

Supplemental Materials and Methods

Proteomic analysis

Sperm from the caudae epididymides of C57BL/6 mice were collected in PBS containing complete protease inhibitor cocktail (Roche Applied Sciences; Indianapolis, IN), and washed two times at 500 g for 5 min at 4°C. Sperm pellets were collected by centrifugation at 13,700 g for 15 min at 4°C, lysed in 1% Triton X-100 buffer (TX) (50 mM Tris-HCl, (pH 9.0), 1 mM EDTA, 1 mM EGTA, 2 mM phenylmethylsulfonyl fluoride (PMSF), 2 mM DTT, one tablet of EDTA-free complete proteinase inhibitor cocktail (Roche Applied Sciences)] or 80 mM n-Octyl-β-D-glucopyranoside buffer (NOG) (10 mM Tris-HCl (pH 7.2), one tablet of complete protease inhibitor cocktail (Roche Applied Sciences)], and incubated for 2 h at 4°C. The sperm solutions were centrifuged at 13,700 g for 15 min at 4°C, and the supernatants were used as samples.

Sperm lysate samples (100 µg) from *Akap4*^{-/-} (Miki et al., 2004) and WT mice were added to 200 µl of DeStreak rehydration solution and loaded on 11-cm Immobiline DryStrip (GE Healthcare Life Sciences, Piscataway NJ), pH3-11NL with IPG buffer pH3-11NL. The first dimension-isoelectric focusing (IEF) was run by IPGphor Isoelectric Focusing System (GE Healthcare Life Sciences) overnight, and the samples were resolved by rehydration for 12 h, at 500 volts/h for 1 h, at 1000 volts/h for 1 h, and at 16,000 volts/h for 2 h. The IPG sample strips were equilibrated twice for 15 min at room temperature (RT) in SDS equilibration buffer (50 mM Tris-Cl, (pH 8.8), 6 M urea, 30 % glycerol (v/v), 2 % SDS, bromophenol blue) containing freshly added 1 % DTT, followed by equilibration in the buffer containing 2.5 % iodoacetamide for 15 min at RT. The equilibrated IPG strips were placed on 8-16 % Criterion Tris-HCl precast gels (Bio-Rad, Hercules, CA), and subjected to second dimensional electrophoresis. After running, gels were fixed and stained overnight using SYPRO Ruby Protein Stains (Life Technologies

Corp., Carlsbad, CA) according to the supplier's directions. The experiments were performed in triplicate.

Gel images were captured (Fluor-S Multimager, Bio-Rad) for analysis and comparison using Image J and PDQuest software. Differentially expressed proteins with 1.5 fold changes in the density level of the spot (Supplemental Fig. S1) were excised manually from gels, digested with trypsin (Promega, Madison, WI) for 8 hour in a Progest In-gel Digester (Genomics Solutions, Bath, UK), and analyzed by MALDI mass spectrometry (MALDI/MS). Samples were lyophilized to dryness and resuspended in 50:50 (v/v) 0.2 % formic acid:acetonitrile, and then 0.3 μ l samples were spotted onto a 192-sample stainless steel MALDI plate and mixed with 0.3 μ l of 33 % saturated α -cyano-hydroxycinnamic acid. Mass spectrometric analyses were performed with an AB 4700 Proteomics Analyzer (Life Technologies) in the positive ion and reflector modes. The MS was internally calibrated using autolytic tryptic peptides and the MS/MS calibrated externally using the fragment ions of the angiotensin I M+H ion (m/z 1296.68). A focus mass of m/z 2000 was used for the MS acquisition. For the MS/MS, 1000 volts were used for the collision energy and argon used as the collision gas with a recharge threshold set at 1.0×10^{-7} torr. Protein identification was performed by interrogating both the MS and the MS/MS using MASCOT and the entire NCBI non-redundant database. Search parameters included an allowance of two missed tryptic cleavages, a 0.06 Da mass tolerance for the MS data, a 0.1 Da mass tolerance for the MS/MS data, and an allowance for variable oxidation of methionine residues.

Supplemental Table S1: PCR Primer Sequences

MGI Gene name	Chromosome (strand; location)	Primer Name (Amplified Region)	Primer (upper: forward; lower: reverse)	Amplified size (bp)	Primer position (upper: forward; lower: reverse)
<i>Eno1</i> (Enolase 1, alpha non-neuron)	4 (+; 149610830-149622988)	<i>Eno1</i> (5'UTR+ORF)	5'- GTG GGA GGC GCT TAG TGC TGC -3' 5'- GAG GCA GCC ACA TCC ATG CCA A -3'	869	31-51 898-878
<i>Gm5506</i> (Predicted gene 5506)	18 (+; 48204989-48208032)	<i>Gm5506_v1</i> (5'UTR+ORF)	5'- GAA GGT GTG GCT GTT TAT TTT AC -3' 5'- CAC CGC AGC TCG GAA GAG ACC -3'	264	1-23 261-241
<i>Eno4</i> (Enolase 4)	19 (+; 59017915-59045909)	<i>Eno4-5'ORF</i> (ORF)	5'- GAG AAC GAC GTC CCG CGC AA -3' 5'- TGG TGG AGG TGC GGG AAC CA -3'	489	117-136 605-586
		<i>Eno4-3'ORF</i> (ORF)	5'- AGC ATG GCC ATA GGC GCT GT -3' 5'- AGC TGC GCT TTT GTG GGT GC -3'	561	741-760 1301-1282
		<i>Eno4-3'UTR</i> (3'UTR)	5'- AGG GCA CAG CCA CTC CAC CA -3' 5'- GGT GCG TGA CTG GGC GTG AA -3'	297	1915-1934 2211-2192
		<i>Eno4-full length</i> (ORF+3'UTR)	5'- GTC CCG CGC AAG CTG GAA GAT -3' 5'- TCC AGA ACC GGG AGG GAG GC -3'	1784	126-146 1909-1890
		<i>Eno4-3'ORF-ex13</i> (ORF including exon 13 nucleotides)	5'- CCG CTT CCG GGA GTG TCT CT -3' 5'- GCC CTG GGA GTC CAG AAC CG -3'	476	1444-1463 1919-1900
<i>Rn18S</i>		<i>Rn18S</i>	5'- GAC CCG GGG AGG TAG TGA CGA-3' 5'- GGA GCT GGA ATT ACC GCG GCT-3'	141	

