## Synthesis, antiplasmodial and antimycobacterial evaluation of new nitroimidazole and nitroimidazooxazine derivatives

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### 1. Synthesis and characterization of compounds 8-16

#### • 1.1 General

Chemical reagents used were supplied by either Sigma-Aldrich<sup>®</sup> or Merck and were used without further purification. The (S)-2-nitro-6,7-dihydro-5H-imidazo[2,1b][1,3]oxazin-6-ol (6) was purchased from AAT Pharmaceuticals, New Jersey, USA. Unless otherwise stated, all the solvents used were anhydrous and were purchased from Sigma-Aldrich, with the exception of THF and diethyl ether which were dried by appropriate techniques. Chromatographic solvents such as Ethyl acetate, dichloromethane and Hexane were purchased from Kimix or Protea Chemicals and were distilled prior to use. Column chromatography was carried out using Merck Kieselgel 60: particle size 70- 230 mesh as a stationary phase. Analytical TLC was carried out using Merck PF<sub>254</sub> aluminium-backed pre-coated silica gel plates and visualized under UV light or iodine vapour. Melting points were determined on a Reichert-Jung Thermovar hot-stage microscope and are uncorrected. Low-resolution mass spectra were obtained by flow-injection (5 mM NH<sub>4</sub> formate pH3 in H<sub>2</sub>O:ACN, no column used) on a AB SCIEX 4000 QTRAP Hybrid triple quadrupole linear ion trap mass spectrometer, coupled with an Agilent 1200 Rapid Resolution (600 bar) HPLC system consisting of a binary pump, degasser, auto sampler and temperature controlled column compartment. Infra-red spectra were recorded on a PerkinElmer Spectrum 100 FT-IR spectrometer in the 4000 – 450 cm<sup>-1</sup> range, with samples either as dichloromethane solution or as KBr discs. NMR spectra were recorded on Bruker 400 MHz or Varian Unity 400 MHz and/or Varian Mercury 300 MHz spectrometers, all chemical shifts are reported in ppm and were referenced using solvent signals (2.50 and 39.4 ppm for DMSO- $d_6$  and 7.26 and 77.0 ppm for CDCl<sub>3</sub>). Chemical shifts ( $\delta$ ) are recorded in parts per million (ppm). Coupling constants, *J*, are measured in Hertz (Hz) and rounded off to one decimal place. Abbreviations used in the assignment of the <sup>1</sup>H NMR spectra are as follows: br (broad), d (doublet), dd (doublet of doublets), m (multiplets), s (singlet) and t (triplet). <sup>13</sup>C NMR chemical shifts are listed without assignment to specific carbon atoms. Purity was determined by HPLC, and all compounds were confirmed to have purity >95%. Sample solutions for purification were prepared at ca. 100 mg/mL in 50% methanol. Injection volumes ranged between 50 🗉 and 1 mL, the mobile phase flow rate was 20 mL/min for all purifications and the column heater was set at 30°C. A mobile phase gradient was used, where mobile phase A, an aqueous solution of 0.1% formic acid, was mixed on-line with mobile phase B, a 0.1% formic acid solution in methanol.

#### **1.2** General procedure<sup>1</sup> for the synthesis of compounds 8a-b

At o °C, under N<sub>2</sub> atmosphere, NaH (60% mineral oil, 1.3 eq) was added to the stirred solution of p, m and o-(bromomethyl)benzaldehydes (1.2 eq) (7a-c) and (S)-2-nitro-6,7-dihydro-5H-imidazo[2,1-b][1,3]oxazin-6-ol (**6**) (1.0 eq) in anhydrous DMF (5 ml). The resultant mixture was maintained at this temperature for 30 minutes, allowed to warm to room temperature and stirred further for 12 hrs. On completion of the reaction as seen on the TLC, the solvent was removed in *vacuo* by azeotropic distillation of toluene to afford crude products **8a-b** (m- and p-based products). The o-(bromomethyl)benzaldehyde reaction did not work as evidence by the recovery of starting materials. The crude products were then purified by column chromatography (on silica gel; elution with DCM: MeOH; 95:05).

4-{[(*S*)-6,7-*dihydro*-2-*nitro*-5H-*imidazo*[2,1-*b*][1,3]*oxazin*-6-*yloxy*]*methyl*}*benzaldehyde* (**8a**) Yield (91 mg) 46%; m. p. 145- 148 °C, R<sub>f</sub> (DCM: MeOH, 95: 05%) 0.27; IR ν<sub>max</sub> (DCM)/cm<sup>-1</sup> 1699 (C=O), 1579 (Ar C=C), 1548 (NO<sub>2</sub>), 1266 (C-O Ether);  $\delta_{\rm H}$  (400 MHz; DMSO-*d*6) 9.99 (1H, s, CHO), 8.02 (1H, s, H1), 7.89 (2H, d, *J* 8.1 Hz, 2 x H7), 7.52 (2H, d, *J* 8.1 Hz, 2 x H6), 4.77 (2H, d, *J* 3.2 Hz, H5), 4.49 (1H, dt, *J* 2.6 and 12.1 Hz, H3), 4.48 (1H, d, *J* 11.9 Hz, H4*a*), 4.28 (3H, m, H2 and H4*b*);  $\delta_{\rm C}$  (101 MHz; DMSO-*d*6) 193.1, 147.6, 145.2, 136.0, 130.0 (2C), 128.2 (2C), 127.1, 118.4, 69.6, 68.4, 62.9 and 47.2; MS (ESI) *m/z* 304.1 (M<sup>+</sup>+H), HPLC purity: 98.9%; t<sub>r</sub>=6.98 min. 4-{[(*S*)-6,7-*dihydro*-2-*nitro*-5H-*imidazo*[2,1-*b*][1,3]*oxazin*-6-*yloxy*]*methyl*}*benzaldehyde* (**8b**) Yield (131 mg) 78%; m. p. 129- 132 °C, R<sub>f</sub> (DCM: MeOH, 95: 05%) 0.22; IR v<sub>max</sub> (DCM)/cm<sup>-1</sup> 1685 (C=O), 1569 (Ar C=C), 1538 (NO<sub>2</sub>), 1245 (C-O Ether);  $\delta_{\rm H}$  (400 MHz; DMSO-*d6*) 10.01 (1H, s, CHO), 7.99 (1H, s, H1), 7.83 (2H, m, H8 and H9), 7.65 (1H, t, *J* 7.7 Hz, H7), 7.60 (1H, d, *J* 7.7 Hz, H6), 4.76 (2H, d, *J* 1.9 Hz, H5), 4.66 (1H, dt, *J* 4.7 and 11.9 Hz, H3), 4.49 (1H, d, *J* 11.6 Hz, H3*a*), 4.28 (3H, m, H2 and H3*b*);  $\delta_{\rm C}$  (101 MHz; DMSO-*d6*) 192.8, 148.3, 143.2, 138.3, 135.9, 133.3, 129.0, 128.8, 127.9, 117.7, 68.9, 67.7, 66.5 and 46.5; MS (ESI) *m/z* 304.2 (M<sup>+</sup>+H), HPLC purity: 93.6%; t<sub>r</sub>=7.29 min.

