

Supporting Information

We enclose this File S1 of Supporting Information containing a detailed account of the classification of bacterial *COX* operons (2 pages), 4 additional Figures and 4 additional Tables.

Classification of bacterial *COX* operons

The terminal oxidases considered here (Fig. 1) belong to the super-class of heme-copper oxygen reductases, which are classified on the basis of structural-functional features of their common catalytic subunit [26], corresponding to *COX1* or *ctaD* for the *aa₃* cytochrome *c* oxidases. Type A includes the majority of these oxidases, while type B includes the *ba₃* oxidases [26,47] that lack *COX3* and *COX4* in their core gene sequence. Type C correspond to the *cbb₃* oxidases, which also lack *COX3* and *COX4*. Finally, NOR includes the nitric oxide reductases, which are considered under the system of N metabolism here (Fig. 1). While maintaining the framework of this classification, we introduce a new subdivision of the *aa₃* cytochrome *c* oxidases which accounts for the large variety in their genomic sequences organized in operons [46,47,51], herein called ‘*COX* operons’ (Figs. 3 and S4, Table S2). We have now classified them systematically in three groups (or sub-types of the overall category A [26]): **type a**, conserving the core sequence *ctaC-F*; **type b**, with the core sequence intermixed with genes for the assembly of the oxidase cofactors, e.g. *CtaB*; and **a-b transition**, showing features intermediate between type a and b and restricted to β- and γ- proteobacteria. Additionally, the *ba₃* oxidases (type B [26]) are maintained as a separated type of *COX* operon (Table S2), even though their gene sequences shows features related to both *COX* operon a-b transition and the operon of *cbb₃* cytochrome *c* oxidases. From the composition and positioning of the genes that are prepended [82] (i.e. upstream) and appended [82] (i.e. downstream) to the core sequence, we distinguish four different *COX* operons of type a. Type **a** alone is characterized by *COX1* fused with *COX3* and the presence of tri-heme *c* cytochromes (Fig. 3 and data not shown). Type **a-I** is associated with the Act (Alternative Complex Three) gene clusters containing homologues of the nitrite reductase *nrfD* [49,50] (Figs. 3A and S4); similarly to *ba₃* oxidases, the combined operon is predominantly present in Rhizobiales and Rhodospirillales (Fig. 6A). Type **a-II** is recognized for the fusion of *COX4* at the C-terminus of *COX1* (Fig. S2), the fusion of *COX2* with a *c*

cytochrome and the frequent association with PQQ-dependent dehydrogenases; its richness in mono- and di-heme *c* cytochromes suggests possible evolutionary relationships with the operon of *cbb₃* oxidases. Finally, type **a-III** characteristically contains a doublet of *ctaE/COX3* proteins (Figs. 3A and S4) and is widely distributed among α -proteobacteria (Table S2). Although *COX* operons of type a are present also in β - and γ -proteobacteria (Tables S2-S4), *COX* operon type **b** is present only in α -proteobacteria, showing no fusion among *COX* subunits and more variation in the contiguity of the gene sequence than in the gene sequence itself. Indeed, while *Beijerinckia* or *Tistrella* have a continuous gene sequence, Rhodobacterales and other taxa present different fragmentations of the operon in separate gene clusters (Figs. 3A and S4A). Of note, previous phylogenetic studies on *aa₃* cytochrome *c* oxidases predominantly used the proteins belonging to *COX* operon type b [6,7,13,47,64].

With regard to the possible evolution of *COX* operons, we hypothesize that an early duplication of *COX4* might have been crucial in the progressive breaking apart of the operon core sequence and the major structural transition of *COX3* from the 5-helices to the 7-helices form (Fig. S3), presumably by fusion of a two-helices *COX4*-like protein. This large molecular change has enhanced *COX3* binding to specific lipids, in particular PG1 and PG2 [60] (see also Table S4). The resulting dense lipo-protein packing could modulate the entry of oxygen in the catalytic centre [60], thereby reducing the affinity for the oxygen substrate. This molecular change affecting the affinity for oxygen appears to correspond to the juncture between *COX* operons type a-II lacking a separate *COX4* subunit and the transition a-b type (Figs. 3 and S3). We surmise that such a junction might have been concurrent with the separation of the β - and γ -lineages from primordial α -proteobacteria (Fig. 3C), an event which occurred around the time when oxygen levels strongly increased on the planet [31].

Supporting Information Figures – and their legend

Figure S1 - Pathways for the bioenergetic evolution of α bacterial not leading to mitochondria. The diagram shows the additional subsets of bioenergetic systems that are not shown in Fig. 1B, including those of *Asaia* and *Saccharibacter* (Table S1B). The asterisk* labels the same subset as in Fig. 1B (main text), but with fewer representative taxa. Underlined organisms are symbionts or pathogens. Each of the six bioenergetic systems presented in Fig. 1 was identified from its catalytic protein subunits and was considered functionally absent when one or more of these subunits were not found in their completeness, as indicated by the profile of their conserved domains (cf. [41]). The functional absence of a given system is represented by an empty square as in Fig. 1B.

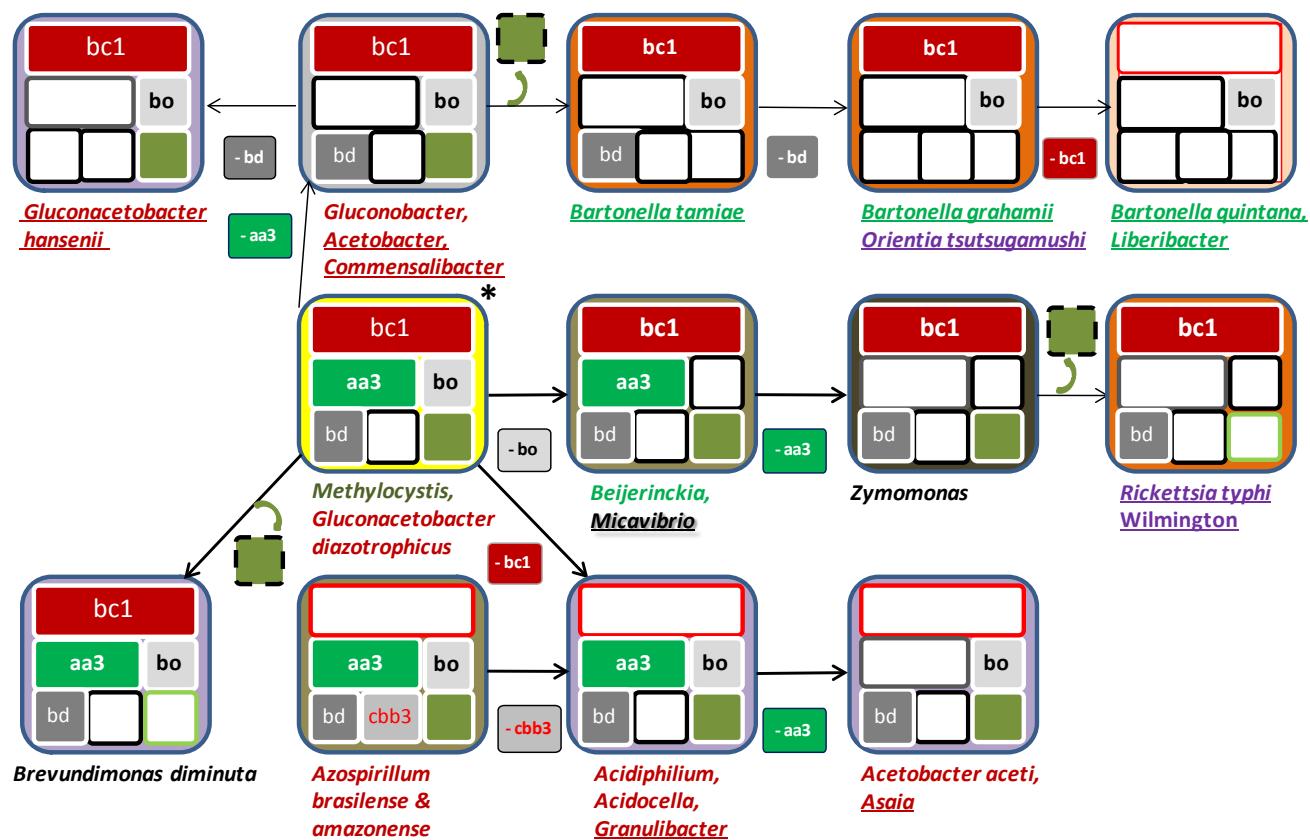


Fig. S2. Sequence analysis to identify the fusion of COX4 subunit with COX1 proteins.

A. Sequences of recognised or putative *COX4* were manually aligned to reference proteins having known 3D structure around the first transmembrane helix (TM1, highlighted in grey): subunit IV of *Thermus caa₃* oxidase (accession: pdb|2YEV [54]) and subunit IV (COX4_pro_2 super family [cl06738]) of *Rhodobacter Sphaeroides aa₃* oxidase (chain D, accession: pdb|1M57 [53]). *Residues in **bold** have positive scores (≥ 0) in the BLOSUM62 substitution matrix [83], those **yellow-highlighted** are identical with either reference protein, while those highlighted in **purple** are identical to *Janibacter COXIV* (accession: ZP_00994995) with scores ≥ 5 [83]. The total count of identities is also highlighted in yellow (**tot**) before the description of the protein on the right. It was used to identify other *COX4*-like proteins such as DUF983 (see Fig. 3A and the section entitled “classification of bacterial *COX* operons”). The minimal count for deeming a protein as “*COX4*-like” was considered to be 10, but several *COX1* proteins exhibited larger numbers of identities. The region of ciliate *COX1* showing similarity with *COX4* partially overlaps the last transmembrane region (TM12) of aligned *COX1*, which is well conserved among all available *COX1* sequences from ciliates. However, the *COX4*-like region in bacterial *COX1* and that of the pathogenic fungus *Zymospetoria* [55] lies outside the conserved domains of other *COX1* proteins. *Azospirillum bras*, *Azospirillum brasiliense*; *Methylobac_extor*, *Methylobacterium extorquens*. **B.** This panel shows the alignment of *COX4* subunits around the second transmembrane helix (TM2), the structure of which is known only for subunit IV of *Thermus caa₃* [54] that was used as the reference for aligning bacterial *COX4* and mtDNA-encoded proteins. In **bold black** are the residues that are identical in the aligned position of at least two *COX4* sequences, or are positive substitutions [83] across at least three aligned *COX4* sequences; they are additionally **yellow-highlighted** when identical between at least one bacterial *COX4* and one mtDNA-encoded protein (cf. A). In **bold dark blue** are the residues that are positive substitutions between bacterial *COX4* and mtDNA-encoded proteins, while those in **bold light blue** are identical or positive substitutions among the aligned mtDNA-coded proteins. This colour labelling enhances the limited similarity between the sequences shown.

