

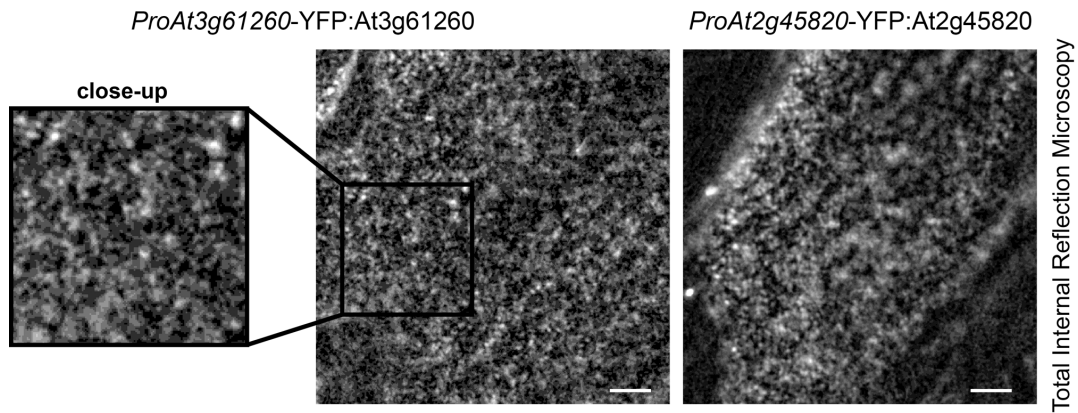
**Supplemental Figure 1 online.**

**Membrane domain localization of different Remorin proteins in *A. thaliana* and *N. benthamiana*.**

**(A)** Expression of At5g61280 under the control of its endogenous promoter in AvrPto DEX inducible *A. thaliana* resulted in labeling of distinct membrane domains. Scale bars indicate 5  $\mu\text{m}$ .

**(B-C)** Heterologous and ectopic expression of the potato REM1.3 (B) and *M. truncatula* SYMREM1 (C) in *N. benthamiana* leaf epidermal cells. Scale bars indicate 5  $\mu\text{m}$ .

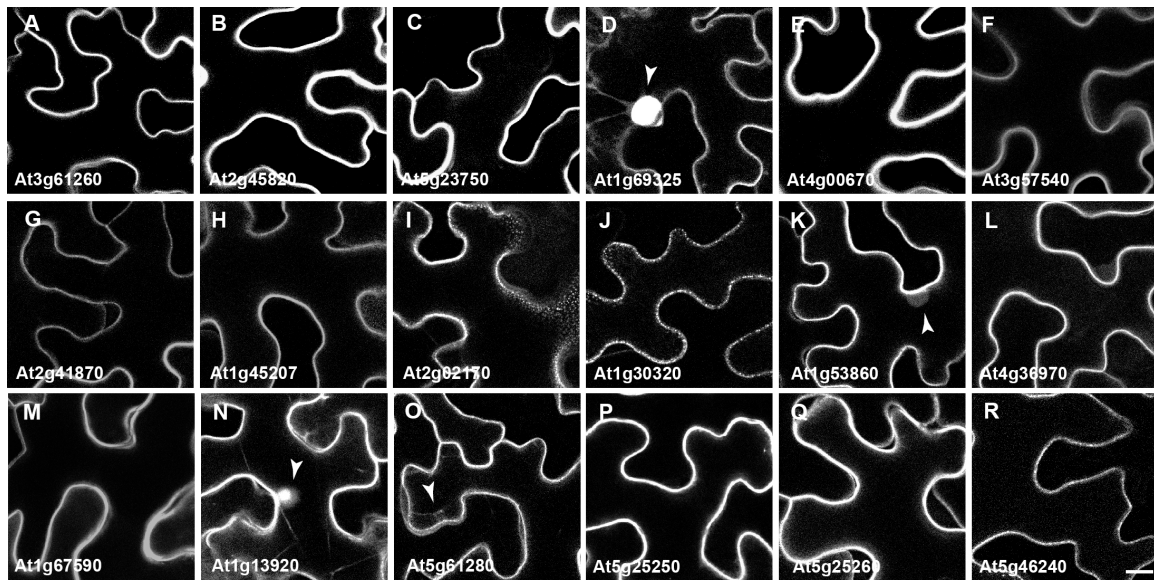
**(D-I)** Ectopic expression of different Remorins in leaf epidermal cells of transgenic *A. thaliana* plants transformed with a dexamethasone inducible AvrPto construct. Scale bars indicate 5  $\mu\text{m}$ .



