### Dechassa et al., Supplementary information

#### **Supplementary Information**

Supplementary Figure S1: Scm3 mediated Cse4-H4 tetramer formation on DNA is independent of Scm3 DNA binding activity and Scm3 is not stably associated with tetrasome after assembly. A. The Nterminal region of Scm3 is critical for DNA binding activity. Scm3 or Scm3<sub>63-189</sub> (1-4 µM) were incubated for 30 min at 25°C with 79 bp DNA (0.5  $\mu$ M) in buffer containing 20 mM Tris-HCl pH 7.5, 300 mM NaCl, 1mM EDTA, 20µg/ml BSA and 4% glycerol. The reactions were analyzed by 5% native PAGE and ethidium bromide staining. Lane 1: DNA only control; lane 2-4: wild type Scm3; lanes 5-7: Scm3 Scm3<sub>63-189</sub>. B. The DNA binding activity of Scm3 is not important for assembly of tetrasome. The experiment was conducted as in Figure 2. A preformed complex of Scm3-Cse4ΔN-H4 or Scm3 Scm3<sub>63-189</sub> -Cse4ΔN-H4, labeled on at H4 either Alexa-488 (green D), or Atto-647N (blue A); or a mixture of the differently labeled complexes (DA) as indicated, was incubated with a five molar excess of 79 bp DNA. Protein-DNA complexes were analyzed by 5% native PAGE followed by fluorescence scanning. Lanes 1: Scm3 Scm363-189 -Cse4ΔN-H4 (D); lanes 2: Scm3 Scm3<sub>63-189</sub> -Cse4ΔN-H4 (A); lane 3: Scm3 Scm3<sub>63-189</sub> -Cse4ΔN-H4 (D) and Scm3 Scm3<sub>63-189</sub> -Cse4 $\Delta$ N-H4 (A); lane 4: Scm3-Cse4 $\Delta$ N-H4 (D); lane 5: Scm3-Cse4 $\Delta$ N-H4 (A); lane 6: Scm3-Cse4AN-H4 (D) and Scm3-Cse4AN-H4 (A); lane 7: Scm3-Cse4AN-H4 (D) and Scm3-Cse4AN-H4 (A, Scm3 labeled with Atto-647N); lane 8: Scm3 (Atto-647N labeled). 1 µM protein and 5 µM 79 bp DNA were used in each reaction. Green, blue and red gel scans are for donor, acceptor and FRET, respectively. \*- Shows the acceptor label is on Scm3.

**Supplementary Figure S2. CENP-A and H4 are compatible to form a heterotypic tetramer. A.** Models for a Cse4-H3 four-helical bundle. The coordinates for Cse4 from the *Kluyveromyces lactis* (Cse4-H4)<sub>2</sub> tetramer structure (pdb 2YFW; (1)) were superimposed onto H3 from the yeast nucleosome structure (pdb 1ID3; (2)) using PYMOL. Only regions of H3 and Cse4 that form the four-helix bundle structure are shown. The residues that are critical for four-helical bundle are shown as sticks. **B.** Alignment of the region shown in (A) of H3 and CENP-A from various species highlights conservation of residues that are involved in four-helical bundle formation (H113, D123, L126 and L/I 130; indicated by black dots) among different organisms.

## Supplementary Figure S3. In vitro reconstitution of heterotypic tetrasomes and nucleosomes.

**A.** Cse4 $\Delta$ N forms a tetramer with His<sub>6</sub>.H3 and two copies of H4. Cse4 $\Delta$ N, His<sub>6</sub>.H3 and H4 were refolded and purified by gel filtration. Fractions 1-5 from the peak were analyzed by SDS-PAGE (inset).

**B.** Complexes containing Cse4 $\Delta$ N-His<sub>6</sub>.H3-H4<sup>2</sup> taken from gel filtration fraction # 3 (A) and control tetramers (His<sub>6</sub>.H3-H4)<sub>2</sub>, (H3-H4)<sub>2</sub> and (Cse4 $\Delta$ N -H4)<sub>2</sub> were reconstituted onto 147 bp 601 DNA by salt dilution to form tetrasomes (lanes 2, 5, 8 and 11). Tetrasomes were reconstituted by salt dilution using 1.8  $\mu$ M 147 bp DNA and 3.0-5.2  $\mu$ M tetramers (1.7-2.9: 1 tetramer to DNA ratio). Nucleosomes were reconstituted as in tetrasomes assembly except that H2A-H2B dimers (3.7-5.6  $\mu$ M) were also included in the reactions. Tet- refers to tetrasomes; Nuc- nucleosomes.

**C.** Tetrasome samples from (B) were purified by Ni-NTA affinity chromatography and analyzed by SDS-PAGE and Coomassie blue staining. IN- input, UB- Ni-NTA unbound, W3- third wash, W4- fourth/last wash, E- elution. Note that only a relatively weak band of Cse4 $\Delta$ N is expected (and observed) due to the enrichment of homotypic (His<sub>6</sub>.H3-H4)<sub>2</sub> tetrasome in addition to heterotypic Cse4 $\Delta$ N- His<sub>6</sub>.H3-H4<sub>2</sub> tetrasomes.

## Supplementary References:

- 1. Cho, U.S. and Harrison, S.C. (2011) Recognition of the centromere-specific histone Cse4 by the chaperone Scm3. *PNAS*, **108**, 9367-9371.
- 2. White, C.L., Suto, R.K. and Luger, K. (2001) Structure of the yeast nucleosome core particle reveals fundamental changes in internucleosome interactions. *Embo J*, **20**, 5207-5218.





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Supplementary Figure 2



# Supplementary Figure 3