

Supporting Information

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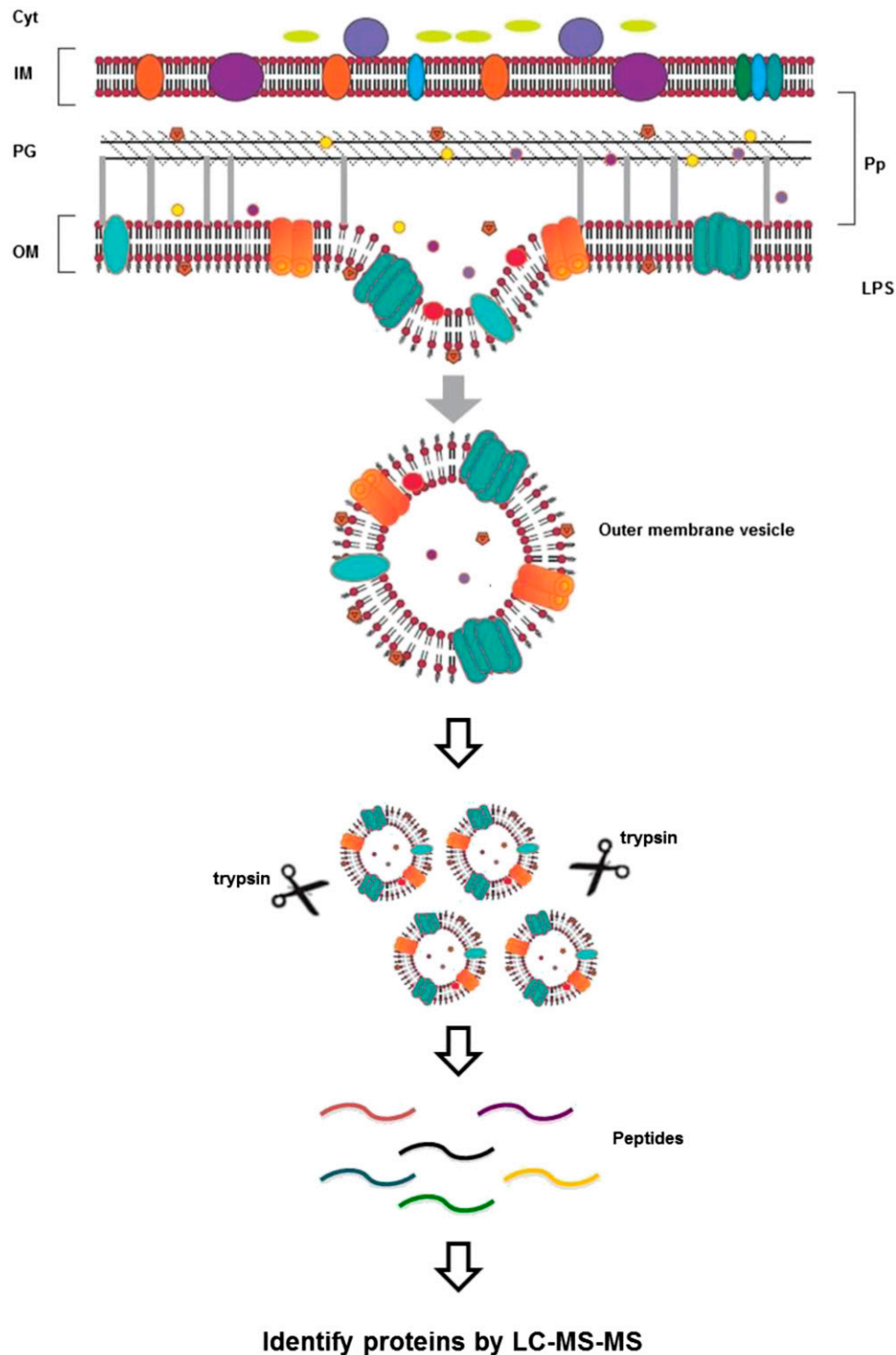


Fig. S1. In-solution digestion approach to characterize outer membrane vesicles (OMVs). OMVs are secreted from the outer membrane and are composed of outer membrane proteins, lipopolysaccharides, and periplasmic proteins (LPS, lipopolysaccharide; Pp, periplasm; OM, outer membrane; PG, peptidoglycan; IM, inner membrane; Cyt, cytosol). OMVs from *Vibrio cholerae* El Tor C6706 were purified from culture media by ultracentrifugation and subjected to overnight trypsin digestion. The resulting peptides were analyzed by LC-MS/MS. Amino acid sequences of peptides were identified by searching obtained spectra against the *V. cholerae* El Tor Inaba N16961 strain database.

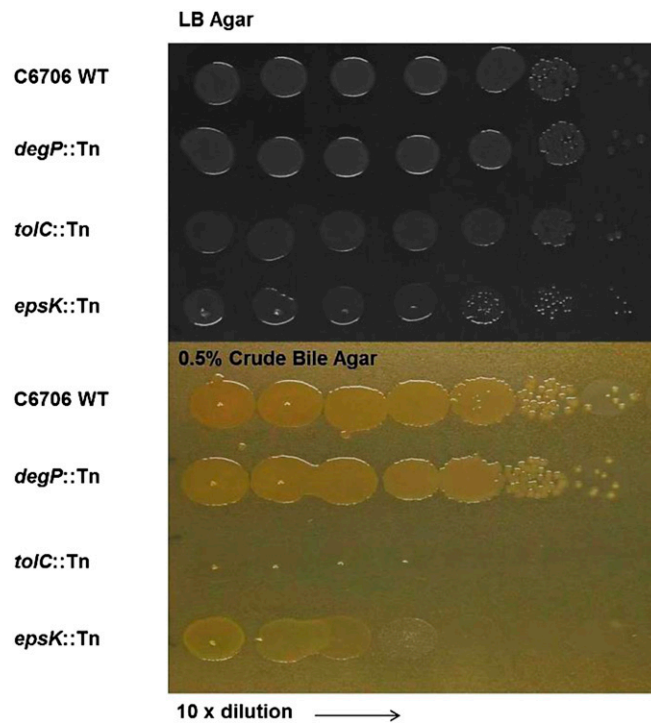


Fig. S3. Sensitivity of *degP* mutant (EC956) on LB agar containing 0.5% crude bile. *V. cholerae* strains were grown in LB until OD_{600} reached 1.0. Cells were serially diluted and then daubed onto both LB agar (row 1–4) or LB agar containing 0.5% crude bile (row 5–8) for colony counting. All TnFGL3 mutants and WT bacteria grew normally on LB agar. *degP* mutant (EC956) grew as well as WT on the bile agar whereas *tolC* mutant (EC2859) did not survive in the presence of bile. The *epsK* mutant (EC903) had severe defects on 0.5% crude bile agar (4 logs). *tolC*, outer membrane protein TolC; *epsK*, general secretion pathway protein K.

Other Supporting Information Files

[Table S1 \(DOCX\)](#)

[Table S2 \(DOCX\)](#)

[Table S3 \(DOCX\)](#)

[Table S4 \(DOCX\)](#)

[Table S5 \(DOCX\)](#)

[Table S6 \(DOCX\)](#)

[Table S7 \(DOCX\)](#)