NCBI Reference sequences: Eno4: NM_178689.4; Eno1:NM_023119.2; Gm5506: see Supplemental Fig. S4
 ORF positions of each gene: Eno4: 33-1889; Eno1:154-1458

Supplemental Table S2. qPCR Primer Sequences

Primer Name (Region)	Primer (upper: forward; lower: reverse)	Amplified size (bp)	Primer Position*
<i>Eno4-5'</i> (<i>Eno4</i> -5'ORF region)	5'- ATG GGA GAT GAA GAC GGC GG-3' 5'- GCA GGC TGG AGG TAG AAG GT-3'	146	33-52 178-159
<i>Eno4-3'ORF</i> (exon13) (<i>Eno4</i> -exon13 region)	5'- GGT GCC CGG TTT ATC AAG TT -3' 5'- CCA TAC TCC CCT CTG GAT GA -3'	105	1656-1675 1760-1741
<i>Gapdh</i>	5'- TGC ACC ACC AAC TGC TTA G-3' 5'- GAT GCA GGG ATG ATG TTC-3'	176	

*NCBI Reference sequences: *Eno4*: NM_178689.4

Supplemental Table S3. Sperm Motility

	% motility			% progress motility		
	5 min	60 min	120 min	5 min	60 min	120 min
<i>Eno4</i> ^{+/+} (n=2)	45.6 ± 8.2	48.2 ± 10.1	37.4 ± 16.9	26.8 ± 3.0	24.4 ± 4.4	17.3 ± 8.0
<i>Eno4</i> ^{Gt/Gt} (n=2)	15.2 ± 2.9	8.0 ± 2.5	1.9 ± 0.2	5.7 ± 1.8 ¹	2.5 ± 1.6	1.1 ± 0.3

¹ p < 0.05 compared to *Eno4*^{+/+}

SUPPLEMENTAL FIGURE LEGENDS

Supplemental Fig. S1. Two-dimensional gel analysis of NOG extract of WT C57BL/6 sperm. The samples were prepared and analyzed as described in Supplemental Methods and Materials. The gel was silver stained and the proteins in the white box were determined by MS/MS analysis to be enolase 4 (ENO4).

Supplemental Fig. S2. Western blotting of kidney, brain, skeletal muscle, testis and sperm, using pan-enolase antibody (ENO) and antibodies to ENO2 and ENO3. Pan-enolase antibody detected a band in all tissues, while antibodies to ENO2 and ENO3 detected a band in brain and skeletal muscle, respectively, but not in sperm. The CBB stained gel for the ENO western blot is shown as a loading control.

Supplemental Fig. S3. Alignment of mouse ENO1, ENO2, ENO3 and ENO4 amino acid sequences. Gaps are show as (-), identities are show as (*), similarities are shown as (:).

Supplemental Fig. S4. Alignment of *Eno4*, *Eno4_v1* and *Eno4_v2* nucleotides and their deduced amino acid sequences. **A)** The underlined ATG in bold in the nucleotide sequences identifies the predicted initiation codons. Gaps are show as (-). **B)** The initiation methionine (**M**) of ENO4 and ENO4_V1 protein sequences is underlined. The predicted initiation methionine (**M**) of ENO4_V2 is shown in bold.

Supplemental Fig. S5. Alignment of *Gm5506* and *Gm5506_v1* nucleotides. Nucleotide sequences in 5' untranslated region of *Gm5506_v1* aligned with *Gm5506*. The underlined ATG identifies the predicted initiation codon. Gaps are show as (-).