#### **1.3** General procedure<sup>2</sup> for the synthesis of compounds 9a-h

The nitroimidazo-oxazine aldehyde **8a-b** (0.099 mmol) and various amine (including 4-amino-7chloroquinoline diamines)<sup>§</sup> (0.207 mmol) were stirred in anhydrous methanol as a suspension at 26 °C for 5 minutes. Thereafter, TMSN<sub>3</sub> (0.207 mmol) was added, followed by the addition of *tert*-butyl isocyanide (0.207 mmol) and the resulting mixture was then further stirred at 40 °C for 12 hrs. Thereafter, the solvent was removed in *vacuo* to afford the crude tetrazoles which were purified by column chromatography (on silica gel; elution with DCM: MeOH; 95:05) and/or HPLC to give the desired products, **9a-h**, in modest to excellent yields.

### (4-{[(*S*)-6,7-*dihydro*-2-*nitro*-5H-*imidazo*[2,1-*b*][1,3]*oxazin*-6-*yloxy*]*methyl*}*phenyl*)(1-tert*butyl*-1H-*tetrazol*-5-*yl*)-N,N-*dimethylmethanamine* (**9a**)

Yield (25 mg) 46%; m. p. 68- 72 °C, R<sub>f</sub> (DCM: MeOH, 95: 05%) 0.32; IR  $\nu_{max}$  (DCM)/cm<sup>-1</sup> 1588 (Ar C=C), 1566 (NO<sub>2</sub>), 1368 (N=N), 1340 (C=N), 1284 (C-O Ether);  $\delta_{\rm H}$  (400 MHz; CDCl<sub>3</sub>) 7.42 (2H, d, *J* 8.0 Hz, 2 x H7), 7.37 (1H, d, *J* 12.9 Hz, H1), 7.27 (2H, d, *J* 8.0 Hz, 2 x H6), 5.19 (1H, s, H8), 4.72 (1H, m, H3), 4.59 (2H, m, H5), 4.35 (1H, d, *J* 12.1 Hz, H4a), 4.17

<sup>&</sup>lt;sup>§</sup> HCl protected amines were neutralized by addition of the base, diisopropyl ethylamine (2 eq)

(3H, m, H2 and H4*b*), 2.29 (6H, s, 2 x H9), 1.69 (9H, s, 3 x H10);  $\delta_{C}$  (101 MHz; CDCl<sub>3</sub>) 154.3, 147.2, 136.9, 136.0, 130.1 (2C), 127.6 (2C), 127.0, 115.0, 70.7, 67.4, 66.7, 65.0, 63.2, 47.5; 42.4 (2C) and 30.2 (3C); MS (ESI) *m/z* 457.3 (M<sup>+</sup>), HPLC purity: 97.5%; t<sub>r</sub>=8.30 min.

N-(4-{[((*S*)-6,7-*dihydro*-2-*nitro*-5H-*imidazo*[2,1-*b*][1,3]*oxazin*-6-*yloxy*)*methyl*]*pheny*}(1-tert*butyl*-1H-*tetrazol*-5-*yl*)*methyl*)-N-*dimethylmethanamine* (**9b**)

Yield (30.2 mg, 63%); m. p. 48- 50 °C, R<sub>f</sub> (DCM: MeOH, 95: 05%) 0.46; IR  $\nu_{max}$  (DCM)/cm<sup>-1</sup> 1601 (Ar C=C), 1536 (NO<sub>2</sub>), 1378 (N=N), 1340 (C=N), 1260 (C-O Ether);  $\delta_{\rm H}$  (400 MHz; CDCl<sub>3</sub>) 7.37 (1H, d, *J* 15.0 Hz, H1), 7.25 (4H,m, 2 x H6 and 2 x H7), 5.61 (1H, s, H8), 4.72 (1H, m, H3), 4.59 (2H, m, H5), 4.34 (1H, d, *J* 12.3 Hz, H4a), 4.16 (3H, m, H2 and H4b), 2.78 (2H, q, *J* 7.1 Hz, H9a), 2.65 (2H, q, *J* 7.1 Hz, H9b), 1.66 (9H, s, 3 x H11), 0.95 (6H, t, *J* 7.1 Hz, 2 x H10);  $\delta_{\rm C}$  (101 MHz; CDCl<sub>3</sub>) 154.7, 147.1, 138.0, 136.4, 136.3, 129.7 (2C), 127.5 (2C), 127.4, 114.9, 70.8, 67.4, 67.3, 60.6, 47.5; 44.8 (2C), 30.3 (3C) and 14.1 (2C); MS (ESI) *m/z* 485.4 (M<sup>+</sup>+H), HPLC purity: 98.9%; t<sub>r</sub>=8.06 min.

(*S*)-6-{4-[(1-tert-*butyl*-1H-*tetrazol*-5-*yl*)(*pyrrolidin*-1-*yl*)*methyl*]*benzyloxy*}-6,7-*dihydro*-2*nitro*-5H-*imidazo*[2,1-*b*][1,3]*oxazine*, (**9c**)

Yield (21 mg) 44%; m. p. 58- 61 °C, R<sub>f</sub> (DCM: MeOH, 95: o5%) o.39; IR  $v_{max}$  (DCM)/cm<sup>-1</sup> 1604 (Ar C=C), 1520 (NO<sub>2</sub>), 1380 (N=N), 1360 (C=N), 1263 (C-O Ether);  $\delta_{\rm H}$  (300 MHz; CDCl<sub>3</sub>) 7.50 (2H, d, *J* 8.1 Hz, 2 x H7), 7.37 (1H, d, *J* 6.7 Hz, H1), 7.26 (2H, d, *J* 8.1 Hz, 2 x H6), 5.20 (1H, s, H8), 4.72 (1H, m, H3), 4.62 (2H, m, H5), 4.34 (1H, d, *J* 12.1 Hz, H4a), 4.17 (3H, m, H2 and H4b), 2.59 (2H, m, H9a), 2.49 (2H, m, H9b), 1.79 (4H, m, 2 x H10), 1.68 (9H, s, 3 x H11);  $\delta_{\rm C}$  (75 MHz; CDCl<sub>3</sub>) 155.5, 147.2, 136.1, 127.4, 123.1, 121.9 (2C), 119.2 (2C), 118.3, 79.9, 67.4, 63.2, 53.0, 52.2, 52.1, 41.2 (2C), 30.2 (3C) and 24.6 (2C); MS (ESI) *m/z* 483.4 (M<sup>+</sup>+H), HPLC purity: 99.6%; t<sub>r</sub>=7.68 min. N-{2-[(4-{[((*S*)-6,7-*dihydro*-2-*nitro*-5H-*imidazo*[2,1-*b*][1,3]*oxazin*-6-*yloxy*)*methyl*]*pheny*}(1-tert-*butyl*-1H-*tetrazol*-5-*yl*)*methylamino*)*ethyl*]}-7-*chloroquinolin*-4-*amine*, (**9d**)

