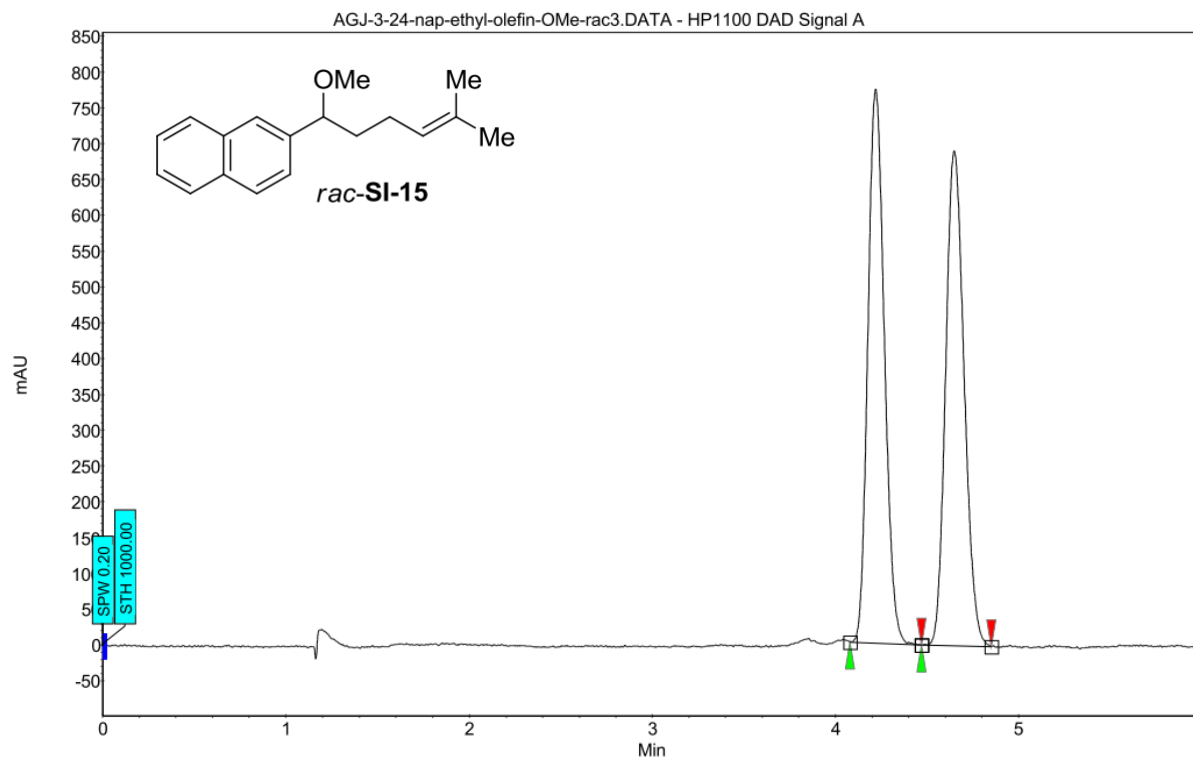
Index	Name	Start	Time	End	RT Offset	Quantity	Height	Area	Area
		[Min]	[Min]	[Min]	[Min]	[% Area]	[μ V]	[μ V.Min]	[%]
1	UNKNOWN	6.30	6.56	7.07	0.00	99.26	1178.6	215.7	99.260
2	UNKNOWN	7.41	7.64	7.93	0.00	0.74	8.1	1.6	0.740
Total						100.00	1186.7	217.3	100.000



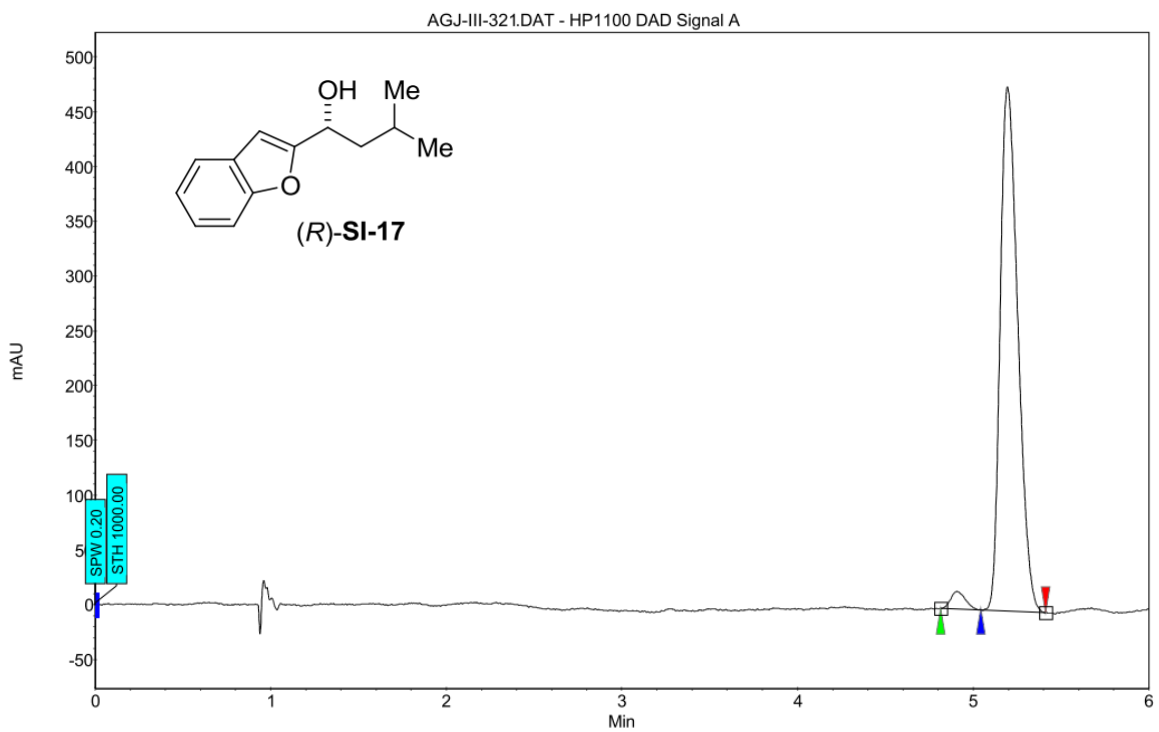
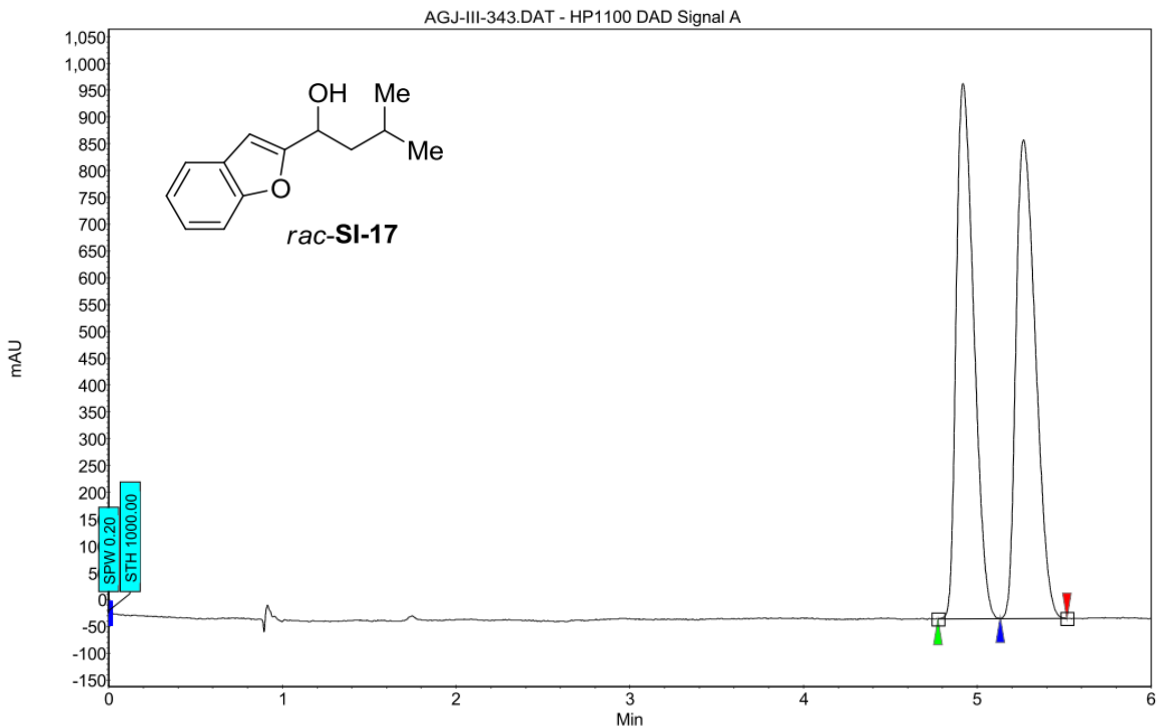
Index	Name	Start	Time	End	RT Offset	Quantity	Height	Area	Area
		[Min]	[Min]	[Min]	[Min]	[% Area]	[μ V]	[μ V.Min]	[%]
1	UNKNOWN	5.07	5.24	5.50	0.00	100.00	497.0	64.9	100.000
Total						100.00	497.0	64.9	100.000



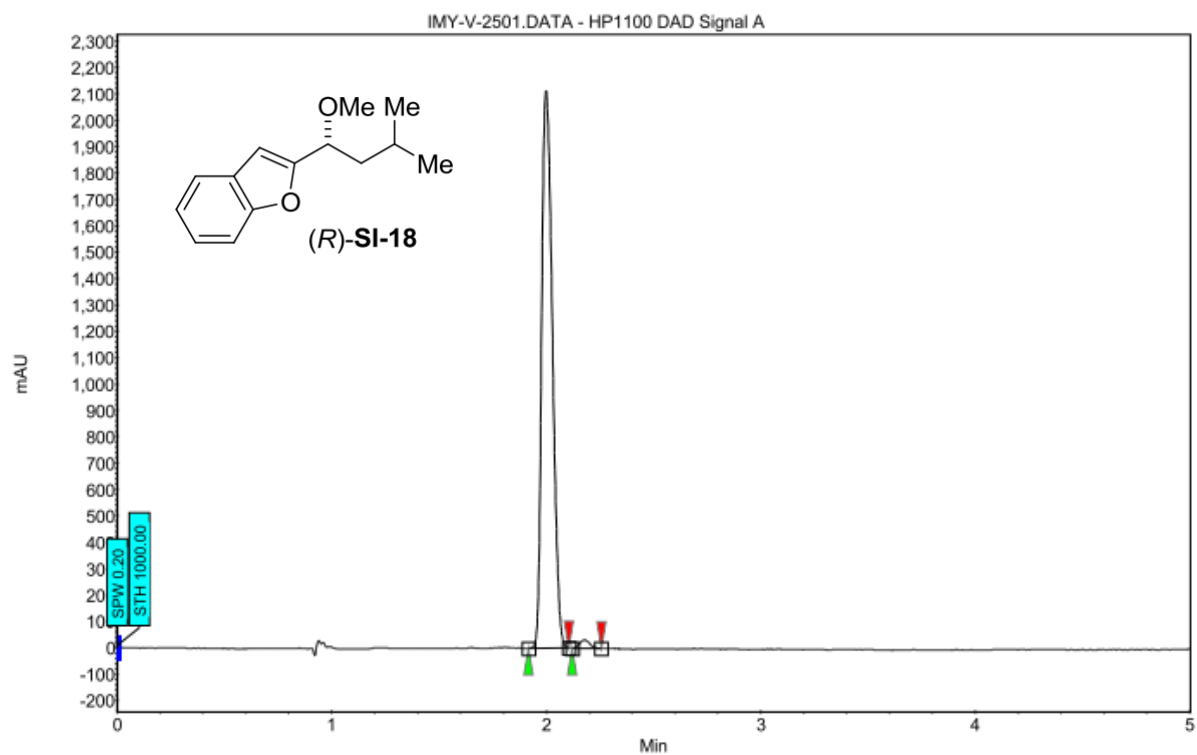
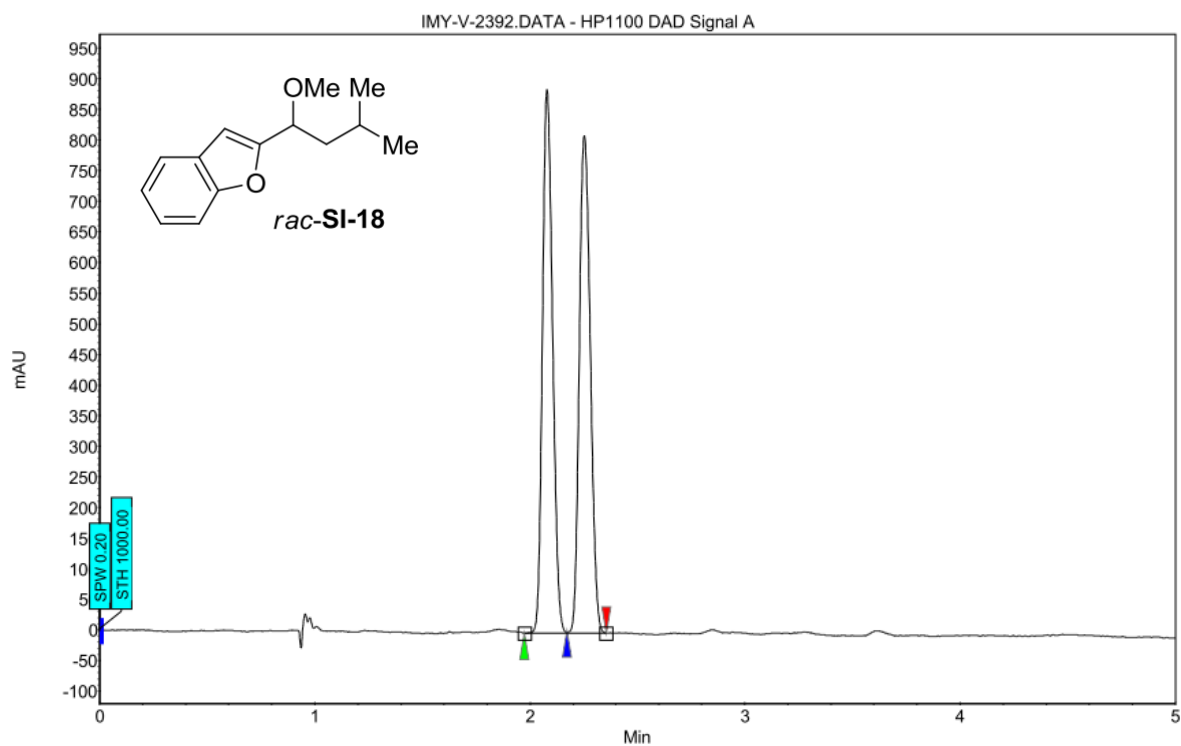
Index	Name	Start	Time	End	RT Offset	Quantity	Height	Area	Area
		[Min]	[Min]	[Min]	[Min]	[% Area]	[μ V]	[μ V.Min]	[%]
2	UNKNOWN	4.73	4.93	5.09	0.00	3.37	75.0	8.6	3.373
1	UNKNOWN	5.13	5.31	5.56	0.00	96.63	1816.9	245.2	96.627
Total						100.00	1891.9	253.8	100.000



Index	Name	Start	Time	End	RT Offset	Quantity	Height	Area	Area
		[Min]	[Min]	[Min]	[Min]	[% Area]	[μ V]	[μ V.Min]	[%]
2	UNKNOWN	4.02	4.16	4.32	0.00	3.08	51.8	4.9	3.080
1	UNKNOWN	4.37	4.58	4.83	0.00	96.92	1396.8	153.4	96.920
Total						100.00	1448.6	158.2	100.000

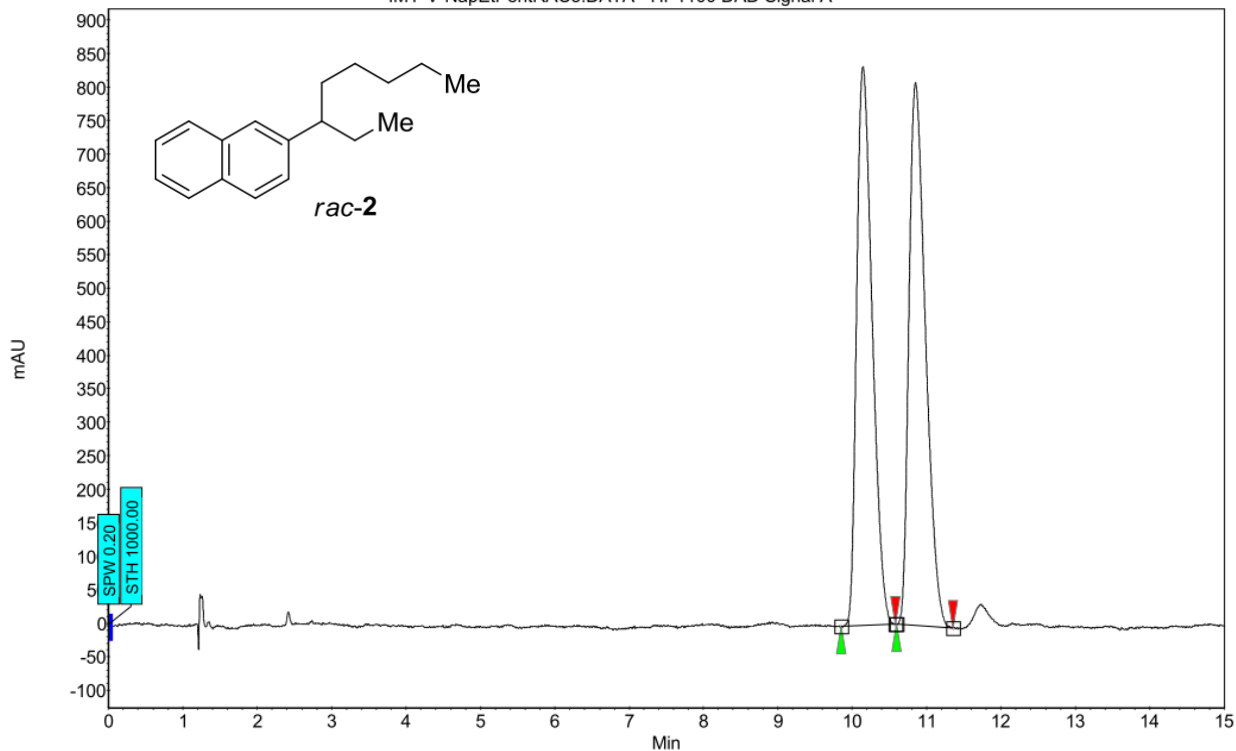


Index	Name	Start Time	End Time	RT Offset	Quantity	Height	Area	Area	
		[Min]	[Min]	[Min]	[% Area]	[μ V]	[μ V.Min]	[%]	
1	UNKNOWN	4.81	4.91	5.04	0.00	2.72	16.2	2.724	
2	UNKNOWN	5.04	5.20	5.41	0.00	97.28	478.6	55.8	
Total						100.00	494.7	57.4	100.000

