

Supporting Information

Synthesis and Antimycobacterial Activity of 2,1'-Dihydropyridomycins

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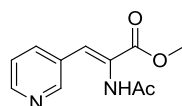
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1. General Methods

All manipulations were conducted under an argon atmosphere using flame-dried glassware and standard syringe/septa and Schlenk techniques. Absolute solvents were purchased from Fluka (absolute over molecular sieves). Commercial chemicals were used without further purification. Solvents for extractions, flash column chromatography (FC) and thin layer chromatography (TLC) were purchased as commercial grade and distilled prior to use. TLC was performed on Merck TLC aluminum sheets (silica gel 60 F254). Spots were visualized with UV light ($\lambda = 254$ nm) or through staining with $\text{Ce}_2(\text{SO}_4)_3$ /phosphomolybdic acid/ H_2SO_4 (CPS), vanillin/ H_2SO_4 or $\text{KMnO}_4/\text{K}_2\text{CO}_3$. Chromatographic purification of products by FC was performed using Fluka silica gel 60 for preparative column chromatography (particle size 40-63 μm).

NMR spectra were recorded on a Bruker Avance 400 MHz NMR spectrometer at 300 K. Chemical shifts (δ) are reported in ppm and are either referenced to the solvent signal as an internal standard (chloroform δ 7.26 ppm for ^1H and δ 77.00 ppm for ^{13}C spectra; DMSO- d_6 δ 2.50 ppm for ^1H and δ 39.43 ppm for ^{13}C spectra). Data are reported as follows: s = singlet, d = doublet, t = triplet, q = quartet, quint = quintet, sext = sextet, m = multiplet, br = broad signal, J = coupling constant in Hz. All ^{13}C -NMR spectra were measured with complete proton decoupling. ^1H - and ^{13}C -signals were assigned using two-dimensional correlation experiments (COSY, HMQC, HMBC). IR spectra were recorded on a Jasco FT/IR-6200 instrument as thin film. Optical rotations were measured on a Jasco P-1020 polarimeter operating at the sodium D line ($\lambda = 589$ nm) and are reported as follows: $[\alpha]_D^T$, concentration (c in g/100 mL) and solvent. Melting points were obtained in open capillary tubes using a Büchi melting point apparatus B-540 and are uncorrected. Mass spectra were recorded by the MS service of the Laboratory of Organic Chemistry (LOC) of the ETH Zürich; HRMS (ESI) spectra were measured on a Bruker Daltonics maxis (UHR-TOF) and HRMS (EI) on a Waters Micromass AutoSpec Ultima instrument.

2. Synthesis of Building Block 5

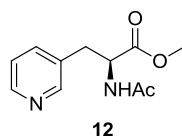


11

Ester 11: Pyridine-3-carbaldehyde (4.38 mL, 46.7 mmol, 1.00 eq.) was added to a mixture of *N*-acetylglycine (5.47 g, 46.7 mmol, 1.00 eq.) and NaOAc (4.21 g, 51.4 mmol, 1.10 eq.) followed by Ac_2O (24.8 mL, 243 mmol, 5.20 eq.) The dark brown mixture was stirred at 115 °C for 18 h. 10 mL MeOH were added (strongly exothermic!), in order to dilute the mixture, which was then poured into 40 mL MeOH containing 1.5 g NaOAc. The dark brown mixture was stirred at RT for 72 h. It was then partitioned between sat. aq. Na_2CO_3 (20 mL) and CHCl_3 (40 mL) and the aq. phase was extracted with

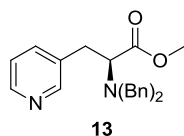
CHCl₃ (3 x 20 mL). The combined org. extracts were concentrated *in vacuo* and the crude product was purified by FC (CH₂Cl₂/MeOH 3% → 10%) to yield **11** (8.06 g, 78%) as yellow crystals which were recrystallized from hexane/EtOAc.

mp: 108-111 °C (Lit.: 110 °C, V. Busetti *et al.*, *J. Cryst. Spectrosc.* **1988**, *18*, 75-85)
¹H-NMR: (400 MHz, DMSO-d₆): δ 9.74 (s, 1H), 8.76 (s, 1H), 8.53 (dd, *J* = 4.8, 1.6 Hz, 1H), 8.02 (dt, *J* = 8.0, 1.7 Hz, 1H), 7.45 (dd, *J* = 8.0, 4.8 Hz, 1H), 7.18 (s, 1H), 3.73 (s, 3H), 2.01 (s, 3H).
¹³C-NMR: (100 MHz, CDCl₃): δ = 168.5, 165.5, 150.7, 149.8, 136.4, 130.4, 128.0, 125.5, 123.5, 53.1, 23.7.
IR: (neat, cm⁻¹): 3237, 2995, 2953, 1721, 1670, 1587, 1567, 1510, 1435, 1371, 1338, 1264, 1219, 1192, 1125, 1025, 985, 808, 764, 733, 705, 634, 609, 521.
HR-MS: (ESI): *m/z* calc. for C₁₁H₁₃N₂O₃ [M+H]⁺ 221.0921, found 221.0920.



Ester 12: Olefin **11** (1.42 g, 6.45 mmol, 1.00 eq.) was dissolved in 75 mL freshly degassed MeOH and HBF₄ (50% in H₂O, 2.22 mL, 9.67 mmol, 1.50 eq.) was added. The solution was transferred into an autoclave and [Rh(COD)(*R,R*-DIPAMP)]BF₄ (4.88 mg, 6.45 μmol, 0.001 eq.) was added. The autoclave was pressurized with H₂ and subsequently vented 5 x before application of the final pressure of 5 bar. The mixture was heated to 50 °C and stirred for 18 h. It was then concentrated *in vacuo*, neutralized with sat. aq. Na₂CO₃ (15 mL) and extracted with CHCl₃ (4 x 20 mL). The crude product was purified by FC (CH₂Cl₂/MeOH 5%) to deliver **12** (1.22 g, 85%) as a yellow solid. The *ee* was determined by chiral HPLC (Daicel Chemical Industries, CHIRALPAK AD-H 0.46x15 cm column, isocratic hexane/*i*PrOH 9:1, 1.0 mL/min, *t_R* major: 10.67 min, *t_R* minor: 15.96 min).

R_f: 0.25 (CH₂Cl₂/MeOH 5%)
***ee*:** 87% (determined by HPLC analysis)
mp: 98-101 °C (Lit.: 101-103 °C, C. Döbler *et al.*, *Tetrahedron: Asymmetry* **1996**, *7*, 117-125)
[α]_D²⁰: +100.3° (*c* 1.19, CHCl₃) (Lit.: +105.1, *c* 1.08, CHCl₃, *Tetrahedron: Asymmetry*, **1996**, *7*, 117-125)
¹H-NMR: (400 MHz, CDCl₃): δ 8.48 (dd, *J* = 4.82, 1.66 Hz, 1H), 8.34 (d, *J* = 1.96 Hz, 1H), 7.44 (dt, *J* = 7.82, 1.95 Hz, 1H), 7.22 (ddd, *J* = 7.81, 4.83, 0.63 Hz, 1H), 6.16 (d, *J* = 6.88 Hz, 1H), 4.90 (ddd, *J* = 7.54, 5.75, 1H), 3.74 (s, 3H), 3.18 (dd, *J* = 14.05, 5.84, 2H), 3.08 (dd, *J* = 14.05, 5.64, 2H), 1.99 (s, 3H).
¹³C-NMR: (100 MHz, CDCl₃): δ 171.8, 169.8, 150.6, 148.7, 136.8, 131.8, 123.6, 53.0, 52.7, 35.3, 23.2.
IR: (neat, cm⁻¹): 3267, 3038, 2954, 1742, 1657, 1541, 1481, 1427, 1373, 1282, 1213, 1176, 1132, 1029, 802, 753, 714, 633, 597.
HR-MS: (ESI): *m/z* calc. for C₁₁H₁₅N₂O₃ [M+H]⁺ 223.1077, found 223.1075.



Ester 13: To acetamide **12** (350 mg, 1.58 mmol, 1.00 eq.) dissolved in 8.0 mL MeOH was added SOCl₂ (741 μL, 9.45 mmol, 5.00 eq.) at 0 °C. The solution was refluxed at 80 °C for 18 h. The mixture was concentrated and dissolved in toluene. The solvent was removed *in vacuo* and the yellow solid was portioned between CHCl₃ (10 mL) and sat. aq. Na₂CO₃ (5 mL). The aq. phase was extracted with CHCl₃ (4 x 15 mL) and the combined org. extracts were concentrated *in vacuo* to yield the free amino ester (crude, 243 mg, 86%) as an orange oil. ¹H- and ¹³C-NMR spectra confirmed the complete transformation to the free amine **27**:

¹H-NMR: (400 MHz, CD₃OD): δ = 8.97 (s, 1H), 8.86 (d, *J* = 5.64 Hz, 1H), 8.68 (d, *J* = 8.08 Hz, 1H), 8.14 (dd, *J* = 8.02, 5.86 Hz, 1H), 4.59 (t, *J* = 7.04 Hz, 1H), 3.83 (s, 3H), 3.60 (dd, *J* = 14.69, 7.48 Hz, 2H), 3.51 (dd, *J* = 14.71, 6.58 Hz, 2H).

¹³C-NMR: (101 MHz, CD₃OD): δ = 169.5, 149.5, 143.6, 142.0, 137.5, 128.8, 54.0, 53.9, 33.8.

To a solution of the above amino ester (234 mg, 1.30 mmol, 1.00 eq.) in 5 mL MeOH and 0.5 mL AcOH, benzaldehyde (791 μL, 7.79 mmol, 6.00 eq.), NaCNBH₃ (163 mg, 2.60 mmol, 2.00 eq.) and molecular sieves (4 Å) were added at RT and the suspension was stirred for 24 h. The mixture was neutralized with sat. aq. NaHCO₃ (5 mL) and extracted with Et₂O (3 x 10 mL). The combined org. extracts were dried over MgSO₄ and concentrated *in vacuo*. The residue was purified by FC (hexane/EtOAc 4:1 → 7:3 → 0:1) to yield **13** (411 mg, 88%) as a colorless oil.

R_f: 0.21 (hexane/EtOAc 4:1)

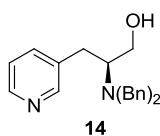
[α]_D²⁰: -88.16° (c 1.20, CHCl₃)

¹H-NMR: (400 MHz, CDCl₃): δ 8.49 (dd, *J* = 4.8, 1.7 Hz, 1H), 8.33 (d, *J* = 1.8 Hz, 1H), 7.31 – 7.18 (m, 11H), 7.14 (ddd, *J* = 7.8, 4.8, 0.8 Hz, 1H), 3.97 (d, *J* = 13.8 Hz, 2H), 3.79 (s, 3H), 3.66 (dd, *J* = 8.6, 6.8 Hz, 1H), 3.57 (d, *J* = 13.9 Hz, 2H), 3.05 (ddd, *J* = 22.9, 14.3, 7.7 Hz, 2H).

¹³C-NMR: (100 MHz, CDCl₃): δ 172.5, 150.9, 147.9, 139.0, 136.8, 133.9, 128.8, 128.4, 127.3, 123.2, 62.0, 54.7, 51.4, 33.1.

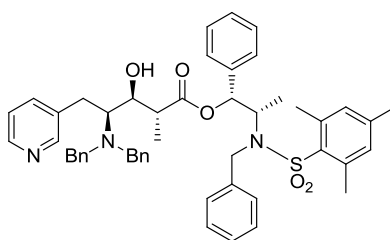
IR: (neat, cm⁻¹): 3028, 2950, 2843, 1730, 1576, 1494, 1479, 1453, 1425, 1374, 1361, 1291, 1214, 1195, 1162, 1128, 1075, 1028, 990, 787, 747, 715, 699.

HR-MS: (ESI): *m/z* calc. for C₂₃H₂₅N₂O₂ [M+H]⁺ 361.1911, found 361.1914.



Alcohol 14: Ester **13** (943 mg, 2.62 mmol, 1.00 eq.) was dissolved in 17 mL Et₂O and the solution was cooled to 0 °C. LAH (199 mg, 5.23 mmol, 2.00 eq.) was added and the suspension was stirred at 0 °C for 30 min and quenched with 2 mL H₂O, 2 mL 10% NaOH, 6 mL H₂O. The mixture was filtered, the filtrate was concentrated *in vacuo* and the residue purified by FC (hexane/EtOAc 2:3) to yield **14** (865 mg, 99%) as a colorless oil.

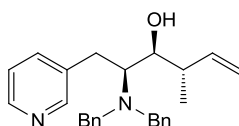
R_f: 0.26 (hexane/EtOAc 2:3)
[α]_D²⁰: + 26.33° (c 1.00, CHCl₃)
¹H-NMR: (400 MHz, CDCl₃): δ 8.38 (dd, *J* = 4.8, 1.6 Hz, 1H), 8.32 (d, *J* = 1.8 Hz, 1H), 7.35 – 7.15 (m, 11H), 7.12 (ddd, *J* = 7.8, 4.8, 0.6 Hz, 1H), 3.87 (d, *J* = 13.3 Hz, 2H), 3.54 – 3.41 (m, 3H), 3.34 – 3.22 (m, 1H), 3.07 – 2.93 (m, 2H), 2.90 – 2.79 (m, 1H), 2.46 – 2.35 (m, 1H).
¹³C-NMR: (100 MHz, CDCl₃): δ 150.4, 147.8, 138.8, 136.3, 134.8, 128.9, 128.6, 127.5, 123.4, 60.7, 60.3, 53.4, 29.2.
IR: (neat, cm⁻¹): 3304 (br.), 3061, 3028, 2930, 2834, 2804, 1578, 1494, 1480, 1453, 1425, 1363, 1129, 1044, 1028, 779, 746, 732, 714, 699.
HR-MS: (ESI): *m/z* calc. for C₂₂H₂₅N₂O [M+H]⁺ 333.1961, found 333.1952.



16

Ester 16: Alcohol **14** (40.0 mg, 332 μmol, 1.00 eq.) was dissolved in 1 mL CH₂Cl₂ and DMP (76.6 mg, 424 μmol, 1.50 eq.) was added at 0 °C. The suspension was stirred at 0 °C for 30 min and then diluted with Et₂O (1 mL). The reaction was quenched with 1 mL DMP workup solution (14 g sodium thiosulfate in 1 L 80% sat. aq. NaHCO₃) and the resulting mixture was stirred at 0 °C for 30 min. The aq. phase was extracted with Et₂O (3 x 5 mL). The combined org. extracts were washed with H₂O and brine (1 x 2 mL each), dried over MgSO₄ and concentrated *in vacuo* at 20 °C. The crude aldehyde **7** was dried at 10⁻³ mbar (RT) for 4 h. (1*R*,2*S*)-2-(*N*-benzyl-2,4,6-trimethylphenylsulfonamido)-1-phenylpropyl propionate (**15**) (75.1 mg, 157 μmol, 1.50 eq.) was dissolved in 1 mL CH₂Cl₂ and NEt₃ (54.4 μl, 392 μmol, 3.75 eq.) was added. The solution was cooled to -78 °C and dicyclohexylboron trifluoromethanesulfonate (112 mg, 345 μmol, 3.30 eq.) in 350 μl hexane was added dropwise during 15 min. The resulting solution was stirred at -78 °C for 3 h. Aldehyde **7** (34.5 mg, 104 μmol, 1.00 eq.) dissolved in 0.5 mL CH₂Cl₂ was added dropwise during 20 min and the solution was stirred at -78 °C for 3 h. The mixture was warmed very slowly to 0 °C and stirred for 1 h. The reaction was then quenched with 1 mL pH 7 buffer, and the mixture was diluted with 4.5 mL MeOH and stirred with 0.45 mL H₂O₂ (50%) at RT for 16 h. The org. solvents were removed *in vacuo* and the residue was taken up in CH₂Cl₂ (5 mL) and H₂O (5 mL). The aq. phase was extracted with CH₂Cl₂ (3 x 5 mL) and the combined org. extracts were dried over MgSO₄. The solvents were removed *in vacuo* and the residue was purified by FC (hexane/EtOAc 3:2) to yield **16** (71.1 mg, 73% over 2 steps) as a 5:1 mixture of isomers (**16** vs. all other isomers). The desired isomer **16** could be isolated from this mixture in pure form by FC with hexane/EtOAc 3:2 as eluent (50.7 mg, 52% over 2 steps). Analytical data are for pure isomer **16**.

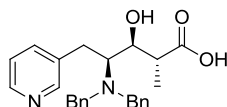
R_f: 0.24 (hexane/EtOAc 3:2)
[α]_D²⁰: + 20.22° (c 1.03, CHCl₃)
¹H-NMR: (400 MHz, CDCl₃): δ 8.48 (d, *J* = 1.7 Hz, 1H), 8.44 (dd, *J* = 4.8, 1.6 Hz, 1H), 7.50 (dt, *J* = 7.8, 1.9 Hz, 1H), 7.38 – 7.05 (m, 19H), 6.89 – 6.81 (m, 4H), 5.80 (d, *J* = 4.0 Hz, 1H), 4.70 (d, *J* = 16.5 Hz, 1H), 4.51 (d, *J* = 16.5 Hz, 1H), 4.19 (d, *J* = 13.1 Hz, 2H), 4.16 – 4.08 (m, 1H), 3.44 (d, *J* = 13.5 Hz, 2H), 3.39 – 3.31 (m, 1H), 3.28 (d, *J* = 3.3 Hz, 1H), 3.13 – 2.99 (m, 3H), 2.93 – 2.84 (m, 1H), 2.48 (s, 6H), 2.27 (s, 3H), 1.13 (d, *J* = 7.0 Hz, 3H), 0.45 (d, *J* = 7.1 Hz, 3H).
¹³C-NMR: (100 MHz, CDCl₃): δ 174.8, 150.6, 147.6, 142.5, 140.3, 139.7, 138.3, 138.2, 136.8, 135.7, 133.5, 132.1, 129.1, 128.4, 128.3, 127.9, 127.5, 127.2, 127.0, 125.9, 123.4, 78.4, 72.9, 59.1, 56.8, 55.6, 48.2, 42.6, 26.9, 22.9, 20.9, 13.1, 12.9.
IR: (neat, cm⁻¹): 3062, 3028, 2979, 2939, 1738, 1604, 1495, 1454, 1378, 1323, 1261, 1205, 1151, 1029, 1013, 931, 857, 751, 730, 699, 661, 568, 538.
HR-MS: (ESI): *m/z* calc. for C₅₀H₅₆N₃O₅S [M+H]⁺ 810.3935, found 810.3934.



18

Olefin 18: Alcohol **14** (408 mg, 1.23 mmol, 1.00 eq.) was dissolved in 9 mL anhydrous DCM and DMP (781 mg, 1.84 mmol, 1.50 eq.) was added at 0 °C. The suspension was stirred at 0 °C for 30 min and then diluted with Et₂O (10 mL). The reaction was quenched with 8 ml DMP workup mix and the solution was stirred for 30 min at 0 °C. The aq. phase was extracted with Et₂O (3 x 15 mL). The combined org. extracts were washed with H₂O and brine (1 x 2 mL each), dried over MgSO₄ and concentrated *in vacuo* at 20 °C. The crude aldehyde **7** was dried at 10⁻³ mbar (RT) for 4 h. A solution of this aldehyde (370 mg, 1.12 mmol, 1.00 eq.) in 10 ml THF was added to CrCl₂ (1.10 g, 8.96 mmol, 8.00 eq.) which had been dried at 200 °C under vacuum for 25 min. This was followed by addition of crotylbromide (576 μl, 5.60 mmol, 5.00 eq.) and the mixture was stirred at RT for 3 h (color change from green to brown). The reaction was quenched with sat. aq. NaHCO₃ (5 mL) and extracted with EtOAc (3 x 10 mL). The combined org. extracts were dried over MgSO₄ and concentrated *in vacuo*. The residue was purified by FC (hexane/EtOAc 1:4) to yield **18** as a yellow oil (308 mg, 67% over 2 steps, 15:1 ratio of isomers).

R_f: 0.26 (hexane/EtOAc 1:4)
¹H-NMR: (400 MHz, CDCl₃): δ 8.40 (dd, *J* = 4.7, 1.1 Hz, 1H), 8.34 (d, *J* = 1.8 Hz, 1H), 7.27 (d, *J* = 7.8 Hz, 1H), 7.22 – 7.00 (m, 11H), 5.52 – 5.40 (m, 1H), 5.04 (dd, *J* = 10.3, 1.4 Hz, 1H), 4.93 (d, *J* = 17.2 Hz, 1H), 3.78 (d, *J* = 14.1 Hz, 2H), 3.69 (d, *J* = 7.3 Hz, 1H), 3.54 (d, *J* = 14.1 Hz, 2H), 2.98 – 2.89 (m, 2H), 2.74 (dd, *J* = 17.9, 8.7 Hz, 1H), 2.23 – 2.10 (m, 1H), 0.78 (d, *J* = 6.8 Hz, 3H).
¹³C-NMR: (100 MHz, CDCl₃): δ 151.0, 147.1, 139.8, 139.8, 136.9, 136.8, 128.7, 128.2, 126.9, 123.0, 117.7, 73.1, 60.2, 54.4, 42.6, 29.2, 16.8.
HR-MS: (ESI): *m/z* calc. for C₂₆H₃₁N₂O [M+H]⁺ 378.2431, found 387.2427.



5

Acid 5:

A) From ester 16: To a solution of **16** (176 mg, 217 μmol , 1.00 eq.) in 5.2 mL MeOH/THF/H₂O 3:2:2 was added LiOH·H₂O (45.6 mg, 1.09 mmol, 5.00 eq.) at RT. The clear solution was stirred at RT for 24 h and diluted with Et₂O (7 mL). The aq. phase was acidified to pH 2 (aq. HCl 1 M) and the cleaved auxiliary was extracted, leaving the product in the aq. phase. The latter was adjusted to pH 7 (sat. aq. NaHCO₃) and the product was extracted with CHCl₃ (4 x 15 mL). The org. phase was dried over MgSO₄ and concentrated *in vacuo* to yield **5** as a yellow, viscous resin (84.1 mg, 96%).

B) From olefin 18: To a solution of AD-mix α (1.51 g) and **18** (136 mg, 351 μmol , 1.00 eq.) in 3.8 mL *t*-BuOH/H₂O 1:1 was added methanesulfonamide (201 mg, 2.11 mmol, 6.00 eq.). The yellow suspension was stirred at RT for 3 d. 0.9 mL H₂O and 1.3 g Na₂SO₃ were added and the mixture was stirred for 30 min. The mixture was extracted with CH₂Cl₂ (3 x 10 mL), the org. phase was dried over MgSO₄ and concentrated *in vacuo*. The residue was filtered through silica (CH₂Cl₂/MeOH 10%) to yield the crude diol (125 mg, 84%) as a white foam. This material (125 mg, 297 μmol , 1.00 eq.) was dissolved in 1.5 mL CH₂Cl₂ and NaIO₄ (254 mg, 1.19 mmol, 4.00 eq.) in 900 μL H₂O was added. The biphasic mixture was stirred at RT for 1 h. The aq. phase was extracted with Et₂O (3 x 5 mL) and concentrated *in vacuo*. The residue was dissolved in 2.88 mL *t*-BuOH/H₂O 4:1, 2,3-dimethylbutene (710 μL , 5.95 mmol, 20.0 eq.), NaClO₂ (134 mg, 1.49 mmol, 5.00 eq.) and NaH₂PO₄ (250 mg, 2.08 mmol, 7.00 eq., dissolved in H₂O) were added, and the clear biphasic mixture was stirred for 45 min at RT. 2 M HCl was added to pH 2 and the aq. phase was extracted with pentane (removal of scavenger). The aq. phase was adjusted to pH 7.5 (NaHCO₃) and extracted with CHCl₃ (4 x 10 mL). The org. phase was dried over MgSO₄ and concentrated to deliver **5** a yellow, viscous resin (64.3 mg, 45% from olefin **18**).

R_f: 0.08 (hexane/EtOAc 3:7)

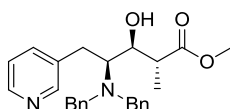
[α]_D²⁰: +35.6° (c 0.540, CHCl₃)

¹H-NMR: (400 MHz, CDCl₃): δ 8.48 (s, 1H), 8.40 (d, *J* = 3.3 Hz, 1H), 7.53 (d, *J* = 7.9 Hz, 1H), 7.29 – 7.04 (m, 11H), 4.05 (d, *J* = 12.8 Hz, 2H), 3.46 (t, *J* = 5.6 Hz, 1H), 3.39 (d, *J* = 13.4 Hz, 2H), 3.04 – 2.89 (m, 3H), 2.80 – 2.64 (m, 1H), 0.84 (d, *J* = 7.1 Hz, 3H).

¹³C-NMR: (100 MHz, CDCl₃): δ 177.7, 149.5, 146.3, 138.8, 137.8, 136.3, 129.2, 128.5, 127.4, 123.8, 73.0, 61.0, 55.1, 42.2, 28.6, 14.6.

IR: (neat, cm⁻¹): 3411, 3062, 3027, 2973, 2936, 2804, 1713, 1494, 1454, 1423, 1376, 1302, 1266, 1196, 1129, 1090, 1075, 1049, 1027, 1007, 983, 751, 700.

HR-MS: (ESI): *m/z* calc. for C₂₅H₂₉N₂O₃ [M+H]⁺ 405.2173, found 405.2173.



22

Ester 22: Acid **5** (7.90 mg, 19.5 μmol , 1.00 eq.) was dissolved in 0.3 mL MeOH and 0.5 mL toluene. TMSCHN₂ (2.0 M in Et₂O, 10.7 μl , 21.5 μmol , 1.10 eq.) was added at 0 °C (after the last drop, the pale yellow color persisted) and the mixture was stirred at 0 °C for 5 min. The reaction was quenched with 3 drops of acetic acid and the solvents were removed *in vacuo*. The residue was purified by FC (hexane/EtOAc 2:3) to yield **22** (6.60 mg, 81%) as a colorless oil.

R_f: 0.25 (hexane/EtOAc 2:3)

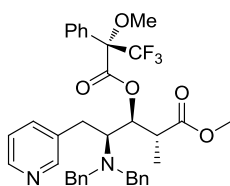
[α]_D²⁰: +17.7° (c 1.39, CHCl₃)

¹H-NMR: (400 MHz, CDCl₃): δ 8.57 (d, *J* = 1.8 Hz, 1H), 8.52 (dd, *J* = 4.8, 1.6 Hz, 1H), 7.59 (dt, *J* = 7.8, 1.9 Hz, 1H), 7.36 – 7.22 (m, 11H), 4.09 (d, *J* = 13.1 Hz, 2H), 3.55 – 3.49 (m, 1H), 3.46 (s, 3H), 3.42 (d, *J* = 13.3 Hz, 2H), 3.16 – 3.05 (m, 2H), 3.04 – 2.96 (m, 1H), 2.90 – 2.81 (m, 1H), 0.81 (d, *J* = 7.1 Hz, 3H).

¹³C-NMR: (100 MHz, CDCl₃): δ 175.4, 150.6, 147.7, 139.1, 136.8, 135.8, 129.2, 128.5, 127.3, 123.4, 73.3, 59.7, 54.9, 51.5, 41.9, 28.5, 14.2.

IR: (neat, cm⁻¹): 3259, 3027, 1733, 1453, 1424, 1262, 1195, 1166, 1129, 1092, 1075, 1028, 1013, 752, 732, 700.

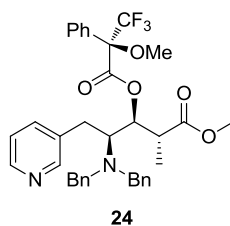
HR-MS: (ESI): *m/z* calc. for C₂₆H₃₀N₂NaO₃ [M+Na]⁺ 441.2149, found 441.2136.



23

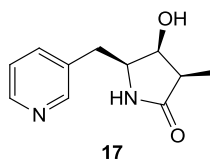
S-Mosher ester 23: To a solution of **22** (6.60 mg, 15.8 μmol , 1.00 eq.) in 0.2 mL CH₂Cl₂ were added pyridine (3.80 μl , 47.3 μmol , 3.00 eq.), DMAP (6.70 mg, 55.2 μmol , 3.50 eq.) and (*R*)-(-)-MTPA-Cl (8.90 μl , 47.3 μmol , 3.00 eq.) at RT. The solution was stirred at RT for 24 h and (*R*)-(-)-MTPA-Cl (17.8 μl , 94.6 μmol , 6.00 eq.) was added. After 3 h, sat. aq. NaHCO₃ (1 mL) was added and the aq. phase was extracted with CH₂Cl₂ (3 x 3 mL). The org. phase was dried over MgSO₄ and concentrated *in vacuo*. The yellow residue was purified by FC (hexane/EtOAc 1:1) to deliver **23** as a colorless film (1.2 mg, 12%).

¹H-NMR: (400 MHz, CDCl₃): δ 8.46 – 8.37 (m, 2H), 7.50 (dt, *J* = 7.9, 1.9 Hz, 1H), 7.44 (d, *J* = 7.3 Hz, 2H), 7.34 – 7.07 (m, 14H), 5.08 (dd, *J* = 10.2, 1.5 Hz, 1H), 3.84 (d, *J* = 13.3 Hz, 2H), 3.36 (s, 3H), 3.25 (dd, *J* = 10.1, 7.0 Hz, 1H), 3.19 (d, *J* = 13.3 Hz, 2H), 3.11 (s, 3H), 3.03 – 2.92 (m, 3H), 2.47 – 2.38 (m, 1H), 0.11 (d, *J* = 6.9 Hz, 3H).



R-Mosher ester 24: To a stirred solution of (2*R*)-3,3,3-trifluoro-2-methoxy-2-phenylpropanoic acid (6.00 mg, 25.6 μ mol, 2.10 eq.), Et₃N (3.73 μ l, 26.8 μ mol, 2.20 eq.), and DMAP (3.30 mg, 26.8 μ mol, 2.20 eq.) in 50 μ l toluene were added 2,4,6-trichlorobenzoyl chloride (4.00 μ l, 25.6 μ mol, 2.10 eq.) and a solution of **22** (5.10 mg, 12.2 μ mol, 1.00 eq.) dissolved in 100 μ l toluene at RT. The white slurry was stirred at RT for 6 h and was heated to 45 °C for 18 h. The yellow mixture was concentrated *in vacuo* and purified by FC (hexane/EtOAc 1:1) to deliver **24** as a colorless film (0.8 mg, 10%).

¹H-NMR: (400 MHz, CDCl₃): δ 8.48 (dd, *J* = 4.8, 1.6 Hz, 1H), 8.35 (t, *J* = 2.8 Hz, 1H), 7.62 (d, *J* = 8.1 Hz, 2H), 7.48 – 7.14 (m, 15H), 5.03 (dd, *J* = 9.9, 2.1 Hz, 1H), 3.75 (d, *J* = 13.5 Hz, 2H), 3.54 (s, 3H), 3.51 (s, 3H), 3.28 – 3.19 (m, 1H), 3.12 (d, *J* = 13.4 Hz, 2H), 2.96 (ddd, *J* = 9.6, 4.0, 2.0 Hz, 1H), 2.80 (dd, *J* = 13.9, 3.7 Hz, 1H), 2.16 (dd, *J* = 14.0, 9.7 Hz, 1H), 0.19 (d, *J* = 6.9 Hz, 3H).



Lactam 17: Ester **22** (15.0 mg, 358 μ mol, 1.00 eq.) was dissolved in 1 mL MeOH and Pd on charcoal (10%, 4.00 mg) was added under Ar. The atmosphere was exchanged with H₂ (1 bar) and the mixture was stirred at RT for 16 h. The suspension was filtered over celite, washed with MeOH and concentrated to yield **17** as a white solid (5.7 mg, 67%). For X-ray crystallography, **17** was crystallized from MeOH at RT by slow evaporation of the solvent.

R_f: 0.15 (CH₂Cl₂/MeOH 5%)

[α]_D²⁰: +2.01° (c 0.570, MeOH)

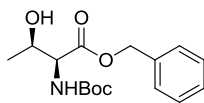
¹H-NMR: (400 MHz, MeOD): δ 8.51 (d, *J* = 1.7 Hz, 1H), 8.40 (dd, *J* = 4.9, 1.5 Hz, 1H), 7.83 (dt, *J* = 7.7, 1.7 Hz, 1H), 7.39 (dd, *J* = 7.7, 4.9 Hz, 1H), 4.08 – 4.04 (m, 1H), 3.87 (ddd, *J* = 8.9, 6.1, 4.0 Hz, 1H), 3.09 (dd, *J* = 13.6, 8.8 Hz, 1H), 2.91 (dd, *J* = 13.6, 6.1 Hz, 1H), 2.54 (qd, *J* = 7.3, 5.1 Hz, 1H), 1.13 (d, *J* = 7.3 Hz, 3H).

¹³C-NMR: (100 MHz, DMSO-d₆): δ 177.2, 150.3, 147.3, 136.7, 134.0, 123.3, 69.5, 58.3, 42.5, 31.7, 8.4.

IR: (neat, cm⁻¹): 3246, 2934, 1679, 1579, 1426, 1335, 1255, 1137, 1027, 994, 714, 638, 404.

HR-MS: (ESI): *m/z* calc. for C₁₁H₁₅N₂O₂ [M+H]⁺ 207.1128, found 207.1132.

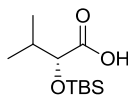
3. Synthesis of Building Blocks 6 and *epi*-6



9

Protected L-Thr 9: L-Thr (2.55 g, 21.4 mmol, 1.00 eq.) was dissolved in 75 mL 50% THF/H₂O. Na₂CO₃ (4.78 g, 45.1 mmol, 2.10 eq.) in 20 mL H₂O was added and the mixture was stirred at RT for 10 min. Boc₂O (5.92 mL, 25.8 mmol, 1.20 eq.) was added and the turbid mixture was stirred at RT for 14 h. It was then diluted with 15 mL H₂O and the pH was adjusted to 4 (aq. HCl 1 M). The aq. phase was extracted with EtOAc (3 x 15 mL), the pH was lowered to 3 (aq. HCl 1 M), NaCl was added to saturation and the aq. phase was again extracted with EtOAc (4 x 20 mL). The combined org. extracts were dried over MgSO₄ and concentrated *in vacuo* to yield L-Boc-Thr (3.79 g, 81% crude) as a colorless foam. The protected amino acid (3.79 g, 17.3 mmol, 1.00 eq.) and benzyl bromide (1.63 mL, 18.9 mmol, 1.05 eq.) were dissolved in 100 mL DMF at 0 °C. Cs₂CO₃ (2.93 g, 8.99 mmol, 0.52 eq.) was added and the suspension was stirred at RT for 20 h. H₂O (15 mL) was added and the mixture was extracted with EtOAc (3 x 20 mL). The combined org. extracts were washed with H₂O (1 x 10 mL) and brine (2 x 10 mL), dried over MgSO₄ and concentrated *in vacuo*. The resulting oil was purified by FC (hexane/EtOAc 9:1 → 3:2) to deliver **9** as a colorless oil (4.59 g, 86%).

