

EXPERIMENTAL

The detergent-free recombinant IN protein was produced in *Escherichia coli* and purified as previously described.¹ All oligonucleotides were synthesized by the phosphoramidite method on an automatic ABI 3400 DNA synthesizer (Applied Biosystems, USA) under conditions recommended by manufacturer and purified by electrophoresis in a 20% polyacrylamide/7 M urea gel. Modified oligonucleotides were additionally purified by RP-HPLC with acetonitrile gradient in 0.1 M ammonium acetate (pH 7). HPLC purification was carried out on AKTA Purifier equipped with Jupiter C18 or C5 column (Phenomenex, size 4.6*250 mm, 5 μ m) with UV-Vis detector. Analytical HPLC was carried out on Agilent 1200 equipped with Ultasphere octyl (Beckman Coulter, size 4.6*250 mm, 5 μ m), 45°C 1 ml/min with diode array detector. 5'-FAM, FAM, TET, HEX phosphoramidites were purchased from Metkinen Chemistry Oy; dT, dA(Bz), dG(iBu), dC(Ac), ddR, 1,3-propanediol phosphoramidites and dT p-methyl phosphoramidite - from ChemGenes, JOE phosphoramidite – from Primetech LLC. Oligonucleotides with eosin and oleic acid at 3'-end were synthesized as described previously.² Phosphorothioate oligonucleotides were synthesized accordingly to Krotz et al.³ Oligonucleotides with methylphosphonate linkages were synthesized as described by Arnold and coworkers.⁴ MALDI MS spectra were registered on AutoFlex (Bruker Вфдецтшсы) using 2,4,6-trihydroxyacetophenone/ ammonium citrate⁵ or 3-hydroxypicolinic acid/ ammonium citrate⁶ as a matrix.

3'-Processing activity assay. A DNA duplex consisting of oligonucleotides U5B (5'-GTGTGGAAAATCTCTAGCAGT-3') and U5A (5'-ACTGCTAGAGATTTTCACAC-3'), and mimicking the end of the HIV-1 U5 LTR was used as an IN substrate. The U5B oligonucleotide (10 pmol) was labeled with T4 polynucleotide kinase (Fermentas, Lithuania) and 50 μ Ci [γ -³²P]ATP (3000 Ci/mmol). After 1 h of incubation at 37°C, the T4 polynucleotide kinase was inactivated by the addition of EDTA followed by heating at 65°C for 5 minutes. The U5B oligonucleotide was then annealed with an equimolar amount of complementary oligonucleotide, U5A. The resulting U5B/U5A duplex was then purified from unincorporated [γ -³²P]ATP by centrifugation through MicroSpin G-25 Columns (GE Healthcare, UK). The ³²P-labeled substrate, U5B/U5A (3 nM), was

incubated in 20 µl of 20 mM Hepes, pH 7.2, 7.5 mM MgCl₂, 1 mM DTT with 50 nM IN, in the presence of increasing concentrations of an oligonucleotide inhibitor (0.01-10 µM) at 37°C for 2 h. The reaction was stopped by adding 80 µl of a stop solution (7 mM EDTA, 0.3 M sodium acetate, 10 mM Tris-HCl, pH 8). IN was extracted by phenol/chloroform, and DNA fragments were precipitated with ethanol. The reaction products were suspended in loading dye (80% formamide, 0.05% bromophenol blue, 0.05% xylene cyanol) and separated by electrophoresis in a 20% polyacrylamide/7 M urea gel. Gel images were recorded on a STORM 840TM PhosphorImager (Molecular Dynamics, USA) and quantified with Image QuANTTM 4.1 software (USA).

REFERENCES

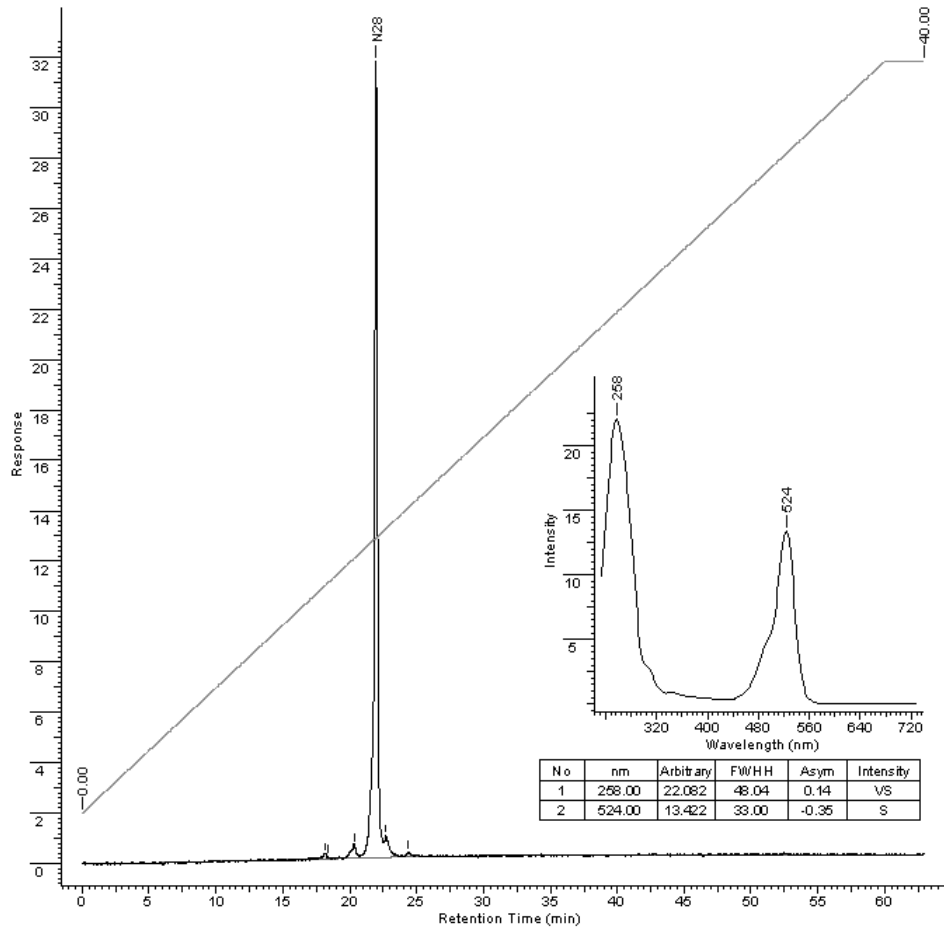
1. Leh, H.; Brodin, P.; Bischerour, J.; Deprez, E.; Tauc, P.; Brochon, J. C.; Le Cam, E.; Coulaud, D.; Auclair, C.; Mouscadet, J. F. Determinants of Mg²⁺-dependent activities of recombinant human immunodeficiency virus type 1 integrase. *Biochemistry* **2000**, *39*, 9285-9294.
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3. Krotz, A. H.; Gorman, D.; Mataruse, P.; Foster, C.; Godbout, J. D.; Coffin, C. C.; Scozzari, A. N. Phosphorothioate Oligonucleotides with Low Phosphate Diester Content: Greater than 99.9% Sulfurization Efficiency with “Aged” Solutions of Phenylacetyl Disulfide (PADS). *Org. Proc. Res. Dev.* **2004**, *8*, 852-858.
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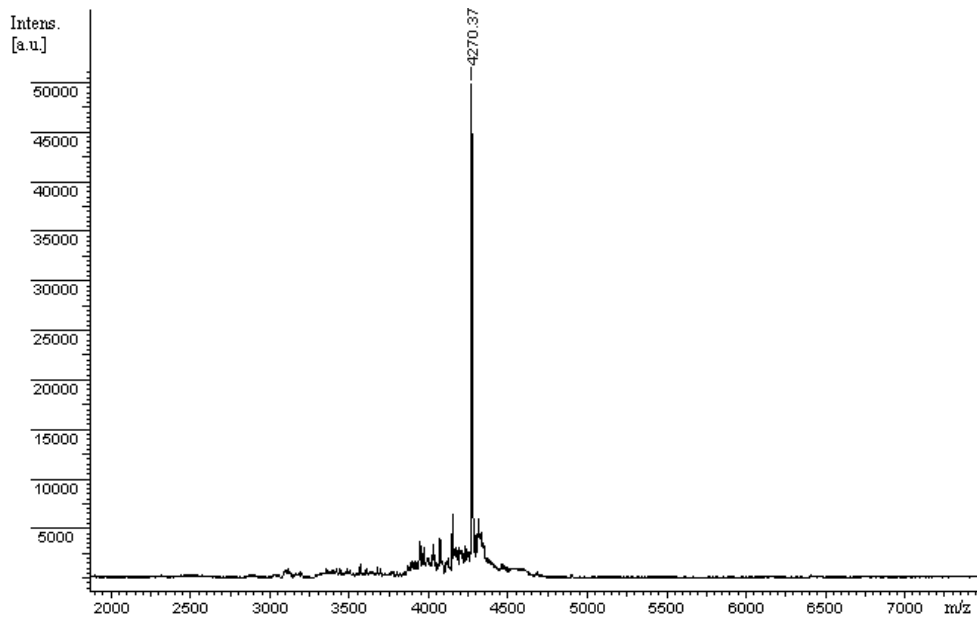
Table 1. MALDI MS analysis of the oligonucleotides used in the study.

N	Conjugate	Calculated mass, M+H ⁺	Found mass, M+H ⁺
1	11D-E	4269.3	4270.4
2	11D-E2	4091.2	4091.4
3	5'-FAM1-11D	4035.8	4036.6
4	5'- FAM2-11D	3924.7	3923.9
5	11D-3'-FITC	3953.7	3954.2
6	HEX-11D	4129.4	4130.2
7	HEX_{Acr}-11D	4128.4	4126.3
8	TET-11D	4060.4	4060.2
9	JOE-11D	4049.6	4049.4
10	11D-Ole	3860.8	3860.9
11	FAM-11D-Ole	4511.4	4512.4
12	11D- FAM-Ole	4511.4	4512.4
13	11DX-E-1	4263.4	4264.2
14	11DX-E-2	4263.4	4263.9
15	11DX-E-3	4263.4	4264.2
16	11DS-E-1	4430.0	4432.9
17	11DS-E-2	4301.5	4302.7
18	11DS-E-3	4301.5	4304.0
19	11DS-E-4	4301.5	4299.5
20	HEX-11-ddR	2788.3	2788.6
21	HEX-11D-ddR-1	3334.7	3337.9
22	HEX-11D-ddR-2	3334.7	3335.2
23	HEX-11-PD	2368.0	2368.9
24	HEX-11D-PD-1	3082.5	3082.1
25	HEX-11D-PD-2	3082.5	3083.1
26	11M-E	4501.4	4501.6
27	HEX-11M	4129.3	4361.9
28	HEX-11MS	4538.2	4537.1
29	HEX-11MS-Ole	5029.8	5028.5

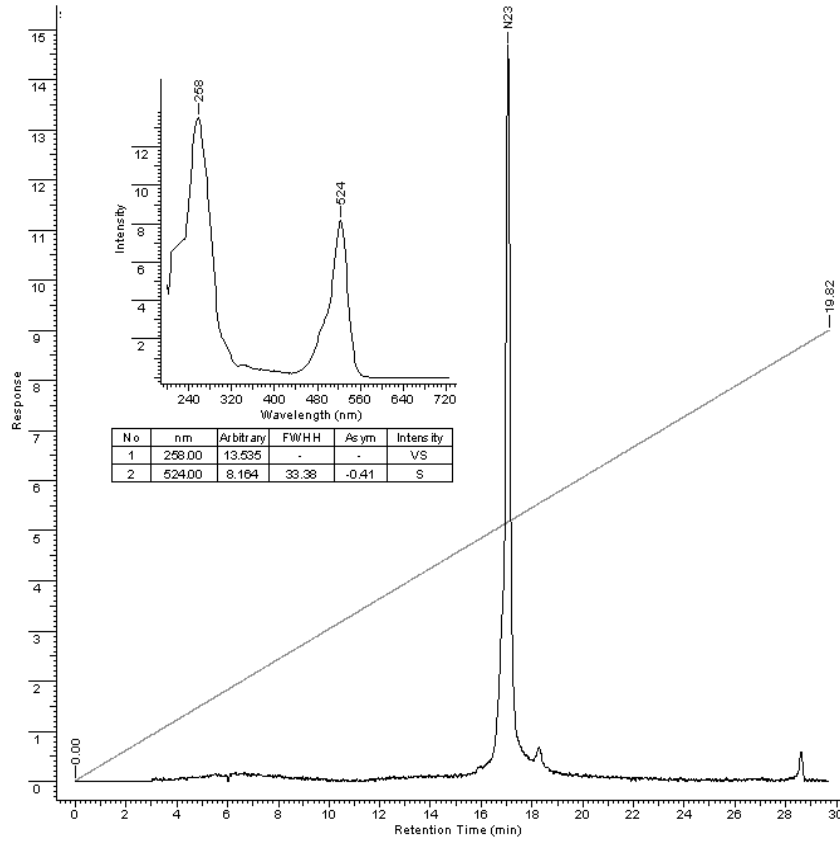
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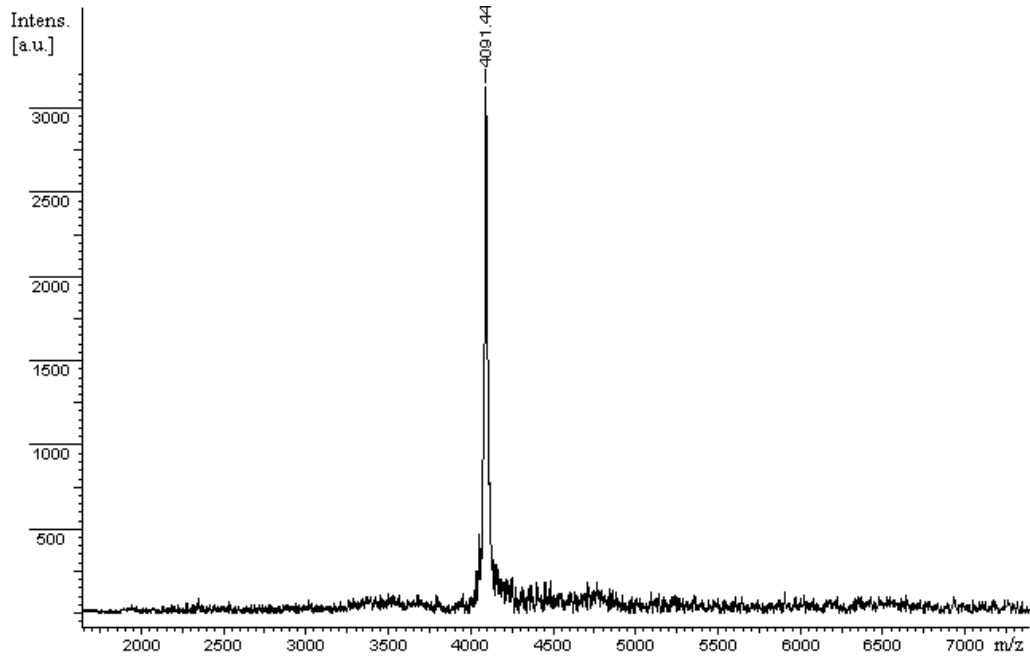
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N28	21.968	500927.625	91.924	0.318	1.000	1.531	76157.781



