Table S1. Primers used in this study

Primers	Sequence (5′-3′)	Applications
perR-upF	ATGGATCCGGTAATCTTTTTTATGCTC	Deletion of *perR*
perR-upR	GATACTGAAGGCTAGAGAGG	Deletion of *perR*
perR-dnF	AAGGATCCAGCAAGAACAAGTGGCTAG	Deletion of *perR*
perR-dnR	CTCATCGTCACGATCCATGTTTG	Deletion of *perR*
mntA-upF	ATAAGGATCCAGCAGGCTGCTAG	Deletion of *mntA*
mntA-upR	CTTCTTGGCTCTGGGGATCA	Deletion of *mntA*
mntA-dnF	ATAAGGATCCGCTACTACAGCATGATG	Deletion of *mntA*
mntA-dnR	CGGTGTTGTTTTCGTCCAAGACAAG	Deletion of *mntA*
perR-com-F	AAGAATTCGTGATAGAAAACCAGCTG	Complementation of *perR*
perR-com-R	ATATGTCGACCTAGCTAGCCACTTGTTC	Complementation of *perR*
mntABC-com-F	AAGAATTCCTTGTTCTCACTTTCATAC	Overexpression of *mntABC*
mntABC-com-R	ATATGTCGACTTATTTAGACAGACCTTCTG	Overexpression of *mntABC*
P*mntABC-Bam*HI	AATAGGATCCGATAATCCCTGTGATAGTCG	*mntABC* luciferase reporter
P*mntABC-Nhe*I	AATAGCTAGCCAAAGGTAAAACAGGCTTG	*mntABC* luciferase reporter
16S-F	AGAGTTTGATCCTGGCTC	qRT-PCR strandard curve of 16S rDNA
16S-R	GATGTTAGGCTTGACTCTGAC	qRT-PCR strandard curve of 16S rDNA
mntH-F	GGTGGCTCTCACATTGTT	qRT-PCR strandard curve of *mntH*
mntH-R	CCAAGGATGGAAAGACCT	qRT-PCR strandard curve of *mntH*
mntA-F	CGTTTCTTAGTTCTGCTTTTG	qRT-PCR strandard curve of *mntA*
mntA-R	CATAAAGTAGTACGACATCATCG	qRT-PCR strandard curve of *mntA*
dpr-F	GGAGCAGGCATTTACAGAGGAAC	qRT-PCR strandard curve of *dpr*
dpr-R	CCAGAGTTAGAAAGAAGAGCATGA	qRT-PCR strandard curve of *dpr*
q16S -F	CGATACATAGCCGACCTGAGAG	qRT-PCR of 16S rDNA
q16S-R	TCCGTCCATTGCCGAAGATTC	qRT-PCR of *16*S rDNA
q*mntH*-F	GGGATTCTCTGTGCTGCTATC	qRT-PCR of *mntH*
q*mntH*-R	GCAACGATGACCGCAATA	qRT-PCR of *mntH*

q*mntA*-F	ACATGAATACGAGCCTCTGCCTG	qRT-PCR of *mntA*
q*mntA*-R	GCCTTCCAAGTAAATCACGTCC	qRT-PCR of *mntA*
q*dpr*-F	GAAGATTTCAATCACGCGCTCCAA	qRT-PCR of *dpr*
q*dpr*-R	TTATACAGCTCGTATCGCTCTT	qRT-PCR of *dpr*