Yield (27 mg) 65%; m. p. 87- 89 °C, R<sub>f</sub> (DCM: MeOH, NH<sub>4</sub>OH, 94: 5.5: 0.5%) 0.16; IR  $v_{max}$  (DCM)/cm<sup>-1</sup> 1610 (Ar C=C), 1550 (NO<sub>2</sub>), 1390 (N=N), 1369 (C=N), 1250 (C-O Ether);  $\delta_{\rm H}$  (400 MHz; DMSO-*d*6) 8.35 (1H, d, *J* 5.4 Hz, Hz'), 8.20 (1H, d, *J* 9.1 Hz, H5'), 7.99 (1H, d, *J* 4.3 Hz, H1), 7.77 (1H, d, *J* 2.2 Hz, H8'), 7.43 (3H, m, 2 x H7 and H6'), 7.26 (2H, d, *J* 8.1 Hz, 2 x H6), 7.18 (1H, t, *J* 5.3 Hz, NH), 6.43 (1H, d, *J* 5.4 Hz, H3'), 5.53 (1H, s, H8), 4.63 (3H, m, H3 and H5), 4.46 (1H, d, *J* 11.9 Hz, H4a), 4.23 (3H, m, H2 and H4b), 3.35 (2H, m, H9), 4.30 (1H, br m, NH), 2.78 (2H, m, H10), 1.62 (9H, s, 3 x H11);  $\delta_{\rm C}$  (101 MHz; DMSO-*d*6) 158.2, 155.8, 151.7 (2C), 149.9, 148.9, 146.5, 138.6, 137.1, 133.2, 128.1 (2C), 127.5 (2C), 127.4 (2C), 123.9, 123.8, 117.8, 98.6, 69.2, 67.7, 66.3, 55.8 46.6, 45.1; 42.5 and 29.2 (3C); MS (ESI) *m*/z 633.5 (M<sup>+</sup>), HPLC purity: 96.2%; t<sub>r</sub>=8.44 min.

# N-{3-[(4-{[((*S*)-6,7-*dihydro*-2-*nitro*-5H-*imidazo*[2,1-*b*][1,3]*oxazin*-6-*yloxy*)*methyl*]*pheny*}(1-tert-*butyl*-1H-*tetrazol*-5-*yl*)*methylamino*)*propyl*]}-7-*chloroquinolin*-4-*amine*, (**9e**)

Yield (20 mg) 47%; m. p. 95- 99 °C, R<sub>f</sub> (DCM: MeOH, NH<sub>4</sub>OH, 94: 5.5: 0.5%) 0.35; IR  $\nu_{max}$  (DCM)/cm<sup>-1</sup> 1613 (Ar C=C), 1576 (NO<sub>2</sub>), 1369 (N=N), 1387 (C=N), 1265 (C-O Ether);  $\delta_{\rm H}$  (400 MHz; CDCl<sub>3</sub>) 8.45 (1H, d, *J* 5.3 Hz, H2'), 7.86 (1H, d, *J* 2.2 Hz, H8'), 7.27 (6H, m, 2 x H7, 2 x H6, H1 and H5'), 7.15 (1H, br s, NH), 6.95 (1H, dd, *J* 9.0 and 2.2 Hz, H6'), 6.29 (1H, d, *J* 5.3 Hz, H3'), 5.30 (1H, s, H8), 4.73 (1H, m, H3), 4.63 (2H, dd, *J* 12.1 and 3.5Hz, H5), 4.36 (1H, d, *J* 12.1 Hz, H4a), 4.18 (3H, m, H2 and H4b), 3.41 (2H, m, H9), 2.88 (1H, m, H1a), 2.79 (1H, m, H1b), 1.92 (2H, m, H10), 1.53 (9H, s, 3 x H12);  $\delta_{\rm C}$  (101 MHz; CDCl<sub>3</sub>) 156.7, 155.2, 152.0 (2C), 150.2, 149.0, 147.0, 144.1, 143.7, 138.3, 137.4, 134.4, 128.4 (2C), 128.3, 128.2, 124.8, 122.1, 117.4, 98.4, 70.5, 67.2, 67.1, 61.6, 59.0, 47.8, 47.4; 43.5 and 29.8 (3C); MS (ESI) *m*/*z* 647.4 (M<sup>+</sup>), HPLC purity: 99.2%; t<sub>r</sub>=8.61 min.

N-{4-[(4-{[((*S*)-6,7-*dihydro*-2-*nitro*-5H-*imidazo*[2,1-*b*][1,3]*oxazin*-6-*yloxy*)*methyl*]*pheny*}(1-tert-*butyl*-1H-*tetrazol*-5-*yl*)*methylamino*)*butyl*]}-7-*chloroquinolin*-4-*amine*, (**9f**)