R_f: 0.31 (hexane/EtOAc 9:1)
[α]_D²⁰: -14.45° (c 1.05, CHCl₃) (Lit.: -19.6°, C. W. Mosher and L. Goodman, *J. Org. Chem.* **1972**, 37, 2928-2933)
¹H-NMR: (400 MHz, CDCl₃): δ 7.37-7.34 (m, 5H), 5.33 (br s, 1H, NH), 5.20 (q, *J* = 11.3 Hz, 2H), 4.35-4.27 (m, 2H), 2.07 (br s, 1H, OH), 1.44 (s, 9H), 1.23 (d, *J* = 6.32 Hz, 3H).
¹³C-NMR: (100 MHz, CDCl₃): δ 171.3, 156.1, 128.6, 128.6, 128.4, 128.2, 80.1, 68.2, 67.2, 58.8, 28.3, 19.9.
IR: (neat, cm⁻¹): 3437, 2978, 2934, 1743, 1715, 1692, 1500, 1456, 1367, 1253, 1160, 112, 1066, 1000, 880, 752, 736, 698.
HR-MS: (ESI): *m/z* calc. for C₁₆H₂₃NNaO₅ [M+Na]⁺ 332.1468, found 332.1471

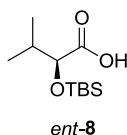


8

(R)-TBS-protected alcohol 8: The compound was prepared according to I. Gonzáles *et al.*, *J. Chem. Soc., Perkin Trans. 1*, **1996**, 1427-1433 while the analytics are compared to: M. Kusakabe *et al.*, *J. Org. Chem.* **1989**, 54, 2085-2091. (*R*)-2-Hydroxy-3-methylbutyric acid (305 mg, 2.58 mmol, 1.00 eq.), TBS-Cl (934 mg, 6.20 mmol, 2.40 eq.) and imidazole (1.07 g, 12.4 mmol, 4.80 eq) were dissolved in 3.5 mL DMF at RT and the mixture was stirred for 24 h. It was then diluted with EtOAc (65 mL), washed with

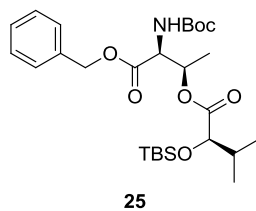
sat. aq. citric acid, sat. aq. NaHCO₃ and brine (3 x 10 mL each), dried over MgSO₄ and concentrated. The resulting oil was dissolved in 22 mL MeOH and the solution was cooled to 0 °C. 650 mg K₂CO₃ in 8 mL H₂O were added and the mixture was stirred at RT for 2.5 h. The solution was adjusted to pH 4 (aq. HCl 1 M) and the aq. phase was extracted with EtOAc (3 x 10 mL). The combined org. extracts were dried over MgSO₄ and concentrated *in vacuo*. The colorless oil was purified by FC (hexane/EtOAc 6:1 → 4:1) to deliver **8** as a colorless oil (339 mg, 57% over 2 steps).

R_f: 0.25 (10% MeOH in CH₂Cl₂)
[α]_D²⁰: +18.31° (c 0.942, CH₂Cl₂)
¹H-NMR: (400 MHz, CDCl₃) δ 4.05 (d, *J* = 4.0 Hz, 1H), 2.16 – 2.01 (m, 1H), 0.97 (d, *J* = 6.9 Hz, 3H), 0.95 – 0.91 (m, 12H), 0.09 (s, 6H).
¹³C-NMR: (101 MHz, CDCl₃) δ 176.9, 76.7, 32.8, 25.7, 18.8, 16.7, -5.2.
HR-MS: (ESI): *m/z* calc. for C₁₁H₂₃O₃Si [M-H]⁻ 231.1422, found 231.1424.



(S)-TBS-protected alcohol *ent*-8: The compound was prepared according to I. González *et al.*, *J. Chem. Soc., Perkin Trans. 1*, **1996**, 1427-1433. (*S*)-2-Hydroxy-3-methylbutyric acid (1.04 g, 8.80 mmol, 1.00 eq.), TBS-Cl (3.19 g, 21.1 mmol, 2.40 eq.) and imidazole (3.64 g, 42.3 mmol, 4.80 eq) were dissolved in 11 mL DMF at RT and the solution was stirred for 24 h. The mixture was diluted with EtOAc (200 mL), washed with sat. aq. citric acid, sat. aq. NaHCO₃ and brine (3 x 40 mL each), dried over MgSO₄ and concentrated *in vacuo*. The resulting oil was dissolved in 70 mL MeOH and the solution was cooled to 0 °C. 2 g K₂CO₃ in 24 mL H₂O were added and the mixture was stirred at RT for 2.5 h. The pH of the solution was adjusted to 4 (aq. HCl 1 M) and the aq. phase was extracted with EtOAc (3 x 20 mL). The combined org. extracts dried over MgSO₄ and concentrated *in vacuo*. The colorless oil was purified by FC (hexane/EtOAc 6:1 → 4:1) to deliver *ent*-**8** as a colorless oil (1.54 g, 75% over 2 steps).

R_f: 0.28 (10% MeOH in CH₂Cl₂)
[α]_D²⁰: -16.45° (c 0.811, CH₂Cl₂)
¹H-NMR: (400 MHz, CDCl₃): δ 10.09 (br. s, 1H), 4.06 (d, *J* = 4.0 Hz, 1H), 2.14-2.04 (m, 1H), 0.98 (d, *J* = 6.2 Hz, 3H), 0.94-0.93 (m, 12H, overlapping signals), 0.09 (s, 6H).
¹³C-NMR: (100 MHz, CDCl₃): δ 176.8, 76.7, 32.8, 25.7, 18.7, 18.2, 16.7, -5.2.
HR-MS: (ESI): *m/z* calc. for C₁₁H₂₃O₃Si [M-H]⁻ 231.1422, found 231.1426.



(R)-TBS-ether 25: To a stirred solution of **8** (250 mg, 1.08 mmol, 1.00 eq.) and Et₃N (449 μl, 3.23 mmol, 3.00 eq.) in 10 mL toluene was added 2,4,6-trichlorobenzoyl chloride (210 μl, 1.35 mmol, 1.25 eq.). As the mixture became turbid (white precipitate) a solution of **9** (350 mg, 1.13 mmol, 1.05 eq.) in 5 mL toluene and DMAP (263 mg, 2.15 mmol, 2.00 eq.) was added at RT. The yellow slurry was stirred at RT for 18 h. Sat. aq. NaHCO₃ (3 mL) was added and the aq. phase was extracted with EtOAc (3 x 10 mL). The combined org. extracts were dried over MgSO₄ and concentrated *in vacuo*. The yellow residue was purified by FC (hexane/EtOAc 9.5:1) to deliver **25** as a colorless oil (411 mg, 73%).

R_f: 0.36, (hexane/EtOAc 9.5:1)

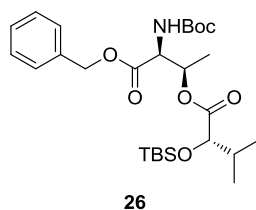
[α]_D²⁰: +44.4° (c 1.41, CHCl₃)

¹H-NMR: (400 MHz, CDCl₃) δ 7.39 – 7.29 (m, 5H), 5.48 (qd, *J* = 6.3, 2.5 Hz, 1H), 5.23 – 5.13 (m, 2H), 5.04 (d, *J* = 12.2 Hz, 1H), 4.46 (dd, *J* = 9.7, 2.4 Hz, 1H), 3.93 (d, *J* = 4.1 Hz, 1H), 2.00 – 1.89 (m, 1H), 1.46 (s, 9H), 1.30 (d, *J* = 6.4 Hz, 3H), 0.92 (s, 12H), 0.82 (d, *J* = 6.8 Hz, 3H), 0.03 (d, *J* = 6.7 Hz, 6H).

¹³C-NMR: (100 MHz, CDCl₃) δ 172.2, 170.0, 155.9, 134.9, 128.6, 128.5, 128.4, 80.3, 76.5, 70.9, 67.6, 57.3, 32.7, 28.3, 25.7, 19.2, 18.3, 17.1, 16.4, -4.9, -5.4.

IR: (neat, cm⁻¹): 3027, 2934, 2805, 1715, 1496, 1455, 1423, 1302, 1266, 1208, 1129, 1075, 1048, 1027, 981, 751, 700, 500, 471, 435.

HR-MS: (ESI): *m/z* calc. for C₂₇H₄₅NNaO₇Si [M+Na]⁺ 546.2858, found 546.2857.



(S)-TBS-ether 26: To a stirred solution of *ent*-**8** (70.5 mg, 303 μmol, 1.00 eq.), Et₃N (169 μl, 1.21 mmol, 4.00 eq.) and DMAP (74.1 mg, 607 μmol, 2.00 eq.) in 2 mL toluene was added 2,4,6-trichlorobenzoyl chloride (71.2 μl, 455 μmol, 1.50 eq.). As the mixture became turbid (white precipitate) a solution of **9** (98.7 mg, 319 μmol, 1.05 eq.) dissolved in 2 mL toluene was added at RT. The yellow slurry was stirred at RT for 18 h. Sat. aq. NaHCO₃ (1 mL) was added and the aq. phase was extracted with EtOAc (3 x 5 mL). The combined org. extracts were dried over MgSO₄ and concentrated *in vacuo*. The yellow residue was purified by FC (hexane/EtOAc 9.5:1) to deliver **26** as a colorless oil (106 mg, 67%).

R_f: 0.41 (hexane/EtOAc 9.5:1)

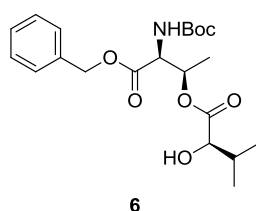
[α]_D²⁰: +1.43 (c 1.36, CHCl₃)

¹H-NMR: (400 MHz, CDCl₃): δ 7.42 – 7.28 (m, 5H), 5.54 – 5.44 (m, 1H), 5.20 (d, *J* = 9.8 Hz, 1H), 5.16 (d, *J* = 12.2 Hz, 1H), 5.05 (d, *J* = 12.2 Hz, 1H), 4.47 (dd, *J* = 9.8, 1.7 Hz, 1H), 3.94 (d, *J* = 4.2 Hz, 1H), 2.03 – 1.92 (m, 1H), 1.45 (s, 9H), 1.30 (d, 3H), 0.92 – 0.90 (m, 12H, overlapping signals), 0.83 (d, *J* = 6.8 Hz, 3H), 0.03 (d, *J* = 6.8 Hz, 6H).

¹³C-NMR: (101 MHz, CDCl₃): δ 172.2, 169.9, 155.9, 135.0, 128.6, 128.5, 128.4, 80.2, 76.6, 70.9, 67.6, 57.3, 32.8, 28.3, 25.7, 19.1, 18.2, 17.0, 16.5, -4.9, -5.4.

IR: (neat, cm⁻¹): 2959, 2931, 2858, 1751, 1722, 1500, 1457, 1386, 1367, 1314, 1251, 1163, 1143, 1112, 1083, 1066, 980, 861, 835, 778, 751, 678.

HR-MS: (ESI): *m/z* calc. for C₂₇H₄₆NO₇Si [M+H]⁺ 524.3038, found 524.3043.



Alcohol 6: To a solution of **25** (378 mg, 722 μmol, 1.00 eq.) in 10 mL THF was added HF•py (30%, 3.1 mL in 2 portions) at 0 °C. The solution was stirred at RT for 16 h. The reaction was quenched with sat. aq. NaHCO₃ (50 mL) and the solution extracted with EtOAc (3 x 15 mL). The combined org. extracts were dried over MgSO₄ and concentrated *in vacuo*. The residue was purified by FC (hexane/EtOAc 4:1) to yield **6** as a colorless oil (254 mg, 86%).

R_f: 0.23 (hexane/EtOAc 4:1)

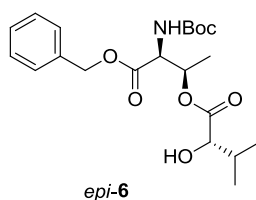
[α]_D²⁰: +29.2° (c 0.765, CHCl₃)

¹H-NMR: (400 MHz, CDCl₃): δ 7.40 – 7.30 (m, 5H), 5.49 (qd, *J* = 6.2, 2.5 Hz, 1H), 5.24 – 5.13 (m, 2H), 5.07 (d, *J* = 12.1 Hz, 1H), 4.52 (dd, *J* = 9.6, 2.4 Hz, 1H), 3.96 (dd, *J* = 5.9, 3.2 Hz, 1H), 2.56 (d, *J* = 6.1 Hz, 1H), 2.01 – 1.89 (m, 1H), 1.46 (s, 9H), 1.33 (d, *J* = 6.4 Hz, 3H), 0.99 (d, *J* = 6.9 Hz, 3H), 0.76 (d, *J* = 6.8 Hz, 3H).

¹³C-NMR: (101 MHz, CDCl₃): δ 173.8, 169.8, 155.8, 134.8, 128.7, 128.7, 128.4, 80.5, 75.1, 72.5, 67.8, 57.1, 31.9, 28.3, 18.9, 16.9, 15.5.

IR: (neat, cm⁻¹): 3460, 2974, 2936, 1740, 1717, 1501, 1456, 1384, 1368, 1346, 1315, 1282, 1248, 1213, 1164, 1136, 1085, 1063, 1031, 997, 698.

HR-MS: (ESI): *m/z* calc. for C₂₁H₃₁NNaO₇ [M+Na]⁺ 432.1993, found 432.1984.

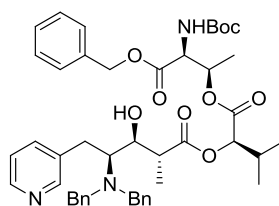


Alcohol *epi*-6: To a solution of **26** (460 mg, 878 μmol, 1.00 eq.) in 12 mL THF was added HF•py (30%, 2 mL in 2 portions) at 0 °C. The solution was stirred at RT for 1 h. More HF•py (30%, 1 mL) was added and the mixture was stirred at RT for 16 h. It was then quenched with sat. aq. NaHCO₃ (55 mL) and extracted with EtOAc (3 x 15 mL). The combined org. extracts were dried over MgSO₄ and

concentrated *in vacuo*. The residue was purified by FC (hexane/EtOAc 4:1) to yield *epi-6* as a colorless oil (344 mg, 96%).

R_f: 0.21 (hexane/EtOAc 4:1)
[α]_D²⁰: +2.58° (c 1.91, CHCl₃)
¹H-NMR: (400 MHz, CDCl₃): δ 7.41 – 7.29 (m, 5H), 5.50 (dd, *J* = 6.3, 2.2 Hz, 1H), 5.23 – 5.05 (m, 3H), 4.52 (dd, *J* = 9.6, 2.0 Hz, 1H), 3.76 (dd, *J* = 5.8, 3.4 Hz, 1H), 2.49 (d, *J* = 5.9 Hz, 1H), 1.98 (qd, *J* = 6.9, 3.4 Hz, 1H), 1.46 (s, 9H), 1.30 (d, *J* = 6.4 Hz, 3H), 0.98 (d, *J* = 6.9 Hz, 3H), 0.80 (d, *J* = 6.8 Hz, 3H).
¹³C-NMR: (100 MHz, CDCl₃): δ 173.8, 169.8, 155.8, 135.0, 128.7, 128.6, 128.5, 80.5, 74.5, 72.1, 67.7, 57.1, 31.9, 28.3, 18.8, 16.8, 15.7.
IR: (neat, cm⁻¹): 3449, 2971, 2936, 1739, 1716, 1500, 1456, 1384, 1367, 1316, 1248, 1213, 1163, 1083, 1062, 1031, 996, 753, 698.
HR-MS: (ESI): *m/z* calc. for C₂₁H₃₁NNaO₇ [M+Na]⁺ 432.1993, found 432.1998.

4. Synthesis of Analogs 2 and 3



19

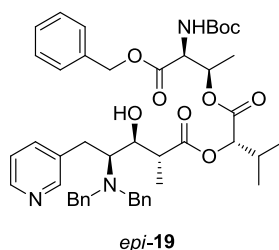
Ester 19: To a stirred solution of **5** (40.0 mg, 98.9 μmol, 1.00 eq.) and 2,4,6-trichlorobenzoyl chloride (27.1 μl, 173 μmol, 1.75 eq.) in 0.6 mL THF was added Et₃N (41.2 μl, 297 μmol, 3.00 eq.) at -78 °C. The mixture was stirred for 5 min and a solution of **6** (44.5 mg, 109 μmol, 1.10 eq.) and DMAP (15.7 mg, 129 μmol, 1.30 eq.) in 0.5 mL toluene was added at -78 °C. The clear solution was stirred at -78 °C for 30 min and then slowly warmed to -35 °C. The turbid mixture was stirred at that temperature for 43 h and allowed to warm to 0 °C for the last 25 min. The reaction was quenched at 0 °C with sat. aq. NaHCO₃ (2 mL). The aq. phase was extracted with EtOAc (3 x 10 mL) and the combined org. extracts were dried over MgSO₄ and concentrated *in vacuo*. The yellow oil was purified by FC (hexane/EtOAc 3:2) to yield **19** as a colorless film (43.0 mg, 50%).

R_f: 0.24 (hexane/EtOAc 3:2)
[α]_D²⁰: +49.3° (c 1.12, CHCl₃)
¹H-NMR: (400 MHz, CDCl₃): δ 8.45 (d, *J* = 1.7 Hz, 1H), 8.35 (dd, *J* = 4.7, 1.2 Hz, 1H), 7.51 (dt, *J* = 7.7, 1.7 Hz, 1H), 7.28 – 7.05 (m, 16H), 5.62 (d, *J* = 10.0 Hz, 1H), 5.46 (qd, *J* = 6.2, 2.5 Hz, 1H), 5.01 (d, *J* = 12.1 Hz, 1H), 4.93 (d, *J* = 12.1 Hz, 1H), 4.57 (d, *J* = 3.8 Hz, 1H), 4.40 (dd, *J* = 10.0, 2.4 Hz, 1H), 4.16 (br. s, 2H), 3.82 (d, *J* = 4.0 Hz, 1H), 3.39 – 3.26 (m, 3H), 3.22 – 2.97 (m, 3H), 2.77 – 2.64 (m, 1H), 2.10 (qd, *J* = 10.6, 6.8 Hz, 1H), 1.30 (s, 9H), 1.11 (d, *J* = 6.4 Hz, 3H), 0.87 (d, *J* = 6.9 Hz, 3H), 0.83 (d, *J* = 6.8 Hz, 3H), 0.16 (d, *J* = 6.8 Hz, 3H).

¹³C-NMR: (100 MHz, CDCl₃): δ 175.8, 170.4, 169.0, 156.1, 150.8, 147.6, 140.1, 137.1, 135.8, 134.8, 129.5, 128.6, 128.5, 128.3, 127.1, 123.5, 80.2, 76.6, 73.9, 72.1, 68.0, 59.3, 57.2, 56.0, 42.1, 30.2, 28.3, 27.3, 18.9, 16.8, 16.8, 13.0.

IR: (neat, cm⁻¹): 3489, 2974, 2936, 2359, 1739, 1717, 1497, 1455, 1367, 1316, 1248, 1217, 1162, 1129, 1086, 1061, 987, 943, 753, 700.

HR-MS: (ESI): *m/z* calc. for C₄₆H₅₈N₃O₉ [M+H]⁺ 796.4168, found 796.4166.



Ester *epi-19*: To a stirred solution of **5** (34.0 mg, 84.1 μmol, 1.00 eq.) and 2,4,6-trichlorobenzoyl chloride (23.0 μl, 147 μmol, 1.75 eq.) in 0.5 mL THF was added Et₃N (35.1 μl, 252 μmol, 3.00 eq.) at -78 °C. The mixture was stirred for 5 min and a solution of *epi-6* (37.9 mg, 92.5 μmol, 1.10 eq.) and DMAP (13.4 mg, 109 μmol, 1.30 eq.) in 0.4 mL toluene was added at -78 °C. The clear solution was stirred at -78 °C for 30 min and then slowly warmed to -35 °C. The turbid mixture was stirred at that temperature for 45 h and then allowed to warm to 0 °C for the last 25 min. The reaction was quenched at 0 °C with sat. aq. NaHCO₃ (2 mL). The aq. phase was extracted with EtOAc (3 x 10 mL) and the combined org. extracts were dried over MgSO₄ and concentrated *in vacuo*. The yellow oil was purified by FC (hexane/EtOAc 3:2) to yield *epi-19* as a colorless film (47.1 mg, 64%).

R_f: 0.21 (hexane/EtOAc 3:2)

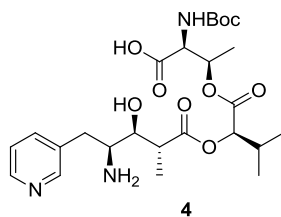
[α]_D²⁰: +45.0° (c 0.960, CHCl₃)

¹H-NMR: (400 MHz, CDCl₃): δ 8.42 (d, *J* = 1.7 Hz, 1H), 8.32 (dd, *J* = 4.8, 1.6 Hz, 1H), 7.50 – 7.43 (m, 1H), 7.27 – 7.03 (m, 16H), 5.43 – 5.33 (m, 1H), 5.25 (d, *J* = 9.5 Hz, 1H), 5.02 (d, *J* = 12.0 Hz, 1H), 4.93 (d, *J* = 12.0 Hz, 1H), 4.64 (d, *J* = 3.9 Hz, 1H), 4.37 (dd, *J* = 9.6, 2.5 Hz, 1H), 4.13 (d, *J* = 12.1 Hz, 2H), 3.62 (d, *J* = 4.0 Hz, 1H), 3.38 – 3.27 (m, 3H), 3.16 – 2.99 (m, 3H), 2.69 (dd, *J* = 8.4, 4.2 Hz, 1H), 2.07 – 1.92 (m, 1H), 1.30 (s, 9H), 1.13 (d, *J* = 6.4 Hz, 3H), 0.74 (d, *J* = 6.9 Hz, 3H), 0.67 (d, *J* = 6.8 Hz, 3H), 0.22 (d, *J* = 6.9 Hz, 3H).

¹³C-NMR: (100 MHz, CDCl₃) δ 174.5, 169.7, 169.6, 155.9, 150.7, 147.5, 140.0, 136.9, 136.0, 134.8, 129.3, 128.6, 128.5, 128.4, 127.1, 123.4, 80.3, 75.6, 73.7, 72.4, 68.0, 58.8, 57.1, 55.8, 43.7, 29.9, 28.3, 26.9, 18.6, 16.7, 16.6, 13.2.

IR: (neat, cm⁻¹): 2975, 2935, 1736, 1497, 1455, 1423, 1368, 1311, 1251, 1215, 1164, 1129, 1086, 1062, 1028, 985, 937, 752, 700.,

HR-MS: (ESI): *m/z* calc. for C₄₆H₅₈N₃O₉ [M+H]⁺ 796.4168, found 796.4163.



Amino acid 4: To a solution of **19** (41.2 mg, 51.8 μmol , 1.00 eq.) in 1 mL MeOH was added Pd on charcoal (10%, 22.0 mg, 20.7 μmol , 0.400 eq.) under Ar. The Ar was exchanged for H₂ (1 bar) and the mixture was stirred at RT for 5 h. The suspension was filtered through celite, the filter cake was washed with MeOH, and the filtrate was concentrated *in vacuo* to yield **4** as a white solid which was used crude in the next step (27.6 mg, quant.).

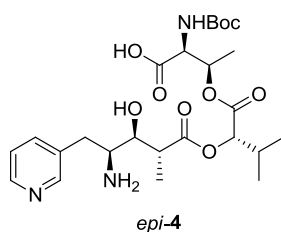
$[\alpha]_D^{20}$: +15.3° (c 1.32, MeOH)

¹H-NMR: (400 MHz, D₂O): δ 8.82 (d, J = 1.4 Hz, 1H), 8.78 (d, J = 5.6 Hz, 1H), 8.65 – 8.58 (m, 1H), 8.10 (dd, J = 8.0, 5.9 Hz, 1H), 5.61 – 5.44 (m, 1H), 4.96 (d, J = 4.2 Hz, 1H), 4.34 (d, J = 2.9 Hz, 1H), 4.09 – 3.99 (m, 1H), 3.88 (t, J = 5.7 Hz, 1H), 3.53 (dd, J = 15.0, 6.0 Hz, 1H), 3.27 (dd, J = 15.0, 8.7 Hz, 1H), 3.07 (p, J = 6.9 Hz, 1H), 2.28 – 2.15 (m, 1H), 1.45 (s, 9H), 1.33 (d, J = 6.4 Hz, 3H), 1.26 (d, J = 7.0 Hz, 3H), 0.95 (d, J = 6.9 Hz, 3H), 0.88 (d, J = 6.8 Hz, 3H).

¹³C-NMR: (100 MHz, D₂O): δ 174.7, 174.1, 170.7, 158.0, 147.6, 141.8, 140.8, 136.0, 127.7, 81.7, 77.8, 73.1, 71.4, 57.9, 53.3, 42.0, 32.5, 29.8, 27.6, 17.8, 16.3, 16.1, 13.4.

IR: (neat, cm⁻¹): 3362, 2974, 2935, 2881, 1722, 1505, 1469, 1369, 1311, 1252, 1168, 1129, 1058, 992, 685, 549.

HR-MS: (ESI): m/z calc. for C₂₅H₄₀N₃O₉ [M+H]⁺ 526.2759, found 526.2756.



Amino acid *epi-4*: To a solution *epi-19* (24.6 mg, 30.9 μmol , 1.00 eq.) in 0.8 mL MeOH was added Pd on charcoal (10%, 13.2 mg, 12.4 μmol , 0.400 eq.) under Ar. The Ar was exchanged for H₂ (1 bar) and the mixture was stirred at RT for 5 h. The suspension was filtered through celite, the filter cake was washed with MeOH, and the filtrate was concentrated *in vacuo* to yield *epi-4* as a white solid which was used crude in the next step (16.4 mg, quant.).

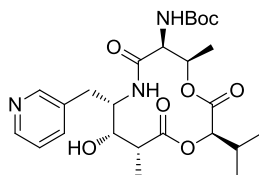
$[\alpha]_D^{20}$: +12.6° (c 1.35, MeOH)

¹H-NMR: (400 MHz, D₂O) δ 8.50 (d, J = 1.7 Hz, 2H), 7.88 (d, J = 7.9 Hz, 1H), 7.52 (dd, J = 7.8, 5.0 Hz, 1H), 5.36 (dt, J = 10.0, 5.9 Hz, 1H), 4.82 – 4.80 (m, 1H), 4.14 – 4.05 (m, 1H), 3.92 – 3.74 (m, 2H), 3.23 (dd, J = 14.5, 6.9 Hz, 1H), 3.08 (dd, J = 14.5, 7.6 Hz, 1H), 2.99 (p, J = 7.0 Hz, 1H), 2.28 – 2.14 (m, 1H), 1.45 – 1.43 (m, 1H), 1.42 (s, 9H), 1.22 (d, J = 7.1 Hz, 6H), 0.94 (t, J = 6.4 Hz, 6H).

¹³C-NMR: (100 MHz, D₂O): δ 175.6, 175.1, 170.6, 157.6, 148.7, 147.5, 139.0, 132.0, 124.9, 81.3, 78.1, 73.9, 70.8, 59.3, 53.5, 42.4, 32.8, 29.7, 27.6, 17.7, 16.6, 16.3, 13.5.

IR: (neat, cm⁻¹): 3401, 2975, 2937, 1720, 1596, 1501, 1389, 1250, 1171, 1131, 1055, 715.

HR-MS: (ESI): m/z calc. for $C_{25}H_{40}N_3O_9$ $[M+H]^+$ 526.2759, found 526.2752.



20

Protected amine 20: To a solution of DIEA (33.9 μ l, 196 μ mol, 2.60 eq.) and HATU (48.7 mg, 128 μ mol, 1.70 eq.) in 30 mL CH_2Cl_2 and 0.3 mL DMF was added a solution of **4** (39.6 mg, 75.3 μ mol, 1.00 eq.) in 20 mL CH_2Cl_2 and 0.2 mL DMF over a period of 4 h at RT (pale yellow color develops). The solution was stirred at RT for 18 h. Sat. aq. $NaHCO_3$ (2 mL) was added and the aq. phase was extracted with CH_2Cl_2 (3 x 5 mL). The combined org. extracts were dried over $MgSO_4$, concentrated *in vacuo*, and the residue was purified by FC (hexane/EtOAc 0.5:10) to deliver **20** as an orange film (24.1 mg, 63%).

R_f: 0.19 (hexane/EtOAc 05:10)

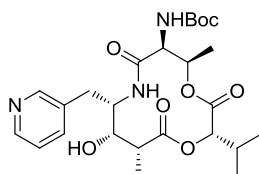
$[\alpha]_D^{20}$: -24.2° (*c* 1.21, MeOH)

1H -NMR: (400 MHz, MeOD): δ 8.46 (d, J = 1.7 Hz, 1H), 8.37 (dd, J = 4.9, 1.4 Hz, 1H), 7.79 – 7.69 (m, 1H), 7.33 (dd, J = 7.7, 5.0 Hz, 1H), 5.27 – 5.12 (m, 1H), 4.68 (d, J = 5.1 Hz, 1H), 4.24 (d, J = 5.9 Hz, 1H), 4.08 (td, J = 7.6, 1.6 Hz, 1H), 3.63 (s, 1H), 3.06 – 2.91 (m, 2H), 2.59 (qd, J = 7.2, 1.0 Hz, 1H), 2.27 – 2.16 (m, 1H), 1.45 (s, 9H), 1.37 (d, J = 7.4 Hz, 3H), 1.29 (d, J = 6.5 Hz, 3H), 0.99 (d, J = 4.2 Hz, 3H), 0.97 (d, J = 4.0 Hz, 3H).

^{13}C -NMR: (100 MHz, MeOD): δ 178.4, 170.9, 169.3, 157.1, 151.1, 148.0, 139.3, 136.3, 125.0, 81.1, 79.1, 75.7, 70.9, 57.7, 56.8, 42.4, 36.4, 31.1, 28.7, 18.7, 18.0, 17.8, 15.0.

IR: (neat, cm^{-1}): 3350, 2974, 2935, 1744, 1717, 1673, 1503, 1459, 1388, 1370, 1251, 1169, 1049, 1023, 847, 558.

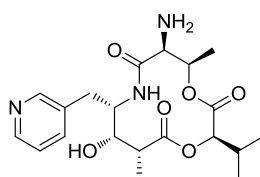
HR-MS: (ESI): m/z calc. for $C_{25}H_{38}N_3O_8$ $[M+H]^+$ 508.2653, found 508.2662.



epi-20

Protected amine epi-20: To a solution of DIEA (23.1 μ l, 87.3 μ mol, 2.60 eq.) and HATU (33.2 mg, 87.3 μ mol, 1.70 eq.) in 20 mL CH_2Cl_2 and 0.2 mL DMF was added a solution of *epi-4* (27.0 mg, 51.4 μ mol, 1.00 eq.) in 15 mL CH_2Cl_2 and 0.1 mL DMF over a period of 4 h at RT (pale yellow color develops). The solution was stirred at RT for 18 h. Sat. aq. $NaHCO_3$ (2 mL) was added and the aq. phase was extracted with CH_2Cl_2 (3 x 5 mL). The combined org. extracts were dried over $MgSO_4$, concentrated *in vacuo*, and the residue was purified by FC (hexane/EtOAc 0.5:10) to deliver *epi-20* as an orange film (20.5 mg, 79%).

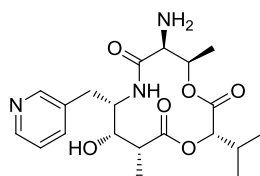
R_f: 0.23 (hexane/EtOAc 05:10)
[α]_D²⁰: -44.9° (c 1.03, MeOH)
¹H-NMR: (400 MHz, MeOD): δ 8.46 (d, *J* = 1.6 Hz, 1H), 8.38 (d, *J* = 3.9 Hz, 1H), 7.78 (d, *J* = 7.8 Hz, 1H), 7.35 (dd, *J* = 7.7, 5.0 Hz, 1H), 4.90 – 4.84 (m, 1H), 4.54 (d, *J* = 2.7 Hz, 1H), 4.35 (d, *J* = 4.6 Hz, 1H), 4.04 (t, *J* = 7.2 Hz, 1H), 3.65 (s, 1H), 3.05 (dd, *J* = 13.6, 6.4 Hz, 1H), 2.95 (dd, *J* = 13.6, 8.0 Hz, 1H), 2.44 (qd, *J* = 7.0, 2.0 Hz, 1H), 2.30 (dq, *J* = 13.5, 6.8 Hz, 1H), 1.45 (s, 9H), 1.31 (d, *J* = 7.2 Hz, 3H), 1.19 (d, *J* = 6.2 Hz, 3H), 1.07 (dd, *J* = 6.8, 4.4 Hz, 6H).
¹³C-NMR: (100 MHz, MeOD): δ 175.5, 173.0, 169.5, 151.1, 148.1, 139.3, 136.2, 130.8, 125.2, 81.2, 79.8, 73.3, 72.1, 57.6, 56.9, 47.5, 37.0, 31.0, 28.7, 19.4, 17.8, 17.2, 13.1.
IR: (neat, cm⁻¹): 3423, 2973, 2933, 1722, 1671, 1492, 1369, 1252, 1169, 1051, 1020, 847, 771, 716, 608, 561, 535, 510, 446.
HR-MS: (ESI): *m/z* calc. for C₂₅H₃₈N₃O₈ [M+H]⁺ 508.2653, found 508.2655.



21

Amine 21: To a solution of **20** (5.00 mg, 9.90 μmol, 1.00 eq.) in 0.6 mL CH₂Cl₂ was added TFA (75.4 μL, 985 mmol, 100 eq.) at 0 °C. The solution was stirred at RT for 3 h. The solvents were removed *in vacuo* to deliver **21** as a yellow oil which was used crude in the next step (10.0 mg, quant.).

[α]_D²⁰: -4.23° (c 0.965, MeOH)
¹H-NMR: (400 MHz, MeOD): δ 8.78 (s, 1H), 8.71 (d, *J* = 5.2 Hz, 1H), 8.51 – 8.44 (m, 1H), 7.94 (dd, *J* = 7.9, 5.7 Hz, 1H), 5.37 – 5.27 (m, 1H), 4.67 (d, *J* = 6.0 Hz, 1H), 4.32 (td, *J* = 7.2, 1.8 Hz, 1H), 4.04 (d, *J* = 5.7 Hz, 1H), 3.67 (t, *J* = 1.6 Hz, 1H), 3.24 (dd, *J* = 13.9, 7.0 Hz, 1H), 3.13 (dd, *J* = 13.9, 7.6 Hz, 1H), 2.65 (qd, *J* = 7.3, 1.2 Hz, 1H), 2.28 – 2.16 (m, 1H), 1.42 (d, *J* = 6.5 Hz, 3H), 1.38 (d, *J* = 7.4 Hz, 3H), 1.01 (d, *J* = 4.7 Hz, 3H), 0.99 (d, *J* = 4.8 Hz, 3H).
¹³C-NMR: (100 MHz, MeOD): δ 178.6, 169.0, 166.9, 147.1, 144.7, 142.3, 130.8, 127.8, 79.6, 75.0, 69.0, 57.4, 55.3, 42.2, 36.8, 30.9, 18.5, 17.9, 17.7, 15.1.
IR: (neat, cm⁻¹): 3358, 2971, 2935, 1745, 1672, 1537, 1472, 1392, 1263, 1173, 1138, 1056, 837, 798, 723, 706, 600, 549, 508, 469, 458.
HR-MS: (ESI): *m/z* calc. for C₂₀H₃₀N₃O₆ [M+H]⁺ 408.2129, found 408.2138.



epi-21

Amine *epi-21*: To a solution of *epi-20* (20.5 mg, 40.4 μmol , 1.00 eq.) in 2 mL CH_2Cl_2 was added TFA (309 μl , 4.04 mmol, 100 eq.) at 0 °C. The solution was stirred at RT for 2.4 h. The solvents were removed *in vacuo* to deliver *epi-21* as a pale yellow oil which was used crude in the next step (22.0 mg, quant.).