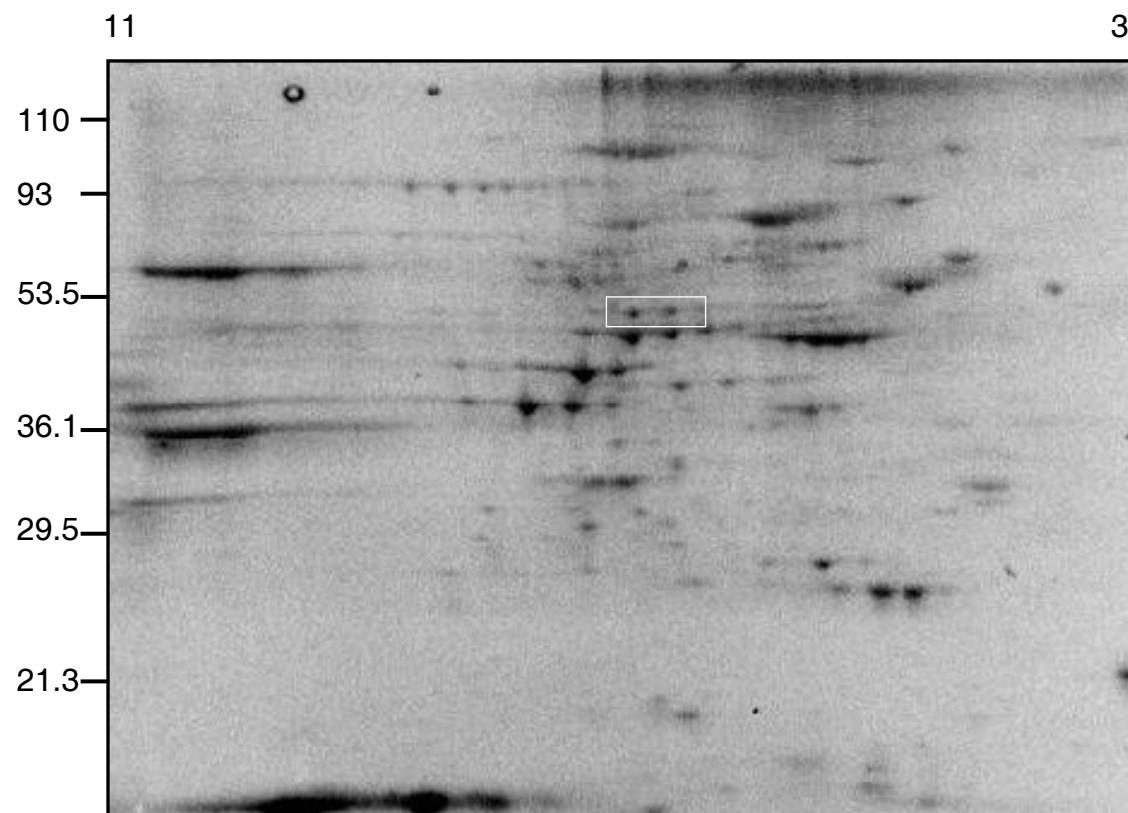
Supplemental Fig. S6. Characterization of *Eno4* gene and of *Eno4* gene trap. **A)** Structure of WT *Eno4* and *Eno4* gene trap alleles. The gene trap vector (including a splice acceptor site (diagonal line box) and β-geo coding region integrated 300 base pair downstream of exon 1 of the *Eno4* gene. The relevant Hind 3 sites and probe positions used for Southern and Northern analysis are indicated. Probe 1 was located in the intron region downstream of the gene trap sequence; Probe 2 was located within the β-geo coding region. **B)** Southern analysis. DNA was isolated from testes of WT mice and of mice heterozygous (*Eno4*^{+/Gt}) or homozygous (*Eno4*^{Gt/Gt}) for the gene trap, digested with Hind 3 and hybridized with probe 1 (left panel) and probe 2 (right panel). The 1.6 and 9.0 Kb bands correspond to *Eno4* wild type and *Eno4* gene trap alleles, respectively. **C)** Northern analysis of mRNA isolated from testis of 3-month-old of WT and *Eno4*^{Gt/Gt} mice using a [³²P]-labeled mouse *Eno4* cDNA probe located in the 3' portion of the ORF. **D)** Western blotting analysis of extracts of testis from 3-month-old WT and *Eno4*^{Gt/Gt} mice with antibodies of ENO4 (left panels) and β-galactosidase (right panels). The antibody to ENO4 detected a band in WT testis, but not in *Eno4*^{Gt/Gt} testis. The antibody to β-galactosidase did not detect a band in WT testis, but detected a band in *Eno4*^{Gt/Gt} testis.

Supplemental Fig. S7. TUNEL assay for apoptosis in testis of WT and *Eno4*^{Gt/Gt} mice. **(A)** TUNEL staining on sections of testis from WT (**A**) and *Eno4*^{Gt/Gt} mice (**B-D**). TUNEL positive cells are shown as a fluorescent signal in the seminiferous tubules. Bars = 25 μm.

Supplemental Fig. S8. Western blotting with sperm from WT and *Eno4*^{Gt/Gt} mice using ENO4, pan-enolase (ENO) and AKAP4 antisera. ENO4 was detected in sperm from WT mice, but not in *Eno4*^{Gt/Gt} mice (left upper panel). ENO was detected with antibody to pan-enolase in WT and *Eno4*^{Gt/Gt} mice; the amount of ENO4 protein detected in *Eno4*^{Gt/Gt} sperm extracts was appreciably lower than in sperm WT mice (left middle panel). AKAP4 was detected in WT and *Eno4*^{Gt/Gt} mice; the amount of band detected with antibody to AKAP4 decreased in *Eno4*^{Gt/Gt} mice. Antibody to α -tubulin was used as a loading control.

