Supplementary Information

First synthetic analogues of diphosphoinositol polyphosphates: interaction with PP-InsP₅ kinase

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Table of Contents

Enzyme assays	S2
Protein expression, purification, crystallisation and structure determination	S2
Data collection and structure refinement statistics	S3
Electron density maps	S4
General chemistry methods	S5
Synthesis and characterisation of compounds 1 to 11	S 6
References	S30

Enzyme Assays. Due to the reversibility of inositol pyrophosphate kinases,¹ the dephosphorylation of 1.5-[PP]₂-InsP₄ by 2.5 µg mL⁻¹ hPPIP5K2^{KD} (residues 1-366) was assayed for some experiments: The enzyme was incubated at 24 °C for 30 min with buffer containing 20 mM Tris-HCl, pH 7.5, 10 mM MgCl₂, 0.1 mM ADP and 0.1 μ M 1,5-[PP]₂-InsP₄ in a 20 μ L assay. The generated ATP was measured by luciferase assay (Invitrogen Catalog number A22066). In other experiments, the reactions were assayed in the forward (ATP-consuming) direction. Either InsP₆ or 5-PA-InsP₅ (40 μ M) was incubated with 20 μ g mL⁻¹ hPPIP5K2^{KD} (residues 1–366) for the indicated times at 37 °C in 20 mM Hepes-NaOH pH 7.2, 50 mM KCl, 1 mg mL⁻¹ bovine serum albumen, 1 mM EDTA disodium salt, 1mM ATP disodium salt, 3 mM MgSO₄, 1 mM DTT, 20 µg mL⁻¹ hPPIP5K2^{KD} in a 250 µL reaction. Following quenching at 100 °C for 3 min, reactions were cooled on ice and then centrifuged at 4 °C, $15,000 \times g$ for 5 min. Supernatants were concentrated in a SpeedVac to approximately 50 µL, then loaded onto a pre-run 16 cm PAGE gel with Orange G loading dye. After electrophoresis, reaction products were stained with Toluidine Blue.² Each lane contained 10 nmol of InsP₆ or 5-PA-InsP₅.

Protein Expression, Purification, Crystallization and Structure Determination. The catalytic domain of hPPIP5K2 (residues 1–366 and 41–366) was sub-cloned, expressed and purified as before.³ The catalytic domain of PPIP5K2 (residues 41– 366) was crystallized by hanging drop vapor diffusion against a well buffer of 12% (w/v) PEG 3350, 20 mM MgCl₂, 0.1 M HEPES, pH 7.0, 1 mM AMPPNP and 2 mM CdCl₂ at 4 °C. The crystals were then soaked with 2 mM compound **2** in a stabilizing buffer containing 22% (w/v) PEG 3350, 10 mM MgCl₂, 0.1 M sodium acetate, pH 5.2 at 4 °C for 3 days. Cryosolvent was prepared by adding 33% ethylene glycol into the soaking buffer. Diffraction data were collected using APS beamlines 22-ID. All data were processed with the program HKL2000.⁴ The structure was determined using rigid body and direct Fourier synthesis, and refined with the equivalent and expanded test sets. The structure was further manually rebuilt with COOT⁵ and refined with PHENIX⁶ and REFMAC⁷ from the CCP4 package. Ligand topology and parameter files were prepared using the PRODRG server.⁸ The molecular graphics representations were prepared with the program PyMol (Schrödinger, LLC). Atomic coordinates and structure factors have been deposited with the Protein Data Bank with accession code 4GB4.

Data collection	
Space group	P2 ₁ 2 ₁ 2 ₁
Cell dimensions (a,b, c (Å))	110.8, 41.5, 89.7
Resolution (Å)*	50-1.9 (1.93)
Rsym [*]	0.062 (0.386)
I/σI [*]	24.6 (3.4)
Completeness (%)*	98.3(100.0)
Redundancy *	5.6
Refinement	
Resolution(Å)*	33.23-1.9 (1.95)
No. reflections	30869
$R_{ m work}^{*}$	19.2 (20.6)
$R_{\rm free}^{*}$	22.2 (26.4)
No. atoms	
Protein	2692
Ligand/ion	74
Solvent	330
B-factors (Å2)	
Protein	28.8
Ligand/ion	43.0
Solvent	41.3
R.m.s. deviations	
Bond length(Å)	0.01
Bond Angle (°)	1.23

Data collection and structure refinement statistics

* The numbers in parentheses are for the highest resolution shell.



Electron density maps. Stereo view of simulated-annealing omit difference map (F_o - F_c) (green) is contoured at 2.5 σ and $2F_o$ - F_c map (blue, contoured at 1.3 σ).