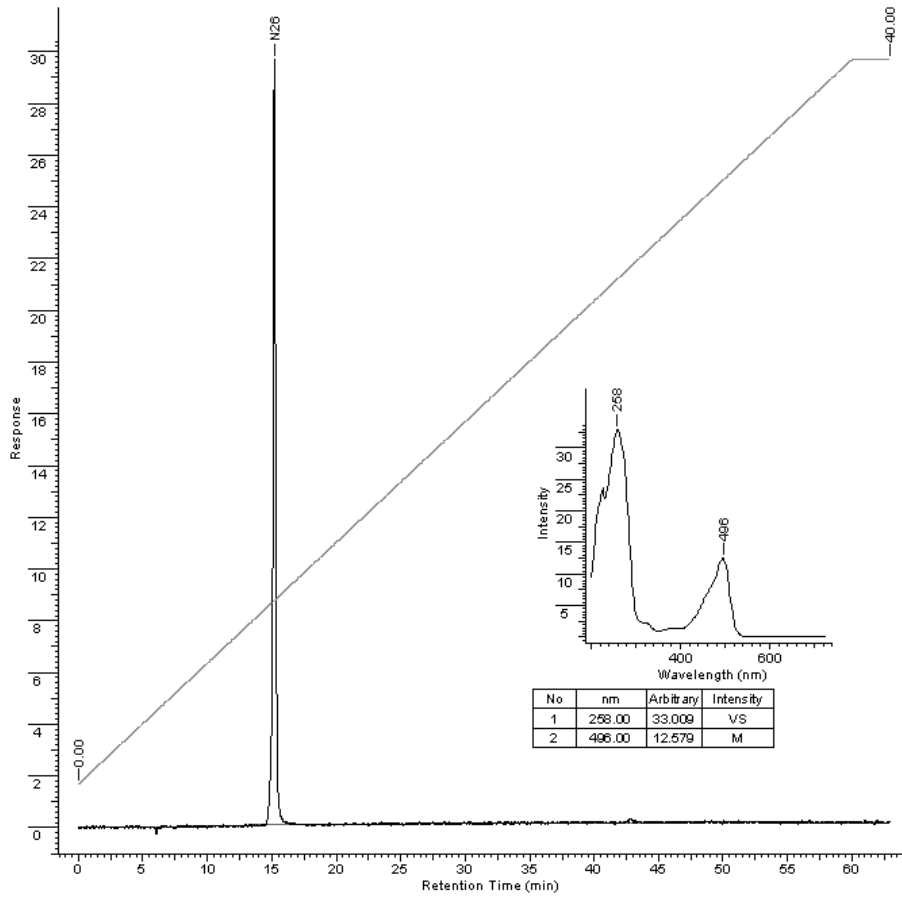
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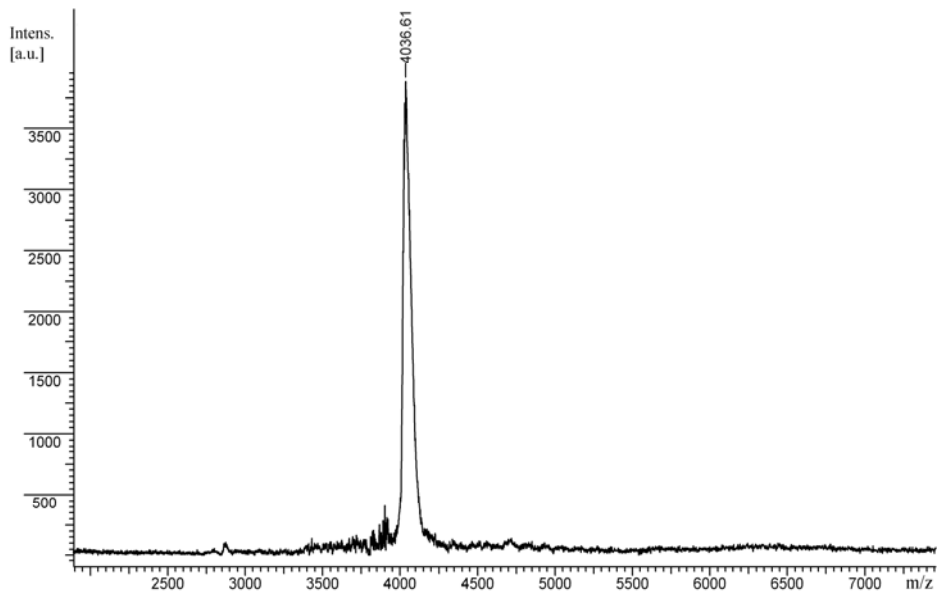
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N23	17.063	239591.859	100.000	0.305	1.095	50088.266



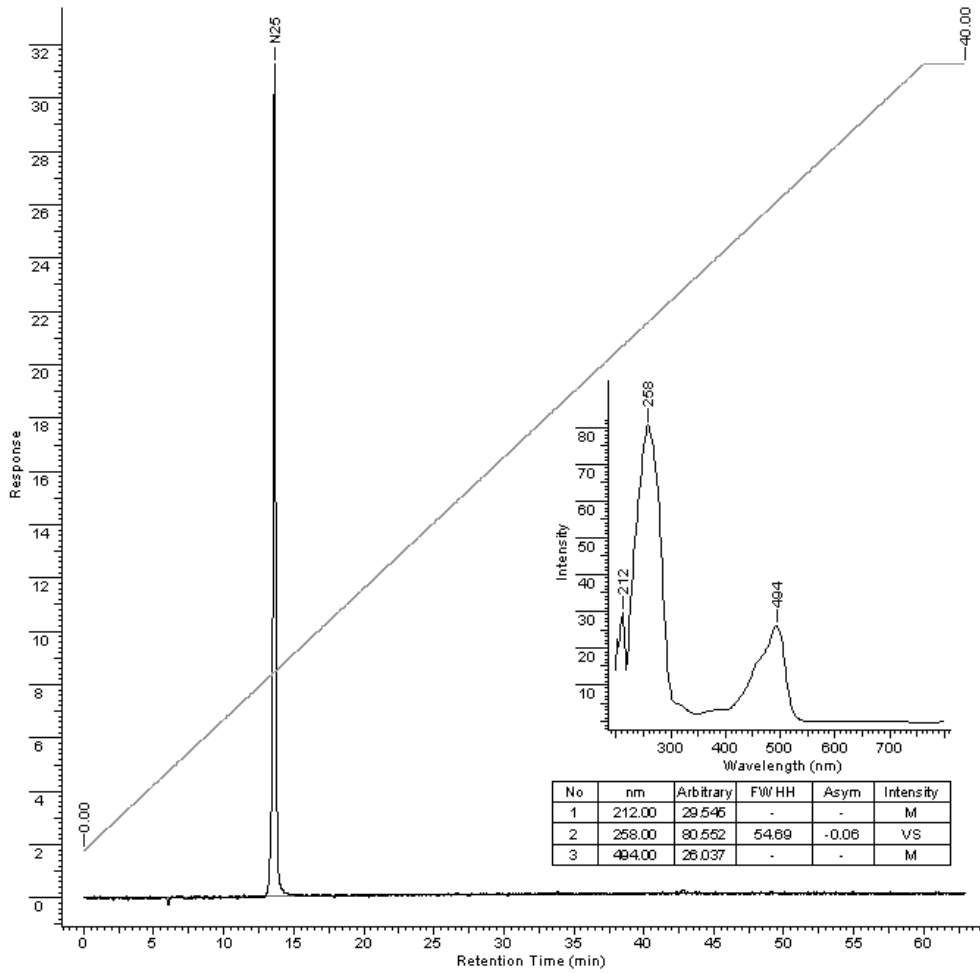
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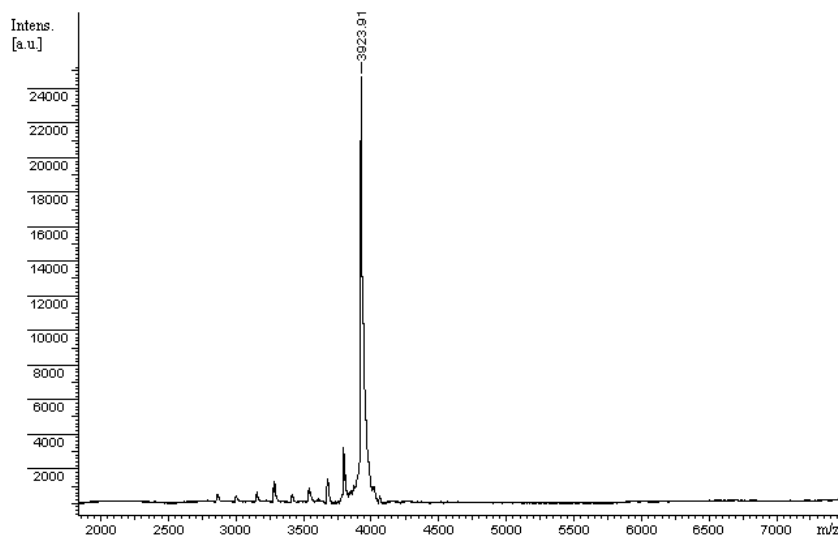
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N26	15.198	400013.188	100.000	0.279	1.000	47611.262



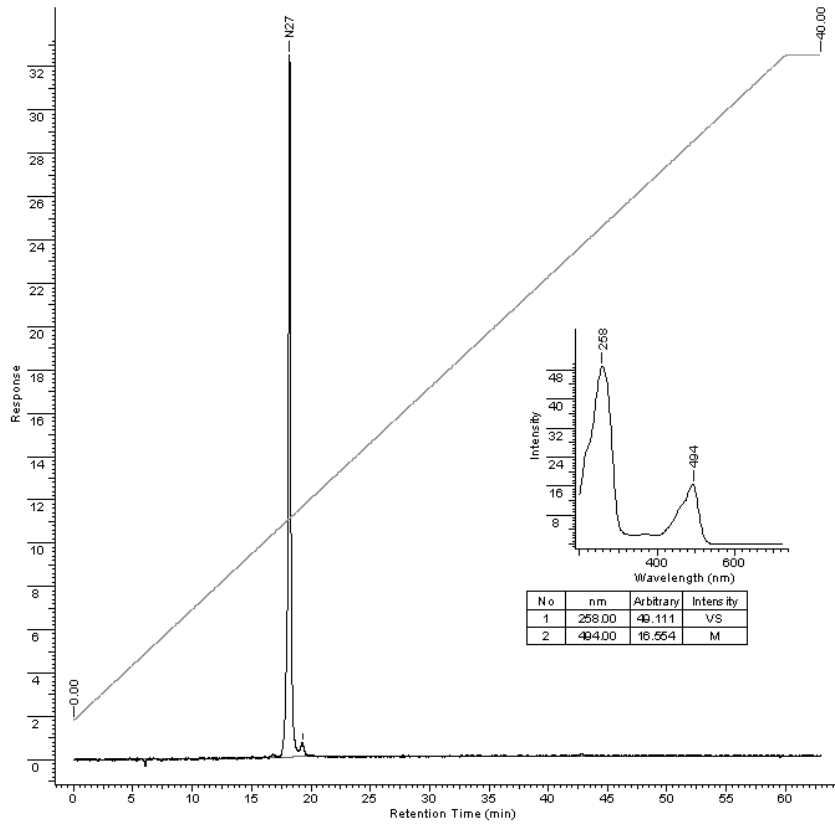
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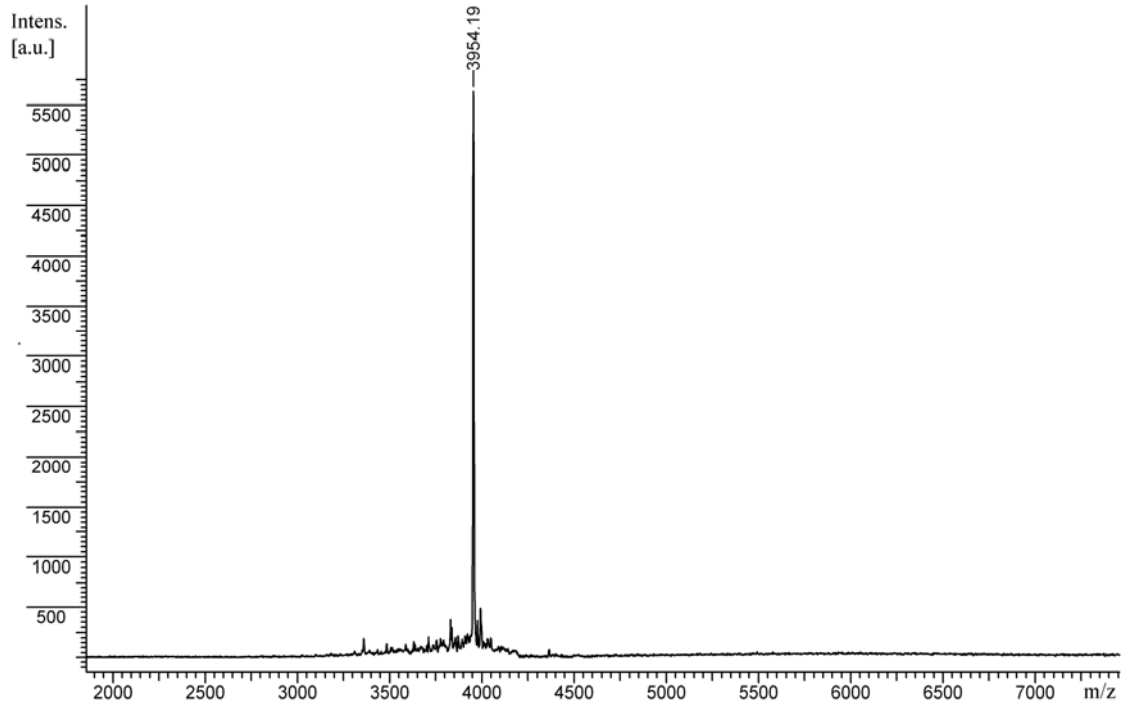
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N25	13.613	374074.500	100.000	0.249	0.923	47913.387



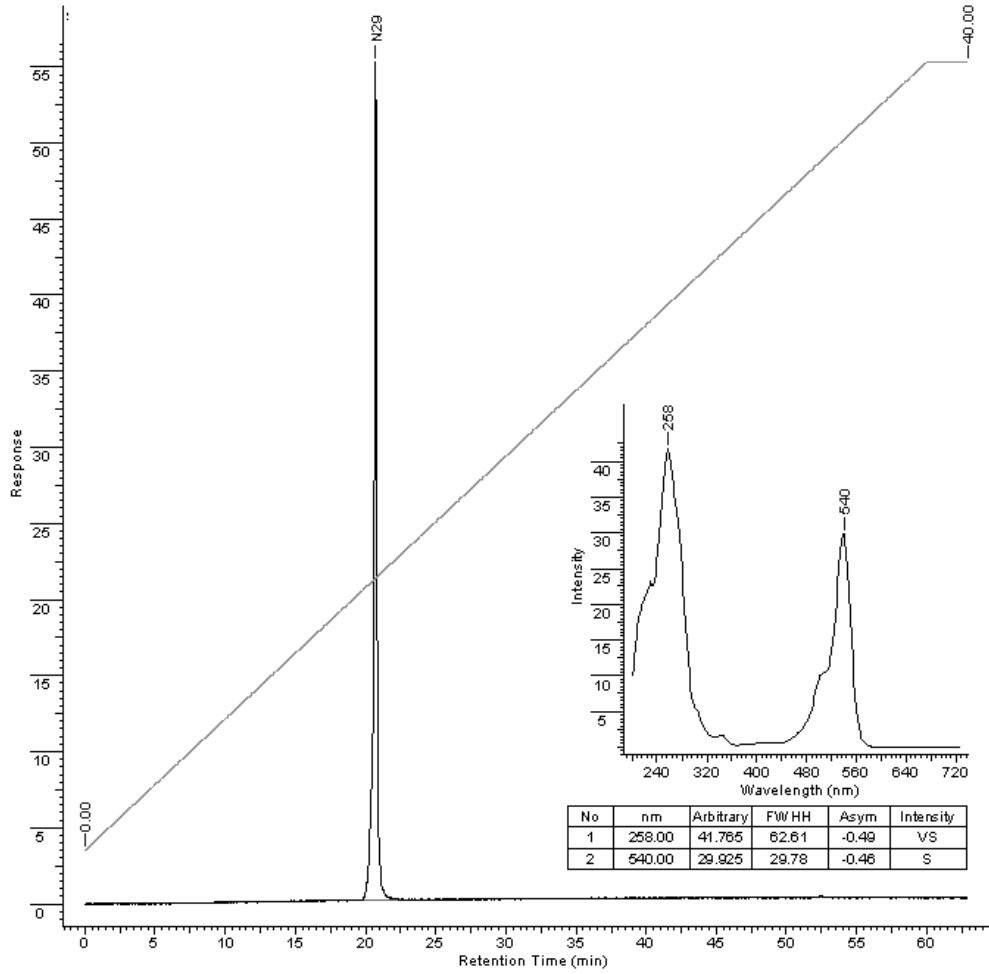
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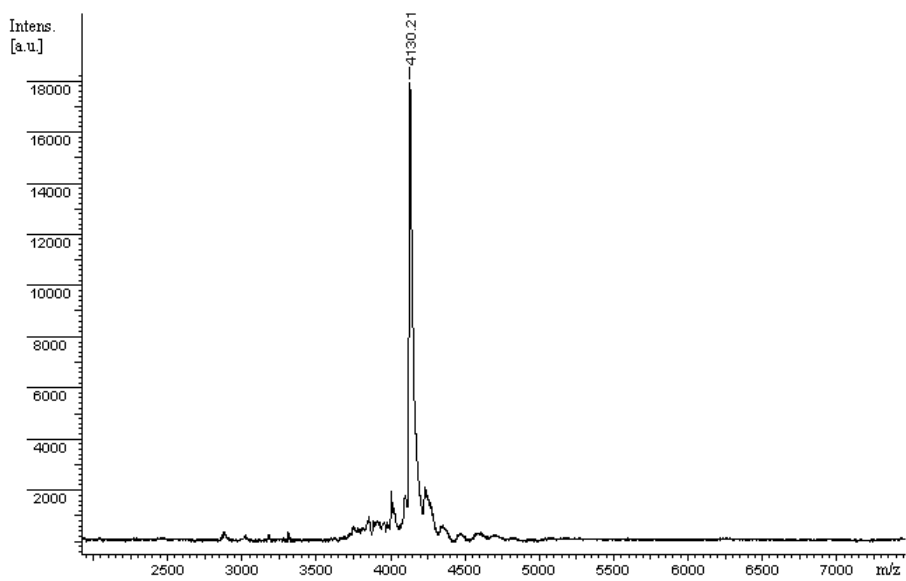
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N27	18.184	503382.969	97.323	0.308	1.067	2.632	55604.047
unknown	19.271	13844.640	2.677	0.517	0.733	-	22194.207