Yield (37 mg) 85%; m. p. 82- 86 °C, R<sub>f</sub> (DCM: MeOH, NH<sub>4</sub>OH, 94: 5.5: 0.5%) 0.32; IR  $v_{max}$  (DCM)/cm<sup>-1</sup> 1619 (Ar C=C), 1555 (NO<sub>2</sub>), 1388 (N=N), 1360 (C=N), 1248 (C-O Ether);  $\delta_{H}$  (400 MHz; CDCl<sub>3</sub>) 8.48 (1H, d, *J* 5.4 Hz, H2'), 7.92 (1H, d, *J* 2.0 Hz, H8'), 7.57 (1H, dd, *J* 9.2 and 2.0 Hz, H6'), 7.33 (1H, d, *J* 6.5 Hz, H1), 7.24 (5H, m, 2 x H7, 2 x H6 and H5'), 6.37 (1H, d, *J* 5.4 Hz, H3'), 5.48 (1H, m, NH), 5.27 (1H, s, H8), 4.67 (1H, m, H3), 4.60 (2H, m, H5), 4.32 (1H, d, *J* 11.7 Hz, H4*a*), 4.12 (3H, m, H2 and H4*b*), 3.32 (2H, m, H12), 2.62 (2H, m, H9), 1.83 (2H, m, H11), 1.69 (2H, m, H10), 1.61 (9H, s, 3 x H13);  $\delta_{C}$  (101 MHz; CDCl<sub>3</sub>) 157.4, 155.5, 151.9 (2C), 149.9, 149.8, 138.9, 137.1, 137.0, 128.6, 128.5, 128.3 (2C), 128.1, 128.0, 125.1, 121.2, 115.0, 99.0, 70.6, 67.3, 67.2, 66.9, 66.8, 58.8, 47.5, 42.9, 30.1 (3C), 27.4 and 26.3; MS (ESI) *m*/z 661.5 (M<sup>+</sup>), HPLC purity: 96.9%; t<sub>r</sub>=8.66 min.

# N-{5-[(4-{[((*S*)-6,7-*dihydro*-2-*nitro*-5H-*imidazo*[2,1-*b*][1,3]*oxazin*-6-*yloxy*)*methyl*]*pheny*}(1-tert-*butyl*-1H-*tetrazol*-5-*yl*)*methylamino*)*pentan*-2-*yl*]}-6-*methoxyquinolin*-8-*amine* (**9g**)

Yield (55 mg) 83% (1:1 diastereomeric mixture); R<sub>f</sub> (DCM: MeOH, 95: o5%) o.48; IR  $\nu_{max}$  (DCM)/cm<sup>-1</sup> 1601 (Ar C=C), 1539 (NO<sub>2</sub>), 1388 (N=N), 1338 (C=N), 1238 (C-O Ester);  $\delta_{H}$  (300 MHz; CDCl<sub>3</sub>) 8.59 (1H, s, H1), 8.50 (1H, dd, *J* 4.2 and 1.6 Hz, H2'), 7.91 (1H, dd, *J* 8.3 and 1.6 Hz, H4'), 7.36 (1H, dd, *J* 8.7 and 4.4 Hz, H3'), 7.31 (2H, m, H7), 7.22 (2H, m, H6), 6.32 (1H, d, *J* 2.5 Hz, H7'), 6.24 (1H, t, *J* 2.8 Hz, H5'), 6.01 (1H, br m, NH), 5.24-5.26 (1H, 2 x s, H8), 4.68 (1H, d, *J* 12.2 Hz, H3), 4.57 (2H, m, H5), 4.31 (1H, d, *J* 12.4 Hz, H4a), 4.12 (3H, m, H2 and H4b), 3.88 (3H, s, OCH<sub>3</sub>), 3.57 (1H, m, H12), 2.56 (2H, m, H9), 1.64 (4H, m, H10 and H11), 1.59-1.60 (9H, 2 x s, 3 x H14), 1.27 (3H, d, *J* 6.3 Hz, H13);  $\delta_{C}$  (75 MHz; CDCl<sub>3</sub>) 159.5, 155.6, 145.0, 144.3, 143.8, 139.7, 136.6, 134.8, 129.9, 128.5 (2C), 128.2, 128.1 (2C), 127.4, 121.8, 114.9, 96.6, 91.6, 70.6, 67.3, 67.2, 66.4, 58.5, 55.2, 47.9; 47.5, 47.4, 34.2, 29.8 (3C), 26.5 and 20.6; MS (ESI) *m/z* 671.6 (M<sup>+</sup>+H), HPLC purity: 98.7%; t<sub>r</sub>=8.52 min.

 $N-\{5-[(3-\{[((S)-6,7-dihydro-2-nitro-5H-imidazo[2,1-b][1,3]oxazin-6-yloxy)methyl]pheny\}(1-tert-butyl-1H-tetrazol-5-yl)methylamino)pentan-2-yl]\}-6-methoxyquinolin-8-amine ($ **9h**)

Yield (140 mg) 79% (1:1 diastereomeric mixture); R<sub>f</sub> (DCM: MeOH, 95: o5%) o.52; IR  $v_{max}$  (DCM)/cm<sup>-1</sup> 1610 (Ar C=C), 1545 (NO<sub>2</sub>), 1390 (N=N), 1340 (C=N), 1228 (C-O Ester);  $\delta_{H}$  (400 MHz; CDCl<sub>3</sub>) 8.50 (1H, s, H1), 8.49 (1H, dd, *J* 4.2 and 1.7 Hz, H2'), 7.90 (1H, dd, *J* 8.3 and 1.6 Hz, H4'), 7.32 (1H, s, H9), 7.31 (2H, m, H3' and H8), 7.20 (2H, m, H6 and H7), 6.32 (1H, d, *J* 2.5 Hz, H7'), 6.24 (1H, dd, *J* 6.8 and 2.7 Hz, H5'), 5.99 (1H, br m, NH), 5.25-5.27 (1H, 2 x s, H10), 4.66 (1H, m, H3), 4.54 (2H, m, H5), 4.26 (1H, m, H4a), 4.09 (3H, m, H2 and H4b), 3.86-3.87 (3H, 2 x s, OCH<sub>3</sub>), 3.59 (1H, m, H14), 2.55 (2H, m, H11), 1.72 (9H, s, 3 x H16), 1.66 (4H, m, H12 and H13), 1.27-1.29 (3H, 2 x d, *J* 5.9 and 6.3 Hz, H15);  $\delta_{C}$  (101 MHz; CDCl<sub>3</sub>) 159.5, 155.7, 145.0, 144.2, 139.7, 137.5, 135.3, 134.8, 129.9, 129.2, 128.1, 127.6, 127.5, 127.0, 121.9, 115.1, 96.7, 91.6, 70.9, 67.7, 66.5, 66.4, 58.6, 55.2, 47.9; 47.8, 47.4, 34.3, 29.9 (3C), 26.5 and 20.6; MS (ESI) *m*/*z* 671.9 (M<sup>+</sup>+H); HPLC purity: 95.6%; t<sub>r</sub>=8.55 min.

#### 1.4 General procedure for the synthesis of compounds 11a-b

To a suspension of **10a/10b** (2.1 mmol) in anhydrous THF (10 ml) under inert nitrogen atmosphere was added TEA (4.2 mmol) and the resulting mixture cooled to below o°C. Methanesulfonyl chloride (2.2 mmol) was then added slowly while keeping the temperature below 5°C. On complete addition of methanesulfonyl chloride the reaction was stirred in an ice bath for 45 minutes. Upon complete consumption of the alcohol, as evidence by TLC, the reaction mixture was diluted with saturated NaHCO<sub>3</sub> solution and extracted with ether (3 x 30 ml). The organic extracts were then dried over MgSO<sub>4</sub>, filtered, and evaporated to give **11a-b** in reasonable purity and good yields.