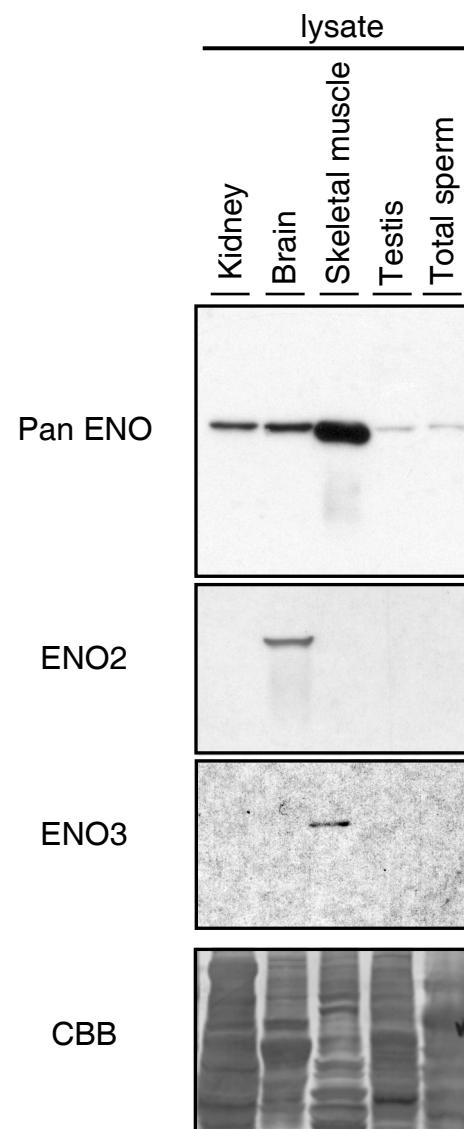
Supplemental Fig. S1

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Supplemental Fig. S2

BOR-Papers in Press. Published as DOI: 10.1095/biolreprod.112.107128



Supplemental Fig. S3

EN01 -----PLAK
EN02 -----PSVL
EN03 -----PKAK
EN04 DATATMAEETLGLLDSIFPTEVIEESAKT

Supplemental Fig. S4

A

Eno4	ACC <u>ATG</u> GGAGATGAAGACGGCGGCCGAGAGGCGGGATCACTAGGGACCTGCAGAAGCTG	60
Eno4_v1	ACC <u>ATG</u> GGAGATGAAGACGGCGGCCGAGAGGCGGGATCACTAGGGACCTGCAGAAGCTG	60
Eno4_v2	-----	
Eno4	AAGCAGCAGGCATGGCGTACTACCAGGAGAACGACGTCCCAGCAAGCTGAAAGATCTG	120
Eno4_v1	AAGCAGCAGGCATGGCGTACTACCAGGAGAACGACGTCCCAGCAAGCTGAAAGATCTG	120
Eno4_v2	-----	
Eno4	CTCAACTCCACCTTCTACCTCAGCCTGCGGATGTCTACGGGACCTGAAGGCAAATAC	180
Eno4_v1	CTCAACTCCACCTTCTACCTCAGCCTGCGGATGTCTACGGGACCTGAAGGCAAATAC	180
Eno4_v2	-----	
Eno4	TTTTCTAAACTGCGAAGCCTCCCTCCATATGCAAAATAGTAGGGAAAACCATACTGGAT	240
Eno4_v1	TTTTCTAAACTGCGAAGCCTCCCTCCATATGCAAAATAGTAGGGAAAACCATACTGGAT	240
Eno4_v2	-----	
Eno4	GGCCTGGGACTTCCTACTCTACAGGTGAAATCTCCTGCACCATTCAAATTTCCCCAAG	300
Eno4_v1	GGCCTGGGACTTCCTACTCTACAGGTGAAATCTCCTGCACCATTCAAATTTCCCCAAG	300
Eno4_v2	-----	
Eno4	TACATCTGTGCCGTGGCGATCCCTACGCACTTTGAAGTCGTGGAGAACGCTTGCCAGAG	360
Eno4_v1	TACATCTGTGCCGTGGCGATCCCTACGCACTTTGAAGTCGTGGAGAACGCTTGCCAGAG	360
Eno4_v2	-----	
Eno4	GCGTTGGATGCAGAGGACTCAGAAAGGGCCAGGCCGTGAACACAGCGGTGCAGTGGATC	420
Eno4_v1	GCGTTGGATGCAGAGGACTCAGAAAGGGCCAGGCCGTGAACACAGCGGTGCAGTGGATC	420
Eno4_v2	-----	
Eno4	AATCAGTCCATCACAGAGGAGCTGTGGGTCTGGTCCCTCCAACCAGGCAGAGGTGGAC	480
Eno4_v1	AATCAGTCCATCACAGAGGAGCTGTGGGTCTGGTCCCTCCAACCAGGCAGAGGTGGAC	480
Eno4_v2	-----	
Eno4	CACTGGCTCAGGACCTTCTTGAACATAAGTACAAGAAGATAAAGAGAGAAAAGAATTG	540
Eno4_v1	CACTGGCTCAGGACCTTCTTGAACATAAGTACAAGAAGATAAAGAGAGAAAAGAATTG	540
Eno4_v2	-----	
Eno4	GAAAAGAGCCAGGAGGAGCTGGTCCGCACCTCACCAGTAACGCTGCCGCCGCC	600
Eno4_v1	GAAAAGAGCCAGGAGGAGCTGGTCCGCACCTCACCAGTAACGCTGCCGCCGCC	600
Eno4_v2	-----	
Eno4	CCGCCGCCTCCCCACCTCCAAAAAGAAAGGCAGAAGCAGGAGGGATACG	660
Eno4_v1	CCGCCGCCTCCCCACCTCCAAAAAGAAAGGCAGAAGCAGGAGGGATACG	660
Eno4_v2	-----GGACGGAGGGATACG	15
Eno4	CTTTGGAGAACCTGTTCACCCCCGGAGCCCCCTGAACCAGTCCTCATGGTAGCATG	720
Eno4_v1	CTTTGGAGAACCTGTTCACCCCCGGAGCCCCCTGAACCAGTCCTCATGGTAGCATG	720
Eno4_v2	CTTTGGAGAACCTGTTCACCCCCGGAGCCCCCTGAACCAGTCCTCATGGTAG ATG	75
Eno4	GCCATAGGCCTGTGTCCTGGCAGTGGCAAAGCCAGCGAACCTGGCAGCGATCCT	780
Eno4_v1	GCCATAGGCCTGTGTCCTGGCAGTGGCAAAGCCAGCGAACCTGGCAGCGATCCT	780
Eno4_v2	GCCATAGGCCTGTGTCCTGGCAGTGGCAAAGCCAGCGAACCTGGCAGCGATCCT	135