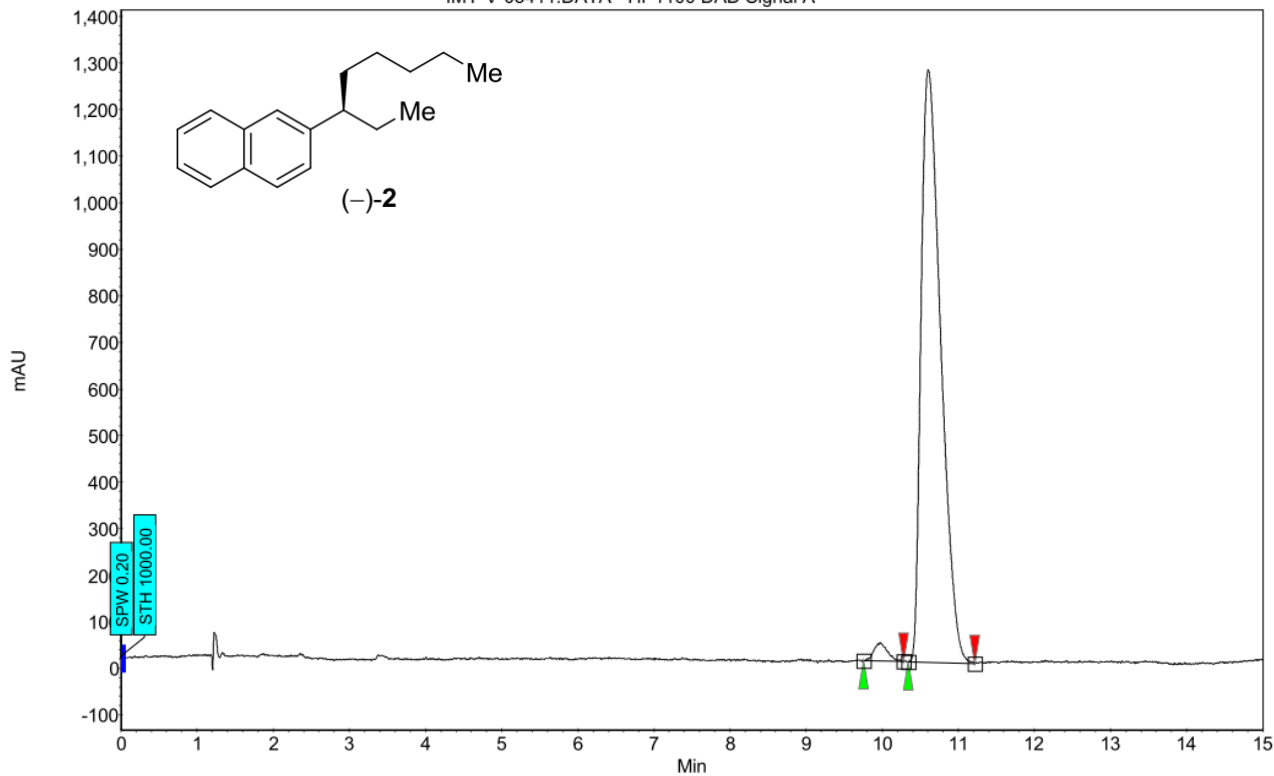


Index	Name	Start Time [Min]	Time [Min]	End [Min]	RT Offset [Min]	Quantity [% Area]	Height [μV]	Area [μV.Min]	Area [%]
1	UNKNOWN	1.92	2.00	2.10	0.00	98.61	2113.6	121.9	98.613
2	UNKNOWN	2.12	2.18	2.26	0.00	1.39	31.1	1.7	1.387
Total						100.00	2144.6	123.6	100.000

IMY-V-NapEtPentRAC5.DATA - HP1100 DAD Signal A

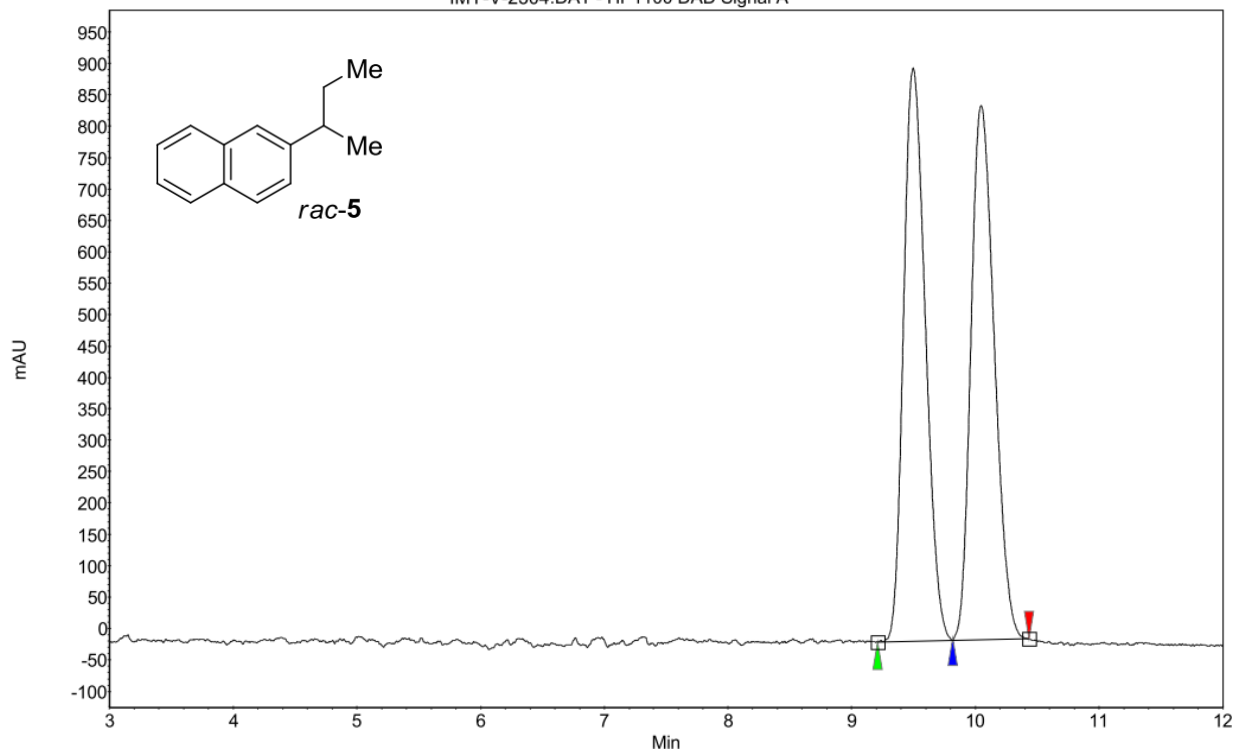


IMY-V-05411.DATA - HP1100 DAD Signal A

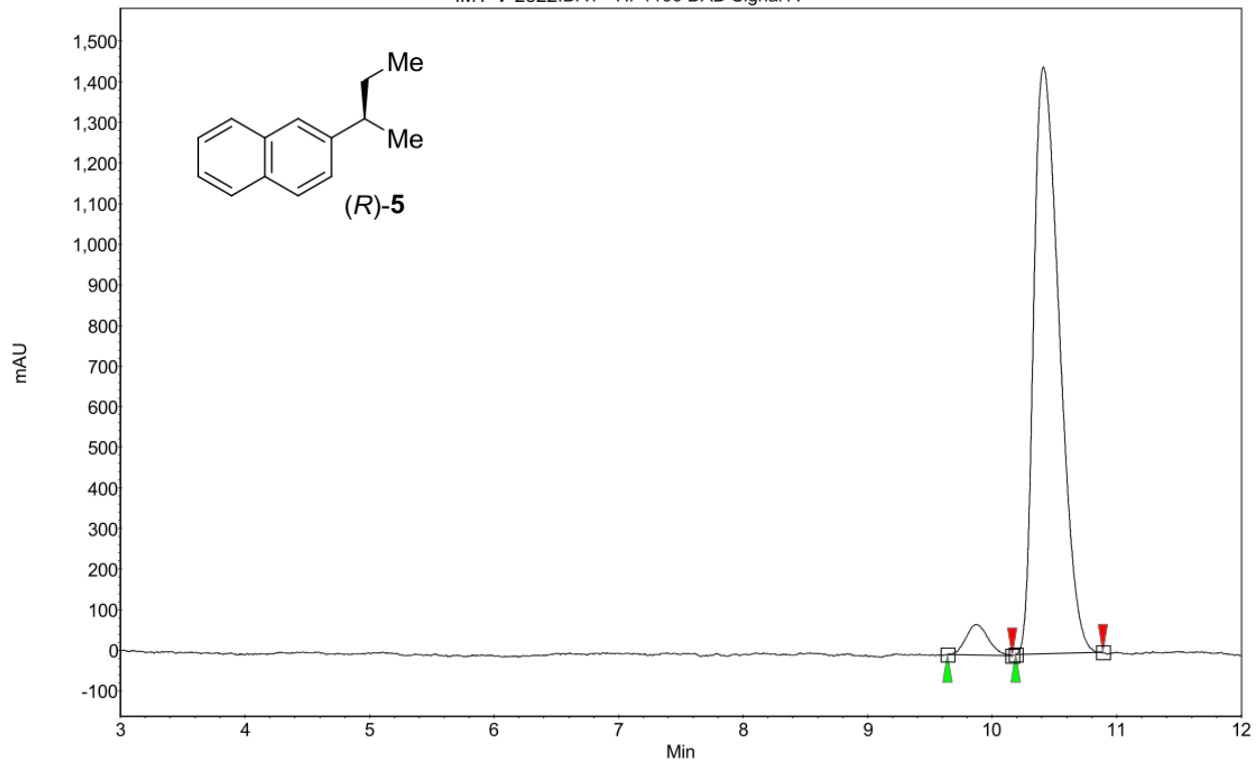


Index	Name	Start	Time	End	RT Offset	Quantity	Height	Area	Area
		[Min]	[Min]	[Min]	[Min]	[% Area]	[μ V]	[μ V.Min]	[%]
1	UNKNOWN	9.76	9.97	10.28	0.00	1.97	38.7	7.7	1.972
2	UNKNOWN	10.34	10.61	11.22	0.00	98.03	1272.5	380.8	98.028
Total						100.00	1311.2	388.5	100.000

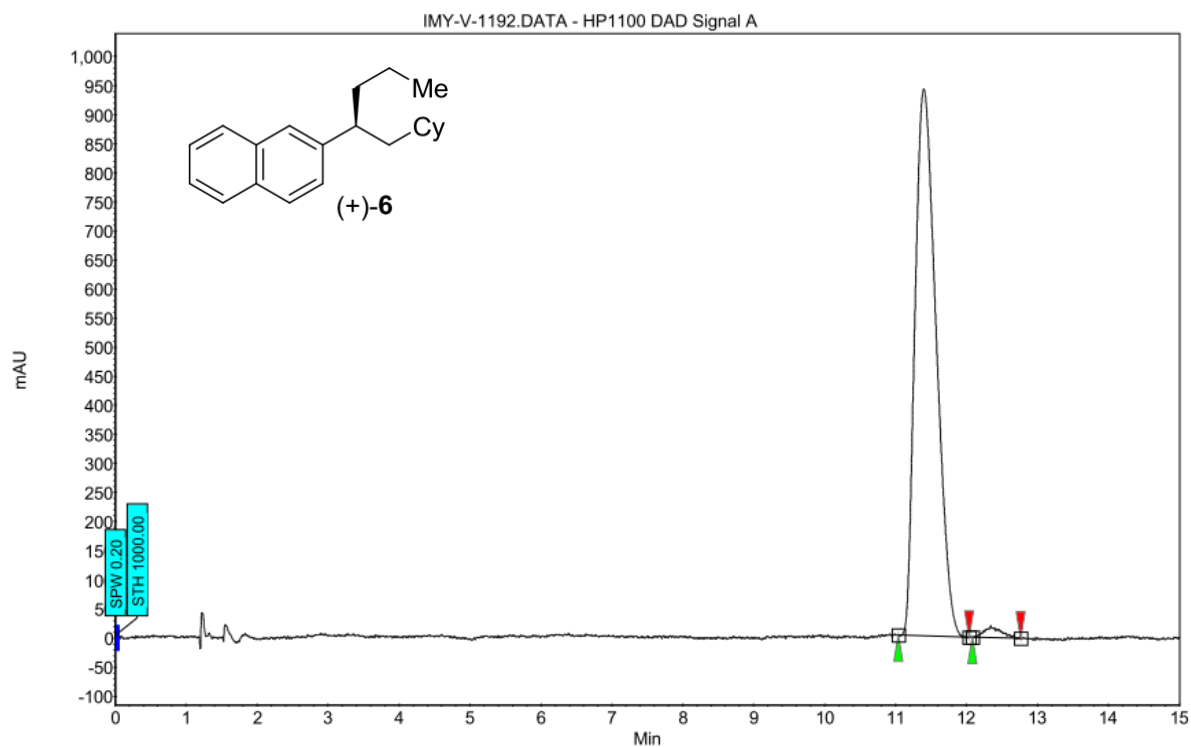
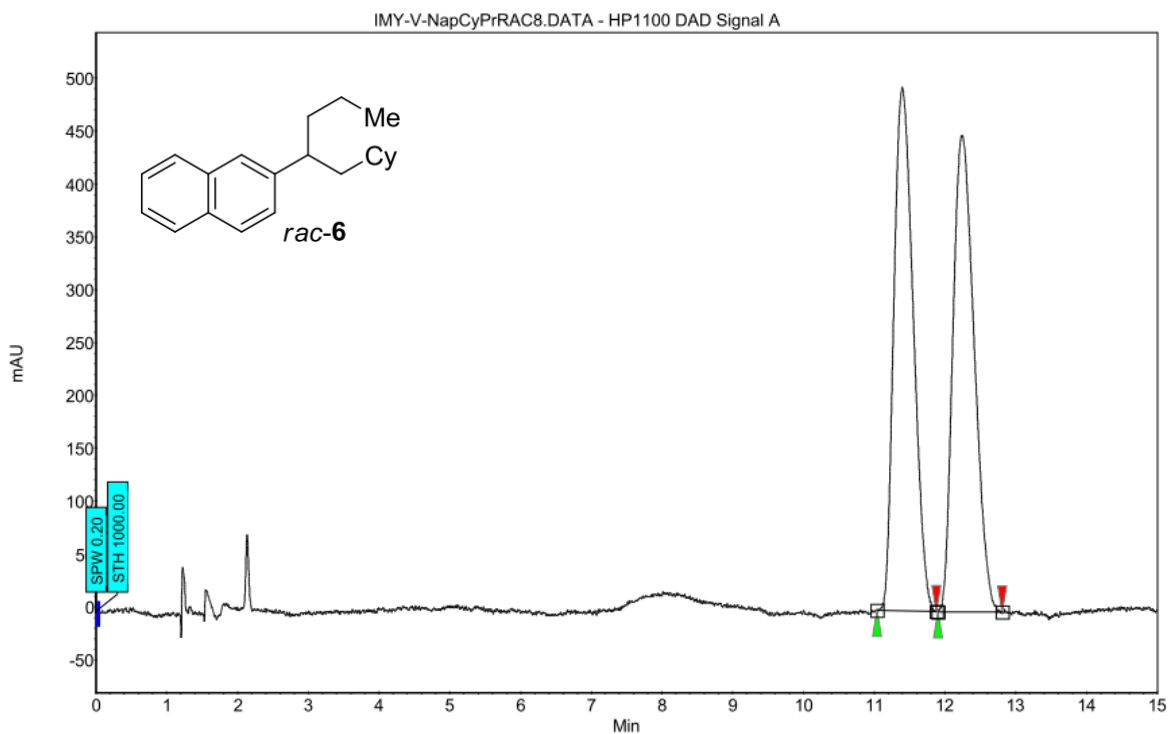
IMY-V-2304.DAT - HP1100 DAD Signal A



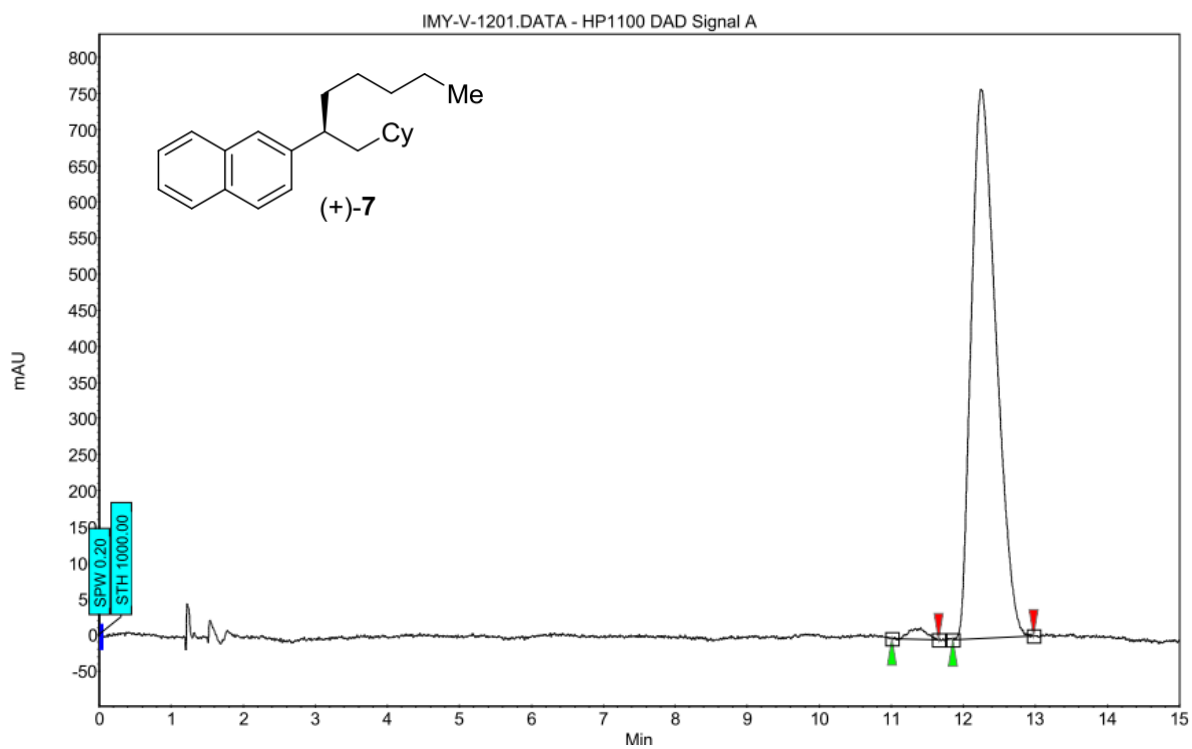
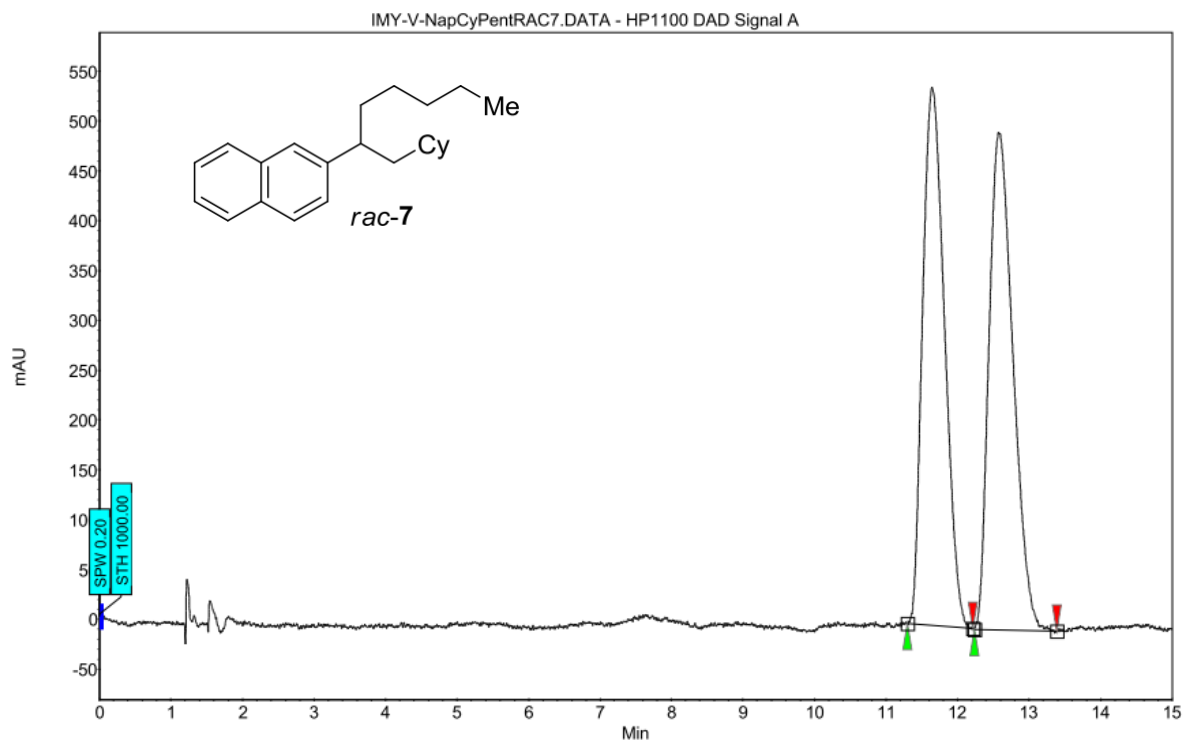
IMY-V-2322.DAT - HP1100 DAD Signal A