#### 2-((7-Chloroquinolin-4-yl)amino)ethyl methanesulfonate (11a)

Yield (430 mg) 65%; m. p. 134-135 °C; R<sub>f</sub> (DCM: MeOH, 9:1%) 0.58;  $\delta_{\rm H}$  (400 MHz; CDCl<sub>3</sub>): 8.46 (1H, d, *J* 5.4 Hz, H2), 7.91 (1H, d, *J* 9 Hz, H5), 7.73 (1H, d, *J* 2.2 Hz, H8), 7.33 (1H, dd, *J* 2.2 and 9 Hz, H6), 6.35 (1H, d, *J* 5.4 Hz, H3), 5.83 (1H, bs, NH), 4.53 (2H, t, *J* 5.9 Hz, 2 x H2'), 3.66 (2H, t, *J* 5.9 Hz, 2 x H1'), 3.08 (3H, s, SO<sub>2</sub>Me).

#### 3-((7-Chloroquinolin-4-yl)amino)propyl methanesulfonate (11b)

Yield (0.4 g) 60%; m. p. 128-129 °C; R<sub>f</sub> (DCM: MeOH, 9:1%) 0.52;  $\delta_{\rm H}$  (400 MHz; CDCl<sub>3</sub>): 8.55 (1H, d, *J* 5.3 Hz, H2), 7.97 (1H, d, *J* 8.9 Hz, H5), 7.70 (1H, d, *J* 2.1 Hz, H8), 7.40 (1H, dd, *J* 2.1 and 8.9 Hz, H6), 6.43 (1H, d, *J* 5.3 Hz, H3), 5.44(1H, bs, NH), 4.41 (2H, t, *J* 5.7 Hz, 2 x H3'), 3.59 (2H, m, 2 x H1'), 3.05 (3H, s, SO<sub>2</sub>Me), 2.18 (2H, m, 2 x H2').

#### 1.5 General procedure for the synthesis of compounds 12a-b

Intermediate **11a**/**11b** (1.66 mmol) was cooled to o°C and *N*-methyl ethanolamine (33.2 mmol) added drop-wise over a period of 10-15 minutes at this temperature. On complete addition of *N*-methyl ethanolamine, TEA (11.25 mmol) was added drop wise to the mixture. The reaction was allowed to warm to room temperature and stirred for one hour. Then the reaction temperature was raised to 45-55°C and further stirred for 4 hours. On completion of the reaction,  $H_2O(50 \text{ ml})$  was added and the reaction extracted with ethyl acetate(3 x 50 ml).The organic layers were combined, washed with brine, dried over MgSO<sub>4</sub>, filtered and evaporation in *vacuo* to afford **12a/b**.

#### 2[{2-[(7-Chloroquinolin-4-yl)amino]ethyl}-(methyl)amino]ethanol (12a)

Yield (0.372 g) 80%; m. p. 106-107 °C; R<sub>f</sub> (DCM: MeOH, 9:1%) 0.14;  $\delta_{\rm H}$  (400 MHz; CD<sub>3</sub>OD): 8.34 (1H, d, *J* 5.6 Hz, H2), 8.06 (1H, d, *J* 9 Hz, H5), 7.77 (1H, d, *J* 2.1 Hz, H8), 7.40 (1H, dd, *J* 2.1 and 9 Hz, H6), 6.43 (1H, d, *J* 5.6 Hz, H3), 5.44 (1H, bs, NH), 3.67 (2H, t, *J* 5.7 Hz, 2 x H4'), 3.44 (2H, t, *J* 6.4 Hz, 2 x H1'), 2.78-2.61 (4H, m, 2 x H2' and H3'),

2.36(3H, s, NCH<sub>3</sub>);  $\delta_c$  (100 MHz; CD<sub>3</sub>OD): 151.1, 151, 148.1, 134.9, 126.1, 124.6, 122.8, 117.3, 98.3, 58.9, 58.8, 55.1, 41.2 and 40.1; MS (ESI) *m/z* 279.1(M<sup>+</sup>); HPLC purity: 98.6%; t<sub>r</sub>=13.4 min.

#### 2[{3-[(7-Chloroquinolin-4-yl)amino]propyl}-(methyl)amino]ethanol (12b)

Yield (0.349 g) 75%; m. p. 115-116 °C; R<sub>f</sub> (DCM: MeOH, 9:1%) 0.12;  $\delta_{\rm H}$  (400 MHz; CD3OD): 8.34 (1H, d, *J* 5.6 Hz, H2), 8.06 (1H, d, *J* 9Hz, H5), 7.77 (1H, d, *J* 2.1 Hz, H8), 7.40 (1H, dd, *J* 2.1 and 9 Hz, H6), 6.43 (1H, d, *J* 5.6 Hz, H3), 5.44 (1H, bs, NH), 3.67 (2H, t, *J* 5.9 Hz, 2 x H5'), 3.41 (2H, t, *J* 6.7 Hz, 2 x H1'), 2.80-2.61 (4H, m, 2 x H3' and H4'), 2.32 (3H, s, NCH<sub>3</sub>), 2.18 (2H, m, 2 x H2');  $\delta_{\rm c}$  (100 MHz; CD<sub>3</sub>OD): 151.3, 151, 148.2, 134.9, 126.1, 124.5, 122.9, 117.4, 98.1, 59, 58.9, 55.7, 41.3(2C), and 25.4; MS(ESI) *m*/*z* 294.2(M<sup>+</sup>+H); HPLC purity: 96.8%; t<sub>r</sub>=11.33 min.

#### 1.6 General procedure for the synthesis of compounds 13a-b

To the suspension of intermediate 12a/12b(0.852 mmol) in dry toluene (3 ml) and dry DMF(0.3 ml) at 0°C was added SOCl<sub>2</sub> solution (8.8 mmol) in dry toluene(3 ml) dropwise in 15 minutes. The temperature of the resultant was raised to reflux for 14 hours. On reaction completion, the solvent was evaporated in *vacuo*, H<sub>2</sub>O (5 ml) added and pH of the resulting mixture adjusted to 7-8 by addition of saturated sodium bicarbonate. The mixture was extracted with ethyl acetate (3 x 50 ml), washed with brine, dried over MgSO<sub>4</sub> and concentrated to give **13a-b**.