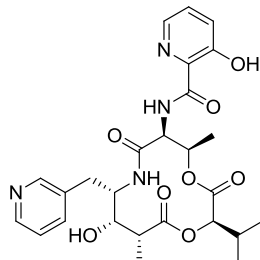
$[\alpha]_D^{20}$: -39.9° (*c* 0.820, MeOH)

$^1\text{H-NMR}$: (400 MHz, MeOD): δ 8.83 (s, 1H), 8.74 (d, $J = 5.6$ Hz, 1H), 8.57 (d, $J = 8.1$ Hz, 1H), 8.01 (dd, $J = 8.0, 5.8$ Hz, 1H), 5.11 – 5.01 (m, 1H), 4.55 (d, $J = 3.0$ Hz, 1H), 4.30 (dd, $J = 12.6, 6.4$ Hz, 1H), 4.22 (d, $J = 4.8$ Hz, 1H), 3.79 (d, $J = 1.9$ Hz, 1H), 3.25 (dd, $J = 13.8, 7.1$ Hz, 1H), 3.17 (dd, $J = 13.9, 7.1$ Hz, 1H), 2.57 (qd, $J = 7.1, 2.3$ Hz, 1H), 2.30 (dq, $J = 13.3, 6.7$ Hz, 1H), 1.34 (d, $J = 7.2$ Hz, 3H), 1.28 (d, $J = 6.0$ Hz, 3H), 1.12 – 0.99 (m, 6H).

$^{13}\text{C-NMR}$: (100 MHz, MeOD): δ 175.4, 169.0, 166.4, 148.9, 143.3, 140.8, 140.6, 128.2, 79.7, 73.8, 70.2, 56.0, 55.7, 37.9, 36.5, 31.0, 19.3, 17.8, 16.9, 13.0.

IR: (neat, cm^{-1}): 2973, 2933, 1735, 1673, 1526, 1473, 1282, 1201, 1135, 1069, 837, 798, 757, 722, 483, 470, 458, 444, 409.

HR-MS: (ESI): m/z calc. for $\text{C}_{20}\text{H}_{29}\text{N}_3\text{NaO}_6$ $[\text{M}+\text{Na}]^+$ 430.1949, found 430.1938.



2

Dihydropyridomycin **2:** To a solution of 3-hydroxypyridine-2-carboxylic acid (1.50 mg, 10.8 μmol , 1.10 eq.), HATU (4.48 mg, 11.8 μmol , 1.20 eq.) and DIEA (5.10 μl , 29.5 μmol , 3.00 eq.) in 100 μl MeCN was added a solution of **21** (4.00 mg, 9.82 μmol , 1.00 eq.) in 0.4 mL MeCN at RT. The mixture was stirred at RT for 18 h. The mixture was diluted with CH_2Cl_2 (1 mL) and sat. aq. NaHCO_3 (0.5 mL). The aq. phase was extracted with CH_2Cl_2 (3 x 2 mL) and the combined org. extracts were dried over MgSO_4 and concentrated *in vacuo*. The remaining green oil was purified by FC ($\text{CH}_2\text{Cl}_2/\text{MeOH}$ 5%) to yield **2** as a colorless film (2.7 mg, 52% over 2 steps). The samples prepared for biological testing were purified by reversed phase HPLC (Symmetry® C18 5 μm 19x100 mm column, gradient: 30% → 100% MeCN in H_2O in 14 min, flow: 25 mL/min, room temperature, $t_R = 6.88$ min) to a purity >98%.

R_f : 0.25 ($\text{CH}_2\text{Cl}_2/\text{MeOH}$ 5%)

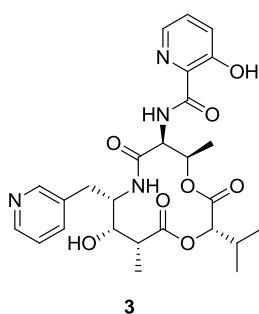
$[\alpha]_D^{20}$: -29.7° (*c* 0.110, MeOH)

¹H-NMR: (500 MHz, DMSO-d₆): δ 11.80 (s, 1H), 8.37 (d, *J* = 1.8 Hz, 1H), 8.30 (d, *J* = 4.7 Hz, 1H), 8.21 – 8.16 (m, 2H), 8.10 (d, *J* = 9.4 Hz, 1H), 7.58 (dd, *J* = 8.5, 4.4 Hz, 1H), 7.55 (dt, *J* = 7.8, 1.9 Hz, 1H), 7.46 (dd, *J* = 8.5, 1.3 Hz, 1H), 7.07 (dd, *J* = 7.7, 4.9 Hz, 1H), 5.29 (p, *J* = 6.4 Hz, 1H), 4.81 – 4.64 (m, 3H), 4.15 – 4.03 (m, 1H), 3.62 (d, *J* = 9.1 Hz, 1H), 2.86 (dd, *J* = 13.5, 6.0 Hz, 1H), 2.77 (dd, *J* = 13.4, 8.7 Hz, 1H), 2.66 – 2.56 (m, 1H), 2.19 – 2.06 (m, *J* = 6.7 Hz, 1H), 1.28 (d, *J* = 7.3 Hz, 3H), 1.21 (d, *J* = 6.5 Hz, 3H), 0.92 (dd, *J* = 6.8, 2.0 Hz, 6H)

¹³C-NMR: (126 MHz, DMSO-d₆): δ 176.2, 168.1, 167.5, 157.7, 150.8, 147.6, 140.6, 137.0, 134.0, 130.8, 130.2, 126.8, 123.4, 77.5, 73.9, 68.8, 56.1, 53.3, 41.4, 35.2, 29.8, 18.5, 17.7, 17.4, 15.0.

IR: (neat, cm⁻¹): 3370, 2971, 2938, 1745, 1650, 1522, 1450, 1386, 1296, 1254, 1168, 1062, 810, 778, 715, 656.

HR-MS: (ESI): *m/z* calc. for C₂₆H₃₃N₄O₈ [M+H]⁺ 529.2293, found 529.2305.



Dihydropyridomycin 3: To a solution of 3-hydroxypyridine-2-carboxylic acid (6.20 mg, 44.5 μmol, 1.10 eq.), HATU (18.5 mg, 48.6 μmol, 1.20 eq.) and DIEA (21.0 μl, 122 μmol, .300 eq.) in 0.4 mL MeCN (dark green) was added a solution of *epi*-**21** (16.5 mg, 40.5 μmol, 1.00 eq.) in 1.7 mL MeCN at RT. The mixture was stirred at RT for 24 h. The mixture was diluted with CH₂Cl₂ (2 mL) and sat. aq. NaHCO₃ (1 mL). The aq. phase was extracted with CH₂Cl₂ (3 x 5 mL) and the combined org. extracts were dried over MgSO₄ and concentrated *in vacuo*. The remaining orange oil was purified by FC (CH₂Cl₂/MeOH 5%) to yield **3** as a colorless film (12.0 mg, 56% over 2 steps). The samples prepared for biological testing were purified by reversed phase HPLC (Symmetry® C18 5 μm 19x100 mm column, gradient: 30% → 100% MeCN in H₂O in 14 min, flow: 25 mL/min, room temperature, *t_R* = 6.78 min) to a purity >98%.

R_f: 0.30 (CH₂Cl₂/MeOH 5%)

[α]_D²⁰: -60.5° (c 0.110, MeOH)

¹H-NMR: (500 MHz, DMSO-d₆): δ 11.84 (s, 1H), 8.58 (d, *J* = 6.4 Hz, 1H), 8.44 (d, *J* = 1.8 Hz, 1H), 8.30 (dd, *J* = 4.8, 1.6 Hz, 1H), 8.18 (dd, *J* = 4.3, 1.2 Hz, 1H), 7.65 (dt, *J* = 7.8, 1.8 Hz, 1H), 7.60 (d, *J* = 3.4 Hz, 1H), 7.56 (dd, *J* = 8.5, 4.4 Hz, 1H), 7.45 (dd, *J* = 8.5, 1.2 Hz, 1H), 7.21 (dd, *J* = 7.7, 4.8 Hz, 1H), 5.49 (s, 1H), 5.02 (p, *J* = 5.9 Hz, 1H), 4.88 (dd, *J* = 6.9, 5.4 Hz, 1H), 4.54 (d, *J* = 5.3 Hz, 1H), 4.05 (q, *J* = 7.6 Hz, 1H), 3.63 (d, *J* = 4.1 Hz, 1H), 2.85 (qd, *J* = 13.5, 7.2 Hz, 2H), 2.50 – 2.44 (m, 1H), 2.28 – 2.17 (m, 1H), 1.23 (d, *J* = 7.1 Hz, 3H), 1.09 (d, *J* = 6.2 Hz, 3H), 1.01 (dd, *J* = 6.7, 4.6 Hz, 6H).

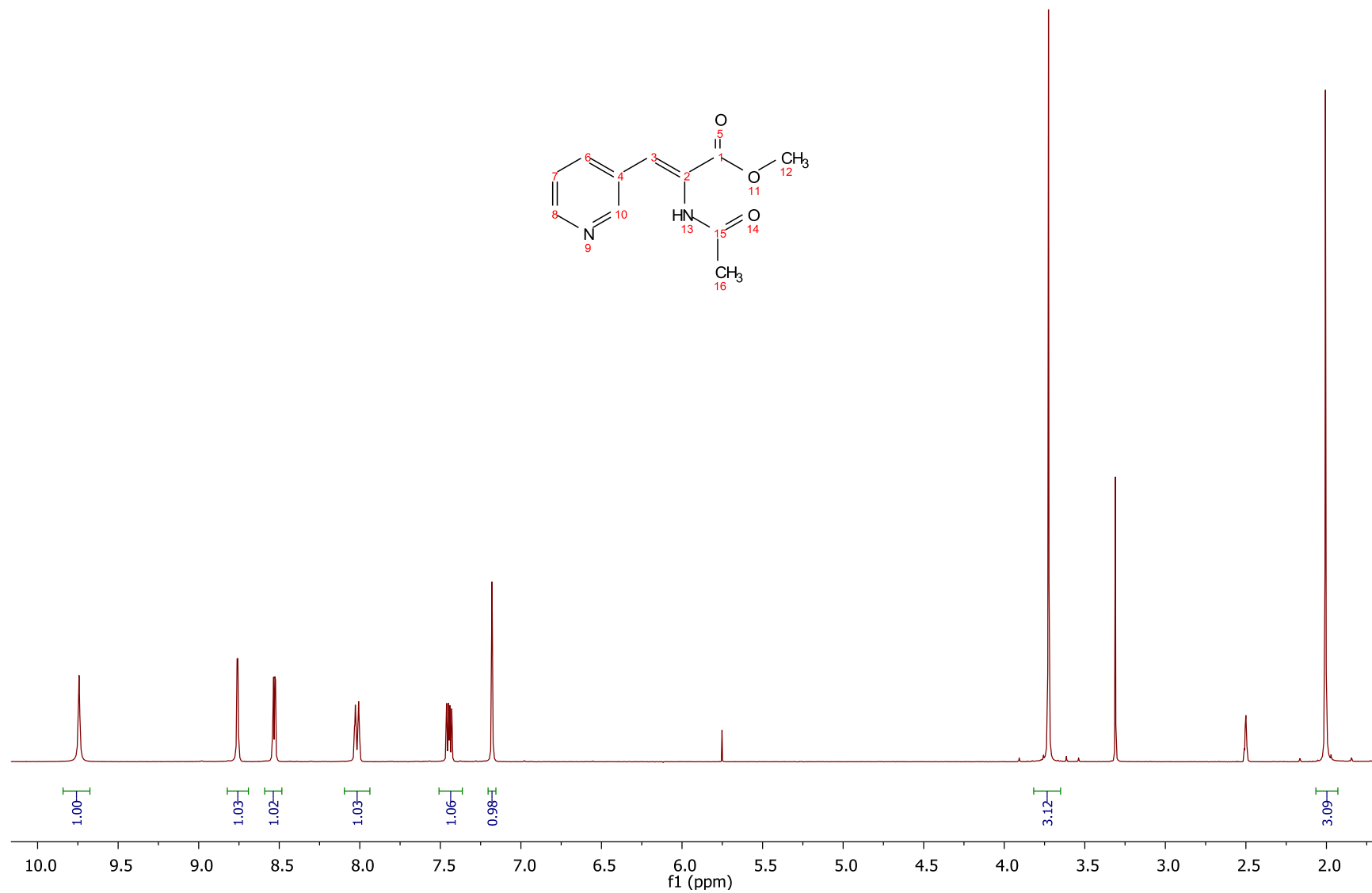
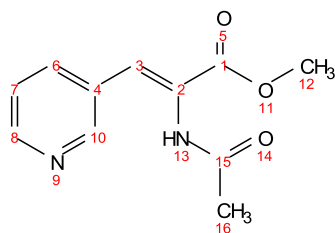
¹³C-NMR: (126 MHz, DMSO-d₆): δ 174.0, 168.3, 168.2, 166.7, 157.7, 150.8, 147.8, 140.7, 137.1, 134.6, 130.9, 130.2, 126.8, 123.7, 78.1, 71.7, 69.7, 55.2, 54.0, 45.8, 36.0, 29.8, 19.1, 17.9, 16.6, 13.1.

IR: (neat, cm^{-1}): 3373, 2977, 2942, 1732, 1650, 1510, 1450, 1293, 1257, 1062, 1026, 1013, 811, 783, 717, 662, 589.

HR-MS: (ESI): m/z calc. for $\text{C}_{26}\text{H}_{33}\text{N}_4\text{O}_8$ $[\text{M}+\text{H}]^+$ 529.2293, found 529.2281.

5. ^1H - and ^{13}C -NMR Spectra of New Comp

11



11

— 168.4
— 165.4

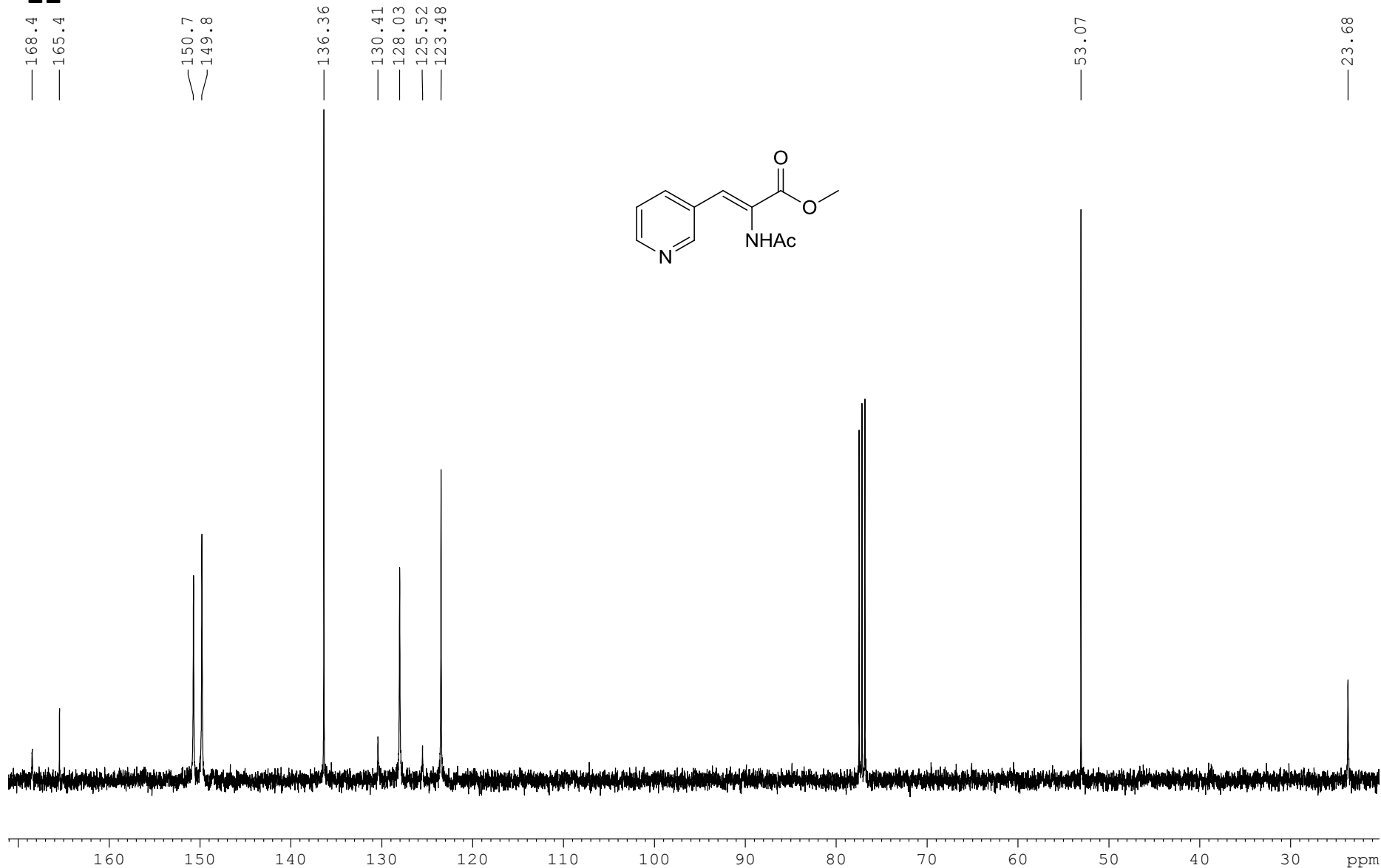
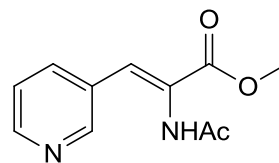
— 150.7
— 149.8

— 136.36

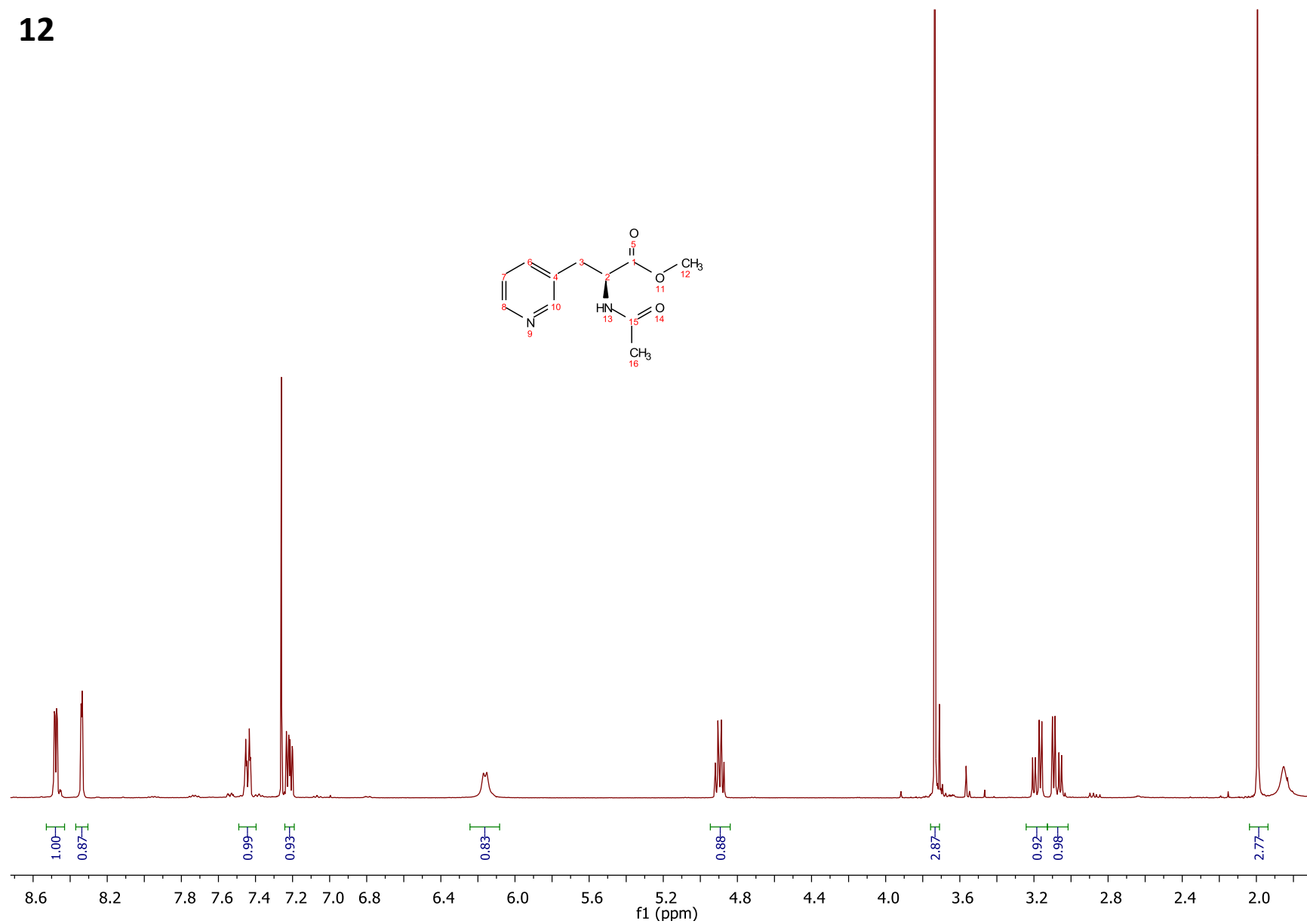
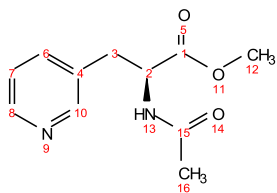
— 130.41
— 128.03
— 125.52
— 123.48

— 53.07

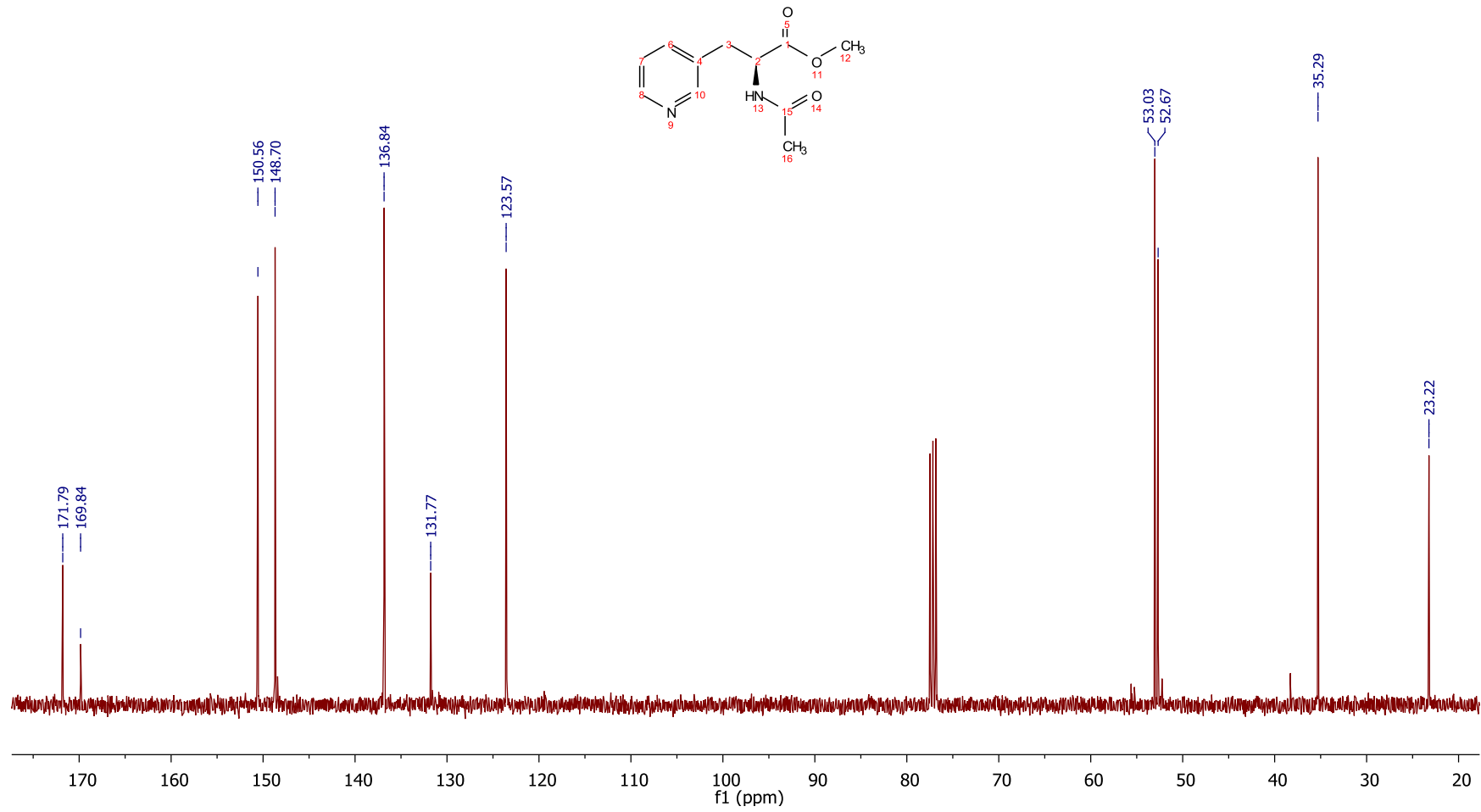
— 23.68



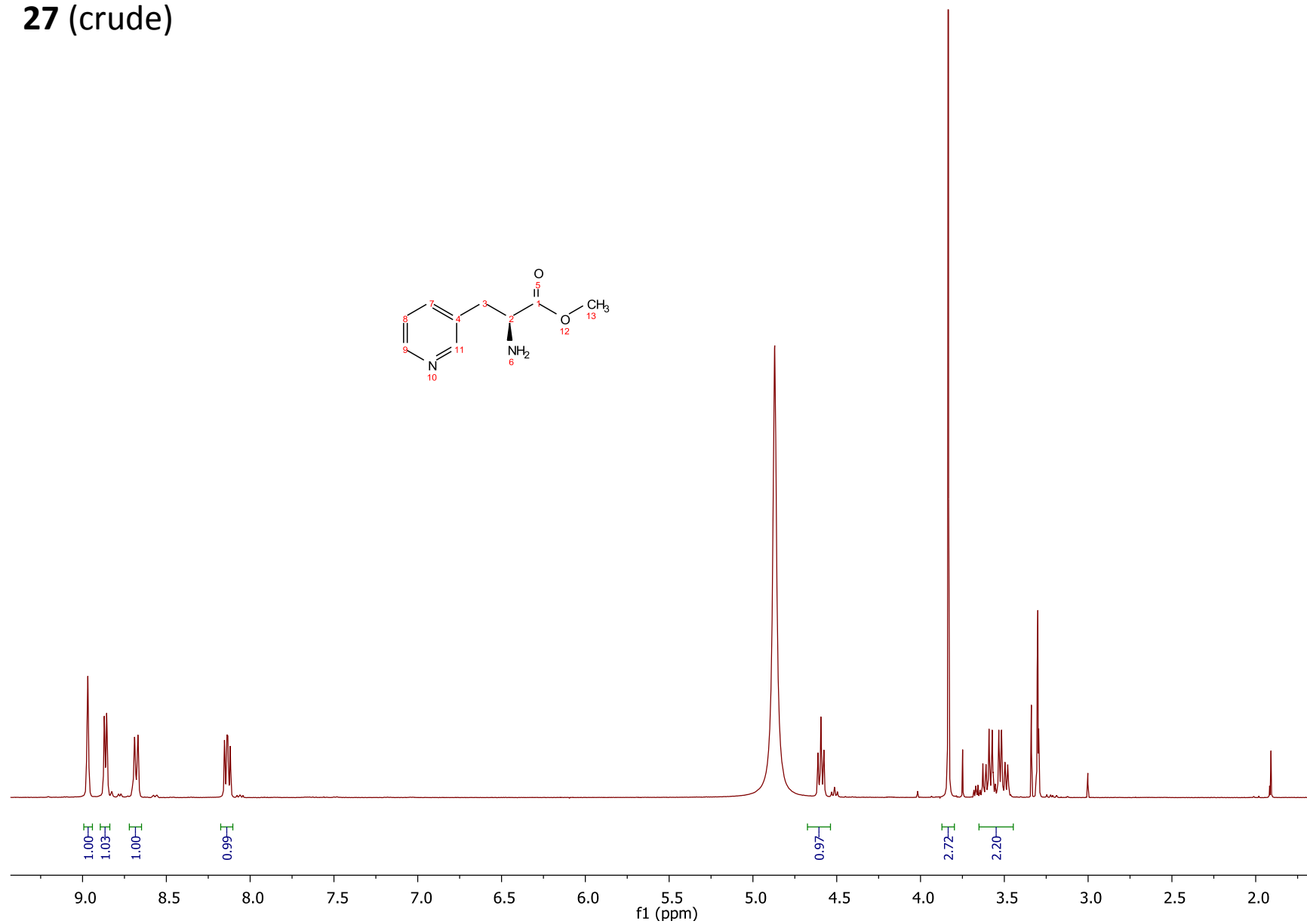
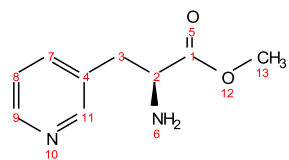
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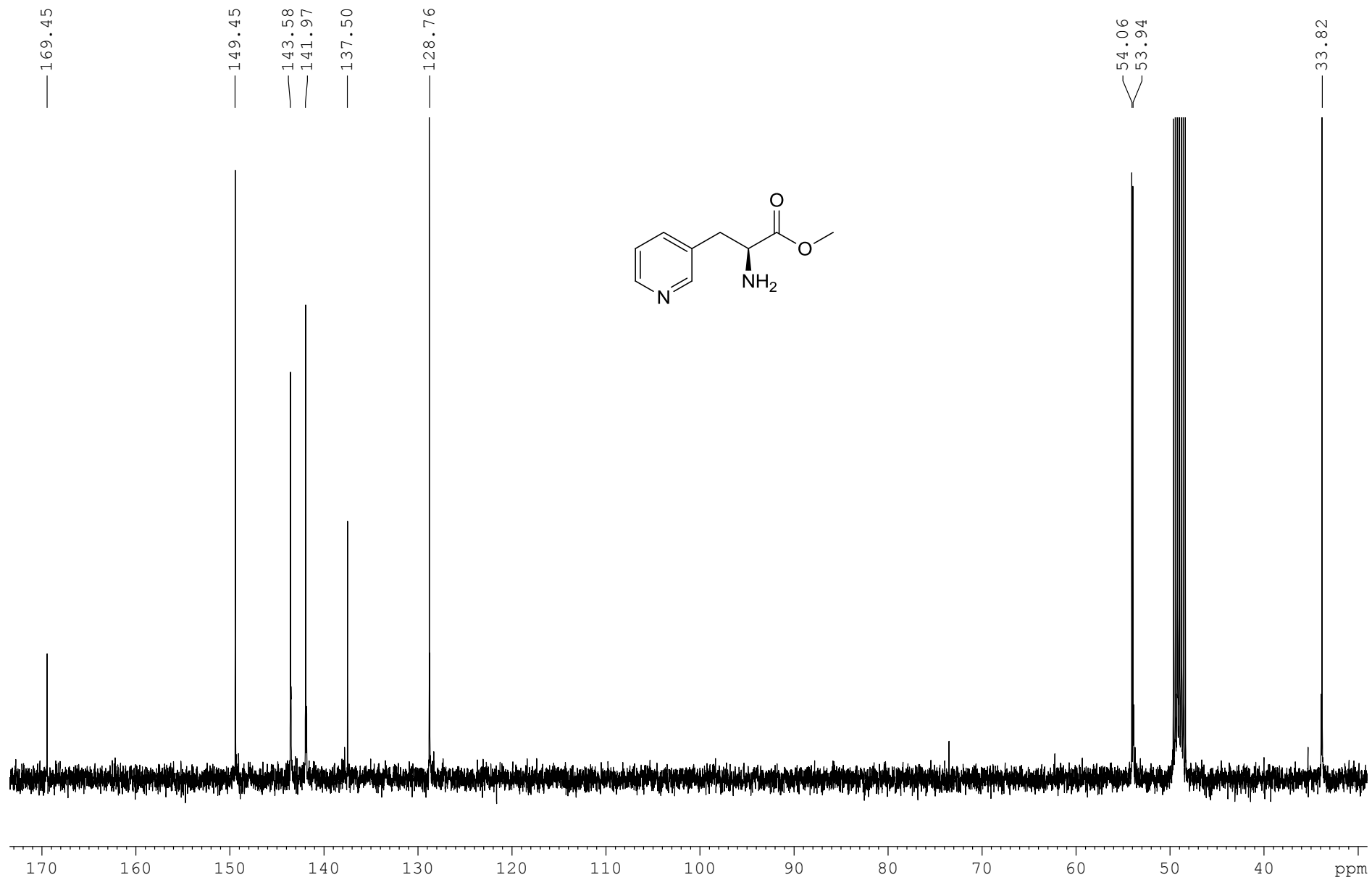
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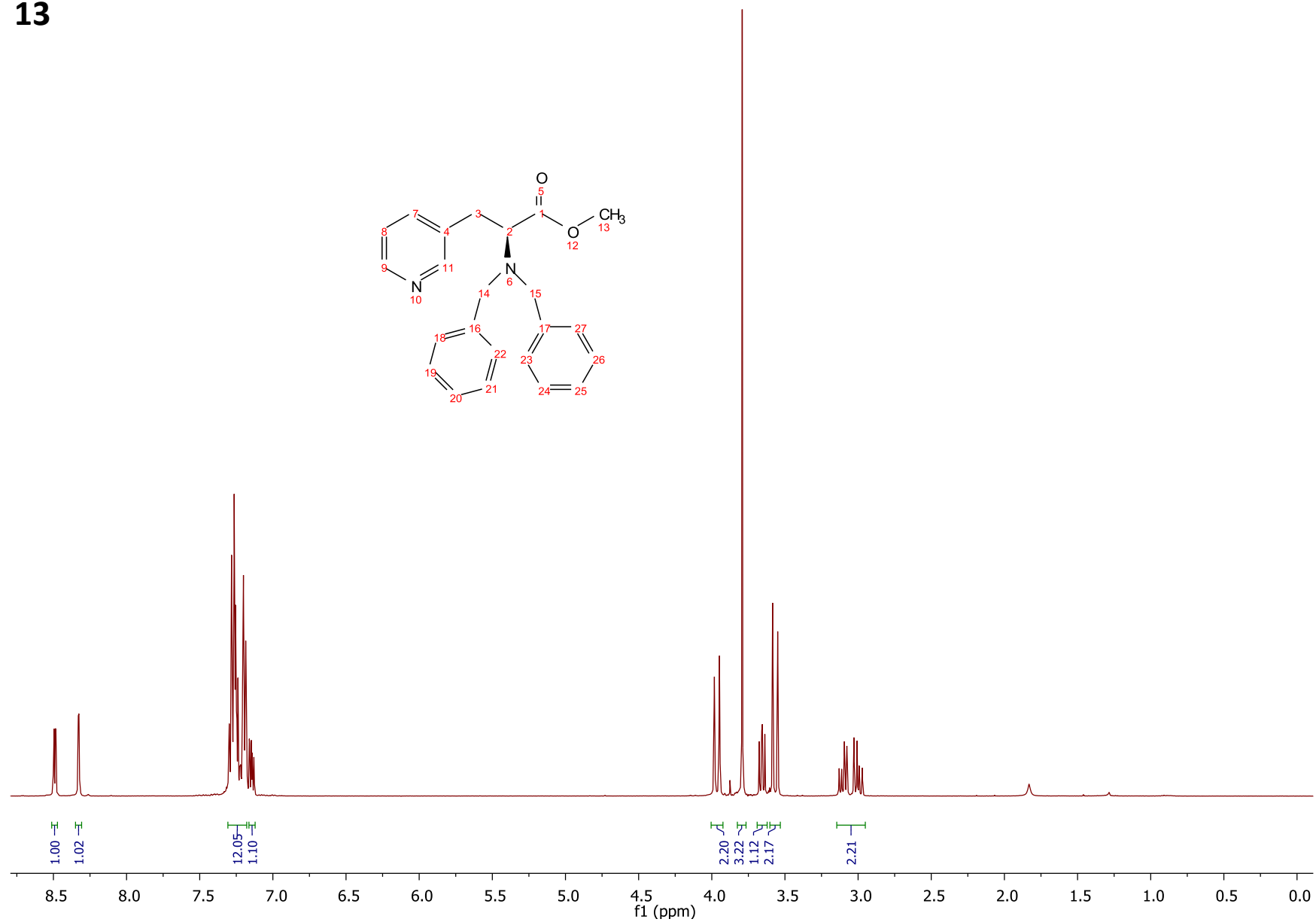
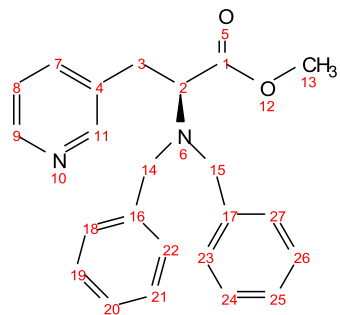
27 (crude)