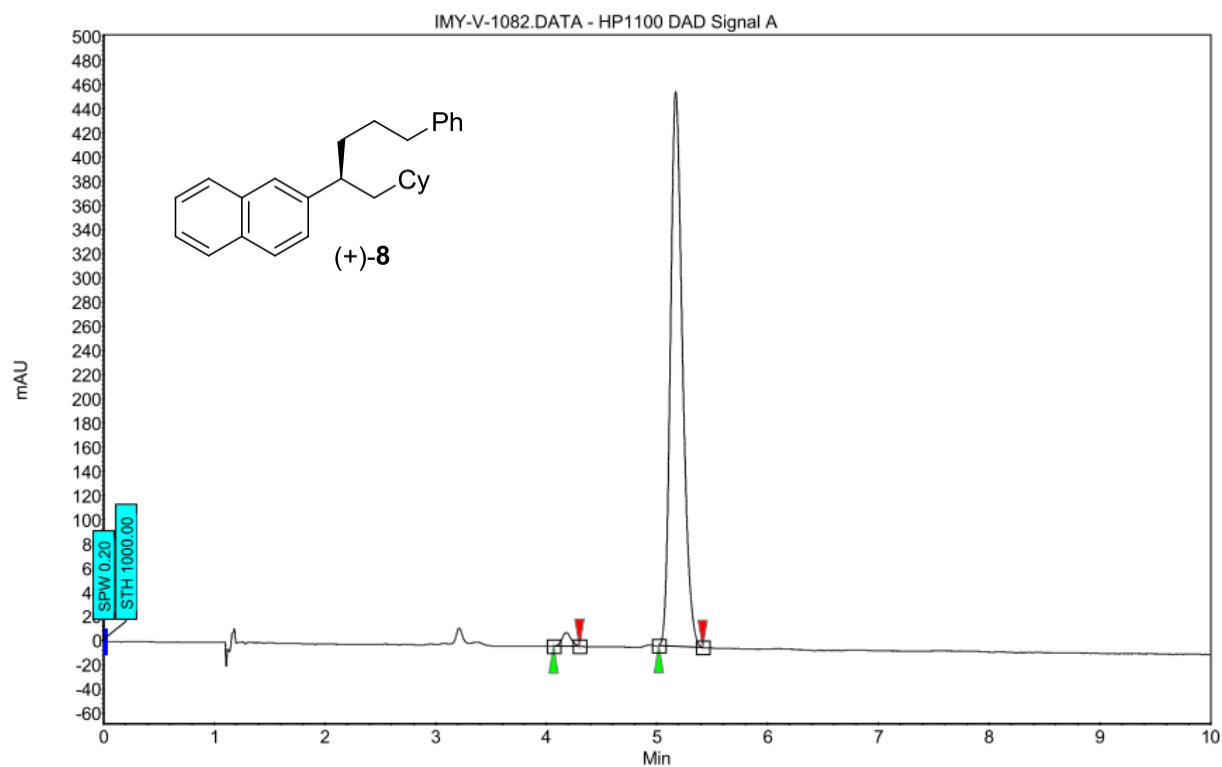
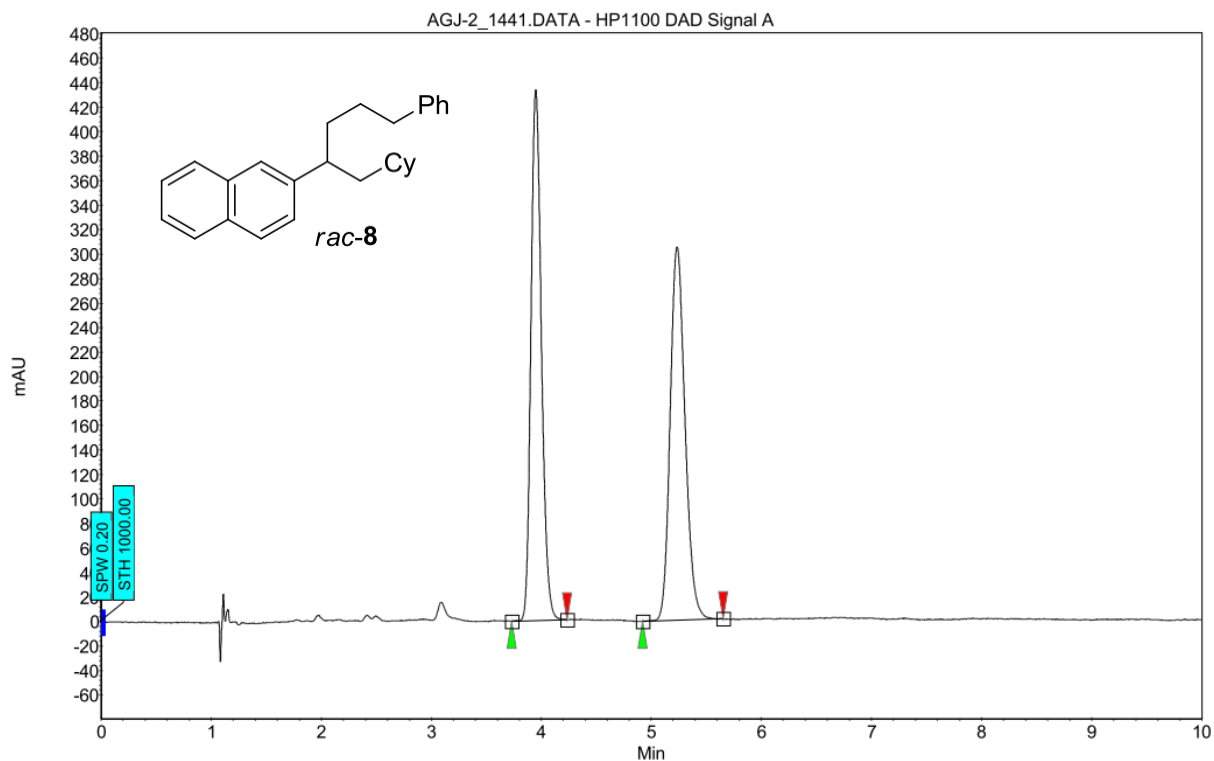
Index	Name	Start	Time	End	RT Offset	Quantity	Height	Area	Area
		[Min]	[Min]	[Min]	[Min]	[% Area]	[μ V]	[μ V.Min]	[%]
1	UNKNOWN	9.64	9.88	10.16	0.00	4.29	75.2	15.4	4.287
2	UNKNOWN	10.19	10.41	10.89	0.00	95.71	1445.1	344.9	95.713
Total						100.00	1520.4	360.4	100.000



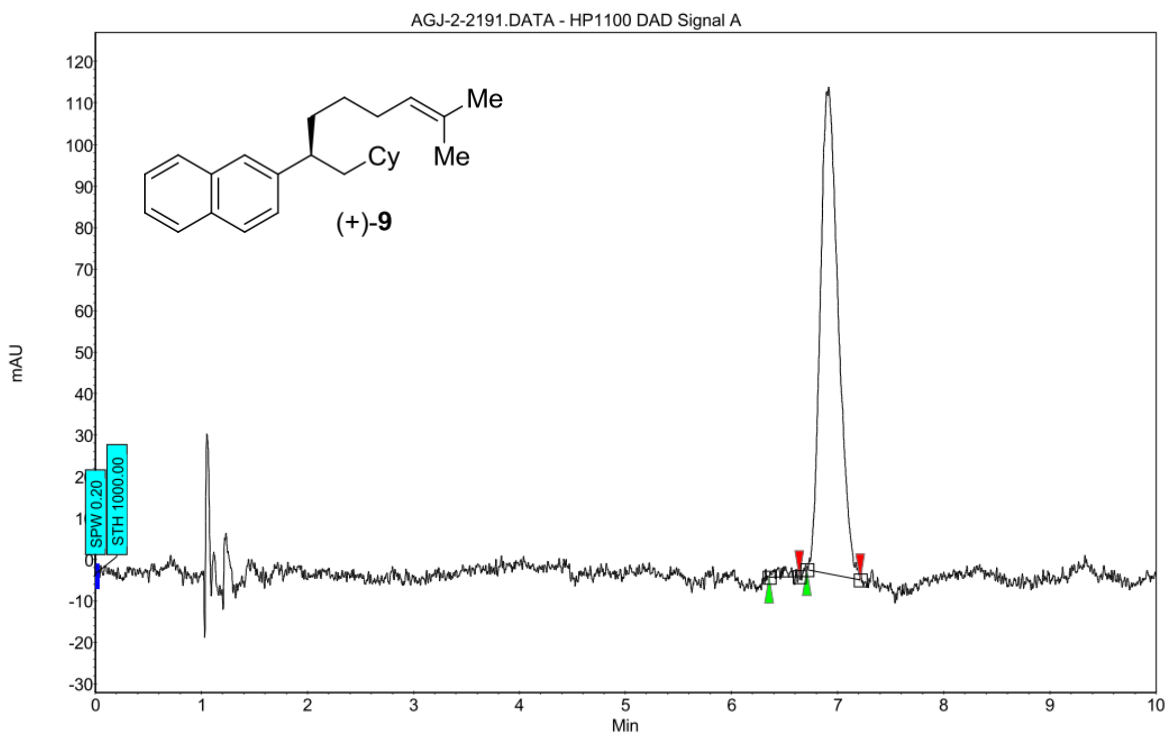
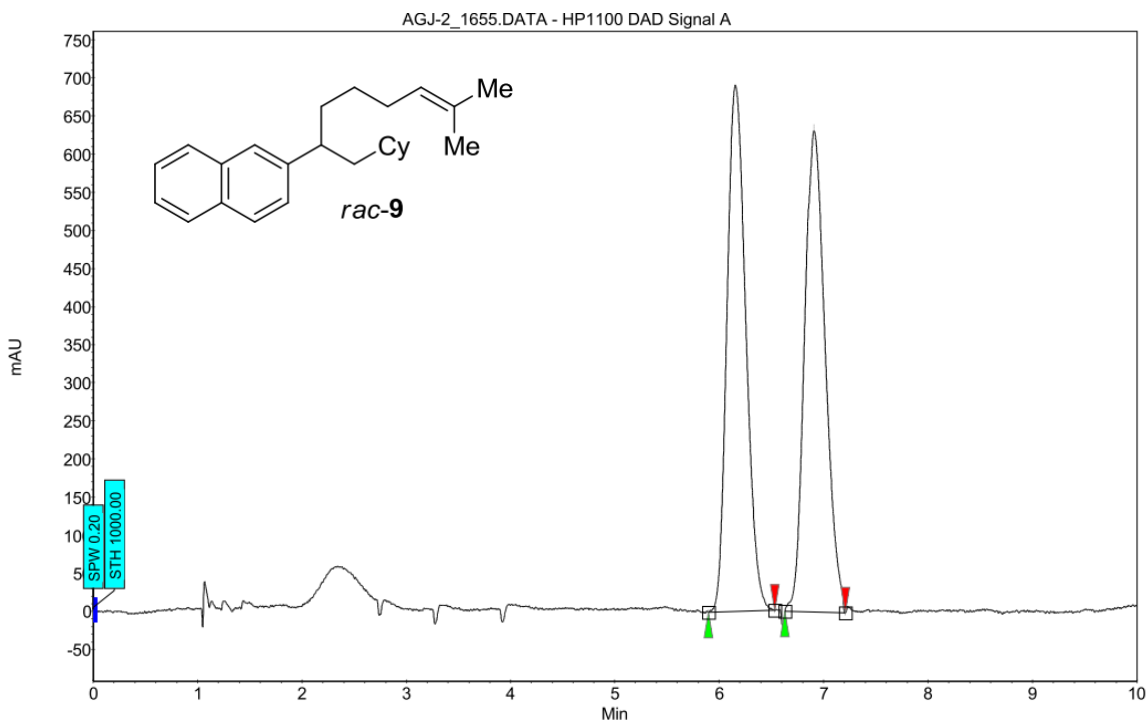
Index	Name	Start	Time	End	RT Offset	Quantity	Height	Area	Area
		[Min]	[Min]	[Min]	[Min]	[% Area]	[μ V]	[μ V.Min]	[%]
1	UNKNOWN	11.04	11.40	12.03	0.00	98.37	939.8	323.2	98.367
2	UNKNOWN	12.08	12.34	12.76	0.00	1.63	20.8	5.4	1.633
Total						100.00	960.6	328.6	100.000



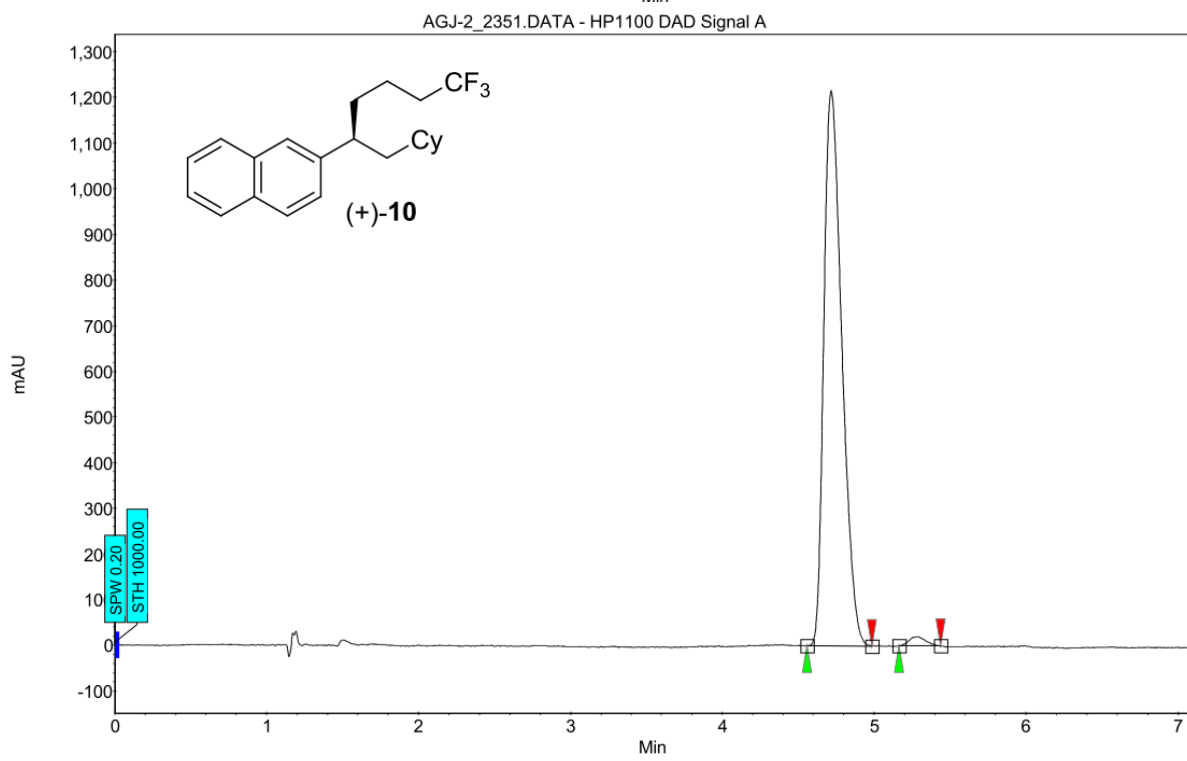
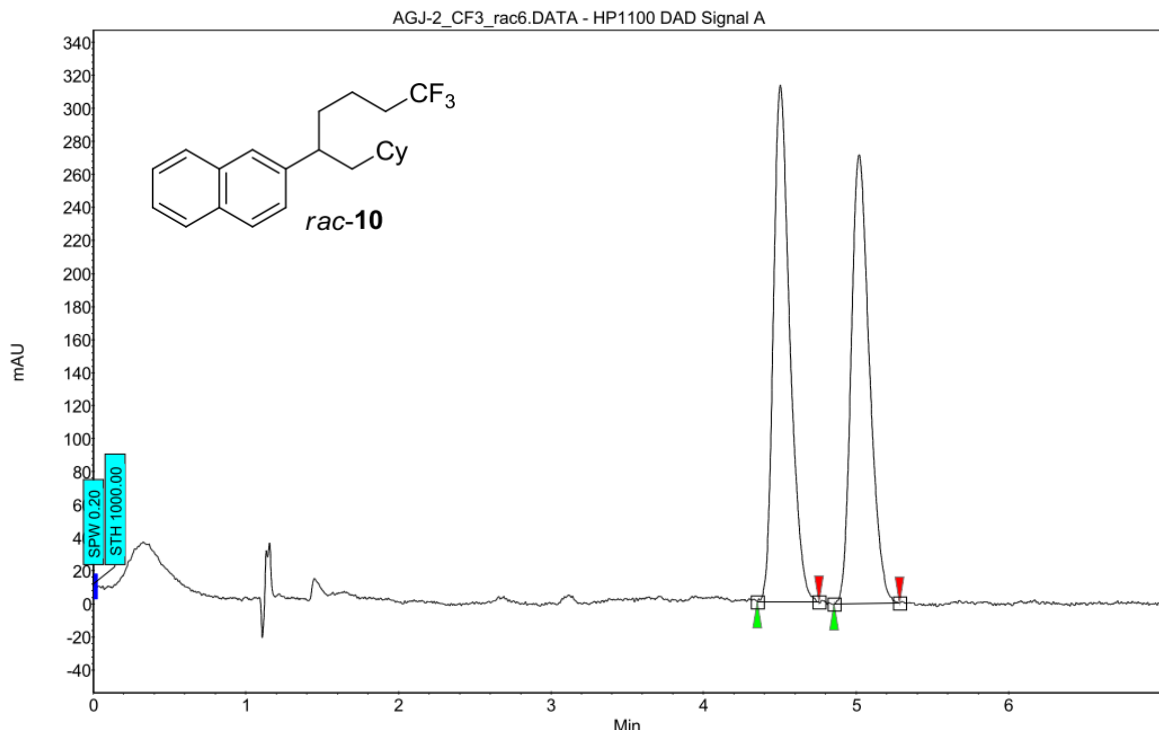
Index	Name	Start	Time	End	RT Offset	Quantity	Height	Area	Area
		[Min]	[Min]	[Min]	[Min]	[% Area]	[μ V]	[μ V.Min]	[%]
2	UNKNOWN	11.01	11.41	11.66	0.00	1.44	16.2	4.4	1.435
1	UNKNOWN	11.85	12.25	12.98	0.00	98.56	759.8	301.9	98.565
Total						100.00	776.0	306.3	100.000