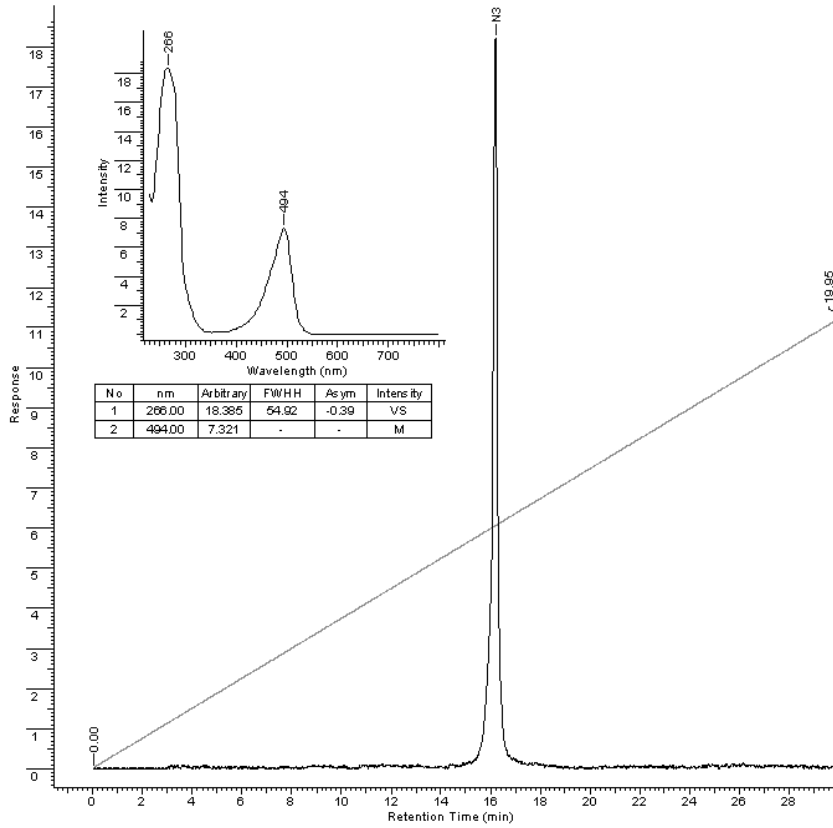
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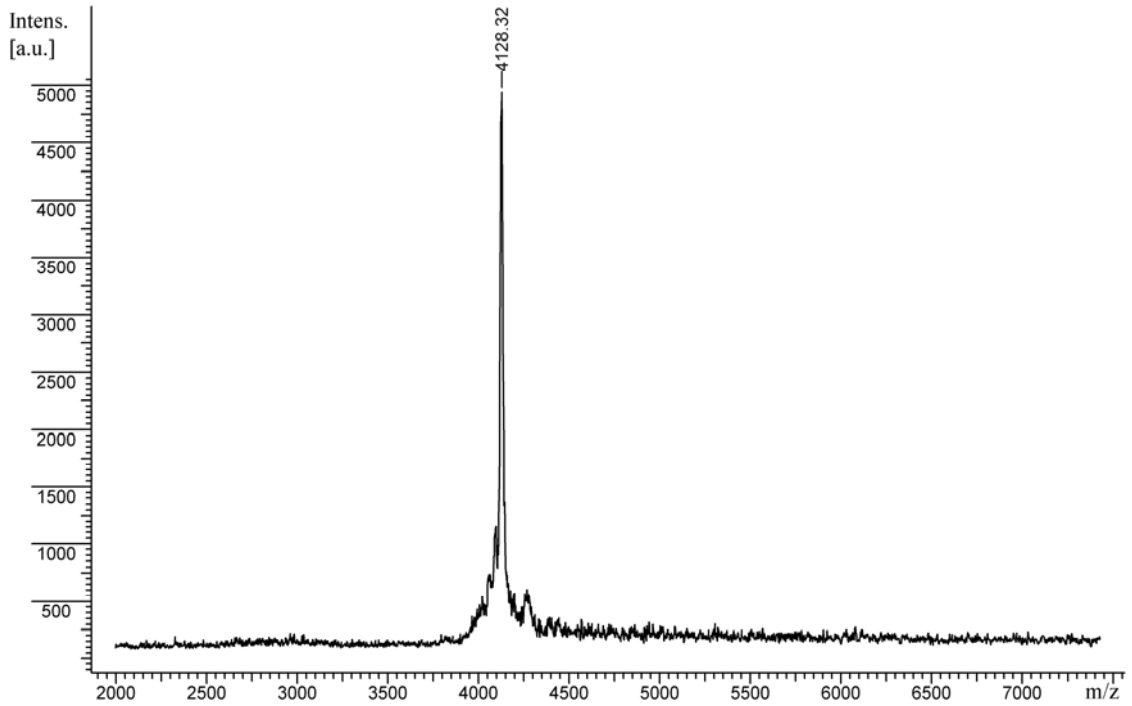
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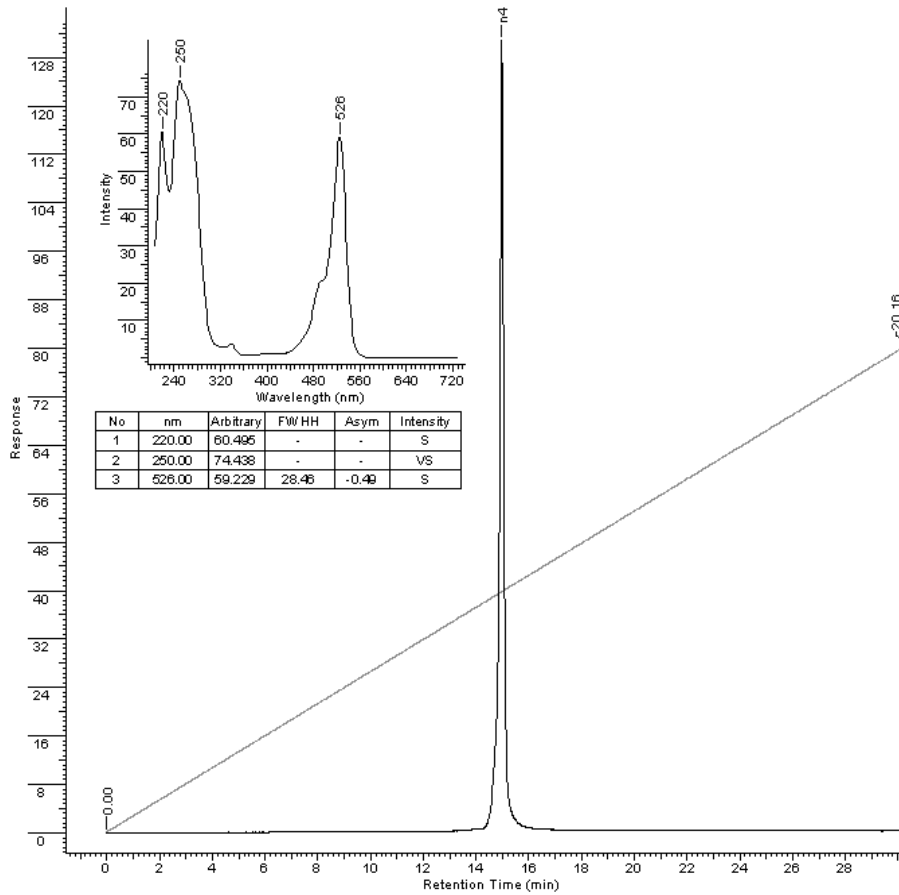
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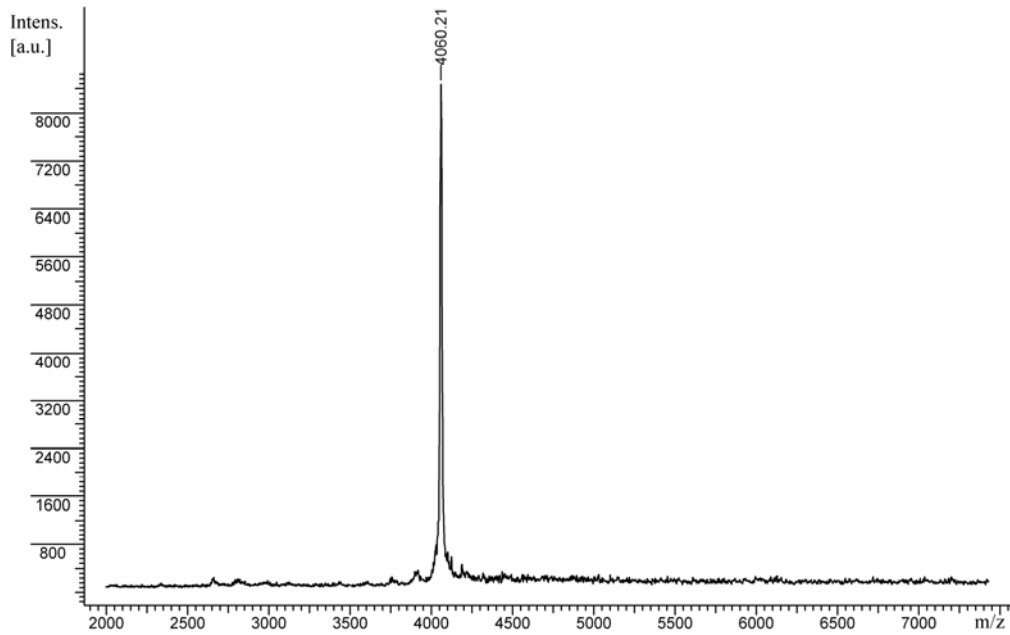
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N3	16.198	272590.375	100.000	0.279	0.867	54066.191



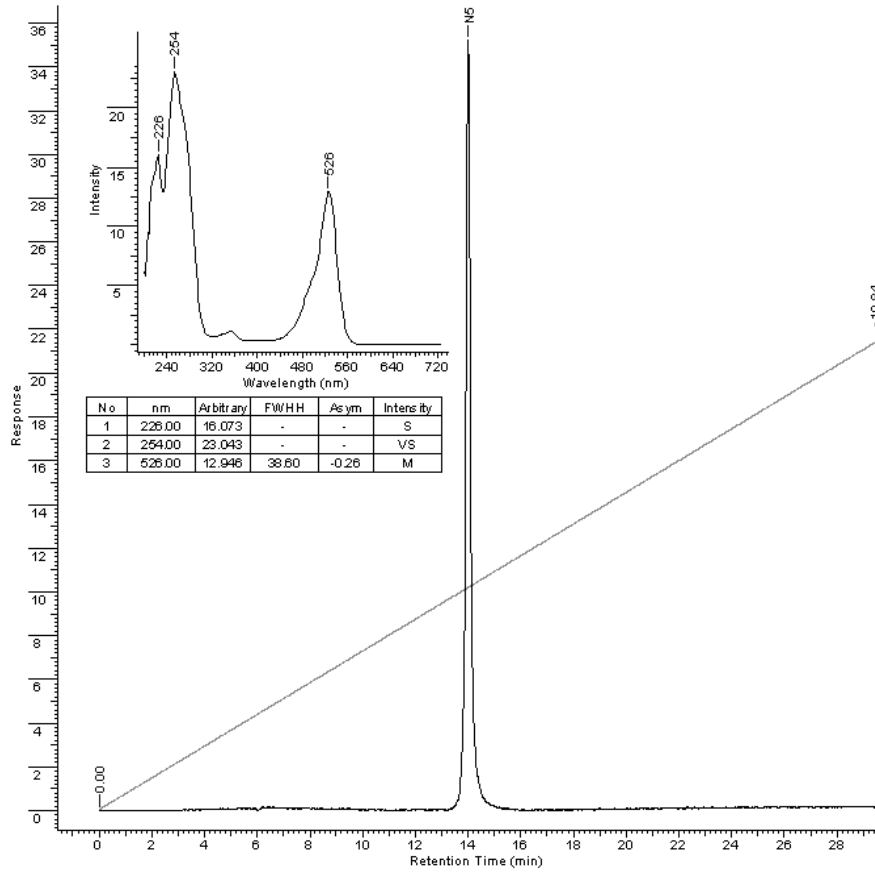
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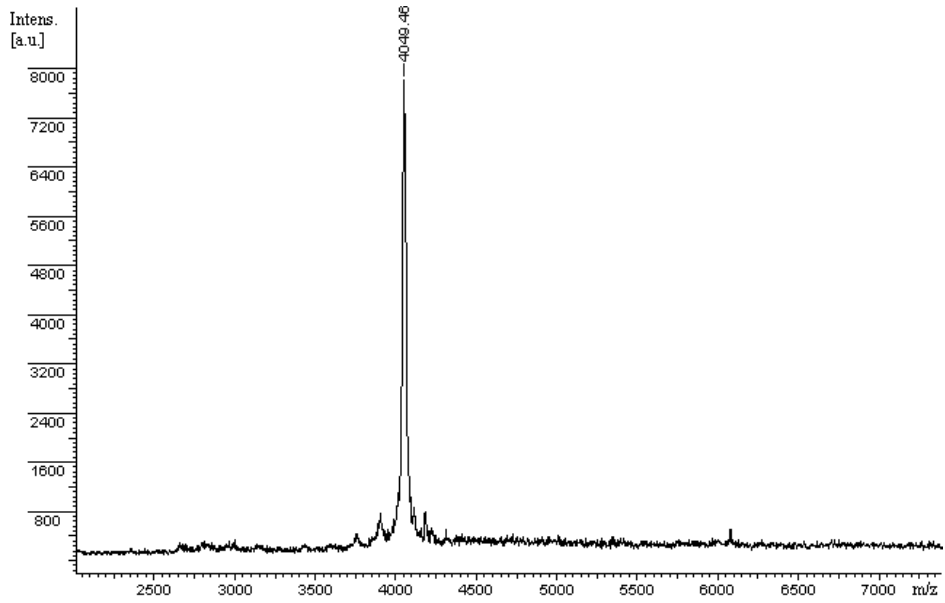
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n4	14.975		1579941.500	100.000	0.260	1.066	53220.902



JOE-11D

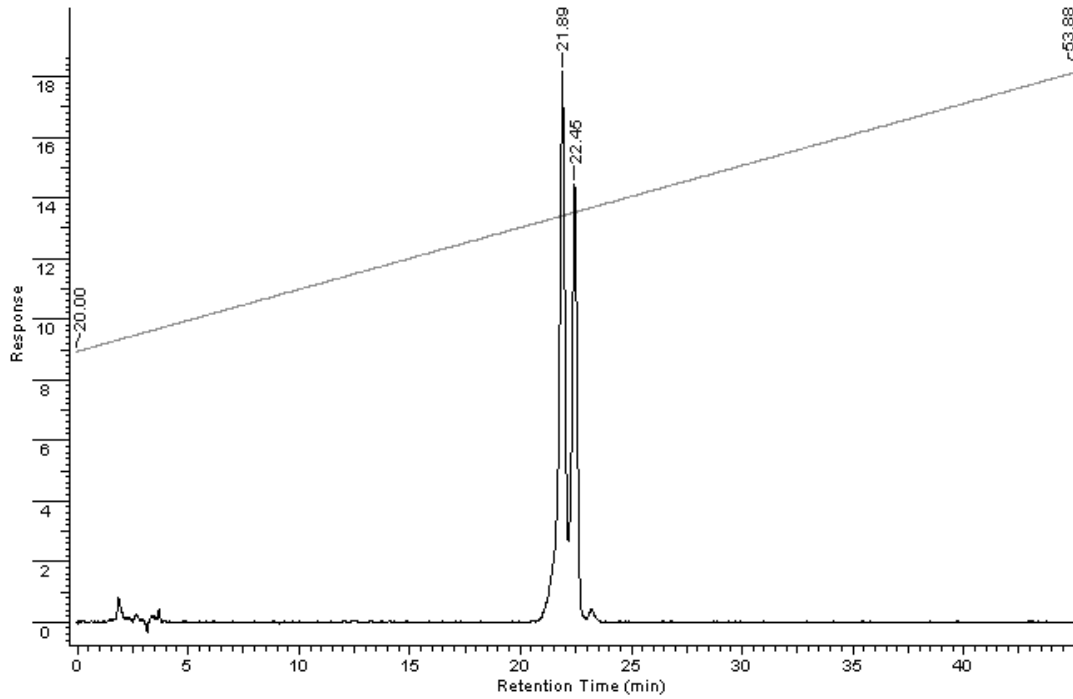


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N5	14.009	455089.594	100.000	0.291	0.952	37183.137

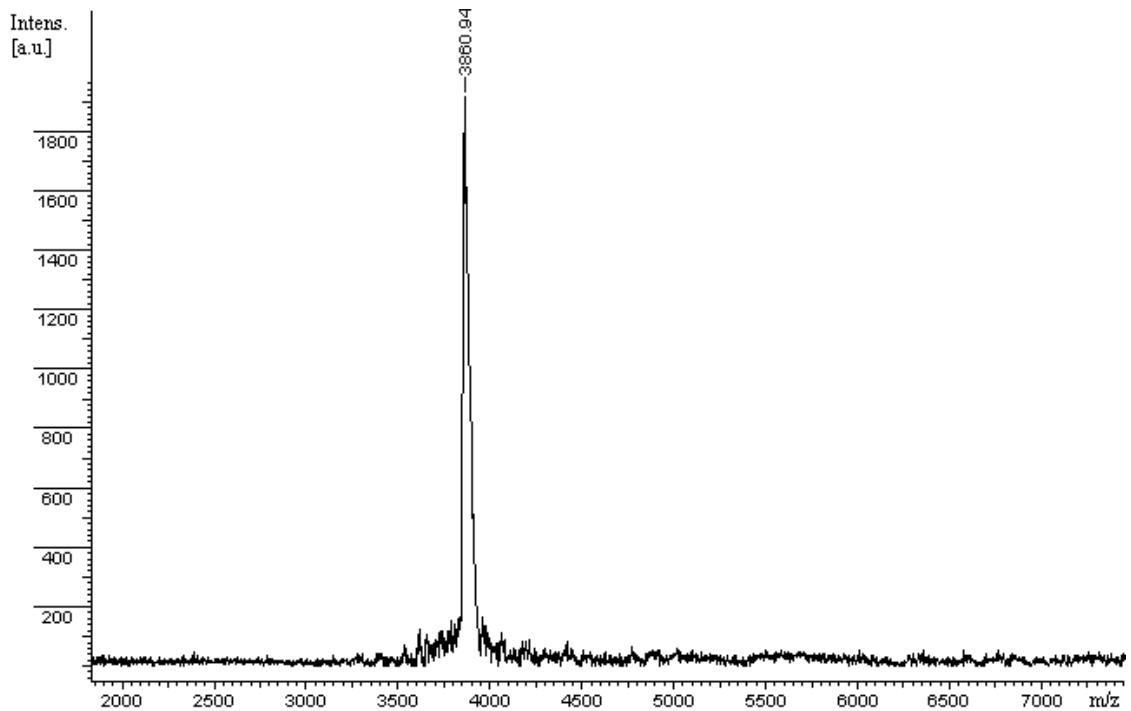


11D-Ole

Double peak on HPLC is a result of the presence of the stereocenter in glycerol moiety (Figure 1). Mass spectrum is recorded for a mixture of peaks.