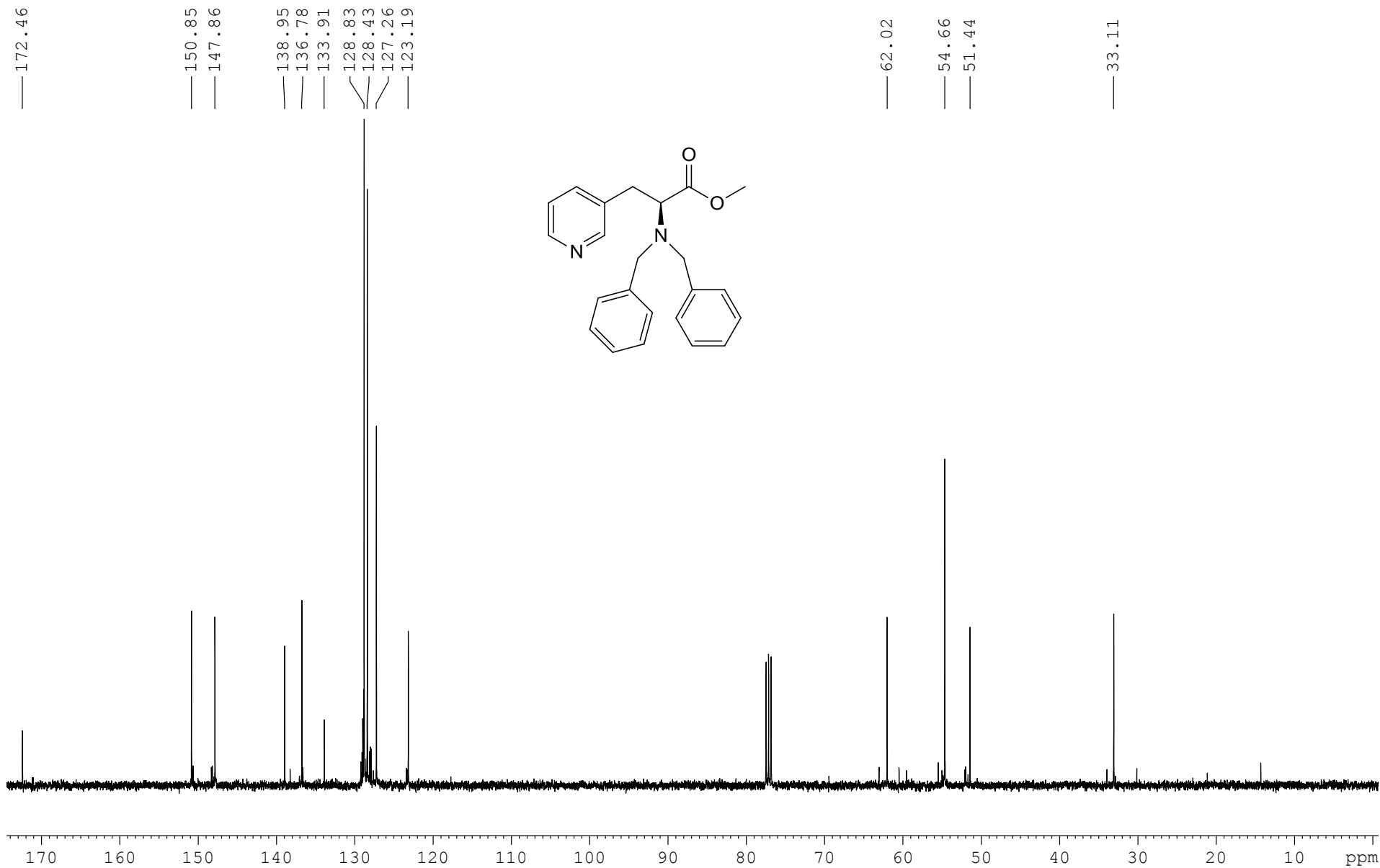
27 (crude)



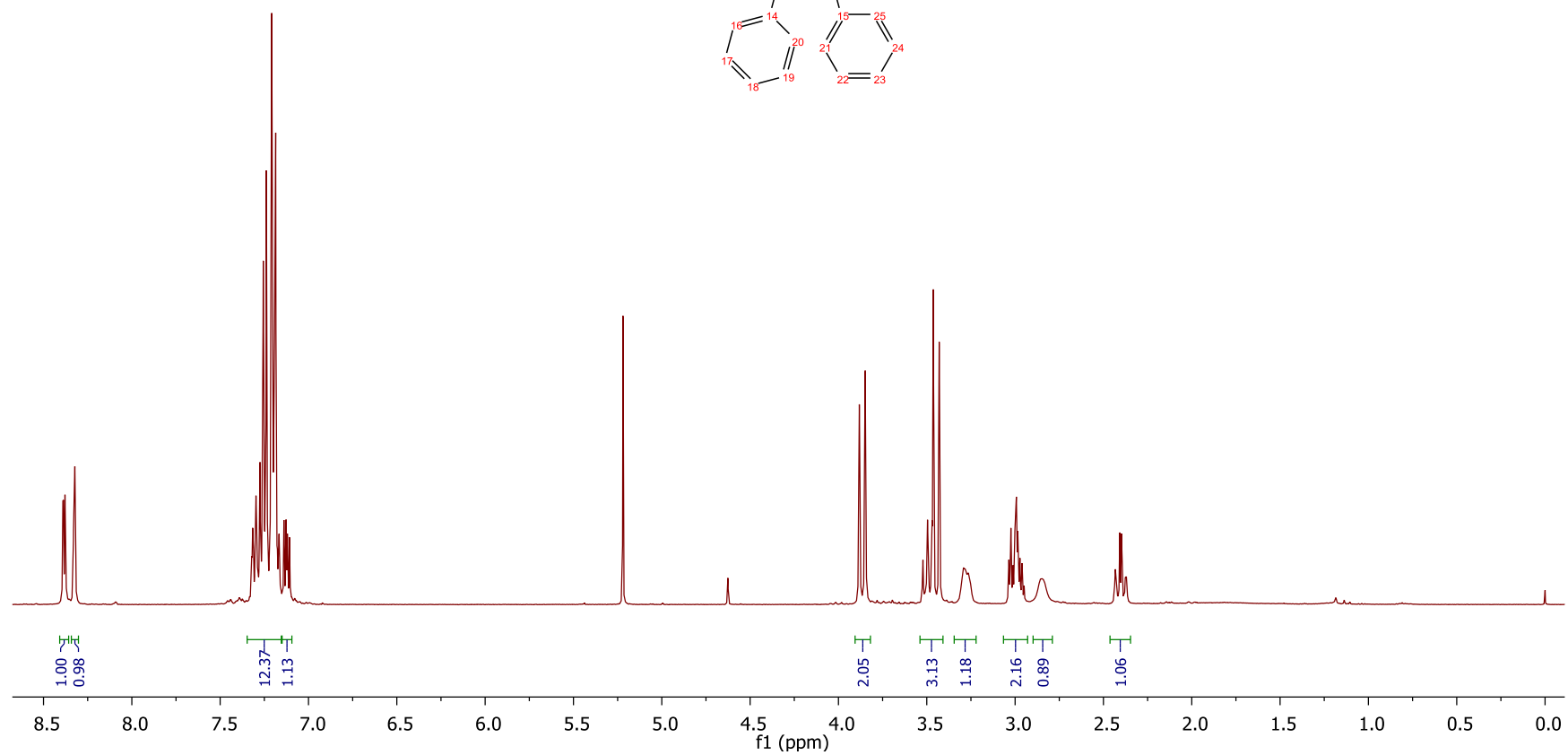
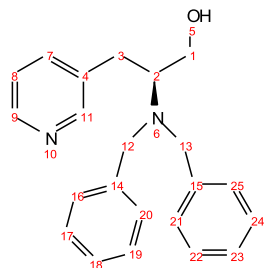
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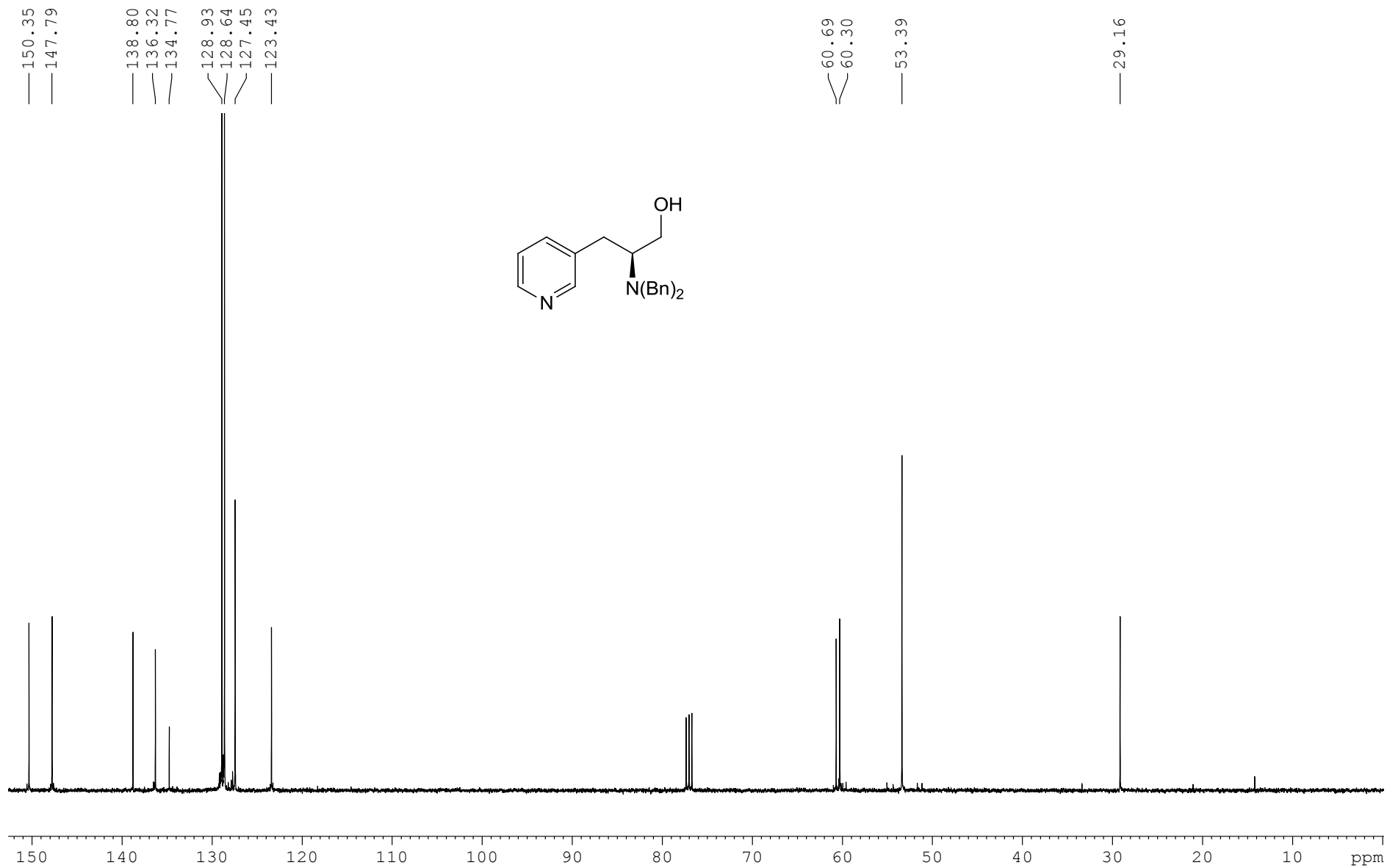
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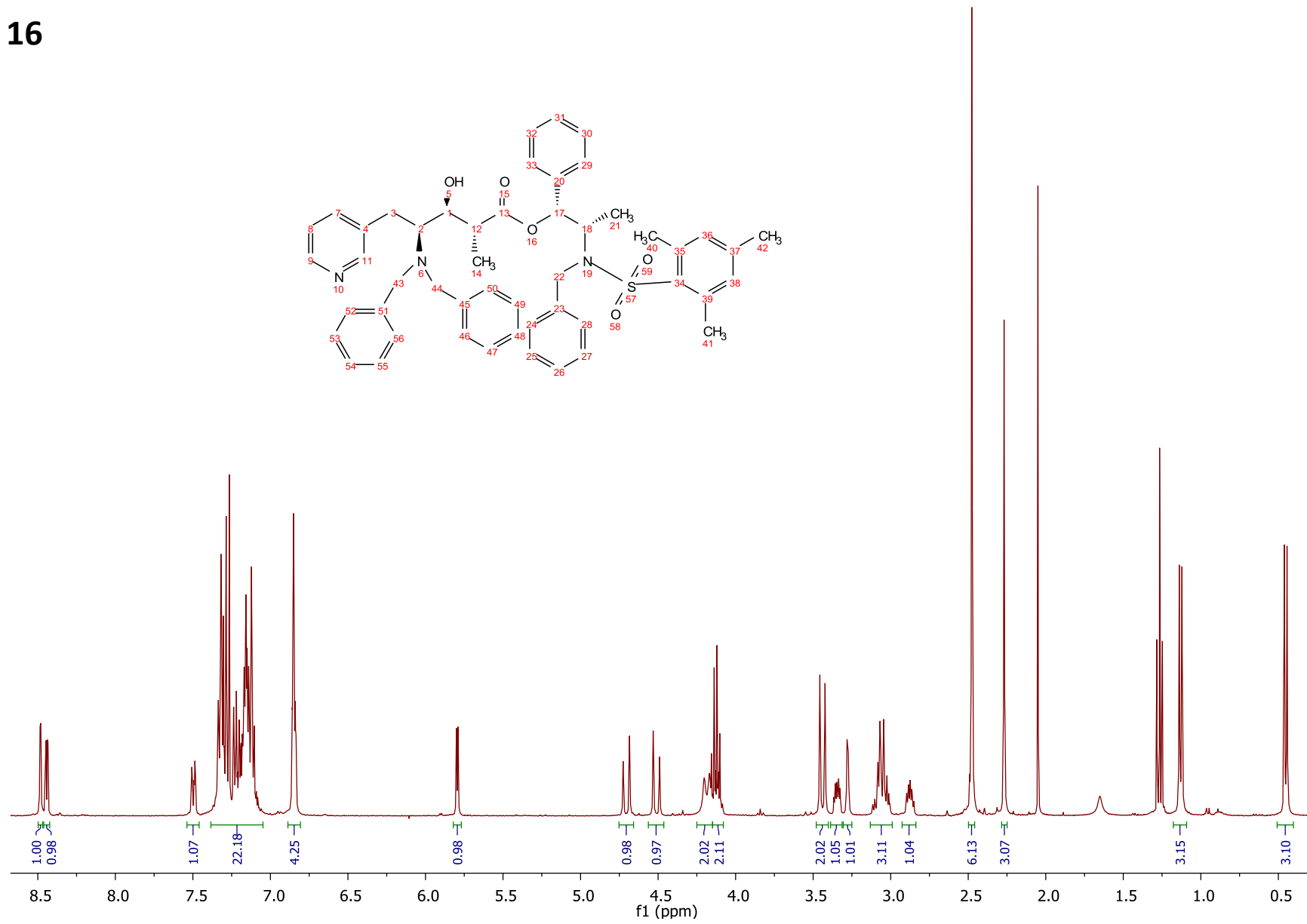
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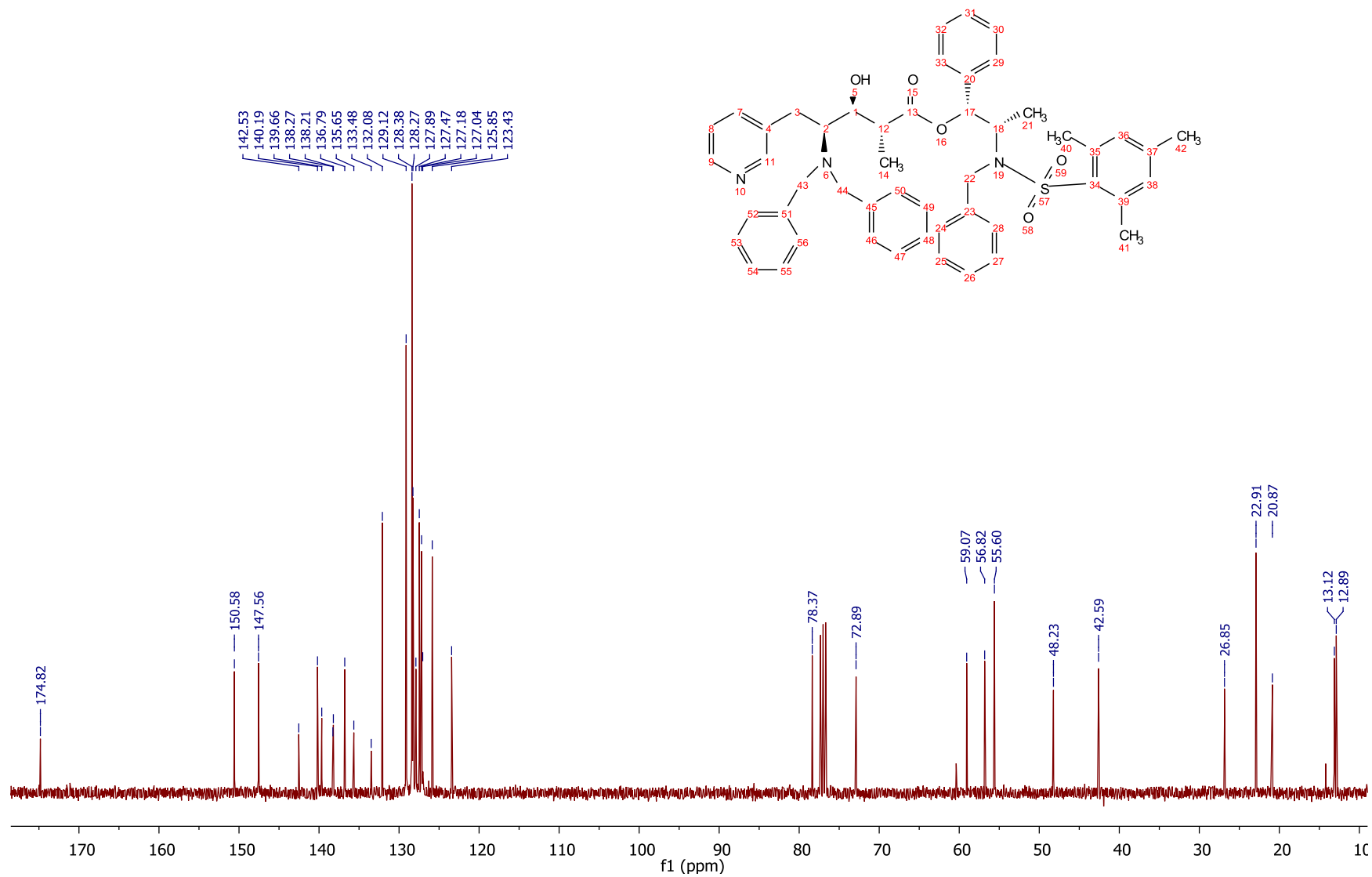
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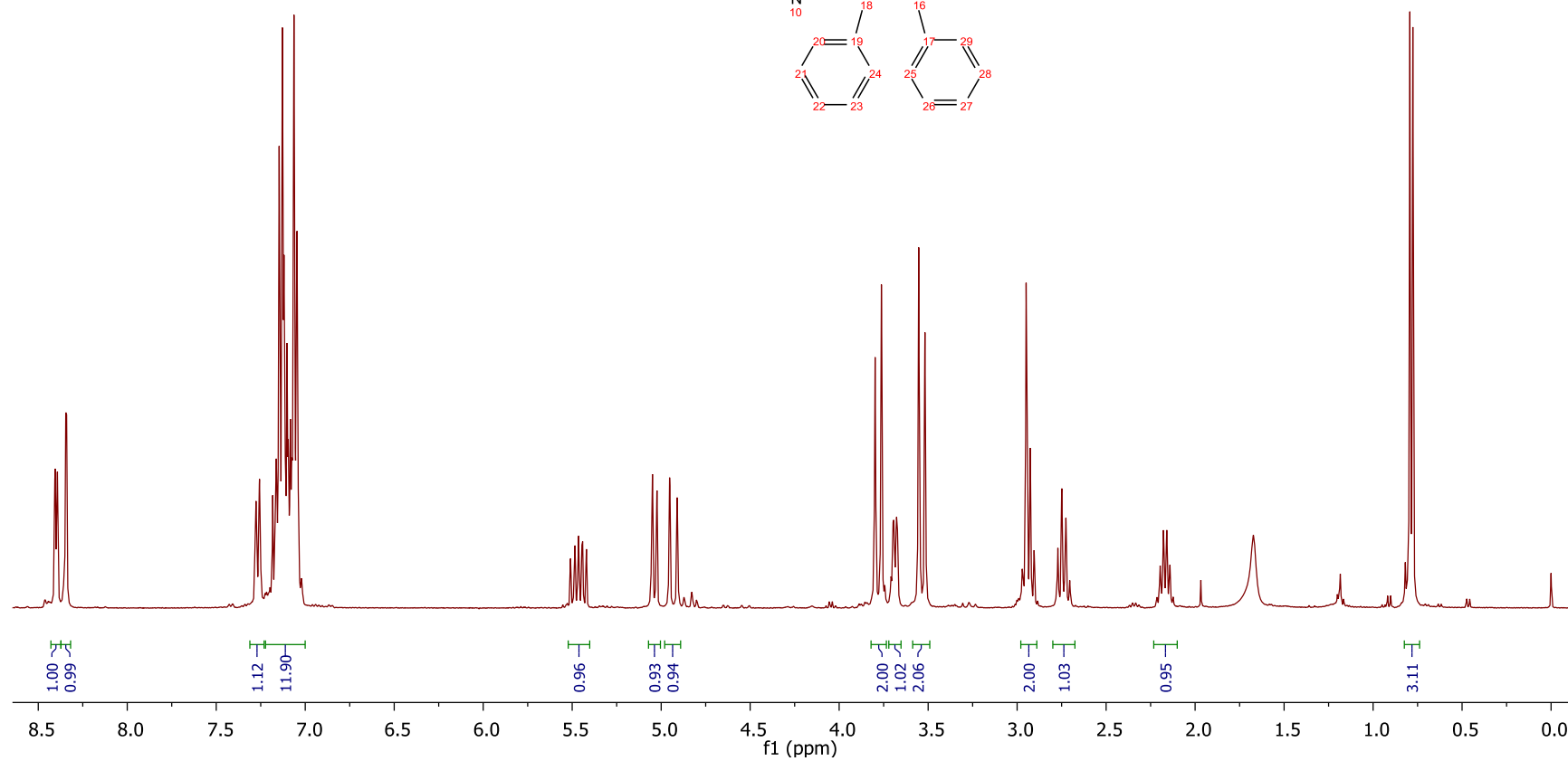
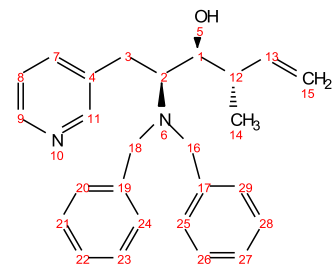
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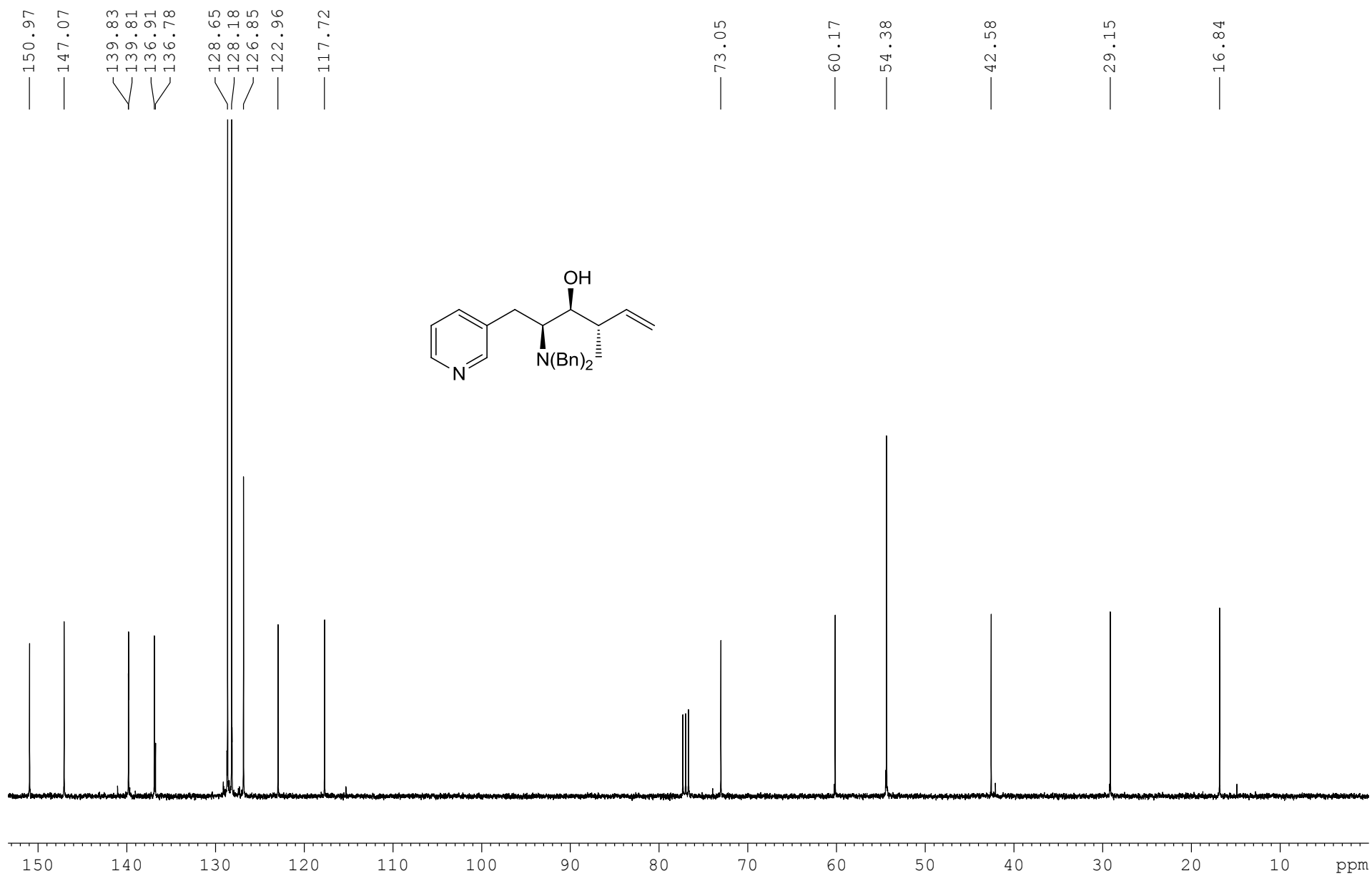
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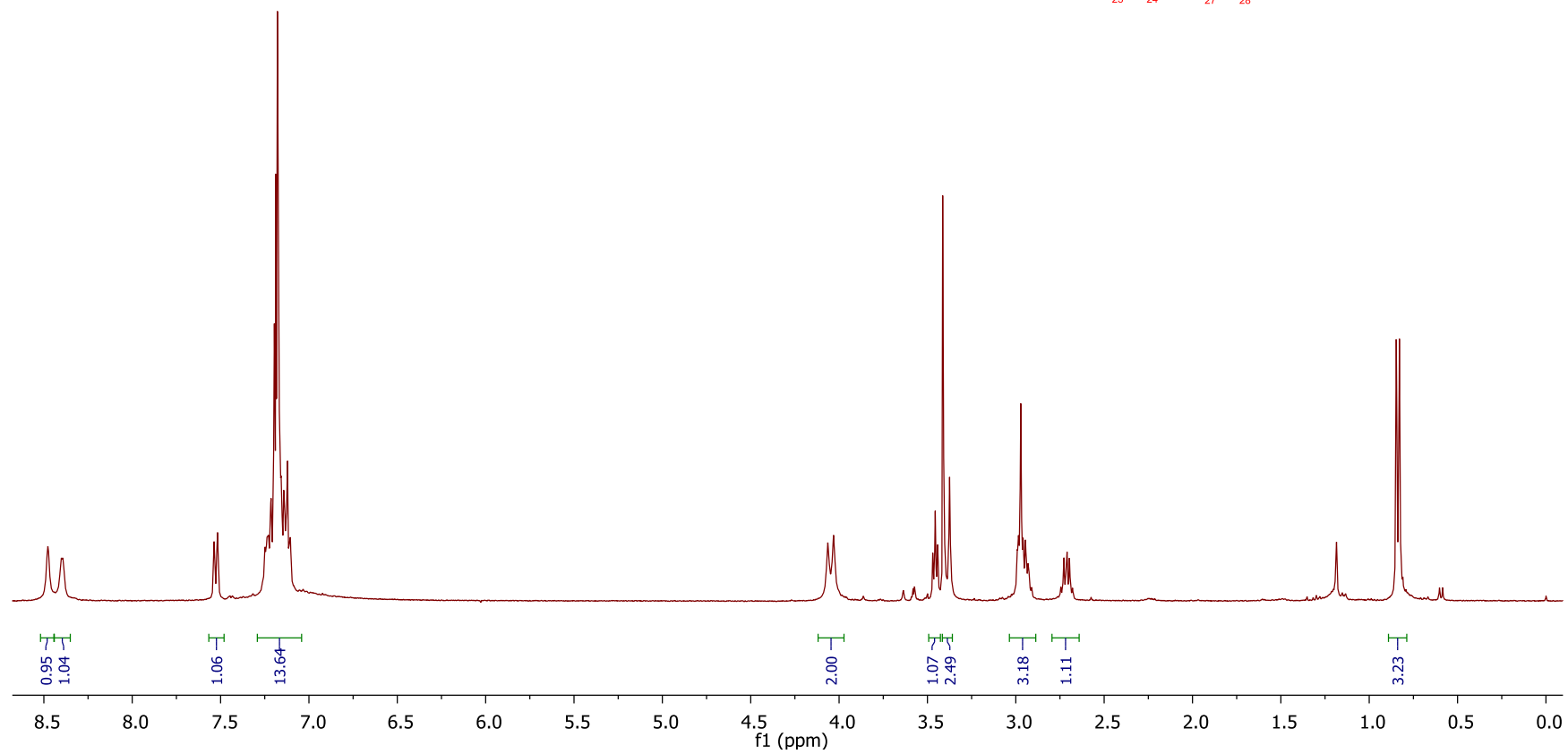
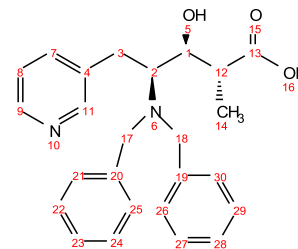
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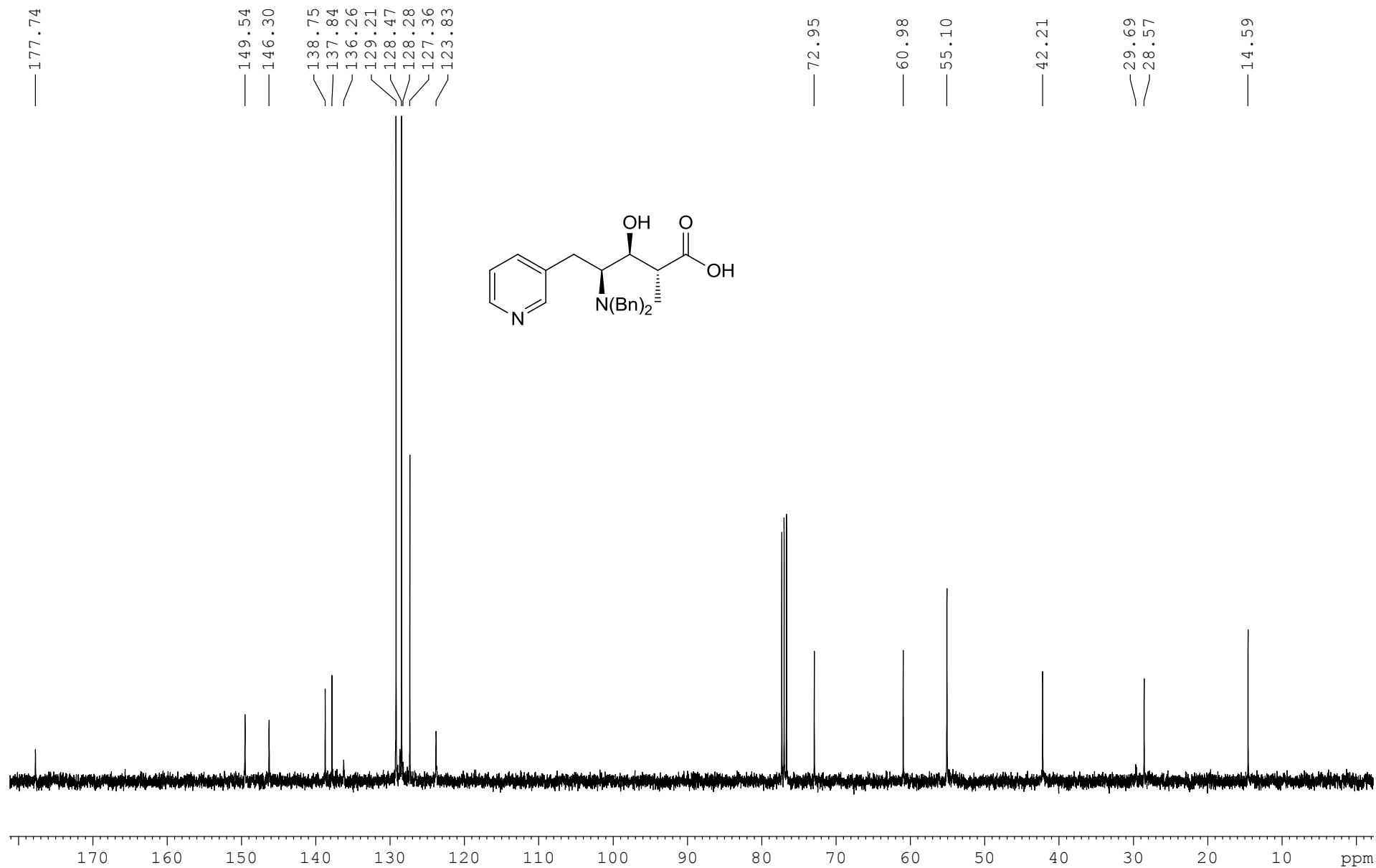
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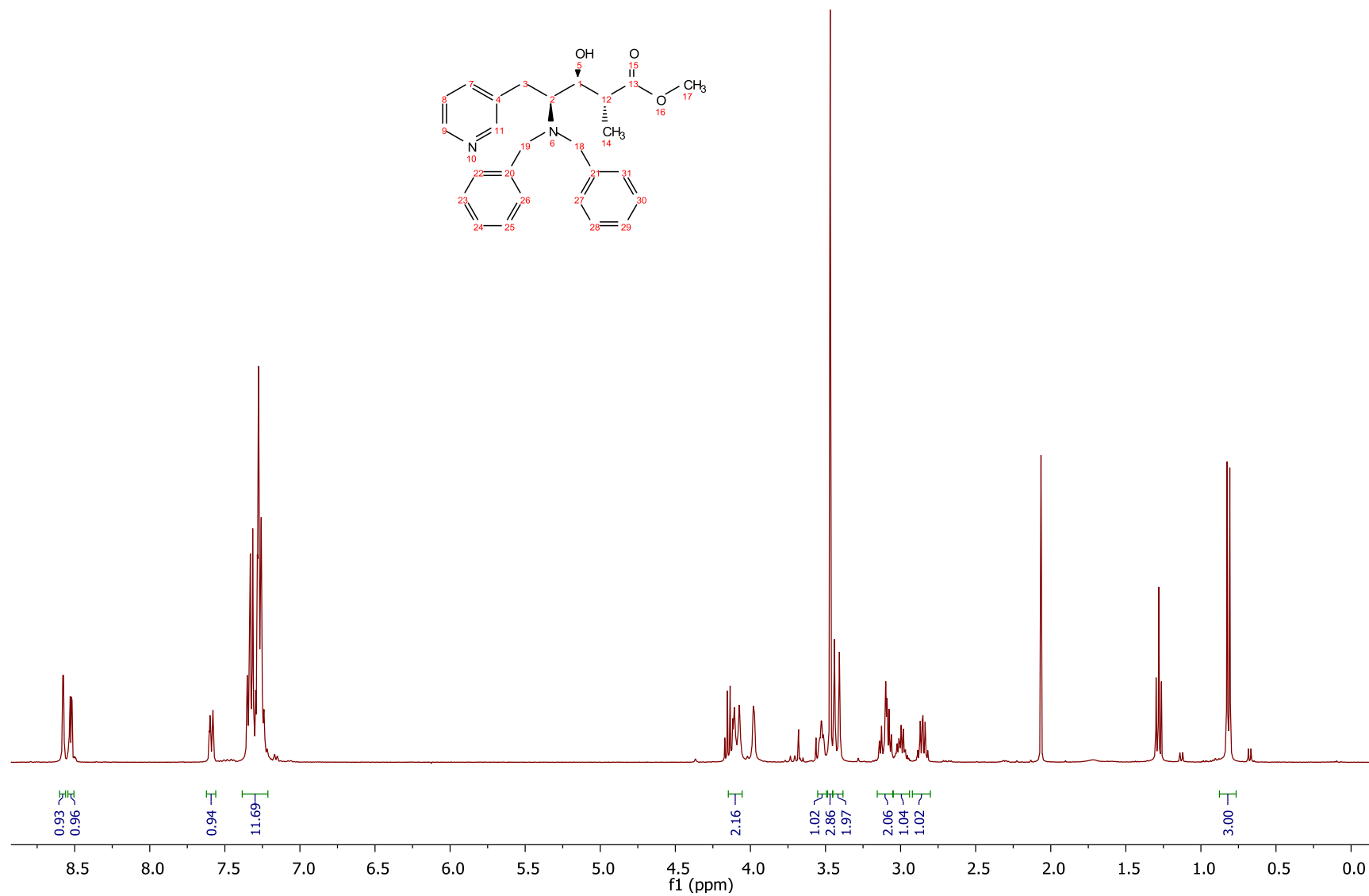
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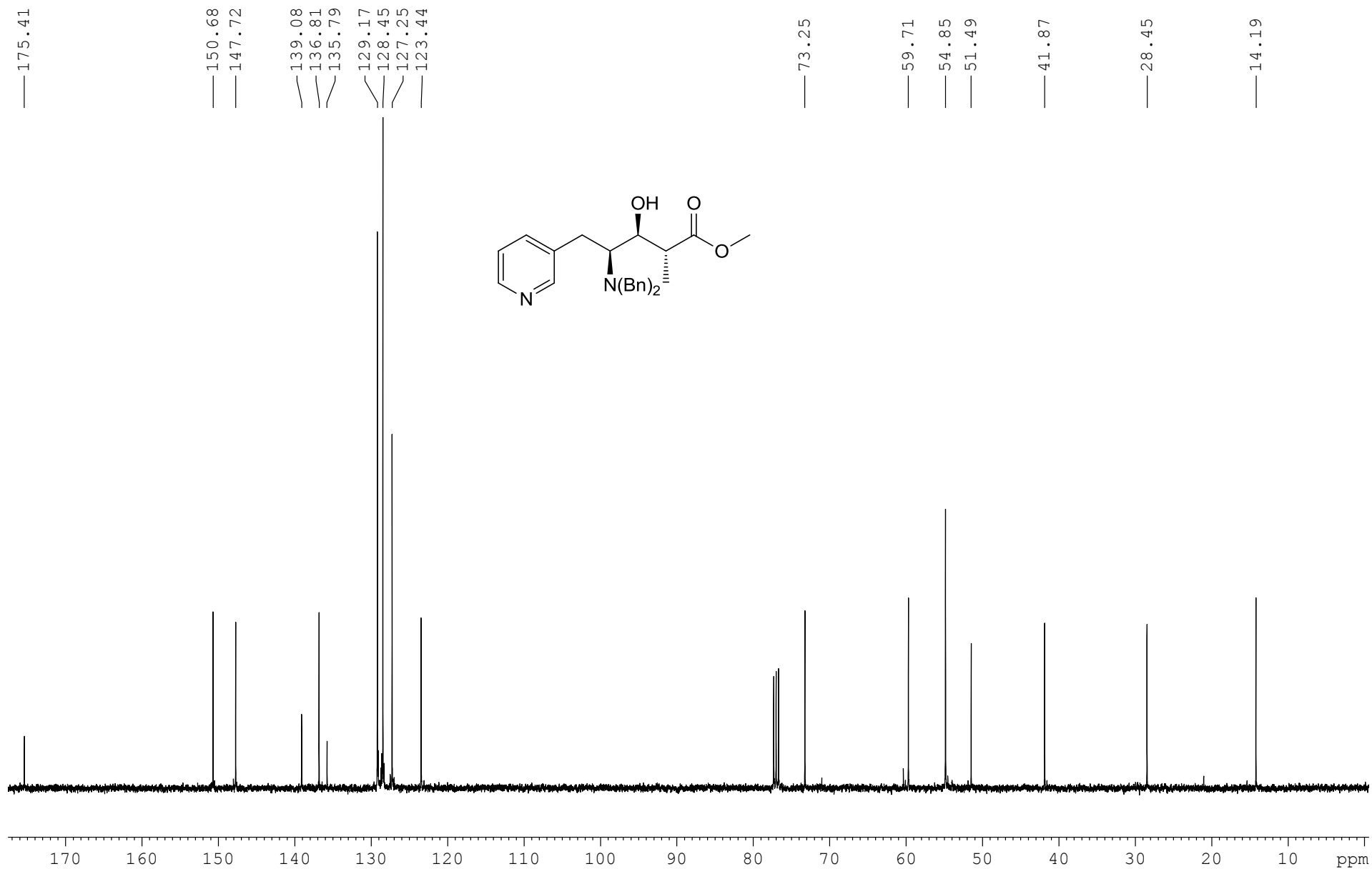
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22