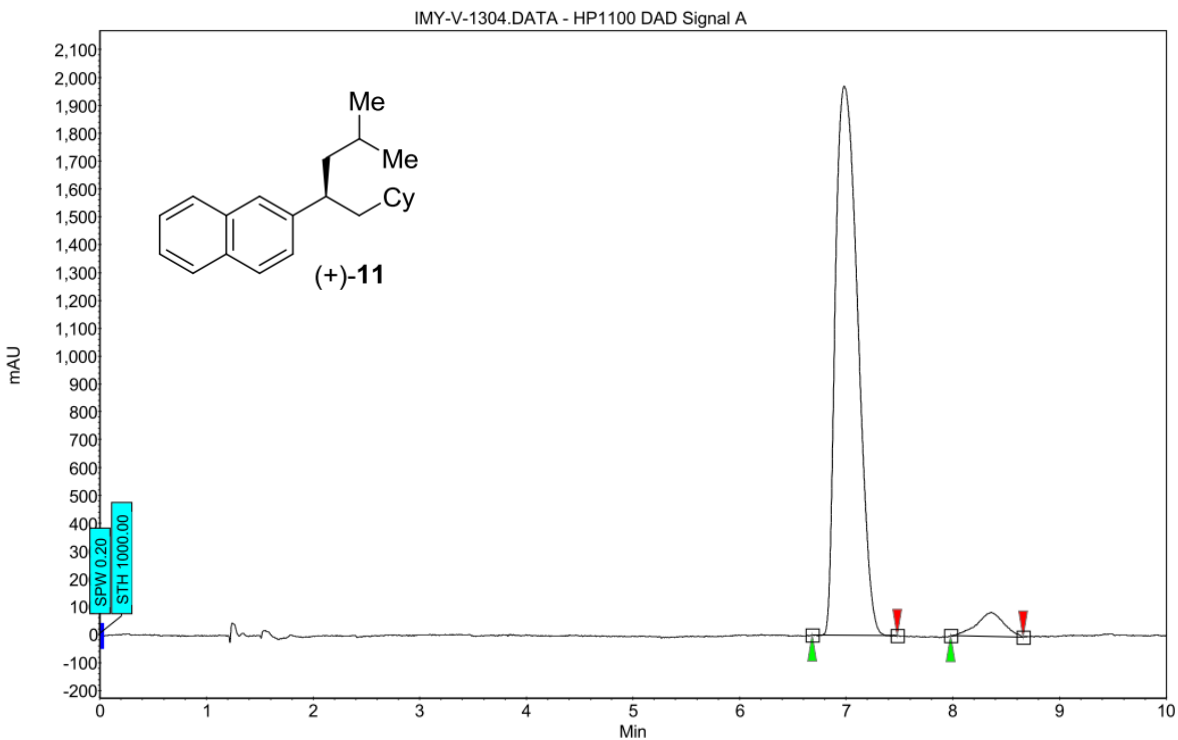
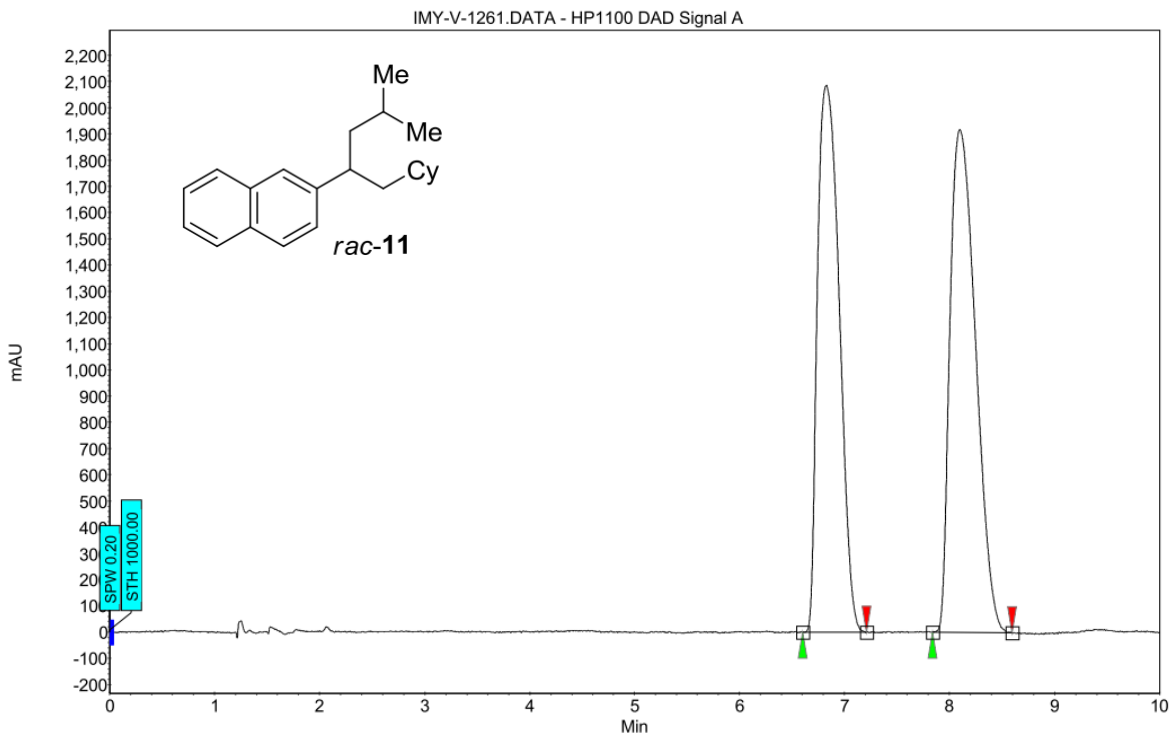
Index	Name	Start	Time	End	RT Offset	Quantity	Height	Area	Area
		[Min]	[Min]	[Min]	[Min]	[% Area]	[μ V]	[μ V.Min]	[%]
1	UNKNOWN	4.07	4.18	4.30	0.00	1.74	11.2	1.0	1.735
2	UNKNOWN	5.02	5.17	5.42	0.00	98.26	458.7	58.1	98.265
Total						100.00	469.9	59.1	100.000



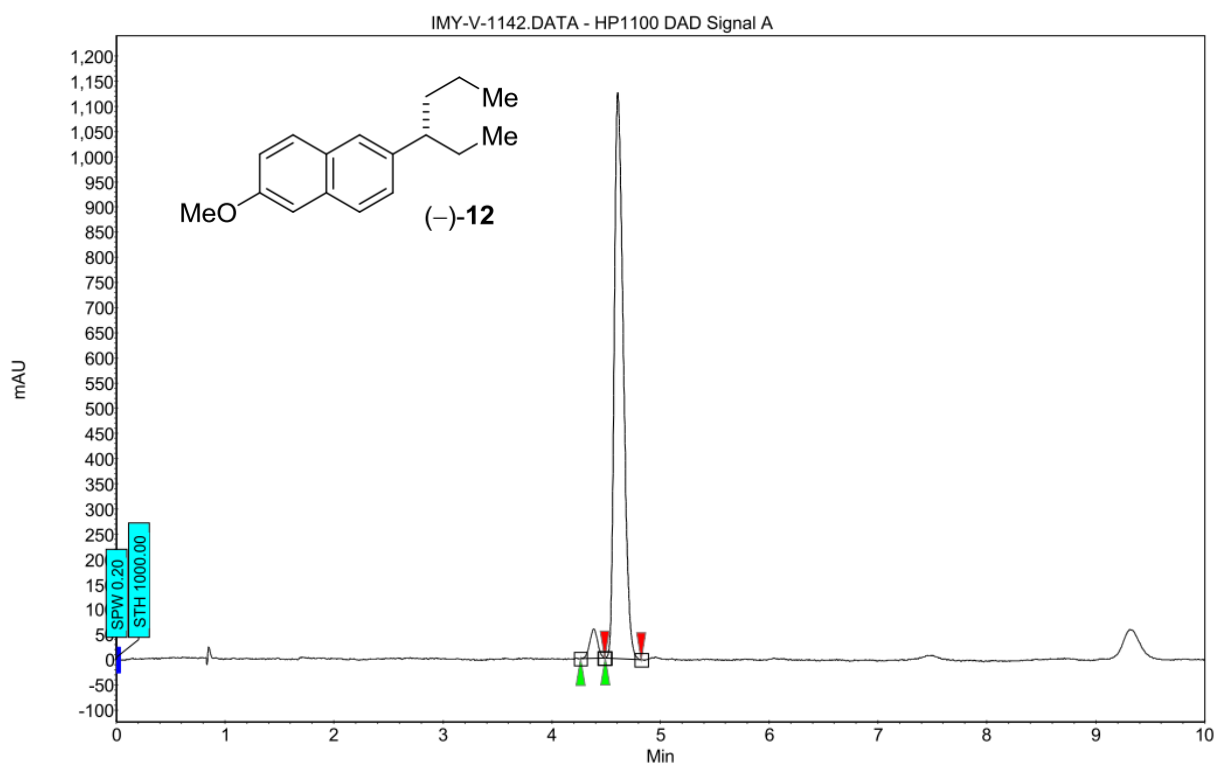
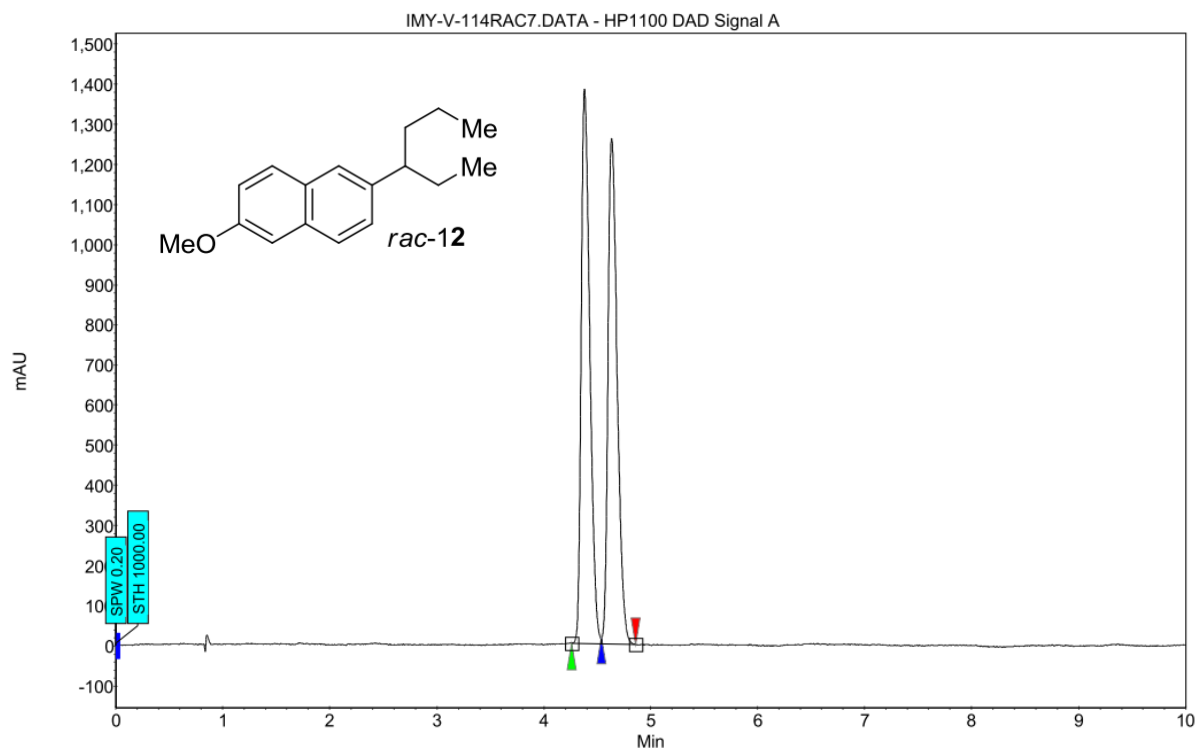
Index	Name	Start	Time	End	RT Offset	Quantity	Height	Area	Area
		[Min]	[Min]	[Min]	[Min]	[% Area]	[μ V]	[μ V.Min]	[%]
2	UNKNOWN	6.35	6.49	6.64	0.00	1.12	2.2	0.3	1.119
1	UNKNOWN	6.71	6.92	7.21	0.00	98.88	117.2	23.0	98.881
Total						100.00	119.4	23.3	100.000



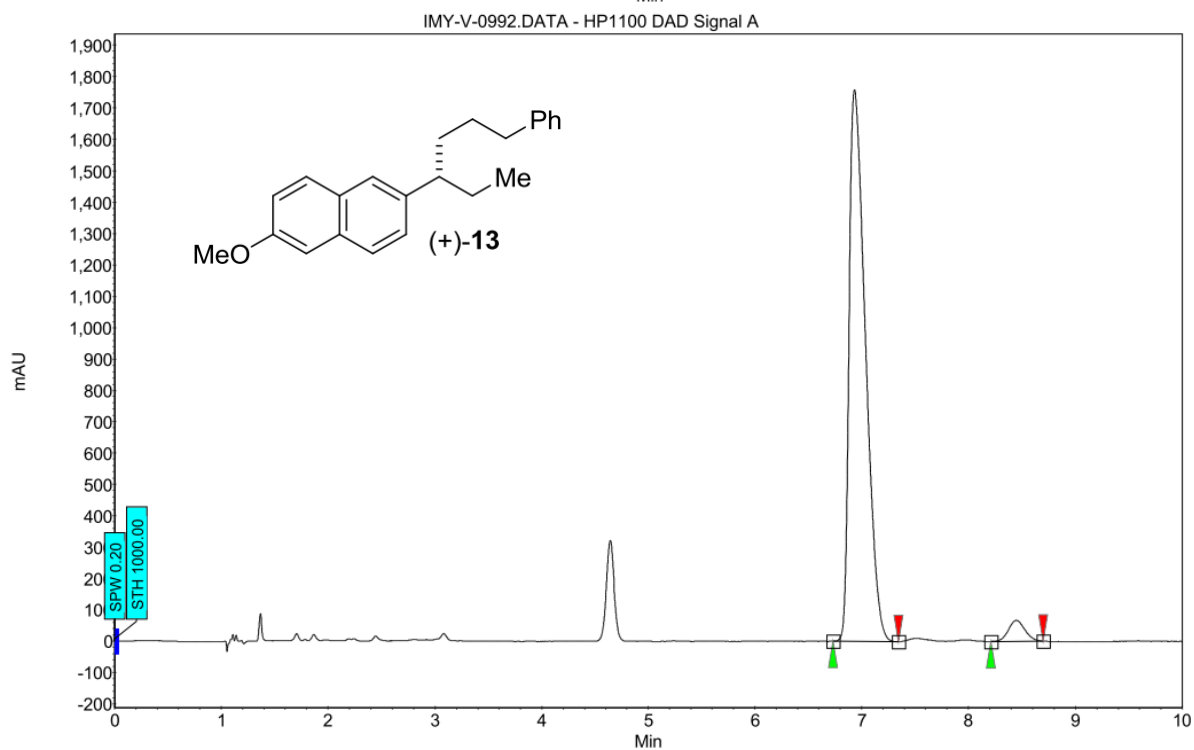
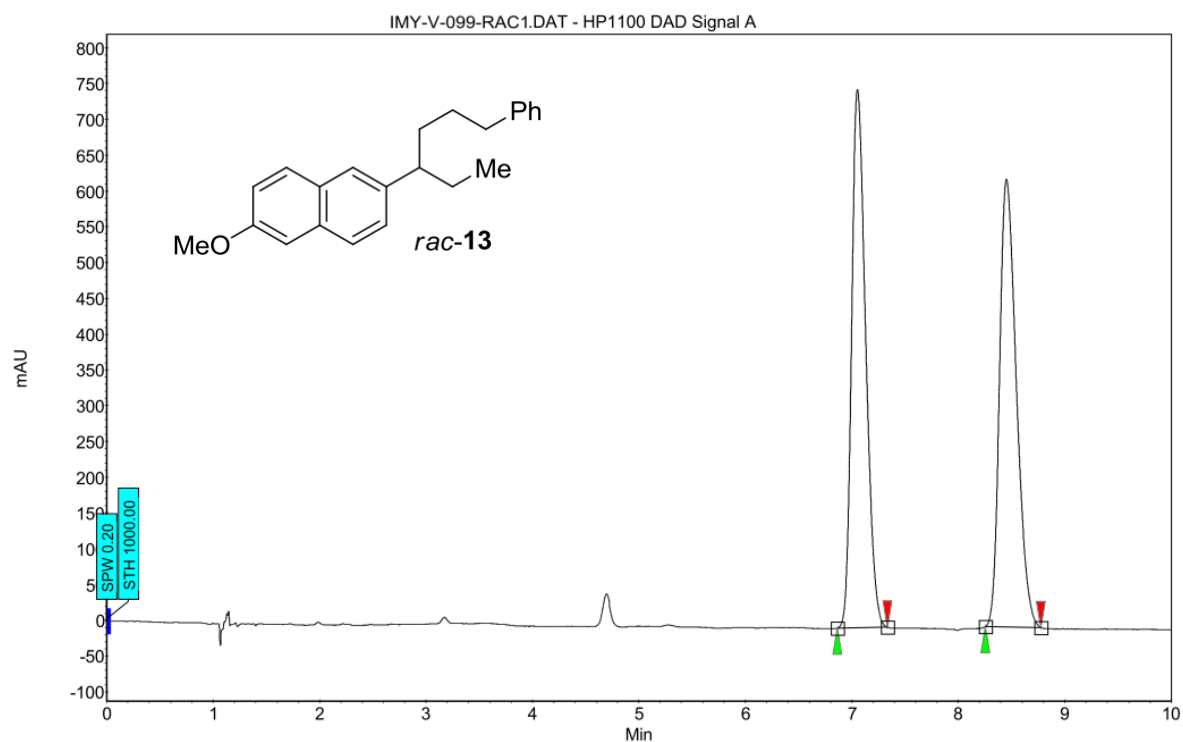
Index	Name	Start	Time	End	RT Offset	Quantity	Height	Area	Area
		[Min]	[Min]	[Min]	[Min]	[% Area]	[μ V]	[μ V.Min]	[%]
1	UNKNOWN	4.56	4.72	4.99	0.00	98.50	1215.7	167.4	98.504
2	UNKNOWN	5.16	5.29	5.44	0.00	1.50	20.2	2.5	1.496
Total						100.00	1235.9	169.9	100.000