Eno4	CTCTACTTAACCTGGCGTCACTGAAGCATGATCAGGAGCAGCGTCAACATTCTATG	840
Eno4_v1	CTCTACTTAACCTGGCGTCACTGAAGCATGATCAGGAGCAGCGTCAACATTCTATG	840
Eno4_v2	CTCTACTTAACCTGGCGTCACTGAAGCATGATCAGGAGCAGCGTCAACATTCTATG	195
Eno4	CCTTTGCTGATGGGATCTGTACTGAGCTGTGGAAAGTCGTACCTGGGAAGCTACATT	900
Eno4_v1	CCTTTGCTGATGGGATCTGTACTGAGCTGTGGAAAGTCGTACCTGGGAAGCTACATT	900
Eno4_v2	CCTTTGCTGATGGGATCTGTACTGAGCTGTGGAAAGTCGTACCTGGGAAGCTACATT	255
Eno4	ATGAAAGAAGTGATTGTATTCCAGTCAGGATTAACAGCAAACAAAGTGTGAGTTG	960
Eno4_v1	ATGAAAGAAGTGATTGTATTCCAGTCAGGATTAACAGCAAACAAAGTGTGAGTTG	960
Eno4_v2	ATGAAAGAAGTGATTGTATTCCAGTCAGGATTAACAGCAAACAAAGTGTGAGTTG	315
Eno4	CTTCTGAAATTCAAGAACAGTTAACAGAGCGATGGAGACGCTTCCACCTCCAAAACAA	1020
Eno4_v1	CTTCTGAAATTCAAGAACAGTTAACAGAGCGATGGAGACGCTTCCACCTCCAAAACAA	1020
Eno4_v2	CTTCTGAAATTCAAGAACAGTTAACAGAGCGATGGAGACGCTTCCACCTCCAAAACAA	375
Eno4	GAGACAAAAAGGGCATAATGGGAGCAAAAGAGCTCAGGCCACCGATCACTGGCAAGGTA	1080
Eno4_v1	GAGACAAAAAGGGCATAATGGGAGCAAAAGAGCTCAGGCCACCGATCACTGGCAAGGTA	1080
Eno4_v2	GAGACAAAAAGGGCATAATGGGAGCAAAAGAGCTCAGGCCACCGATCACTGGCAAGGTA	435
Eno4	TCTCACCTTGGCTTTAACCATCAACTACGACGCCATAGAGCAGCCACTGCTGCTGCTT	1140
Eno4_v1	TCTCACCTTGGCTTTAACCATCAACTACGACGCCATAGAGCAGCCACTGCTGCTGCTT	1140
Eno4_v2	TCTCACCTTGGCTTTAACCATCAACTACGACGCCATAGAGCAGCCACTGCTGCTGCTT	495
Eno4	CAGGGGATCTGCTAAACCTGGGTTGAACTGGGAGTGAACCTCCATCTAGCCATCAAC	1200
Eno4_v1	CAGGGGATCTGCTAAACCTGGGTTGAACTGGGAGTGAACCTCCATCTAGCCATCAAC	1200
Eno4_v2	CAGGGGATCTGCTAAACCTGGGTTGAACTGGGAGTGAACCTCCATCTAGCCATCAAC	555
Eno4	TGTGCTGGCATGAGCTGATGGACTACAGTAAAGGAAGTACGAAGTGATGGTGGCACC	1260
Eno4_v1	TGTGCTGGCATGAGCTGATGGACTACAGTAAAGGAAGTACGAAGTGATGGTGGCACC	1260
Eno4_v2	TGTGCTGGCATGAGCTGATGGACTACAGTAAAGGAAGTACGAAGTGATGGTGGCACC	615
Eno4	CACAAAAGCGCAGCTGAGATGGTAGGAGCTCTACGTGGACCTGATCAACAAGTATCCTCC	1320
Eno4_v1	CACAAAAGCGCAGCTGAGATGGTAGGAGCTCTACGTGGACCTGATCAACAAGTATCCTCC	1320
Eno4_v2	CACAAAAGCGCAGCTGAGATGGTAGGAGCTCTACGTGGACCTGATCAACAAGTATCCTCC	675
Eno4	ATAATCGCTTAATTGATCCTTCAGGAAGGAGGACGCCGAGCAGTGGACAGCCTCTAT	1380
Eno4_v1	ATAATCGCTTAATTGATCCTTCAGGAAGGAGGACGCCGAGCAGTGGACAGCCTCTAT	1380
Eno4_v2	ATAATCGCTTAATTGATCCTTCAGGAAGGAGGACGCCGAGCAGTGGACAGCCTCTAT	735
Eno4	GCTGCTCTGGCTTCCCGGTGCTACCTAATTGCAGGCGCCGCTCCGGAGTGTCTCTAAA	1440
Eno4_v1	GCTGCTCTGGCTTCCCGGTGCTACCTAATTGCAGGCGCCGCTCCGGAGTGTCTCTAAA	1440
Eno4_v2	GCTGCTCTGGCTTCCCGGTGCTACCTAATTGCAGGCGCCGCTCCGGAGTGTCTCTAAA	795
Eno4	CTCCTAGAGTGCAGAAATATAAGCACCCCTGAAATCCCACGGACTGATCATAAAGCACACA	1500
Eno4_v1	CTCCTAGAGTGCAGAAATATAAGCACCCCTGAAATCCCACGGACTGATCATAAAGCACACA	1500
Eno4_v2	CTCCTAGAGTGCAGAAATATAAGCACCCCTGAAATCCCACGGACTGATCATAAAGCACACA	855
Eno4	AACCAAACCACAATGTCTGACTTGGTGGAAATAACCCATCTTATCAACGTAAGAAGCTC	1560
Eno4_v1	AACCAAACCACAATGTCTGACTTGGTGGAAATAACCCATCTTATCAACGTAAGAAGCTC	1560
Eno4_v2	AACCAAACCACAATGTCTGACTTGGTGGAAATAACCCATCTTATCAACGTAAGAAGCTC	915
Eno4	CTGGCCGTCTTGGAAAGCACAGACTCGGAGTCCTCTGATGACAGCCTTGTGATTTGGCT	1620
Eno4_v1	CTGGCCGTCTTGGAAAGCACAGACTCGGAGTCCTCTGATGACAGCCTTGTGATTT----	1616
Eno4_v2	CTGGCCGTCTTGGAAAGCACAGACTCGGAGTCCTCTGATGACAGCCTTGTGATTTGGCT	975
Eno4	GTTGGATTGGTGCCTGGTTATCAAGTGGGGGTCTTCTGGGGTGAACGGATGACC	1680