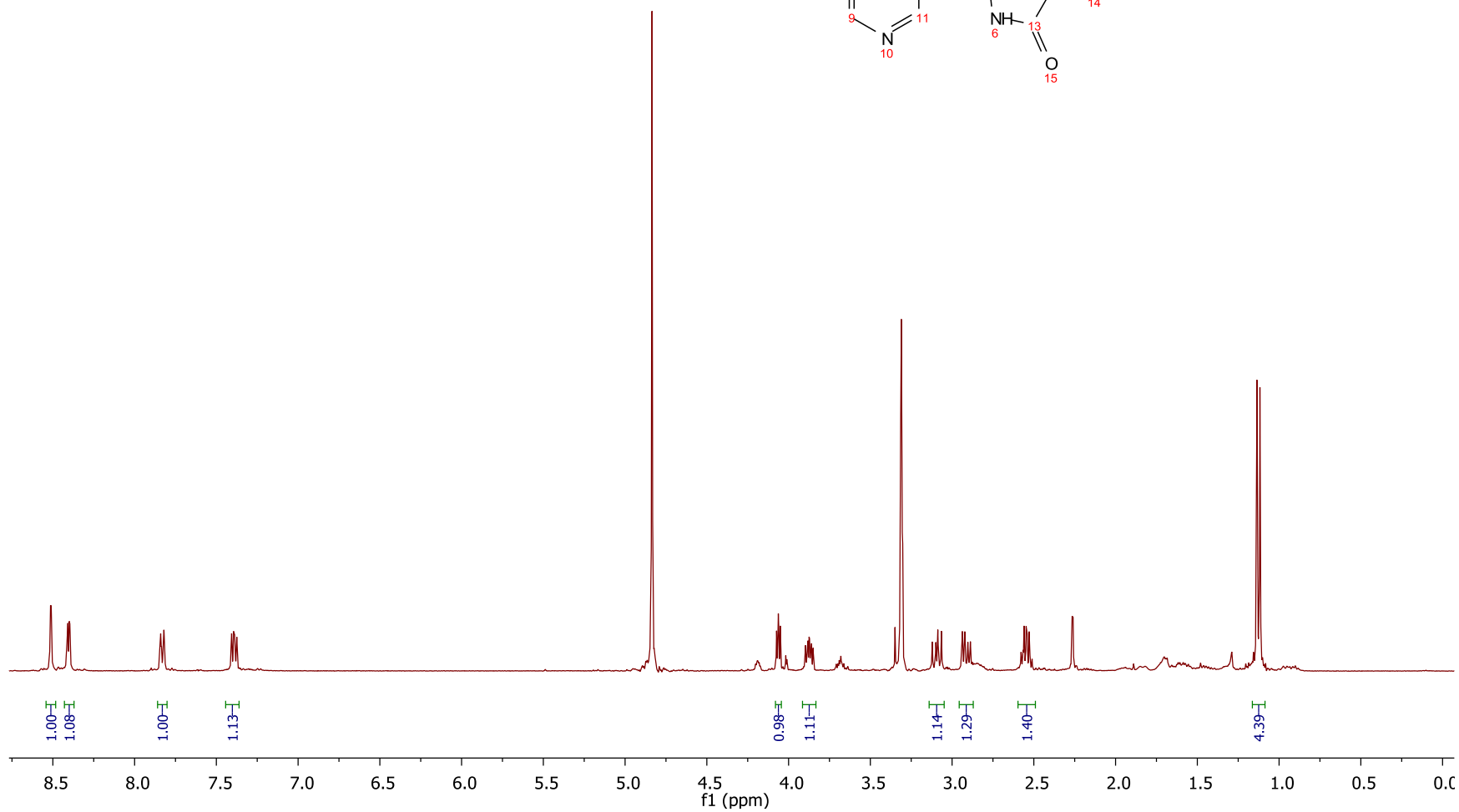
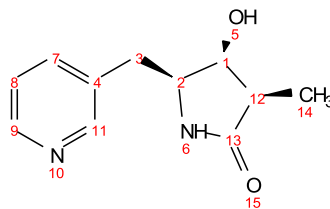


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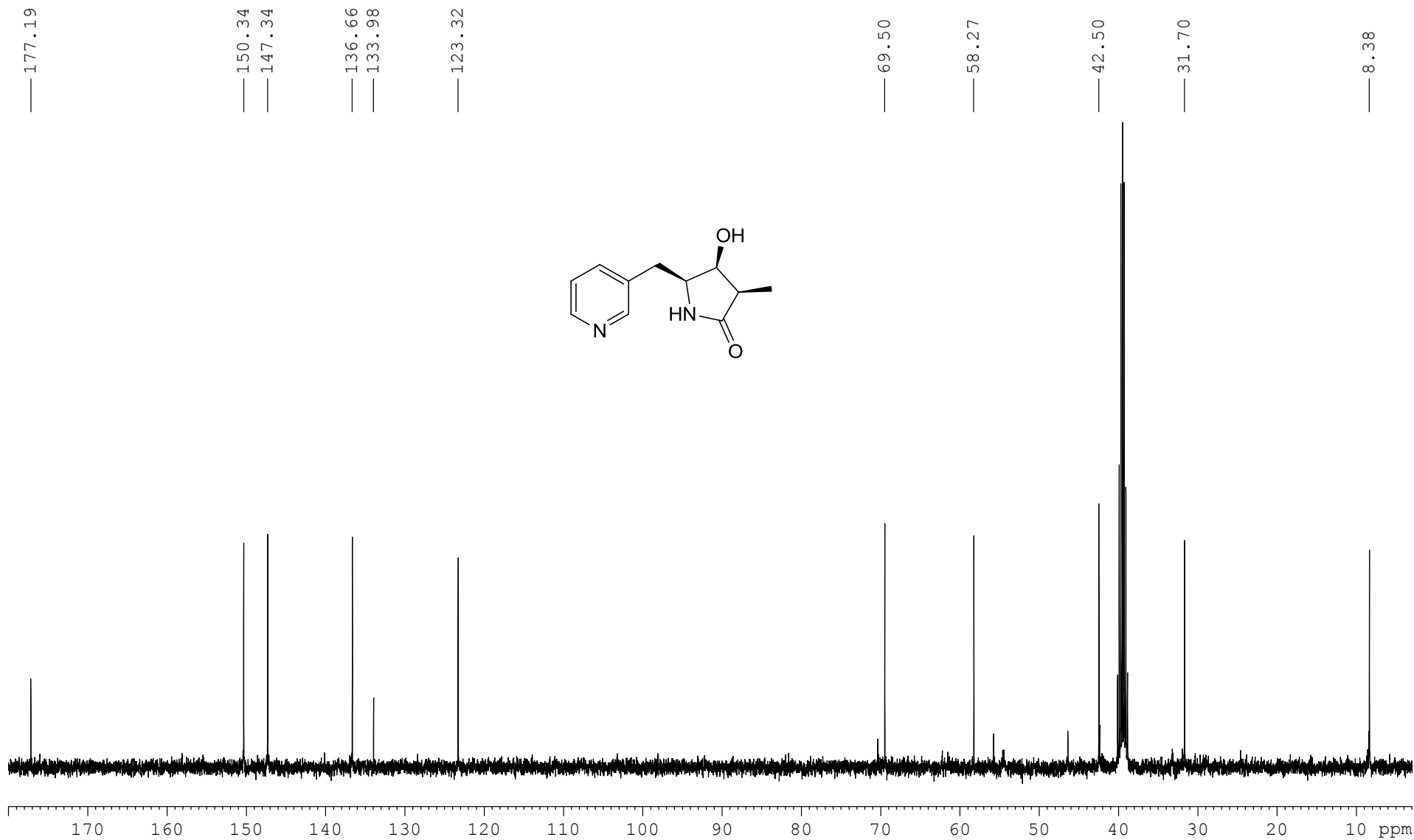


17

5

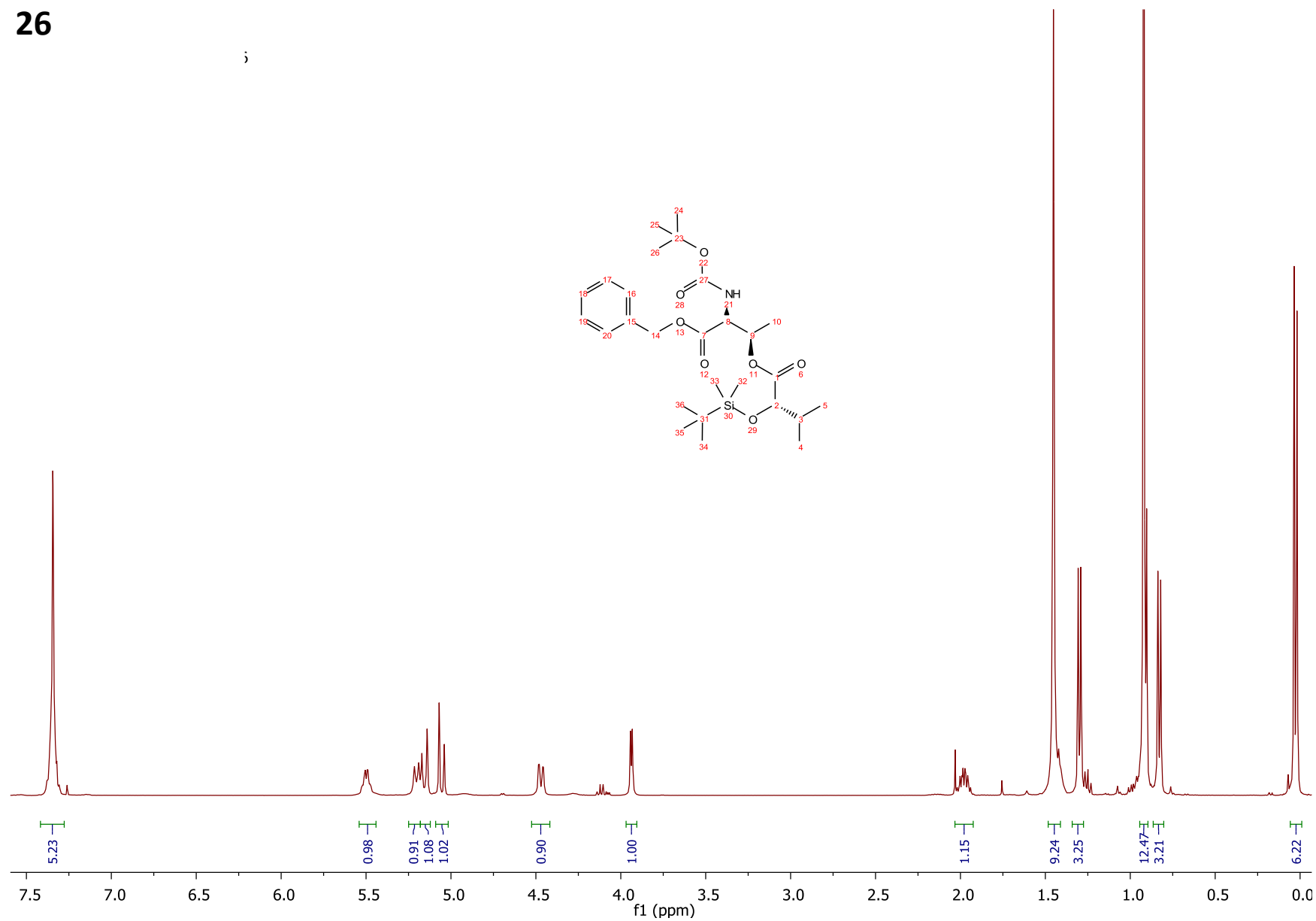


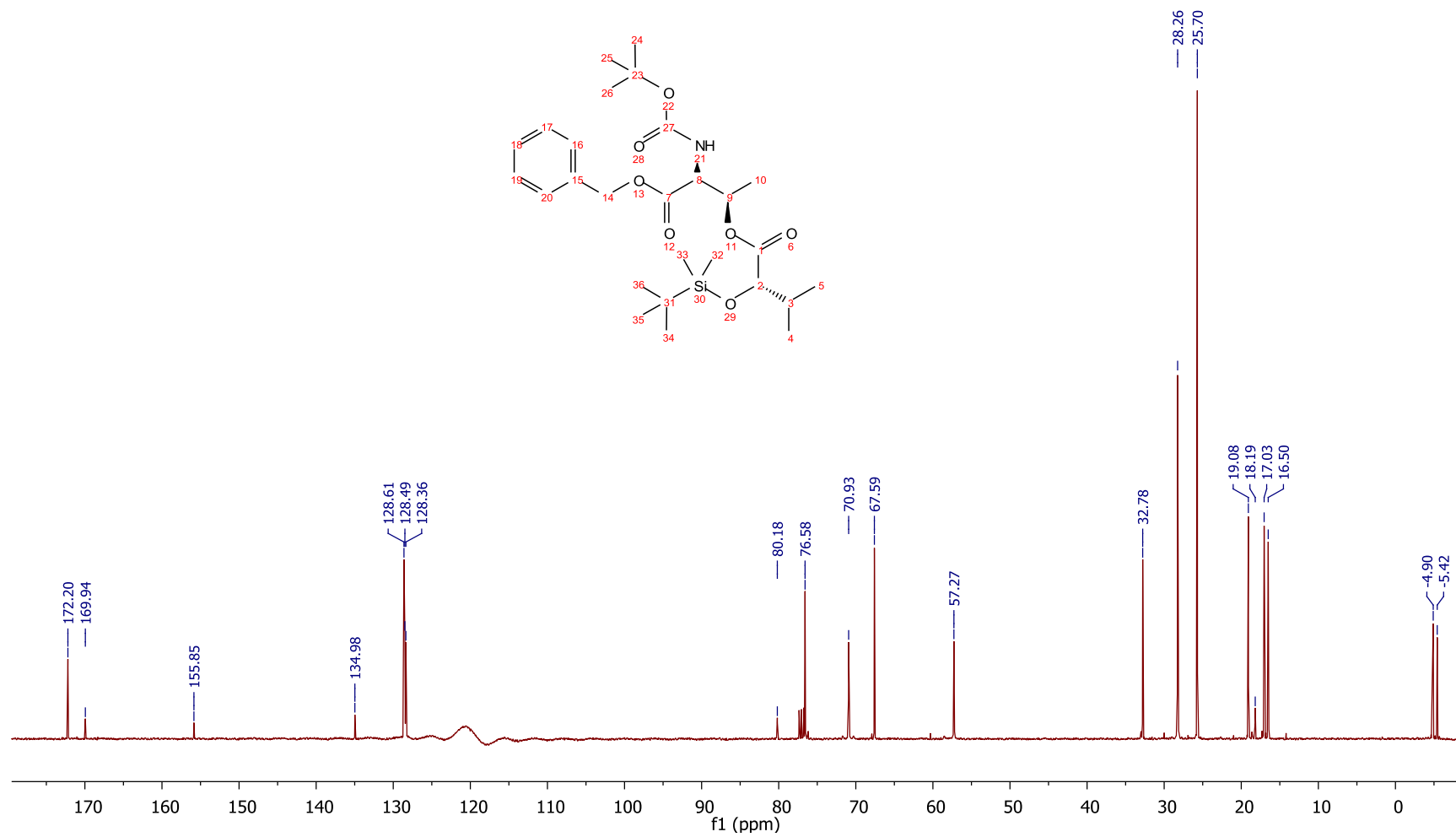
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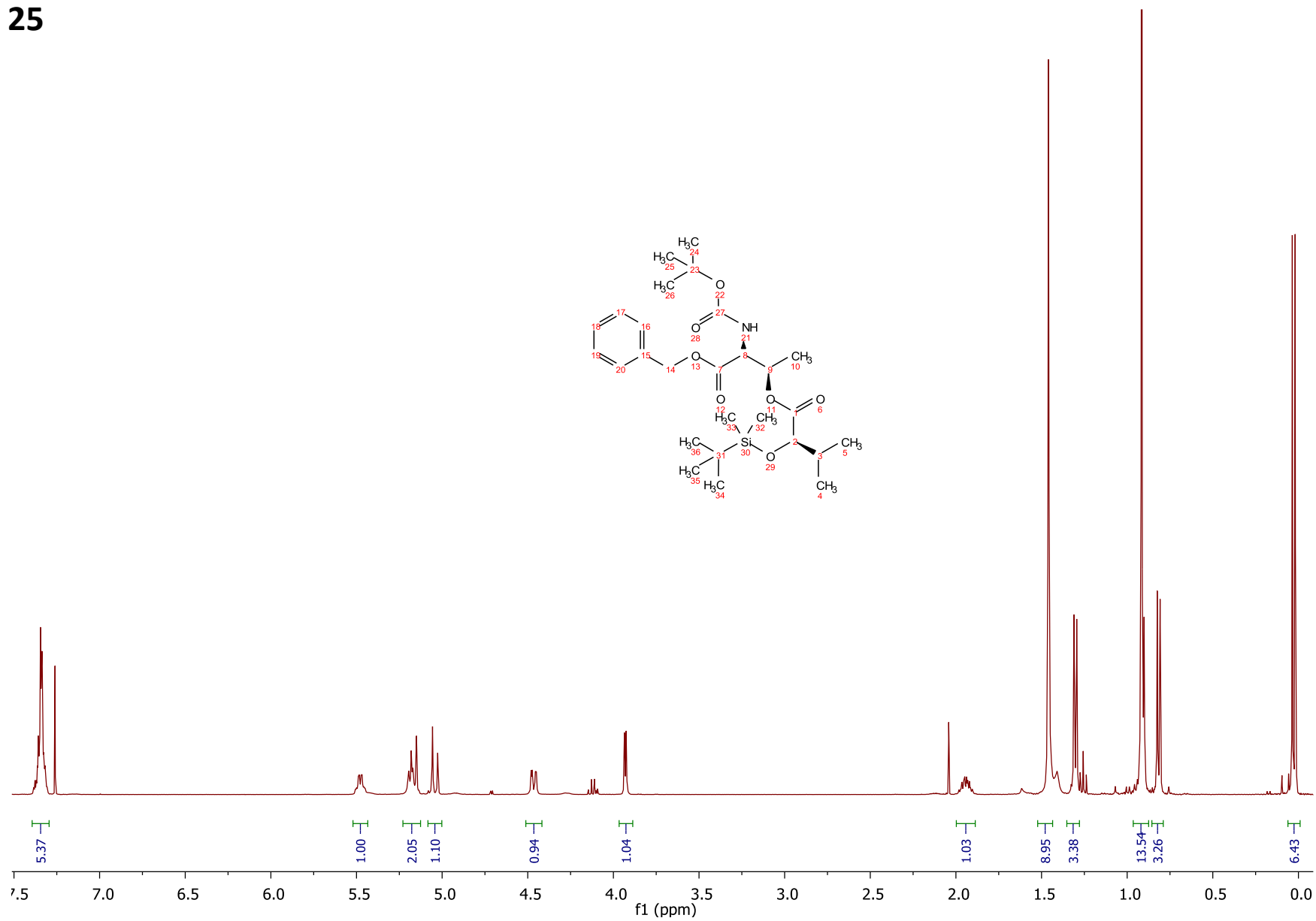
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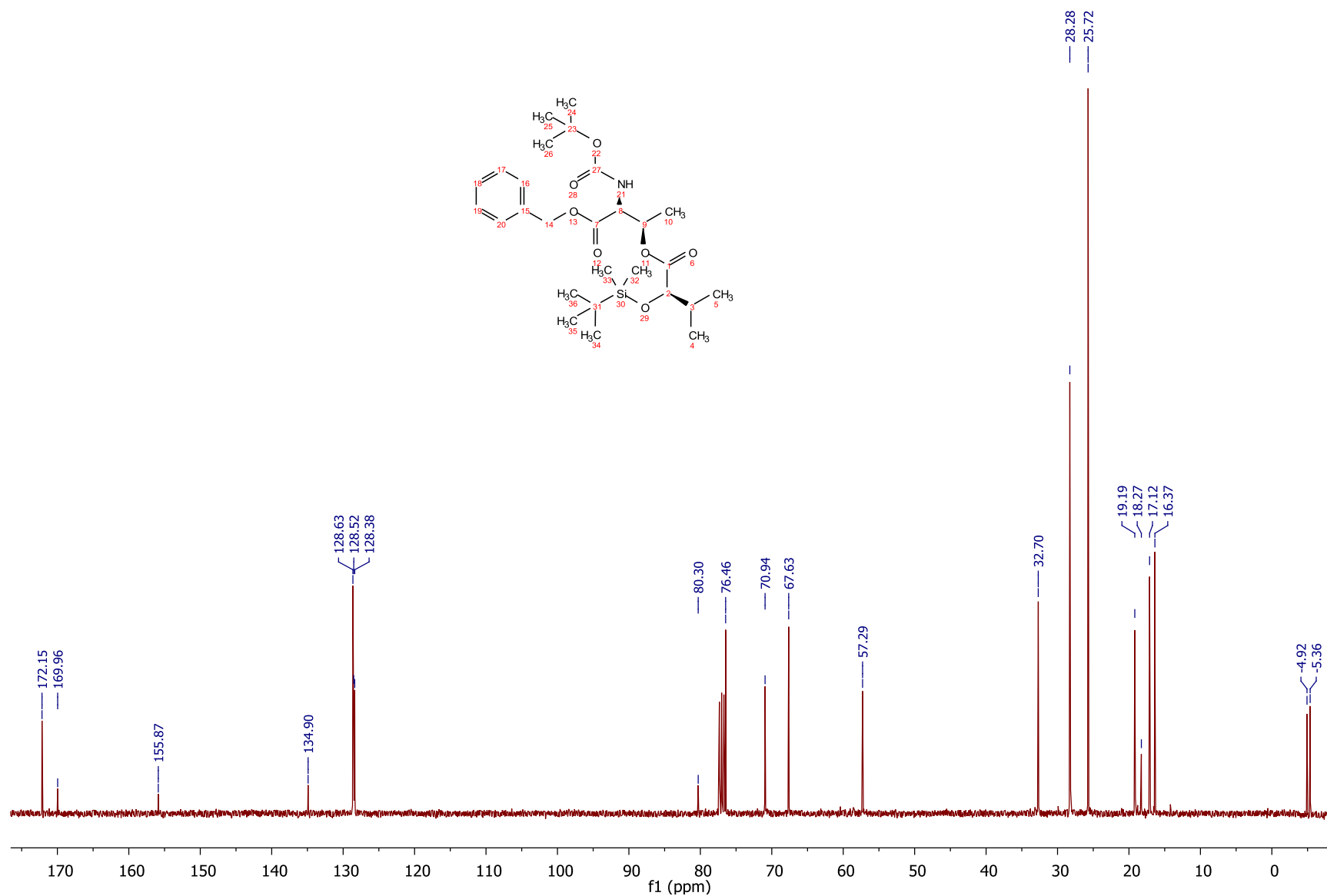