A

| Accession: | | TM1 | | | | | | | | | | | |
|--------------|------|--|----|------------|-----------|----------------|--------|-------------------------|------------------|------------|--------------------|------------|------|
| pdb 1M57 J | 12 | HVAGSMSDITTQQEKT TAG FVVRMVT A AVVIVAA L FLALANA | 51 | aa3 | subunit | IV | 3D | Rhodobacter_sphaeroides | | | | | |
| pdb 2YEV | 2 | - VYIALF ALGAALTVLFYFLILNP--RVLTT E GTFD----- | 34 | caa3 | subunit | IV | 3D | Thermus | | | | | |
| | | | | ≥0* | identical | tot | | | | | | COX operon | type |
| YP_006592792 | 10 | SVASLM F ALLA E ELAAT F S--FP G WGRSGVAVIAAMVGIA A F G E | 24 | 14 | 3 | 17 | Cox | 4 | Methylocystis | SC2 | | a-I | |
| YP_005030786 | 582 | TIWPLLS A LTAVT I LGSI-FTP N ALLVW-GAVPLAVGLV-G W FW | 22 | 14 | 8 | 22 | Cox | 1 | Azospirillum | bras | | a-II | |
| WP_009763216 | 630 | SIW P PLIAAIATA- I LF I GSM-FTP N ALLVW-G S VL L AL A LM-G W FW | 22 | 12 | 8 | 20 | Cox | 1 | Microvirga | | | a-II | |
| YP_006593115 | 588 | SM P LLIAA- T VI T IGSI-FTP N AVLVW-G C ALVA-AP L IG W FW | 21 | 11 | 8 | 19 | Cox | 1 | Methylocystis | SC2 | | a-II | |
| YP_001637594 | 589 | SIW P PLA A LA V AVG-AT I IGSI-FTP N AVLVW-G A PP I AA A ALT-G W FW | 25 | 11 | 7 | 18 | Cox | 1 | Methylobac_extor | PA1 | | a-II | |
| YP_006373319 | 588 | SFRPLAAA I FIG-T C PI A AT-FEW N W L AAAG-G I IGIGV I GL G WLW | 17 | 4 | 7 | 11 | Cox1/3 | | Tistrella | mobilis | | a-II | |
| YP_00994995 | 89 | SW W PL L AG S C A -F I FL G LA- V GW W LV A AL V P A LV-G W FW | 23 | 12 | 13 | 25 | Cyt_c | ox | IV | Janibacter | | caa3 | |
| YP_001648742 | 593 | VGDVTSRSQIYS A LIHSMT V MP T PNPT L LG-G I QLTGNA I IV W SA | 18 | 9 | 3 | 12 | Cox | 1 | Zymoseptoria | tritici | mtDNA | | |
| gb AB151751 | 577 | Q F YL G -F S G M P-RRIHD P Y V FM G HSMS T T G -HF I TL I -G I M P | 20 | 8 | 5 | 13 | Cox | 1 | Tetrahymena | pigmentosa | mtDNA | | |
| YP_003734424 | 632 | M F FL G -F S G L P-RRIHD P FA F LG G HL G AT C G -HF L TM I -G V B | 20 | 7 | 5 | 12 | Cox | 1 | Paramecium | caudatum | mtDNA | | |
| AEV66698 | 1209 | M W L G -Y A G M P-RRV D Y P FA G GH S VIS A G-H M LN V A-G LM A | 21 | 6 | 5 | 11 | Cox | 1 | Oxytricha | trifallax | mtDNA | | |
| ACX30949 | 850 | M W L G -Y L G M P-RRV D Y P AA G GH S VL S AA H M I --T V G I L A | 21 | 6 | 5 | 11 | Cox | 1 | Moneuploites | minuta | mtDNA | | |
| AAA71993 | 472 | H L L G L- A G M P-RRIPD P DAY L H F N L ISS Y GS F VL V ST I -M F | 14 | 1 | 2 | 3 | Cox | 1 | Physarum | mtDNA | - negative control | | |
| | | End of TM12 of mitochondrial COX1 | | | | | | | | | | | |
| | | | | identities | >5* | also with COX4 | | | | | | | |

B

| Accession: | TM2 | | | |
|--------------|------|--|------|-----------------------------------|
| pdb 2YEV | 35 | -----LRFVLFMILLILLAA-GTVALMLIGKAHHIL----- | 66 | Thermus <i>caa3</i> Subunit IV |
| YP_006592792 | 52 | --FMDLRQEGVVVRL-FAAAALLWLVILLGLGALDPMTRTLPTVTIAVP----- | 97 | Methylocystis SC2 COX operon a-I |
| WP_019086563 | 46 | --FMRIITRAP-ALAFIFAVGGFLWLSILLALGSLSFTRTNFVGDILSSTAHDPRTPDTAP----- | 105 | Gluconacetobacter europaeus a-I |
| YP_002961361 | 60 | -LFMRLNRADHLVRL-TAACGLFWLAILFALTLTDTLSRLANT----- | 100 | Methylobac_extor AM1 & PA1 a-II |
| WP_009763218 | 53 | --LSLFGGFLWSRS-AAAAAATPRTDTTGAGRPHRFMALIGVSIALLFTVIIIL-LHGVALVQFCGER----- | 116 | Microvirga COX operon a-II |
| WP_014239625 | 65 | --SWRARQPGKGRADSDP-[GNRRFVAGI]-SAGTAAFFVLILAQLAAA--VFLTGCR----- | 113 | Azospirillum_bras COX operon a-II |
| YP_001769834 | 54 | -VAASLGGSYASWRACQRPEADATTVPAPDGSRATSLSDPEGGRRPHLIAAIGVLVSLLFAVVIVGVHG----- | 122 | Methylobacterium sp. 4-46 a-II |
| YP_006591292 | 56 | -LFILV-AVLVALRS-QFRSASPFPVAHSTILAAIAGATLFPALALPHCA----- | 107 | Methylocystis SC2 COX operon a-II |
| gb ABI5175 | 615 | -FFLMIFD\$HIERRACTSTTLGLPWRWKRYSY-YIFKIRYLQHNKSKMNGIP----- | 667 | COX1 Tetrahymena pigmentosa mtDNA |
| AEV66698 | 1249 | -FFIMIFD\$--LROAKAATRNFGVNRVNTRL-NFYIYEIARLTYYQOKALFLFRIIKSSNLKLNKNY----- | 1320 | COX1 Oxytricha trifallax mtDNA |

Fig. S3. Gene sequence of additional COX operons in diverse bacteria. The reference gene name for each cluster is indicated on the right of the figure. Symbols identify the same proteins as in Fig. 3A, with the addition of  protein related to nucleotide exchange factor EF-TS. These short proteins were recognised after alignment to the sequence with known 3D structure of Chain A, dimerization domain of Ef-Ts From *Thermus thermophilus* (Accession: pdb|1TFE|A) using a sequence analysis similar to that shown in Fig. S2.

Hypothetical steps in the evolution of COX operons are indicated.

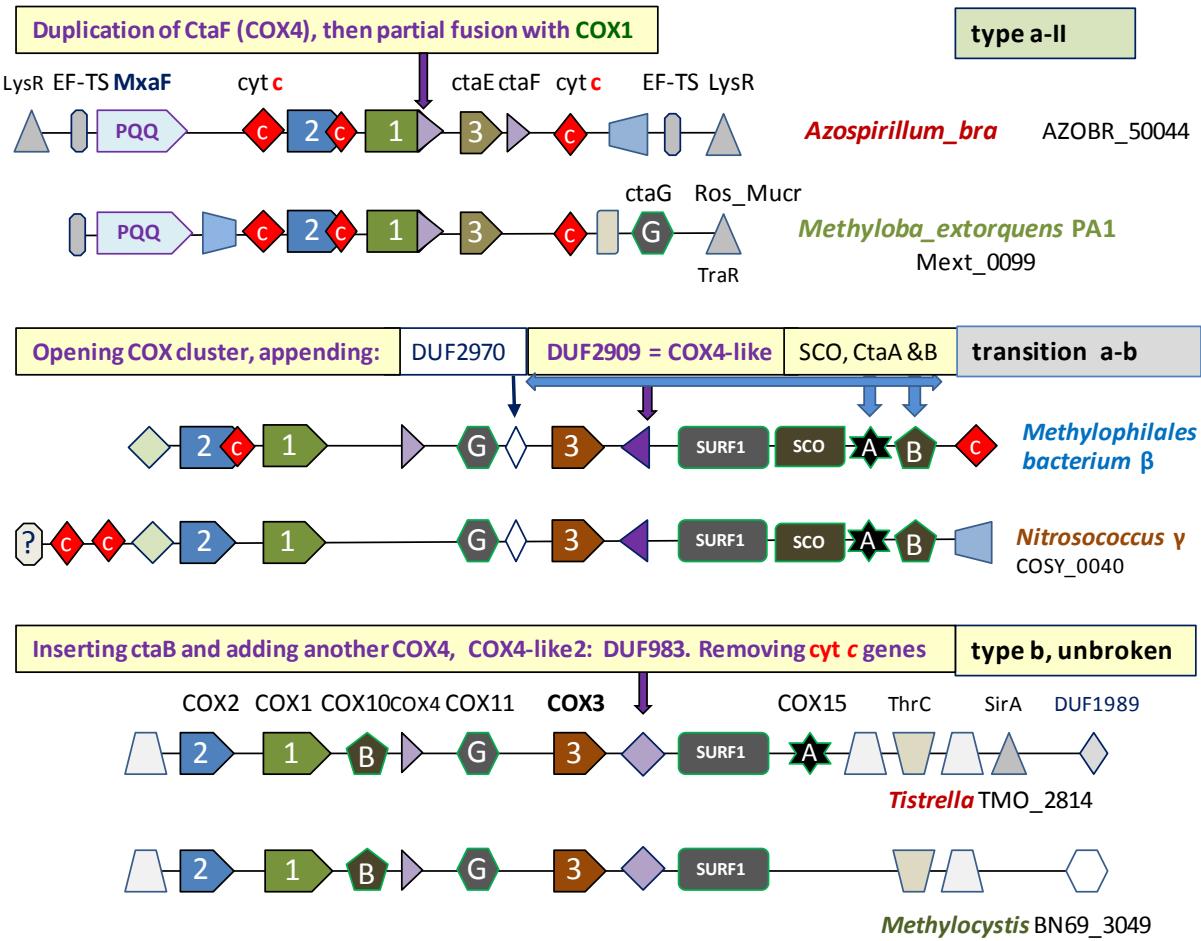


Figure S4. Structure-based alignment of bacterial and mitochondrial “Rieske” ISP.

