

Supporting Information

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Noncompetitive Inhibition of Indolethylamine *N*-Methyltransferase by *N,N*-Dimethyltryptamine (DMT) and *N,N*-Dimethylaminopropyltryptamine

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Figure Legend

Figure S1: A representative thin layer chromatography showing the end products of rabINMT methylation reactions. The INMT methylation of tryptamine shows two products corresponding to N-methyl (MMT) and N,N-dimethyltryptamine (DMT) as detected by autoradiography. Cold tryptamine, MMT, and DMT were added to each sample after the reaction was quenched with potassium borate and prior to extraction. Samples were spotted on silica plate and separated using 12 n-Butanol: 5 H₂O: 3 acetic acid solvent system. A control condition was performed in the absence of rabbit lung homogenate. The relative location of each compound on TLC plates was determined separately. Co-migration of cold MMT and DMT with the radioactive products was confirmed and shown by the dotted lines around UV-active spots. MMT, N-methyltryptamine; DMT, N,N-dimethyltryptamine; RLH, Rabbit lung homogenates. Rf values for DMT is 0.39; MMT, 0.48; and Tryptamine, 0.57.

Figure S2: Sequence alignment of human and rabbit INMT. Amino acids highlighted in red are identical between two species and those highlighted in yellow show sequence similarity. The blue box highlights the loop region identified by *in silico* docking of DMT and PDAT. The proposed allosteric N-terminal Helix-loop-Helix region is not within the active site of the enzyme as assessed by the position of SAH in the co crystal structure of hINMT (PDB code 2A14).

Figure S3: Binding mechanism of DMT for rabINMT. **A.** A repeat experiment of the Michaelis-Menten plot for DMT in the absence or presence of cold DMT (100 μ M). Concentrations of tryptamine varied from 100 μ M to 1000 μ M. **B.** Lineweaver-Burk

transformation of the data presented in A. Velocity (V) was calculated by determining the [¹⁴C]DMT formed over the time course of 60 min) and [S] represents the tryptamine concentrations. Inset shows a close up image of part B showing the intersections of the two lines. In the absence of inhibitor, the mean K_m (from two separate experiments) for DMT is $852.2 \pm 61.35 \mu\text{M}$ and $1618 \pm 275.5 \mu\text{M}$ in the presence of cold DMT. The K_m values were determined using the Michaelis-Menten analyses from Graphpad Prism and standard error of the mean was calculated from two separate experiments performed in duplicates.

Figure S4: Binding mechanism of PDAT for rabINMT. **A.** A repeat experiment for the Michaelis-Menten plot for DMT in the absence or presence of 100 μM of PDAT. Concentrations of tryptamine varied from 100 μM to 1000 μM . **B.** Lineweaver-Burk transformations of the data presented in A. Velocity (V) was calculated by determining the ¹⁴C-DMT formed over the time course of 60 min) and [S] represents the tryptamine concentrations. Inset shows a close up image of part B showing the intersections of the two lines. In the absence of inhibitor, the mean K_m (from two separate experiments) for DMT is $499.6 \pm 68.2 \mu\text{M}$ and $523.4 \pm 85.4 \mu\text{M}$ in the presence of 100 μM PDAT. The K_m values were determined using the Michaelis-Menten analyses from Graphpad Prism and standard error of the mean was calculated from two separate experiments performed in duplicates.

Figure S5: Schematic representation of the kinetic mechanisms of binding of DMT (A) and PDAT (B). **A.** The inhibition of INMT by DMT fits a mixed-noncompetitive mechanism model. **B.** The inhibition of INMT by PDAT follows a pure non-competitive model.

