

Figure S1. *Runx1* is required for the normal development of Flt3⁺ MDPs but dispensable for mature DC development. (A - C) BM cells from *Runx1*-CKO (*Vav1*-iCre⁺ *Runx1*^{F/F}, Cre⁺) and control (*Runx1*^{F/F}, Cre⁻) mice were analyzed for expression of lineage markers (B220, CD11b, MHC-II, CD16/32), c-kit, Sca1, CD150, and CD135 (Flt3). Frequencies of Flt3⁺ Lin⁻ Sca1⁺ c-kit⁺ (LMPP, A), Flt3⁺ Lin⁻ Sca1⁻ c-kit⁺ (MDP/CLP) and Flt3⁺ Lin⁻ Sca1⁻ c-kit^{lo/-} (CDP) populations (B) are shown with rectangle gates. Data from *Vav1*-iCre⁺ *Cbfb*^{F/F} and *Vav1*-iCre⁺ *Cbfb*^{F/F} mice analyzed at the same time are shown for comparison. Statistical analysis from three mice is shown as mean and standard deviation in (C). (D) Splenocytes from *Runx1*-CKO (*Vav1*-iCre *Runx1*^{F/F}) or control (*Runx1*^{F/F}) mice were analyzed for expression of B220, CD11c, MHC-II, CD11b, and CD24. CD11c⁺MHC-II⁺ cDCs in B220⁻ gated splenocytes and CD11b and CD24 expression in cDCs are shown with rectangle gates and percentages. Data represents 6 mice per genotype. (E) Flt3 expression in Runx1-deficient LMPPs. A histogram overlay shows CD135 (Flt3) expression in LMPPs (Lin⁻ Sca1⁺ c-kit⁺ Flt3⁺) from *Vav1*-iCre⁺ *Runx1*^{F/F} (blue histograms) and control Cre (–) Runx1^{F/F} (red histograms) mice. Data shown in Figure 4D of Cai *et al.* (*PLoS ONE*, 2011;6(12):e28430) were reanalyzed.

Figure S2

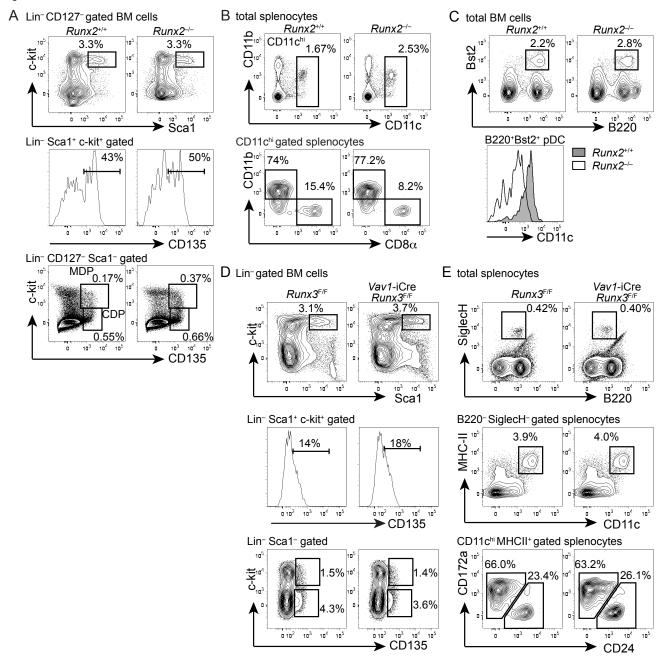


Figure S2. Runx2 is required for CD11c expression in pDCs, but neither Runx2 nor Runx3 are necessary for cDC differentiation. (A) Frequencies of MPP, LMPP, Flt3⁺ MDP, and CDP populations in irradiation chimeric mice reconstituted with $Runx2^{+/+}$ and $Runx2^{-/-}$ fetal liver cells. Data represent analysis of four recipient mice per genotype. (B) Splenocytes from Runx2^{+/+} and Runx2^{-/-} fetal liver chimeras were analyzed for CD11b, CD11c and CD8α expression. CD11chi DCs are gated with rectangles in upper panels with percentages of total splenocytes. Bottom panels show CD11b and CD8α expression in CD11chi cells, and percentages of CD8α-CD11b+DCs and CD8α+CD11b-DCs are shown. (C) BM cells from $Runx2^{+/+}$ and $Runx2^{-/-}$ fetal liver chimeras were analyzed for B220, Bst2 and CD11c expression. Bst2⁺ B220⁺ pDCs are shown with rectangles and their percentages of total BM cells are shown. The histogram depicts CD11c expression in gated pDCs. Data is representative of 4 mice with similar results. (D) Frequencies of MPP, LMPP, Flt3⁺ MDP and CDP populations in Vav1-iCre Runx3^{F/F} and control Cre(-) Runx3^{F/F} mice. (E) Splenocytes from Vav1-iCre Runx3^{F/F} and control Cre(-) Runx3^{F/F} mice were analyzed for expression of B220, SiglecH, MHCII, CD11c, CD24 and CD172a expression. B220^{int} SiglecH⁺ pDCs and B220⁻ MHCII⁺ CD11c^{hi} cDCs are gated with rectangles with percentages in total splenocytes shown. Bottom panels show CD24 and CD172a expression in B220⁻ MHCII⁺ CD11c^{hi} cells, and percentages of CD172a⁺ (CD8α⁻ CD11b⁺) DCs and CD24⁺ (CD8α⁺ CD11b⁻) DCs are shown. Data represent analysis of four mice per genotype with similar results.

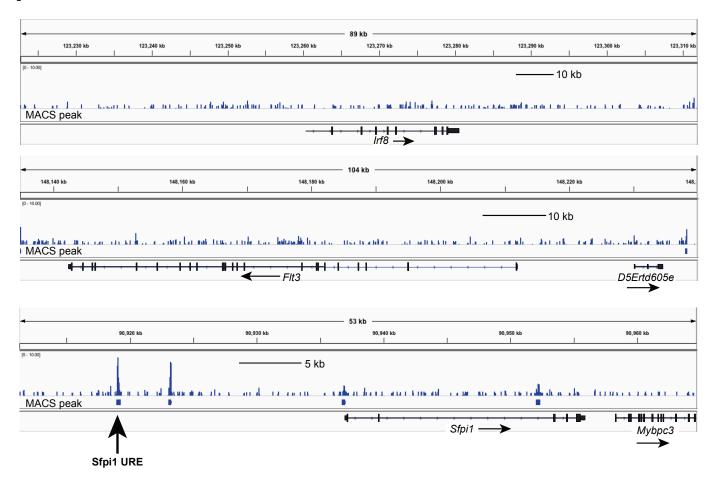


Figure S3. Reanalysis of a published dataset of genome-wide Runx1 binding profiles in a murine hematopoietic progenitor model cell line. Runx1 ChIP sequencing data from GSM552241 were mapped to the mouse genome mm9, and binding peaks were called using bowtie2-2 1.0 with a P-value of 1 x 10⁻⁶. Histograms of Runx1-pulled binding tags encompassing *Irf8*, *Flt3* and *Sfpi1* loci are shown.