The protein sequences of various ISP of the *bc₁* complex were aligned following structures available from various sources matching the alignment gaps or insertions with the most refined 3D data [68-71]. The limits of secondary structures (alpha helices, highlighted in green, and beta sheets, highlighted in purple) were deduced from a consensus of the latest coordinates deposited in the NCBI databanks [68-71]. Common insertions and deletions (Indels [72]) between mitochondrial and bacterial sequences are consecutively labelled CIMit1-7 (cf.

Fig. 7A). The C terminus of some sequences is truncated at the residue indicated by the numeral before the slash. Key residues for the iron-sulfur cluster, including Y165 influencing its redox potential [71], are in **bold**. Note that *Nitrospira*, *Nitrosomonas*, *Nitrosococcus* and *Methylocystis* are metabolically related by ammonia/methane autothropy. The organisms follow established phylogenetic distance from top to bottom according to the following taxonomic groups and species. **Cyanobacteria:** *Synechocystis* (*b₆f* complex), *Synechocystis* sp. PCC 6803, 192 aa; **Nitrospirales:** *Nitrospira*, *Candidatus Nitrospira defluvii* [73], 183 aa ; ***ε*-proteobacteria:** Epsilon, *Helicobacter pylori*, 167 aa; **Planctomycetes:** *Kuenenia_2*, *Candidatus Kuenenia stuttgartiensis* (in-operon Kuste3096 [66]), 173 aa ; *Schlesneria_2*, *Schlesneria paludicula* DSM 18645 (accession: ZP_11092182), 189 aa. ***γ*-proteobacteria:** *Nitrosoc*, *Nitrosococcus watsonii* C-113, 201 aa; **Frateuria**, *Frateuria aurantia*, 201 aa; ***β*-proteobacteria:** *Nitrosomonas*, *Nitrosomonas europaea* ATCC 19718, 201 aa; **Beta**, *Neisseria meningitidis* MC58, 193 aa. ***α*-proteobacteria:** *Methylocy_1* & *_2*, *Methylocystis* sp. SC2 [84], *_1* in-operon, 176 aa , *_2* in isolated gene cluster, 209 aa; *Methylob_r*, *Methylobacterium radiotolerans* JCM 2831, 189 aa; **Nitrobacter**, *Nitrobacter hamburgensis* ISP2, 219 aa; **Gluc_dia**, *Gluconacetobacter diazotrophicus* PA1 5 (in isolated gene cluster), 221 aa; **Saccharib**, *Saccharibacter* sp. (Chouaia *et al* [22]), 223 aa; *Glu_oxyd*, *Gluconobacter oxydans* H24, 218 aa; **Beijerinckia**, *Beijerinckia indica*, 172 aa; **RoseobacterA2**, *Roseobacter litoralis* petA2 in-operon, 186 aa; **Maricaulis_1**, *Maricaulis maris* in-operon, 207 aa; **Micavibrio**, *Micavibrio aeruginosavorus* [25], 185 aa; **Magnetococcus**, *Magnetococcus marinus* [76], 178 aa; **Rickettsia**, *Rickettsia felis*, 177 aa. **Mitochondria: Acanthamoeba**, *Acanthamoeba castellanii*, 235 aa; *S_cerevisiae*, *Saccharomyces cerevisiae*, mature 185 aa (3D structure available [85]); Chicken, *Gallus gallus*, mature 192 aa (3D structure available [68]). C-terminal extensions are highlighted in pale blue with some conserved residues in gray.

Rieske ISP, N-terminal part: UCR TM domain, hinge & beta1-3 plus major TURN

| | | | | | | | | | | |
|----------------------|----|---------------------------------|----------------------------------|---|---|---|--|--|---------|---------|
| <i>Synechocystis</i> | 6 | LKFRRFIMTQ | --ISGSPDPVVDLGRQQFMNL | --TFTGITGVAAGALY | --PAVKYLIPPSGGSGGGVTAKDALGNDVKV | TEFLASHNAGDRVLAQGLKGDP | | | | |
| <i>Nitrospira</i> | 18 | VGSRRTFHW | --VTVAAAVVGVLAVPLLS | --SLISPAFMRR | --RAWIDVGPVDELAPAGRPTQLDHVTVRDGW | QEATSHKAV----- | | | | |
| <i>Epsilon</i> | 3 | DIORRDFLGM | --SLASVTAIGAIASLVMKLTWDPPLPSVSAG | -- | --FTTIDVANMQEGOFSTVEWRCKPVVILKRSKKEGFNNNEKRDFKI | ----- | | | | |
| <i>Kuenenia_2</i> | 21 | TISRRNFLTL | --AGWGLFFAAIGAYLSQLFCYKGFYPKVLF | --EPSPRFFVGEPKQFPVGSVTTLKSRKIFVVVRDAD | -- | ----- | | | | |
| <i>Schlesneria_2</i> | 12 | TPPRRFLA | --ISSVVIIGFITLIPVSGIAFFLDPVLRTR | --TFKKGCDSEGFLFVANLSDLDDDGTPRLFLPDDGTPLRF | QMRADKQDAWNVV | | | | | |
| <i>Methylocy_2</i> | 8 | DPLRRDVLYV | --ATGACAVVATAAAIWPLI | --ALNPDDASSVAAGA | --PIDVFLGGLEPGKKVVRWRGLPVAIFRRPDAALEKLKDPLVLL | --ALADP | | | | |
| <i>Saccharib</i> | 22 | RPTRRVLT | --GSVALGGAGACTLAAFPVLSLRRTRASAST | --QEETVLDISVQDLEPGAHKIVWWQCMPPVAIQRRTPEMIRALEDPALLA | --QLLDP | | | | | |
| <i>Glu_oxyd</i> | 20 | GSPRRDVLAT | --VTVAGCAGACAVAYFLISLNGTRAHVGESD | --ILDVDSLTLKAGQOIVVWRCWPVFVQRTPEMLKTLDPAILO | --KLRDP | | | | | |
| <i>Gluc_dia</i> | 20 | GTGRRDFLTL | --VTLAAGGAGAAAFAWPFLDSLRS | --PVDVDVSKLPQCGQITVVWHGSPVFIHTHPTESLARLQEAALAA | --RLRDP | | | | | |
| <i>Nitrosomonas</i> | 10 | MSGRRDFLIV | --ATSVAGAGAGAVATPFLISMMP | --SERAKAAGA | --PVEVDISKLEPGMLLTAEWCRKVVVWLKRTPMELDNLEKLNSQL | --ADP | | | | |
| <i>Beta</i> | 6 | MGNGRRFLTL | --ATCGAGGAALCVAUTPFPVAFSFFP | --AGA | --AVEVDSK1EAGOLLLTAEWQCKPIVWLNRDQQQLKDLKGLNGEL | --TNSDP | | | | |
| <i>Nitrosoc</i> | 7 | DQGKRRFLTA | --ATTVVGGVGMGIFTAVPFVFSMQP | --SERAQAGA | --PVEVDISNLEPGHIMTVEWRCKPVWILRRTPEELESIEALTEED | --ILRDP | | | | |
| <i>Frateuria</i> | 6 | DGGRRFLAV | --STSVVGAGLAAGAAVBFAS | --SMQPBARARSLAA | --PQVQDLAGIAQEQMVITYAWRCOPVVFVHSAROLALLQKHDORL | --LDP | | | | |
| <i>Nitrobacter</i> | 17 | SPTRRDFLII | --ATAAVGTGVAAVTPMIQLEP | --DKA | --PAVLELSQLQVPGQSVWRWRIDPILVINRSDAALKTLQDPKLLA | --QLSDP | | | | |
| <i>Methylob_r</i> | 13 | DGSRRDFLFL | --ATGAGLAVGAGAAAWPLIA | --SMAP | --DADTAAGA | --PVEVDTIPLIQDGQIVNVFWRCKLIVFVRLTAKEVADMKA | PLS | | | |
| <i>Rickettsia</i> | 11 | QTTRRDFMVL | --TASSVAAVGAVCTLWPLVLSLNP | --SADVL | --LALS | --SIEVDSLNSIAVGQTVTIVKWKQCKPVFITNRPDKIAEARAVKMS | --ELIDP | | | |
| <i>Maricaulis_1</i> | 14 | EPSRRDFIYI | --AAGGVAAVGGGLVAVPFII | --QMNP | --AADDLALG | --SIRVDSVSVVPGSEITVMMWRGCPVFIHRHTEEEIAEARAVELD | --ALPD | | | |
| <i>RoseobacterA2</i> | 8 | EGTRRDFLYY | --ATAGAGAVATGAAIWPFLI | --QMNPA | --DASTL | --VADLALS | --SIRVDSVSVVPGSEITVMMWRGCPVFIHRHTEEEIDAARAVDIA | --DLPDQ | | |
| <i>Magnetococcus</i> | 8 | DENRRDFLIM | --ATGAVGAVGVAGAAWPFI | --SWSP | --ADAVL | --LSLAA | --TTEVSVAAIAEGQMINTMWQCKPVFILHNRNAQEINLARAKTD | DKATDLVP | | |
| <i>Methylocy_1</i> | 12 | EPSRRDILYI | --ATGAAAGAAAAGMWWPLIA | --QMNP | --DASTL | --LALA | --STEVL | --LSSVPPQIVTVKWKLCKPVFVRRRTQEEIDAARAVDIA | --ELRDP | |
| <i>Beijerinckia</i> | 8 | APTRRDFLFI | --ATGAMGVVAAGSVLWPLVLI | --QMNPA | --DASTL | --AAA | --TTEVDIGNIAEGQIVTVKWKRCKPVFVRRRTKKEISDAEAVPV | --ELRDP | | |
| <i>Micavibrio</i> | 16 | GGTRRDFLFL | --TAGAGFAGGAGIFSVPFI | --SMNP | --AADT | --LAMA | --STEVL | --LSPVEVGQSITVWRCWPVFIHRHTEEEIAARA | VDVS | --SLRDP |
| <i>Acanthamoeba</i> | 62 | ESGGRAFSYMLVLTAGVG | --YAAAKHTVIKFLI | --SMNP | --HAN | --RAMA | --NVEVIVSNTIAEQTIMTVKWRCKPLFIRHRTAEEIADAESAPLT | --DMRDP | | |
| <i>S_cerevisiae</i> | 50 | ADGRGSYAYFMVGAMGLLSSACAKSTVETFI | --SMTA | --TADV | --LAMA | -- | --KVEVNLAAIPLGKNNVVKWQCKPVFIRHRTPEHEIOEANSVDM | --ALKDP | | |
| <i>Chicken</i> | 29 | SEDRKGSYLVLTATACVATAYAKNVVTQFIS | --SSLAS | --SADVL | --LALS | -- | --KIEIKLSDIPECKNVAFKWRCKPLFVFRHRTQAEINQEAEVDVS | --KLRDP | 120 | |

