Supporting Information

Pharmacophore-Based Virtual Screening versus Docking-Based Virtual Screening: a Benchmark Comparison against Eight Targets

Zhi Chen^a, Honglin Li^{b,*,}, Qijun Zhang^a, Xiaoguang Bao^a, Kunqian Yu^a, Xiaomin Luo^a, Weiliang Zhu^a, Hualiang Jiang^{a,b,*}

^a Center for Drug Discovery and Design, State Key Laboratory of Drug Research, Shanghai Institute of Materia Medica, Chinese Academy of Sciences, Shanghai 201203, China

^bSchool of Pharmacy, East China University of Science and Technology, Shanghai 200237, China.

Supporting Methods

The detailed procedure of virtual screening using DOCK, GOLD and Glide is described below.

DOCK. For each target, the Connolly molecular surface was calculated with a probe radius of 1.4 Å. The residues around the co-crystallized ligand within 6 Å were defined as binding site with Sybyl 6.8. Within the binding site, spheres were generated therein with the SPHGEN program encoded in DOCK4.0. SPHGEN output the spheres in clusters. Cluster 1 was selected for each target. Compounds in the databases were docked into the binding site, allowing for ligand flexibility, using the grid-based energy scoring option for minimization after initial placement in the site. The box for the scoring grid was defined such that all spheres were enclosed with an extra 7 Å added in each dimension. Scoring grids for contact and energy scores were calculated with a grid spacing of 0.3 Å. The bump check was set such that atoms closer than 75 percent of the van der Waals radii of the respective atoms were rejected. A 6-12 Lennard-Jones van der Waals potential was used along with a Coulomb potential using a distance-dependent dielectric coefficient of 4 to simulate solvation effects. The energy cutoff was set to 10 Å. The radii used were those in the vdw.defn set.

GOLD. The GOLD docking program of Version 3.1 was evaluated in this study. The active site was defined to encompass any protein atom included in a 10-Å-radius sphere centered on the mass of the co-crystallized ligand in the complex structure. 7-8 times speedup setting was selected in virtual screening. All the variable parameters were set as the default values. The compounds were flexibly docked into the binding site of the targets, and the pose with the best GoldScore for each compound was adopted.

Glide. The Glide of version 4.0 was used to screen the databases. Target protein protonation was performed with the pprep script in Glide. Glide constructed a grid that defined the ligand-binding site search region, which was centered on the co-crystallized ligand in the complex structure. The grid was defined as an enclosing box with 15 Å in all three dimensions. The standard precision (SP) docking mode was selected. The ligands in the MOL2 format in databases were transformed into MAE format with mol2convert utility encoded in the program, and then docked flexibly into the binding site. The pose with the best GlideScore for each ligand was saved for further analysis.

Supporting Tables and Figures

Table S1. Chemical Structures of the Actives Used in This Study. The DrugBank NumbersWere Used to Name the Actives.

Compound	Structure	Compound	Structure	Compound	Structure
ACE					
DB00178		DB00519		DB00584	O O O O O O O O O O O O O O O O O O O
DB00492		DB00542		DB00691	
DB00790		DB00881		DB01089	
DB02032	HO O HS N	DB04184	ОТОН	DB03740	
DB01180		DB01348	O NHOO N S S S		

AChE





DacA







Table S1 (continued)



Figure S1. Refined pharmacophore models for the eight targets represented in Catalyst form: hydrogen bond acceptor (HBA), hydrogen bond donor (HBD), hydrophobic (HP), and excluded volume (gray sphere).



Figure S2. Actives mapping to the pharmacophore model and fitting to the binding pocket of ACE. Actives in the 5% of the highest ranks of the entire database produced by the pharmacophore-based method were used in the mapping and fitting. Pharmacophore model was represented in Catalyst form: hydrogen bond acceptor (HBA, green), hydrogen bond donor (HBD, purple), hydrophobic group (HP, cyan), and excluded volume (EV, gray). Binding conformations of the actives were isolated from the results of virtual screens. Actives in the docking models were represented in stick model. Ligand bound in the crystal structure was used as reference molecule (white), and actives docked by DOCK, GOLD and Glide were displayed in red, green, and blue colors, respectively.





DB00178 Fit=3.38







F



DB02032 Fit=3.79

Figure S3. (A) Structural superposition of ACE actives after pharmacophore mapping and binding pocket fitting. The conformation of the actives predicted by DOCK (B), GOLD (C), and Glide (D) are also displayed.



Figure S4. Actives mapping to the pharmacophore model and fitting to the binding pocket of AChE. The figure caption is the same as Figure S2.



DB00944 Fit=2.95













DB00981 Fit=2.13

Figure S5. Structural superposition of AChE actives after pharmacophore mapping and binding pocket fitting. The figure caption is the same as Figure S3.



Figure S6. Actives mapping to the pharmacophore model and fitting to the binding pocket of AR. The figure caption is the same as Figure S2.



Figure S6 (continued)



DB02478 Fit=2.71









DB02901 Fit=3.06

Figure S7. Structural superposition of AR actives after pharmacophore mapping and binding pocket fitting. The figure caption is the same as Figure S3.



Figure S8. Actives mapping to the pharmacophore model and fitting to the binding pocket of DHFR. The figure caption is the same as Figure S2.



Figure S8 (continued)



Figure S9. Structural superposition of DHFR actives after pharmacophore mapping and binding pocket fitting. The figure caption is the same as Figure S3.



Figure S10. Actives mapping to the pharmacophore model and fitting to the binding pocket of DacA. The figure caption is the same as Figure S2.



Figure S11. Structural superposition of DacA actives after pharmacophore mapping and binding pocket fitting. The figure caption is the same as Figure S3.



Figure S12. Actives mapping to the pharmacophore model and fitting to the binding pocket of HIV-pr. The figure caption is the same as Figure S2.





Figure S13. Structural superposition of HIV-pr actives after pharmacophore mapping and binding pocket fitting. The figure caption is the same as Figure S3.