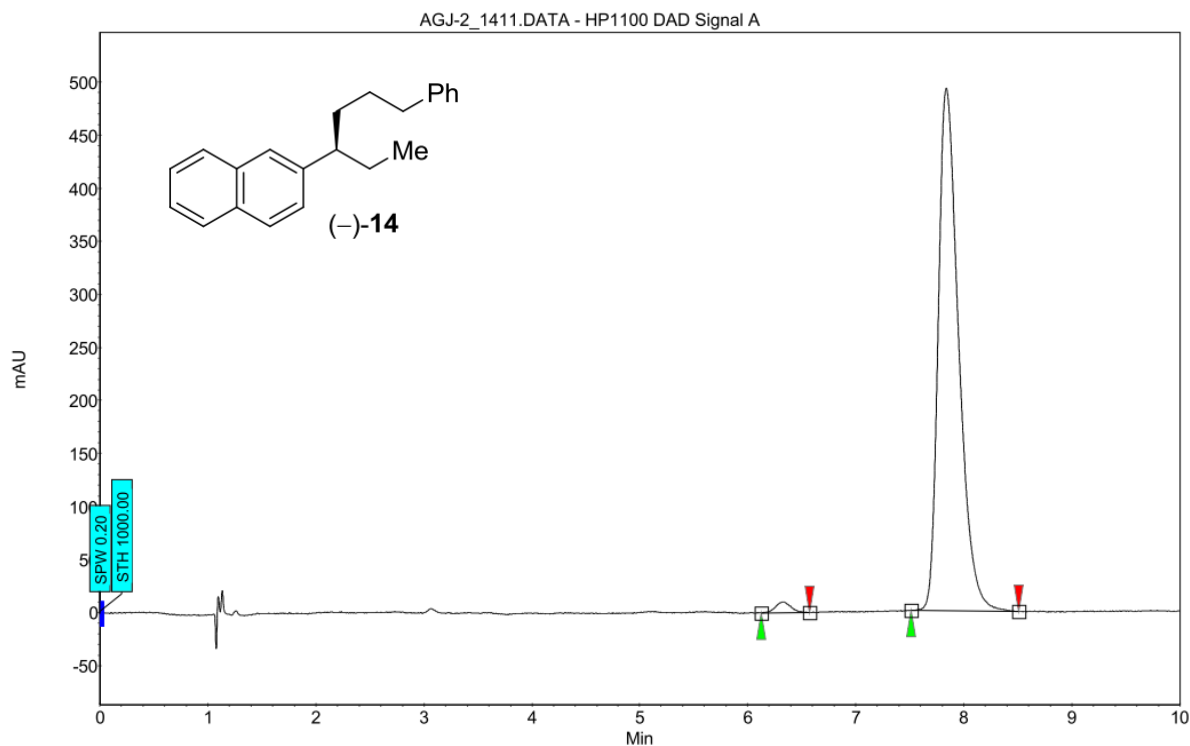
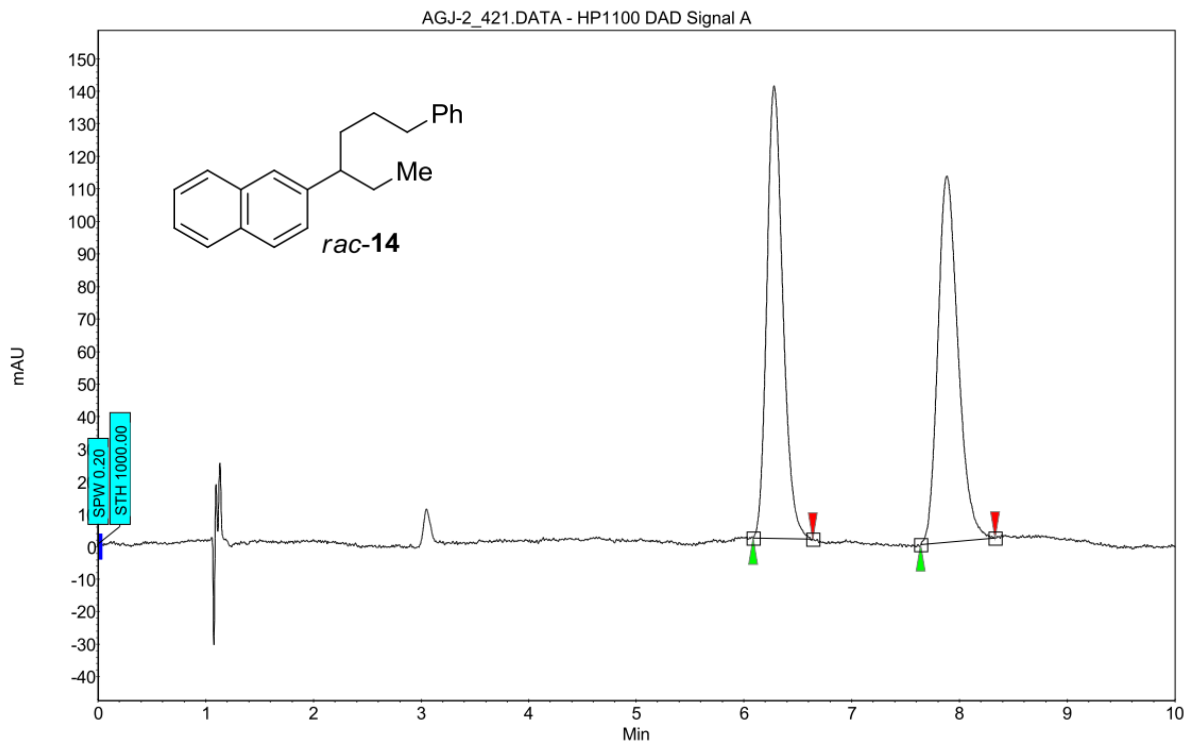
Index	Name	Start [Min]	Time [Min]	End [Min]	RT Offset [Min]	Quantity [% Area]	Height [μ V]	Area [μ V.Min]	Area [%]
1	UNKNOWN	6.68	6.98	7.48	0.00	94.98	1972.0	477.3	94.985
2	UNKNOWN	7.98	8.36	8.66	0.00	5.02	86.7	25.2	5.015
Total						100.00	2058.6	502.5	100.000



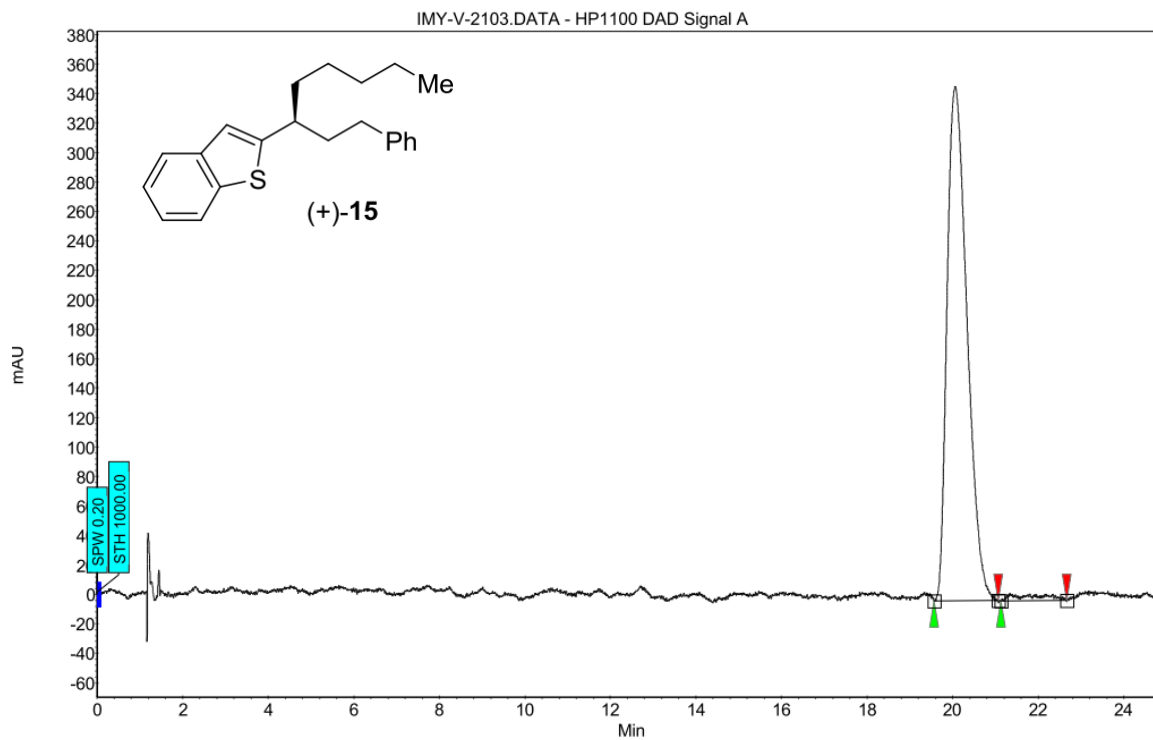
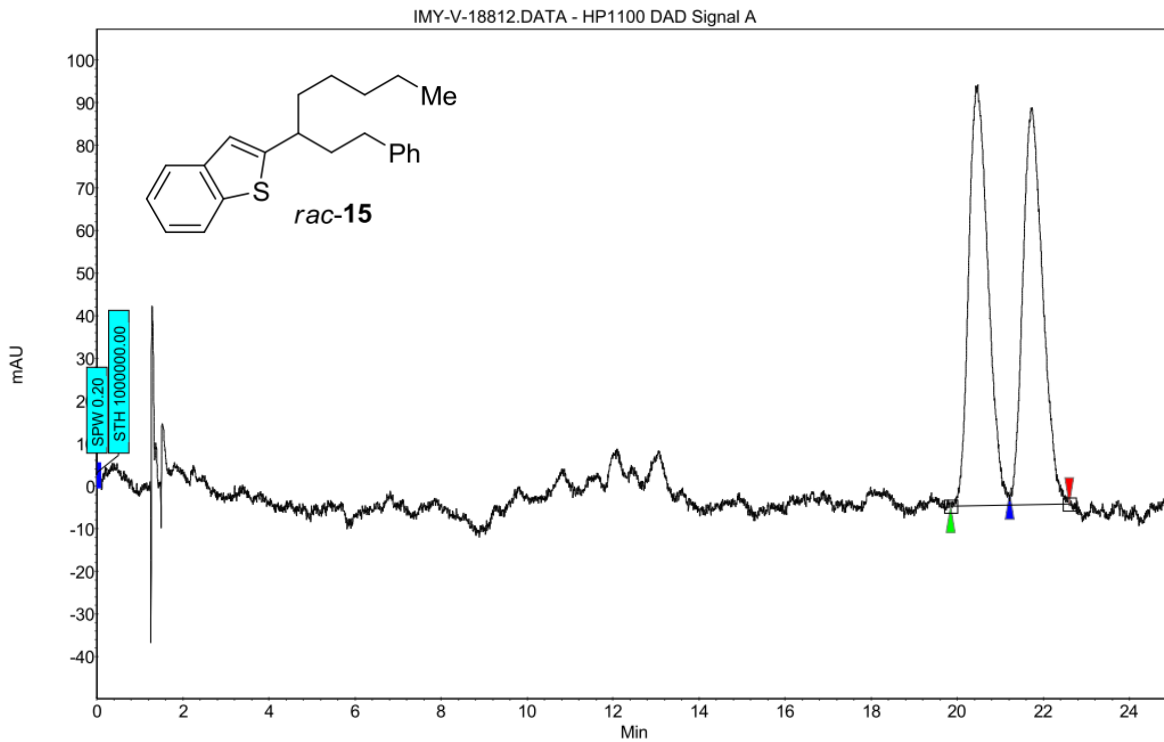
Index	Name	Start	Time	End	RT Offset	Quantity	Height	Area	Area
		[Min]	[Min]	[Min]	[Min]	[% Area]	[μ V]	[μ V.Min]	[%]
1	UNKNOWN	4.27	4.39	4.48	0.00	4.24	58.1	4.9	4.239
2	UNKNOWN	4.49	4.61	4.83	0.00	95.76	1124.7	110.8	95.761
Total						100.00	1182.8	115.7	100.000



Index	Name	Start	Time	End	RT Offset	Quantity	Height	Area	Area
		[Min]	[Min]	[Min]	[Min]	[% Area]	[μ V]	[μ V.Min]	[%]
1	UNKNOWN	6.73	6.93	7.34	0.00	96.27	1758.4	310.2	96.265
2	UNKNOWN	8.21	8.45	8.70	0.00	3.73	67.1	12.0	3.735
Total						100.00	1825.5	322.3	100.000

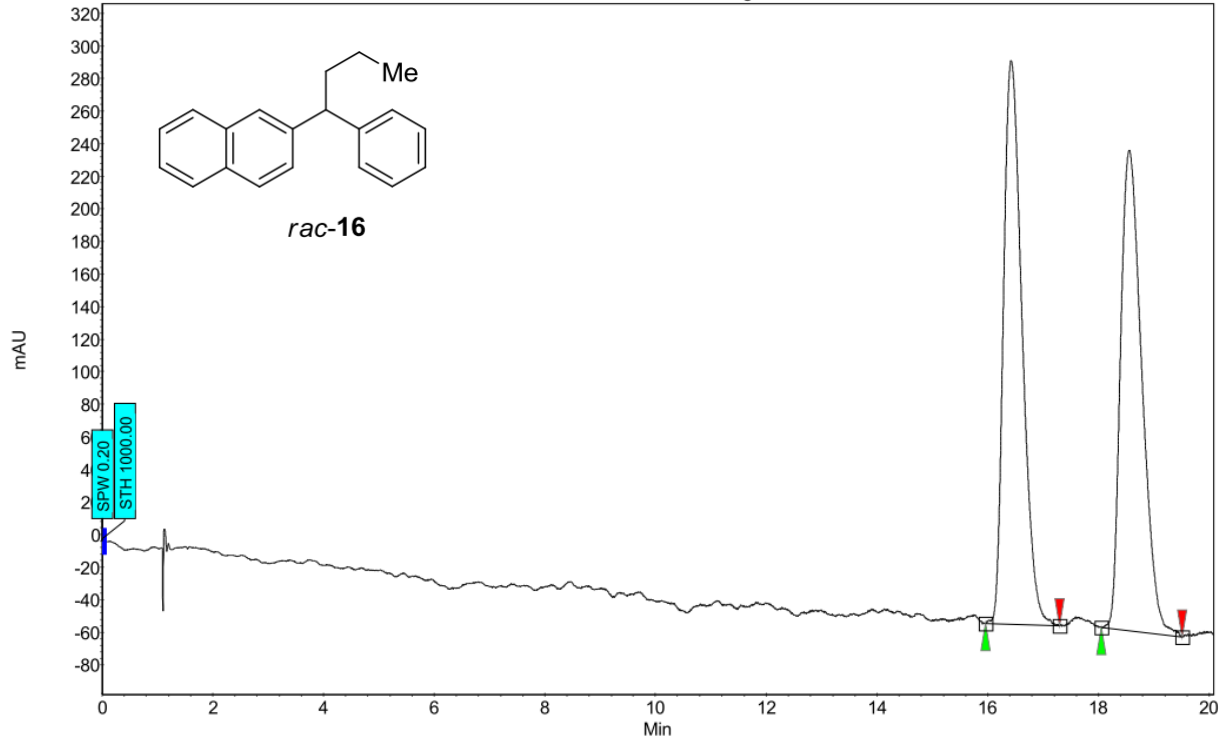


Index	Name	Start	Time	End	RT Offset	Quantity	Height	Area	Area
		[Min]	[Min]	[Min]	[Min]	[% Area]	[μ V]	[μ V.Min]	[%]
1	UNKNOWN	6.12	6.32	6.57	0.00	1.49	10.0	1.6	1.489
2	UNKNOWN	7.51	7.84	8.51	0.00	98.51	492.5	107.7	98.511
Total						100.00	502.5	109.3	100.000

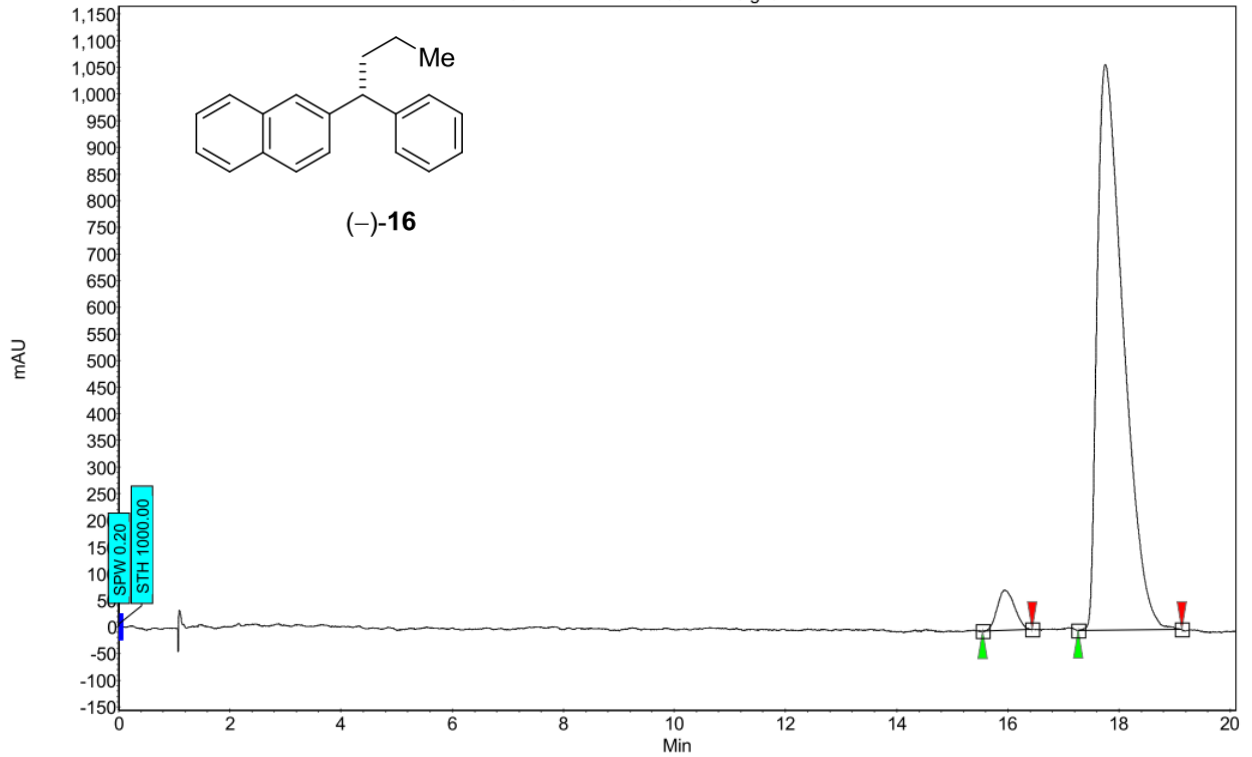


Index	Name	Start	Time	End	RT Offset	Quantity	Height	Area	Area
		[Min]	[Min]	[Min]	[Min]	[% Area]	[μ V]	[μ V.Min]	[%]
1	UNKNOWN	19.56	20.06	21.06	0.00	97.88	349.2	188.1	97.876
2	UNKNOWN	21.13	21.35	22.66	0.00	2.12	5.3	4.1	2.124
Total						100.00	354.5	192.2	100.000