General Chemistry Methods

Chemicals were purchased from Sigma-Aldrich, Acros, Alfa Aesar or Fluka and used without further purification. Anhydrous solvents from Sigma-Aldrich were used without further treatment. TLC was performed on precoated plates (Merck Aluminum sheets silica 60 F₂₅₄, art No. 5554). Chromatograms were visualised under UV light and by dipping plates into either phosphomolybdic acid in EtOH or alkaline KMnO₄ solution, followed by heating. Ion exchange chromatography was performed on an LKB-Pharmacia Gradifrac medium pressure ion-exchange chromatograph using Q Sepharose Fast Flow resin and a gradient of 0 to 100% 2.0 moldm⁻³ aqueous triethylammonium bicarbonate (TEAB). Proton ¹H NMR, COSY, HMBC and HMQC spectra were recorded on Bruker Avance III (400 MHz and 500 MHz) spectrometers. Proton chemical shifts are reported in ppm (δ) relative to internal tetramethylsilane (TMS, δ 0.0 ppm) or with the solvent reference relative to TMS employed as the internal standard (DMSO-d₆: 2.50 ppm; D₂O: 4.79 ppm). The following abbreviations are used to describe resonances: br, broad; s, singlet; d, doublet; dd, double doublet; q, quartet; m, multiplet; t, triplet. ¹³C and DEPT spectra were recorded on Bruker Avance III (100 MHz and 126 MHz) spectrometers with complete proton decoupling. Carbon chemical shifts are reported in ppm (δ) relative to TMS with the respective solvent resonance as the internal standard. ³¹P NMR NMR spectra were recorded on a Bruker Avance III (162 MHz) spectrometer with complete proton decoupling. Phosphorus chemical shifts are reported in ppm (δ) relative to an 85% H₃PO₄ external standard (H₃PO₄, δ 0.0 ppm). Melting points were determined using a Reichert-Jung Thermo Galen Kofler block or a Stanford Research Systems Optimelt MPA100 automated melting point system and are uncorrected. Microanalysis was carried out at the University of Bath microanalysis service. Mass spectra were recorded at the SERC Mass Spectrometry Service Centre, Swansea, and at the University of Bath on VG Autospec or MicroTOF instruments. Flash column chromatography was performed on an ISCO CombiFlash Rf automated flash chromatography system using RediSep Rf disposable flash columns.

1,6:3,4-bis-[O-(2,3-dimethoxybutane-2,3-diyl)]-myo-inositol (4)

To a stirred suspension of myo-inositol (25.0 g, 139 mmol) in MeOH (250 mL) was added trimethyl orthoformate (100 mL), butanedione (25 mL, 285 mmol) and (±)-10camphorsulphonic acid (0.5 g). The mixture was heated under N₂ at reflux for 96 h and the resulting cherry-red suspension was allowed to cool before triethylamine (1 mL) was added. The precipitate was filtered off, washed with MeOH (200 mL) and allowed to dry, giving crude diol 4 as a white solid (22.4 g, 85% pure by ¹H NMR). This material was crystallised from boiling CHCl₃/MeOH (1:1 v/v, 650 mL) and dried under vacuum at 60 °C to give 4 as colourless crystals (15.4 g, 37.7 mmol, 27%); $R_{\rm f}$ 0.24 (EtOAc); R_f 0.29 (2:1 CHCl₃-acetone); mp > 300°C with sublimation and decomposition; ¹H NMR (DMSO-d₆, 400 MHz) δ 1.17 (6 H, s, CH₃), 1.18 (6 H, s, CH₃), 3.13 (6 H, s, OCH₃), 3.15 (6 H, s, OCH₃), 3.25 (1 H, dt, *J* = 5.6, 9.3 Hz, H-5), 3.35 (2 H, dd, J = 10.2, 2.4 Hz, H-1 and H-3), 3.67 (2 H, dd, J = 9.8, 9.8 Hz, H-4 and H-6), 3.76 (1 H, dt, J = 4.6, 2.4 Hz, H-2), 4.99 (1 H, d, J = 4.8 Hz, OH-2), 5.05 (1 H, d, J = 5.6 Hz, OH-5); ¹³C NMR (DMSO-d₆) δ 17.59 (Me), 17.60 (Me), 47.08 (OMe), 47.36 (OMe), 67.59 (C-2), 68.37 (C-1 and C-3), 69.16 (C-5), 69.31 (C-4 and C-6), 98.40 (BDA quaternary C), 98.96 (BDA quaternary C); HRMS (m/z) [M + Na]⁺ calcd. for C₁₈H₃₂O₁₀ 431.1888; found 431.1880; Anal. Calcd for C₁₈H₃₂O₁₀; C 52.93, H, 7.90; found C 52.6, H, 7.92.