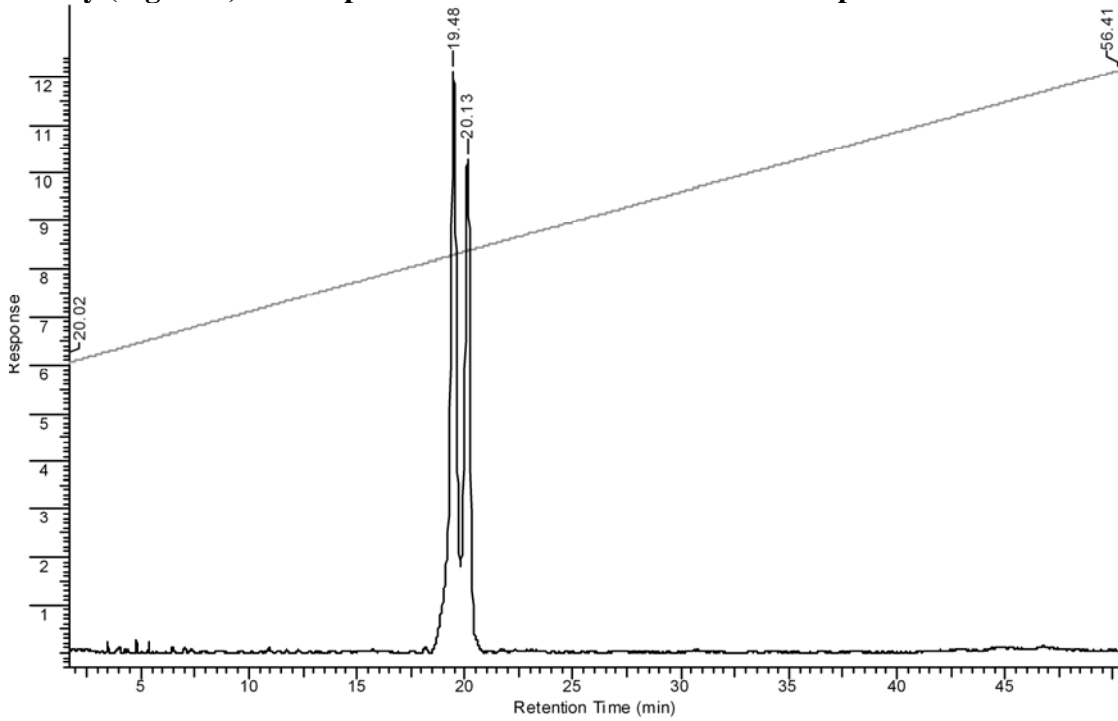


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N6	21.894	344811.125	56.923	0.388	0.950	1.463	50929.844
unknown	22.447	236705.625	39.076	0.368	0.947	1.896	59481.328

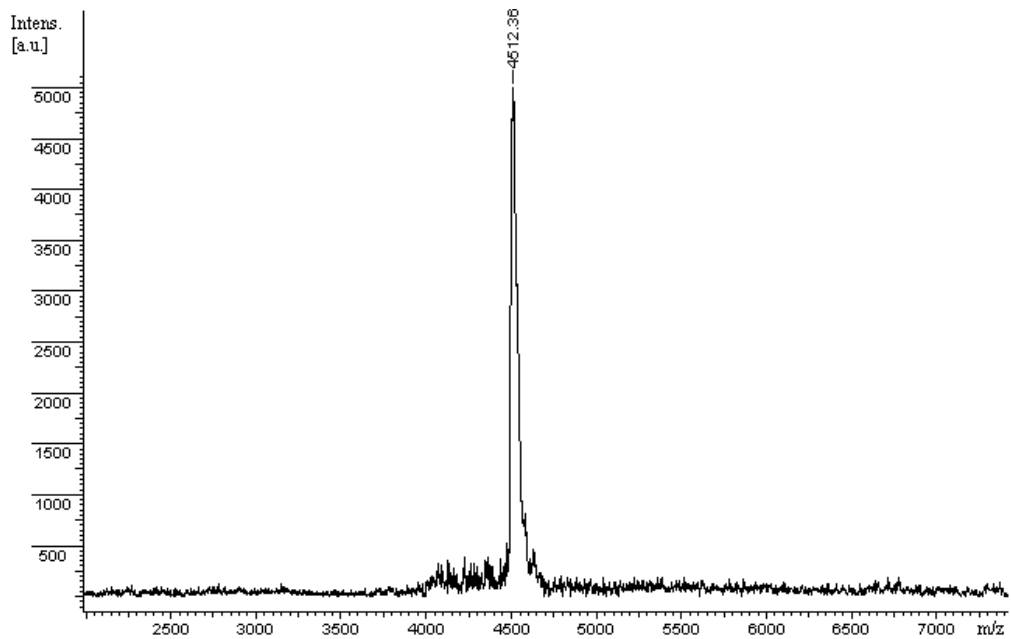


FAM-11D-Ole

Double peak on HPLC is a result of the presence of the stereocenter in glycerol moiety (Figure 1). Mass spectrum is recorded for a mixture of peaks.

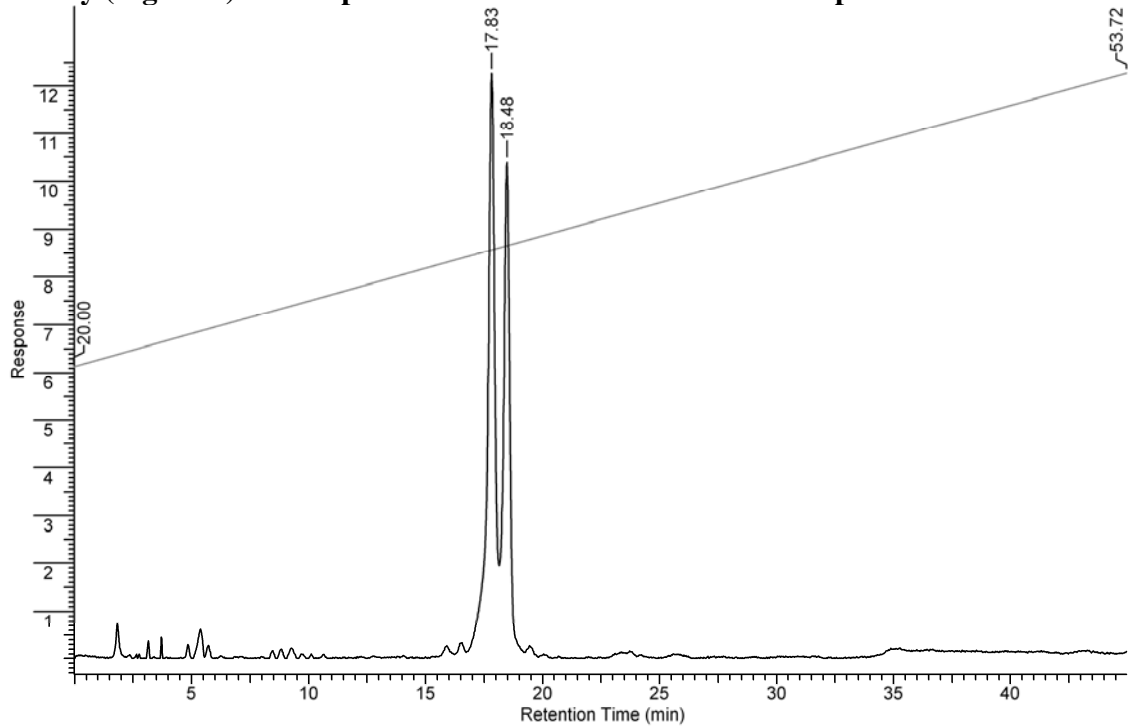


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N7	19.480	298653.354	54.870	0.435	0.952	1.357	39799.359
unknown	20.139	215639.571	39.618	0.424	0.818	-	44090.024

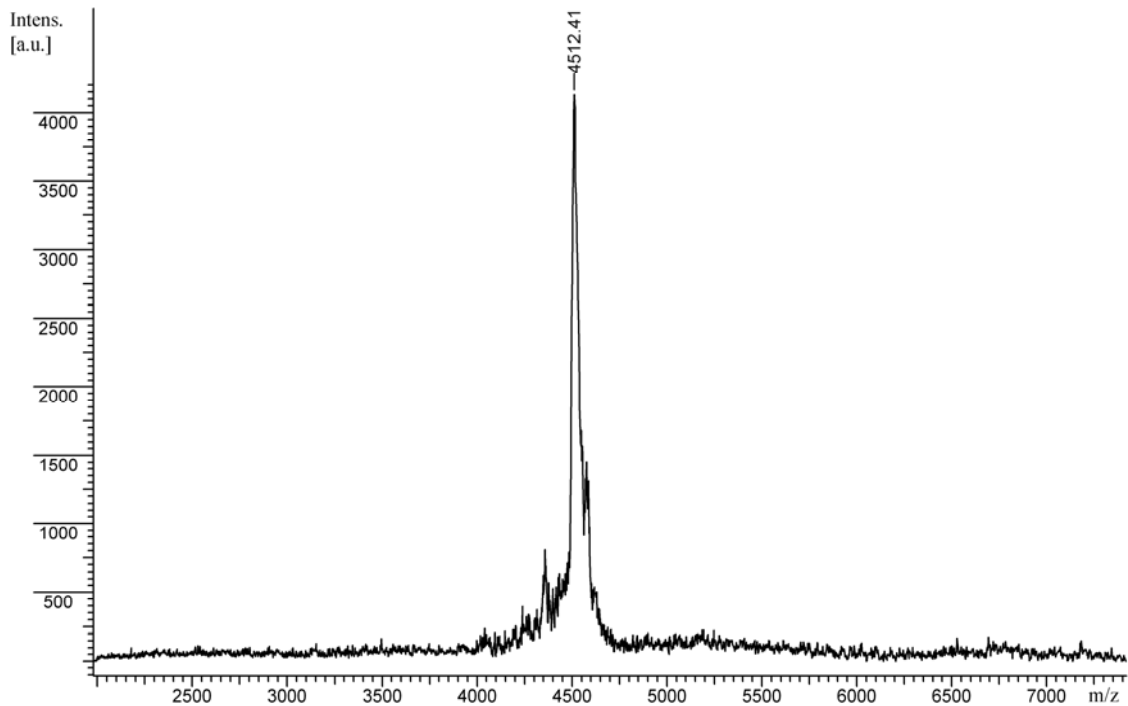


11D-FAM-Ole

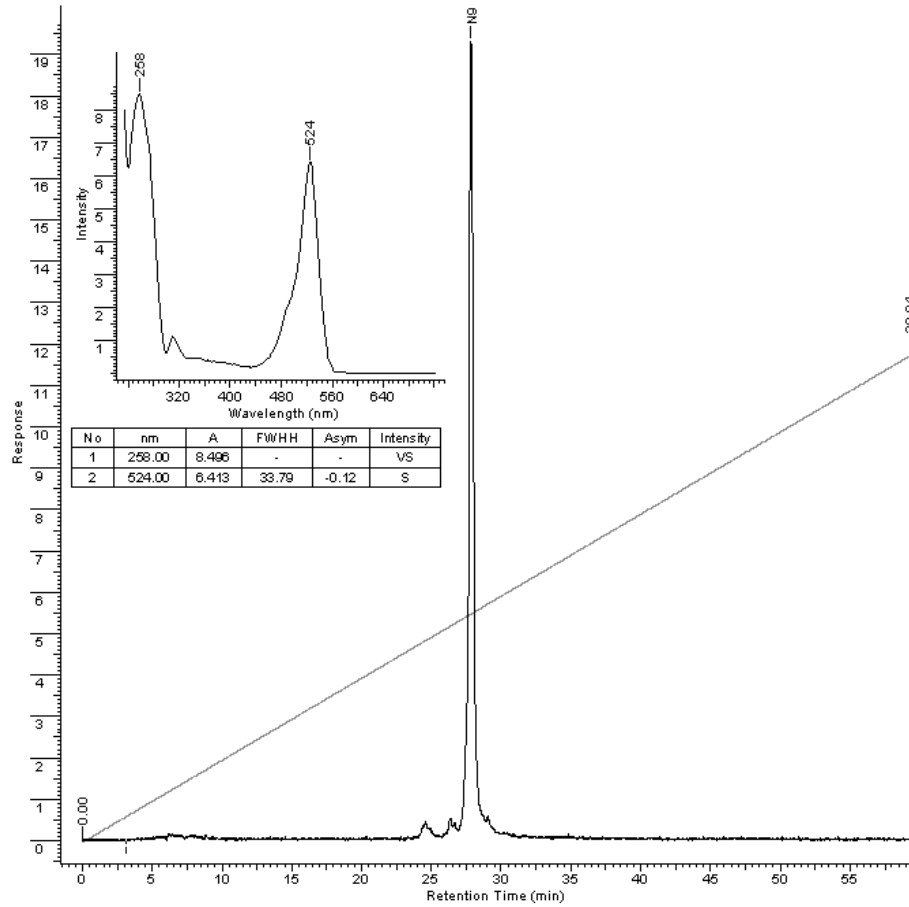
Double peak on HPLC is a result of the presence of the stereocenter in glycerol moiety (Figure 1). Mass spectrum is recorded for a mixture of peaks.



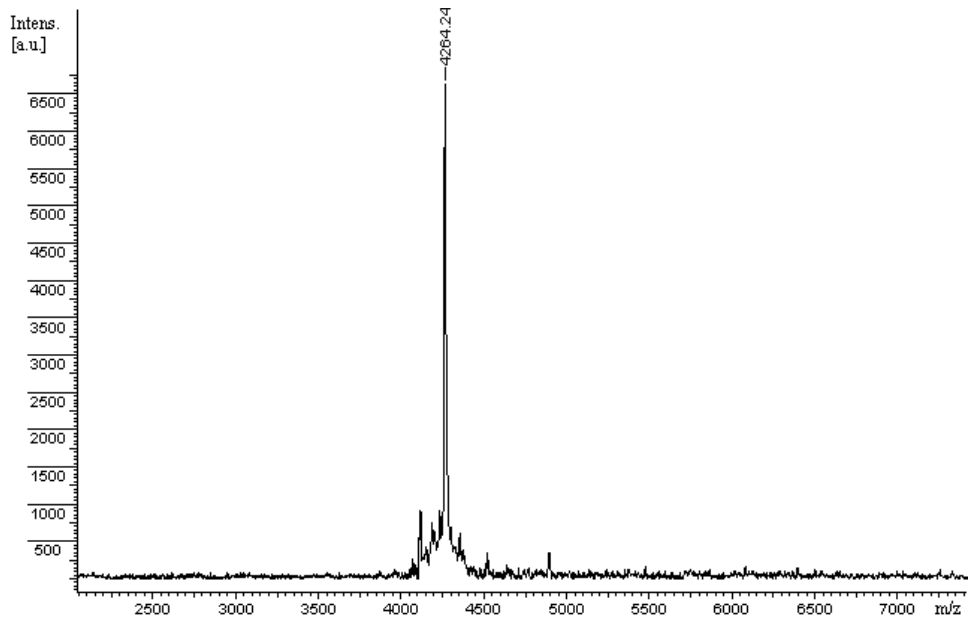
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N8	17.828	255549.922	45.452	0.428	0.955	1.545	27778.529
unknown	18.481	184714.344	32.853	0.418	0.909	-	31289.764