**Supplemental Figure 2 online.**

**Total Internal Reflection Microscopy (TIRFM) of upper plasma membrane planes.**

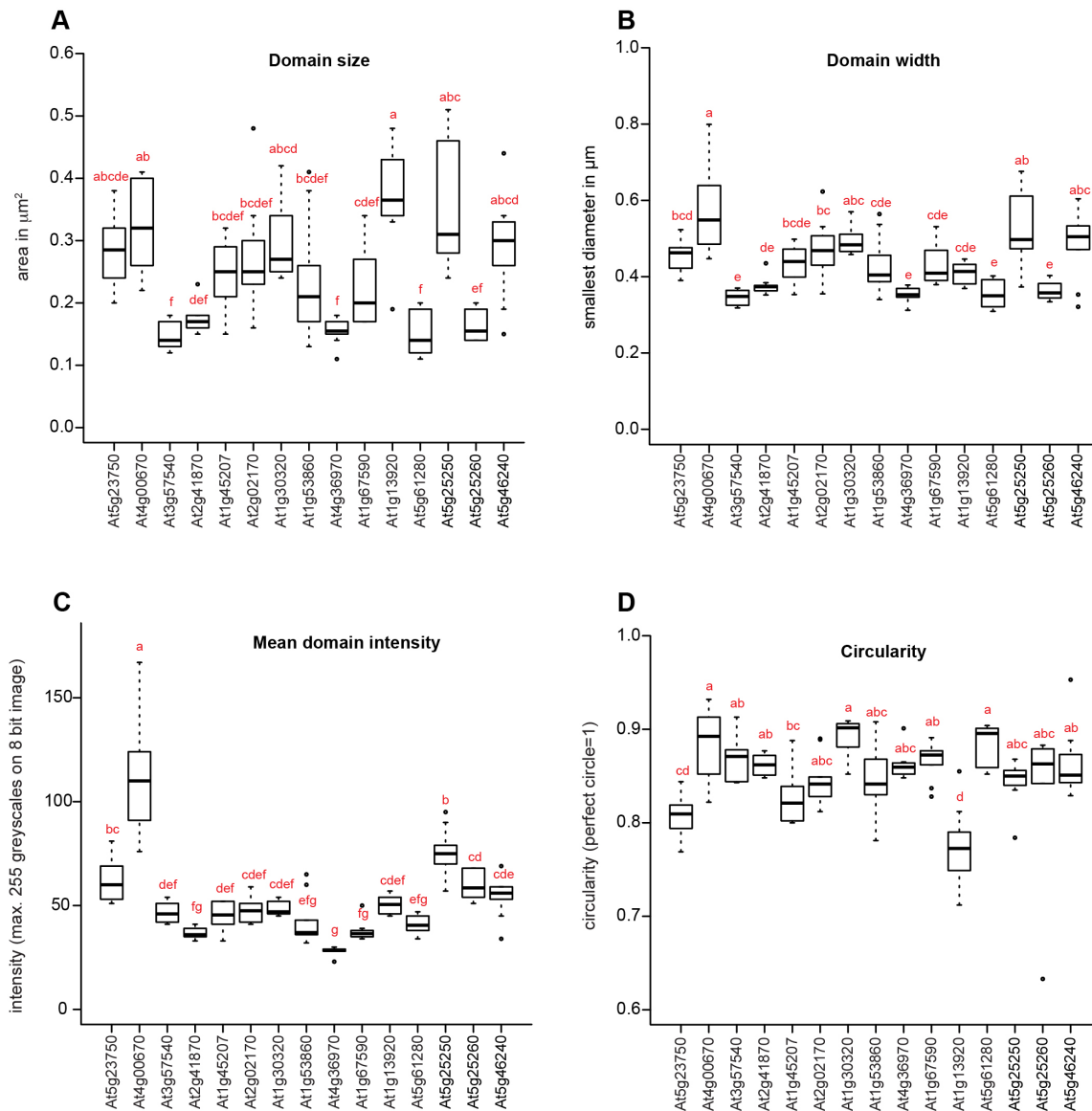
Null mutants expressing *At3g61260* and *At2g45820* as YFP fusion proteins were assessed by TIRFM. Both proteins localized to small yet highly mobile domains. Scale bars indicate 2  $\mu\text{m}$ .



