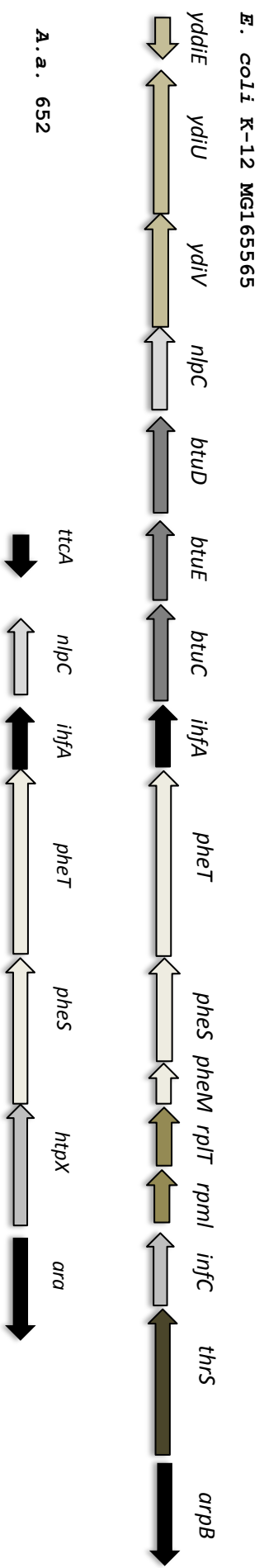
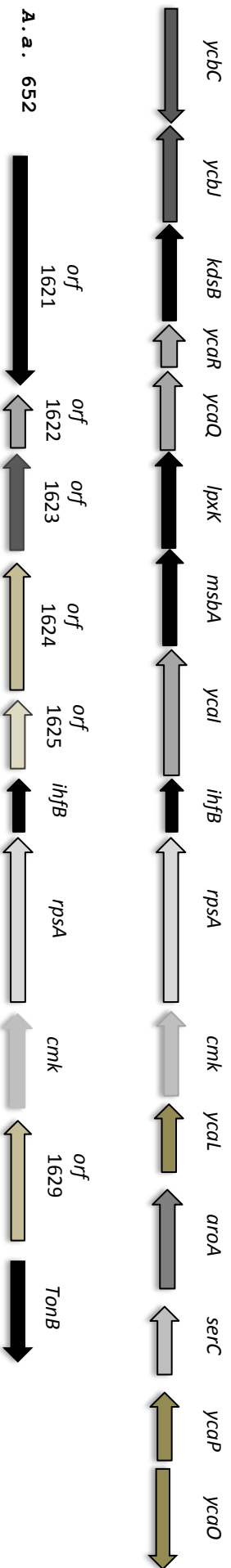


Figure S1

A



E. coli K-12 MG165565



B

<i>A. a.</i> IHF α	3	LTKVELLAENLIEK-FHLSKREAKDIVESFEEIRVALETGNDVKLSGFNGFELRDKASRP	61
		+TK EL E L++K ++ + ++ V+++ E+++ IE G +++ GFG+E L + R	
<i>A. a.</i> IHF β	1	MTKSELLIELLVQKNSNIIPKHYEEAVKAILLEQMSYVLEHGERIEVRGFGSFSLHCRQPRI	60
<i>A. a.</i> IHF α	62	GRNPKTGESVPSARRVVVEFKPGQKLRNRVE	92
		GRNPKTGE V + A+ V EK G++LR RV+	
<i>A. a.</i> IHF β	61	GRNPKTGEQVKLIDAKCVPEYFKAGKELRERVD	91

Figure S1. (A) Genomic organization of the *ihfA* and *ihfB* loci in *A. actinomycetemcomitans* compared to the related loci in *Escherichia coli*. **(B)** Protein sequence alignment of *A. actinomycetemcomitans* IHF α and IHF β . Residue numbers are shown to the right of each protein sequence.