2[{2-[(7-Chloroquinolin-4-yl)amino]ethyl}-(methyl)amino]ethylchloride (13a)

Yield (0.191g) 60%; m. p. 100-101 °C;  $R_f$  (DCM: MeOH, 8:2%) 0.4;  $\delta_H$  (400 MHz; CD<sub>3</sub>OD): 8.34 (1H, d, *J* 5.6 Hz, H2), 8.02 (1H, d, *J* 9 Hz, H5), 7.75 (1H, d, *J* 2.1 Hz, H8), 7.40 (1H, dd, *J* 2.1 and 9 Hz, H6), 6.44 (1H, d, *J* 5.6 Hz, H3), 5.42 (1H, bs, NH), 3.56 (2H,

t, J 5.7 Hz, 2 x H4'), 3.42 (2H, t, J 6.4 Hz, 2 x H1'), 2.78-2.56 (4H, m, 2 x H2' and H3'), 2.36 (3H, s, NCH<sub>3</sub>);  $\delta_c$  (100 MHz; CD<sub>3</sub>OD): 151.1, 151, 148.1, 134.9, 126.1, 124.6, 122.8, 117.3, 98.3, 58.8, 55.1, 41.2, 40.1 and 39.8; MS(ESI) m/z 298.2(M<sup>+</sup>+H).

2[{3-[(7-Chloroquinolin-4-yl)amino]propyl}-(methyl)amino]ethylchloride (13b)

Yield (0.213g) 67%; m. p. 105-106 °C; R<sub>f</sub> (DCM: MeOH, 8:2%) 0.44;  $\delta_{\rm H}$  (400 MHz; CD<sub>3</sub>OD): 8.33 (1H, d, *J* 5.6 Hz, H2), 8.01 (1H, d, *J* 9 Hz, H5), 7.76 (1H, d, *J* 2.1 Hz, H8), 7.38 (1H, dd, *J* 2.1 and 9 Hz, H6), 6.50 (1H, d, *J* 5.6 Hz, H3), 5.44 (1H, bs, NH), 3.65 (2H, t, *J* 6.5 Hz, 2 x H5'), 3.42 (2H, t, *J* 6.7 Hz, 2 x H1'), 2.80-2.61 (4H, m, 2 x H3' and H4'), 2.32 (3H, s, NCH<sub>3</sub>), 1.90 (2H, m, 2 x H2');  $\delta_{\rm c}$  (100 MHz; CD<sub>3</sub>OD): 151.3, 150.8, 148, 134.8, 126, 124.4, 122.7, 117.2, 98.1, 58.7, 55.3, 41.2, 40.9, 40.8 and 25.4; MS(ESI) m/z 312.1(M<sup>+</sup>+H).

#### 1.7 General procedure for the synthesis of compounds 14a-b

To the solution of 2-methyl-4/5-nitroimidazole (0.385 mmol) and **13a/13b** (0.385 mmol) in dry DMF(20ml) at room temperature was added anhydrous  $K_2CO_3(1.15 \text{ mmol})$ . The reaction mass was then stirred for 6 hours at 100-110 °C. On completion, DMF was evaporated and the crude dissolved in MeOH: EtOAc mixture (1:8 ratio), dried over MgSO<sub>4</sub>, and concentrated. The obtained crude was then purified by column chromatography, eluting with 15-20% MeOH in EtOAc, to afford compounds **14a-b** as regioisomeric mixture.

2[{2-[(7-Chloroquinolin-4-yl)amino]ethyl}-(methyl)amino]ethyl[2-(2-methyl-4/5nitroimidazole-1-yl (14a)

Yield (72mg) 60%; R<sub>f</sub> (DCM: MeOH, 8:2) 0.38; δ<sub>H</sub> (400 MHz; DMSO): 8.34(1H, d, *J* 5.4 Hz, H2), 8.27 (1H, s, H5'), 8.19 (1H, d, *J* 9 Hz, H5), 7.74 (1H, d, *J* 2.2 Hz, H8), 7.39 (1H, d, *J* 2.1 and 9 Hz, H6), 6.30 (1H, d, *J* 5.6 Hz, H3), 4.07 (2H, t, *J* 6 Hz, 2 x H4'), 3.12 (2H,

t, *J* 6.5 Hz, 2 x Hi'), 2.66 (2H, t, *J* 6.1 Hz, 2 x H4'), 2.43 (2H, t, *J* 6.7 Hz, 2 x H3'), 2.34 (3H, s, NCH<sub>3</sub>), 2.23 (3H, s, CH<sub>3</sub>);  $\delta_c$  (100 MHz; DMSO): 152.3, 150.5, 149.5, 145.5, 133.8, 129.3, 128.5, 127.9, 124.5, 124.4, 122.9, 117.9, 98.9, 56.9, 55.3, 44.8, 42.2 and 11.4; MS(ESI) m/z 389.2(M<sup>+</sup>+H); HPLC purity: 93.8%; t<sub>r</sub>=8.26 min.

2[{3-[(7-Chloroquinolin-4-yl)amino]propyl}-(methyl)amino]ethyl[2-(2-methyl-4/5nitroimidazole-1-yl (14b)

Yield (0.080g) 51%; R<sub>f</sub> (DCM: MeOH, 8:2%) 0.32;  $\delta_{\rm H}$  (400 MHz; CD<sub>3</sub>OD): 8.33(1H, d, *J* 5.6 Hz, H2), 8.03(1H, s, H6'), 7.98 (1H, d, *J* 9 Hz, H5), 7.73 (1H, d, *J* 2.1 Hz, H8), 7.34 (1H, dd, *J* 2.1 and 9 Hz, H6), 6.37 (1H, d, *J* 5.6 Hz, H3), 5.44 (1H, bs, NH), 4.08(2H, t, *J* 6 Hz, 2 x H5'), 3.22 (2H, t, *J* 6.9 Hz, 2 x H1'), 2.74 (2H, t, *J* 6 Hz, 2 x H4'), 2.54 (2H, *J* 6.7 Hz, 2 x H3'), 2.36 (3H, s, NCH<sub>3</sub>), 2.32 (3H, s, CH<sub>3</sub>), 1.80 (2H, m, 2 x H2');  $\delta_{\rm c}$  (100 MHz; CD<sub>3</sub>OD): 151.4, 150.7, 147.8, 145.5, 135, 125.9, 124.6, 122.8, 122.7, 121, 117.2, 98.1, 56.7, 55.2, 44.6, 40.9, 40.7, 25.6 and 11.4; MS(ESI) m/z 403.1(M<sup>+</sup>+H); HPLC purity: 98.4%; t<sub>r</sub>=8.48 min.