Eno4_v1	-----	
Eno4_v2	GTTGGATTCGGTGCCCGGTTATCAAGTTGGGGGTCTTCGGGGTGAACGGATGACC	1035
Eno4	AAATACAACCGCCTCTTGCTATAGAGGAAGAACCATCCAGAGGGAGTATGGGTTTC	1740
Eno4_v1	-----	GGTTTC
Eno4_v2	AAATACAACCGCCTCTTGCTATAGAGGAAGAACCATCCAGAGGGAGTATGGGTTTC	1095
Eno4	AGTGAAGAACACAATTTCTTCAAGAGGGATGCTACTGCCACAATGGCTGAGGAA	1800
Eno4_v1	AGTGAAGAACACAATTTCTTCAAGAGGGATGCTACTGCCACAATGGCTGAGGAA	1682
Eno4_v2	AGTGAAGAACACAATTTCTTCAAGAGGGATGCTACTGCCACAATGGCTGAGGAA	1155
Eno4	ACTCTGGGCTCCTGGACTCCATCTCCCCACAGAGGTGATAAGAGGAATCGGCTAAAACA	1880
Eno4_v1	ACTCTGGGCTCCTGGACTCCATCTCCCCACAGAGGTGATAAGAGGAATCGGCTAAAACA	1742
Eno4_v2	ACTCTGGGCTCCTGGACTCCATCTCCCCACAGAGGTGATAAGAGGAATCGGCTAAAACA	1215
Eno4	TGA	1883
Eno4_v1	TGA	1745
Eno4_v2	TGA	1218

B

ENO4	M GDEDGGRRGGITRDLQKLKQQAMAYYQENDVPRKLEDLLNSTFYLQPADVYGHANYFS	60
ENO4_v1	M GDEDGGRRGGITRDLQKLKQQAMAYYQENDVPRKLEDLLNSTFYLQPADVYGHANYFS	60
ENO4_v2	-----	
EN04	KLAKPPSICKIVGKTILDGLGLPTLQVEISCTIQNFPKYICAVAIPTHFEVVENALPEAL	120
EN04_v1	KLAKPPSICKIVGKTILDGLGLPTLQVEISCTIQNFPKYICAVAIPTHFEVVENALPEAL	120
EN04_v2	-----	
EN04	DAEDSERAQAVNTAVQWINQSITEELWGLVPSNQAEDHRLRTFFEHKVQEDKERKELEK	180
EN04_v1	DAEDSERAQAVNTAVQWINQSITEELWGLVPSNQAEDHRLRTFFEHKVQEDKERKELEK	180
EN04_v2	-----	
EN04	SQEELVPAPPPVTLPPPPPPPPSKKKQKAGRDTLLEKPVSPPPEPVLHGSMAI	240
EN04_v1	SQEELVPAPPPVTLPPPPPPPPSKKKQKAGRDTLLEKPVSPPPEPVLHGSMAI	240
EN04_v2	-----	5
EN04	GAVSLAVAKASATLASDPLYLTLASLKHDQEQPSTFSMPLLMGSVLSCGSSPGKLHLMK	300
EN04_v1	GAVSLAVAKASATLASDPLYLTLASLKHDQEQPSTFSMPLLMGSVLSCGSSPGKLHLMK	300
EN04_v2	GAVSLAVAKASATLASDPLYLTLASLKHDQEQPSTFSMPLLMGSVLSCGSSPGKLHLMK	65
EN04	EVICIPSPGLTAKQSVELLIEIQKQVNRAMETLPPPQETKKGHNGSKRAQPPITGVSH	360
EN04_v1	EVICIPSPGLTAKQSVELLIEIQKQVNRAMETLPPPQETKKGHNGSKRAQPPITGVSH	360
EN04_v2	EVICIPSPGLTAKQSVELLIEIQKQVNRAMETLPPPQETKKGHNGSKRAQPPITGVSH	125
EN04	LGCLTINYDAIEQPLLLLQGICSNLGLELGVNFHLAINCAGHELMDSKGKYEVVGTHK	420
EN04_v1	LGCLTINYDAIEQPLLLLQGICSNLGLELGVNFHLAINCAGHELMDSKGKYEVVGTHK	420
EN04_v2	LGCLTINYDAIEQPLLLLQGICSNLGLELGVNFHLAINCAGHELMDSKGKYEVVGTHK	185
EN04	SAAEMVELYVDLINKYPSIIALIDPFRKEDAEQWDSLTYAALASRCYLIAGAASGSVSKLL	480
EN04_v1	SAAEMVELYVDLINKYPSIIALIDPFRKEDAEQWDSLTYAALASRCYLIAGAASGSVSKLL	480
EN04_v2	SAAEMVELYVDLINKYPSIIALIDPFRKEDAEQWDSLTYAALASRCYLIAGAASGSVSKLL	245