Table S1. Strains and plasmids used in this work

Strain or plasmid	Derived, relevant genotype ^a or characteristics ^b	Source or reference
<i>Escherichia coli</i>		
XL1-Blue MRF'	$\Delta(mcrA)183 \Delta(mcrCB-hsdSMR-mrr)173$ <i>endA1 supE44 thi-1 recA1 gyrA96 relA1 lac</i> [F' <i>proAB. lacI^qZΔM15 Tn10 (Tc^r)</i>]	Stratagene
LMG194	F ⁻ $\Delta(lacIPOZY)X74$ <i>galE galK thi rpsL ΔphoA</i> (PvuII) <i>ara714 leu::Tn10</i>	Invitrogen
BL21	F ⁻ <i>ompT hsdS_B(r_B⁻ m_B⁻)gal dcm rne131 (DE3)</i>	Novagen
Top10	F ⁻ <i>mcrA Δ(mrr-hsdRMS-mcrBC)</i> Φ80 <i>lacZΔM15 ΔlacX74 recA1 araD139</i> $\Delta(ara leu) 7697 galU galK rpsL (Str^r)$ <i>endA1 nupG</i>	Invitrogen
<i>Aggregatibacter actinomycetemcomitans</i>		
652	Wild type, serotype c	(30)
652-TE78	652, $\Delta ihfA$	This study
652-TE79	652, $\Delta ihfB$	This study
652-TE-23	652, <i>IsrR::IsrR23-lacZ</i>	This study
652-TE78-23	652, $\Delta ihfA IsrR::IsrR23-lacZ Sp^r$	This study
652-TE79-23	652, $\Delta ihfB IsrR::IsrR23-lacZ Sp^r$	This study
652-TE-75	652, <i>IsrA::IsrA75-lacZ</i>	This study

652-TE78-75	652, $\Delta ihfA$ <i>lsrA::lsrA75-lacZ</i> Sp ^r	This study
652-TE79-75	652, $\Delta ihfB$ <i>lsrA::lsrA75-lacZ</i> Sp ^r	This study
Plasmid		
pYA3883	Amp ^r , expression vector	(48)
pET28a+	Km ^r , pBR322 expression vector	Novagen
pJT3	Km ^r , <i>lacZ</i> promoterless	(47)
pJT7	pYGK, Km ^r , cloning vector	Torres-Escobar et al., submitted
pJT1	Sp ^r , suicide vector	(47)
pJT10	Sp ^r , suicide vector	Torres-Escobar et al., submitted
Derived from pYA3883		
pATE76	Amp ^r , P _{BAD} <i>ihfA</i> -(AUI)-(6X His)	This study
pATE77	Amp ^r , P _{BAD} <i>ihfB</i> -(AUI)-(6X His)	This study
Derived from pET28a+		
pATE80	Km ^r , pET28a+ <i>ihfB</i> -(AUI)-(6X His)	This study
Derived from pJT1		
pATE78	Sp ^r , flanking region to <i>ihfA</i>	This Study
pATE79	Sp ^r , flanking region to <i>ihfB</i>	This Study
Derived from pJT3		
pATE23	Km ^r , P _{lSrR -1 to 255} - <i>lacZ</i>	(30)
pATE68	Km ^r , P _{lSrR -82 to -255} - <i>lacZ</i>	(30)
pATE69	Km ^r , P _{lSrR -1 to 193} - <i>lacZ</i>	This study
pATE70	Km ^r , P _{lSrR -1 to 143} - <i>lacZ</i>	This study
pATE100	Km ^r , P _{lSrR -1 to 414} - <i>lacZ</i>	This study
pATE71	Km ^r , P _{lSrA -1 to 249} - <i>lacZ</i>	(30)
pATE73	Km ^r , P _{lSrA -88 to 414} - <i>lacZ</i>	This study
pATE74	Km ^r , P _{lSrA -43 to 414} - <i>lacZ</i>	This study

pATE75	Km ^r , P _{l_{SrA}-1 to 414} - <i>lacZ</i>	This study
Derived from pJT7		
pATE92	Km ^r , P _{ihf} - <i>infA</i>	This study
pATE93	Km ^r , P _{ihf} - <i>infB</i>	This study
Derived from pJT10		
pATE94	Sp ^r , <i>l_{SrR}::l_{SrR}23-lacZ</i>	This study
pATE95	Sp ^r , <i>l_{SrA}::l_{SrA}75-lacZ</i>	This study