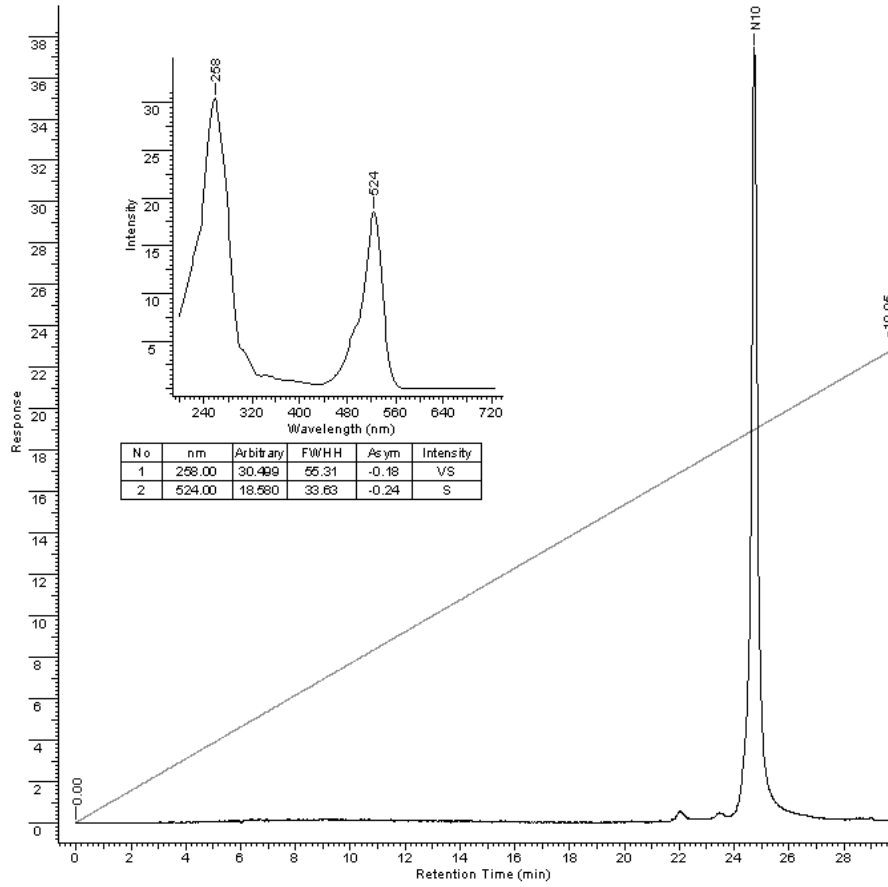
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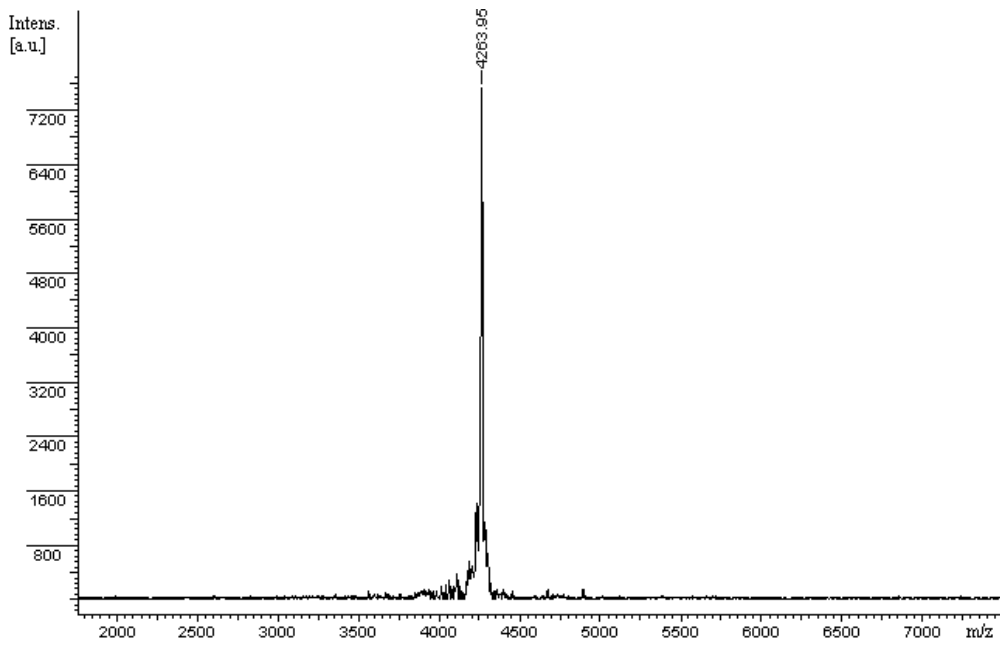
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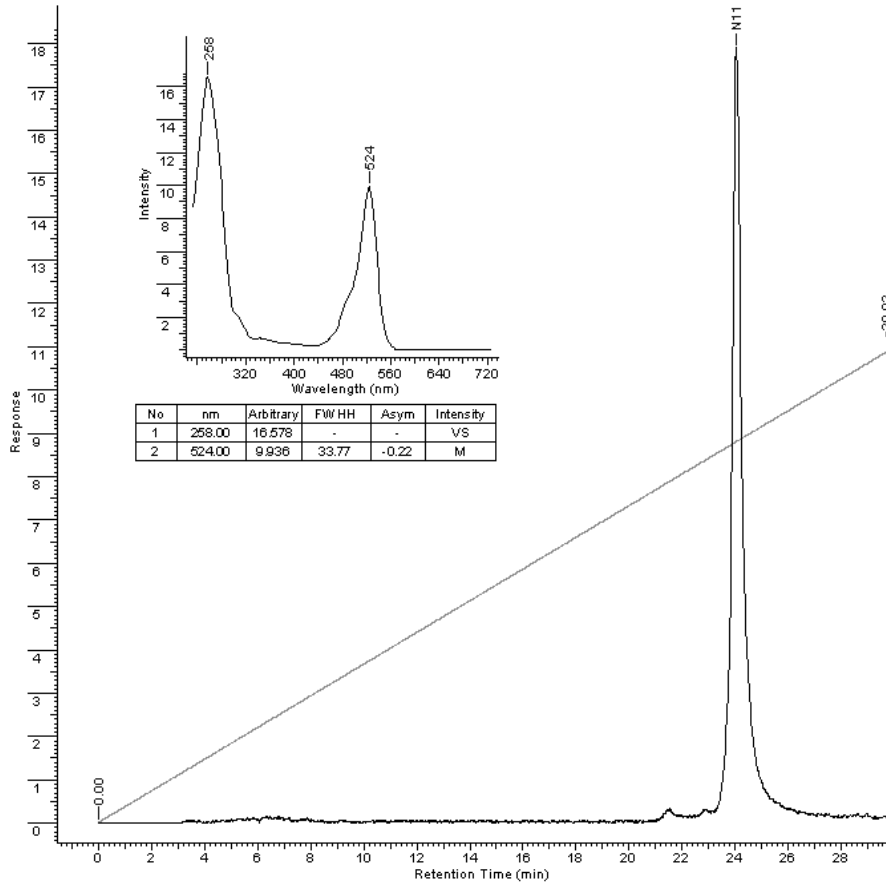
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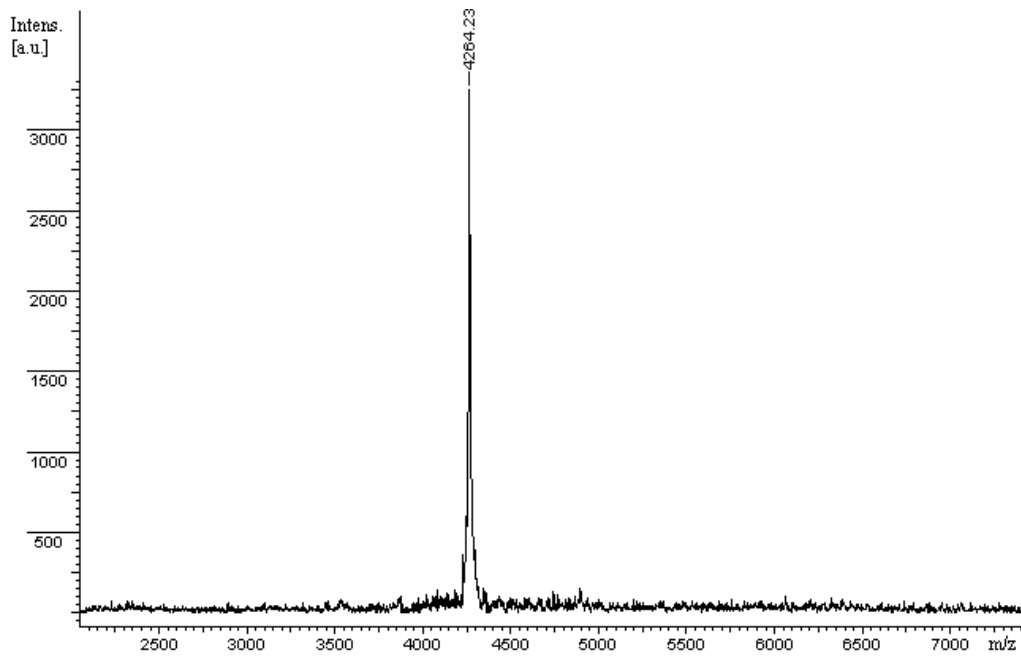
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N10	24.739	675369.750	100.000	0.366	1.020	73193.102



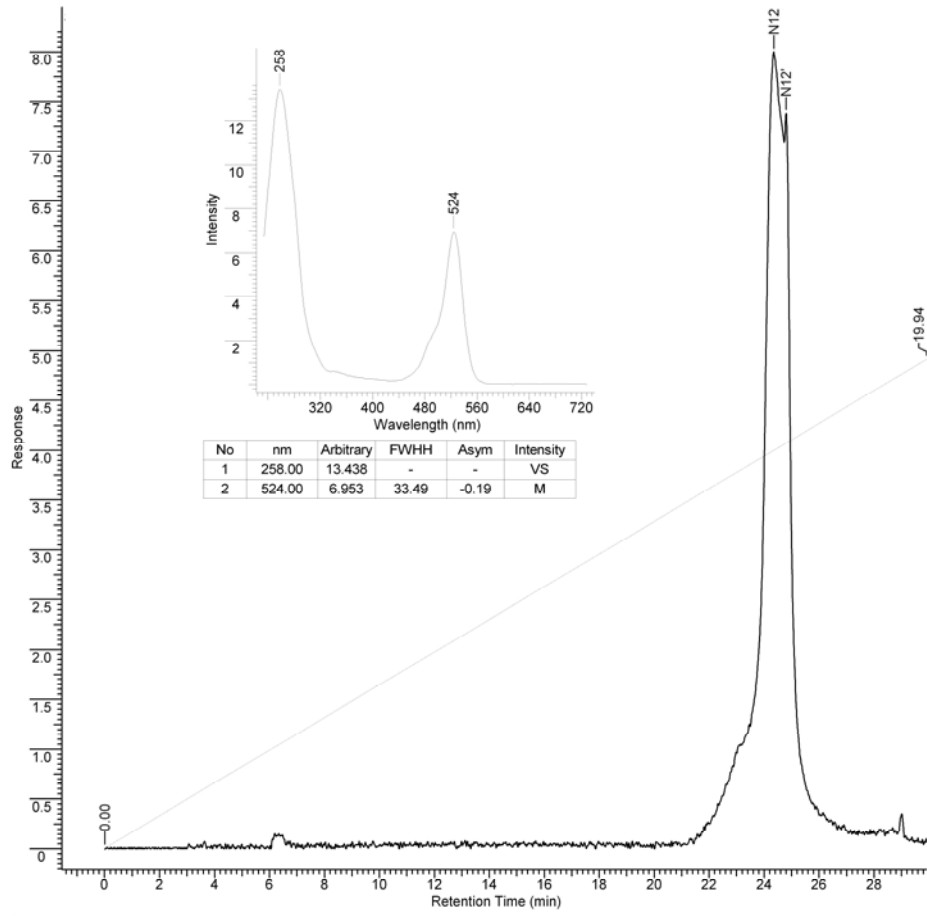
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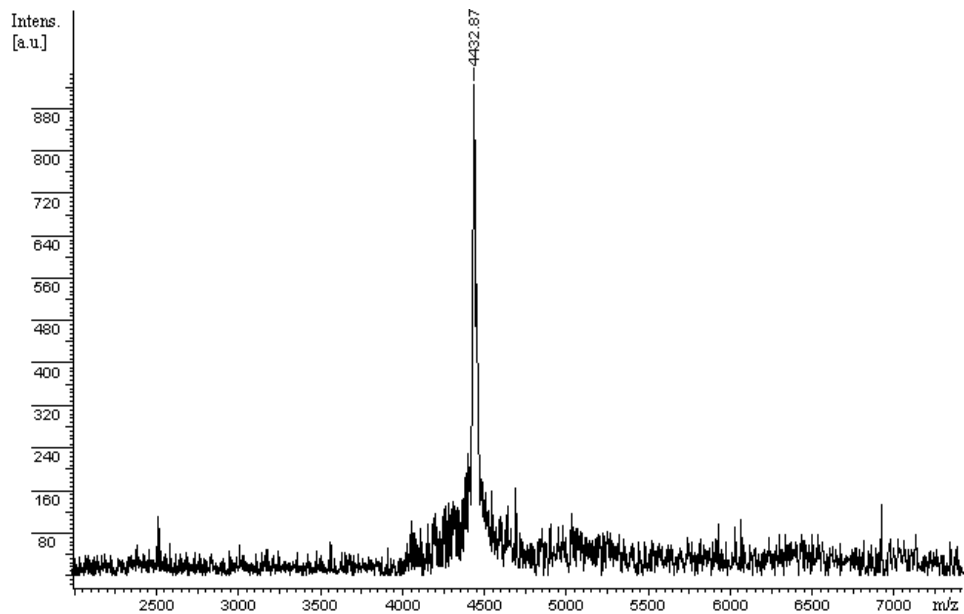
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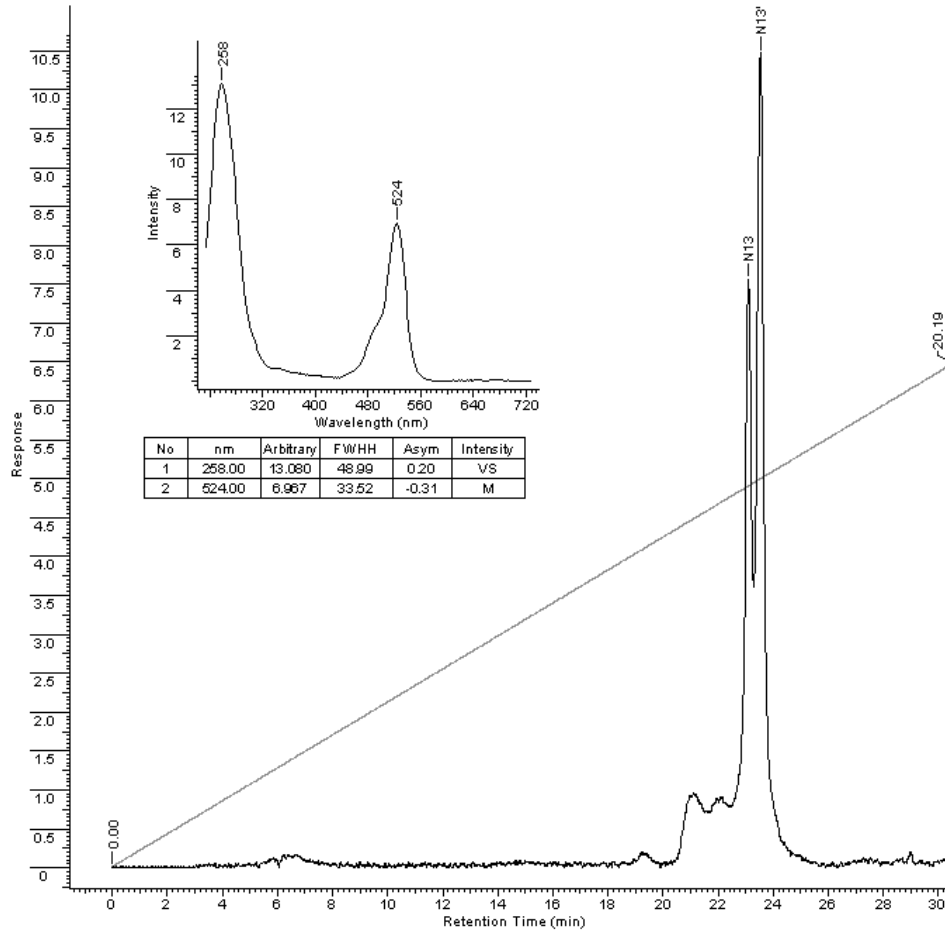
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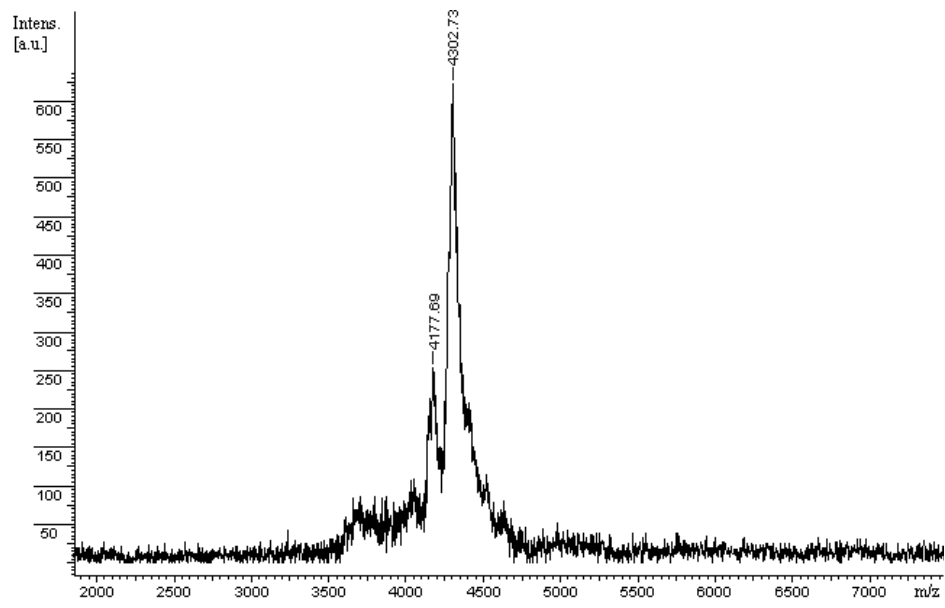
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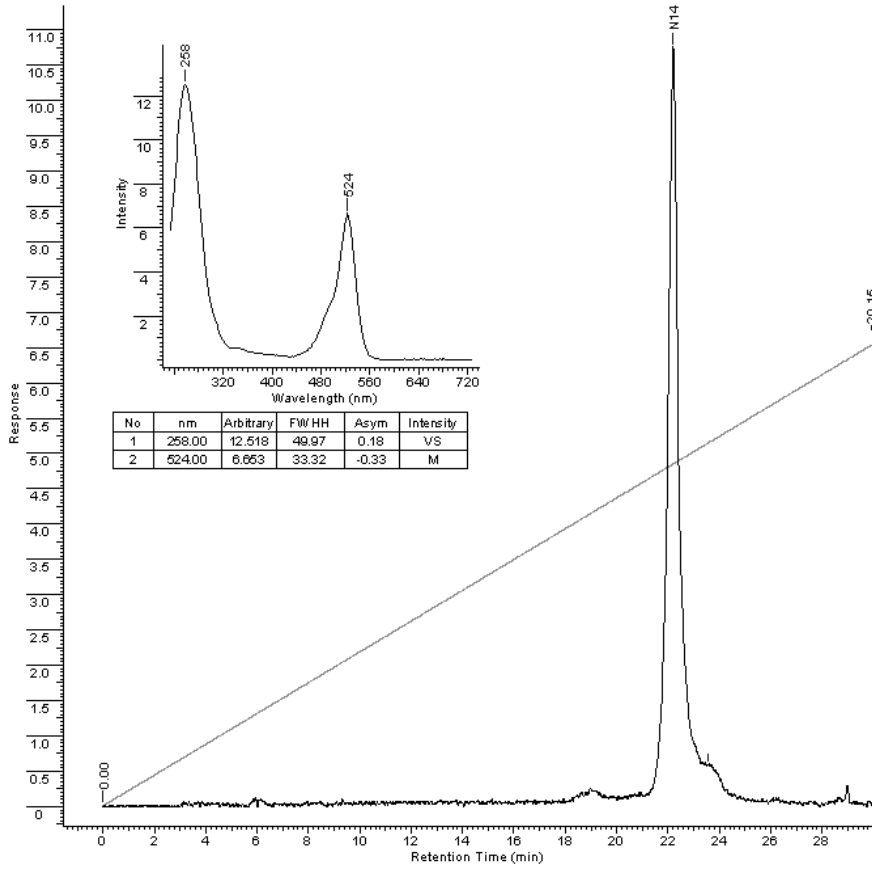
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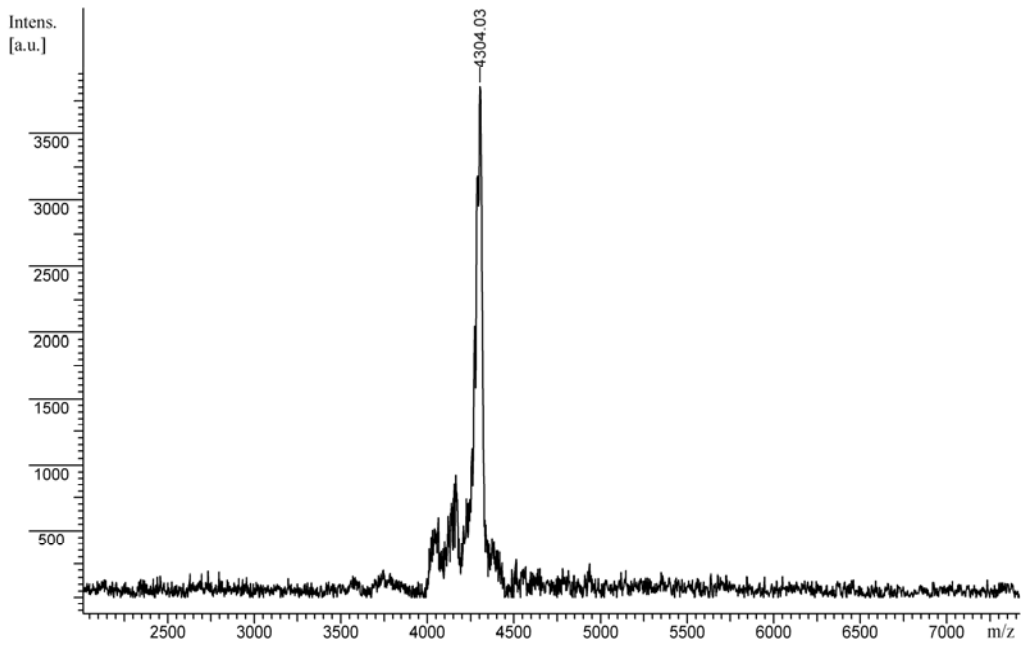
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N13	23.103	149400.844	41.727	0.430	1.094	0.978	46187.863
N13	23.536	207242.899	58.273	0.466	0.891	-	42667.832