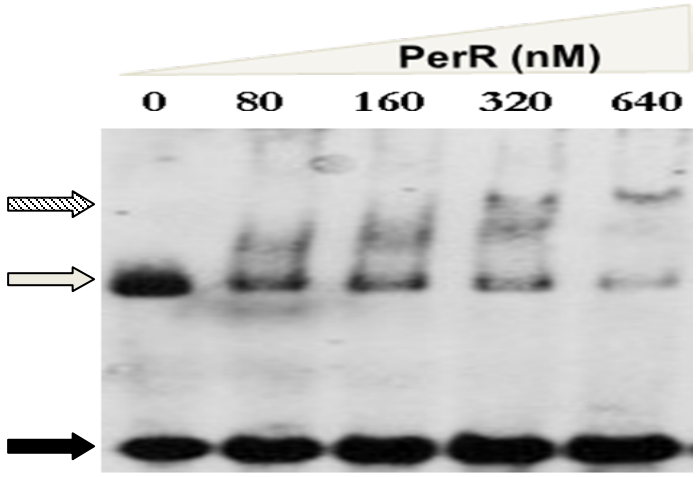

Fig. S1. EMSA detects the binding of GST-PerR protein to *dpr* promoter. Recombinant GST-PerR protein was overexpressed in *Escherichia coli* BL21(DE3)PLySs strain and purified as recommended by GE Healthcare. DNA fragment including intact *dpr* promoter (229 bp) was PCR amplified with 5' biotin labeled primers and gel purified. A labeled 50-bp DNA fragment, which contains the putative binding site of the response regulator ComE (*com* box) located in the promoter region of *comC* gene, was included as the negative control. The 20 µl reaction mixtures contained various concentrations of PerR protein (ranging from 0-640 nM), each of 0.2 nM biotin-labeled *dpr* promoter and *com* box fragment, 20 mM Tris-HCl (pH 8.0), 5% glycerol, 1mM dithiothreitol, 50 mM KCl, 5 mM $MnCl_2$, 50 µg/ml BSA, 40 ng/µl poly(dI-dC), 1U/µl catalase. After incubation for 20 min at 20°C, the reaction mixtures were separated on a 6% polyacrylamide gel. The DNA-protein complex was transferred onto a nylon membrane, and the biotin-labeled DNA was detected by LightShift Chemiluminescent EMSA Kit (Pierce, Rockford, IL). The added PerR concentrations are indicated above the lanes. Shaded arrow indicates

shifted migration position of the PerR-bound promoter fragment; gray arrow indicates the free DNA fragment, and the black arrow indicates the negative-control DNA (*com* box).

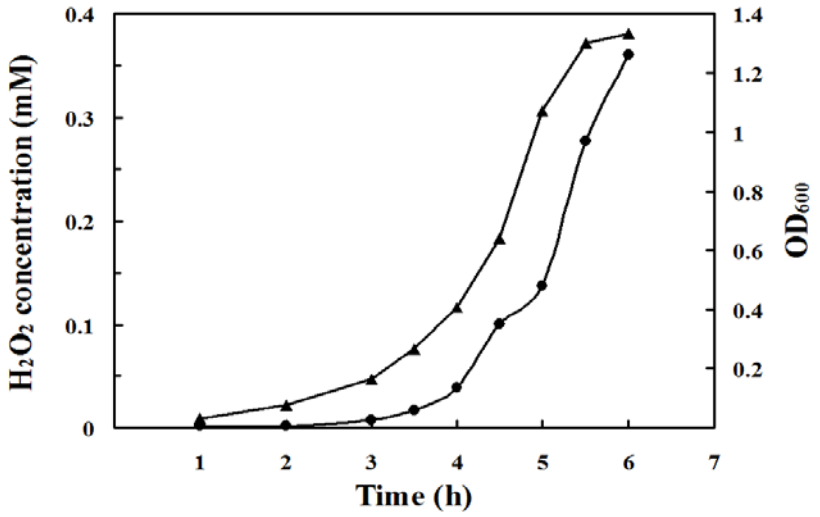

Fig. S2. Hydrogen peroxide production by *Streptococcus oligofermentans* growing statically in BHI culture. Overnight culture of *S. oligofermentans* wild strain was diluted at a 1:40 ratio into fresh BHI medium and incubated statically. Cultures were sampled at the time points shown in the figure, and then optical density at 600 nm and H_2O_2 production were measured as described in "Materials and Methods". ▲, OD_{600}; ●, H_2O_2 concentration. Data are representative of three independent experiments.