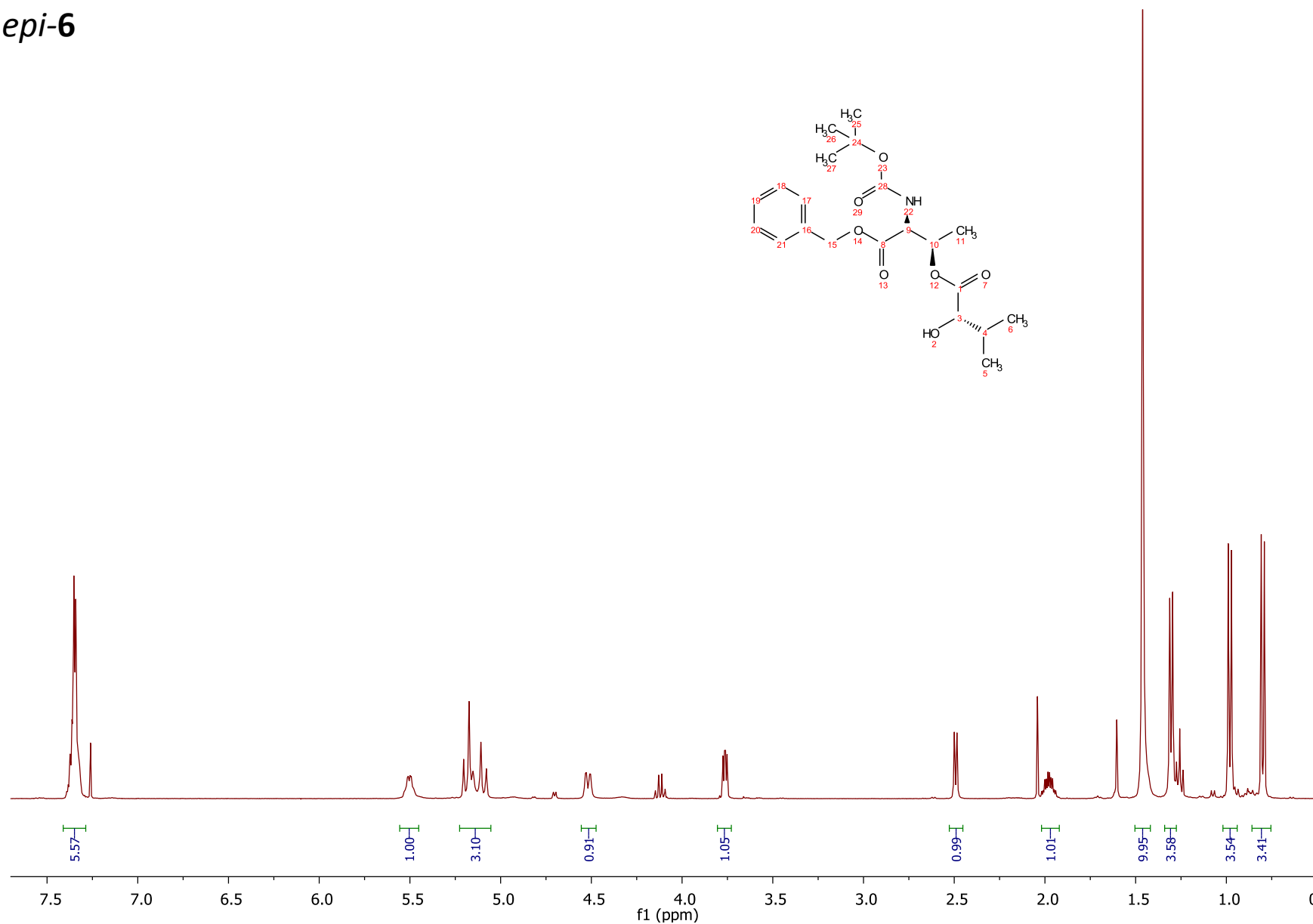
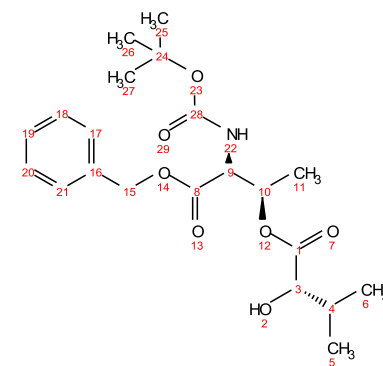
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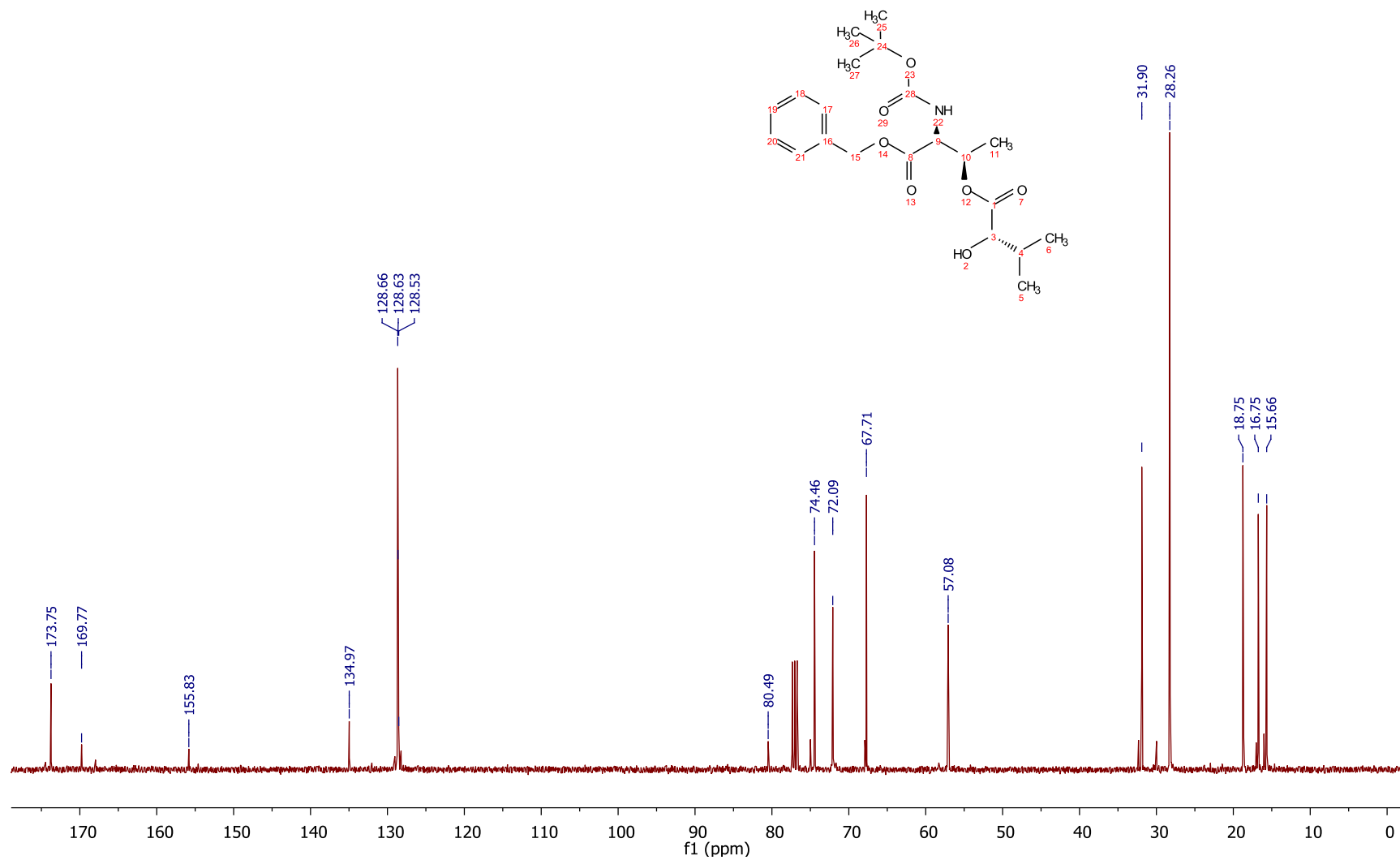
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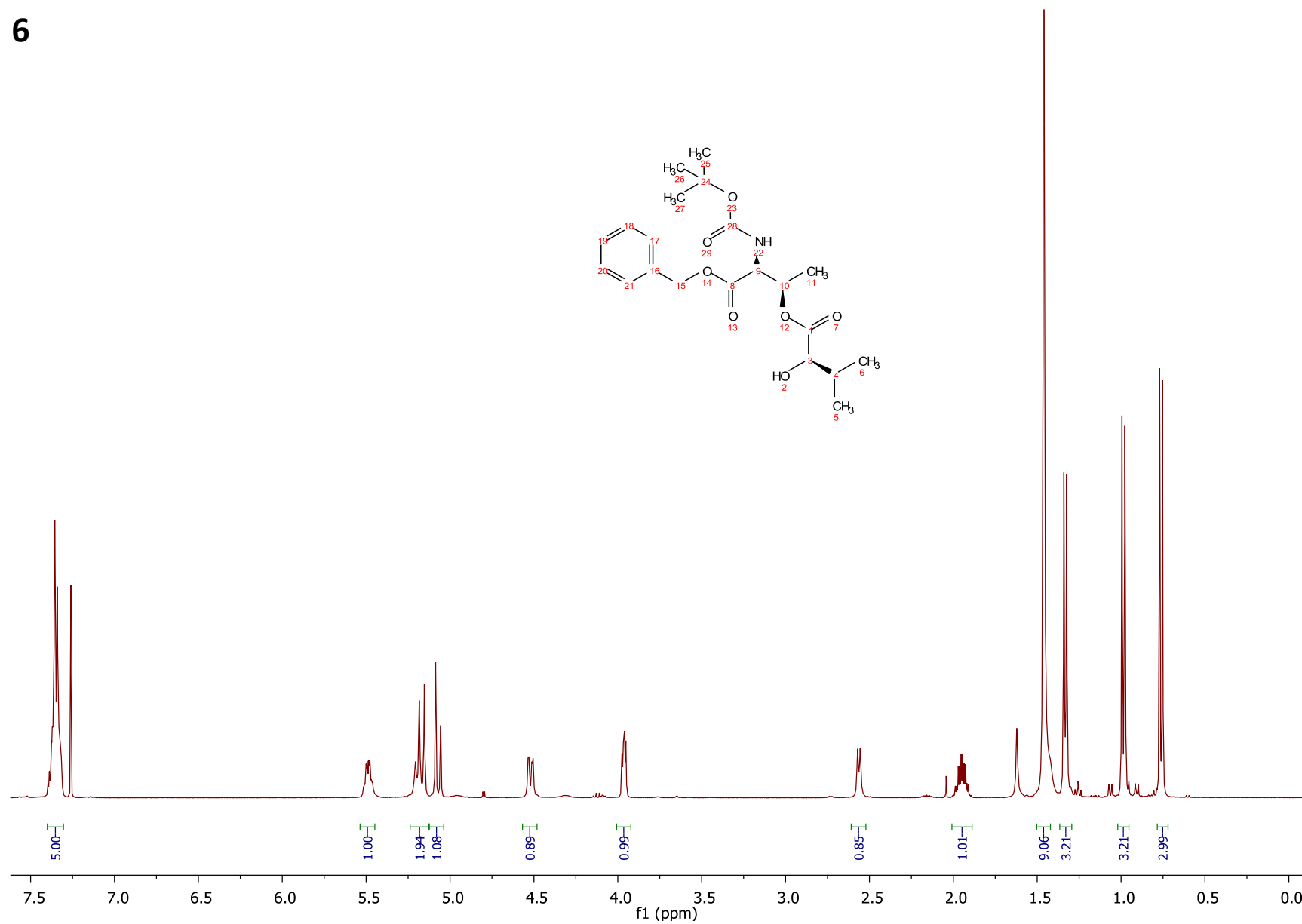
epi-6



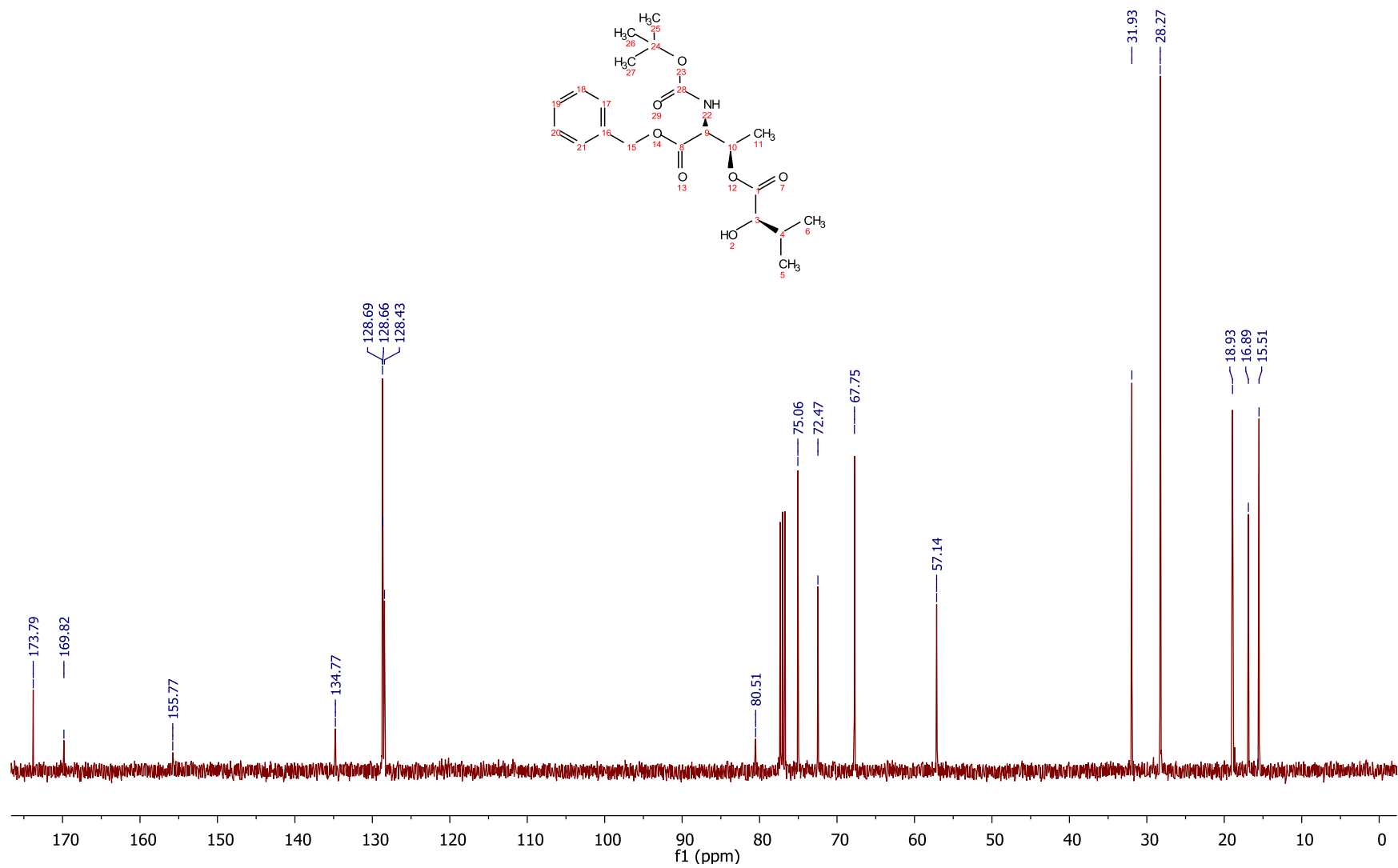
epi-6



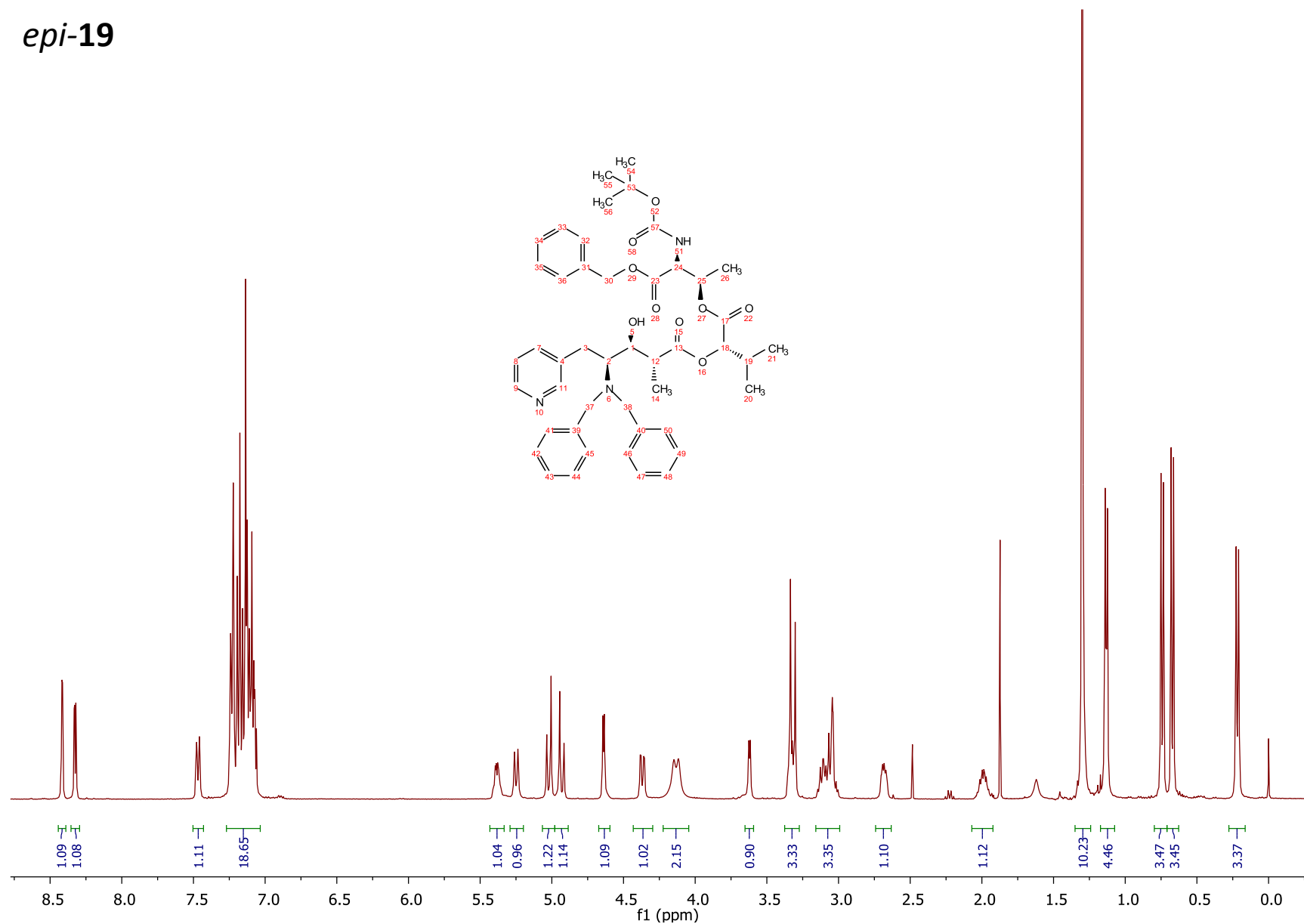
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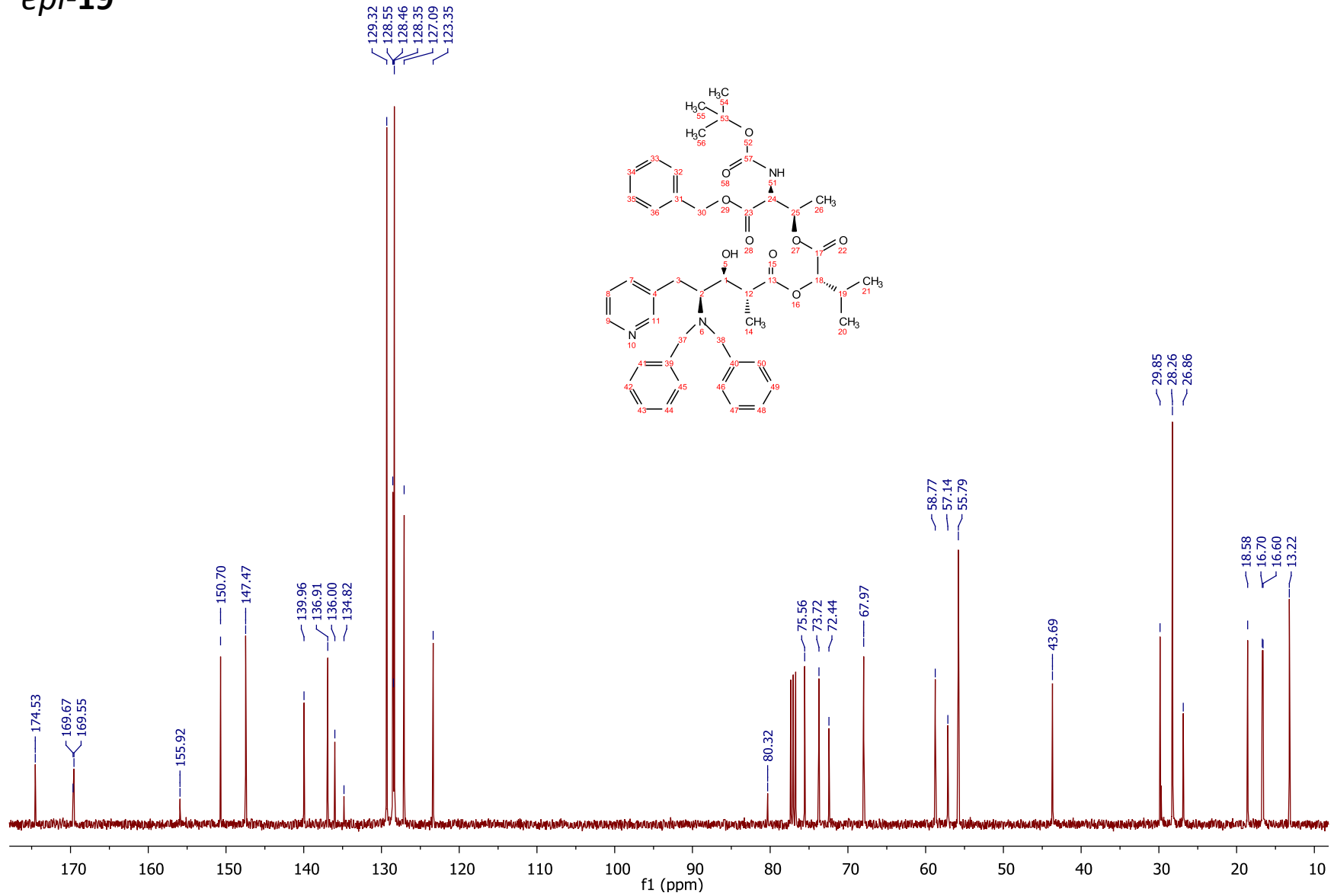
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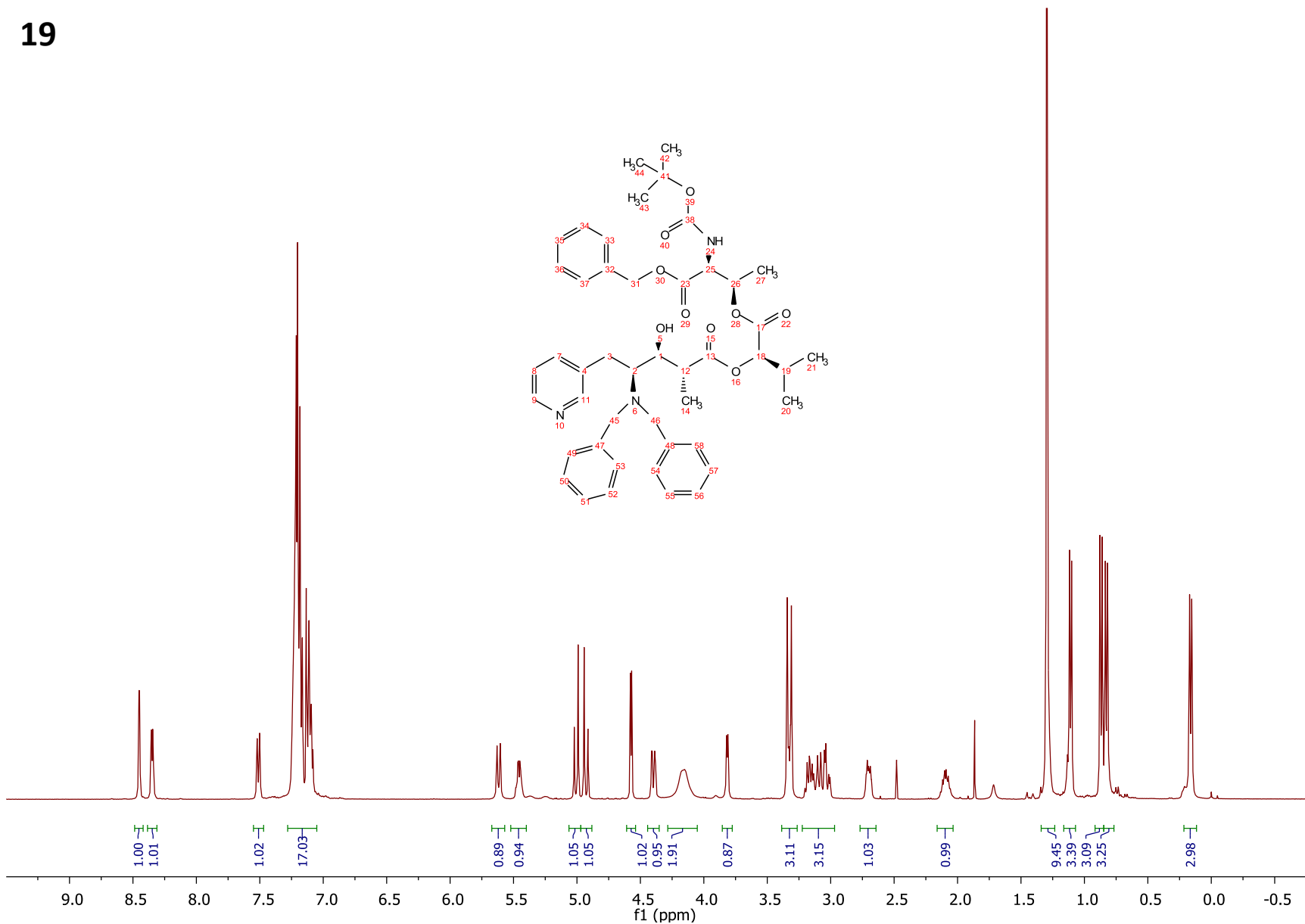
epi-19



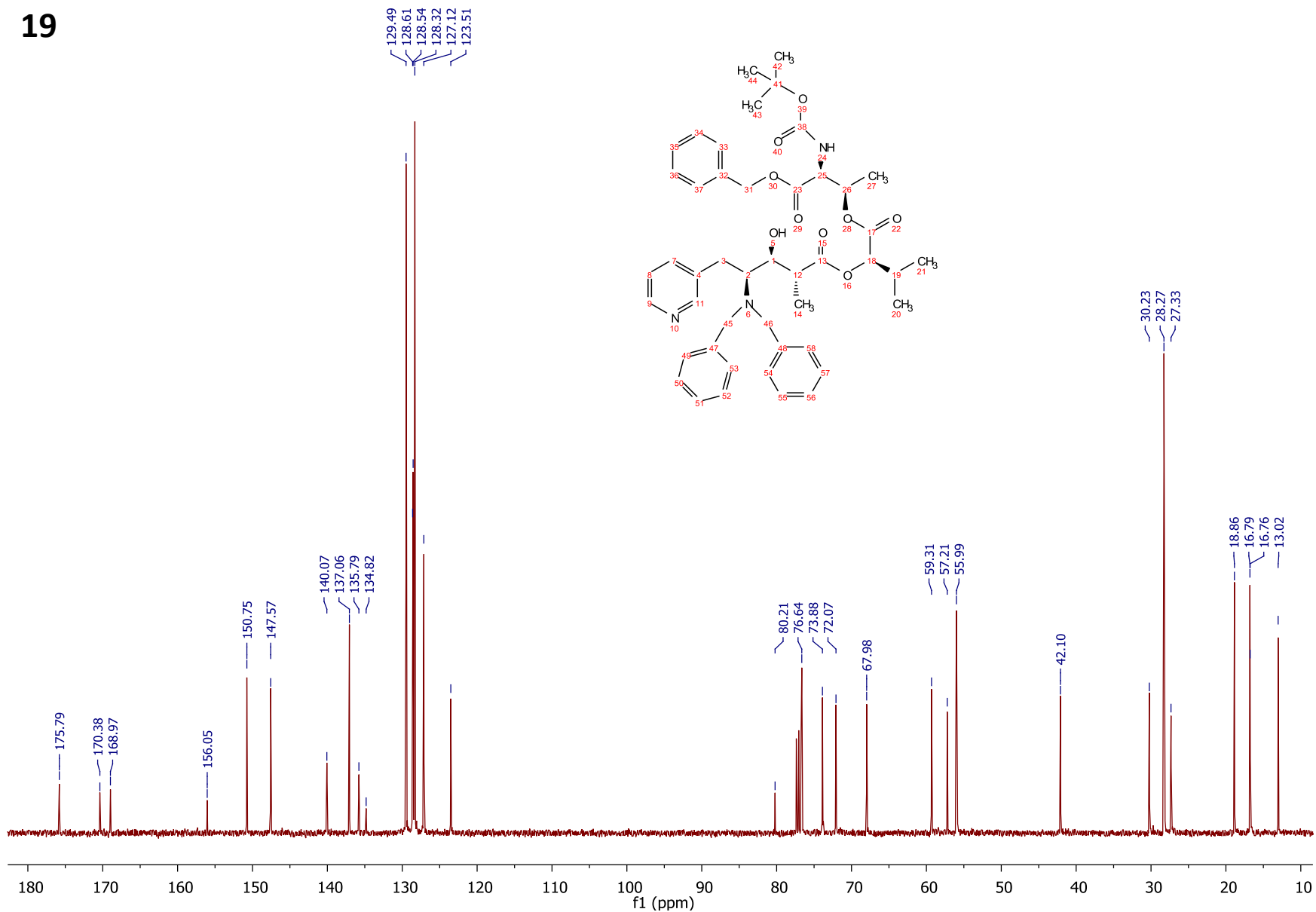
epi-19



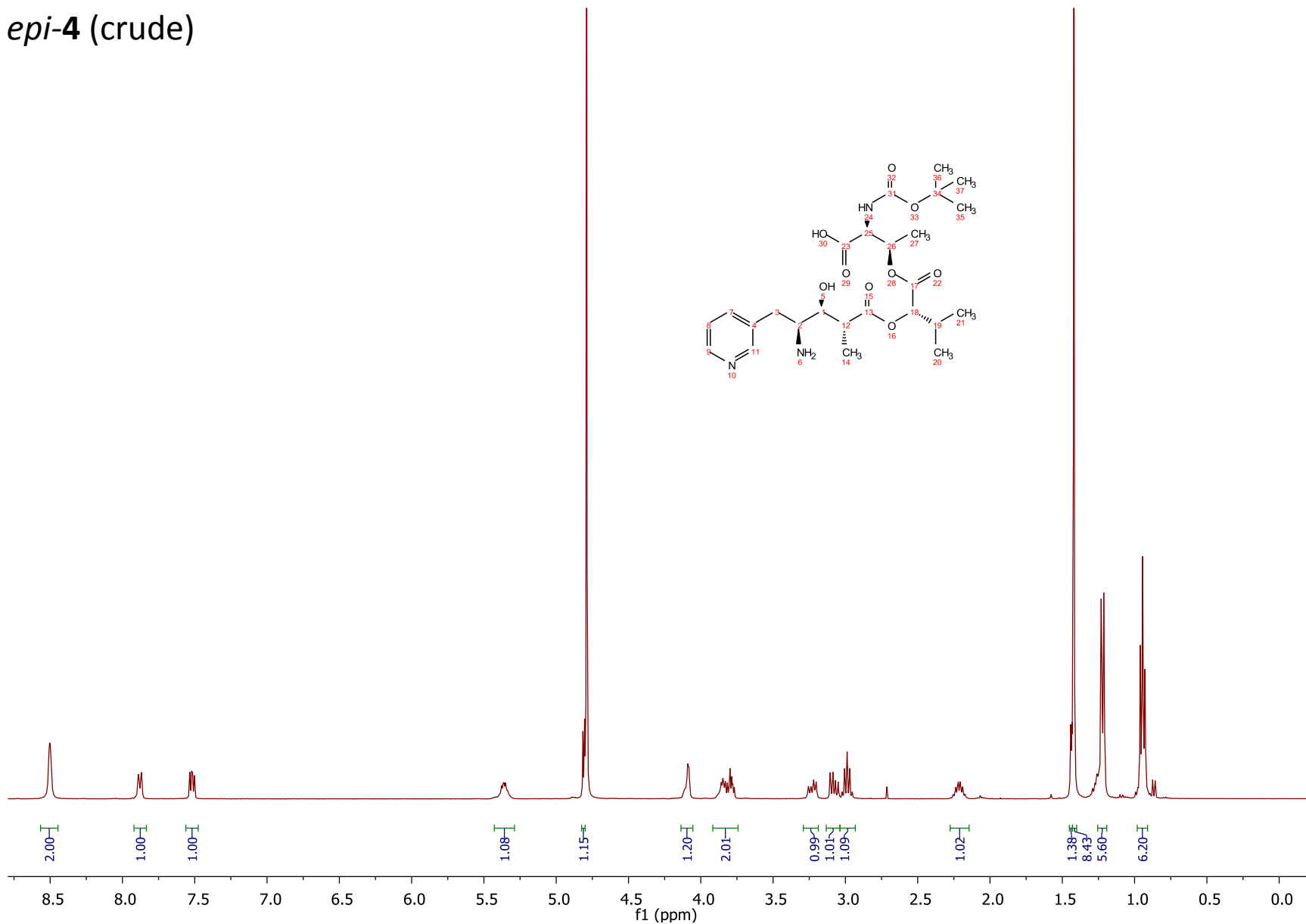
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19

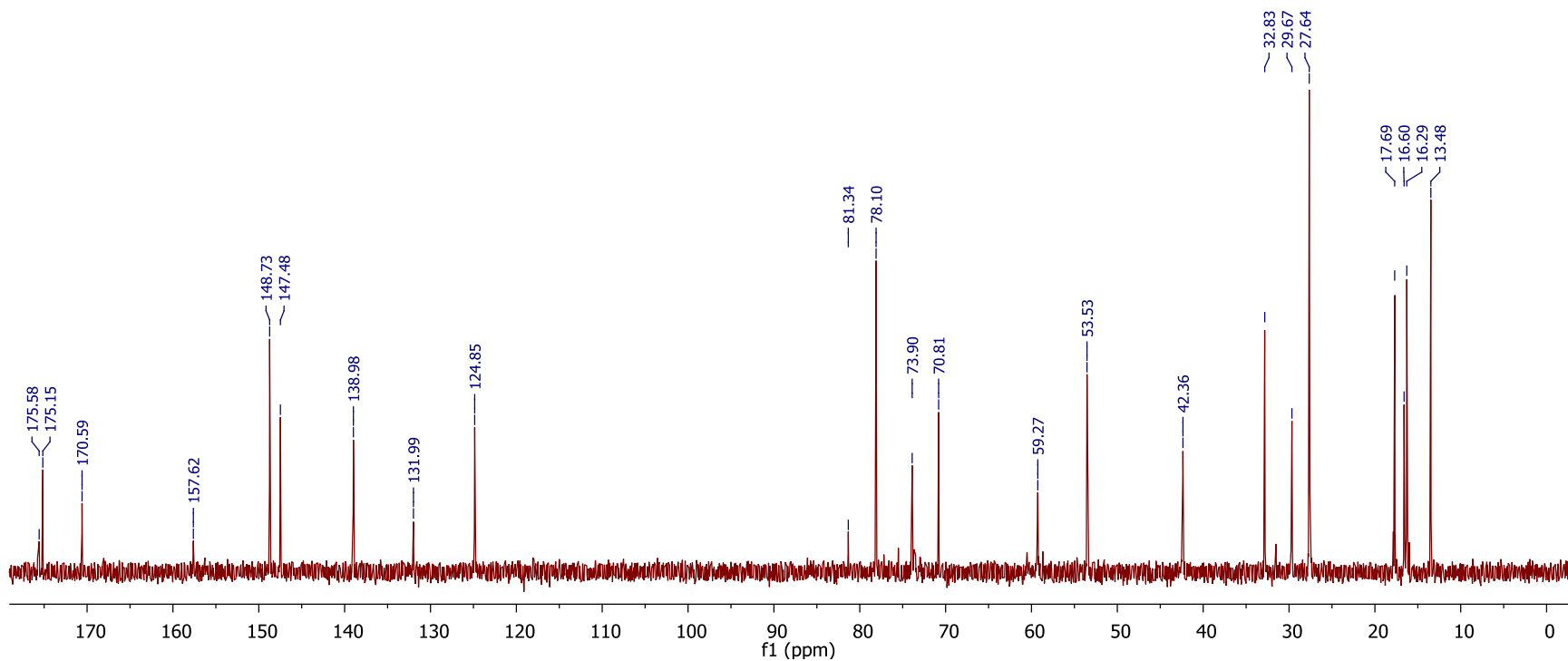
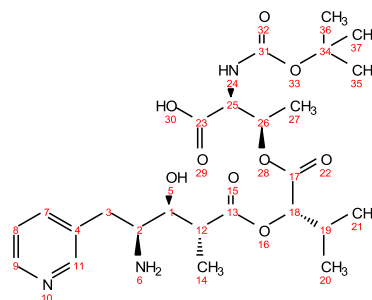


epi-4 (crude)

