

Supporting Information

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Table S1. Oligonucleotide primers for gene amplification analyses

Name	Sequence (5'-3')	Purpose
TransE2rcF	GGAGACTATTCCAAGATAACATTAACG	Genomic PCR
TransE2rcR	AGTATCAGTGCTCGGAGGACAG	
eGFP-F	GCTGGAATACAACACTACAACCTCGC	Genomic PCR
eGFP-R	CCAGCTGCACCGATCCATC	
Actin-F	AAGGCCAACCGTGAGAAGATGACT	Genomic PCR
Actin-R	GCTCGTTGCCAATGGTGATAC	
Aegyptin-F	CTATTCCAAGGTTTGTGATCATTTTC	Genomic PCR
Aegyptin-R	GATTGGTTTTTCGTTGGCTTGAGAC	
dsAegyptin-1F	AAGAAACGACCGATGATGCT	qPCR
dsAegyptin-1R	GCCAGCATCCTTATCTCCAG	qPCR
dsAegyptin-2F	TGTCTGGTAGGCATTGTGCT	
dsAegyptin-2R	AGACTCTCCGCTTGCATCAT	qPCR
Ribosomal S7R	TCGTGGACGCTTCTGCTTGTTG	
Ribosomal S7F	GGGACAAATCGGCCAGGCTATC	

Fig. S1. Primary sequences of the pMos_dsAegyptin and pMos_30KExGM transgenes.

[Fig. S1](#)

Fig. S2. Genomic integration and expression in vivo of the dsAegyptin transgene. (A) The insertion of the transgene into the *Aedes aegypti* genome was analyzed by genomic PCR. Amplification reactions were carried out on genomic DNA from mosquitoes by using different primer sets as described in *Materials and Methods*. PCR products were resolved on agarose gel, stained with ethidium bromide and visualized under UV light. Amplicons corresponding to eGFP and dsAegyptin demonstrate the transgene genome insertion. (B) Expression of the reporter gene eGFP was analyzed under fluorescent microscopy. Accumulation of eGFP is limited to the lateral distal lobes of transgenic mosquitoes demonstrating the functionality of the transgene. (C) No fluorescence in the salivary glands of the parental strain Higgs was detected.

[Fig. S2](#)