Features RR 1 α turn less in bacteria
 Con.Indels CIMit1 CIMit2

Rieske ISP, C-terminal extrinsic part: Iron Sulfur cluster domain to C terminus

| | | | | | | | | | |
|----------------------|-----|--------------|-------------------------|--------------|-----------------------------|--------------------|------------------------|-------------------------|-------------|
| <i>Synechocystis</i> | | TYIVVQGD | --TIANYGINAVCTHLGCCVVPW | -- | NASENKFCMPCCHGSQYNAEGKVRG | --PAPLSSLALAHATVTD | --DKLVLSTWTETDFTD | --EDPWA | 192 |
| <i>Nitrospira</i> | | WAVKPQGD | --TVFSPICTHLGCGYRW | -- | --DDAEKKFLCPCHGSSFDVKGDLGCG | --PAPRPLDLRPAKVEG | --RRLLVMYKDFKSGLRDSVEL | | 183 |
| <i>Epsilon</i> | 79 | --GGSVFTAA | --IQCITHLGCIPTY | -- | --QDEEKFGFLCPCHGCGFTSDGVNIA | --GATPPRPF | --DPPFKI | --EGTKITFGEAGAE | .. 158/167 |
| <i>Kuenenia_2</i> | | --VRDADS | --FKAISVVC | --THLGCCAVEF | --SKEKNLIFEC | --CHGSKSYV | --RNGVNFAG | --PAPRPLDHLQML | --DNGKLAVID |
| <i>Schlesneria_2</i> | | ANQT | --IGTIYLRKMPGNQ | --IYI | --QENSKSFLCPCHGCSKFDLGER | --TNTKIPRGLD | --LTD | --SKTDISGRVWVVKYQEFOCGV | TQKGAVS 189 |
| <i>Methylocy_2</i> | | DSEVL-QQP | --RYARNWHR | --SIDP | --TIVYAVLVGVCTHLGCCP | --IYPTPSGA | --EPT | --ADWP | 173 |
| <i>Saccharib</i> | | --R | --YARNWHR | --S | --LCPHGSQF | --DQDAGVRLRN | --PAPYNL | --PVPYRMI | 199/209 |
| <i>Glu_oxyd</i> | | QSRIR-QQP | --REAAENAHRS | --LRP | --DGYVVTICTHLGCCP | --PYQNTIS | --NIT | --SDR | 212/223 |
| <i>Gluc_dia</i> | | ESCIF-QQP | --KDATNWHR | --SPD | --IGVMI | --GTC | --PHQF | --DHTIRL | 207/218 |
| <i>Nitrosomonas</i> | | SSGAL-QQP | --PYAANNWHR | --I | --KPEFGVVG | --VGI | --CTHLGCC | --PTYS | 210/221 |
| <i>Beta</i> | | DSQRD-QQP | --PYAQNHTRS | --IKPE | --I | --LVVLGV | --C | --TANWP | 201 |
| <i>Nitrosoc</i> | | NSDAE-QQP | --EYAKNETR | --S | --I | --KPNL | --V | --G | 193 |
| <i>Frateuria</i> | | QSEVESQQP | --PFAENIYRS | --I | --KPE | --LLV | --G | --C | 201 |
| <i>Nitrobacter</i> | | DSKVD-QQP | --AWARNPHRS | --QRP | --A | --WLV | --G | --P | 201 |
| <i>Methylob_r</i> | | QSSEF-QQP | --EYAHNWHRS | --SAN | --PEY | --GV | --G | --F | 210/219 |
| <i>Rickettsia</i> | | --A | --FADP | --R | --V | --G | --C | --P | 189 |
| <i>RoseobacterA2</i> | | LAENANIAP | --D | --A | --S | --DENR | --S | --E | 177 |
| <i>Maricaulis_1</i> | | DARND-NLPAGV | --P | --T | --D | --ANR | --V | --R | 186 |
| <i>Magnetococcus</i> | | QDANRVL-- | --PEFLIV | --KGN | --T | --LGC | --V | --G | 200/207 |
| <i>Micavibrio</i> | | --PEWLV | --V | --G | --I | --C | --H | --D | |
| <i>Methylocy_1</i> | | QDADR | --VKK | --D | --E | --WLV | --I | --V | 235 |
| <i>Beijerinckia</i> | | QDADR | --VKK | --D | --E | --WLV | --I | --V | 185 |
| <i>Acanthamoeba</i> | | QDADR | --VKK | --D | --E | --WLV | --I | --V | 196/272 |
| <i>S_cerevisiae</i> | | QDADR | --VKK | --D | --E | --WLV | --I | --V | |
| <i>Chicken</i> | 121 | QHDLDRV | --KK | --PEW | --V | --L | --G | --V | |

features Bac Insert C+H+C+P αβyinsert G C CHGS Y165 C-terminus
 Con.Indels CIMit 3a 3b CIMit4 CIMit5 CIMit6 CIMit7

Supporting Information Tables

These tables contain large datasets that do no fit the recommended format of the text and are therefore presented as pasted images; they are also available as Excel files, if requested.

Table S1 – Genomic distribution of bioenergetic systems in α -proteobacteria.

A. The genomes of *ca.* 120 α -proteobacterial organisms were studied from the latest version of the genome NCBI database <http://www.ncbi.nlm.nih.gov/genome/browse/> - accessed on 14 March 2014, verifying also the completeness of genomic data (*). Reconstruction of the various bioenergetic systems (see text) was deduced by combining genomic information with biochemical and microbiological data. The organisms are listed following the left-right sequence in the model of Fig. 1B. Two major types of *bd* oxidases are classified as *bd*-I or CIO [25,79]. The organisms directly shown in Fig. 1B are yellow highlighted and those proposed to be relatives of mitochondria are shown in italics with pertinent references (including [86,87]). Underlined organisms are symbionts or pathogens. **B.** This table lists the organisms that have been analysed but are not included in the model of Fig. 1B, also because they are in parallel paths of evolution with respect to the mitochondrial subset of bioenergetic systems. The organisms highlighted in pale yellow are shown in Fig. 1, while other annotations are the same as in **A**. Complementary information is in Table S2.