**Supplemental Figure 3 online.**

**Confocal images of secant planes illustrate plasma membrane localization of all marker proteins.**

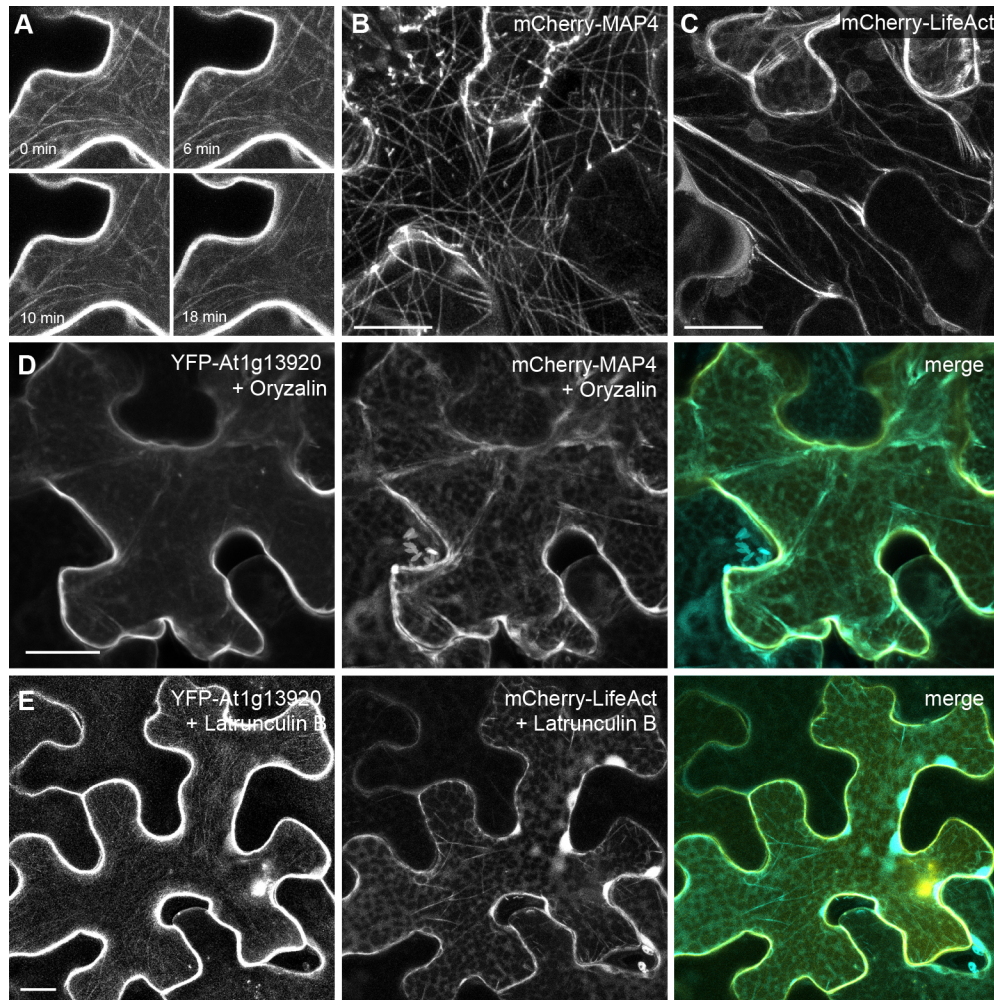
Eighteen different marker proteins were expressed in *N. benthamiana* leaf epidermal cells. Secant plane images are shown to demonstrate localization of the proteins to the plasma membrane. Arrowheads point towards partial nuclear and minor cytosolic localization of At1g69325, At1g53860, At1g13920 and At5g61280. Scale bar indicates 10  $\mu$ m.



