

# Evaluation and Structural Basis for the Inhibition of Tankyrases by PARP inhibitors

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## Content:

**Experimental procedures** contains details of the used proteins, assays and crystallization

**Supplementary Table 1** contains details of the compound library.

**Supplementary Table 2** contains details of the crystallographic data and refinement.

## EXPERIMENTAL PROCEDURES

**Chemical collection.** Tested compounds were purchased from Alexis Biochemicals, Sigma-Aldrich, Calbiochem, JS Research Chemicals Trading, Maybridge and Selleck Biochemicals. Details of the tested compounds are provided in Supplementary Table 1. The purity of all the compounds was  $\geq 95\%$  based on GC, HPLC, NMR or TLC analysis by the suppliers. When such information was not available, the compounds were identified and the purity was verified with HPLC.

**Protein Expression.** The expression and purification of ARTD5 (residues 1030-1317) and ARTD6 (residues 873-1161) for biochemical assays and ARTD6 (residues 946-1162) for crystallography was carried out as previously reported.<sup>1-3</sup>

**Activity assay.** Enzymatic assays were conducted as reported earlier using automodification reaction of tankyrases with 500 nM NAD<sup>+</sup> as a substrate.<sup>1</sup> Compounds were screened at 10  $\mu$ M and 500 nM concentrations in triplicate and the best compounds were selected for further studies. The dose response curves were measured for the selected compounds. The observed fluorescence values were used for fitting or when intrinsic compound fluorescence was observed at high concentrations the fluorescence was normalized to a conversion percentage using the controls. The data points were measured in quadruplicate and all IC<sub>50</sub> values were measured at least three times and fitted individually.

**Crystallization, Data collection and Refinement.** The ARTD6 catalytic domain was crystallized as previously reported.<sup>2</sup> The crystals were soaked in the precipitant solution supplemented with 250 mM NaCl and the inhibitors (**16**, **18**: 100  $\mu$ M, **10**: 1 mM, **17**: 5 mM or **29**: 10 mM). Crystals were incubated for 24 hours (**16**, **17**, **18**) or 90 days (**10**, **29**) and were dipped into a cryosolution containing additional 20% glycerol and 250 mM NaCl before flash cooling in liquid nitrogen. Data and refinement statistics are shown in Supplementary Table 2.

**Supplementary Table 1.** PARP inhibitor like compounds tested.

No.	Compound names	SMILES	Supplier
1 <sup>4</sup>	3AB	<chem>NC(=O)c1cccc(N)c1</chem>	Alexis Biochemicals
2 <sup>5</sup>	Niacinamide, Nicotinamide	<chem>NC(=O)c1cccnc1</chem>	Alexis Biochemicals
3 <sup>4</sup>	Benzamide	<chem>NC(=O)c1ccccc1</chem>	Alexis Biochemicals
4 <sup>6</sup>	Veliparib, ABT-888	<chem>C[C@@]1(CCCN1)c1nc2c(cccc2[nH]1)C(N)=O</chem>	Alexis Biochemicals
5 <sup>7</sup>	5-aminoisoquinolinone, 5-AIQ	<chem>Nc1cccc2c1cc[nH]c2=O</chem>	Alexis Biochemicals
6*	3-methyl-5- aminoisoquinolinone, 3-methyl-5-AIQ	<chem>Cc1cc2c(N)cccc2c(=O)[nH]1</chem>	Alexis Biochemicals
7 <sup>5</sup>	4-amino-1,8- naphthalimide, 4-ANI	<chem>Nc1ccc2C(=O)NC(=O)c3cccc1c23</chem>	Alexis Biochemicals
8 <sup>7</sup>	DPQ	<chem>O=C1NCCc2c(OCCCCN3CCCCC3)cccc12</chem>	Alexis Biochemicals
9 <sup>8</sup>	DR2313	<chem>Cc1nc(=O)c2CSCCc2[nH]1</chem>	Alexis Biochemicals
10 <sup>9</sup>	EB-47	<chem>Nc1ncnc2n(cnc12)[C@@H]1O[C@@H]([C@@H](O)[C@H]1O)C(=O)N1CCN(CC(=O)Nc2cccc3C(=O)NCc23)CC1</chem>	Alexis Biochemicals
11 <sup>5</sup>	4-Hydroxyquinazoline, 4-HQN	<chem>O=c1nc[nH]c2ccccc12</chem>	Alexis Biochemicals
12 <sup>10</sup>	INH2BP	<chem>Nc1ccc2oc(=O)ccc2c1</chem>	Alexis Biochemicals
13 <sup>5</sup>	1,5-Isoquinolinediol, DHQ	<chem>Oc1cccc2c1cc[nH]c2=O</chem>	Alexis Biochemicals
14 <sup>11</sup>	Minocin, Minocycline	<chem>CN(C)[C@H]1[C@@H]2C[C@@H]3Cc4c(ccc(O)c4C(O)=C3C(=O)[C@]2(O)C(=O)C(=C(/N)O)C1=O)N(C)C</chem>	Alexis Biochemicals
15 <sup>12</sup>	NU1025	<chem>Cc1nc(=O)c2ccc(O)c2[nH]1</chem>	Alexis Biochemicals
16 <sup>5</sup>	Phenanthridinone	<chem>O=c1[nH]c2ccccc2c2ccccc12</chem>	Alexis Biochemicals
17 <sup>13</sup>	PJ-34	<chem>CN(C)CC(=O)Nc1ccc2[nH]c(=O)c3ccccc3c2c1</chem>	Alexis Biochemicals
18 <sup>14</sup>	TIQ-A	<chem>O=c1[nH]c2sccc2c2ccccc12</chem>	Alexis Biochemicals
19 <sup>15</sup>	Paraxanthine, 1,7- dimethylxanthine	<chem>Cn1cnc2[nH]c(=O)n(C)c(=O)c12</chem>	Sigma-Aldrich
20 <sup>16</sup>	CNQ	<chem>NC(=O)c1cccc2ncc(nc12)-c1ccc(Cl)cc1</chem>	Calbiochem