A

| Table S1a | Organism/Name - underlined, endocellular/parasite proposed mitochondrial relative with Reference Shown in Fig. 1B | Order/Group | BIONERGETIC SYSTEMS | | | | | NOTES | Genome* complete | Photosynthetic |
|---|---|-------------|---------------------|---------------|---------------|---------|-------|---|---------------------|----------------|
| | | | 1. bc1 | 4. aa3 | 2. bd - types | 5. cbb3 | 3. bo | | | |
| All 6 bioenergetic systems | | | | | | | | | | |
| <i>Paracoccus denitrificans</i> PD1222 [17] | Rhodobacterales | YES | YES | YES | YES | YES | NO | Nitrate and nitrite reductase AND NOR; CIO bd oxidase; bo3 as bo oxidase | YES | |
| <i>Agrobacterium tumefaciens</i> FZ [86] | Rhizobiales | YES | YES | YES -bd-I | YES | YES | NO | Nitrate and Nitrite reductase, NOR | YES | |
| <i>Ochrobactrum anthropi</i> ATCC 49188 [6,7] | Rhizobiales | YES | YES | YES -bd-I | YES | YES | NO | Nitrate and Nitrite reductase, NOR | YES | |
| <i>Bradyrhizobium</i> ORS 375 [20] | Rhizobiales | YES | YES | YES | YES | YES | NO | Nitrate and Nitrite reductase, NOR like oxidase; CIO bd oxidase; 2 ISP forms | YES | |
| <i>Rhodopseudomonas palustris</i> HoA2 [6,7] | Rhizobiales | YES | YES | YES - 2 types | YES | YES | NO | Nitrate and Nitrite reductase, NOR; CIO bd oxidase; 2 ISP forms | YES | |
| <i>Methylobacterium nodulans</i> | Rhizobiales | YES | YES | YES | YES | YES | NO | Nitrate and Nitrite reductase ; CIO bd oxidase | YES | |
| <i>Microviga</i> sp. WSM3557 | Rhizobiales | YES | YES | YES - 2 types | YES | YES | NO | Multiple COX operons; Nitrate and Nitrite reductase ; CIO bd oxidase | YES | |
| <i>Rhizobium leguminosarum</i> bv. <i>trifoli</i> WSM2304 | Rhizobiales | YES | YES | YES -bd-I | YES | YES | NO | Nitrate and nitrite reductase AND NOR | YES | |
| <i>Mesorhizobium loti</i> MAFF303099 | Rhizobiales | YES | YES | YES | YES | YES | NO | Nitrate and Nitrite reductase, NOR; CIO bd oxidase | YES | |
| <i>Brucella suis</i> 1330 | Rhizobiales | YES | YES | YES -bd-I | YES | YES | NO | Nitrate and nitrite reductase AND NOR | YES | |
| <i>Methyloccella silvestris</i> | Rhizobiales | YES | YES | YES | YES | YES | NO | YES Methane monooxygenase; Nitrate reductase; assimilatory Nitrite reductase ; CIO bd oxidase | YES | |
| <i>Hyphomicrobium denitrificans</i> ATCC 51888 | Rhizobiales | YES | YES | YES | YES | YES | NO | Nitrate and Nitrite reductase AND NOR; CIO bd oxidase | YES | |
| <i>Parvibaculum lavamentivorus</i> DS-1 | Rhizobiales | YES | YES | YES | YES | YES | NO | Nitrate and nitrite reductase AND NOR; CIO bd oxidase | YES | |
| <i>Tistrella mobilis</i> KA081020-065 [48] | Rhodospirillales | YES | YES | YES - 2 types | YES | YES | NO | Nitrate and Nitrite reductase; CIO bd oxidase | YES | |
| <i>Thalassospira profundimaris</i> WP0211 | Rhodospirillales | YES | YES | YES | YES | YES | NO | Nitrate and Nitrite reductase; CIO bd oxidase | NO | |
| <i>Rhodospirillum centenum</i> SW | Rhodospirillales | YES | YES | YES - 3 types | YES | YES | NO | Nitrate reductase periplasmic and Nitrite reductase; CIO bd oxidase; 2 ISP forms | YES | |
| <i>Caulobacter</i> [18] | Caulobacterales | YES | YES | YES -bd-I | YES | YES | NO | Nitrate and Nitrite reductase | YES | |
| <i>Asticcacaulis eccentricus</i> | Caulobacterales | YES | YES | YES -bd-I | YES | YES | NO | Nitrate and Nitrite reductase AND NOR | YES | |
| <i>Novosphingobium pentarmatovorans</i> | Sphingomonadales | YES | YES | YES -bd-I | YES | YES | NO | Nitrate and Nitrite reductase | YES | |
| minus 1, bd | | | | | | | | | | |
| <i>Sphingopyxis alaskensis</i> RB2256 | Sphingomonadales | YES | YES | NO | YES | YES | NO | assimilatory Nitrate & Nitrite reductase | YES | |
| minus 1, cbb3 | | | | | | | | | | |
| <i>Glucanacetobacter</i> europeaus LMG 18494 | Rhodospirillales | YES | YES | YES - 3 types | NO | YES | NO | Nitrate and Nitrite reductase;2 CIO bd oxidase; 2 ISP forms | YES | |
| <i>Glucanacetobacter diazotrophicus</i> PA1 5 | Rhodospirillales | YES | YES | YES | NO | YES | NO | Nitrate and Nitrite reductase; CIO bd oxidase; 2 ISP forms | YES | |
| <i>Azospirillum lipofluorium</i> 4B | Rhodospirillales | YES | YES | YES | NO | YES | NO | COX not validated as aa3; Nitrite reductase; CIO bd oxidase | YES | |
| <i>Acetobacteraceae bacterium</i> AT-5844 | Rhodospirillales | YES | YES | YES - 2 types | NO | YES | NO | Nitrate and Nitrite reductase; CIO bd oxidase | NO | |
| <i>Methylcystis</i> SC2 | Rhizobiales | YES | YES | YES - 2 bd-I | NO | YES | NO | Multiple COX operons; Methane monooxygenases; Nitrate and Nitrite reductase; 2 ISP forms | YES | |
| <i>Methylbacterium radiotolerans</i> JCM 2831 | Rhizobiales | YES | YES | YES | NO | YES | NO | Nitrate reductase periplasmic; Nitrite reductase; 3 CIO bd oxidases | YES | |
| <i>Methylbacterium extorquens</i> (4 strains) | Rhizobiales | YES | YES | YES - 2 types | NO | YES | NO | Nitrate reductase periplasmic; Nitrite reductase; 2 or 3 CIO bd oxidase | YES | |
| <i>Methylsinus trichosporinus</i> | Rhizobiales | YES | YES | YES | NO | YES | NO | Methane monooxygenase; Nitrate and Nitrite reductase | NO | |
| <i>Nitrothobacter hamburgensis</i> X14 | Rhizobiales | YES | YES | YES | NO | YES | NO | Nitrite oxidoreductase and assimilatory reductase , NOR like oxidase; CIO bd oxidase; 2 ISP forms | YES | |
| <i>Azorhizobium caulinodans</i> ORS 571 | Rhizobiales | YES | YES | YES - 2 types | NO | YES | NO | Nitrite reductase; CIO bd oxidase | YES | |
| <i>Sinorhizobium meliloti</i> AK83 | Rhizobiales | YES | YES | YES | NO | YES | NO | COX not validated as aa3; Nitrate reductase periplasmic, Nitrite reductase | YES | |
| <i>Phyllobacterium</i> sp. YR51 | Rhizobiales | YES | YES | YES -bd-I | NO | YES | NO | Nitrate and Nitrite reductase | YES | |
| <i>Pelagibacter halotolerans</i> B2 | Rhizobiales | YES | YES | YES -bd-I | NO | YES | NO | Nitrate and Nitrite reductase | YES | |
| <i>Parvularcula bermudensis</i> HTCC2503 [18] | Parvularculales | YES | YES | YES -bd-I | NO | YES | NO | Nitrate reductase; CIO bd oxidase; also subunit II of a non functional CIO bd oxidase | YES | |
| minus 1, bo | | | | | | | | | | |
| <i>Rhodobacter</i> <i>sphaeroides</i> ATCC 17025 [6-8] | Rhodobacterales | YES | YES | YES | NO | NO | NO | Nitrate and Nitrite reductase AND NOR; CIO bd oxidase; bo present in other strains | YES | |
| <i>Rhodopseudomonas palustris</i> BisA53 [6,7] | Rhizobiales | YES | YES | YES | NO | NO | NO | Nitrate and Nitrite reductase, NOR; 3 ISP forms ; CIO bd oxidase | YES | |
| <i>Xanthobacter</i> <i>autotrophicus</i> | Rhizobiales | YES | YES | YES | NO | NO | NO | Nitrate reductase and assimilatory Nitrite reductase ; CIO bd oxidase | YES | |
| <i>Afipia felis</i> | Rhizobiales | YES | YES | YES | NO | NO | NO | Nitrate and Nitrite reductase, NOR like cyt c oxidase; CIO bd oxidase; 2 ISP forms | NO | |
| <i>Brucella abortus</i> S19 | Rhizobiales | YES | YES | YES | NO | NO | NO | Nitrite reductase only | YES | |
| <i>Caenispillum</i> <i>salinarum</i> AK4 | Rhodospirillales | YES | YES | YES | NO | NO | NO | Nitrate and Nitrite reductase, NOR; 2 CIO bd oxidases; 2 ISP forms | NO | |
| <i>Sphingobium</i> sp. SYK-6 | Sphingomonadales | YES | YES | YES -bd-I | YES | NO | NO | bo oxidase present in other Sphingobium species; assimilatory Nitrate and Nitrite reductase | YES | |
| minus 2, middle | | | | | | | | | | |
| <i>Mariculis maris</i> | Rhodobacterales | YES | YES | NO | YES | NO | NO | Nitrate reductase, respiratory; 2 ISP forms | YES | |
| <i>Roseobacter</i> <i>litoralis</i> | Rhodobacterales | YES | YES | NO | YES | NO | NO | Nitrate reductase and assimilatory Nitrite reductase; 3 ISP forms | YES | |
| <i>Magnetospirillum</i> <i>magneticum</i> AMB-1 | Rhodospirillales | YES | YES | NO | YES | NO | NO | Nitrate reductase periplasmic, assimilatory Nitrite reductase ; NOR | YES | |
| <i>Micovibrio aeruginosavorus</i> ARL-13 [19] | Rhodospirillales? | YES | YES | YES | NO | NO | NO | Ectoparasite of bacteria; Nitrate reductase, respiratory; CIO bd oxidase | YES | |
| <i>Beijerinckia</i> <i>indica</i> subsp. <i>indica</i> | Rhizobiales | YES | YES | YES | NO | NO | NO | Nitrate reductase and assimilatory Nitrite reductase ; CIO bd oxidase | YES | |
| minus 2, bottom | | | | | | | | | | |
| <i>Rhodospirillum rubrum</i> [20] | Rhodospirillales | YES | NO | YES -bd-I | YES | NO | NO | Nitrite reductase in various strains of the species; NOR; 2 ISP forms | YES | |
| <i>Rhodospirillum photometricum</i> DSM 122 | Rhodospirillales | YES | NO | YES -bd-I | YES | NO | NO | Nitrite reductase only; 2 ISP form | YES | |
| <i>Oligotropha carboxidovorans</i> OM4 | Rhizobiales | YES | NO | YES | YES | NO | NO | Nitrate reductase respiratory, Nitrite reductase; CIO bd oxidase | YES | |
| minus 3, top | | | | | | | | | | |
| <i>Rickettsia prowazekii</i> Rp22 [4-7.87] | Rickettsiales | YES | YES | NO | NO | NO | NO | NO N metabolism; CIO bd oxidase | YES | |
| <i>Rickettsia</i> <i>ellis</i> UNWVAC02 [4-7.88] | Rickettsiales | YES | YES | NO | NO | NO | NO | NO N metabolism; CIO bd oxidase | YES | |
| <i>Wolbachia</i> sp. wRi | Rickettsiales | YES | YES | NO | NO | NO | NO | NO N metabolism; CIO bd oxidase | YES | |
| <i>Wolbachia</i> <i>endosymbiont</i> of <i>Drosophila melanogaster</i> | Rickettsiales | YES | YES | NO | NO | NO | NO | NO N metabolism; CIO bd oxidase | YES | |
| <i>Candidatus</i> <i>Odyssea</i> <i>thesallonicensis</i> L13 | Rickettsiales | YES | YES | YES -bd-I | NO | NO | NO | NO N metabolism | NO | |
| <i>alpha</i> <i>proto</i> <i>bacterium</i> BA1199 | Rickettsiales | YES | YES | YES | NO | NO | NO | CIO bd oxidase | NO | |
| Mitochondrion, fungi & protists | eukaryotes | YES | YES | NO | NO | NO | NO | Assimilatory Nitrate and Nitrite reductases | YES | |
| minus 3, bottom | | | | | | | | | | |
| <i>Magnetococcus</i> <i>marinus</i> MC-1 | Magnetococcales | YES | NO | NO | YES | NO | NO | COX1 similar to NOR, but COX2 and COX3 missing; Nitrate reductase periplasmic | YES | |
| <i>Candidatus</i> <i>Midichlorian</i> <i>mitochondrii</i> IrlcVA [21] | Rickettsiales | YES | NO | YES | YES | NO | NO | NO N metabolism; CIO bd oxidase | YES | |
| <i>Candidatus</i> <i>Endolsooclinum</i> <i>faukneri</i> LS | Rhodospirillales | YES | NO | NO | YES | NO | NO | Nitrite reductase | YES | |
| minus 4, middle | | | | | | | | | | |
| <i>Anaplasma marginale</i> str. St. Maries | Rickettsiales Anap. | YES | YES | NO | NO | NO | NO | NO N metabolism | YES | |
| <i>Wolbachia</i> <i>endosymbiont</i> of <i>Culex quinquefasciatus</i> NIV | Rickettsiales Anap. | YES | YES | NO | NO | NO | NO | NO N metabolism | YES | |
| <i>Ehrlichia canis</i> str. Jake | Rickettsiales Anap. | YES | YES | NO | NO | NO | NO | NO N metabolism | YES | |
| <i>Neorickettsia</i> <i>ristici</i> str. Illinois | Rickettsiales Anap. | YES | YES | NO | NO | NO | NO | NO N metabolism | YES | |
| <i>Candidatus</i> <i>Pelagibacter</i> <i>ubique</i> HTCC1062 [13,14] | SAR clade | YES | YES | NO | NO | NO | NO | NO N metabolism - a sulfite reductase remotely related to assimilatory nitrite reductases | YES | |
| <i>Candidatus</i> <i>Puniceispirillum</i> <i>marinum</i> IMCC1322 [13,14] | SAR11 clade | YES | YES | NO | NO | NO | NO | NO N metabolism | NO | |
| SAR116 cluster HMB100 [13] | SAR11 clade | YES | YES | NO | NO | NO | NO | NO N metabolism | | |
| Mitochondrion - metazoans | eukaryotes | YES | YES | NO | NO | NO | NO | NO N metabolism | YES | |