#### 1.8 General procedure for the synthesis of compounds 15a-b

Following the synthetic protocol used in synthesizing **14a/14b**, intermediate **11a/11b** (0.498 was used in synthesizing regioisomeric mixture of compound **15a-b**.

(7-Chloroquinolon-4-yl)-[2-(2-methyl-4/5-nitroimidazol-1-yl)-ethyl]-amine (15a)

Yield (0.115 g) 70%; R<sub>f</sub> (DCM: MeOH, 8:2%) 0.34;  $\delta_{\rm H}$  (300 MHz; CD<sub>3</sub>OD): 8.40 (1H, d, *J* 5.7 Hz, H2), 8.06 (1H, s, H3'), 8.02 (1H, d, *J* 9 Hz, H5), 7.83 (1H, d, *J* 2.1 Hz, H8), 7.48 (1H, dd, *J* 2.1 and 9 Hz, H6), 6.65 (1H, d, *J* 5.7 Hz, H3), 4.37 (2H, t, *J* 7.1 Hz, 2 x H2'), 3.38 (2H, t, *J* 6.6 Hz, 2 x H1'), 2.29 (3H, s, CH3);  $\delta_{\rm c}$  (100 MHz; DMSO): 153.5, 150.2, 149.1, 145.9, 145.6, 134.9, 127.8, 125, 124.2, 122.9, 117.8, 100.1, 47.3, 43.4 and 13.6; MS(EI) m/z 331.2(M<sup>+</sup>); HPLC purity: 94.2%; t<sub>r</sub>'=9.6 min.

Yield (0.158 g) 72%; R<sub>f</sub> (DCM: MeOH, 8:2%) 0.30;  $\delta_{\rm H}$  (400 MHz; CD<sub>3</sub>OD): 8.36 (1H, d, *J* 5.7 Hz,H2), 8.09 (1H, s, H4'), 8.06 (1H, d, *J* 8.9 Hz, H5), 7.79 (1H, d, *J* 2.1 Hz, H8), 7.41 (1H, dd, *J* 2.1 and 8.9 Hz, H6), 6.54(1H, d, *J* 5.7 Hz, H3), 4.17 (2H, t, *J* 7.1 Hz, 2 x H3'), 3.48 (2H, t, *J* 6.6 Hz, 2 x H1'), 2.37 (3H, s, CH3), 2.26 (2H, m, 2 x H2');  $\delta_{\rm c}$  (100 MHz; CD<sub>3</sub>OD): 151, 150.8, 149.9, 145.2, 135, 131.2, 126, 124.7, 122.6, 120.3, 117.2, 98.3, 44.5, 39.4, 28 and 11.2; MS(EI) m/z 345.1(M<sup>+</sup>); HPLC purity: 97.3%; t<sub>r</sub>=8.43 min.

#### 1.9 General procedure for the synthesis of compound 16

The solution of 2-nitroimidazooxazine alcohol (0.174 mmol) in dry DMF (5 ml) under  $N_2$  was cool to -50°C, NaH (0.174 mmol) was added slowly, and the reaction mass stirred for 15-20 minutes at this temperature. Intermediate **11b** (0.159 mmol) was then added, the temperature of the resultant mass raised to room temperature and further stirred overnight. On completion, DMF was removed in *vacuo*, and the crude diluted with H<sub>2</sub>O (10 ml) followed by extraction with EtOAc(3 x 20m l). The combined organic extract was washed with brine, dried over MgSO<sub>4</sub> and concentrated. The resultant crude was purified by column chromatography, eluting with 35% MeOH in DCM, to give the title compound **16** as yellow solid.

(6S)-2-Nitro-6[(7-chloroquinolon-4-aminopropyl)oxy]-6,7-dihydro-5H-imidazo[2,1b][1,3]oxazine (**16**)

Yield (0.015g) 27%; m. p. 115-116 °C; R<sub>f</sub> (DCM: MeOH, 8:2%) 0.45;  $\delta_{\rm H}$  (300 MHz; CD<sub>3</sub>OD): 8.28(1H, d, *J* 6 Hz, H2), 8.02 (1H, d, *J* 9 Hz, H5), 7.74 (1H, d, *J* 2.1 Hz, H8), 7.66 (1H, s, H7'), 7.40 (1H, dd, *J* 2.1 and 9 Hz, H6), 6.49 (1H, d, *J* 6 Hz, H3), 5.44 (bs, 1H, NH), 4.65 (1H, dt, *J* 2.3 and 2.4 Hz, H4'), 4.42 (1H, d, *J* 12 Hz, H5a'), 4.26-4.10 (3H, m, H5'a, H6'), 3.77 (2H, m, 2 x H3'), 3.43 (2H, t, *J* 6.8 Hz, 2 x H1'), 2.04 (2H, m, 2 x H2');  $\delta_{\rm c}$ 

(100 MHz; CD<sub>3</sub>OD): 152.2, 148.9, 147.7, 145.9, 136, 125.2, 124.5, 123, 116.8, 116.2, 98.1, 67.9(2C), 67.5, 66.4, 46.7, 40.1 and 28.2; MS(EI) m/z 403.1( $M^+$ ); HPLC purity: 98.1%;  $t_{r'}$ =10.12 min.

#### 1.10 Structures of quinoline diamines 17a-c



## 2. In vitro assays 2.1 Antimalarial assays<sup>3</sup>

In vitro activity against erythrocytic stages of *P. falciparum* was determined using a <sup>3</sup>Hhypoxanthine incorporation assay, using the chloroquine and pyrimethamine resistant K1 strain that originate from Thailand and the standard drug chloroquine (Sigma C6628). Compounds were dissolved in DMSO at 10 mg/mL and added to parasite cultures incubated in RPMI 1640 medium without hypoxanthine, supplemented with HEPES (5.94 g/l), NaHCO<sub>3</sub> (2.1 g/l), neomycin (100 U/mL), Albumax<sup>R</sup> (5 g/l) and washed human red cells A<sup>+</sup> at 2.5% haematocrit (0.3% parasitaemia). Serial drug dilutions of eleven three-fold dilution steps covering a range from 100 to 0.002 µg/ml were prepared. The 96-well plates were incubated in a humidified atmosphere at 37 C; 4% CO<sub>2</sub>, 3% O<sub>2</sub>, 93% N<sub>2</sub>. After 48 h 50 µl of <sup>3</sup>H-hypoxanthine (=0.5 µCi) was added to each well of the plate. The plates were incubated for a further 24 h under the same conditions. The plates were then harvested with a Betaplate<sup>TM</sup> cell harvester (Wallac, Zurich, Switzerland), and the red blood cells transferred onto a glass fibre filter then washed with distilled water. The dried filters were inserted into a plastic foil with 10 mL of scintillation fluid, and counted in a Betaplate<sup>TM</sup> liquid scintillation counter (Wallac, Zurich, Switzerland). IC<sub>50</sub> values were calculated from sigmoidal inhibition curves by linear regression using Microsoft Excel.