2-O-benzyl-1,6:3,4-bis-[O-(2,3-dimethoxybutane-2,3-diyl)]-myo-inositol (5)

To a stirred suspension of diol (20.4 g, 50 mmol, dried in vacuo at 60 °C) in dry DMF (200 mL) was added sodium hydride (2.4 g of a 60% suspension in oil, 60 mmol). After 30 min, the mixture was cooled to 0 °C and benzyl bromide (6.5 mL, 55 mmol) was added dropwise over 30 min. The mixture was allowed to reach room temperature and stirring was continued for a further 18 h. Water (5 mL) was added and the mixture was concentrated under reduced pressure. The residue was taken up in CH₂Cl₂ (300 mL) and washed with water (300 mL). The cloudy organic layer was dried over MgSO₄, filtered and concentrated to give an off-white solid (~23 g). The residue was taken up in CH₂Cl₂, adsorbed onto silica (70 g) and purified by flash chromatography (0 to 100% EtOAc in petroleum ether) to give pure 2-O-benzyl ether 5 as a white solid (13.6 g, 27.3 mmol, 55%). Crystals from boiling EtOH, m.p. 214-216 °C; ¹H NMR (CDCl₃, 400 MHz) δ 1.31 (12 H, s, CH₃), 2.40 (1 H, d, J 2.0 Hz, OH-5), 3.23 (6 H, s, OCH₃), 3.28 (6 H, s, OCH₃), 3.54 (2 H, dd, J 10.2, 2.4 Hz, H-1 and H-3), 3.66 (1 H, td, J 9.4, 2.0, Hz, H-5), 3.81 (1 H, t, J 2.4 Hz, H-2), 4.08 (2 H, dd, J 10.0, 9.8 Hz, H-4 and H-6), 4.86 (2 H, s, OCH₂Ph), 7.22-7.26 (1 H, m, Ph), 7.29–7.34 (2 H, m, Ph), 7.50–7.53 (2 H, m, Ph); ¹³C NMR (CDCl₃, 101 MHz) δ 17.69 and 17.76 (CH₃), 47.90 and 47.95 (OCH₃), 69.09 (C-1 and C-3), 69.39 (C-4 and C-6), 70.55 (C-5), 73.79 (PhCH₂O), 76.19 (C-2), 99.13 (BDA quaternary C), 99.62 (BDA

quaternary C), 126.93 (*para-C* of *PhC*H₂O), 127.64 (*C*H of Ph), 127.84 (*C*H of Ph), 139.60 (*ipso-C* of *Ph*CH₂O); MS *m/z* (+ve ion FAB, relative intensity); 499.3 [(M + H)⁺, 20%], 497.3 (30), 488.3 (80), 467.3 (50), 101.1 (60), 91.1 [(C_7H_7)⁺, 100%]; HRMS (*m/z*) [M + Na]⁺ calcd. for C₂₅H₃₈O₁₀; 521.2357; found 521.2337; Anal. Calcd for C₂₅H₃₈O₁₀; C 60.23, H, 7.68; found C 60.3, H, 7.44.



2-*O*-benzyl-1,6:3,4-bis-[*O*-(2,3-dimethoxybutane-2,3-diyl)]-*myo*-inositol 5-*O*-bis(benzyloxy)phosphorylacetate (6)

To a solution of 2-O-benzyl-1,6:3,4-bis-[O-(2,3-dimethoxybutane-2,3-diyl)]-mvoinositol (5) (500 mg, 1.00 mmol) and dibenzylphosphonacetic acid⁹ (352 mg, 1.10 mmol) in dry dichloromethane (10 mL) was added EDAC (230 mg, 1.20 mmol). The solution was stirred at room temperature under N₂ for 16 h. TLC (ethyl acetate/petrol 1:1) showed conversion of alcohol ($R_f 0.32$) into a less polar product ($R_f 0.42$). The solution was diluted with dichloromethane (50 mL), washed with 0.5 M HCl, saturated aqueous NaHCO₃ and brine (50 mL of each), dried over MgSO₄ and concentrated. The residue was purified by flash chromatography (ethyl acetate in petroleum ether, 0 to 100%) to give 6 as a colourless oil (757 mg, 0.945 mmol, 95%), which slowly crystallised; m.p. 135.5–137.5 °C (from chloroform/petroleum ether); ¹H NMR (CDCl₃, 400 MHz) δ 1.16 (6 H, s, CH₃), 1.28 (6 H, s, CH₃), 3.03 (2 H, d, ²J_{HP} 21.6 Hz, CH₂P), 3.15 (6 H, s, OCH₃), 3.21 (6 H, s, OCH₃), 3.61 (2 H, dd, J 10.2, 2.4 Hz, H-1 and H-3), 3.82 (1 H, t, J 2.4 Hz, H-2), 4.20 (2 H, dd, J 10.1. 9.9 Hz, H-4 and H-6), 4.85 (2 H, s, OCH₂Ph), 5.16 (1 H, t, J 9.9 Hz, H-5), 5.03-5.12 (4 H, m, POCH₂Ph), 7.23–7.34 (13 H, m, Ph), 7.51–7.54 (2 H, m, Ph); ¹³C NMR (CDCl₃, 101 MHz) δ 17.62 and 17.66 (CH₃), 34.42 (¹J_{CP} 136.8 Hz, CH₂P), 47.69 and 47.88 (OCH_3) , 67.37 (C-4 and C-6), 68.06 (${}^2J_{CP}$ 6.0 Hz, PhCH₂OP), 69.18 (C-1 and C-3), 72.43 (C-5), 74.04 (PhCH₂O), 76.12 (C-2), 99.12 (BDA quaternary C), 99.65 (BDA quaternary C), 127.06 (para-C of PhCH₂O), 127.78 (CH of Ph), 127.89 (ortho-C of PhCH₂OP), 127.92 (CH of Ph), 128.41 (meta-C of PhCH₂O), 128.56 (meta-C of *PhC*H₂OP), 135.96 (${}^{3}J_{CP}$ 6.5 Hz, *ipso-C* of *PhC*H₂OP), 139.48 (*ipso-C* of *PhC*H₂O), 164.35 (${}^{2}J_{CP}$ 5.3 Hz, C=O); ${}^{31}P$ NMR (CDCl₃, 162 MHz) δ 20.39; HRMS (*m/z*) [M + Na^{+}_{1} calcd. for $C_{41}H_{53}O_{14}P$; 823.3065; found 823.3031; analysis (calcd., found for C₄₁H₅₃O₁₄P): C (61.49, 61.5), H (6.67, 6.70).