**Supplemental Figure 4 online.**

**Quantification of membrane domain parameters of all marker proteins.**

Boxplots representing ten independent images of single protein expressing *N. benthamiana* leaf epidermal cells were used to quantify domain parameters as described in the Methods section. Letters indicate results of a one-way analysis of variance (ANOVA) followed by a Tukey HSD test.



**Supplemental Figure 5 online.**

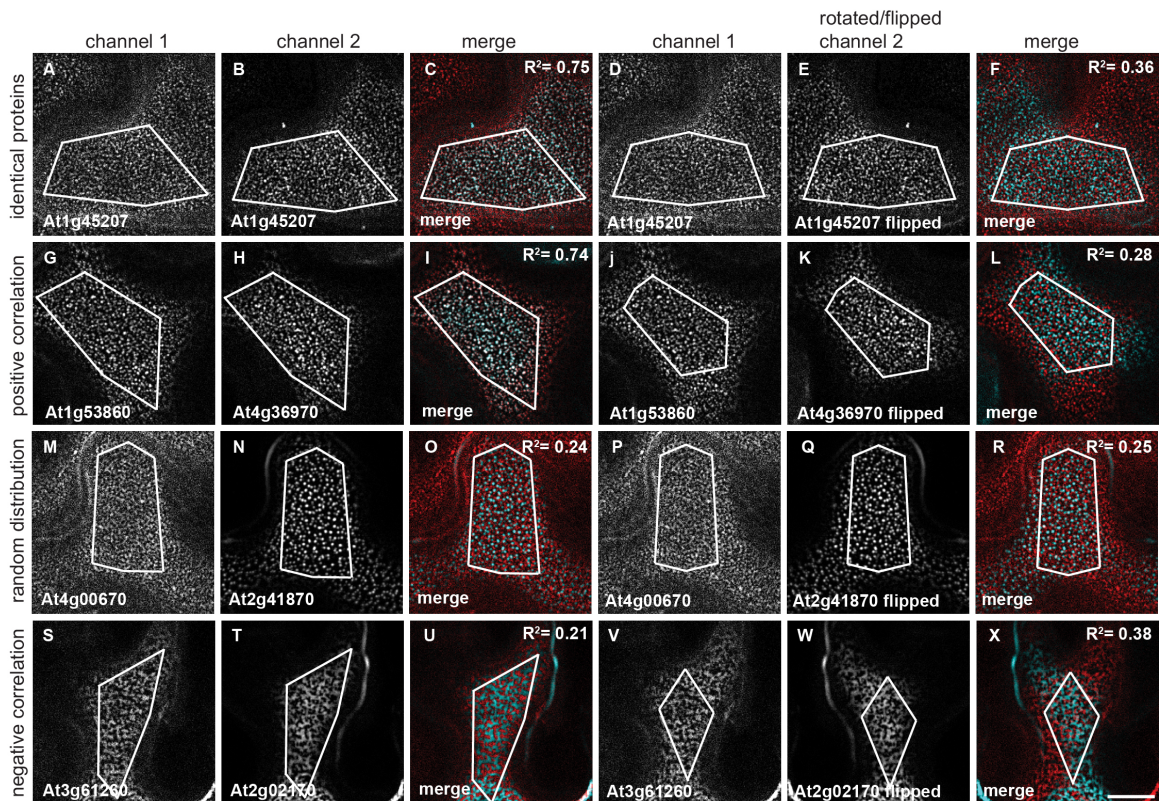
**Filament-like localization of At1g13920 is dependent on microtubules.**

(A) Time lapse experiments show that filament-like structures that are frequently labeled by At1g13920 are stable over an observation period of at least 20 minutes.