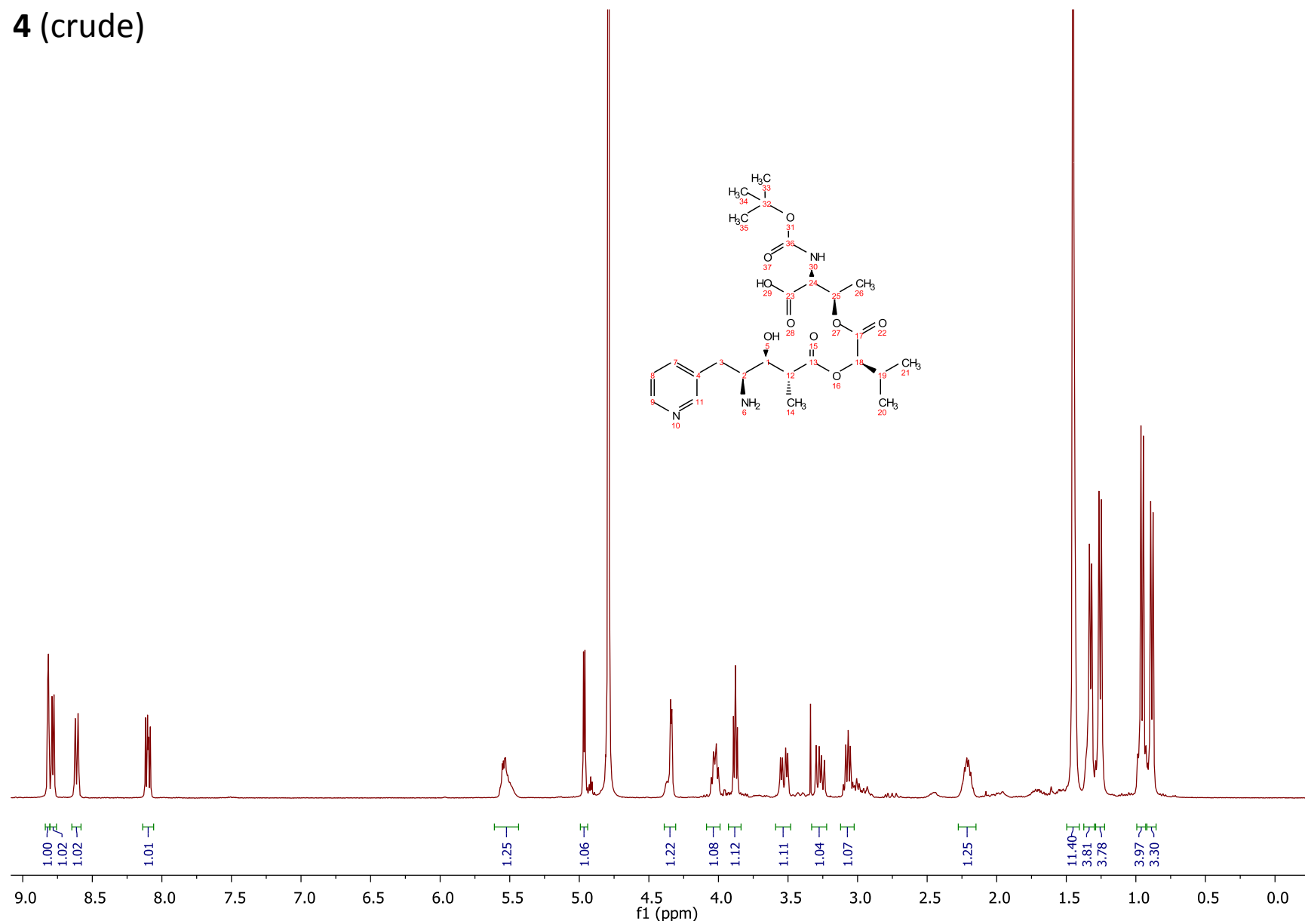


epi-4 (crude)

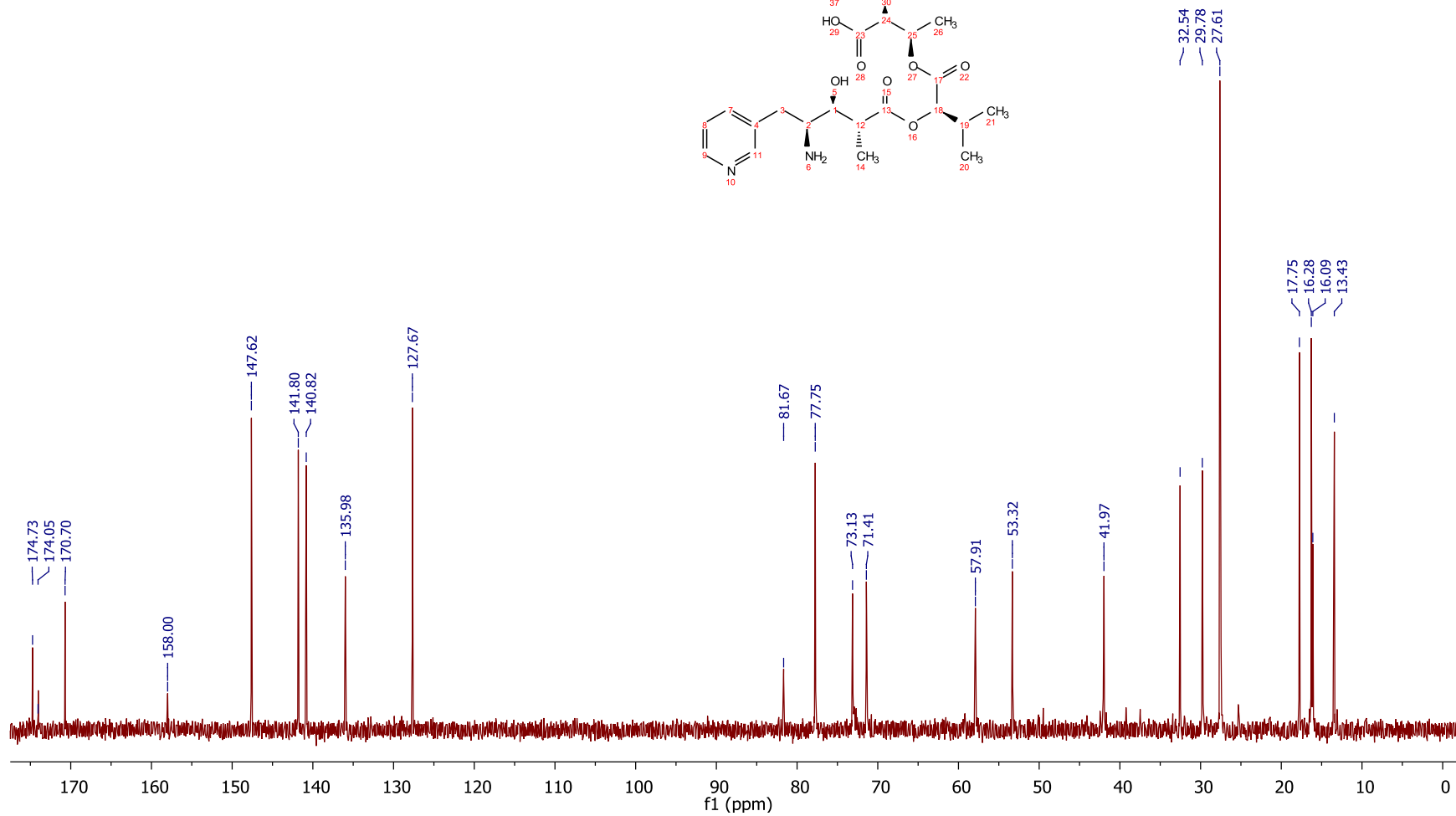
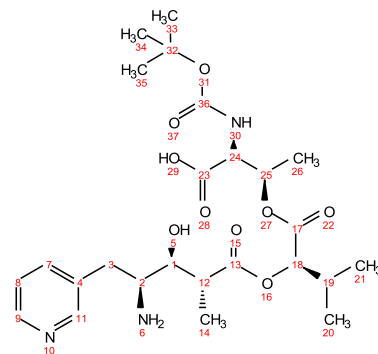
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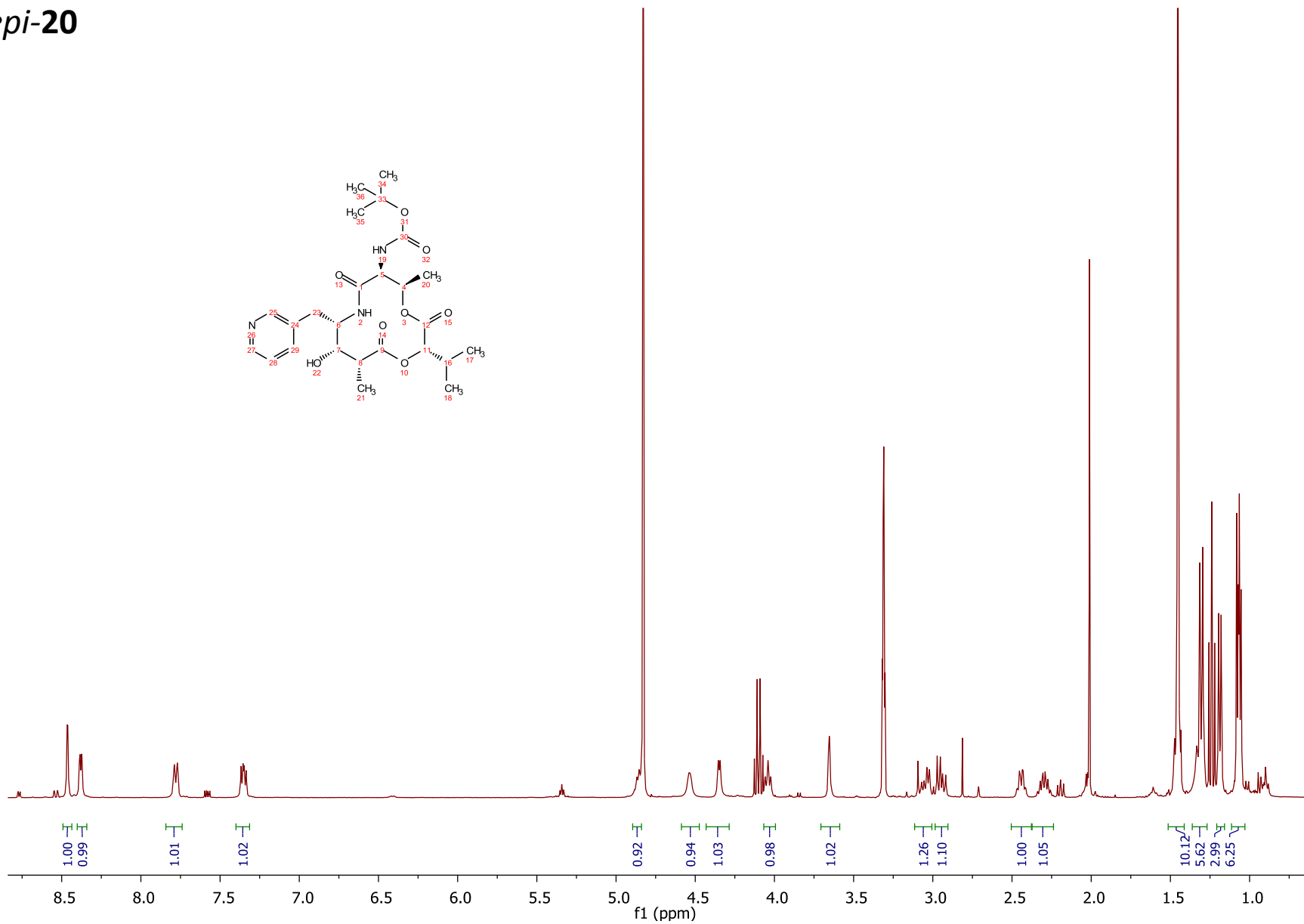
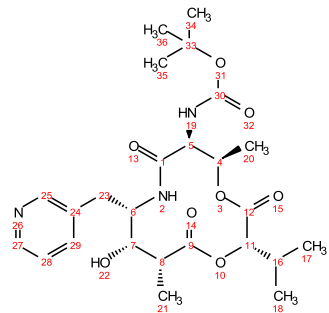
4 (crude)



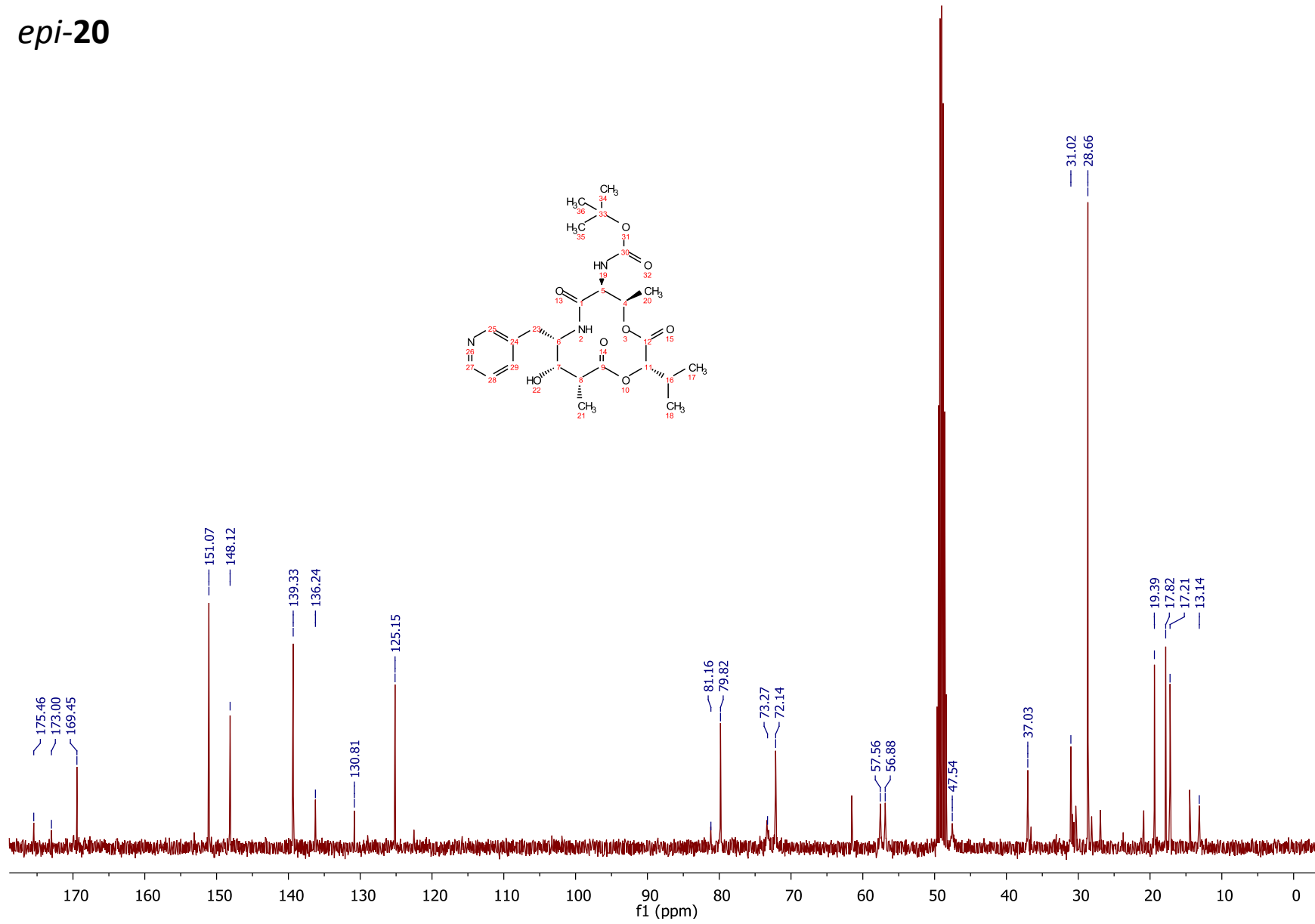
4 (crude)



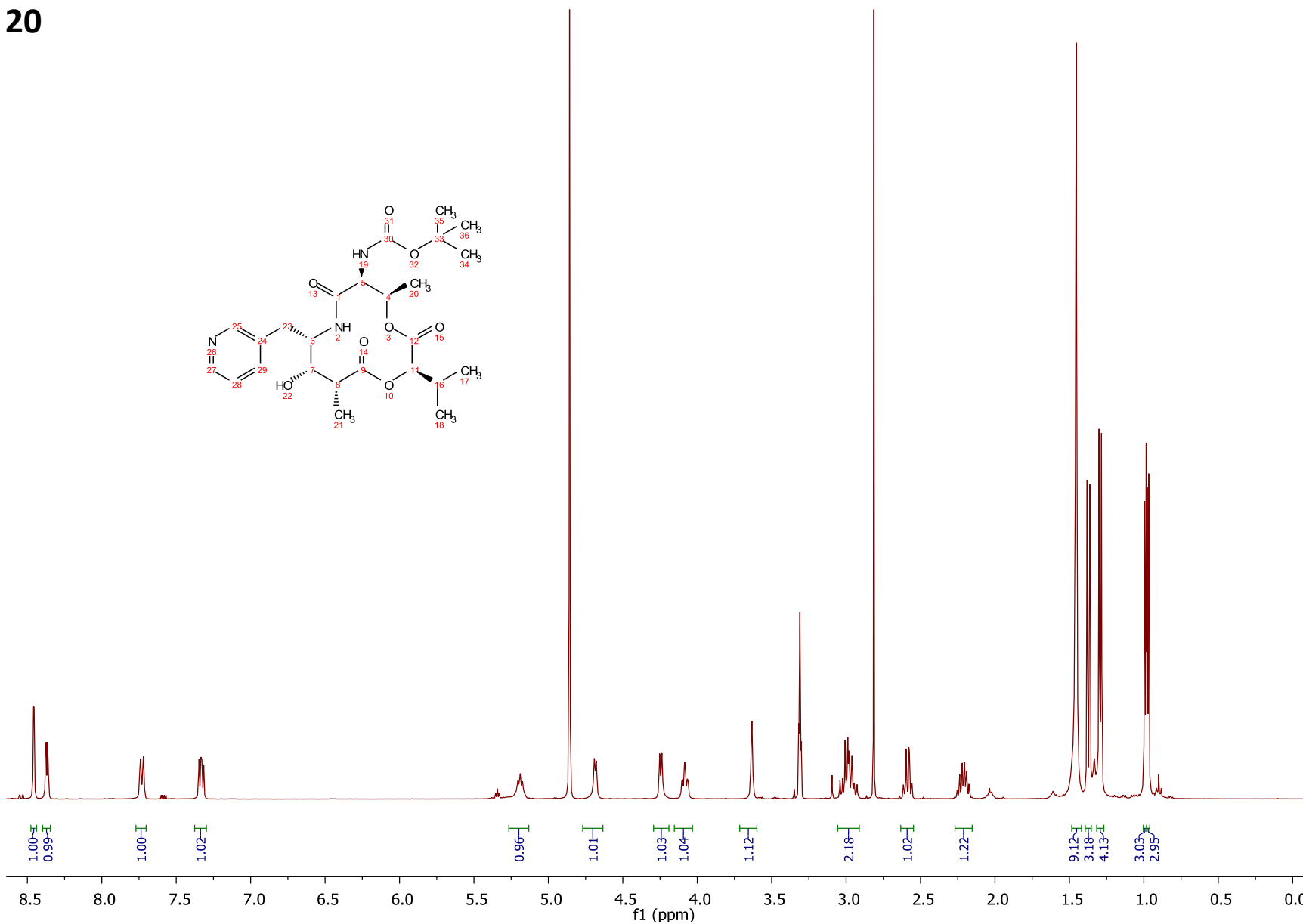
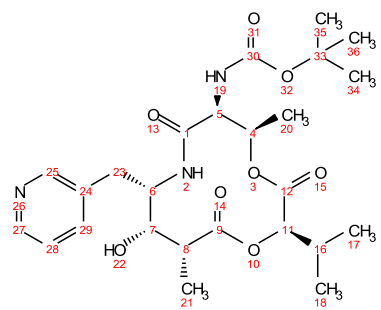
epi-20



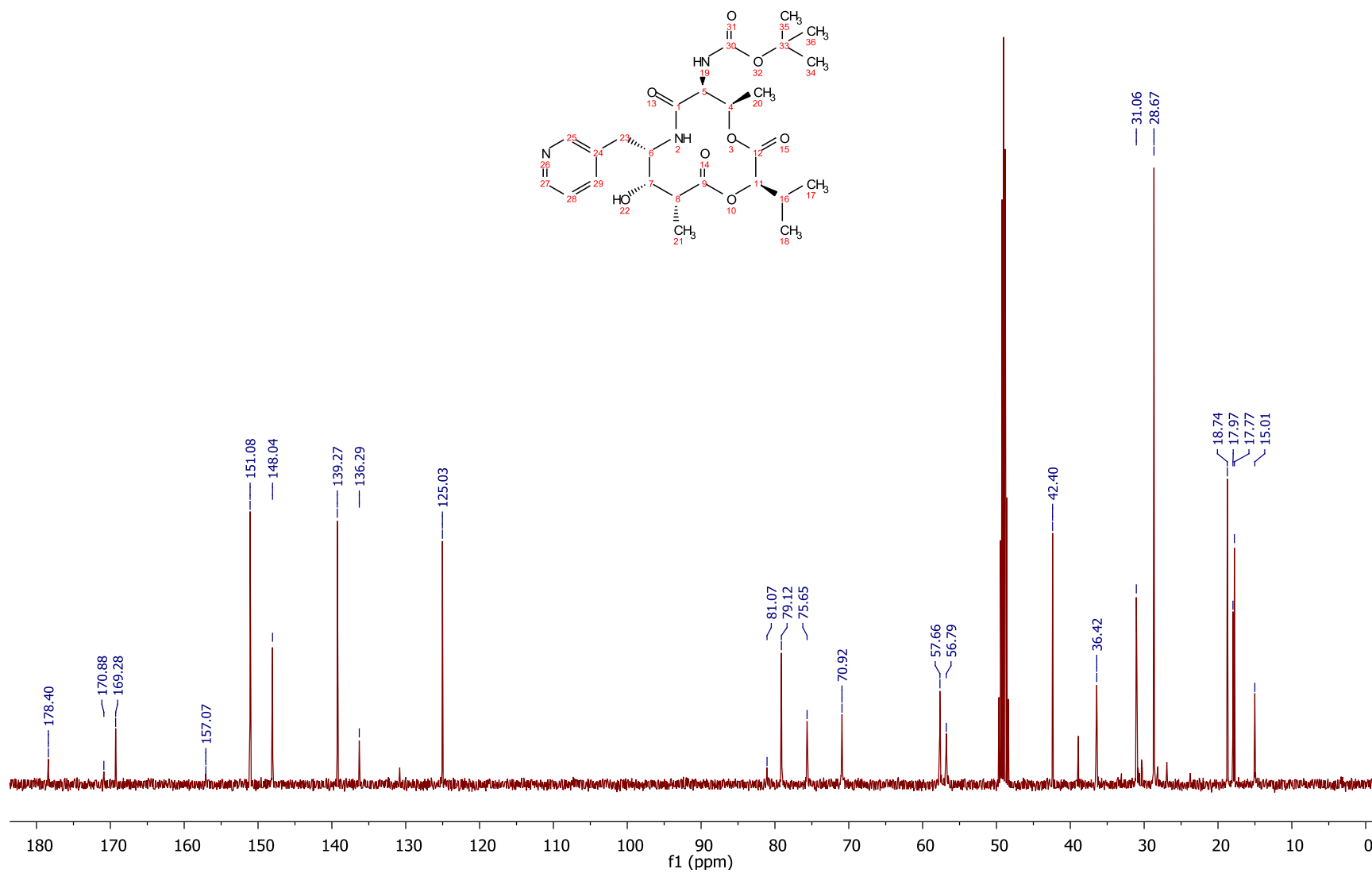
epi-20



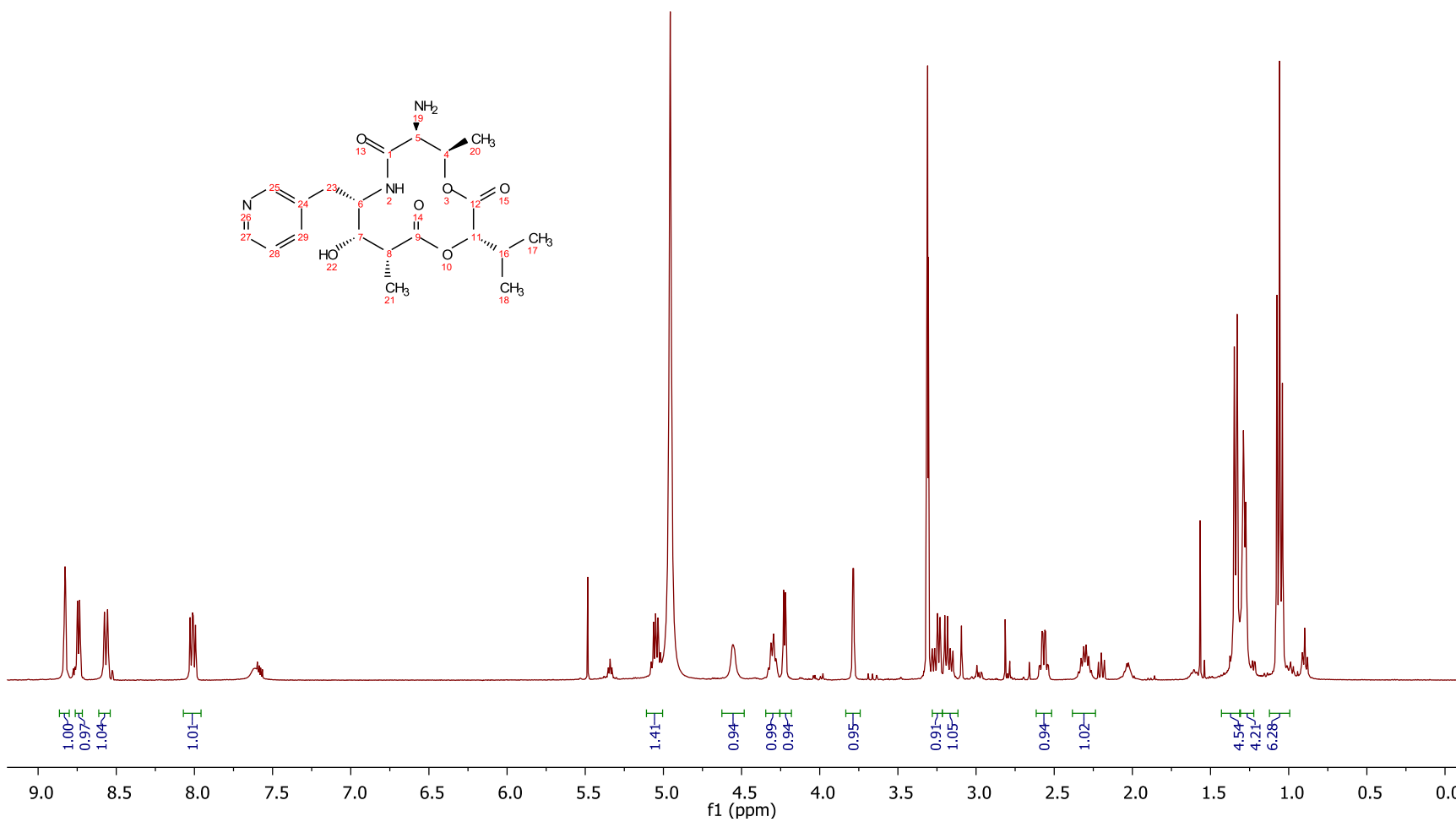
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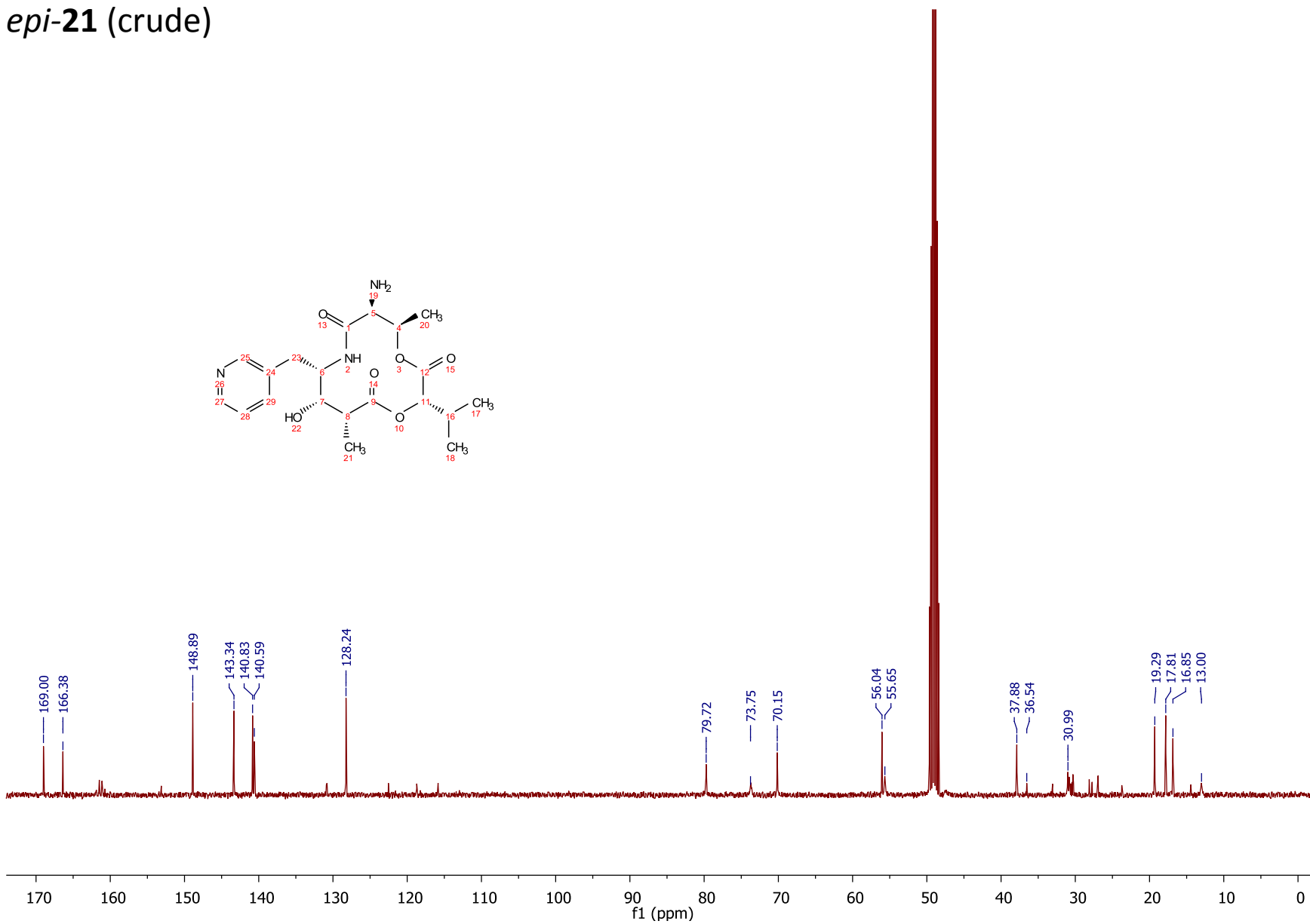
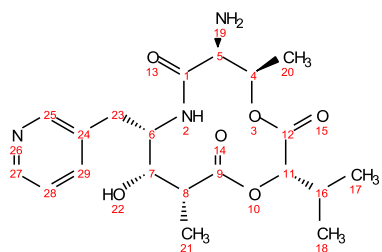
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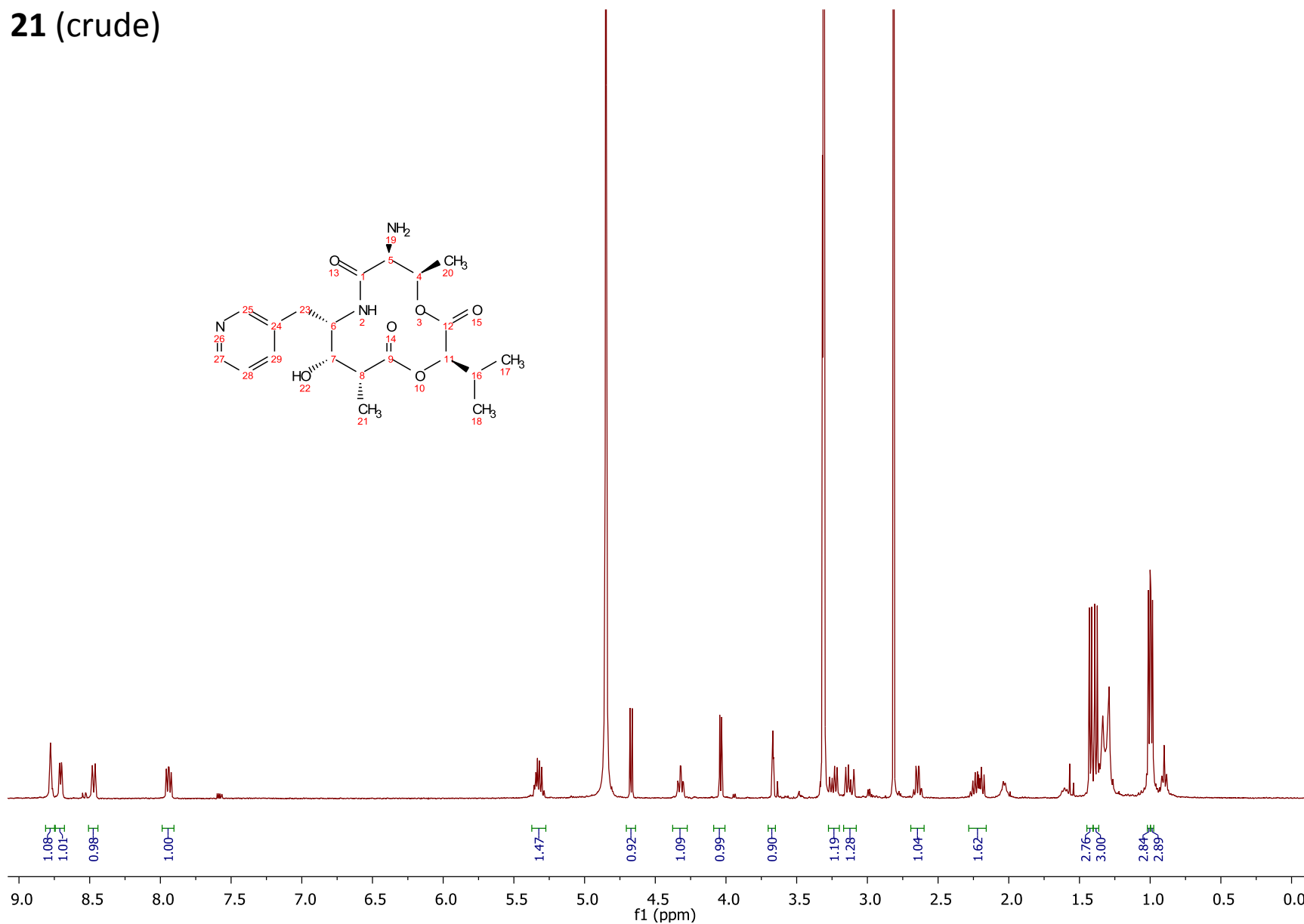
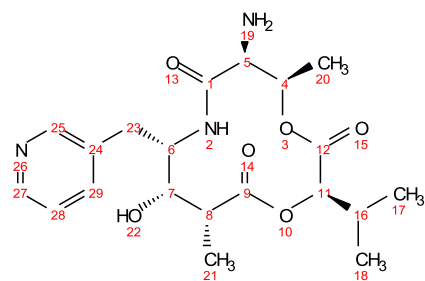
epi-21 (crude)