2-O-benzyl-myo-inositol 5-O-bis(benzyloxy)phosphorylacetate (7)

To a sample of 6 (244 mg, 0.305 mmol) was added 95% aqueous TFA (3 mL). The solution was stirred at room temperature for 5 min (yellow colour of butanedione will appear within seconds; it is important to keep reaction time short to minimise monodebenzylation of the dibenzylphosphonate group) and the solvents were then rapidly removed by evaporation in vacuo (no heat) to give a solid residue. TLC (dichloromethane/methanol 10:1) showed conversion of starting material ($R_{\rm f}$ 0.70) into a major more polar product ($R_f 0.22$) together with traces of other polar products. The solid was dissolved in methanol, adsorbed onto silica (1 g) and purified by flash chromatography, eluting with a gradient of methanol (0 to 20%) in dichloromethane to give the tetraol 7 as a colourless oil (108 mg, 0.189 mmol, 62%); ¹H NMR (CDCl₃, 400 MHz) δ 3.03 (2 H, d, ²J_{HP} 21.5 Hz, CH₂P), 3.57 (2 H, dd, J 9.8, 2.7 Hz, H-1 and H-3), 3.83–3.88 (6 H, m, H-4, H-6 and 4 × OH), 3.98 (1 H, t, J 2.7 Hz, H-2), 4.83 (2 H, s, OCH₂Ph), 4.84 (1 H, t, J 9.5 Hz, H-5), 4.93–5.04 (4 H, m, POCH₂Ph), 7.22–7.34 (15 H, m, Ph); 13 C NMR (CDCl₃, 101 MHz) δ 35.21 (${}^{1}J_{CP}$ 131.4 Hz, CH₂P), 68.73 (²J_{CP} 6.5 Hz, PhCH₂OP), 71.85 (C-4 and C-6), 72.43 (C-1 and C-3), 75.38 (PhCH₂O), 78.41 (C-5), 79.11 (C-2), 127.68 (para-C of PhCH₂O), 127.82 (para-C of PhCH₂OP), 128.13 (ortho-C of PhCH₂OP), 128.41 (ortho-C of PhCH₂O), 128.68 (meta-C of *PhCH*₂O), 128.74 (*meta-C* of *PhCH*₂OP), 135.35 (${}^{3}J_{CP}$ 6.1 Hz, *ipso-C* of *PhCH*₂OP),

138.69 (*ipso-C* of *Ph*CH₂O), 165.72 (${}^{2}J_{CP}$ 5.8 Hz, C=O); ${}^{31}P$ NMR (CDCl₃, 162 MHz) δ 22.71; HRMS (*m*/*z*) [M + Na]⁺ calcd. for C₂₉H₃₃O₁₀P; 595.1704; found 595.1682.





