

1 **Supporting Information**

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3 **Title: Cytosolic DNA-mediated, STING-dependent pro-inflammatory gene**
4 **induction necessitates canonical NF- κ B activation through TBK1**

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8 **Supporting Figure legends**

9 **Fig. S1. STING ligands-mediated signaling response in MEFs.** (A) Primary MEFs (1
10 $\times 10^5$ cells/well) derived from wild-type (STING^{+/+}) or STING-deficient mice
11 (STING^{-/-}) were stimulated with 10 μ g /ml of canonical 3'-5' cyclic-GMP-AMP
12 (cGAMP) for the times indicated. The expression level of STING, IRF3 phosphorylated
13 at Ser³⁹⁶ (p-IRF3), NF κ B p65 phosphorylated at Ser⁵³⁶ (p-NF- κ Bp65), NF- κ Bp65,
14 TBK1 phosphorylated at Ser¹⁷² (p-TBK1), and β -actin was determined by
15 immunoblotting. (B) Nuclear translocation of NF- κ Bp65 is detected by cellular
16 fractionation. STING^{+/+} and STING^{-/-} MEFs were stimulated with 10 μ g /ml of
17 dsDNA90 for 3 h, and then cells were separated into cytosol and nuclear fractions. Each
18 fraction was concentrated and subjected to immunoblotting with indicated antibodies.
19 (C) STING^{+/+} and STING^{-/-} MEFs were stimulated with 10 μ g /ml of dsDNA90, 5 μ g
20 /ml of Poly(dA:dT), 5 μ g /ml of Poly(I:C), or 200 μ M of DMXAA for 6 h, and then
21 total RNA was extracted from these cells, and the expression level of mRNA of
22 IFN β , Ccl5, IL-6 and CXCL10 was determined by qRT-PCR. Data were normalized to

23 the levels of GAPDH mRNA. Error bars indicate the standard deviations (SD).

24 **Fig. S2. HSV-1-mediated signaling response in MEFs.** (A) Primary MEFs (1×10^5
25 cells/well) derived from wild-type (STING^{+/+}) or STING-deficient mice (STING^{-/-}) were
26 stimulated with 5 $\mu\text{g}/\text{ml}$ of dsDNA representing the genomes of HSV-1 or purified
27 HSV-1 at an moi of 10 PFU/ml for the times indicated. The expression level of STING,
28 IRF3 phosphorylated at Ser³⁹⁶ (p-IRF3), NF κ B p65 phosphorylated at Ser⁵³⁶
29 (p-NF- κ Bp65), NF- κ Bp65, TBK1 phosphorylated at Ser¹⁷² (p-TBK1), TBK1, and
30 β -actin was determined by immunoblotting.

31 **Fig. S3. TBK1 does not involve in the dsDNA-mediated MAPKs activation.** (A)
32 TBK1^{+/+} and TBK1^{-/-} MEFs were stimulated with 10 $\mu\text{g}/\text{ml}$ of dsDNA90, or 5 $\mu\text{g}/\text{ml}$ of
33 Poly(I:C) for the times indicated. The expression level of ERK1/2 phosphorylated at
34 Thr²⁰²/Tyr²⁰⁴ (p-ERK1/2), ERK1/2, SAPK/JNK phosphorylated at Thr¹⁸³/Tyr¹⁸⁵
35 (p-SAPK/JNK), JNK1/2, p38 phosphorylated at Thr¹⁸⁰/Tyr¹⁸² (p-p38), p38, and c-Jun
36 phosphorylated at Ser⁶³ (p-cJun) was determined by immunoblotting. (B) 293T cells
37 were transfected with empty vector (EV) and Flag-tagged TBK1 (TBK1-FL) for 24 h,
38 and then cell extracts were subjected to immunoblotting with indicated antibodies.

39 **Fig. S4. IKKi/IKK ϵ and NIK does not involve in the STING-dependent,**
40 **dsDNA-mediated signaling response in MEFs.** RNAi in MEFs using the non-specific
41 (NS) and IKKi/IKK ϵ (A) or NIK (B) were stimulated with 5 $\mu\text{g}/\text{ml}$ of dsDNA90 for the
42 times indicated. The expression level of STING, IRF3 phosphorylated at Ser³⁹⁶
43 (p-IRF3), NF κ B p65 phosphorylated at Ser⁵³⁶ (p-NF- κ Bp65), NF- κ Bp65, TBK1
44 phosphorylated at Ser¹⁷² (p-TBK1), TBK1, NF- κ Bp52, NF- κ Bp100 phosphorylated at

45 Ser^{866/870} (p-NF-κBp100), and β-actin was determined by immunoblotting. (C) RNAi in
46 MEFs using the non-specific (NS) and IKKi/IKKε (upper panels) or NIK (lower panels)
47 were stimulated with 5 μg /ml of dsDNA90 for 6 h, and then total RNA was extracted
48 from these cells, and the expression level of mRNA of IL-6, CXCL10, and
49 IKKi/IKKε or NIK were determined by real-time PCR, respectively. Real-time PCR
50 data were normalized to the amount of GAPDH mRNA.