IMY-V-143-rac1.DATA - HP1100 DAD Signal A

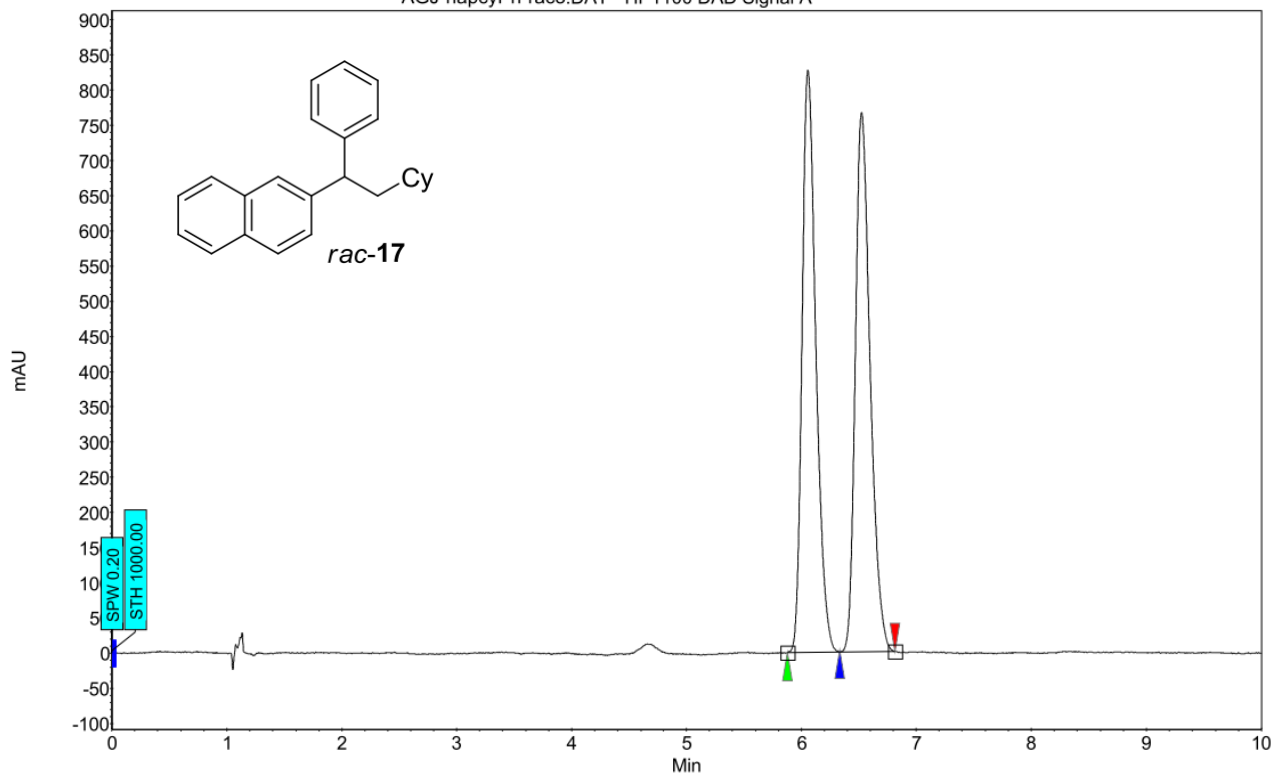


IMY-V-143111.DATA - HP1100 DAD Signal A

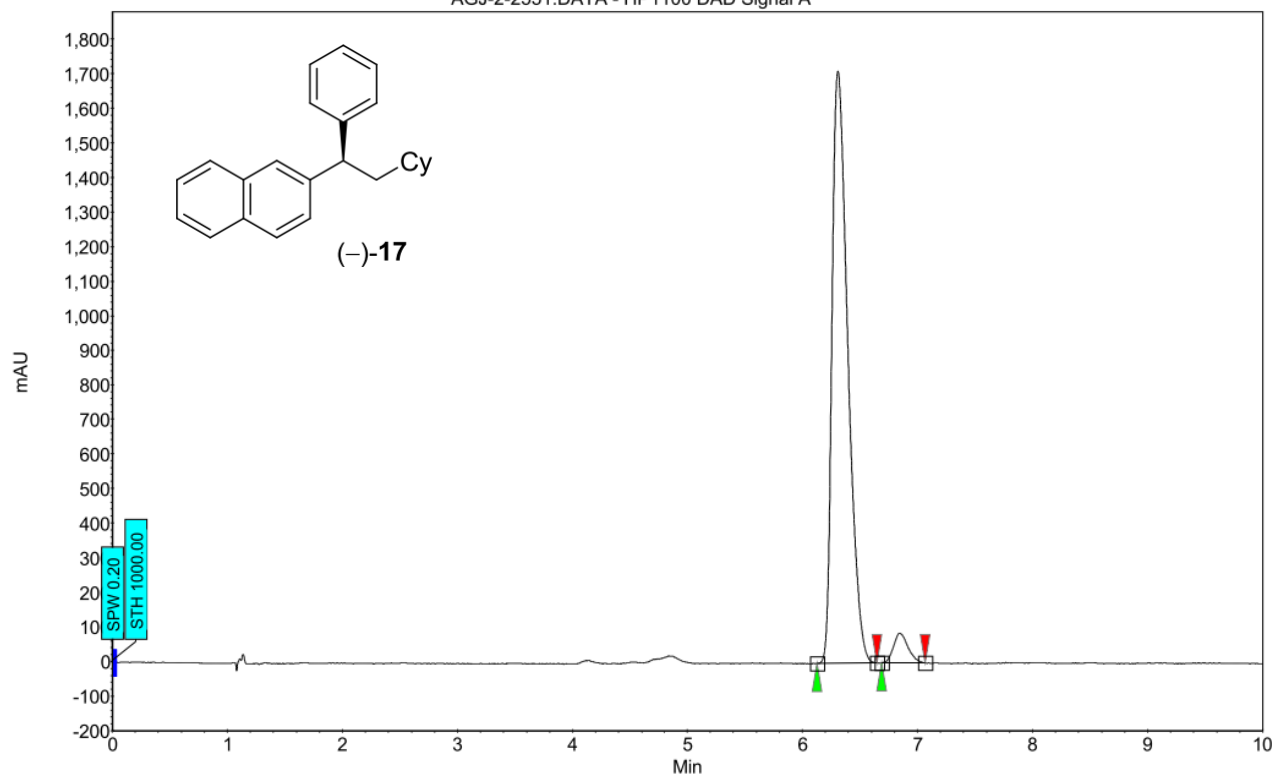


Index	Name	Start [Min]	Time [Min]	End [Min]	RT Offset [Min]	Quantity [% Area]	Height [μV]	Area [μV.Min]	Area [%]
1	UNKNOWN	15.55	15.95	16.44	0.00	4.46	75.3	27.0	4.458
2	UNKNOWN	17.27	17.75	19.13	0.00	95.54	1061.4	578.6	95.542
Total						100.00	1136.7	605.5	100.000

AGJ-napcyPh-rac3.DAT - HP1100 DAD Signal A

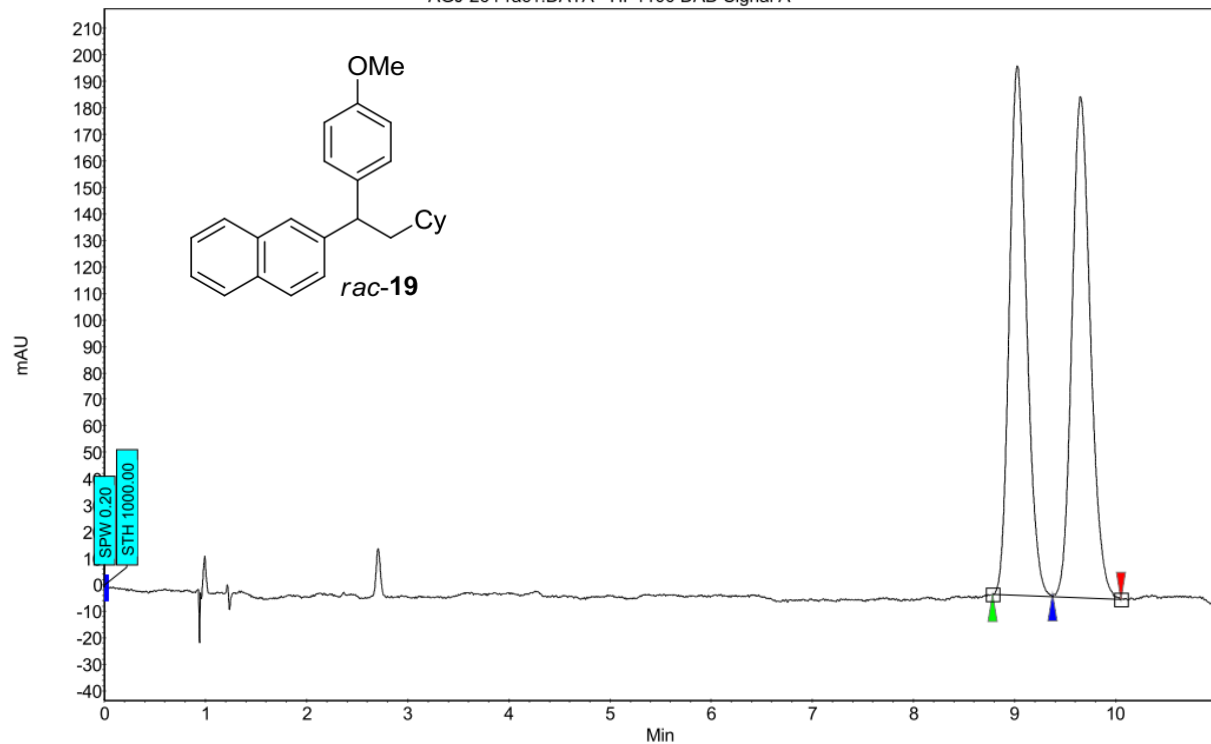


AGJ-2-2551.DATA - HP1100 DAD Signal A

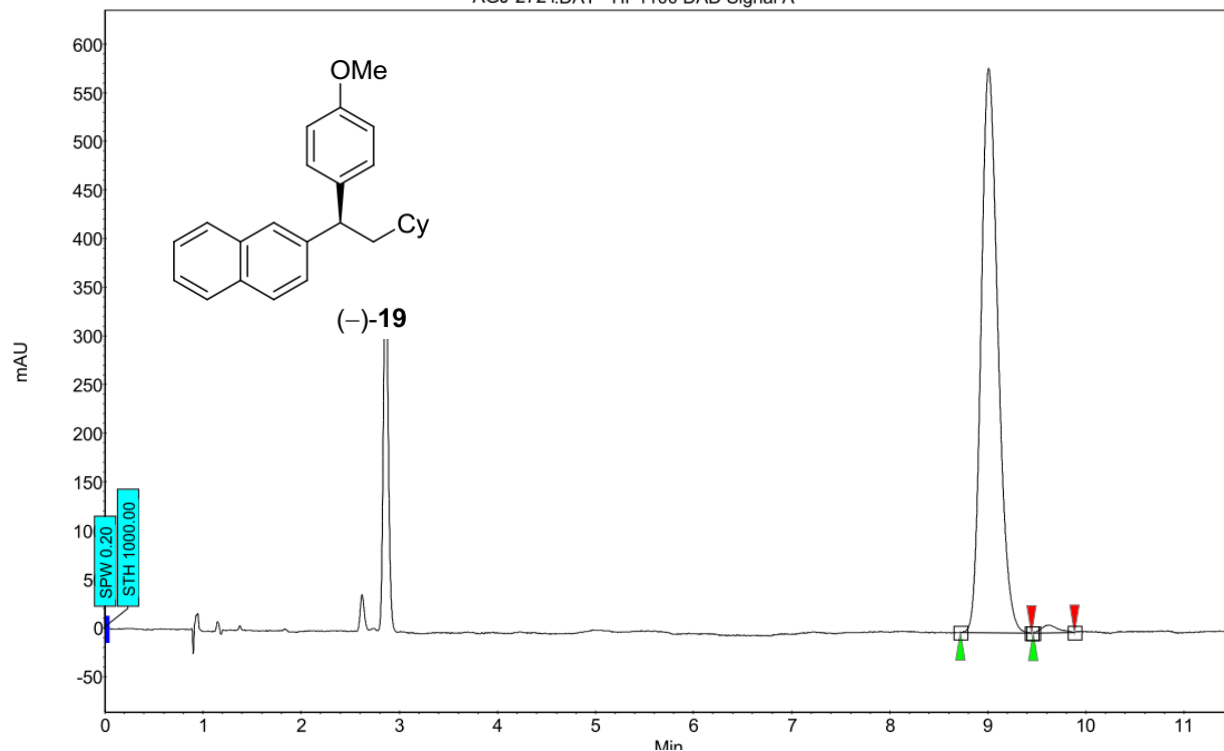


Index	Name	Start	Time	End	RT Offset	Quantity	Height	Area	Area
		[Min]	[Min]	[Min]	[Min]	[% Area]	[μ V]	[μ V.Min]	[%]
1	UNKNOWN	6.13	6.31	6.65	0.00	95.60	1711.1	272.1	95.596
2	UNKNOWN	6.69	6.85	7.07	0.00	4.40	85.5	12.5	4.404
Total						100.00	1796.6	284.6	100.000

AGJ-264-rac1.DAT - HP1100 DAD Signal A

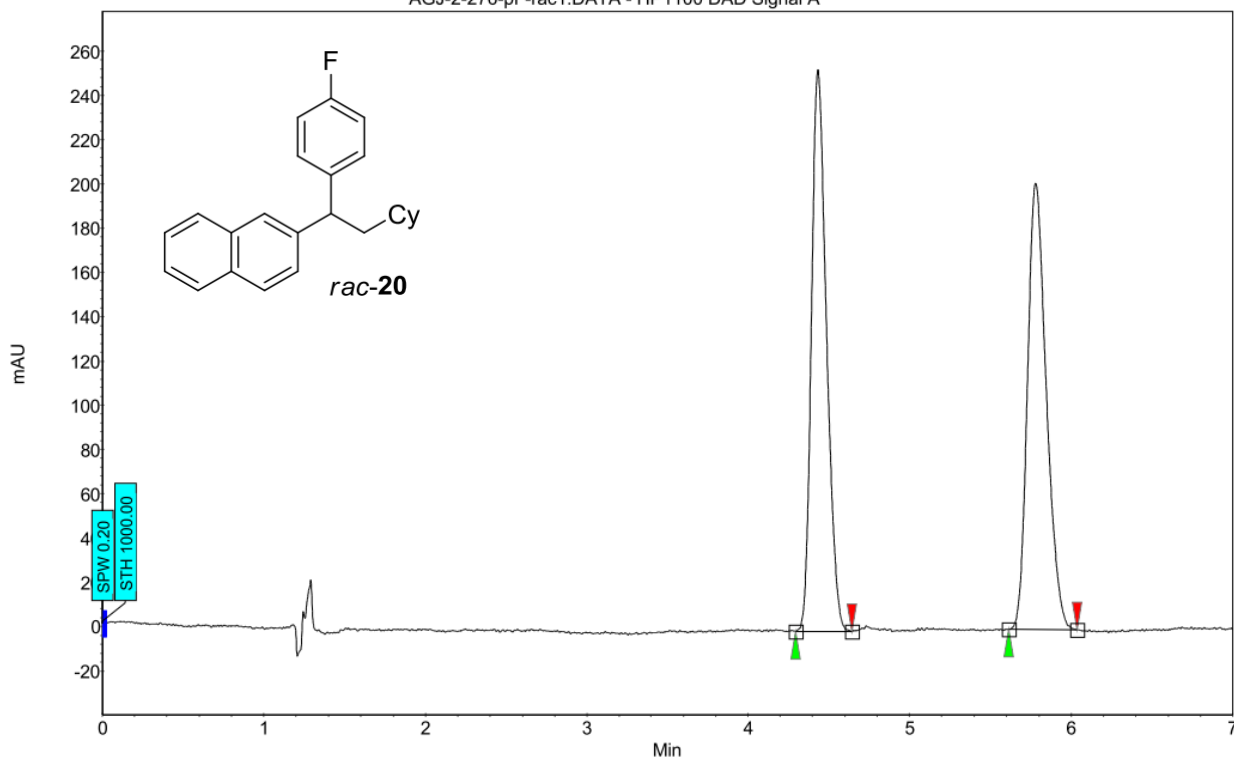


AGJ-2721.DAT - HP1100 DAD Signal A

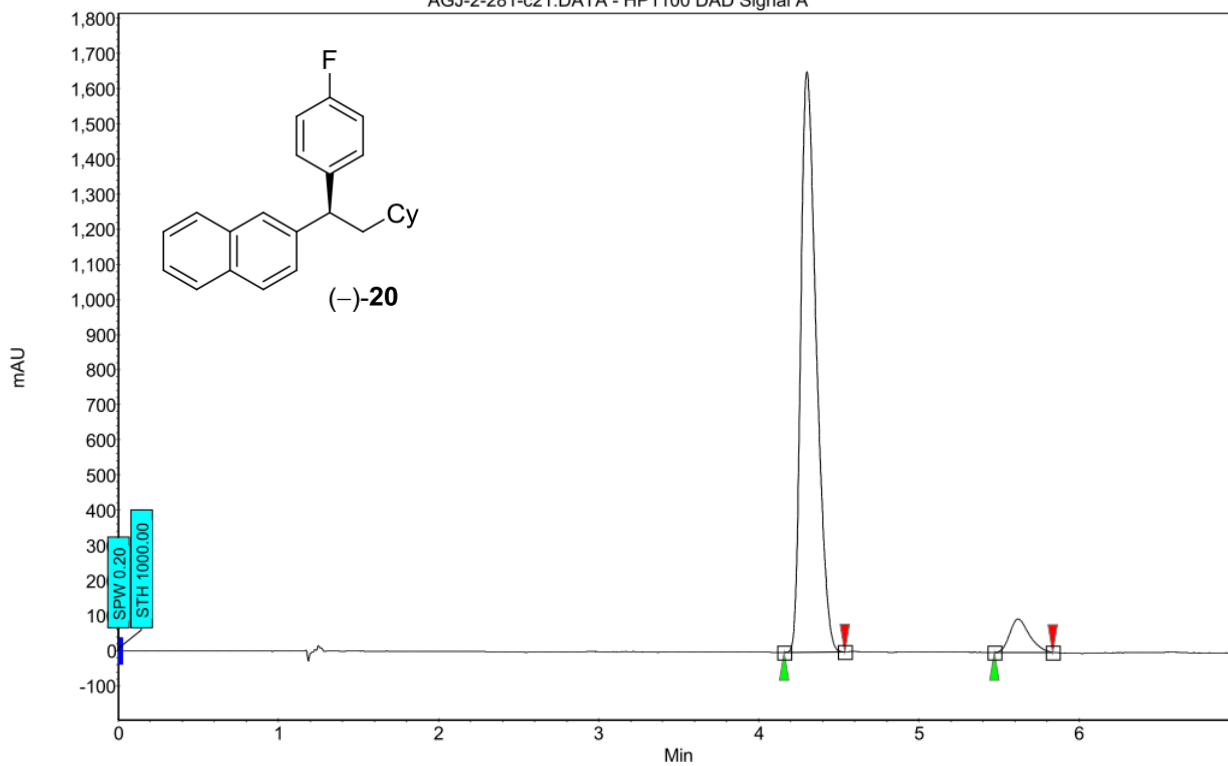


Index	Name	Start	Time	End	RT Offset	Quantity	Height	Area	Area
		[Min]	[Min]	[Min]	[Min]	[% Area]	[μ V]	[μ V.Min]	[%]
1	UNKNOWN	8.72	9.01	9.44	0.00	98.68	579.9	115.5	98.681
2	UNKNOWN	9.46	9.63	9.88	0.00	1.32	8.0	1.5	1.319
Total						100.00	587.9	117.0	100.000