No.	Compound names	SMILES	Supplier
21 <sup>17</sup>	Olaparib, KU-0059436, AZD-2281	<chem>Fc1ccc(Cc2n[nH]c(=O)c3ccccc23)cc1C(=O)N1CCN(CC1)C(=O)C1CC1</chem>	JS Research Chemicals Trading
22 <sup>18</sup>	BYK204165	<chem>Cn1cccc1\C=C1/C(=O)NC(=O)c2ccccc12</chem>	Sigma-Aldrich
23 <sup>19</sup>	XAV939	<chem>FC(F)(F)c1ccc(cc1)-c1nc(=O)c2CSCCc2[nH]1</chem>	Maybridge
24 <sup>8</sup>	RF03877	<chem>Oc1nc(nc2CCSCc12)-c1cccn1</chem>	Maybridge
25 <sup>20</sup>	RF03876	<chem>Oc1nc(nc2CCSCc12)C1CC1</chem>	Maybridge
26 <sup>21</sup>	IWR-1	<chem>O=C(Nc1cccc2cccnc12)c1ccc(cc1)N1C(=O)[C@H]2[C@@H]3C[C@@H](C=C3)[C@H]2C1=O</chem>	Sigma-Aldrich
27 <sup>21</sup>	IWP-2	<chem>Cc1ccc2nc(NC(=O)CSc3nc4CCSc4c(=O)n3-c3ccccc3)sc2c1</chem>	Sigma-Aldrich
28 <sup>22</sup>	Iniparib, BSI-201	<chem>NC(=O)c1ccc(l)c(c1)N(=O)=O</chem>	Selleck Biochemicals
29 <sup>23</sup>	AG014699, Rucaparib	<chem>CNCc1ccc(cc1)-c1[nH]c2cc(F)cc3C(=O)NCCc1c23</chem>	Selleck Biochemicals
30 <sup>24</sup>	UPF1035	<chem>O=C(Oc1cccc2c1cc[nH]c2=O)c1ccccc1</chem>	Alexis Biochemicals
31 <sup>24</sup>	UPF1069	<chem>O=C(COc1cccc2c1cc[nH]c2=O)c1ccccc1</chem>	Alexis Biochemicals
32 <sup>5</sup>	Flavone	<chem>O=c1cc(oc2ccccc12)-c1ccccc1</chem>	Sigma-Aldrich

**Supplementary Table 2.** Data and refinement statistics.

<b>Compound</b>	<b>10</b>	<b>16</b>	<b>17</b>	<b>18</b>	<b>29</b>
<b>PDB code</b>	4BJ9	4AVU	4BJB	4AVW	4BJC
<b>Data quality</b>					
Beamline	Diamond I04-1	ESRF ID14-1	ESRF ID23-2	ESRF ID14-1	Diamond I04-1
Wavelength (Å)	0.92	0.93340	0.87260	0.93340	0.92
Space group	C222 <sub>1</sub>	C222 <sub>1</sub>	P 4 <sub>1</sub> 2 <sub>1</sub> 2	C222 <sub>1</sub>	P 4 <sub>1</sub> 2 <sub>1</sub> 2
Cell dimensions a, b, c (Å)	90.56, 97.78, 120.03	93.37, 96.69, 116.33	66.33, 66.33, 121.39	91.57, 97.99, 117.95	65.30, 65.30, 122.91
Resolution (Å)	30-2.05 (2.10-2.05)	67- 2.40 (2.46-2.40)	30-2.30 (2.36-2.30)	67- 2.15 (2.21-2.15)	30-2.20 (2.26-2-20)
R <sub>merge</sub>	0.091 (0.742)	0.126 (0.587)	0.123 (1.032)	0.115 (0.579)	0.111 (0.879)
I / σI	13.1 (2.3)	9.7 (2.4)	18.0 (3.0)	10.2 (2.4)	15.6 (2.6)
Completeness (%)	99.8 (99.8)	99.7 (99.7)	99.9 (100.0)	99.2 (98.9)	98.6 (99.4)
Redundancy	6.4 (6.0)	3.7 (3.7)	10.2 (10.5)	3.7 (3.7)	12.6 (12.0)
CC1/2 (%)	99.8 (75.4)	99.1 (71.7)	99.8 (86.1)	99.3 (73.3)	99.9 (83.4)
<b>Refinement</b>					
Reflections	32035	19819	12007	27534	13240
R <sub>work</sub> / R <sub>free</sub>	0.171 / 0.197	0.182 / 0.244	0.199 / 0.255	0.187 / 0.228	0.195 / 0.247
<b>B-factors</b>					
Protein	35.2	20.8	39.9	20.6	43.3
Compound	54.9	16.7	53.0	13.9	41.4
Zn	32.7	20.3	41.0	19.9	42.0
SO <sub>4</sub>	45.0	31.8	49.9	33.5	69.6
Water	40.6	18.8	37.7	21.6	44.5
Glycerol	37.4	35.7	46.3	24.4	-
PEG	-	45.9	-	43.4	-
<b>R.m.s.d.</b>					
Bond lengths (Å)	0.010	0.012	0.005	0.011	0.005
Bond angles (°)	1.4	1.5	1.0	1.5	1.0
<b>Ramachandran plot</b>					
Most favorable regions (%)	98.0	98.0	96.4	98.0	97.4
Additionally allowed regions (%)	2.0	2.0	3.6	2.0	2.6

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§ This study