2-O-benzyl-myo-inositol 1,3,4,6-tetrakis(dibenzylphosphate) 5-

bis(benzyloxy)phosphorylacetate (8)

To a solution of tetraol 7 (90 mg 0.157 mmol) in dry dichloromethane (3 mL) under N_2 added 5-phenyl-1*H*-tetrazole (138)mg 0.942 mmol) was and bis(benzyloxy)diisopropylaminophosphine (0.3 mL, 0.892 mmol). The mixture was stirred at room temperature for 1 h and then cooled to -78 °C, before mCPBA (57%, 380 mg, 1.25 mmol) was added. The mixture was allowed to warm to room temperature and then diluted with EtOAc (30 mL). The clear solution was washed with 10% aq. Na₂SO₃ solution (2×30 mL), dried over MgSO₄ and concentrated. The residue was purified by flash chromatography (EtOAc in petroleum ether 0 to 100%) to give 8 as a colourless oil (240 mg, 0.149 mmole, 95%); TLC (EtOAc): $R_f = 0.16$; ¹H NMR (CDCl₃, 400 MHz) δ 3.19 (2 H, d, ²J_{HP} 18.0 Hz, CH₂P), 4.25 (2 H, ddd, J 10.0, 7.8 and 2.4 Hz, H-1 and H-3), 4.73 (1 H, t, J 2.4 Hz, H-2), 4.74 (2 H, s, OCH₂Ph), 4.86–5.05 (22 H, m, H-4, H-6 and POCH₂Ph), 5.20 (1 H, t, J 9.5 Hz, H-5), 7.14–7.27 (55 H, m, Ph); ¹³C NMR (CDCl₃, 101 MHz) δ 32.97 (¹J_{CP} 151.2 Hz, CH₂P), 67.65 (${}^{2}J_{CP}$ 6.5 Hz, PhCH₂OPCH₂), 69.65–69.92 (with ${}^{3}J_{CP}$ couplings, PhCH₂OPO), 71.54 (C-5), 75.31 (broad, C1, C-3, C-4 and C-6), 76.12 (PhCH₂O), 77.16 (C-2), 127.43–128.57 (CH of Ph), 135.36–135.73 (overlapping signals with ${}^{3}J_{CP}$ couplings, *ipso-C* of *Ph*CH₂OP), 136.33 (*ipso-C* of *Ph*CH₂O), 166.15 (C=O); 31 P NMR (CDCl₃, 162 MHz) δ -1.97 [2 P, (BnO)₂P(O)O], -1.37, [2 P, (BnO)₂P(O)O], 21.10 [1 P, $(BnO)_2P(O)CH_2$]; HRMS (m/z) [M + Na]⁺ calcd. for $C_{85}H_{85}O_{22}P_5$; 1635.4113; found 1635.4103.





1,6:3,4-bis-[*O*-(2,3-dimethoxybutane-2,3-diyl)]-*myo*-inositol 5-*O*bis(benzyloxy)phosphorylacetate (9)

To a stirred suspension of finely ground 4 in dry dichloromethane (10 mL) was added a catalytic amount of DMAP (15 mg) and dibenzylphosphonoacetic acid⁹ (380 mg, 1.18 mmol) followed by DCC (248 mg, 1.20 mmol). TLC (dichloromethane/ethyl acetate 1:1) showed conversion of diol (R_f 0.18) into a UV-active product (R_f 0.30) with traces of less polar products. After 2 h, the suspension was filtered through a plug of Celite to remove DCU and unreacted 4. The filtrate was concentrated to give a white residue, which was purified by flash chromatography (ethyl acetate in dichloromethane, 0 to 100%) to give the product 9 as a white solid (521 mg, 0.733 mmol, 73% slightly contaminated with 4. A small portion of 9 was recrystallised from diisopropyl ether/chloroform, m.p. 194–196 °C; ¹H NMR (CDCl₃, 400 MHz) δ 1.17 (6 H, s, CH₃), 1.30 (6 H, s, CH₃), 2.38 (1 H, broad d, OH-2), 3.05 (2 H, d, ²J_{HP} 21.6 Hz, CH₂P), 3.17 (6 H, s, OCH₃), 3.23 (6 H, s, OCH₃), 3.63 (2 H, dd, J 10.2, 2.6 Hz, H-1 and H-3), 4.04 (1 H, broad s, H-2), 4.15 (2 H, dd, J 10.1. 9.9 Hz, H-4 and H-6), 5.04–5.13 (4 H, ABX, J_{AB} 11.8 Hz, ³J_{HP} 8.6 and 8.0 Hz, POCH₂Ph), 5.17 (1 H, t, J 9.9 Hz, H-5), 7.30–7.34 (10 H, m, Ph); 13 C NMR (CDCl₃, 101 MHz) δ 17.60 and 17.61 (CH₃), 34.42 (¹J_{CP} 137.0 Hz, CH₂P), 47.77 and 48.01 (OCH₃), 66.84 (C-4 and C-6), 68.11 (²J_{CP} 6.1 Hz, PhCH₂OP), 68.58 (C-1 and C-3), 68.83 (C-2), 71.89 (C-5), 99.29

(BDA quaternary C), 100.10 (BDA quaternary C), 127.95 (*ortho-C* of *Ph*CH₂OP), 128.48 (*para-C* of *Ph*CH₂OP), 128.60 (*meta-C* of *Ph*CH₂OP), 135.98 (${}^{3}J_{CP}$ 6.5 Hz, *ipso-C* of *Ph*CH₂OP), 164.38 (${}^{2}J_{CP}$ 5.1 Hz, C=O); 31 P NMR (CDCl₃, 162 MHz) δ 20.30; HRMS (*m/z*) [M + Na]⁺ calcd. for C₃₄H₄₇O₁₄P; 733.2596; found 733.2579; analysis (calcd., found for C₃₄H₄₇O₁₄P): C (57.46, 57.4), H (6.67, 6.63).