B

| Organism/Name shown in Fig. S1 | Order/Group | BIOENERGETIC SYSTEMS | | | | | Nitrogen (N) metabolism [see also Table 1]; NOR, nitric oxide reductase; CIO, Cyanide-insensitive bd oxidase [79] | Genome* complete | Photosynthetic |
|---|------------------|----------------------|--------|---------------|---------|-------|---|---------------------|----------------|
| | | 1. bci | 4. aa3 | 2. bd -types | 5. cbb3 | 3. bo | | | |
| All 6 bioenergetic systems | | | | | | | | | |
| <i>Starkeya novella</i> DSM 506 | Rhizobiales | YES | YES | YES | YES | YES | Nitrate and Nitrite reductase; CIO bd oxidase | YES | |
| <i>Rhodovulum</i> sp. PH10 | Rhodobacterales | YES | YES | YES - 2 types | YES | YES | Nitrite reductase; 2 CIO bd oxidases; 3 ISP forms | NO | YES |
| <i>Azospirillum</i> sp. BS10 | Rhodospirillales | YES | YES | YES -bd-I | YES | YES | Nitrate reductase periplasmic and Nitrite reductase | YES | |
| <i>Roseomonas cervicalis</i> ATCC 49957 | Rhodospirillales | YES | YES | YES - 3 types | YES | YES | NOR; 2 CIO bd oxidases | NO | |
| <i>Thalassospira xiamenensis</i> M-5 = DSM 17429 | Rhodospirillales | YES | YES | YES | YES | YES | Nitrate reductase periplasmic, assimilatory Nitrite reductase; CIO bd oxidase | NO | |
| minus 1 | | | | | | | | | |
| <i>Azospirillum amazonense</i> Y2 | Rhodospirillales | NO | YES | YES - 2 types | YES | YES | No ISP in bci1; Nitrate reductase periplasmic, Nitrite reductase; CIO bd oxidase | NO | |
| <i>Azospirillum brasiliense</i> Sp245 | Rhodospirillales | NO | YES | YES | YES | NO | NO cytochrome b nor c1; Nitrate reductase periplasmic, Nitrite reductase; CIO bd oxidase | YES | |
| <i>Roseomonas</i> sp. BS | Rhodospirillales | YES | YES | YES - 2 types | NO | YES | Nitrate and Nitrite reductase; CIO bd oxidase | NO | |
| <i>Magnetospirillum gryphiswaldense</i> MSR-1 v2 | Rhodospirillales | YES | YES | YES | YES | NO? | COX possibly working also as ubiquinol oxidase; Nitrate and nitrite reductase; CIO bd oxidase | YES | |
| <i>Oceanibaculum indicum</i> P24 | Rhodospirillales | YES | YES | YES -bd-I | YES | NO | Nitrate reductase | NO | |
| <i>Glucanacetobacter</i> oboediens 174Bp2 | Rhodospirillales | YES | NO | YES - 3 types | YES | YES | assimilatory Nitrite reductase; 2 CIO bd oxidases; 2 ISP forms | NO | |
| <i>Fulvimonas pelagi</i> | Rhizobiales | YES | YES | YES | NO | YES | Nitrate and Nitrite reductase; CIO bd oxidase | NO | YES |
| <i>Polyorphomium</i> gilvum SL03B-26A1 | Polyphymorph | YES | YES | YES - 2 types | YES | NO | Nitrate and Nitrite reductase; CIO bd oxidase | YES | |
| minus 2 | | | | | | | | | |
| <i>Acetobacter pomorum</i> | Rhodospirillales | YES | NO | YES - 2 types | NO | YES | Nitrate reductase; CIO bd oxidase | NO | |
| <i>Acetobacter pasteurianus</i> IFO 3283-01-42C (9 strains) | Rhodospirillales | YES | NO | YES - 2 types | NO | YES | Nitrate reductase and assimilatory Nitrite reductase ; CIO bd oxidase | YES | |
| <i>Glucanacetobacter xylinus</i> NBRC 3288 | Rhodospirillales | YES | NO | YES | NO | YES | Nitrate and Nitrite reductase ; CIO bd oxidase; 2 ISP forms | YES | |
| <i>Glucanacetobacter oxydans</i> G21H | Rhodospirillales | YES | NO | YES | NO | YES | COX not functional; assimilatory Nitrite reductase; CIO bd oxidase | YES | |
| <i>Glucanobacter morifer</i> | Rhodospirillales | YES | NO | YES | NO | YES | COX not functional; Nitrite reductase; 2 CIO bd oxidases | NO | |
| <i>Saccharibacter</i> sp. | Rhodospirillales | YES | NO | YES | NO | YES | COX not functional; Nitrite reductase; CIO bd oxidase | YES | |
| <i>Saccharibacter florcola</i> | Rhodospirillales | YES | NO | YES -bd-I | NO | YES | COX not functional; Nitrite reductase | | |
| <i>Commenzialibacter intestini</i> A911 | Rhodospirillales | YES | NO | YES -bd-I | NO | YES | COX not functional; Nitrite reductase; CIO bd oxidase; 2 ISP forms | NO | YES |
| <i>Acidiphilum</i> cryptum JF-5 | Rhodospirillales | NO | YES | YES 2 types | NO | YES | cytochrome b deranged and NO ISP; assimilatory Nitrate and Nitrite reductase; CIO bd oxidase | YES | |
| <i>Acidocella</i> | Rhodospirillales | NO | YES | YES 2 types | NO | YES | cytochrome b & ISP deranged; respiratory Nitrate reductase & assimilatory Nitrite reductase; CIO bd oxidase | YES | |
| <i>Granulibacter bethesdensis</i> CGDNH1 | Rhodospirillales | NO | YES | YES | NO | YES | assimilatory Nitrate and Nitrite reductase; CIO bd oxidase | YES | |
| <i>Brevundimonas diminuta</i> | Caulobacterales | YES | YES | YES -bd-I | NO | YES | NO metabolism | NO | |
| <i>Phaeospirillum molischianum</i> | Rhodospirillales | YES | NO | YES -bd-I | YES | NO | Nitrite reductase only; 2 ISP forms | YES | YES |
| <i>Nitratireducotor indicus</i> C115 | Rhizobiales | YES | NO | YES -bd-I | YES | NO | assimilatory Nitrate reductase, Nitrite reductase | NO | |
| <i>Rhodomicrobium vanneleii</i> ATCC 17100 | Rhizobiales | YES | NO | YES | YES | NO | Nitrite reductase; CIO bd oxidase | YES | YES |
| <i>Magnetospirillum magnetotacticum</i> | Rhodospirillales | YES | YES | NO | YES | NO? | COX possibly working also as ubiquinol oxidase; Nitrate and nitrite reductase | NO | |
| minus 3 | | | | | | | | | |
| <i>Acetobacter aceti</i> | Rhodospirillales | NO | NO | YES | NO | YES | COX 1 short; Nitrite reductase; 2 CIO bd oxidases, 1 partial | NO | |
| <i>Asaia</i> sp. platycody | Rhodospirillales | NO | NO | YES -bd-I | NO | YES | COX not functional; assimilatory Nitrite reductase | NO | |
| <i>Asaia</i> sp. SF2.1 | Rhodospirillales | NO | NO | YES | NO | YES | nitrite and nitrate reductase; CIO bd oxidase | | |
| <i>Glucanacetobacter hansenii</i> ATCC 23769 | Rhodospirillales | YES | NO | NO | NO | YES | Nitrite reductase | YES | |
| <i>Zymomonas mobilis</i> subsp. <i>mobilis</i> ZM4 | Sphaeromonadales | YES | NO | YES | NO | NO | Nitrite/sulfite reductase only; CIO bd oxidase | YES | |
| <i>Bartonella tamiae</i> (2 strains) | Rhizobiales | YES | NO | YES | NO | YES | NO N metabolism - fragments of proteins related to Nitrite reductase; CIO bd oxidase | NO | |
| minus 4 | | | | | | | | | |
| <i>Orientia tsutsugamushi</i> str. Boryong | Rickettsiales | YES | YES | NO | NO | NO | NO N metabolism | YES | |
| <i>Rickettsia</i> typhi str. Wilmington | Rickettsiales | YES | NO | YES | NO | NO | NO metabolism; CIO bd oxidase | YES | |
| <i>Bartonella grahamii</i> as4app | Rhizobiales | YES | NO | NO | NO | YES | NO N metabolism | YES | |
| minus 5 | | | | | | | | | |
| <i>Bartonella quintana</i> (2 strains) | Rhizobiales | NO | NO | NO | NO | YES | NO N metabolism | YES | |
| <i>Bartonella claridgeiae</i> 73 | Rhizobiales | NO | NO | NO | NO | YES | NO N metabolism | YES | |
| <i>Bartonella bacilliformis</i> KC583 | Rhizobiales | NO | NO | NO | NO | YES | NO N metabolism | YES | |
| <i>Bartonella henselae</i> | Rhizobiales | NO | NO | NO | NO | YES | NO N metabolism | YES | |
| <i>Candidatus Liberibacter asiaticus</i> str. psy62 | Rhizobiales | NO | NO | NO | NO | YES | NO N metabolism | YES | |

Table S2. Diverse gene clusters for aa_3 -type oxidase in α -proteobacteria. The table lists the diverse types of *COX* operons (Fig. 3A). *COX1* proteins recognised as ba3-like_Oxidase_I [cd01660] [41] are under the column ba3[^] and correspond to class B [26]. Concatenated operons are framed in blue and connected by a thick line. Incomplete (or ‘dead’ [82]) operons, indicated by the asterisk*, lack one or more of core subunits *ctaC-E* (Fig. 3A). Functional capacity of the oxidase has been deduced also from biochemical studies [88,89].