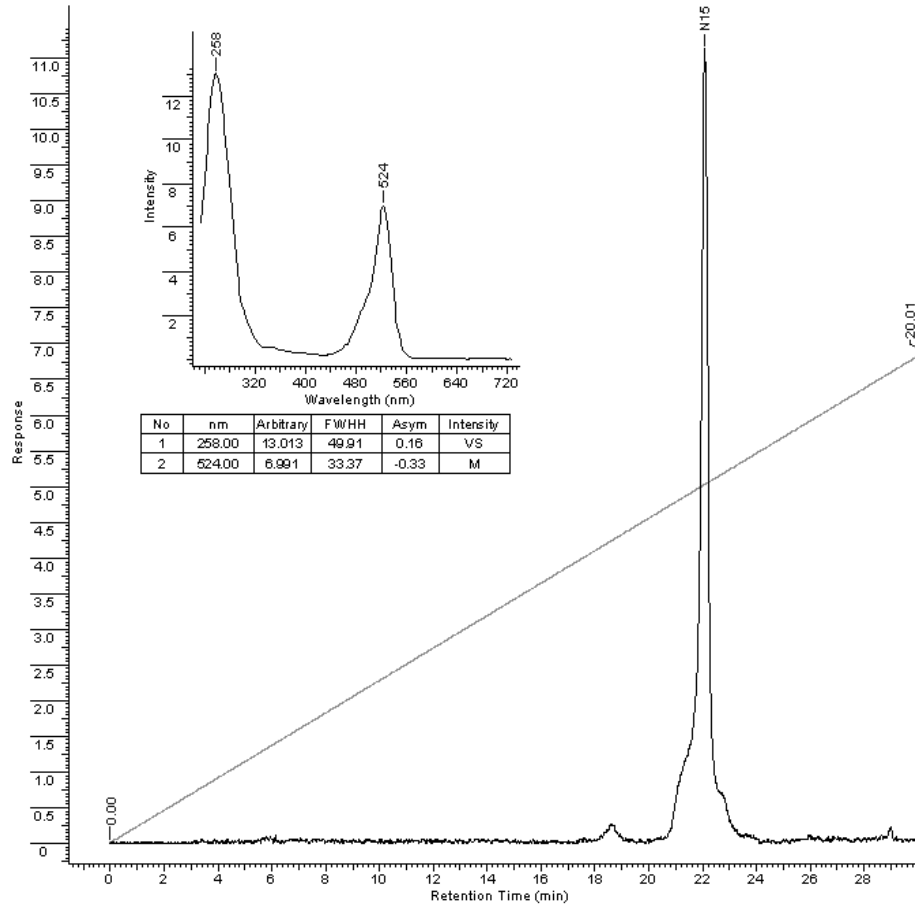
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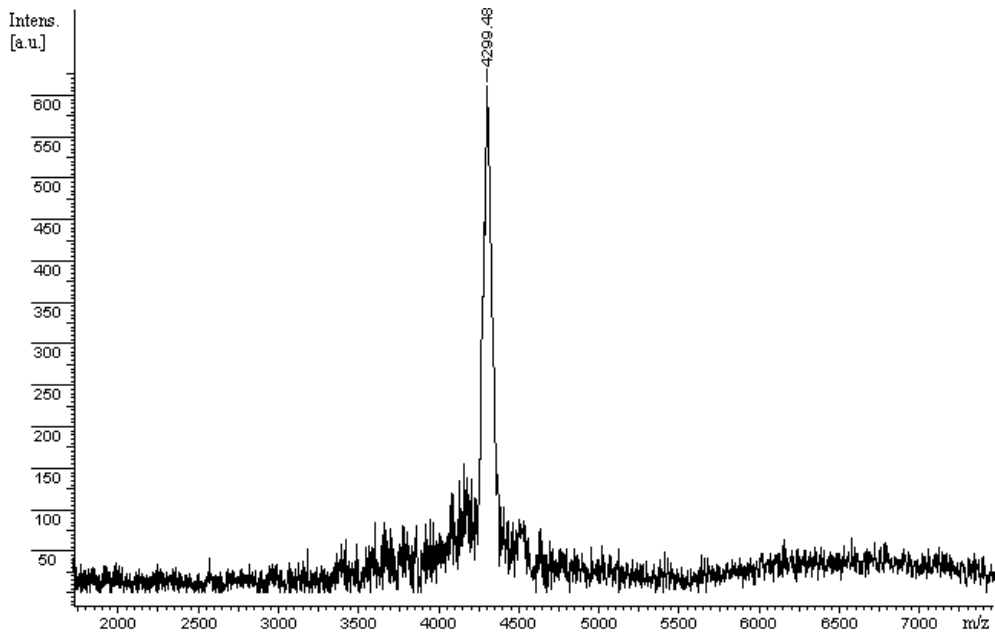
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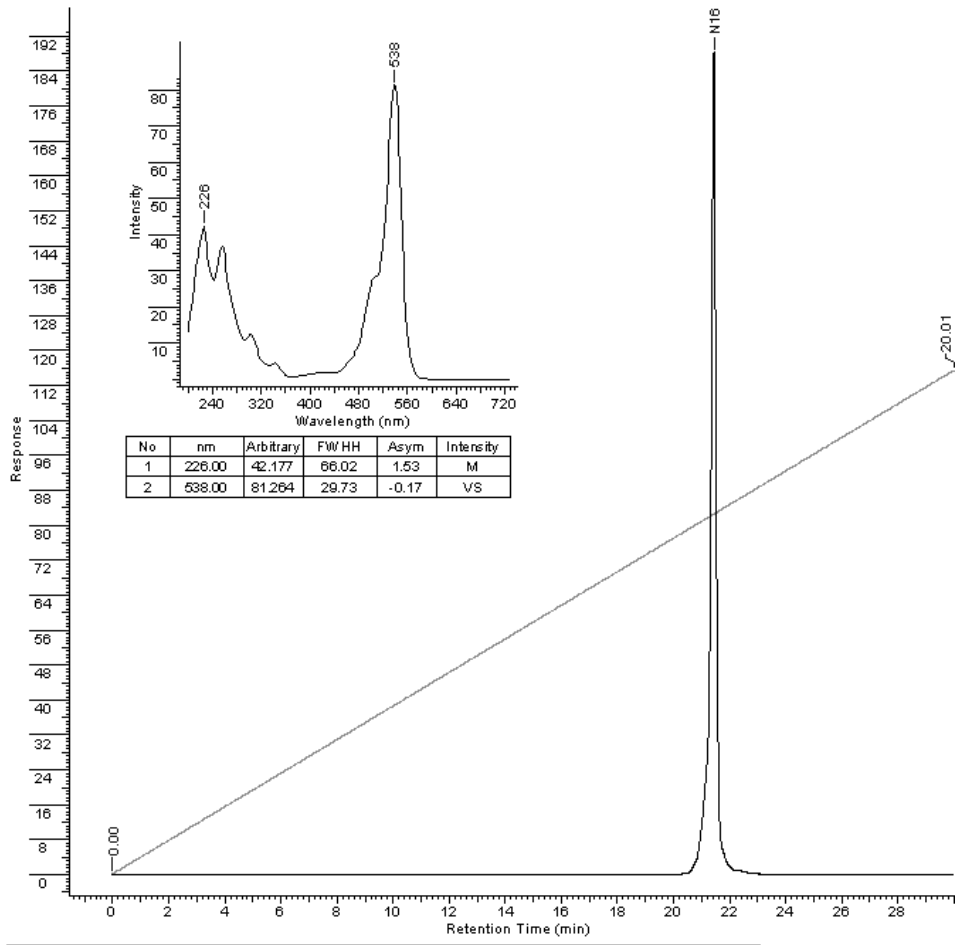
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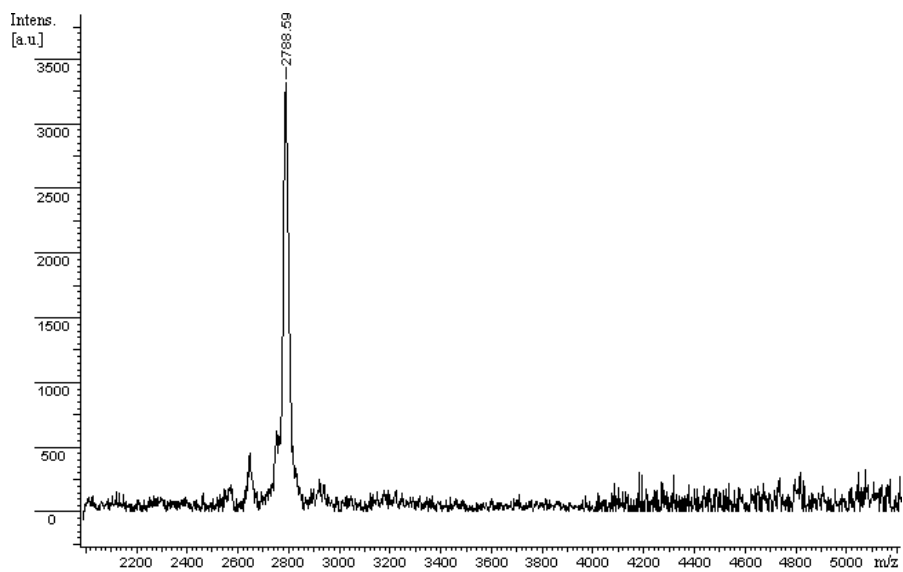
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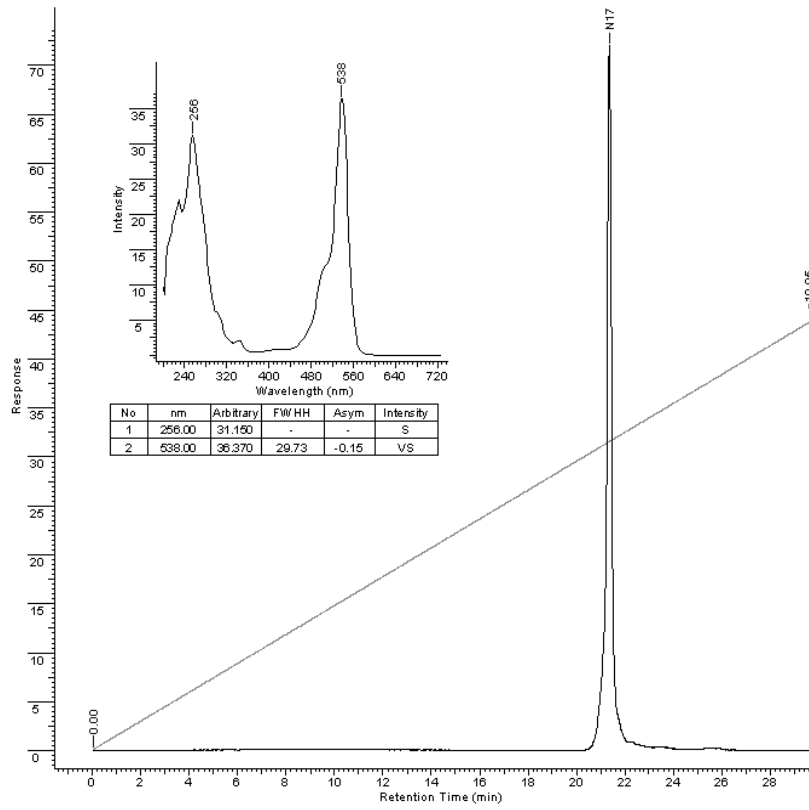
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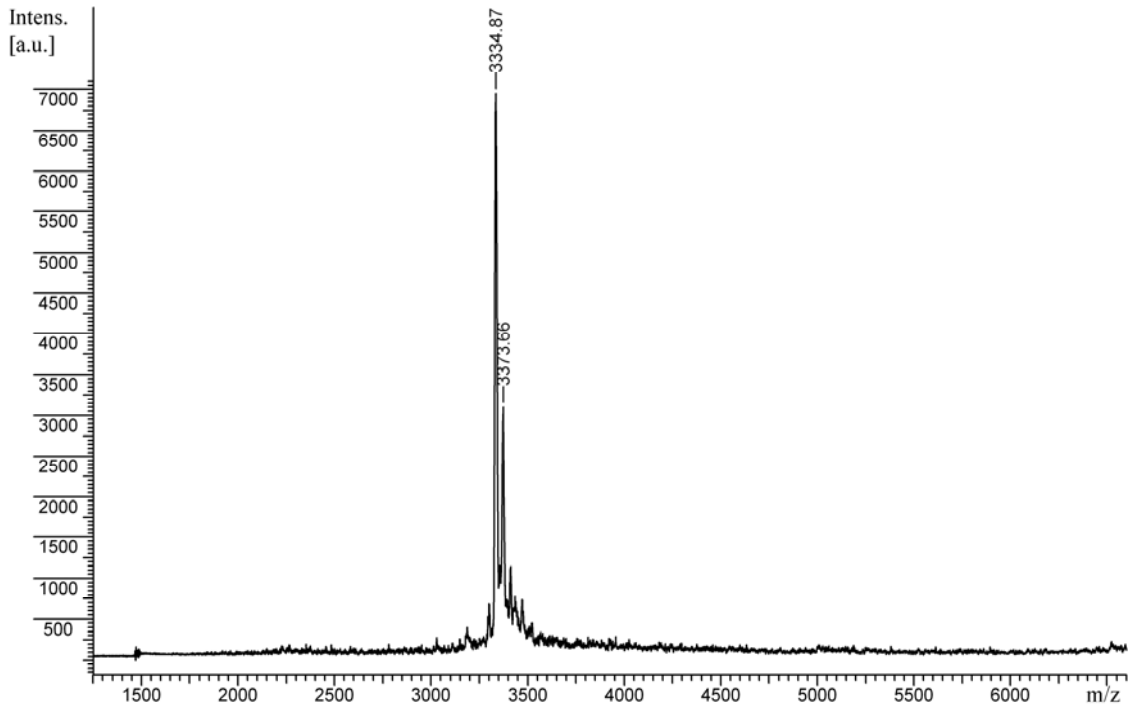
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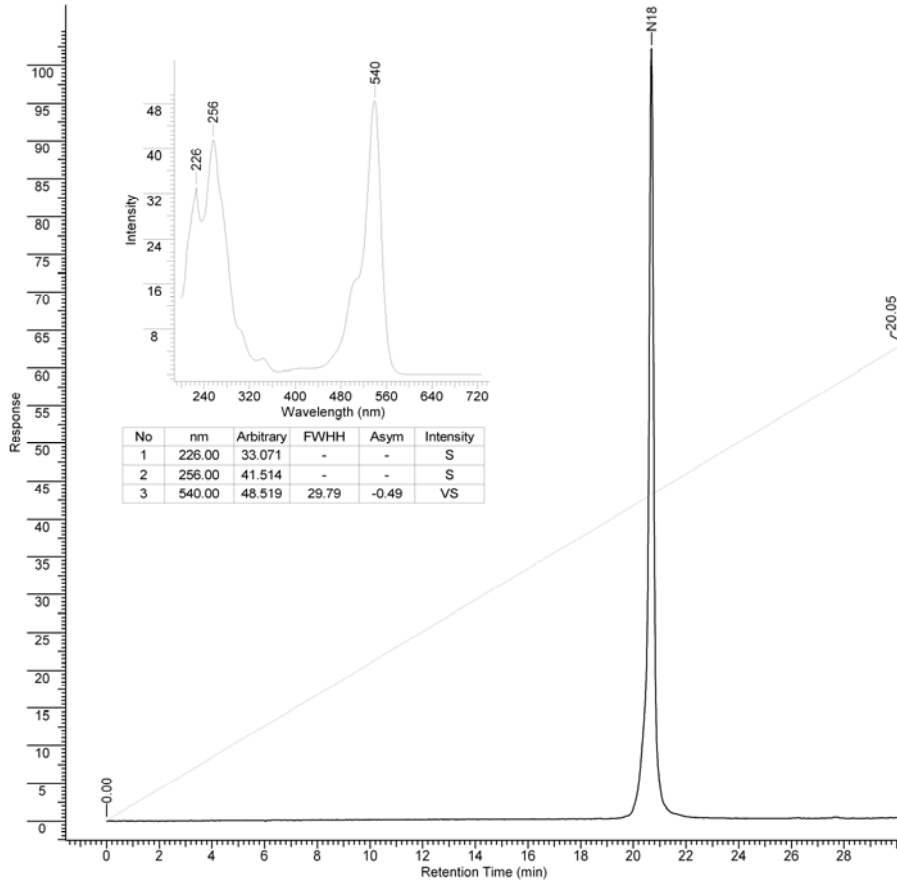
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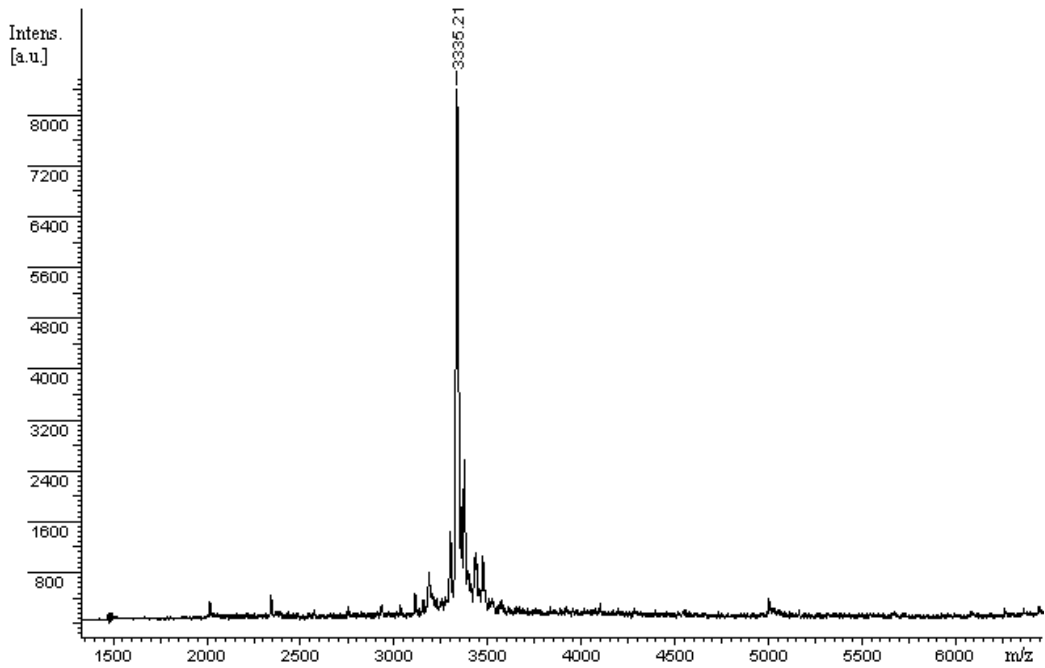
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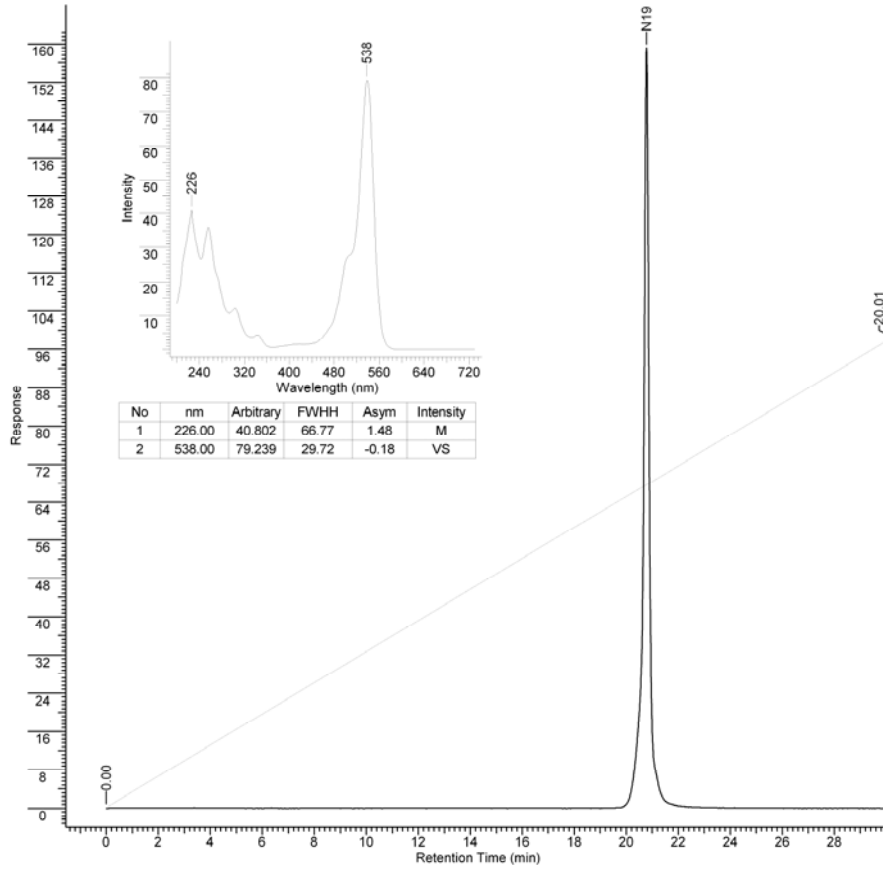
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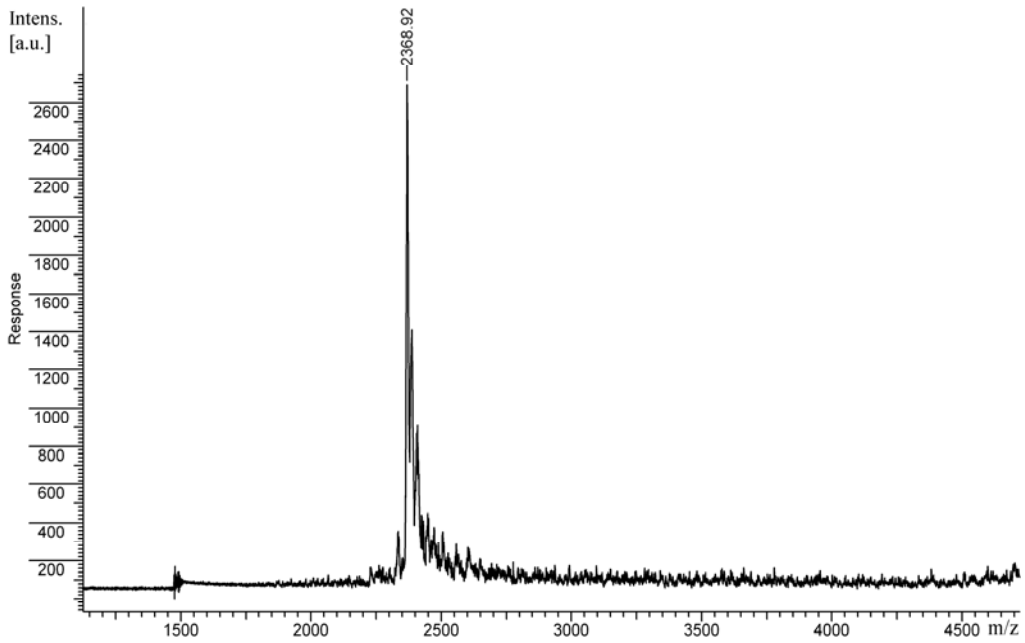
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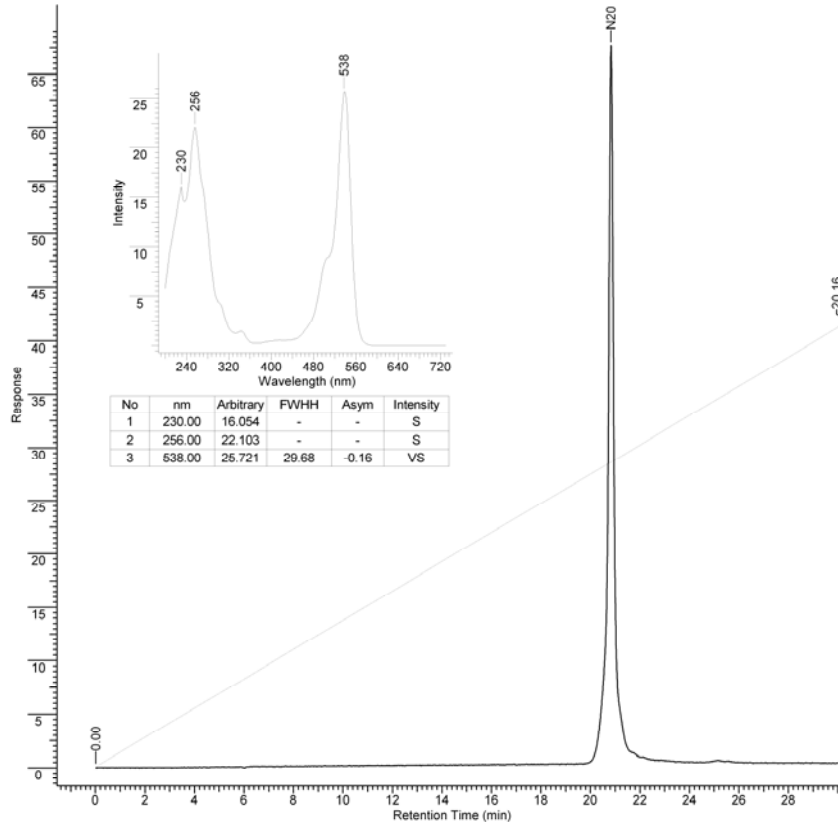
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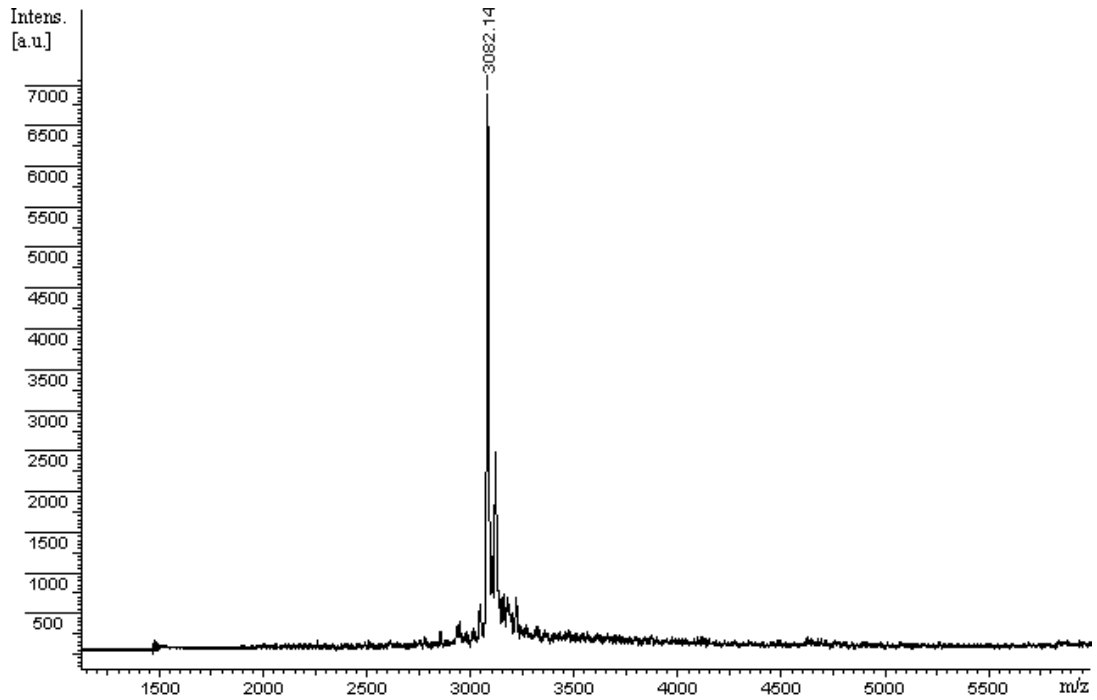
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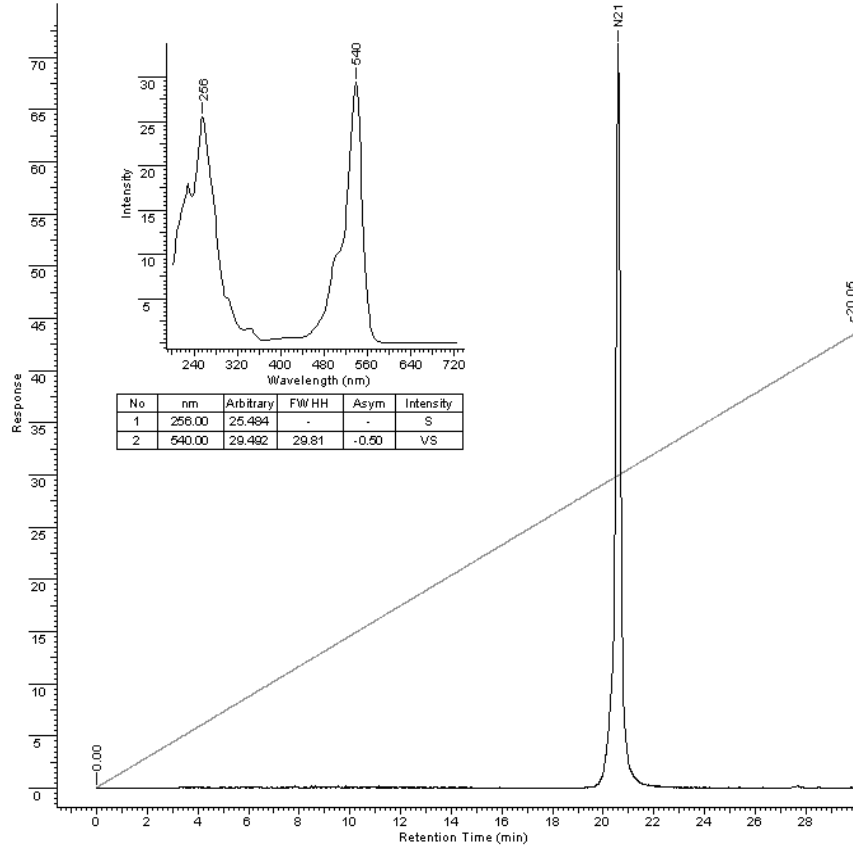
HEX-11D-PD1