myo-inositol 5-O-bis(benzyloxy)phosphorylacetate (10)

To a sample of 9 (350 mg, 0.492 mmol) was added 95% aqueous TFA (5 mL). The solution was stirred at room temperature for 5 min (yellow colour of butanedione will appear within seconds; it is important to keep reaction time short to minimise monodebenzylation of the dibenzylphosphonate group) and the solvents were then rapidly removed by evaporation in vacuo (no heat) to give a solid residue. TLC (dichloromethane/methanol 8:1) showed conversion of starting material ($R_{\rm f}$ 0.80) into a more polar product ($R_{\rm f}$ 0.20) together with traces of other polar products. The solid was dissolved in methanol, adsorbed onto silica (1 g) and purified by flash chromatography, eluting with a gradient of methanol (0 to 20%) in dichloromethane to give pentaol 10 as a colourless oil, which slowly crystallised (119 mg, 0.247 mmol, 50%, m.p. 152.5–154.5 °C (from ethanol); ¹H NMR [(CD₃)₂SO, 400 MHz] δ 3.25– 3.29 (4 H, m, buried, CH₂P, H-1 and H-3), 3.60 (ddd, J 9.6, 9.6, 4.8 Hz, H-4 and H-6), 3.72–3.77 (1 H, m, H-2), 4.56 (2 H, d, J 5.8 Hz, OH-1 and OH-3), 4.60 (1 H, t, J 9.6 Hz, H-5), 4.64 (1 H, d, J 3.5 Hz, OH-2), 4.73 (2 H, d, J 4.8 Hz, OH-4 and OH-6), 5.03–5.11 (4 H, m, POCH₂Ph), 7.31–7.38 (10 H, m, Ph); ¹H NMR [(CD₃)₂SO/D₂O, 400 MHz] δ 3.26 (2 H, d, ²J_{HP} 20.8 Hz, CH₂P), 3.265 (2 H, dd, J 9.7, 2.7 Hz, H-1 and H-3), 3.58 (2 H, dd, J 9.7. 9.5 Hz, H-4 and H-6), 3.74 (t, J 2.7 Hz, H-2), 4.60 (1 H, t, J 9.6 Hz, H-5), 5.00–5.09 (4 H, m, POCH₂Ph), 7.32–7.38 (10 H, m, Ph); ¹³C NMR

[(CD₃)₂SO/D₂O, 100 MHz] δ 34.05 (¹J_{CP} 132.8 Hz, CH₂P), 67.71 (²J_{CP} 6.1 Hz, PhCH₂OP), 70.56 (C-4 and C-6), 71.53 (C-1 and C-3), 72.54 (C-2), 78.92 (C-5), 128.09 (*ortho-C* of *Ph*CH₂OP), 128.58 (*para-C* of *Ph*CH₂OP), 128.75 (*meta-C* of *Ph*CH₂OP), 136.39 (³J_{CP} 6.6 Hz, *ipso-C* of *Ph*CH₂OP), 165.33 (²J_{CP} 5.1 Hz, C=O); ³¹P NMR [(CD₃)₂SO/D₂O, 162 MHz] δ 22.25; HRMS (*m*/*z*) [M + Na]⁺ calcd. for C₂₂H₂₇O₁₀P; 505.1234; found 505.1219; analysis (calcd., found for C₂₂H₂₇O₁₀P): C (54.77, 54.5), H (5.64, 5.62).







myo-inositol 1,2,3,4,6-pentakis(dibenzylphosphate) 5bis(benzyloxy)phosphorylacetate (11)