AGJ-2-276-pF-rac1.DATA - HP1100 DAD Signal A

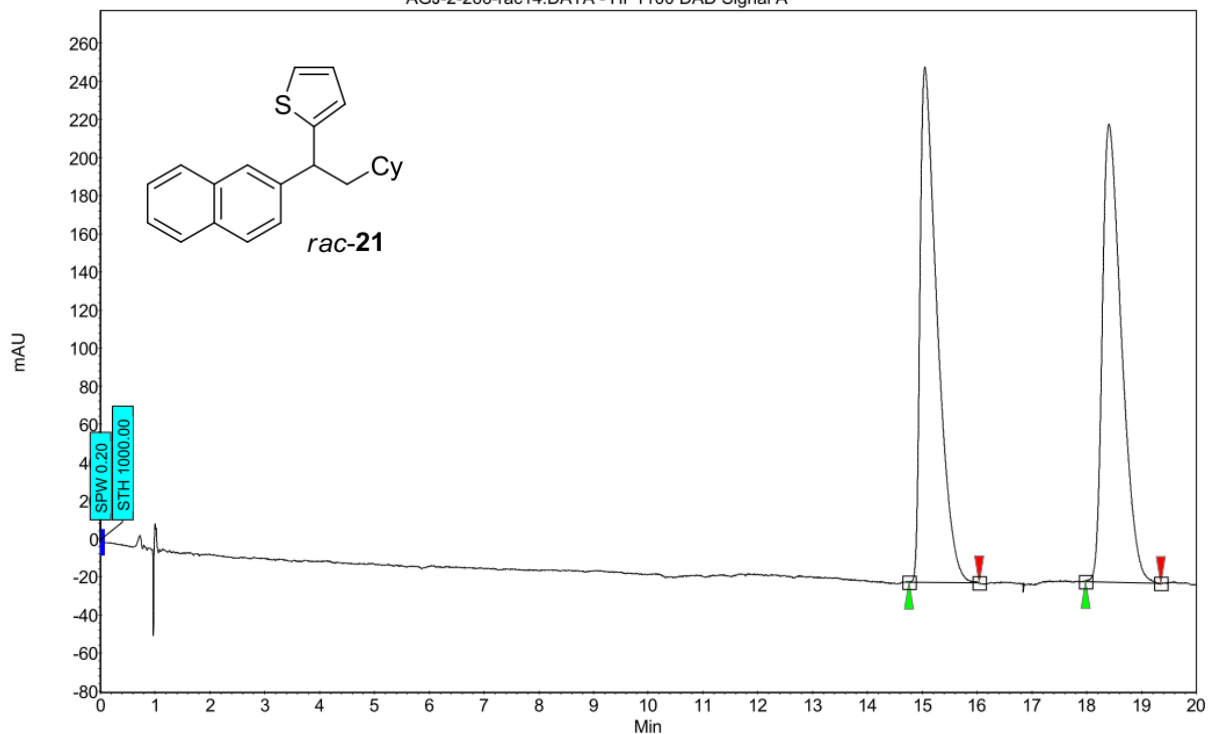


AGJ-2-281-c21.DATA - HP1100 DAD Signal A

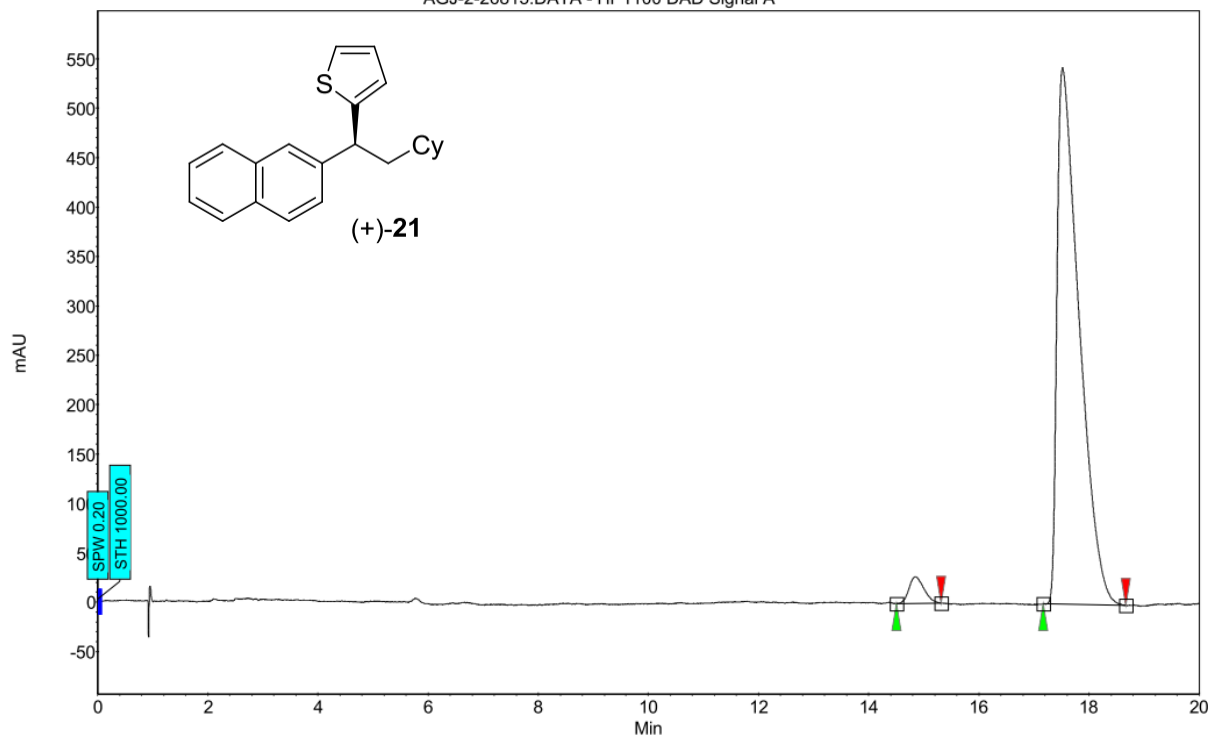


Index	Name	Start	Time	End	RT Offset	Quantity	Height	Area	Area
		[Min]	[Min]	[Min]	[Min]	[% Area]	[μV]	[μV.Min]	[%]
1	UNKNOWN	4.16	4.30	4.54	0.00	93.26	1651.3	185.9	93.264
2	UNKNOWN	5.47	5.62	5.84	0.00	6.74	94.7	13.4	6.736
Total						100.00	1746.0	199.4	100.000

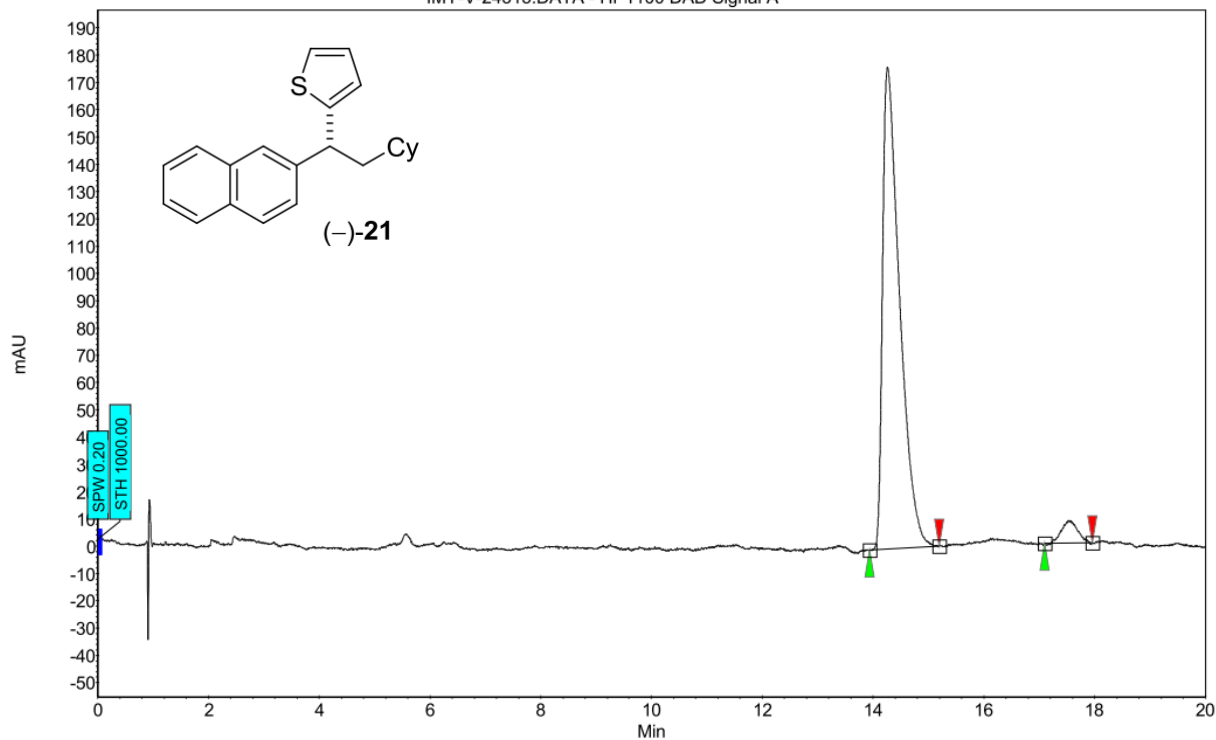
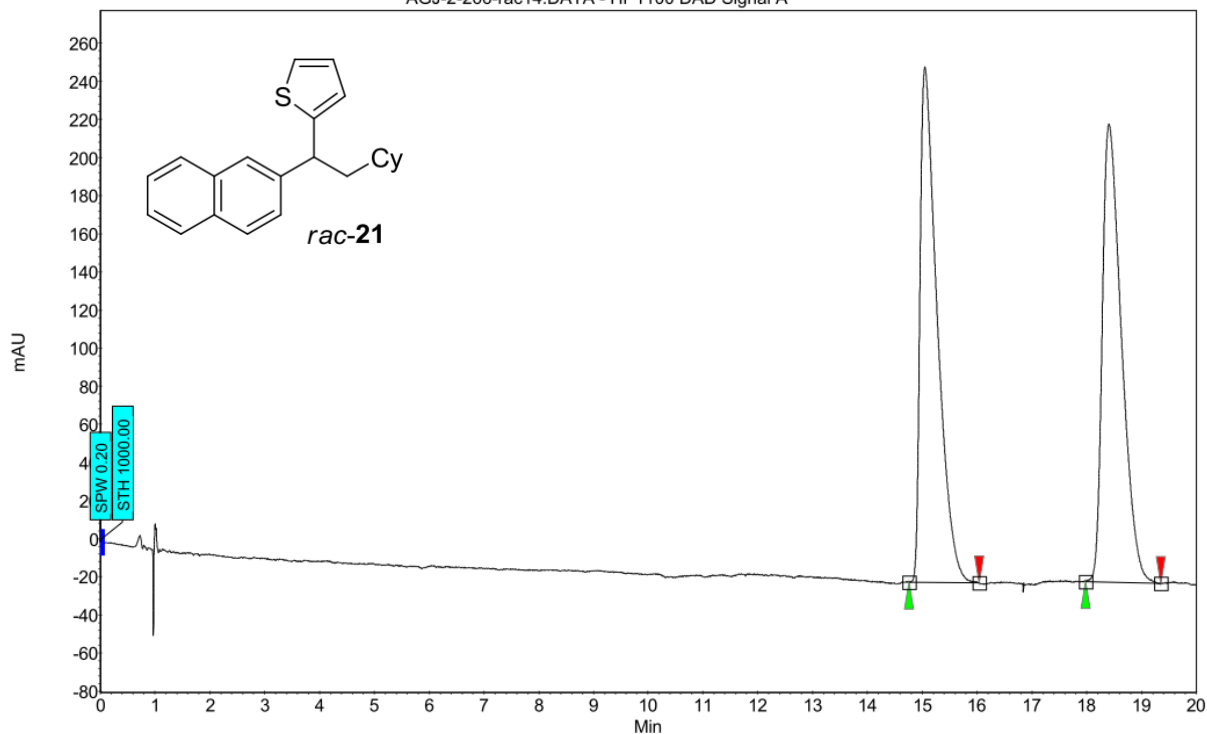
AGJ-2-266-rac14.DATA - HP1100 DAD Signal A



AGJ-2-26815.DATA - HP1100 DAD Signal A

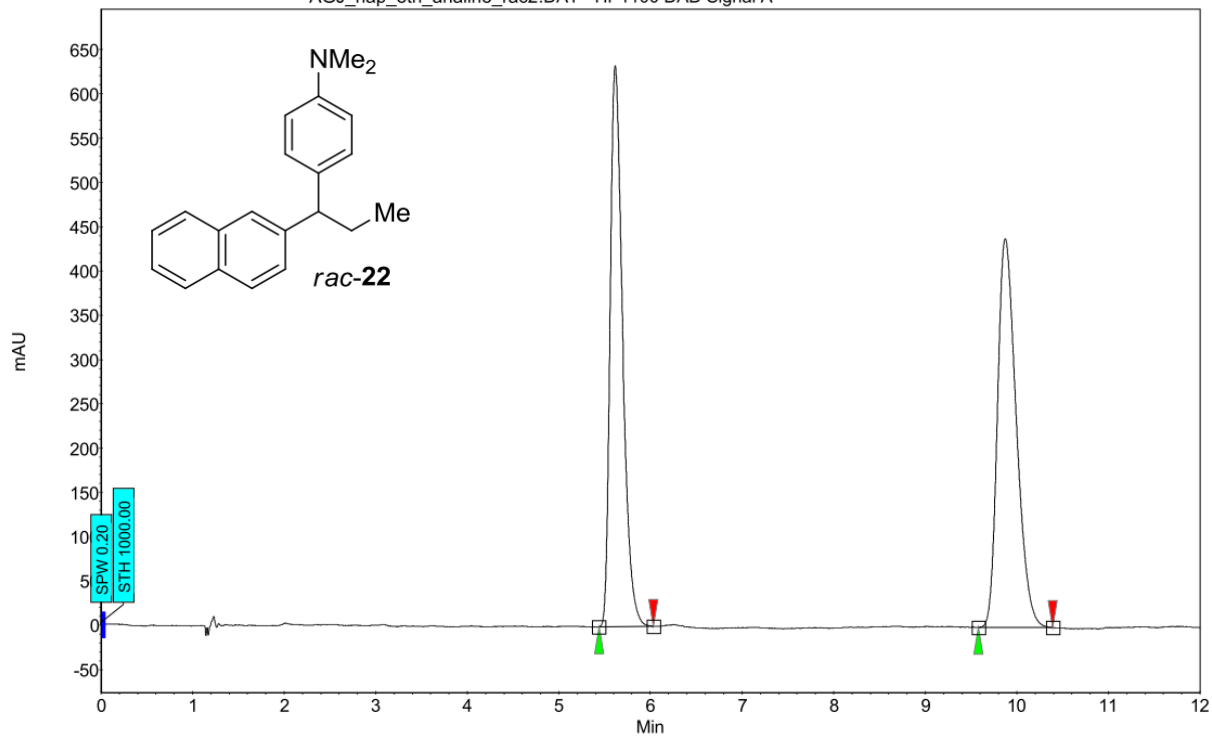


Index	Name	Start	Time	End	RT Offset	Quantity	Height	Area	Area
		[Min]	[Min]	[Min]	[Min]	[% Area]	[μ V]	[μ V.Min]	[%]
2	UNKNOWN	14.50	14.85	15.31	0.00	3.24	26.9	8.4	3.236
1	UNKNOWN	17.17	17.52	18.67	0.00	96.76	543.4	252.1	96.764
Total						100.00	570.3	260.5	100.000

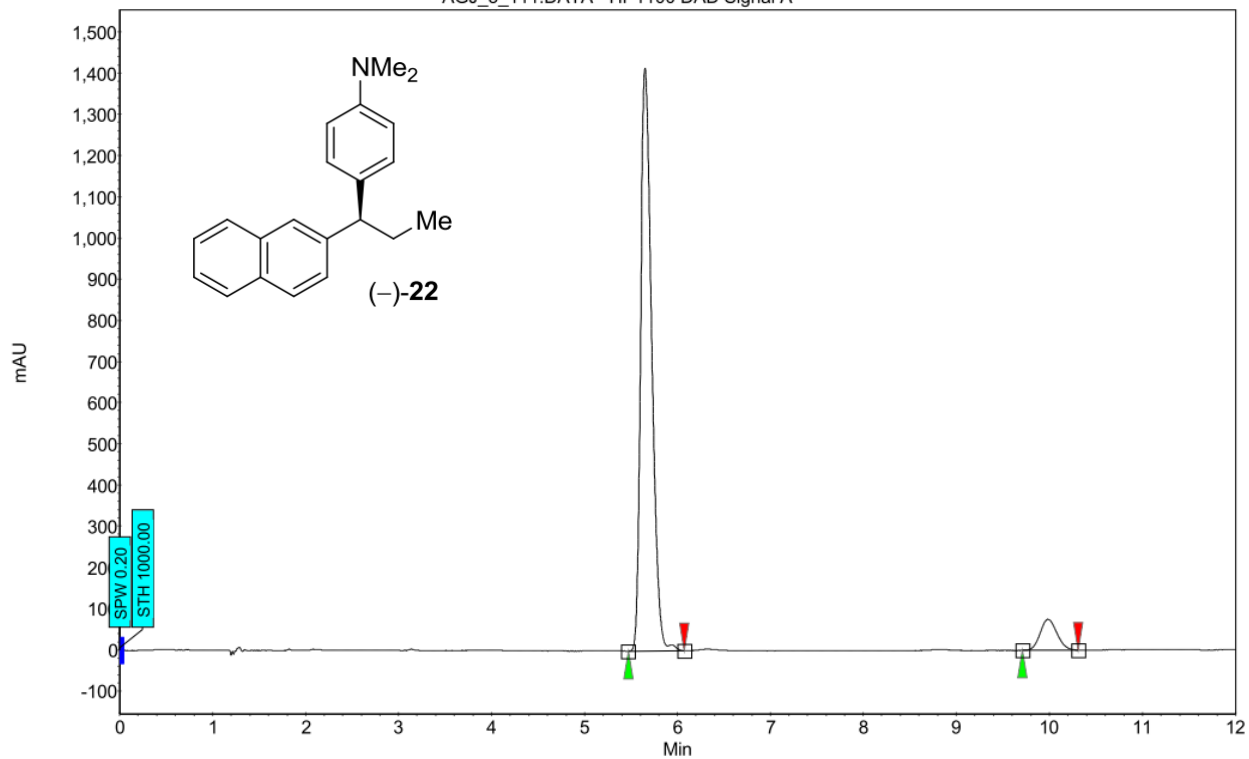


Index	Name	Start	Time	End	RT Offset	Quantity	Height	Area	Area
		[Min]	[Min]	[Min]	[Min]	[% Area]	[μ V]	[μ V.Min]	[%]
1	UNKNOWN	13.94	14.26	15.20	0.00	95.79	176.7	62.8	95.788
2	UNKNOWN	17.10	17.53	17.96	0.00	4.21	8.4	2.8	4.212
Total						100.00	185.2	65.6	100.000