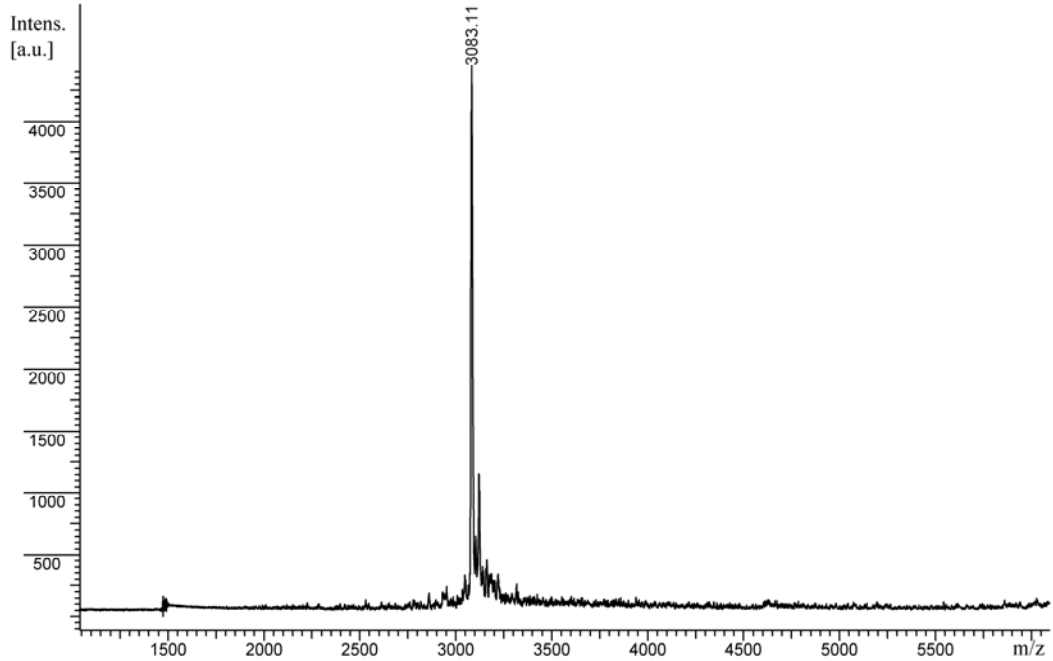
Name	tR	Peak Area (Y units)	Area Percent	Width	Asymmetry	Plate Number
N20	20.831	1156564.000	100.000	0.342	1.073	59188.613