To a stirred suspension of pentaol 10 (65 mg 0.135 mmol) in dry dichloromethane (3 mL) under N₂ was added 5-phenyl-1H-tetrazole (146 mg 1.00 mmol) and bis(benzyloxy)diisopropylaminophosphine (0.3 mL, 0.9 mmol). The mixture was stirred at room temperature for 2 h and then cooled to -78 °C, before MCPBA (57%, 400 mg, 1.32 mmol) was added. The mixture was allowed to warm to room temperature and then diluted with EtOAc (30 mL). The clear solution was washed with 10% aq. Na₂SO₃ solution (2×30 mL), dried over MgSO₄ and concentrated. The residue was purified by flash chromatography (EtOAc in petroleum ether 0 to 100%) to give **11** as a colourless oil which slowly crystallised (202 mg, 0.113 mmole, 84%); m.p. 100–102 °C (from diethyl ether/petroleum ether); TLC (EtOAc): $R_{\rm f} = 0.14$; ¹H NMR (CDCl₃, 400 MHz) δ 3.19 (2 H, d, ²J_{HP} 18.0 Hz, CH₂P), 4.31–4.38 (2 H, m, H-1 and H-3), 4.87-5.08 (24 H, m, H-4, H-6 and POCH₂Ph), 5.12-5.17 (2 H, m, POCH₂Ph), 5.25 (1 H, t, J 9.6 Hz, H-5), 5.60 (1 H, dt, J 9.1, 2.3 Hz, H-2), 7.12–7.27 (55 H, m, Ph); ¹³C NMR (CDCl₃, 101 MHz) δ 33.00 (¹J_{CP} 151.2 Hz, CH₂P), 67.68 $(^{2}J_{CP} 6.0 \text{ Hz}, \text{PhCH}_{2}\text{OPCH}_{2}), 69.75-70.15 \text{ (with } ^{3}J_{CP} \text{ couplings}, \text{PhCH}_{2}\text{OPO}), 71.38$ (C-5), 73.51 (C-1 and C-3), 74.71 (C-4 and C-6), 76.07 (C-2), 127.89-128.50 (CH of Ph), 135.54–135.69 (overlapping signals with ${}^{3}J_{CP}$ couplings, *ipso-C* of *Ph*CH₂OP), 136.33 (³J_{CP} 6.4 Hz, *ipso-C* of *Ph*CH₂OP), 166.07 (C=O); ³¹P NMR (CDCl₃, 162 MHz) δ -2.35 [1 P, (BnO)₂P(O)O], -1.55 [2 P, (BnO)₂P(O)O], -1.05 [2 P, $(BnO)_2 P(O)O]$, 20.87 [1 P, $(BnO)_2 P(O)CH_2$]; analysis (calcd., found for C₉₂H₉₂O₂₅P₆): C (61.95, 62.0), H (5.20, 5.17).





myo-inositol 1,3,4,6-tetrakisphosphate-5-phosphorylacetate (1)

To a solution of 8 (85 mg, 52 µmol) in MeOH (8 mL) and deionised water (2 mL) was added palladium hydroxide on activated charcoal (Fluka, 20%, 50% water, 50 mg). The suspension was shaken in a Parr hydrogenator under H_2 (50 p.s.i.) for 18 h. The catalyst was removed by filtration through a PTFE filter, giving a colourless solution, which was concentrated under reduced pressure. A ¹H NMR spectrum in D₂O of the residue at this stage showed no residual aromatic signals, indicating that hydrogenolysis was complete. The residue was purified by ion-exchange chromatography on Q Sepharose Fast Flow resin eluting with a gradient of aqueous TEAB (0 to 2.0 moldm⁻³) to give the triethylammonium salt of 1 (44 mg, 41 μ mol, 79%); ¹H NMR (D₂O, 500 MHz, HDO ref at 4.79) δ 1.20 (~39 H, t, J 7.4 Hz, CH₃ of TEA⁺), 2.89 (2 H, d, ²J_{HP} 20.7 Hz, CH₂P), 3.13 (~25 H, q, J 7.4 Hz, CH₂ of TEA⁺), 4.17 (2 H, ddd, J 9.8, 9.7, 2.6 Hz, H-1 and H-3), 4.30 (1 H, t, J 2.6 Hz, H-2), 4.50 (2 H, ddd, J 9.6, 9.5, 9.5 Hz, H-4 and H-6), 5.04 (1 H, t, J 9.6 Hz, H-5); ¹³C NMR (D₂O, 126 MHz) δ 8.14 (CH₃ of TEA⁺), 37.30 (¹J_{CP} 118.1 Hz, CH₂P), 46.50 (CH₂ of TEA⁺), 70.66 (C-2), 73.63 (C-5), 74.06 (J_{CP} 5.7, 2.0 Hz, C-1 and C-3), 74.67 (J_{CP} 5.7, 5.6 Hz, C-4 and C-6), 169.40 (${}^{2}J_{CP}$ 6.4 Hz, C=O); ${}^{31}P$ NMR (CD₃OD, 162 MHz) δ 0.40 (2 P,

phosphate P), 0.89 (2 P, phosphate P), 12.20 (1 P, phosphonate P); HRMS (m/z) [M]⁻ calcd. for C₈H₁₈O₂₂P₅; 620.8983; found 620.8988.





myo-inositol 1,2,3,4,6-pentakisphosphate-5-phosphorylacetate (2)