#### 2.2 Antimycobacterial assays

#### Culture preparation of the mycobacterium

*Mycobacterial tuberculosis* H<sub>37</sub>Rv drug-sensitive strain was grown on Middlebrook 7H10 (Merck) supplemented with 0.5% glycerol and 10% Middlebrook Oleic acid Dextros catalase (OADC) enrichment (Merck) for five days at 37 °C, without shaking. To prepare the suspension for inoculation, the cultures were vortexed, left for 45 seconds to allow heavy particles to settle, and followed by dilution.

#### **Broth microdilution method assay**

All minimum inhibitory concentrations (MICs) were determined by the broth microdilution method.<sup>4</sup> A stock culture of Mtb H<sub>37</sub>RvMA<sup>5</sup> was grown to OD<sub>600</sub> o.6 - 0.7 in Middlebrook 7H9 broth (Difco) supplemented with 0.05% Tween-80, 0.2% glycerol, and albumin/NaCl/ glucose (ADC) complex. The culture was diluted 1:500 in 7H9-based medium before aliquoting 50  $\mu$ L into each well of a 96-well plate (Rows 2-12). The compounds were dissolved in DMSO to make stock solutions of 12.8 mM and were diluted in 7H9-based medium to a final concentration of 640  $\mu$ M. 100  $\mu$ L of each compound was added to the first row of wells of the 96-well plate. After pipet mixing and use of a multichannel pipet, 50  $\mu$ L was removed from each well in the first row and added to the second row. 2-Fold dilution in this manner was carried out to give ten dilutions of each compound assayed (160-0.078  $\mu$ M). Rifampicin and kanamycin were used as positive controls, while 5% DMSO and 7H9-based medium were

employed as negative controls. The plates were incubated for 2 weeks at 37 °C, and the  $MIC_{99}$  values were read macroscopically using an inverted plate reader at Day 7 and Day 14 post inoculation.  $MIC_{99}$  values were recorded as the lowest concentration of compound that resulted in inhibition of growth of more than 99% of the bacterial population at Day 7.

## 2.3 Cytotoxicity assays<sup>6</sup>

Assays were performed in 96-well microtiter plates, each well containing 100 µl of RPMI 1640 medium supplemented with 1% L-glutamine (200mM) and 10% fetal bovine serum, and 4000 L-6 cells (a primary cell line derived from rat skeletal myoblasts). Serial drug dilutions of eleven three-fold dilution steps covering a range from 100 to 0.002 µg/mL were prepared. After 70hours of incubation the plates were inspected under an inverted microscope to assure growth of the controls and sterile conditions. 10µl of Alamar Blue was then added to each well and the plates incubated for another 2 hours. Then the plates were read with a Spectramax Gemini XS microplate fluorometer (Molecular Devices Cooperation, Sunnyvale, CA, USA) using an excitation wave length of 536 nm and an emission wave length of 588 nm. The IC50 values were calculated by linear regression (Huber 1993) from the sigmoidal dose inhibition curves using SoftmaxPro software (Molecular Devices Cooperation, Sunnyvale, CA, USA).

### **3** References

1. Jiricek, J.; Patel, S.; Keller, T. H.; Barry III, C. E.; Dowd, C. S. Nitroimidazole compounds. *PCT Int. Appl.* (2008), WO20080275035.

2. Tukulula, M.; Little, S.; Gut, J.; Rosenthal, P. J.; Wan. B.; Franzblau, S. G.; Chibale, K. The design, synthesis, *in silico* ADME profiling, antiplasmodial and antimycobacterial evaluation of new arylamino quinoline derivatives. *Eur. J. Med. Chem.* Available online September 7, **2012**; doi:10.1016/j.ejmech.2012.08,047.

3. (i) Desjardins, R. E.; Canfield, C. J.; Haynes, J. D.; Chulay. J. D. Quantitatuve assessment of antimalarial activity *in vitro* by semiautomated microdilution technique. *Antimicrob. Agents Chemother.* **1979**, *16*, 710-716; (ii) Thaithong, S.; Beale, G. H.; Chutmongkonkul, M. Variability in drug susceptibility amongst clones and isolates of *Plasmodium falciparum. Trans. Royal Soc. Trop. Med. Hygiene*, **1988**, *77*, 33-36.

4. (i) Domenech, P., Reed, M. B.; Barry III, C. E. Contribution of the *Mycobacterium tuberculosis* MmpL protein family to virulence and drug resistance. *Infect. Immun.* **2005**, *73*, 3492-501; (ii) Ioerger, T. R.; Feng, Y.; Ganesula, K.; Chen, X.; Dobos, K. M.; Fortune, S.; Jacobs, Jr., W. R.; Mizrahi, V.; Parish, T.; Rubin, E.;. Sassetti, C; Sacchettini, J. C. Variation among genome sequences of H37Rv strains of *Mycobacterium tuberculosis* from multiple laboratories. J. Bacteriol. **2010**, *192*, 3645-3653; (iii) Leite, C. Q.; Beretta, L. A.; Anno, I. S.; Telles, M. A. Standardization of broth microdilution method for *Mycobacterium tuberculosis*. *Mem Inst Oswaldo Cruz*, **2000**, *95*, 127-129.

5. Kim, P.; Zhang, L.; Manjunatha, U. H.; Singh, R.; Patel, S.;. Jiricek, J.; Keller, T. H.; Boshoff, H. I.; Barry III, C. E.; Dowd, C. S. Structure-activity relationships of antitubercular nitroimidazoles. 1. Structural features associated with aerobic and anaerobic activities of 4- and 5-nitroimidazoles. *J. Med. Chem.* **2009**, 52, 1317-1328.

6. (i) Page, C. M.; Noel, C. A new fluorometric assay for cytotoxicity measurements *in vitro. J. Oncology*, **1993**, *3*, 473-476; (ii) Ahmed, S. A.; Gogal, R. M.; Walsh, J. E. A new rapid and simple non-radioactive assay to monitor and determine the proliferation of lymphocytes: An altenative to [3H]thymidine incorporation assay. *J. Immunol. Methods*, **1994**, 170, 211-224.

## 4. NMR spectra for compounds 8-16



ppm (t1)

















ppm (t1)