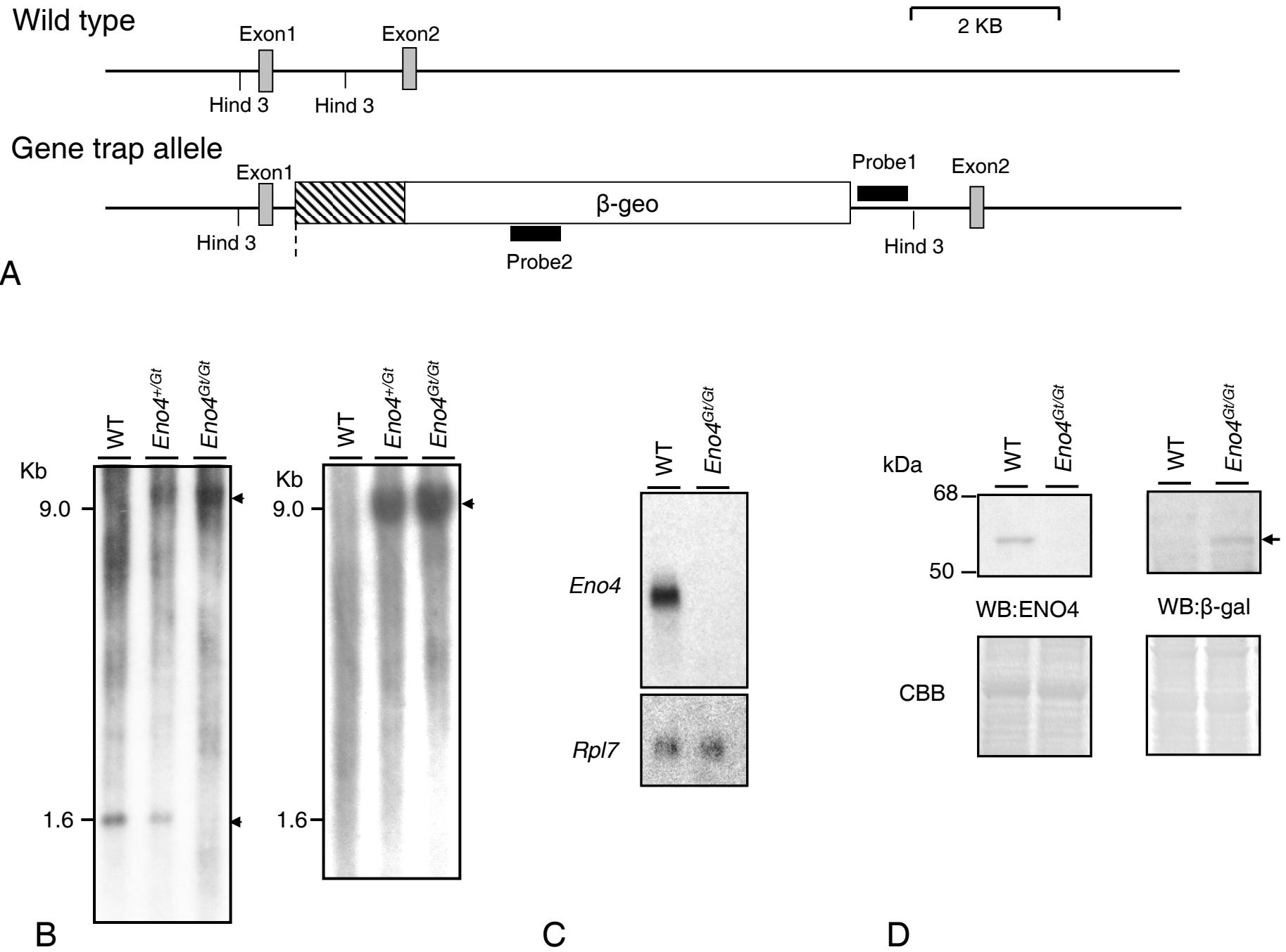
EN04	ECRNISTLKGHLIKHTNQTTMSDLVEITHLINGKKLLAVFGSTDSESSDDSLVDLAVG	540
EN04_v1	ECRNISTLKGHLIKHTNQTTMSDLVEITHLINGKKLLAVFGSTDSESSDDSLVDLVSV	540
EN04_v2	ECRNISTLKGHLIKHTNQTTMSDLVEITHLINGKKLLAVFGSTDSESSDDSLVDLAVG	305
EN04	FGARFIKLGGLSRGERMTKYNRLLAIEEELIQRGVWGFSEEHNFSSFFQEDATATMAEETL	600
EN04_v1	KNTIFLSFKRMLLPQWLRK1LGSWTPSSPQR-----	572
EN04_v2	FGARFIKLGGLSRGERMTKYNRLLAIEEELIQRGVWGFSEEHNFSSFFQEDATATMAEETL	365
EN04	GLLDSIFPTEVIEESAKT	618
EN04_v1	-----	572
EN04_v2	GLLDSIFPTEVIEESAKT	388