^a In the descriptions of the genotype, P represents promoter, and the subscript refers to the promoter fragment driving *lacZ* transcription. For consistency, nucleotide -1 is the first base upstream from the *l_{SrR}* or *l_{SrA}* start codon. ^b Amp^r ampicillin resistance, Km^r kanamycin resistance, Sp^r spectinomycin resistance, Tc^r tetracycline resistance.

TABLE S2. Oligonucleotides used in this work

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Related product, name	Sequence 5' - 3'
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–	
Upstream region <i>IsrR</i>	
ATE-6R <i>Bam</i> HI	CTTGATTATCCG <u>TGGATCCC</u> ATTAATTCGTTC
ATE-172F <i>Kpn</i> I	TCAAATACTAG <u>GGTACCT</u> CAGGAATGCACAAAATT TC
ATE-173F <i>Kpn</i> I	TCTGGATAATAT <u>GGTACCT</u> ATCTAAAAATCGCCT
ATE-242F <i>kpn</i> I	GCCGGCGATAATTTTCAT <u>GGTACC</u> AGCGTGGATTTCCCCGCCCG
Upstream region <i>IsrA</i>	
ATE-7F <i>Kpn</i> I	CTTGATTATCCG <u>TGGTACCC</u> ATTAATTCGTTC
ATE-165R <i>Bam</i> HI	GCTTTCAGCACTT <u>CGGATCCC</u> ATGCCGAAAACG
ATE-166F <i>Bam</i> HI	CGCCGAGCAGTGCGTGGGAT <u>CCCATT</u> TACCGGC
<i>ihfA</i>-AUI-6His	
ATE-188F <i>Nco</i> I	GCTTATTTGAGAGATTA <u>ACCATGGC</u> ATTAATACTAAAGTAG
ATE-189R <i>Apal</i>	CAGTAGCCAATTTATTAG <u>GGGCC</u> CAGCTTTCGGTTTGACC
<i>ihfB</i>-AUI-6His	
ATE-190F <i>Nco</i> I	GAGTTAGGAGAAT <u>CCATGGC</u> AAAGTCAGAACTTATTG
ATE-191R <i>Apal</i>	CCGTTATGAAAAGTTAGTTAGTAAAAAATTAG <u>GGCCCT</u> GCTGCATAAACGTC
ATE192F <i>Nco</i> I	CAGGAGGAATTA <u>ACCATGGC</u> AAAGTCAGAACTTATTGAAC
ATE193R <i>Xho</i> I	CTCGTTATTTAATGATGATGATGATGATG <u>CTCGAGG</u> ATGTAGCGGTACGTGTC

Deletion of *ihfA*

ATE-157F *NheI* GGACGAAAAGGCCGCTAGCGGCGTGCACCAAG
ATE-134R *XhoI* CGAGTTCTACTTTAGTTAATGTCATCTCGAGAGTTTAATCTCTC
ATE-135F *XhoI* CAAACCGAAAGCTTAACTCGAGTAAATTGGCTACTGC
ATE-158R *SacI* CTAGAAATTGTATGTGAGAGCTCTTAAATTCGTCTTGCC