| Group and organism | COX operon $\alpha\alpha_3$ -type cytochrome c oxidase | | | | | type b | a-b transition | NOTES |
|---|--|-----|------------|-------|------------------|----------------------------|----------------|--|
| | type a | a-I | a-II | a-III | ba3 ^A | | | |
| Beta proteobacteria | | | | | yes* | | | |
| <i>Nitrosomonas europea</i> | yes | | yes | | YES | | yes | |
| <i>Burkholderia xenovorans</i> LB400 | yes | | yes | | | | yes | |
| <i>Variovorax paradoxus</i> S110 | | | | | | | yes | |
| <i>Methylophilales bacterium</i> HTCC2181 | | | | | | | yes | |
| Alpha proteobacteria | | | | | | | | |
| Rhodospirillales | | | | | | | | |
| <i>Gluconacetobacter europeus</i> LMG 18494 | | yes | | | | yes | | |
| <i>Gluconacetobacter diazotrophicus</i> PAI 5 | yes | yes | | | | yes | | |
| <i>Acetobacteraceae bacterium</i> AT-5844 | yes | | yes | yes | | yes | | |
| <i>Roseomonas</i> sp. B-5 | yes | yes | | | | yes | | incomplete genome |
| <i>Roseomonas cervicalis</i> | yes | | | | | yes | | incomplete genome |
| <i>Tistrella mobilis</i> | | yes | yes | | | yes | | |
| <i>Caenospirillum salinarum</i> | | yes | | | YES | yes | | |
| <i>Magnetospirillum gryphiswaldense</i> MSR-1 | yes - dead* | | | | yes* | yes | | |
| <i>Magnetospirillum magnetotacticum</i> | | | | | yes* | yes - 1 dead* & 1 complete | | |
| <i>Magnetospirillum magneticum</i> AMB-1 | | | | | yes* | yes - 1 dead* & 1 complete | | |
| <i>Magnetospirillum</i> sp. SO-1 | | | | | yes* | yes - 1 dead* & 1 complete | | |
| <i>Azospirillum brasiliense</i> | yes | | yes | | | yes | | |
| <i>Azospirillum lipoferum</i> | yes | | | | | yes | | |
| <i>Rhodospirillum centenum</i> SW | yes | | | | | yes | | |
| <i>Azospirillum amazonense</i> | | | | | | yes | | |
| Rickettsiales | | | | | | | | |
| <i>Anaplasma phagocytophilum</i> HZ | | | | | | yes - 2 (1 split, 1 dead*) | | |
| <i>Ehrlichia ruminantium</i> str. Welgevonden | | | | | | yes - 2 (1 split, 1 dead*) | | |
| Rhizobiales | | | | | | | | |
| <i>Nitrobacter winogradsky</i> | yes | | | | | yes - 2 | | |
| <i>Nitrobacter hamburgensis</i> | | yes | | | | yes | | |
| <i>Bradyrhizobium</i> sp. S2321 | | | | | YES | yes | | plus other 3 <i>Bradyrhizobium</i> species |
| <i>Bradyrhizobium</i> sp. WSM471 | yes - dead* | | yes | yes | YES | yes | | |
| <i>Bradyrhizobium</i> sp. ORS 375 | | yes | yes - 2 | | | yes | | |
| <i>Bradyrhizobium japonicum</i> USDA 6 | | | yes | | | yes | | plus other 3 <i>Bradyrhizobium</i> species |
| <i>Bradyrhizobium</i> sp. ORS 285 | | | yes | | | yes | | |
| <i>Bradyrhizobium</i> sp. ORS 278 | | | | | YES | yes | | |
| <i>Rhodopseudomonas palustris</i> BisA53 (& DX-1) | | | | | | yes | | |
| <i>Afipia felis</i> | | | | | | yes | | plus other 3 <i>Rhodopseudomonas</i> strain |
| <i>Afipia birigiae</i> | | | | | | yes | | incomplete genome |
| <i>Methylocystis</i> sp. SC2 | | | | | | yes | | incomplete genome |
| <i>Methylocystis parvus</i> | | | | | | yes | | |
| <i>Methylosinus trichosporium</i> | | | | | | yes | | |
| <i>Microvirga</i> | | | | | | yes | | |
| <i>Methylobacterium extorquens</i> PA1 | | | | | | yes | | |
| <i>Methylobacterium extorquens</i> DM4 | | | | | YES | yes | | plus other 3 <i>Methylobacterium</i> species |
| <i>Methylobacterium</i> sp. 4-46 | | | | | | yes | | |
| <i>Methylobacterium nodulans</i> | | | | | | yes | | |
| <i>Methylobacterium radiotolerans</i> | | | | | | yes | | |
| <i>Methylobacterium populi</i> | | | | | | yes | | plus another <i>Methylobacterium</i> species |
| <i>Chelatococcus</i> sp. GW1 | yes - dead* | | yes | yes | | yes | | incomplete genome |
| <i>Methylocella silvestris</i> | | | yes | | | yes | | |
| <i>Methyloferula stellata</i> | | | yes | | | yes | | |
| <i>Beijerinckia indica</i> | | | | | yes* | yes | | incomplete genome |
| <i>Starkeya novella</i> | | | | | | yes | | |
| <i>Pseudaminobacter salicylatoxidans</i> | | | | | | yes | | |
| <i>Aurantimonas manganoxydans</i> | | | | | | yes | | |
| <i>Pelagibacterium halotolerans</i> B2 | | | | | | yes | | |
| <i>Parviculum</i> lavamintivorans DS-1 | | | | | | yes | | |
| <i>Sinorhizobium meliloti</i> 1021 | | | | | | yes | | |
| <i>Chelatovorans</i> sp. BC1 | | | | | | yes | | |
| <i>Sinorhizobium medicae</i> WSM419 | | | | | | yes | | |
| <i>Mesorhizobium opportunistum</i> WSM2075 | | | | | | yes | | plus other 3 <i>Mesorhizobium</i> species |
| <i>Rhizobium leguminosarum</i> bv. trifolioli WSM1325 | yes | yes | yes | yes | | yes | | plus other 5 <i>Rhizobium</i> species |
| <i>Rhizobium etli</i> CFN 42/mimosae | | | | | YES | yes | | plus other 4 <i>Rhizobium</i> species |
| <i>Mesorhizobium ciceri</i> biovar biserrulae WSM1271 | | | | | | yes | | |
| Rhodobacterales | | | | | | | | |
| <i>Rhodobacter sphaeroides</i> 2.4.1 | yes | | | | | yes | | |
| <i>Oceanoballus guishaninsula</i> | | yes | | | | yes | | |
| <i>Rhodovulum</i> sp. PH10 | | yes | | | | yes | | |
| <i>Roseobacter litoralis</i> | | | | | | yes | | |
| Other groups | | | | | | | | |
| <i>Brevundimonas subvibrioides</i> | yes | | | | | yes | | |
| <i>Phenyllobacterium zucineum</i> HLK1 | yes | | | | | yes | | |
| <i>Sphingobium</i> sp. SYK-6 | yes | | | | | yes | | |
| <i>Sphingobium chlorophenolicum</i> L-1 | | | yes, dead* | yes | | yes | | plus other 4 <i>Sphingobium</i> species |
| <i>Sphingobium</i> sp. UT265 | | | | | | yes | | |
| <i>Novosphingobium</i> pentaromaticivorans | | | | | | yes | | |
| <i>Novosphingobium</i> sp. PP1Y | | | | | | yes | | |
| <i>Candidatus Pelagibacter</i> sp. IMCC9063 | | | | | yes | yes | | plus several marine metagenome organisms |
| Micavirio | | | | | | yes | | |

Table S3 – Phylogenetic distribution of the main characters of *COX* gene operons.

We constructed a matrix of 11 independent characters (indicated concisely on top of the columns) that could differentiate the gene sequence of *COX* subunits in the mitochondria of some protists from the gene sequence of bacterial *COX* operons (Table S2). The cumulative phenetic analysis indicate that *COX* operon type a-II of methylotrophs and *Tistrella* (highlighted) share the largest number of characters with *COX* gene clusters of protist mitochondria (F. Comandatore and C. Bandi, unpublished).

| organism and COX1 length | | | | | | | | | | | | | | total characters vs. mitos |
|---------------------------------------|-----------------|----------------|---------|-------------|----------------------|---------------|-----------|-------------|-----------------|------------|---------|---------------------------|----|----------------------------|
| | CHARACTERS--> | 1 | 2 | CtaC-F core | 3 | 4 | 5 | 6 | 7 | 8 | 9 | 10 | 11 | |
| | COX operon type | COX2COX1 | COX3 in | E90 in COX3 | Cox3 fused with COX1 | COX3 with 7TM | COX11COX3 | COX4 NOT in | COX4 fused COX1 | COX2 fused | no ctAB | in operon ABC transporter | | |
| gamma proteobacteria | | | | | | | | | | | | | | |
| Methylococcus capsulatus, | 541 aa | transition a-b | 1 | 1 | 0 | 0 | 0 | 1 | 0 | 0 | 1 | 1 | 0 | 5 |
| Methylbacter marinus, | 642 aa | a | 1 | 1 | 0 | 1 | 0 | 0 | 0 | 1 | 1 | 1 | 0 | 6 |
| beta proteobacteria | | | | | | | | | | | | | | |
| Methylophilales bacterium , | 531 aa | transition a-b | 1 | 0 | 1 | 0 | 0 | 0 | 0 | 0 | 1 | 0 | 0 | 3 |
| Variovorax paradoxus, | 662 aa | a-ll | 1 | 1 | 1 | 0 | 0 | 0 | 0 | 1 | 1 | 1 | 1 | 6 |
| alpha proteobacteria | | | | | | | | | | | | | | |
| Acidiphilum cryptum | | transition a-b | 1 | 0 | 1 | 0 | 1 | 0 | 0 | 0 | 0 | 0 | 0 | 3 |
| Rhodopseudomonas palustris Ha2, | | a | 1 | 1 | 1 | 0 | 1 | 0 | 0 | 0 | 0 | 0 | 0 | 4 |
| Methylocystis sp. SC2, | 636 aa | a-ll | 1 | 1 | 1 | 0 | 0 | 0 | 0 | 1 | 1 | 1 | 0 | 7 |
| Mesorhizobium opportuniticum, | 658 aa | a-ll | 1 | 1 | 1 | 0 | 0 | 0 | 0 | 1 | 1 | 1 | 0 | 7 |
| Microvirga sp. WSM3558, | 670 aa | a-ll | 1 | 1 | 1 | 0 | 0 | 0 | 0 | 1 | 1 | 1 | 0 | 7 |
| Methylocella sylvestris, | 650 aa | a-ll | 1 | 1 | 1 | 0 | 0 | 0 | 0 | 1 | 1 | 1 | 1 | 7 |
| Methylobacterium extorquens PA1, | 638 aa | a-ll | 1 | 1 | 1 | 0 | 0 | 0 | 1 | 1 | 1 | 1 | 1 | 9 |
| Rhizobium leguminosarum bv. Trifolii, | 637 aa | a-ll | 1 | 1 | 1 | 0 | 0 | 0 | 1 | 1 | 1 | 1 | 0 | 8 |
| <i>Tistrella mobilis</i> , | 839 aa | a-ll | 1 | 1 | 1 | 1 | 0 | 0 | 0 | 1 | 1 | 1 | 0 | 7 |
| Azospirillum brasiliense, | 645 aa | a-ll | 1 | 1 | 1 | 0 | 0 | 0 | 0 | 1 | 1 | 1 | 0 | 6 |
| Beijerinckia indica subsp. indica, | 555 aa | b | 1 | 0 | 1 | 0 | 1 | 1 | 1 | 0 | 0 | 0 | 0 | 5 |
| Micavibrio aeruginovorus, | 559 AA | b | 1 | 0 | 1 | 0 | 1 | 1 | 1 | 0 | 0 | 0 | 0 | 5 |
| Magnetspirillum magneticum AMB-1 | | b | 1 | 0 | 1 | 0 | 1 | 1 | 1 | 0 | 0 | 0 | 0 | 5 |
| Mariculis maris | | b | 1 | 0 | 1 | 0 | 1 | 1 | 1 | 0 | 0 | 0 | 0 | 5 |
| Rhoseobacter litoralis | | b | 0 | 0 | 1 | 0 | 1 | 1 | 0 | 0 | 0 | 0 | 0 | 3 |
| Wolbachia wrl | | b | 1 | 0 | 1 | 0 | 1 | 0 | 1 | 0 | 0 | 0 | 0 | 4 |
| Rickettsia felis | | b | 0 | 0 | 1 | 0 | 1 | 0 | 1 | 0 | 0 | 0 | 0 | 3 |
| eukaryotes (mitochondria) | | | | | | | | | | | | | | |
| Dictyostelium | | | 1 | 0 | 1 | 0 | 1 | 0 | 1 | 0 | 1 | 1 | 0 | 6 |
| Jakobida | | | 1 | 0 | 1 | 0 | 1 | 1 | 0 | 0 | 0 | 1 | 1 | 7 |
| Tetrahymena | | | 1 | 0 | 1 | 0 | 0 | 0 | 0 | 1 | 0 | 1 | 0 | 4 |
| Oxytrichia | | | 1 | 1 | 1 | 1 | 1 | 0 | 0 | 1 | 1 | 1 | 0 | 8 |
| CHARACTERS | | 1 | 4 | 6 | 3 | 5 | 7 | 8 | 8 | 9 | 11 | | | |