Supplemental Fig. S5

Gm5506	-----GAGGTCTGATACCAACACAGACACTGCTAGAACTAT	38
Gm5506_v1	GAAGGTGTGGCTGTTATTTACAGTCTGTTAGAAAAAAACAGAGAGTGCTGCCGGT	60
Gm5506	TGTGGTTGGTCTAATGCTGAATTATGTTAACGGGTCCTCAAATTGCATGAGAATCT	98
Gm5506_v1	ACAGGGTCGCCTCGCTCTGCTTAAGGCTCCCTCGGTGTCACGGCACCCCTTTCT	120
Gm5506	GCATGTAAGAACGATCCTACTGCCAGAAATTGCC <u>ATG</u> TCTATTCTCAGGATCCACGCC	158
Gm5506_v1	CTTGCTTGAGCGATCCTACTGCCAGAAATTGCC <u>ATG</u> TCTATTCTCAGGATCCACGCC	180
Gm5506	AGAGAGATCTTGACTCCGTGGAAATCCCCTGTTGAGGTCGATCTGTACACCGCAAAA	218
Gm5506_v1	AGAGAGATCTTGACTCCGTGGAAATCCCCTGTTGAGGTCGATCTGTACACCGCAAAA	240
Gm5506	GGTCTCTTCCGAGCTGCCAGCGGTGCGTCCACTGGCATCTACGAGGCCCTAGAA	278
Gm5506_v1	GGTCTCTTCCGAGCTGCCAGCGGTGCGTCCACTGGCATCTACGAGGCCCTAGAA	300
Gm5506	CTCCGAGACAATGATAAGACCCGTTCATGGGAAGGGTGTCTCACAGGCTGTTGAGCAC	338
Gm5506_v1	CTCCGAGACAATGATAAGACCCGTTCATGGGAAGGGTGTCTCACAGGCTGTTGAGCAC	360
Gm5506	ATCAATAAAACTATTGCGCCTGCTCTGGTTAGCAAGAAAGTGAATGTTGTGGAGCAAGAG	398
Gm5506_v1	ATCAATAAAACTATTGCGCCTGCTCTGGTTAGCAAGAAAGTGAATGTTGTGGAGCAAGAG	420
Gm5506	AAGATTGACAAGCTGATGAT	418
Gm5506_v1	AAGATTGACAAGCTGATGAT	440

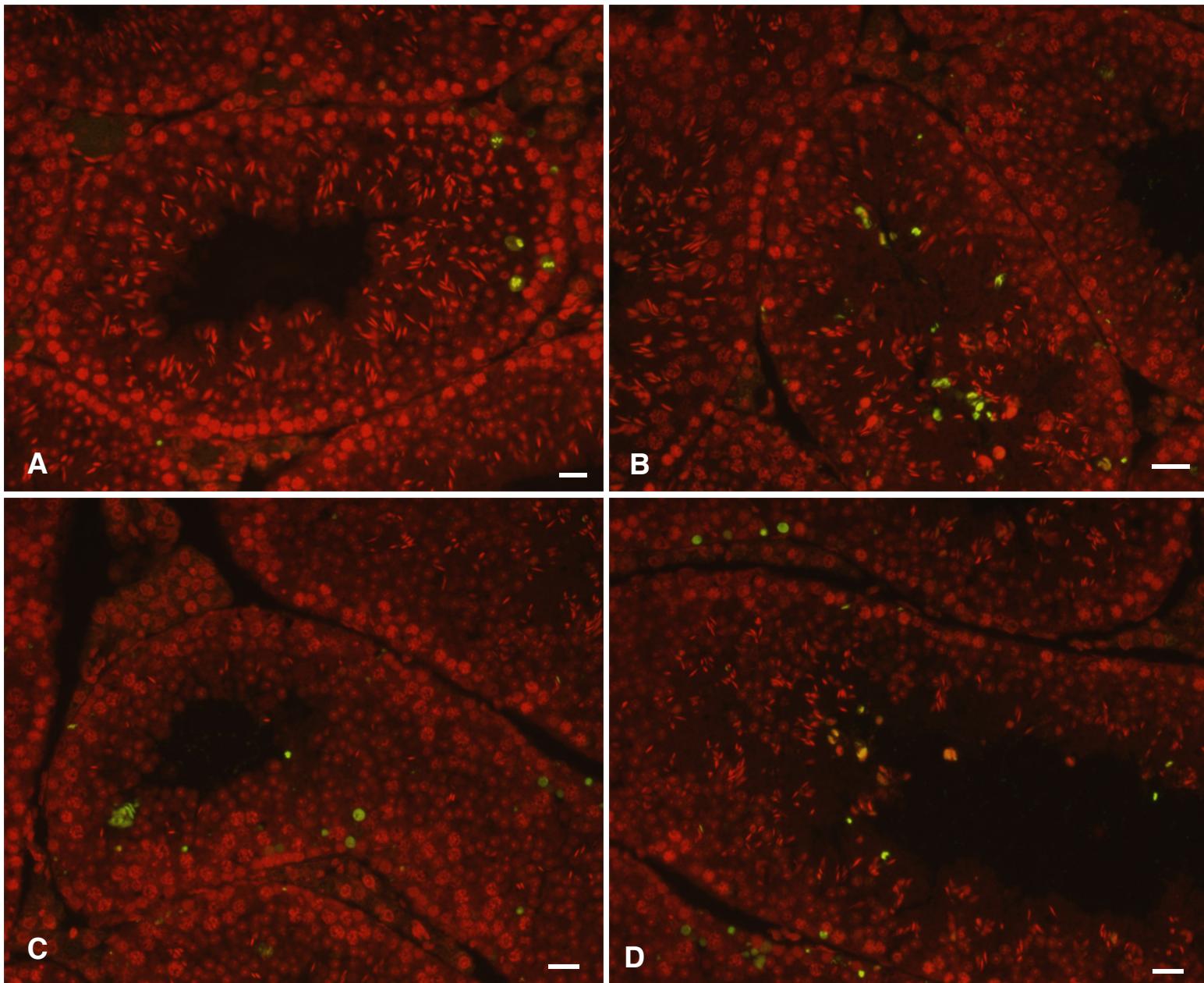
Supplemental Fig. S6

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Supplemental Fig. S7

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Supplemental Fig. S8

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