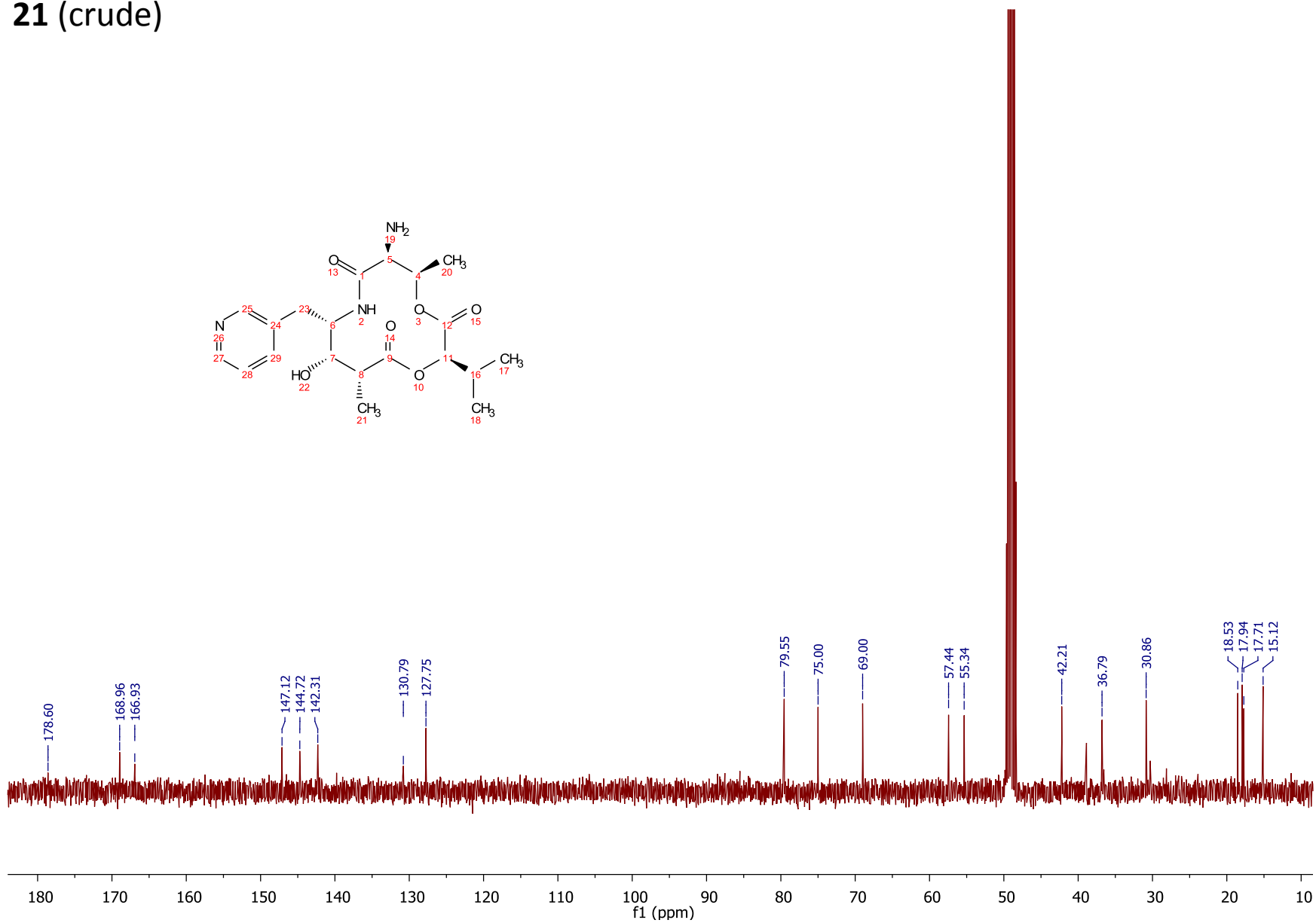
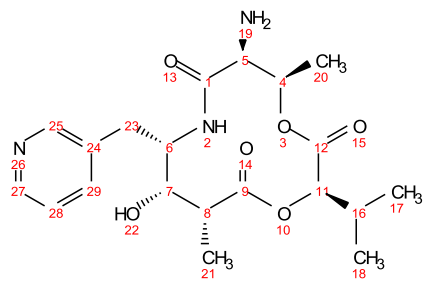
epi-21 (crude)



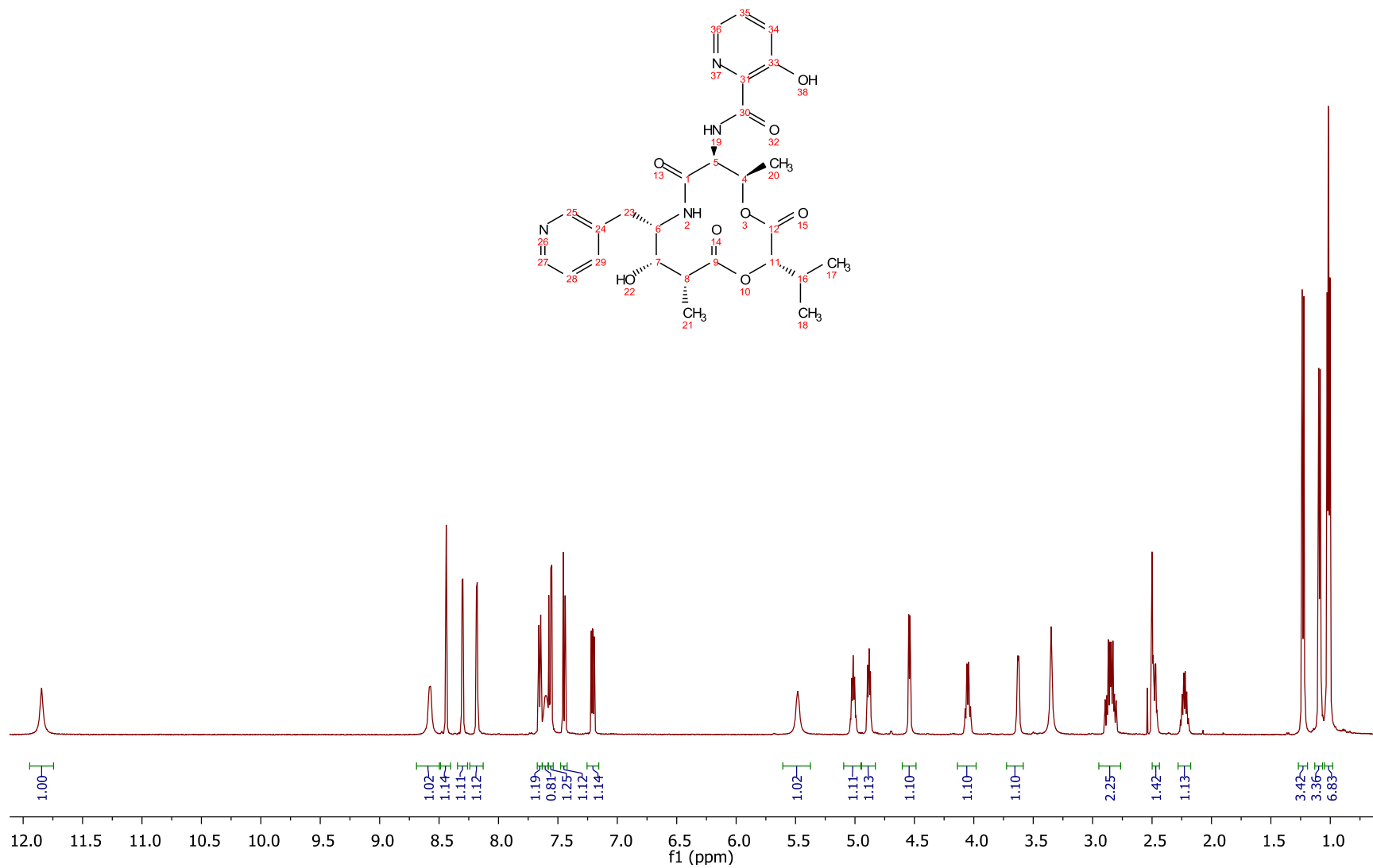
21 (crude)



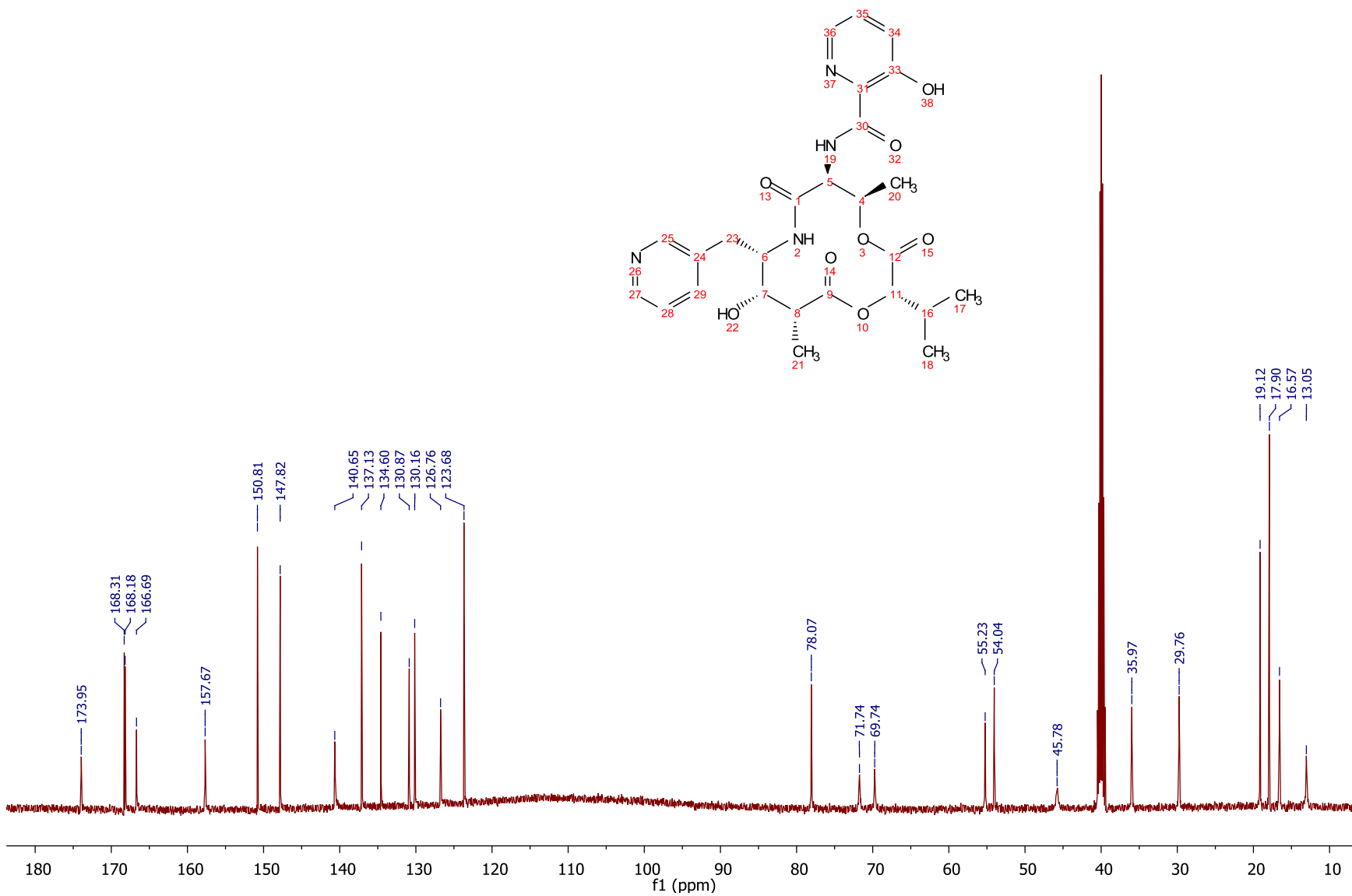
21 (crude)



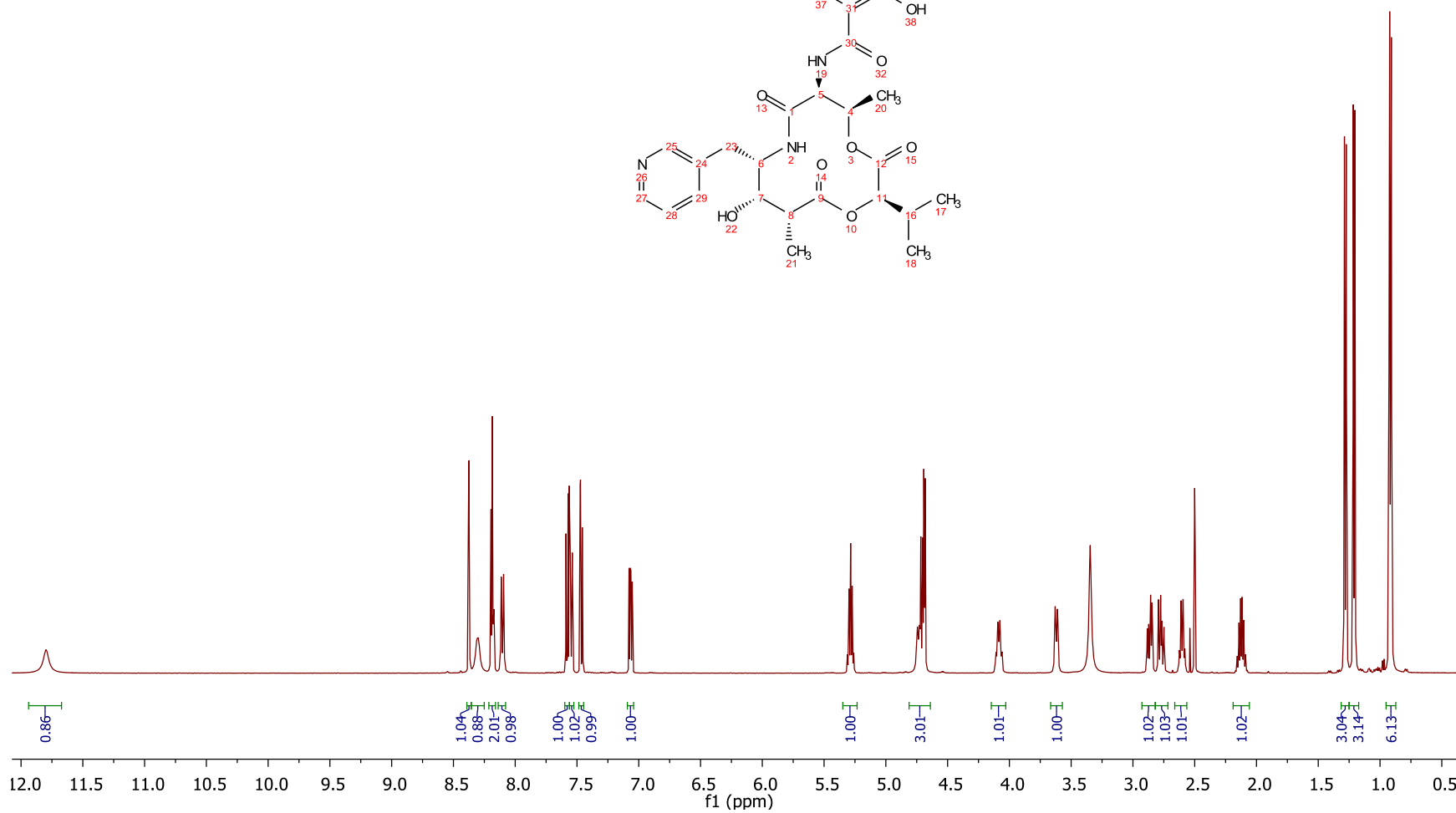
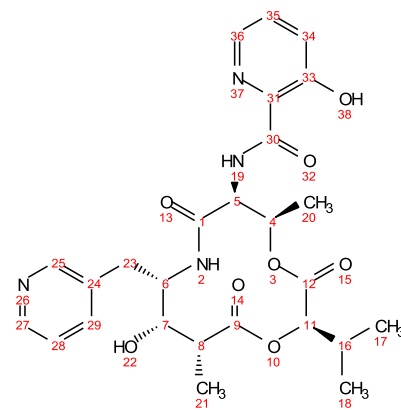
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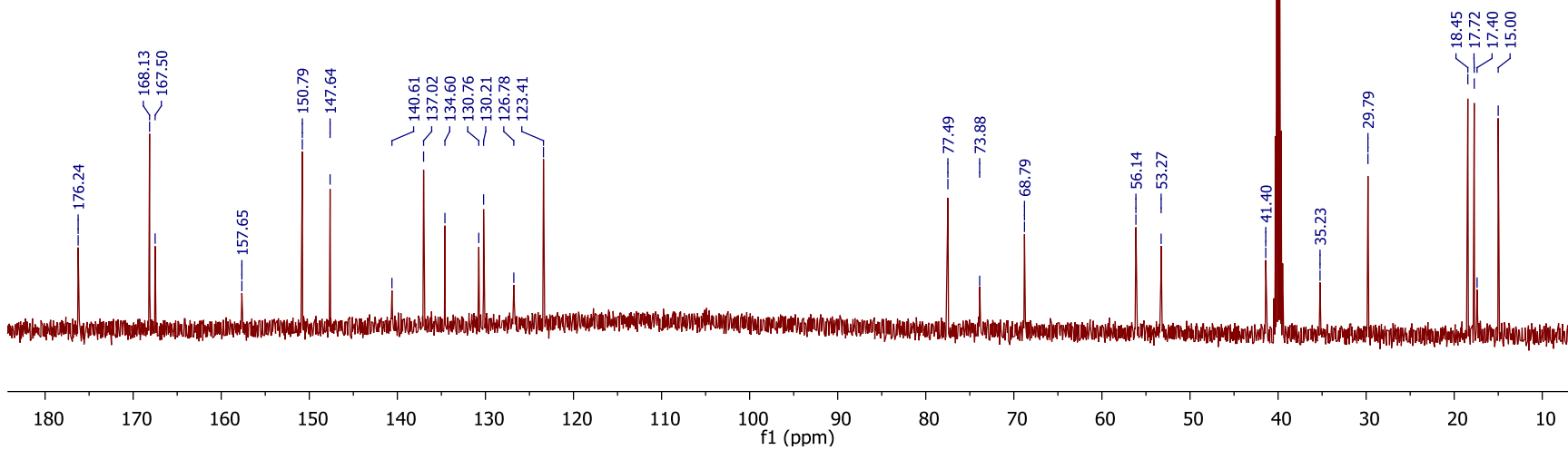
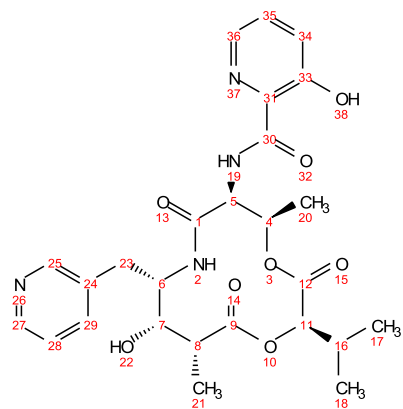
3



2



2



6. Biological Evaluation

Determination of the minimal inhibitory concentration (MIC) of 1, 2, and 3.^[1]

The drug susceptibility of *Mycobacterium tuberculosis* strain H37Rv was determined using the resazurin microtitre assay (REMA) (Palomino JC, Martin A, Camacho M, Guerra H, Swings J, Portaels F, Resazurin microtiter assay plate: simple and inexpensive method for detection of drug resistance in *Mycobacterium tuberculosis*, *Antimicrob Agents Chemother* **2002**, *46*, 2720-2722). Briefly, bacteria were diluted from frozen stocks to an OD₆₀₀ of 0.0001, and grown in a 96-well plate in the presence of serial compound dilutions. After 10 generations (7 days for *M. tuberculosis*) bacterial viability was determined using 10 µL of resazurin (0.025 % (w/v), and calculated as a percentage of resazurin turnover in the absence of compound. The MIC was determined as the minimal concentration of compound that caused background resazurin reduction

Steady state kinetics and inhibition of InhA.^[1]

Inhibition of InhA activity was investigated using InhA(S94A) at 60 nM. Kinetic parameters were determined by following NADH oxidation every min for 30 min at 340 nm using a TECAN FL200 spectrophotometer. All reactions were performed at 25°C in 30 mM PIPES (pH 6.5), 150 mM NaCl and 10% glycerol in a 384 well plate. After addition of variable concentrations of NADH, reactions were initiated by adding 2-trans-dodecenoyl-CoA to a final concentration of 40 µM (250 µM in one experiment). Steady state *K_m* for NADH was determined by measuring enzyme kinetics at different NADH concentrations (0 – 800 µM). NADH *K_m* and pyridomycin and analogs *K_i*'s were determined by measuring enzyme kinetics with both different NADH concentrations (33, 40, 50, 66, 75, 100, 150 µM) and different pyridomycin and analog concentrations (0, 1.25, 2.5, 5, 10, 20, 30 and 40 µg/mL). Firstly the rate of NADH oxidation was determined by linear regression of the 340 nm absorbance values (enzyme velocity) using Graphpad Prism. Then the data was analysed to determine NADH *k_m* and compound *K_i* values (competitive inhibition model).

[1] R. C. Hartkoorn, C. Sala, J. Neres, F. Pojer, S. Magnet, R. Mukherjee, S. Uplekar, S. Boy-Rottger, K.-H. Altmann, S. T. Cole, *EMBO Mol. Med.* **2012**, *4*, 1032-1042.

7. Crystallographic Data for 17 (CCDC 905295)

The crystal structure of compound **17** is deposited at the Cambridge Crystallographic Data Centre with the CCDC reference number 905295.

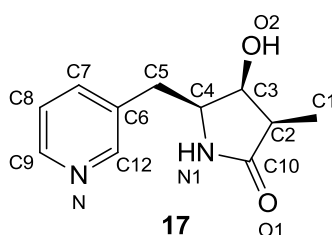
Table 1. Crystal data and structure refinement for **17** (905295).

Identification code	905295
Empirical formula	C ₁₁ H ₁₄ N ₂ O ₂

Formula weight	206.24
Temperature	100(2) K
Wavelength	1.54178 Å
Crystal system, space group	Orthorhombic, P2(1)2(1)2(1)
Unit cell dimensions	a = 4.7626(3) Å alpha = 90 deg. b = 8.9582(6) Å beta = 90 deg. c = 23.7884(19) Å gamma = 90 deg.
Volume	1014.92(12) Å ³
Z, Calculated density	4, 1.350 Mg/m ³
Absorption coefficient	0.769 mm ⁻¹
F(000)	440
Crystal size	0.13 x 0.02 x 0.01 mm
Theta range for data collection	3.72 to 67.18 deg.
Limiting indices	-3 <= h <= 5, -9 <= k <= 10, -26 <= l <= 27
Reflections collected / unique	5482 / 1686 [R(int) = 0.0858]
Completeness to theta	= 67.18 93.5 %
Absorption correction	None
Max. and min. transmission	0.9939 and 0.9086
Refinement method	Full-matrix least-squares on F ²
Data / restraints / parameters	1686 / 0 / 192
Goodness-of-fit on F ²	0.873
Final R indices [I > 2sigma(I)]	R1 = 0.0574, wR2 = 0.1438
R indices (all data)	R1 = 0.0850, wR2 = 0.1640
Absolute structure parameter	-0.2(6)
Largest diff. peak and hole	0.235 and -0.289 e.Å ⁻³

Table 2. Atomic coordinates ($\times 10^4$) and equivalent isotropic displacement parameters ($\text{Å}^2 \times 10^3$) for 905295.

U(eq) is defined as one third of the trace of the orthogonalized Uij tensor.



	x	y	z	U(eq)
O(1)	-5307(6)	-4179(3)	-2578(1)	39(1)
O(2)	-6768(5)	-3236(3)	-1022(1)	38(1)
N(1)	-7490(7)	-2169(4)	-2172(1)	32(1)
C(1)	-7597(11)	-5964(5)	-1654(2)	42(1)
C(2)	-8835(9)	-4477(5)	-1828(2)	32(1)
C(3)	-9203(9)	-3324(5)	-1367(2)	34(1)

C(4)	-9456(9)	-1872(5)	-1702(2)	32(1)
C(5)	-8598(9)	-453(5)	-1395(2)	36(1)
C(6)	-10607(9)	16(4)	-937(2)	32(1)
C(7)	-12521(9)	1173(4)	-1020(2)	35(1)
C(8)	-14268(9)	1614(5)	-581(2)	38(1)
C(9)	-14117(9)	877(5)	-80(2)	36(1)
C(10)	-7006(8)	-3628(4)	-2242(2)	33(1)
N	-12308(8)	-257(4)	15(1)	40(1)
C(12)	-10597(9)	-646(5)	-413(2)	37(1)

Table 3. Bond lengths [Å] and angles [deg] for 905295.

O(1)-C(10)	1.240(5)
O(2)-C(3)	1.422(5)
O(2)-H(2)	1.09(5)
N(1)-C(10)	1.338(5)
N(1)-C(4)	1.483(5)
N(1)-H(1)	0.97(5)
C(1)-C(2)	1.515(6)
C(1)-H(1A)	1.06(4)
C(1)-H(1B)	1.00(6)
C(1)-H(1C)	0.99(6)
C(2)-C(3)	1.518(6)
C(2)-C(10)	1.519(5)
C(2)-H(2A)	1.05(4)
C(3)-C(4)	1.531(6)
C(3)-H(3)	1.04(4)
C(4)-C(5)	1.522(6)
C(4)-H(4)	1.05(4)
C(5)-C(6)	1.510(5)
C(5)-H(5A)	1.01(4)
C(5)-H(5B)	0.99(5)
C(6)-C(12)	1.379(6)
C(6)-C(7)	1.395(6)
C(7)-C(8)	1.392(6)
C(7)-H(7)	0.94(4)
C(8)-C(9)	1.363(6)
C(8)-H(8)	0.98(6)
C(9)-N	1.351(6)
C(9)-H(9)	1.00(5)
N-C(12)	1.351(5)
C(12)-H(12)	0.98(5)
C(3)-O(2)-H(2)	114(3)
C(10)-N(1)-C(4)	112.2(4)
C(10)-N(1)-H(1)	130(3)
C(4)-N(1)-H(1)	117(3)
C(2)-C(1)-H(1A)	117(2)
C(2)-C(1)-H(1B)	111(3)

H(1A)-C(1)-H(1B)	105(4)
C(2)-C(1)-H(1C)	109(3)
H(1A)-C(1)-H(1C)	107(4)
H(1B)-C(1)-H(1C)	108(4)
C(1)-C(2)-C(3)	116.4(3)
C(1)-C(2)-C(10)	113.3(3)
C(3)-C(2)-C(10)	101.2(3)
C(1)-C(2)-H(2A)	115(2)
C(3)-C(2)-H(2A)	106(2)
C(10)-C(2)-H(2A)	104(2)
O(2)-C(3)-C(2)	111.2(3)
O(2)-C(3)-C(4)	108.5(3)
C(2)-C(3)-C(4)	102.1(3)
O(2)-C(3)-H(3)	108(2)
C(2)-C(3)-H(3)	116(2)
C(4)-C(3)-H(3)	111(2)
N(1)-C(4)-C(5)	110.0(3)
N(1)-C(4)-C(3)	101.0(3)
C(5)-C(4)-C(3)	116.0(4)
N(1)-C(4)-H(4)	114(2)
C(5)-C(4)-H(4)	106(2)
C(3)-C(4)-H(4)	110(2)
C(6)-C(5)-C(4)	114.1(3)
C(6)-C(5)-H(5A)	110(2)
C(4)-C(5)-H(5A)	108(2)
C(6)-C(5)-H(5B)	112(2)
C(4)-C(5)-H(5B)	107(3)
H(5A)-C(5)-H(5B)	105(3)
C(12)-C(6)-C(7)	116.7(4)
C(12)-C(6)-C(5)	122.0(4)
C(7)-C(6)-C(5)	121.2(4)
C(8)-C(7)-C(6)	119.7(4)
C(8)-C(7)-H(7)	120(2)
C(6)-C(7)-H(7)	120(2)
C(9)-C(8)-C(7)	119.1(4)
C(9)-C(8)-H(8)	122(3)
C(7)-C(8)-H(8)	119(3)
N-C(9)-C(8)	123.0(4)
N-C(9)-H(9)	115(3)
C(8)-C(9)-H(9)	122(3)
O(1)-C(10)-N(1)	125.6(4)
O(1)-C(10)-C(2)	126.4(4)
N(1)-C(10)-C(2)	108.0(3)
C(9)-N-C(12)	116.8(4)
N-C(12)-C(6)	124.6(4)
N-C(12)-H(12)	114(2)
C(6)-C(12)-H(12)	121(2)

Symmetry transformations used to generate equivalent atoms: 0

Table 4. Anisotropic displacement parameters ($\text{\AA}^2 \times 10^3$) for 905295.

The anisotropic displacement factor exponent takes the form:

$$-2 \pi^2 [h^2 a^{*2} U_{11} + \dots + 2 h k a^* b^* U_{12}]$$

	U11	U22	U33	U23	U13	U12
O(1)	34(2)	46(2)	38(2)	0(1)	4(1)	4(1)
O(2)	24(2)	51(2)	38(2)	-1(2)	-6(1)	-3(1)
N(1)	31(2)	35(2)	31(2)	1(2)	3(2)	1(2)
C(1)	38(3)	39(2)	49(3)	7(2)	7(2)	-3(2)
C(2)	25(2)	36(2)	36(2)	3(2)	4(2)	-4(2)
C(3)	16(2)	49(2)	37(2)	-1(2)	-4(2)	-3(2)
C(4)	16(2)	46(2)	34(2)	-2(2)	7(2)	0(2)
C(5)	25(2)	45(3)	39(2)	-1(2)	7(2)	-3(2)
C(6)	25(2)	38(2)	34(2)	-4(2)	4(2)	-4(2)
C(7)	29(2)	37(2)	38(2)	-3(2)	0(2)	-3(2)
C(8)	28(2)	44(2)	42(3)	-5(2)	-5(2)	0(2)
C(9)	25(2)	48(2)	36(2)	-7(2)	2(2)	-1(2)
C(10)	24(2)	40(2)	36(2)	0(2)	-3(2)	-2(2)
N	36(2)	51(2)	34(2)	0(2)	7(2)	0(2)
C(12)	35(2)	39(2)	38(2)	-3(2)	4(2)	-2(2)

Table 5. Hydrogen coordinates ($\times 10^4$) and isotropic displacement parameters ($\text{\AA}^2 \times 10^3$) for 905295.

	x	y	z	U(eq)
H(2)	-6770(110)	-4020(60)	-670(20)	63(15)
H(2A)	-10780(90)	-4550(40)	-2031(15)	28(10)
H(1A)	-7420(90)	-6790(50)	-1972(16)	34(10)
H(5A)	-8400(80)	370(40)	-1680(15)	24(10)
H(4)	-11550(80)	-1710(40)	-1825(14)	28(10)
H(5B)	-6670(100)	-620(50)	-1251(17)	40(13)
H(3)	-10920(80)	-3470(40)	-1105(15)	23(9)
H(1)	-7010(100)	-1330(50)	-2411(18)	49(13)
H(1B)	-5650(130)	-5840(50)	-1506(19)	54(14)
H(9)	-15350(110)	1130(50)	250(20)	57(14)
H(1C)	-8770(120)	-6390(60)	-1350(20)	71(17)
H(12)	-9360(100)	-1500(50)	-328(17)	44(12)
H(7)	-12660(80)	1640(40)	-1374(16)	25(10)
H(8)	-15680(130)	2390(60)	-650(20)	70(16)

Table 6. Torsion angles [deg] for 905295.

C(1)-C(2)-C(3)-O(2)	45.4(5)
C(10)-C(2)-C(3)-O(2)	-77.9(4)
C(1)-C(2)-C(3)-C(4)	160.9(3)
C(10)-C(2)-C(3)-C(4)	37.7(4)
C(10)-N(1)-C(4)-C(5)	143.6(4)
C(10)-N(1)-C(4)-C(3)	20.5(4)
O(2)-C(3)-C(4)-N(1)	82.0(4)
C(2)-C(3)-C(4)-N(1)	-35.5(4)
O(2)-C(3)-C(4)-C(5)	-36.9(5)
C(2)-C(3)-C(4)-C(5)	-154.4(3)
N(1)-C(4)-C(5)-C(6)	175.3(3)
C(3)-C(4)-C(5)-C(6)	-70.9(5)
C(4)-C(5)-C(6)-C(12)	79.5(5)
C(4)-C(5)-C(6)-C(7)	-102.2(5)
C(12)-C(6)-C(7)-C(8)	0.8(6)
C(5)-C(6)-C(7)-C(8)	-177.5(4)
C(6)-C(7)-C(8)-C(9)	-1.5(6)
C(7)-C(8)-C(9)-N	1.0(6)
C(4)-N(1)-C(10)-O(1)	-174.8(4)
C(4)-N(1)-C(10)-C(2)	3.6(5)
C(1)-C(2)-C(10)-O(1)	26.6(6)
C(3)-C(2)-C(10)-O(1)	152.0(4)
C(1)-C(2)-C(10)-N(1)	-151.8(4)
C(3)-C(2)-C(10)-N(1)	-26.4(4)
C(8)-C(9)-N-C(12)	0.2(6)
C(9)-N-C(12)-C(6)	-1.0(6)
C(7)-C(6)-C(12)-N	0.4(6)
C(5)-C(6)-C(12)-N	178.8(4)

Symmetry transformations used to generate equivalent atoms: 0