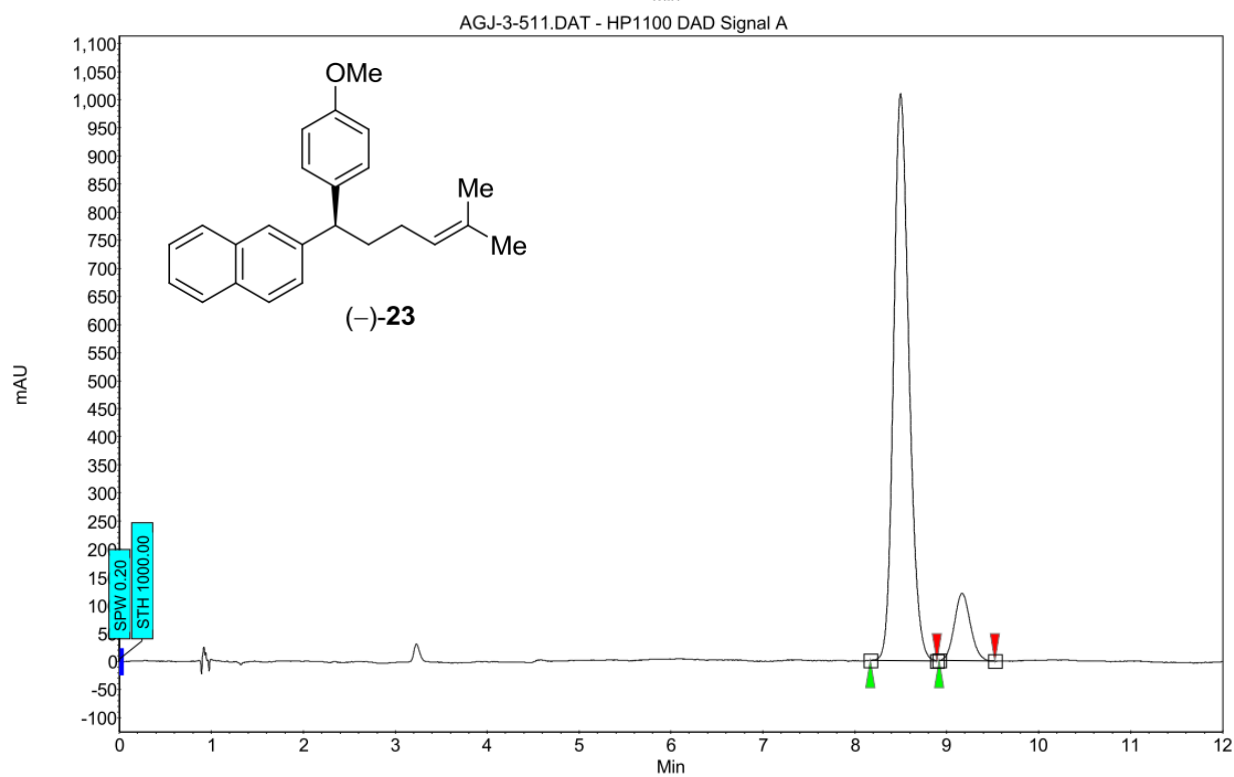
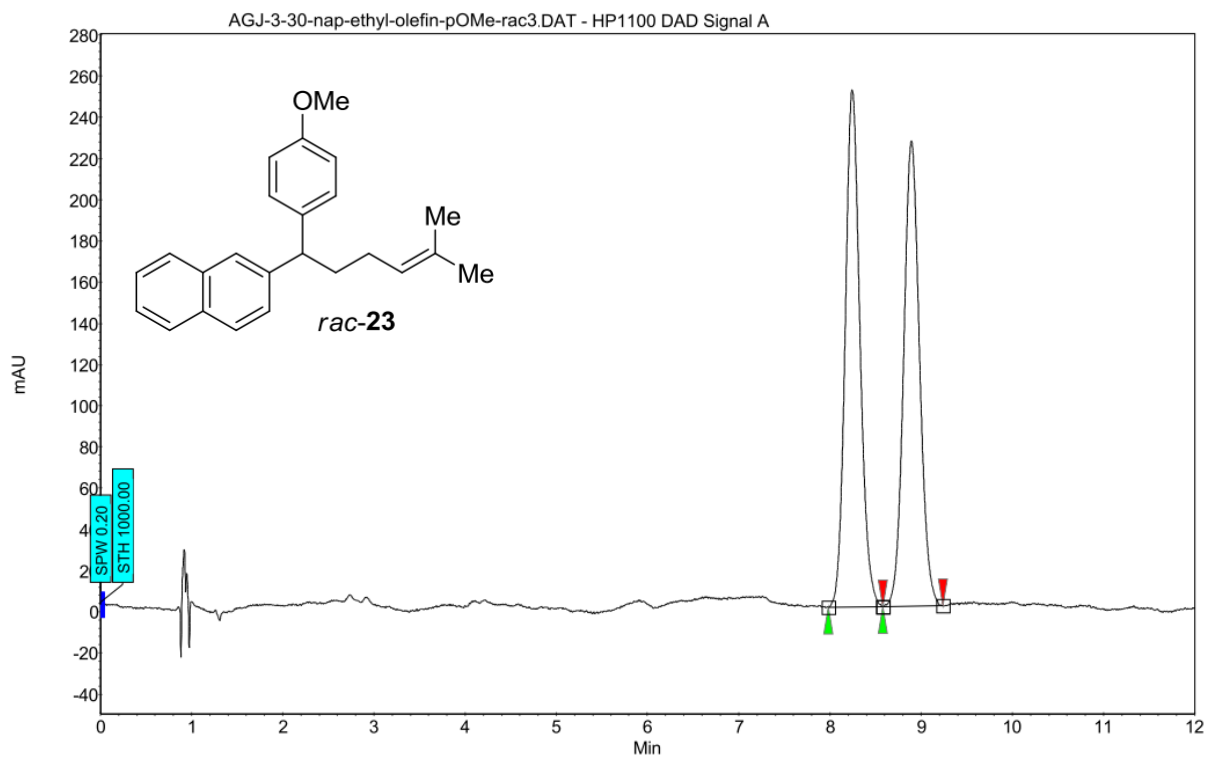
AGJ_nap_eth_analine_rac2.DAT - HP1100 DAD Signal A



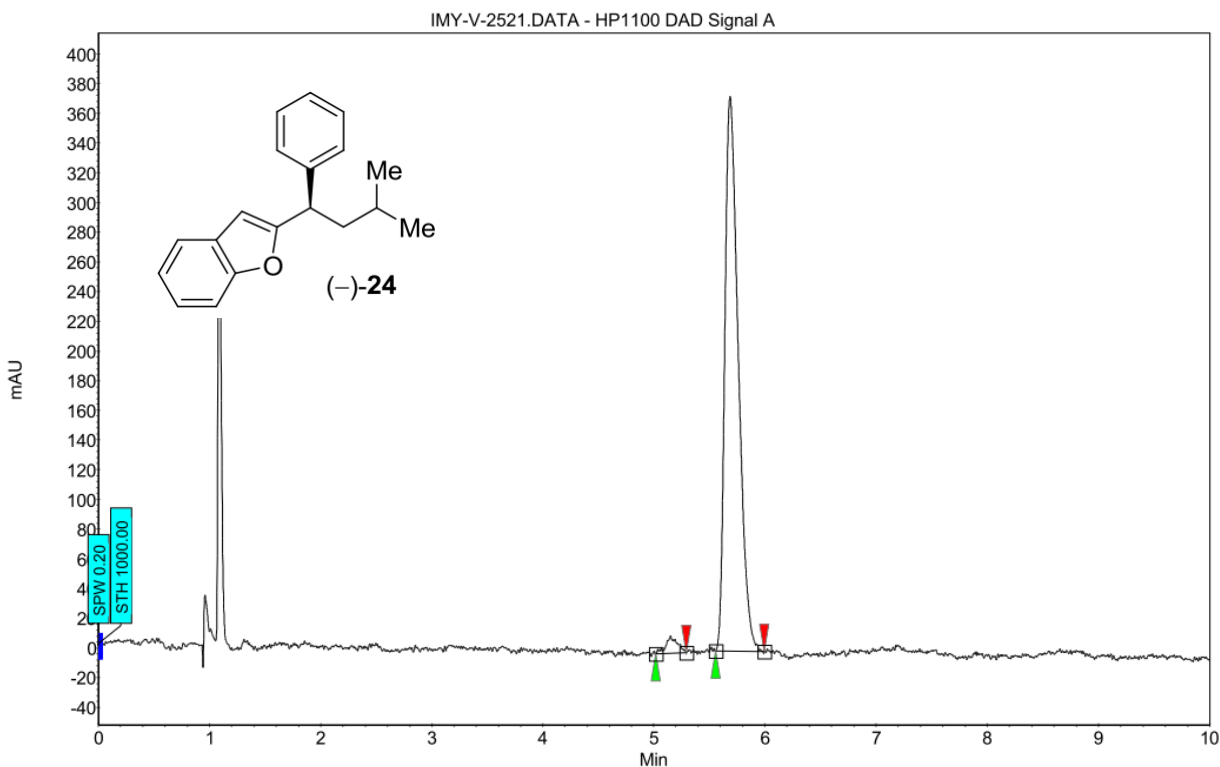
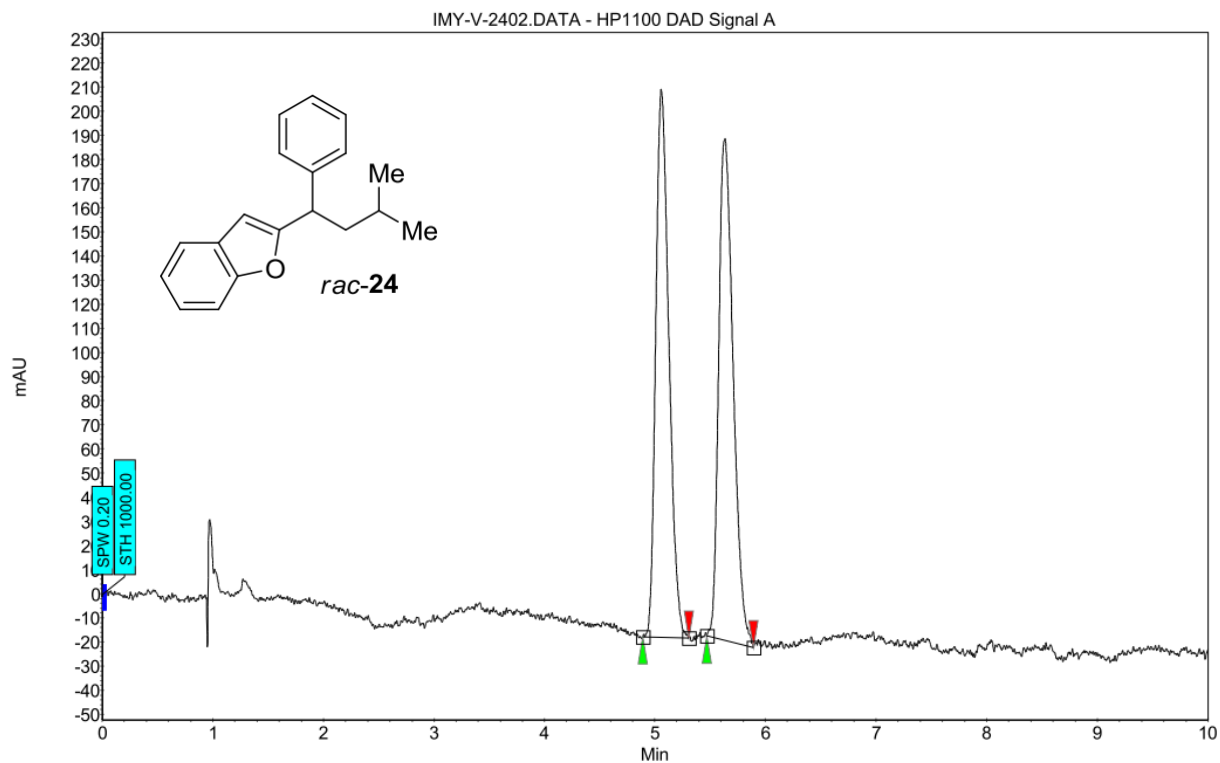
AGJ_3_111.DAT - HP1100 DAD Signal A



Index	Name	Start	Time	End	RT Offset	Quantity	Height	Area	Area
		[Min]	[Min]	[Min]	[Min]	[% Area]	[μ V]	[μ V.Min]	[%]
1	UNKNOWN	5.47	5.65	6.08	0.00	92.72	1414.5	200.0	92.717
2	UNKNOWN	9.71	9.98	10.31	0.00	7.28	75.1	15.7	7.283
Total						100.00	1489.6	215.7	100.000

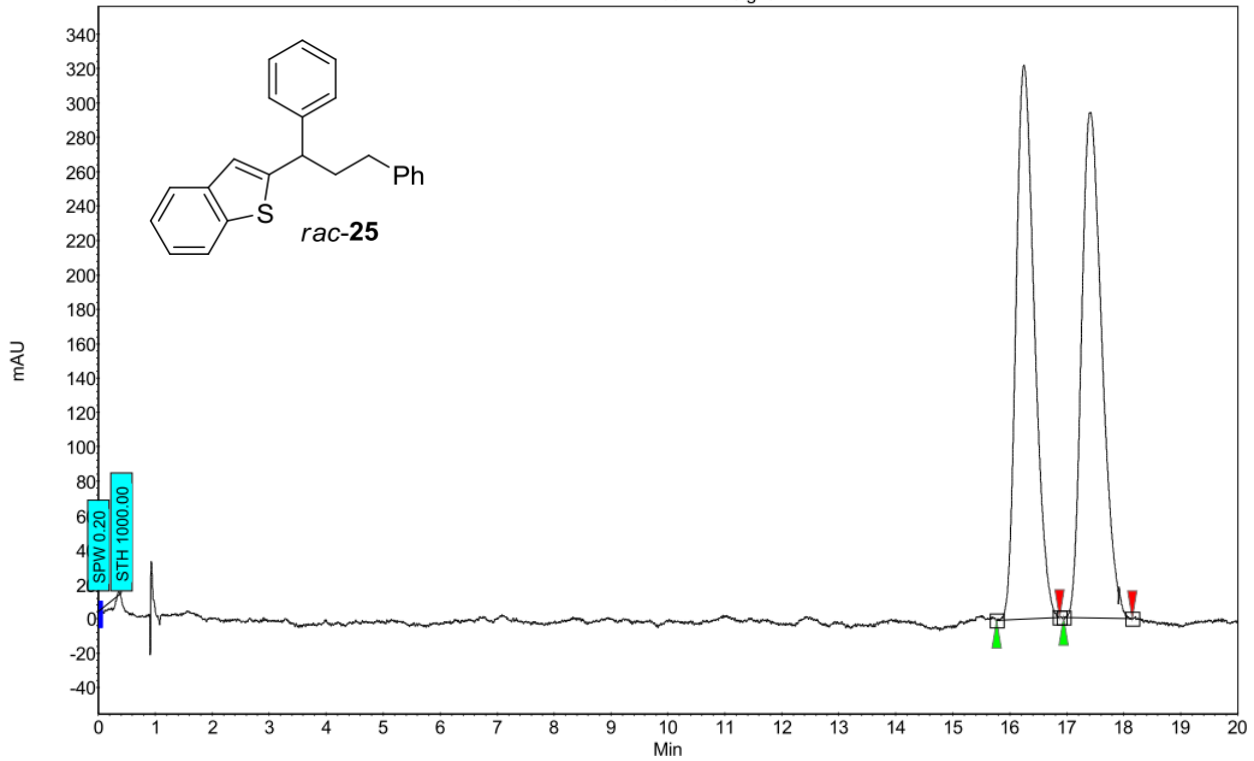


Index	Name	Start	Time	End	RT Offset	Quantity	Height	Area	Area
		[Min]	[Min]	[Min]	[Min]	[% Area]	[μ V]	[μ V.Min]	[%]
1	UNKNOWN	8.17	8.50	8.89	0.00	89.04	1009.3	195.4	89.036
2	UNKNOWN	8.91	9.17	9.52	0.00	10.96	119.6	24.1	10.964
Total						100.00	1128.9	219.5	100.000

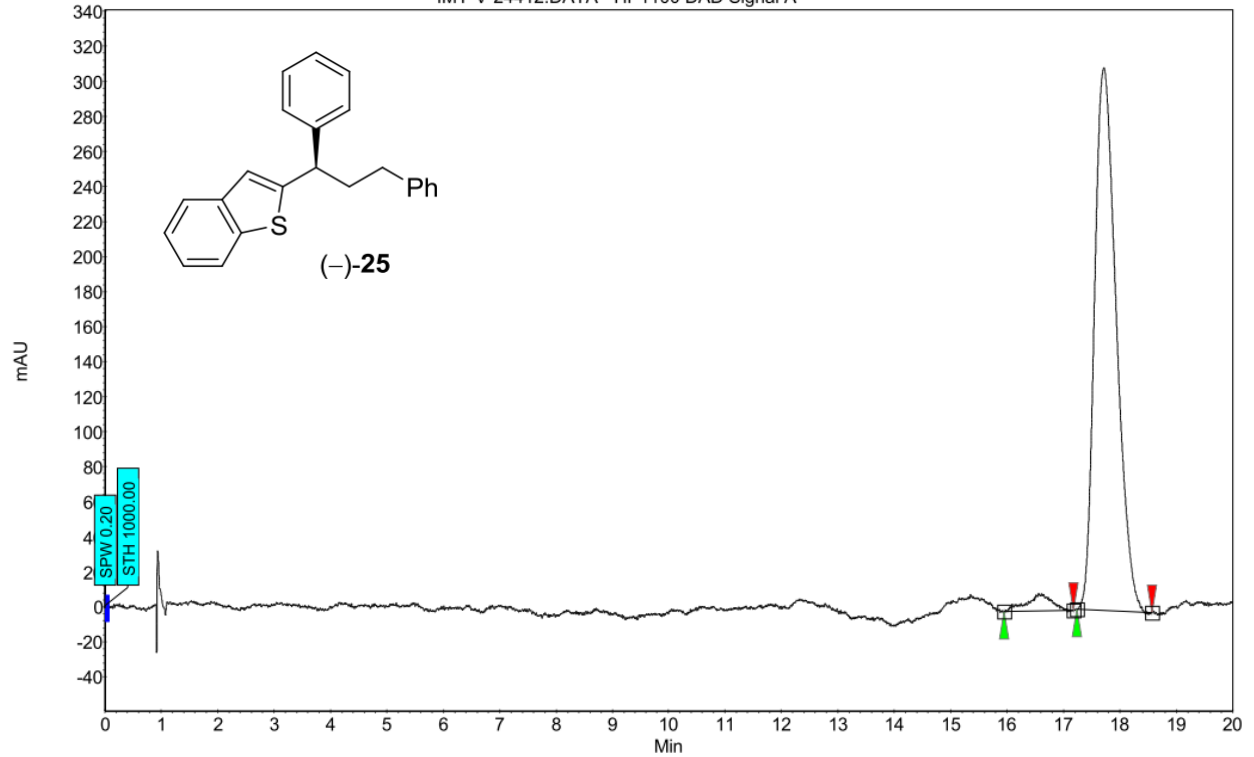


Index	Name	Start	Time	End	RT Offset	Quantity	Height	Area	Area
		[Min]	[Min]	[Min]	[Min]	[% Area]	[μ V]	[μ V.Min]	[%]
2	UNKNOWN	5.02	5.15	5.29	0.00	2.50	11.5	1.3	2.499
1	UNKNOWN	5.55	5.69	5.99	0.00	97.50	373.5	52.3	97.501
Total						100.00	385.0	53.6	100.000

IMY-V-22811.DATA - HP1100 DAD Signal A



IMY-V-24412.DATA - HP1100 DAD Signal A



Index	Name	Start	Time	End	RT Offset	Quantity	Height	Area	Area
		[Min]	[Min]	[Min]	[Min]	[% Area]	[μ V]	[μ V.Min]	[%]
1	UNKNOWN	15.95	16.57	17.17	0.00	3.31	9.8	4.7	3.313
2	UNKNOWN	17.24	17.72	18.57	0.00	96.69	309.8	138.2	96.687
Total						100.00	319.6	142.9	100.000