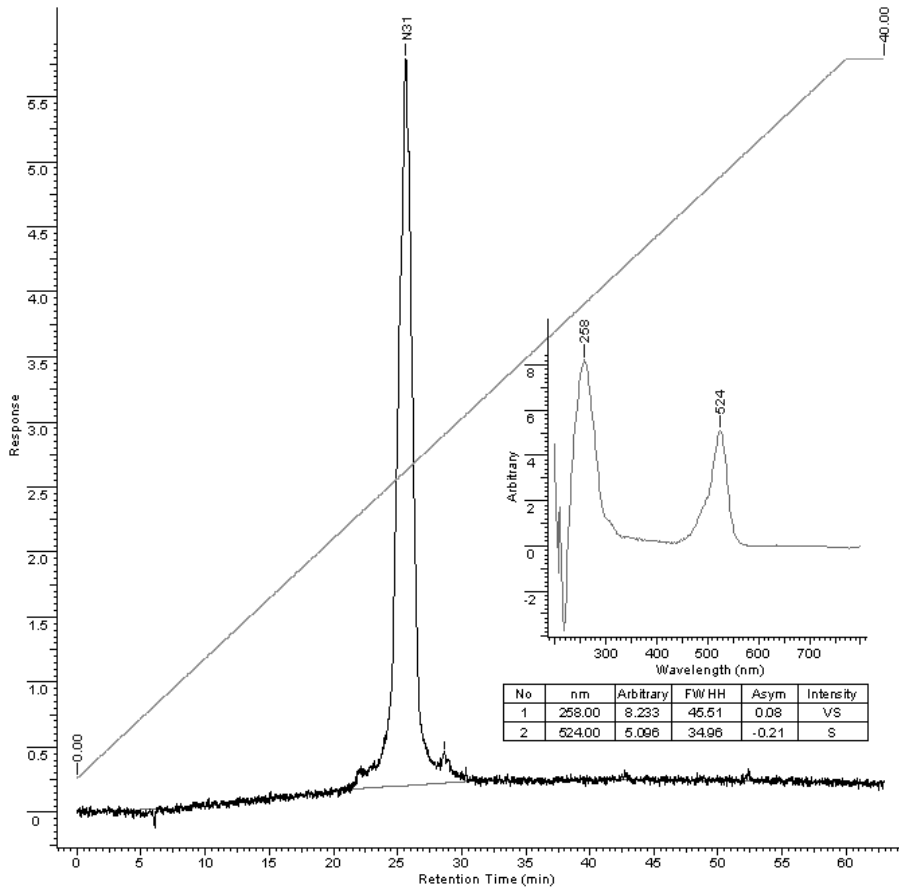
HEX-11D-PD2



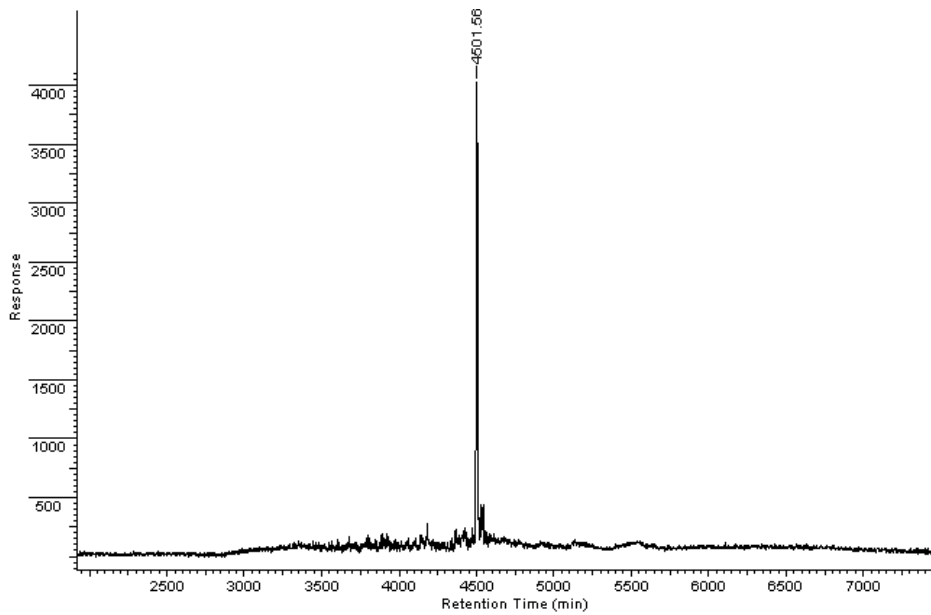
Name	tR	Peak	Area (Y units)	Area Percent	Width	Asymmetry	Plate Number
N21	20.593		1065941.500	100.000	0.322	1.037	66270.320



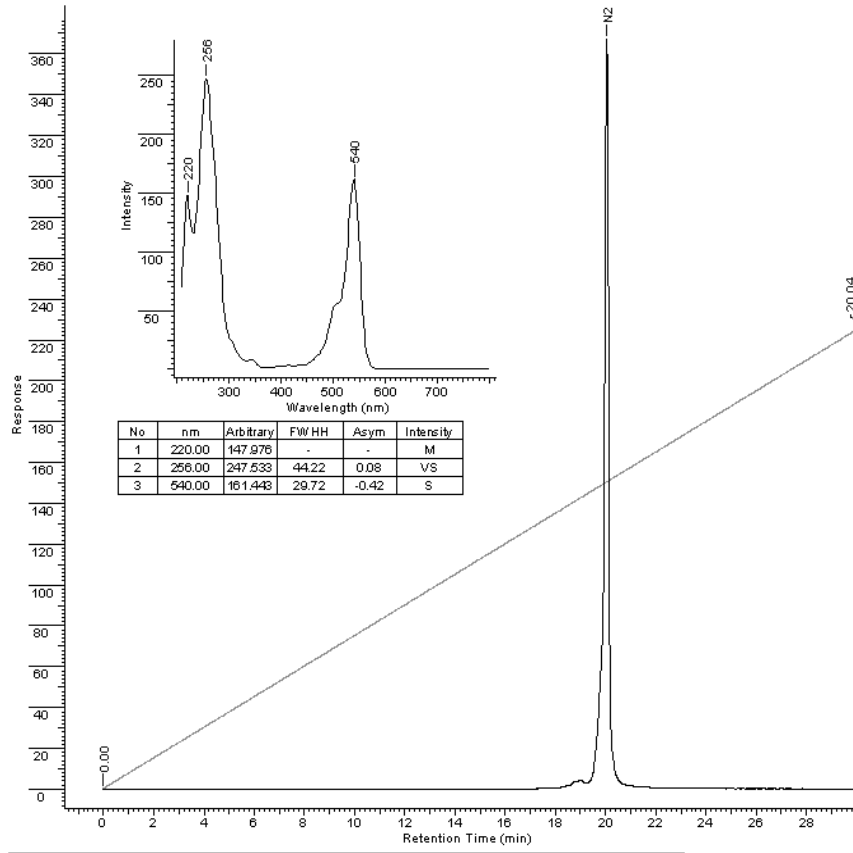
11M-E



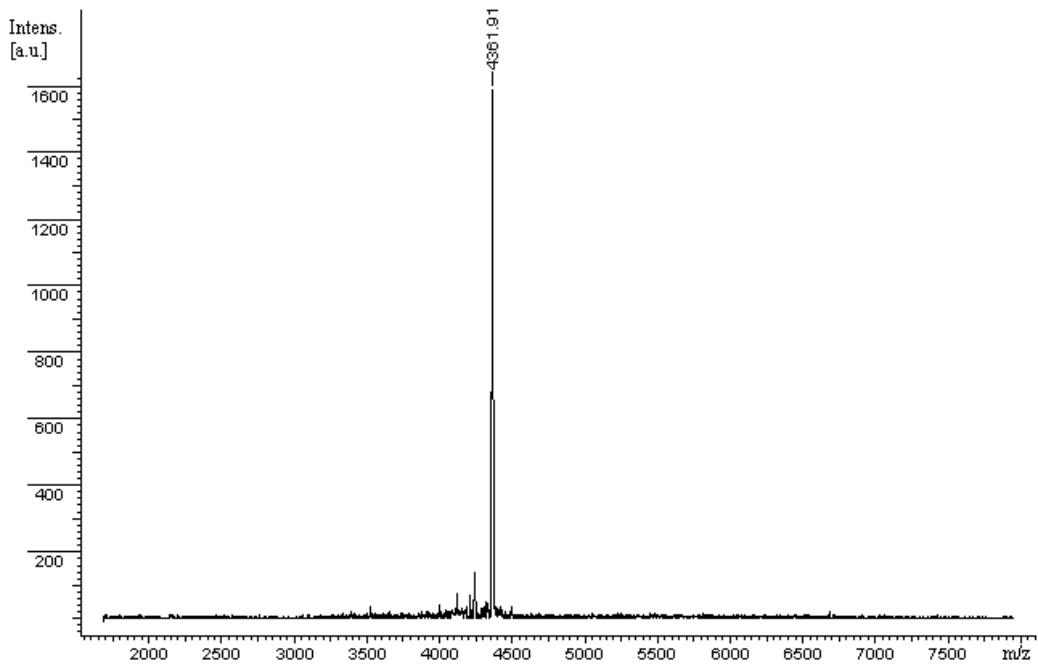
No.	Name	tR	Peak Area (Y units/mg)	Area Percent
1	N31	28.633	439455.000	98.671
2	unknown	28.633	5905.575	1.329



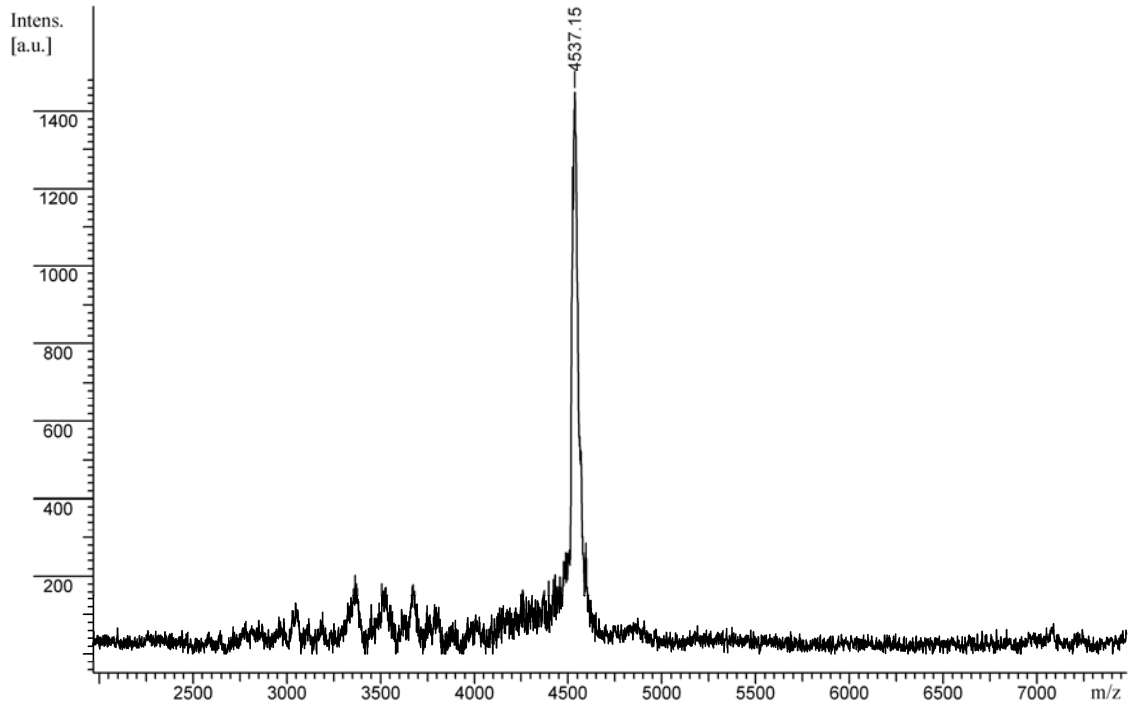
HEX-11M



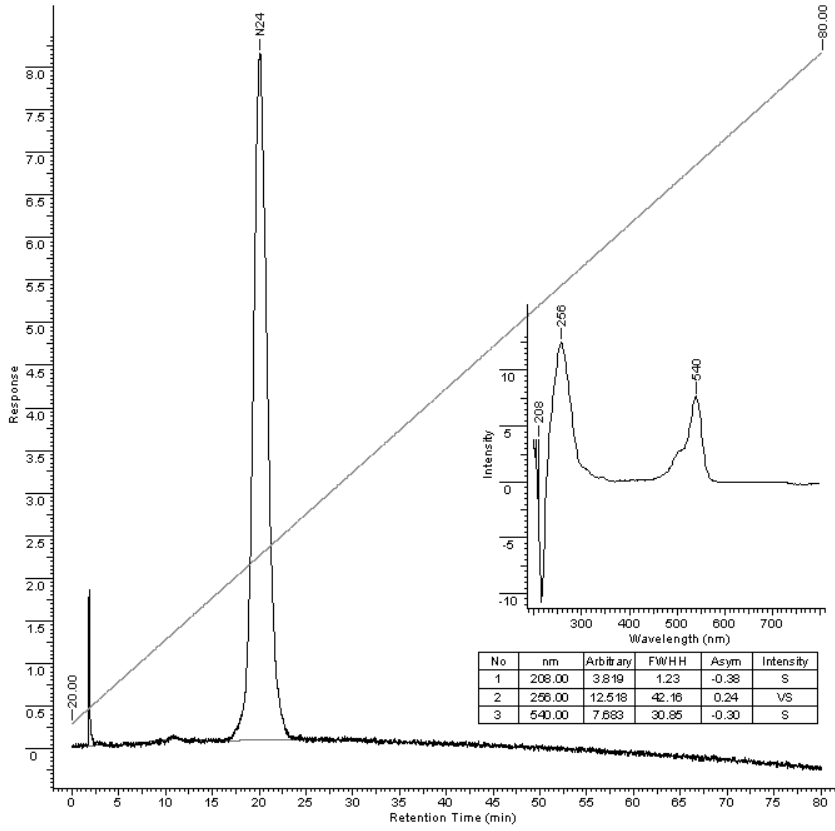
Name	tr	Peak Area (Y units)	Area Percent	Width	Asymmetry	Plate Number
N2	20.047	4743072.500	100.000	0.277	0.891	83963.758



HEX-11MS



HEX-11MS-Ole



Name	IR	Peak Area (Y units/ms)	Area Percent	Width	Asymmetry	Plate Number
N24	20.064	836683.625	100.000	2.378	1.173	1138.895