(B-C) Cells co-expressing the microtubule marker protein MAP4 (B) and the actin labeling peptide LifeAct (C) before treatment with oryzalin. Scale bars indicate 20  $\mu\text{m}$ .

(D) Oryzalin treatment depolymerized microtubules and led to a loss of the At1g13920-labeled filaments and its predominant cytosolic localization. Scale bar indicates 20  $\mu\text{m}$ .

(E) Depolymerization of the actin cytoskeleton by incubation in latrunculin B did not alter localization of At1g13920 but disrupted the majority of actin filaments. Scale bar indicates 20  $\mu\text{m}$ .



**Supplemental Figure 6 online.**

**Image processing and randomization for quantitative analysis of co-localizations.**

To make any quantitative and robust statements on co-localization between two proteins, squared overlap coefficients ( $R^2$ ) were calculated (C, I, O, U). For this, the channel 2 image was flipped or rotated (E, K, Q, W) to randomize the image.  $R^2$  values were calculated for a new region of interest that contained image information of both channels (F, L, R, X). When mean  $R^2$  values were significantly higher than the corresponding randomized values, pairs were ranked as positive correlations (co-localization). Pairs without any significant difference between these two values are randomly distributed and those where the randomized  $R^2$  was significantly higher than the corresponding original  $R^2$  value were ranked as negative correlation (mutual exclusion). Scale bars indicate 5  $\mu\text{m}$ .

**Supplemental Table 1 online.****Data from quantitative image analysis of co-localization experiments.**

Protein pairs were grouped as 'positive co-localization' when  $R^2$  was significantly higher ( $p < 0.05$ ) compared to  $R^2_{\text{random}}$ , as 'random co-localization' when  $R^2$  was not significantly different compared to  $R^2_{\text{random}}$  and as 'exclusion' when  $R^2$  was significantly lower ( $p < 0.05$ ) compared to  $R^2_{\text{random}}$ . Significant levels were calculated by Student ttest. rd= random; std.err= standard error; n= number of independent images that were analyzed.

Protein pair	sq. Manders		Pearson		n	sq. Manders		Pearson		n	ttest $R^2$	ttest Rr
	$R^2$	std. err.	Rr	std. err.		rd $R^2$	rd std. err.	rd Rr	rd std. err.			
At1g45207xAt1g45207	0,752	0,020	0,661	0,026	11	0,364	0,021	0,016	0,012	10	3,47E-11	6,20E-15
At4g00670xAt4g00670	0,718	0,022	0,617	0,038	15	0,360	0,016	0,017	0,010	19	8,04E-14	1,30E-16
At4g36970xAt4g36970	0,711	0,022	0,582	0,021	9	0,432	0,011	-0,003	0,006	11	6,46E-10	8,82E-17
At1g30320xAt1g30320	0,687	0,018	0,607	0,021	17	0,365	0,013	0,054	0,015	11	9,02E-13	5,67E-17
At2g02170xAt2g02170	0,653	0,007	0,524	0,015	13	0,352	0,008	0,002	0,008	15	1,67E-20	2,08E-22
At2g41870xAt2g41870	0,635	0,009	0,332	0,012	8	0,476	0,005	-0,005	0,008	10	1,94E-11	2,93E-12
At1g67590xAt1g67590	0,618	0,010	0,397	0,022	9	0,418	0,011	-0,015	0,010	11	1,28E-10	4,98E-13
At3g57540xAt3g57540	0,609	0,006	0,303	0,012	8	0,459	0,005	-0,003	0,006	10	7,65E-13	5,32E-14
At3g61260xAt3g61260	0,498	0,007	0,027	0,007	10	0,477	0,007	-0,017	0,004	11	3,66E-02	3,15E-05