Deletion of *ihfB*

ATE-159F *NheI* CAAACCTAACGCTAGCACACAATTCGCCGAAACCC
ATE-142R *XhoI* CAATAAGTTCTGACTTTGTCATCTCGAGATATTCTCCTAACTC
ATE-143F *XhoI* GACGTTTATGCAGCATAACTCGAGTTTTTTACTAACTAACTTTTCATAACGGC
ATE-160R *SacI* CGGAGAGACTTTAGAGCTCTGCAATAAGATTTGCTG

PCR verification of $\Delta ihfA$

ATE137 CAGCAGGCGGAGATAAATTAACCC
ATE140 CTGTTGCTTAAAAGCGCAATCGCC

PCR verification of $\Delta ihfB$

ATE145 GCCCGTATCGTTCATTTATCTGTTCG
ATE148 GGTAGAAAGCTGGAATTGGCTTTGCG

Cloning *IhfA*

ATE-216F *KpnI* GGTCGCACACATTTCTCGGTACCGAGAAGTCGCCCGTG
ATE-218R *XbaI* CGCCCATTCGTATTGCTGTTGCTTCTAGAAAAGCGCAATCGCC

Cloning *ihfB*

ATE-219F *KpnI* GGCTGATTTAACTAACGAAGTGGTACCTTGCCGGCGATG

ATE-221R *Xba*I

GGTAATGATTTGATCATTGTTCTAGAAATGGTAACGGCAACCAG

EMSA

I-sr f I-84	GTGCGGTCAGTTTTCCGATG
I-sr r I-85	CGCCGGCATT CAGCCACAAG
I-sr f II-68	ATTTTCCTTCACCGTTAAATTCGG
I-sr r II-69	ACGCACTGCTCGGCGGTAAC
I-sr f III-62	GAGCTGATGGGCTTTATTCG
I-sr r III-64	GCCCCAAACATCGGTAAGTCG
I-sr f IV-74	GCCGGCGATAATTTTCATCAG
I-sr r IV-75	CATTTGTTCACTCTATGCGGC
I-sr f V-70	GCACTTCACCGGCATATAAG
I-sr r V-71	GAAATTTTGTGCATTCCTGAC
I-sr f VI-78	GGAAAAATCAATGGCTTTCAG
I-sr r VI-79	CCAGATTTGTGATCTAGTTCTCA
I-sr f VII-63	CGACTTACCGATGTTTCGGGC
I-sr r VII-66	GTAAAAGCATGATGGAGATCACT
I-sr f VIII-80	GATATTAAGCATTTTGGCTCTG
I-sr r VIII-81	GTTTTTTATTATTATCAATTGTTCTTACCG
I-sr f IX-76	GCCGCATAGAATGAACAAATG
I-sr r IX-77	GATTTTGGGTATTATTCATTTG
I-sr f X-72	TGAGAACTAGATCACAAATCTGG
I-sr r X-73	CTTTGGATCGTTTCTTGATTATCC
I-sr f XI-65	AGTGATCTCCATCATGCTTTTAC
I-sr r XI-67	CAGGTTATCGTGATAATAAAAACC

lsr f XII-82	GAACGAATTAATGACGGATAATCAAG
lsr r XII-83	GAATTGATCTGGACTTTAATAATGCCTG
lsr f Yp-98	CCATGGAGACTGTTCTCATG
lsr r Yp-99	CATCACCCGGCTGTTTACCCGC

Gene Racer

ATE-208R	GCATCTGATTCGGTTTAAGCAACGACATCAGCATTTGTG
ATE-210R	ATAAGCCCACGCATAATTTCCACCAGCTGTTGATCGGCAAC

PCR verification of integration

ATE-208R	GCATCTGATTCGGTTTAAGCAACGACATCAGCATTTGTG
ATE-210R	ATAAGCCCACGCATAATTTCCACCAGCTGTTGATCGGCAAC
MDJR-123R	GGGTTTTCCAGTCACGACGTTGTAAAACGAC
MDJR-148F	GGTCTGACAGTCGACTTTCGTTGTTGAATACATG