To a solution of 11 (129 mg, 72.3 µmol) in MeOH (15 mL) and deionised water (2 mL) was added palladium hydroxide on activated charcoal (Fluka, 20%, 50% water, 20 mg). The suspension was stirred vigorously under an atmosphere of hydrogen (balloon) for 16 h. The catalyst was removed by filtration through a PTFE syringe filter to give a colourless solution, which was neutralised with aqueous TEAB (1 M, 1 mL) and then concentrated under reduced pressure. A ¹H NMR spectrum in D_2O of the residue at this stage showed no residual aromatic signals, indicating that hydrogenolysis was complete. The residue was purified by ion-exchange chromatography on Q Sepharose Fast Flow resin eluting with a gradient of aqueous TEAB (0 to 2.0 moldm⁻³) to give the triethylammonium salt of 2 (89 mg, 66 μ mol, 92%); ¹H NMR (D₂O, 500 MHz, HDO ref at 4.79) δ 1.27 (~48 H, t, J 7.4 Hz, CH₃ of TEA⁺), 2.97 (2 H, d, ²J_{HP} 20.9 Hz, CH₂P), 3.18 (~31 H, q, J 7.4 Hz, CH₂ of TEA⁺), 4.30 (2 H, broad t, J 9.7 Hz, H-1 and H-3), 4.57 (2 H, ddd, J 9.6, 9.5, 9.6 Hz, H-4 and H-6), 4.87 (1 H, dt, J 9.7, 2.5 Hz, H-2), 5.14 (1 H, t, J 9.6 Hz, H-5); ¹³C NMR (D₂O, 100 MHz) δ 8.31 (CH₃ of TEA⁺), 37.53 (¹J_{CP} 119.6 Hz, CH₂P), 46.70 (CH₂ of TEA⁺), 73.49 (with J_{CP} couplings, C-1 and C-3), 73.84 (broad, C-5), 74.71 (J_{CP} 5.8, 5.8 Hz, C-4 and C-6), 75.75 (²J_{CP} 5.8 Hz C-2), 169.65 (²J_{CP} 6.5 Hz, C=O; ³¹P NMR (CD₃OD, 162 MHz) δ –0.45 (1 P, phosphate P), 0.18 (2 P, phosphate P), 0.44 (2 P, phosphate

P), 12.17 (1 P, phosphonate P); HRMS (m/z) [M]⁻ calcd. for C₈H₁₉O₂₅P₆; 700.8647; found 700.8620.





2-O-benzyl-myo-inositol 1,3,4,6-tetrakisphosphate-5-phosphorylacetate (3)

To a solution of 8 (122 mg, 75 µmol) in MeOH (12 mL) and aqueous TEAB (1.0 moldm⁻³, pH 7.5, 3 mL) was added palladium hydroxide on activated charcoal (Fluka, 20%, 50% water, 20 mg). The suspension was stirred vigorously under an atmosphere of hydrogen (balloon) for 0.5 h. The catalyst was removed by filtration through a PTFE syringe filter to give a colourless solution, which was concentrated under reduced pressure. A ³¹P NMR spectrum in CD₃OD of the residue at this stage showed only three peaks and the corresponding ¹H NMR spectrum showed that the benzylphosphate esters had been removed, while the benzyl ether was still intact. The residue was re-dissolved in deionised water and lyophilised to give the triethylammonium salt of **3** (73 mg, 59 µmol, 79%); ¹H NMR (D₂O, 400 MHz, HDO ref at 4.79) δ 1.30 (~41 H, t, J 7.4 Hz, CH₃ of TEA⁺), 2.99 (2 H, d, ²J_{HP} 20.9 Hz, CH₂P), 3.21 (~27 H, q, J 7.4 Hz, CH₂ of TEA⁺), 4.34 (2 H, ddd, J 9.7 Hz, H-1 and H-3), 4.44 (1 H, br s, H-2), 4.64 (2 H, ddd, J 9.6, 9.5, 9.6 Hz, H-4 and H-6), 4.99 (2 H, s, OCH₂Ph, 5.15 (1 H, t, J 9.6 Hz, H-5);7.40–7.44 (1 H, m, para-H of Ph), 7.47–7.51 (2 H, m, meta-H of Ph), 7.62–7.64 (2 H, m, ortho-H of Ph); 13 C NMR (D₂O, 100 MHz) δ 8.30 (CH₃ of TEA⁺), 37.50 (¹J_{CP} 118.1 Hz, CH₂P), 46.69 (CH₂ of TEA⁺), 73.99 (broad, C-5), 74.47 (with J_{CP} couplings, C-1 and C-3), 75.23 (J_{CP} 5.6, 5.6 Hz, C-4 and

C-6), 76.15 (OCH₂Ph), 79.35 (C-2), 128.04 (*para*-C of Ph), 128.43 (Ph), 128.65 (Ph), 138.49 (*ipso*-C of Ph), 169.61 (${}^{2}J_{CP}$ 6.6 Hz, C=O); ${}^{31}P$ NMR (CD₃OD, 162 MHz) δ 0.32 (2 P, phosphate), 0.78 (2 P, phosphate), 12.15 (1 P, phosphonate); HRMS (*m/z*) [M]⁻ calcd. for C₁₅H₂₄O₂₂P₅; 710.9453; found 710.9451.





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