Identical proteins

At4g36970xAt1g53860	0,737	0,024	0,698	0,032	12	0,282	0,016	0,012	0,007	12	1,52E-13	4,70E-16
At1g67590xAt2g02170	0,717	0,013	0,605	0,024	11	0,382	0,010	0,027	0,009	10	3,42E-14	5,11E-15
At4g36970xAt1g45207	0,633	0,029	0,435	0,054	11	0,412	0,015	0,030	0,008	11	1,33E-06	3,51E-07
At1g30320xAt2g02170	0,628	0,017	0,382	0,029	8	0,411	0,021	0,017	0,009	11	6,54E-07	1,37E-10
At1g67590xAt1g30320	0,572	0,025	0,487	0,036	13	0,238	0,014	-0,010	0,009	12	4,69E-11	6,11E-12
At2g41870xAt3g57540	0,569	0,007	0,217	0,014	10	0,465	0,005	-0,005	0,003	10	1,65E-10	1,21E-11
At1g53860xAt1g45207	0,513	0,012	0,222	0,014	15	0,408	0,012	-0,004	0,006	15	7,80E-07	7,68E-15
At1g67590xAt1g53860	0,492	0,013	0,217	0,016	10	0,385	0,011	-0,002	0,004	10	6,83E-06	8,75E-11
At1g53860xAt2g02170	0,446	0,008	0,251	0,010	12	0,302	0,009	-0,002	0,010	13	3,40E-11	9,97E-15
At2g02170xAt1g45207	0,434	0,009	0,286	0,009	11	0,275	0,009	0,001	0,005	11	5,84E-11	8,13E-18
At4g36970xAt3g57540	0,423	0,017	0,070	0,016	9	0,384	0,009	-0,007	0,004	21	3,70E-02	7,92E-07
At1g67590xAt3g57540	0,401	0,009	0,104	0,012	13	0,330	0,009	-0,004	0,008	9	3,44E-05	1,71E-06
At4g36970xAt1g30320	0,366	0,028	0,188	0,031	12	0,274	0,021	0,015	0,011	12	1,94E-02	5,43E-05
At1g53860xAt1g30320	0,298	0,010	0,139	0,016	10	0,238	0,014	-0,010	0,009	12	3,66E-03	3,36E-08

positive co-localization

Supplemental Table 1 online (continued)

Protein pair	sq. Manders		Pearson		n	sq. Manders		Pearson		n	sq. Manders	Pearson
	R <sup>2</sup>	std. err.	Rr	std. err.		rd R <sup>2</sup>	rd std. err.	rd Rr	rd std. err.		ttest R <sup>2</sup>	ttest Rr
At2g02170xAt3g57540	0,424	0,009	-0,002	0,005	9	0,409	0,014	-0,007	0,007	9	4,00E-01	5,60E-01
At1g45207xAt3g57540	0,417	0,009	0,039	0,006	19	0,396	0,008	-0,006	0,003	19	8,24E-02	3,84E-08
At4g36970xAt2g02170	0,381	0,013	-0,002	0,010	10	0,366	0,011	-0,005	0,004	13	3,84E-01	7,88E-01
At1g53860xAt2g41870	0,374	0,009	0,003	0,007	10	0,366	0,012	-0,005	0,006	12	5,87E-01	4,01E-01
At1g67590xAt2g41870	0,361	0,019	0,087	0,010	13	0,312	0,022	-0,005	0,006	10	1,10E-01	4,88E-08
At1g67590xAt1g45207	0,354	0,029	0,058	0,011	10	0,322	0,024	-0,011	0,003	10	3,90E-01	1,60E-05
At2g41870xAt3g61260	0,353	0,007	-0,027	0,005	12	0,356	0,009	-0,018	0,007	12	8,37E-01	2,39E-01
At1g45207xAt2g41870	0,353	0,010	0,045	0,021	12	0,333	0,010	0,012	0,008	12	3,33E-01	1,60E-01
At1g45207xAt3g61260	0,318	0,011	-0,019	0,003	11	0,319	0,012	-0,020	0,004	11	9,54E-01	8,85E-01
At2g41870xAt4g00670	0,239	0,021	-0,031	0,013	11	0,246	0,017	-0,007	0,005	11	7,88E-01	1,10E-01
At1g30320xAt1g45207	0,133	0,008	0,050	0,007	18	0,116	0,009	0,002	0,005	17	1,53E-01	1,28E-06
At1g30320xAt3g57540	0,132	0,005	0,014	0,007	12	0,122	0,005	-0,011	0,004	12	1,03E-01	5,25E-03
At2g02170xAt2g41870	0,410	0,005	-0,042	0,009	10	0,436	0,006	-0,003	0,006	10	2,89E-03	2,79E-03
At1g53860xAt3g61260	0,373	0,011	-0,117	0,011	15	0,434	0,004	-0,005	0,006	14	2,52E-05	2,52E-09
At4g36970xAt3g61260	0,296	0,014	-0,128	0,026	11	0,353	0,006	0,010	0,008	12	9,82E-04	2,84E-05
At1g67590xAt4g36970	0,286	0,012	-0,009	0,012	13	0,385	0,011	-0,002	0,004	10	6,41E-06	6,29E-01
At1g67590xAt3g61260	0,208	0,013	-0,368	0,027	10	0,357	0,008	-0,019	0,009	11	5,12E-09	8,21E-11
At2g02170xAt3g61260	0,207	0,016	-0,432	0,047	15	0,383	0,012	-0,012	0,010	14	2,29E-09	4,98E-09
At4g00670xAt3g61260	0,193	0,014	-0,136	0,017	11	0,247	0,016	0,004	0,009	11	2,00E-02	4,04E-07
At1g30320xAt4g00670	0,191	0,023	-0,153	0,021	12	0,265	0,018	0,003	0,009	12	1,70E-02	6,06E-07
At1g30320xAt3g61260	0,153	0,006	-0,106	0,032	10	0,192	0,011	-0,017	0,010	10	5,61E-03	1,64E-02
At1g67590xAt4g00670	0,102	0,011	-0,291	0,019	13	0,217	0,015	0,004	0,008	13	1,60E-06	2,46E-13