Table S4 – Conserved phospholipid binding sites in COX3 proteins.

The alignment in Fig. 4A was enlarged and the residues corresponding to the PL-binding sites and E90 (close to O2 entry in beef COX3 [60]) were considered conserved when producing positive substitutions [83] (bold amino acid symbols in white background). Other substitutions are highlighted in pale brown while identities are identified as yes. Organisms are abbreviated as in Fig. 4.

| in phylogenetic/phenetic order | | Organism and protein | protein | Accession | length (aa) | PL binding sites (beef numbering) | | | | | | | | | | | | PE2 | | | | | | | | | | | | PL binding total* |
|------------------------------------|----------------|----------------------|---------|-----------|----------------|-----------------------------------|-----|-----|-----|------|------|--------|--------|--------|--------|------|------|------|------|------|-----------|--------|--------|-----------|--|--|--|--|--|----------------------|
| O2 | E90 | | | | | PG1 | W57 | W58 | E64 | F86 | H71 | T66 | F214 | R221 | F/V233 | G234 | Y241 | H231 | Y181 | Y182 | I185 | F198 | F203 | G82 & M83 | | | | | | |
| <i>Methylocystis</i> sp. SC2 | COX1-3 | YP_006591291 | 827 | L | yes | yes | N | G | L | P | C | A | T | S | L | P | yes | A | gap | A | A | 0 | 3 | | | | | | | |
| <i>Rhodo_palu_BisA53</i> | COX1-3 | YP_782773 | 841 | yes | P | G | A | G | R | Y | gap | S | A | M | I | T | M | L | G | yes | 0 | 2 & 2 | | | | | | | | |
| <i>Methylocystis</i> sp. SC2 | COX3a | YP_006592539 | 236 | V | M | E | yes | V | yes | gap | A | K | T | R | yes | L | W | R | gap | Y | V | 0 | 3 & 3 | | | | | | | |
| <i>Variovorax_parEPS</i> | COX3b | WP_018905232 | 234 | D | gap | gap | gap | yes | Y | gap | yes | K | S | I | yes | N | F | S | gap | yes | yes | 0 | 5 & 4 | | | | | | | |
| <i>Methylocystis</i> sp. SC2 | COX3 | YP_006592793 | 213 | yes | gap | gap | gap | I | yes | G | L | K | N | S | yes | R | W | R | A | yes | L | 0 | 3 & 3 | | | | | | | |
| <i>Roseomonas</i> sp. B5 | COX3 | WP_019459297 | 217 | yes | gap | gap | A | yes | yes | E | V | Q | M | P | yes | Q | yes | H | gap | Y | L | 1 | 5 & 4 | | | | | | | |
| <i>Glucanacetobacter_europaeus</i> | COX3 | WP_019086564 | 210 | yes | gap | gap | yes | yes | yes | P | L | yes | P | A | yes | R | W | H | A | Y | L | 2 | 7 & 4 | | | | | | | |
| <i>Variovorax_parS110</i> | COX3 | YP_002942615 | 201 | yes | gap | gap | gap | L | yes | gap | D | L | V | D | yes | R | F | A | W | W | L | 1 | 3 & 4 | | | | | | | |
| <i>Methylocystis</i> sp. SC2 | COX3 | YP_006593114 | 242 | yes | gap | gap | gap | yes | Y | gap | D | L | S | D | yes | R | F | A | W | W | L | 1 | 3 & 4 | | | | | | | |
| <i>Azospirillum_brasiliense</i> | COX3 | YP_005030787 | 202 | yes | gap | gap | gap | yes | yes | gap | D | L | Q | D | yes | R | F | T | W | W | yes | 1 | 5 & 3 | | | | | | | |
| <i>Nitrosococcus_oceani</i> | COX3 | YP_345015 | 288 | yes | yes | F | yes | yes | gap | E | M | yes | yes(F) | A | yes | yes | yes | G | D | Y | yes | 2 | 11 & 3 | | | | | | | |
| <i>Methylophili_bac</i> | COX3 | WP_008107990 | 299 | yes | yes | F | yes | yes | K | R | M | yes | yes(F) | A | yes | D | yes | M | N | Y | yes | 2 | 10 & 3 | | | | | | | |
| <i>Thermus_thermophilus</i> | COX1-3 | YP_143578 | 791 | yes | yes | A | yes | yes | yes | S | V | L | gap | T | yes | yes | F | yes | H | W | yes | 2 | 10 & 3 | | | | | | | |
| mitochondria | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | |
| <i>Plasmadium_yoelii</i> | COX3 | XP_729225 | 263 | yes | F | V | yes | T | gap | L | yes | D | yes(V) | V | yes | S | L | G | gap | H | T | 2 | 6 & 2 | | | | | | | |
| <i>Monoeuploites_minuta</i> | COX1-3 | ACX30949 | 1203 | S | gap | F | L | I | gap | yes | Y | L | I | T | V | F | F | D | L | yes | 1 | 4 & 5 | | | | | | | | |
| <i>Oxytricha_trifallax</i> | COX1-3 | AEV66698 | 1331 | S | gap | F | Y | gap | F | I | Y | yes | Y | N | yes | W | F | D | yes | yes | N | 1 | 5 & 6 | | | | | | | |
| alpha, sister group of mitos | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | |
| <i>Beijerinckia_indica</i> | COX3 | YP_001831864 | 287 | yes | yes | yes | yes | yes | yes | E | yes | yes | yes | yes | yes | Q | F | A | D | yes | yes | 2 | 14 & 3 | | | | | | | |
| <i>Caenispirillum</i> | COX3 | WP_009539949 | 267 | yes | yes | yes | yes | yes | yes | M | yes | yes | S | I | W | R | yes | G | A | Y | yes | 2 | 11 & 3 | | | | | | | |
| <i>Ricket_prow_Rp22</i> | COX3 | YP_005998599 | 278 | yes | yes | yes | yes | yes | yes | I | yes | yes | L | yes | yes | G | yes | yes | E | Y | yes | 2 | 14 & 1 | | | | | | | |
| <i>Wolbachia_Dra_sim</i> | COX3 | YP_007886075 | 275 | yes | yes | yes | yes | yes | yes | I | yes | yes | L | C | yes | D | yes | yes | F | Y | yes | 1 | 11 & 1 | | | | | | | |
| <i>Paracoccus</i> | COX3 | YP_918077 | 273 | yes | yes | yes | yes | yes | yes | E | yes | yes | yes | yes | yes | Q | yes | S | L | Y | yes | 2 | 14 & 4 | | | | | | | |
| <i>Roseobacter</i> | COX3 | YP_004690241 | 273 | yes | yes | yes | yes | yes | yes | K | yes | yes | yes | yes | yes | K | yes | S | L | yes | yes | 2 | 15 & 2 | | | | | | | |
| <i>Tistrella</i> | COX3 | YP_006372231 | 269 | yes | yes | yes | yes | yes | yes | Q | yes | yes | I | yes | yes | K | yes | S | T | Y | yes | 2 | 13 & 2 | | | | | | | |
| <i>Acidiphilium</i> | COX3 | YP_001235282 | 283 | yes | yes | yes | yes | yes | yes | R | yes | yes | yes | yes | yes | R | yes | T | P | yes | yes | 2 | 15 & 1 | | | | | | | |
| sister group2: mitochondria | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | |
| <i>Dictyostelium</i> | COX3 | NP_050071 | 435 | yes | yes | yes | D | yes | yes | L | T | V | Q | I | yes | N | yes | yes | gap | yes | yes | 2 | 11 & 1 | | | | | | | |
| <i>Acanthamoeba</i> | COX3 | NP_042532 | 329 | yes | yes | yes | yes | yes | yes | yes | yes | yes | Q | yes(F) | yes | yes | yes | yes | G | gap | H | L | 1 | 13 & 1 | | | | | | |
| <i>Andulicia</i> | COX3 | YP_007890482 | 267 | yes | yes | yes | yes | yes | yes | yes | yes | yes | yes | yes | yes | yes | yes | yes | G | gap | H | yes | 1 | 15 & 1 | | | | | | |
| <i>Bos_taurus</i> (beef) | COX3 | AAZ16929 | 260 | yes | yes | yes | yes | yes | yes | yes | yes | yes | yes | yes | yes | yes | yes | yes | yes | yes | yes | yes | 2 | 19 | | | | | | |
| Organism and protein | protein operon | aa | E90 | W57 | W58 | E64 | F86 | H71 | T66 | F214 | R221 | F/V233 | G234 | Y241 | H231 | Y181 | Y182 | I185 | F198 | F203 | G82 & M83 | total* | | | | | | | | |