random co-localization

exclusion



## Supplemental Table 2 online.

## List of primers used in this study.

## cloning of overexpression constructs

gene ID	forward primer	reverse primer	vector
At3g61260	CACC <b>AT</b> GGCGGAGGAAACAGAAAGA	<b>T</b> TAAGAAACATCCACAAGTTGCCTT	2
At2g45820	CACC <b>AT</b> GGCGGAGGAGCAAAAAGAC	<b>T</b> TAAGAAACATCCACACGTTGCCTT	2
At5g23750	GGGGACAAGTTTGTACAAAAAAGCAGGCTCC <b>AT</b> GGCTGAAGAGGAACCG	GGGGACCACCTTTGTACAAGAAAGCTGGGTT <b>T</b> TAATGCATCCGAAAAGC	3
At1g89325	GGGGACAAGTTTGTACAAAAAAGCAGGCTCC <b>AT</b> GAACGAATCCACAGTGC	GGGGACCACCTTTGTACAAGAAAGCTGGGTT <b>T</b> CAAGGCATGTAGAGGGTTTCC	3
At4g00670	GGCGCGCTACC <b>AT</b> GGAGCCAAATATCCGATCC	CGTTTAAACC <b>T</b> TAAGAGCAGCTCAAAAGATGA	1
At3g57540	GGCGCGCTACC <b>AT</b> GTGACTTTGTACGGTCA	CGTTTAAACC <b>T</b> TAAGAAAGAGAGAAGAATGATC	1
At2g41870	GGCGCGCTACC <b>AT</b> GCTGACTCTTTACCATCAAG	CGTTTAAACC <b>T</b> TAAGAAAGAGAGAAGAAGGAGC	1
At1g45207	GGCGCGCTACC <b>AT</b> GCGTCCGAGTCATCGTAC	CGTTTAAACC <b>T</b> TAAGATACATGGCAGGTGAAGC	1
At2g02170	GGCGCGCTACC <b>AT</b> GGATTACGAACGAATCGG	GGCGCGCT <b>T</b> TAAGAAACAAAGCTAAAGC	1
At1g30320	GGCGCGCTACC <b>AT</b> GGATTACGAGAGGATACAG	CGTTTAAACC <b>T</b> TAAGAAACCAACCACAACA	1
At1g53860	GGCGCGCTACC <b>AT</b> GGACTTCAACAAGAAACAG	CGTTTAAACC <b>T</b> TAAGAAAGATATTTGC	1
At4g36970	GGGGACAAGTTTGTACAAAAAAGCAGGCTCC <b>AT</b> GAGAAAGACTTCTGTTTC	GGGGACCACCTTTGTACAAGAAAGCTGGGTT <b>T</b> CAAGAGCAGAAAGAAGATTTTC	3
At1g67590	CACC <b>AT</b> GAGATCTAGTGTAGAAG	<b>T</b> TATGACACCAACACGAG	2
At1g13920	GGGGACAAGTTTGTACAAAAAAGCAGGCTCC <b>AT</b> GGATACTTAATCAAGC	GGGGACCACCTTTGTACAAGAAAGCTGGGTT <b>T</b> CAAGAAACAGCATGCATTTTC	3
At5g61280	GGGGACAAGTTTGTACAAAAAAGCAGGCTCC <b>AT</b> GGATAATTTGGTTAAGC	GGGGACCACCTTTGTACAAGAAAGCTGGGTT <b>T</b> CAAGTAACCCGAAAGCAGAAA	3
KAT1	GGGGACAAGTTTGTACAAAAAAGCAGGCTTA <b>AT</b> GTCGATCTCTTGGACTCG	GGGGACCACCTTTGTACAAGAAAGCTGGGTT <b>T</b> ATTGATGAAAAATACAATGATCACC	3
FLOT1A	TTTGGTCTCTCACC <b>AT</b> GTTCAAAGTTGCAAGAGC	AAAGGTCTCACCTTCTGCTGCGAGTCACTTGC	4
FLOT1B	TTTGGTCTCTCACC <b>AT</b> GTTCAAAGTTGCAAGAGC	AAAGGTCTCACCTTCTGCTTAGAGTACCGATCC	4
SYMREM1	AGGCGCGCTACC <b>AT</b> GGAAAGATCGAAAAACAAC	AGGCGCGCT <b>T</b> ACTGAAAAACCTTAAACC	1

## cloning of Pro:YFP-ORF constructs

ProAt3g61260	TTTGGTCTCTCACC <b>AT</b> GGCGGCTCGTTG	AAAGGTCTCTTGTGTCAGTCGCGGCTCTCAGCC	4
At3g61260	TTTGGTCTCGAAT <b>AT</b> GGCGGAGGAACAG	AAAGGTCTCACCTTT <b>T</b> AGAAACATCCACAAGTTGC	4
ProAt2g45820	TTTGGTCTCTCACC <b>AT</b> GGCTGATCTCTTCAAAATC	AAAGGTCTCTTGTCTGTCTCTCAGCCGAAAGAAAG	4
At2g45820	TTTGGTCTCAGAA <b>AT</b> GGCGGAGGAGCAAAAAG	AAAGGTCTCACCTTT <b>T</b> AGAAACATCCACAGCTTGC	4
ProAt4g36970	TTTGGTCTCTCACC <b>AT</b> GCGTTCATCGTCTGTA	AAAGGTCTCTTGTCTGTGTTGGTTCTCAAGAAACAATAATC	4
At4g36970	TTTGGTCTCAGAA <b>AT</b> GGAAAGACTTCTGTTTC	AAAGGTCTCACCT <b>T</b> ACTGAGAGCAAGAAGATTTTC	4
free YFP	TTTGGTCTCTCAG <b>AT</b> GCTGAGCAAGGCGSAGS	AAAGGTCTCTATCTCTTGTACAGCTCTGCTCATGC	4

## qPCR primers

At4g36970	GAACATGAACAACAACAGGG	CGAGGAGCAAGCAAGTCATG
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## legend

1= for cloning into pksi

2= for cloning into pENTR-D

3= for cloning into pDONR207

4= for cloning into pENTR-D Bsal

START and STOP codons are depicted in red; Pro=promoter