

Supplementary Figure Legends

Figure S1. (a) Western blot of purified SRAP 1-236, purified SRAP 13-236 and whole cell lysates from HEK293 and HeLa cells. Probed with anti-SRAP antibody Abcam ab72552. (b) Western blot of purified SRAP 1-236, purified SRAP 13-236 and MCF-7 whole cell lysate probed with anti-SRAP antibody Abcam ab72407. (c) Histogram of RNA-seq reads around the two candidate SRAP start codons M1 and M13. Each black bar represents one nucleotide. The height of each bar is proportional to the number of reads mapped to that specific nucleotide. The positions of the two candidate start codons are marked in green.

Figure S2. Anomalous difference Fourier map. Map was computed with anomalous differences measured at 1.5 Å wavelength to 4.0 Å resolution and model phases. Cartoon drawing of B subunit of the SRAP carboxy-terminal domain, with Met and Cys side chains drawn as stick models. Sulfurs are orange. Anomalous map (magenta) contoured at 3 sigma. Molecular drawings were made with PyMOL (<http://www.pymol.org>).

Figure S3. EMSA of SRA RNA fragments by SRAP(V106M-215). Arrows show positions of unshifted RNAs. Experimental conditions same as in Figure 2 of main text.

Figure S4. Individual siRNAs in pool 1 have different effects on MCF-7 cell growth. In each case, approximately 200K MCF-7 cells were transfected with 25 nM corresponding siRNA. 3 days after transfection, cell number was examined by flow cytometry (n=2). * denotes the siRNA that has the most deleterious effect on cell growth.

Figure S5. Response of pS2 and PR expression to different levels of 17β-estradiol. Top graph, endogenous SRA RNA levels; middle and lower graphs, pS2 and PR mRNA levels, respectively. Lane 1: MCF-7 cells were grown in complete medium. Lane 2-11: MCF-7 cells were grown in hormone-free medium for 4 days and then allowed to continue for one day in hormone-free medium (Lane2), complete medium (Lane 3), hormone-free medium with 17β-estradiol and fulvestrant (Lane 4-7) or hormone-free medium with just 17β-estradiol (Lane 8-11). The RNA

levels of SRA, pS2 and PR were examined by RT-qPCR, using GAPDH mRNA as internal control (SRA primer: SRA-4).

Figure S6. Examination of the expression level of pS2 and PR at different time points after SRA knockdown. MCF-7 cells were transfected with 25 nM corresponding siRNA pool. Cells were harvested 1, 2 and 3 days after transfection. RNA level of pSR (left) and PR (right) was examined by RT-qPCR, with GAPDH mRNA as internal control (n=3).

Supplementary Tables

Table S1. Sequences of siRNAs in SRA siRNA pool 1 and their corresponding scrambled control siRNAs.

SRA siRNA	Forward (5' to 3')	Reverse (5' to 3')
Pool 1 #1 SRA siRNA	ucacuuggcuccuucuuatt	uaagaaggagccaagugatt
Pool 1 #1 control siRNA	cuuaccuuuuggucaccuctt	gaggugaccaaagguaagtt
Pool 1 #2 SRA siRNA	agagggaguucauguguuatt	uaacacaugaacuccucutt
Pool 1 #2 control siRNA	uucguuaggagauguagagtt	cucuacaucuccuaacgaatt
Pool 1 #3 SRA siRNA	ggaaaguugucaauaccugtt	cagguauugacaacuucctt
Pool 1 #3 control siRNA	ccugaauaggaagucaguutt	aacugacuuccuauucaggtt
Pool 1 #4 SRA siRNA	gguaggaguuaaaagauuatt	uaaucuuuaacuccuacctt
Pool 1 #4 control siRNA	ggagaaaggguauuuuuatt	uaauaauuacccuucucctt

Table S2. Sequences of RT-qPCR primers.

Gene	Forward (5' to 3')	Reverse (5' to 3')
GAPDH*	acagcaacaggggtgggac	gaccattgctggggctggg
SRA-1 (Exon 2)	ccgccgcagtttcatac	cttggaagcagcgagcg
SRA-2 (Exon 2,3 junction)	aggcgcctcgtgcttac	tctctgatgcggggactc
SRA-3 (Exon 3,4 junction)	gtggccacacaaggaagc	tcctccagcccactgttc
SRA-4 (Exon 4)	gacatcagccgacgcc	ctgcaccagtagagccattc
SRA-5 (Exon 4,5 junction)	ctctactggtgcaagagctttc	ccatgagggagcgggtg
SRA-6 (Exon 4,5 junction)	aggaacagtgggctggag	gtggcttgaagctcttgc
SRA-7 (Exon 5)	acatccaccgctcctc	cattggctgcctcctctg
pS2	catcgacgtcctccagaagag	ctctgggactaatcaccgtgctg
PR	cgcgctctaccctgcactc	tgaatccggcctcaggtagtt
GREB1*	gcttgccgctctagaaggt	gtgcagctggaaagggttt
MSRB2	cgcctttcagtgggatctac	ccagtgccagagcagttactt
RACGAP1	tctgtttgagcagcttgtgc	tcctctgccactttttacgg
C3orf58	cttcgcaacctcaaggactc	aaatggccaaccttcatcag
NMD3	caaccaccaggaacttgat	aaagcctgcatctacaagcc
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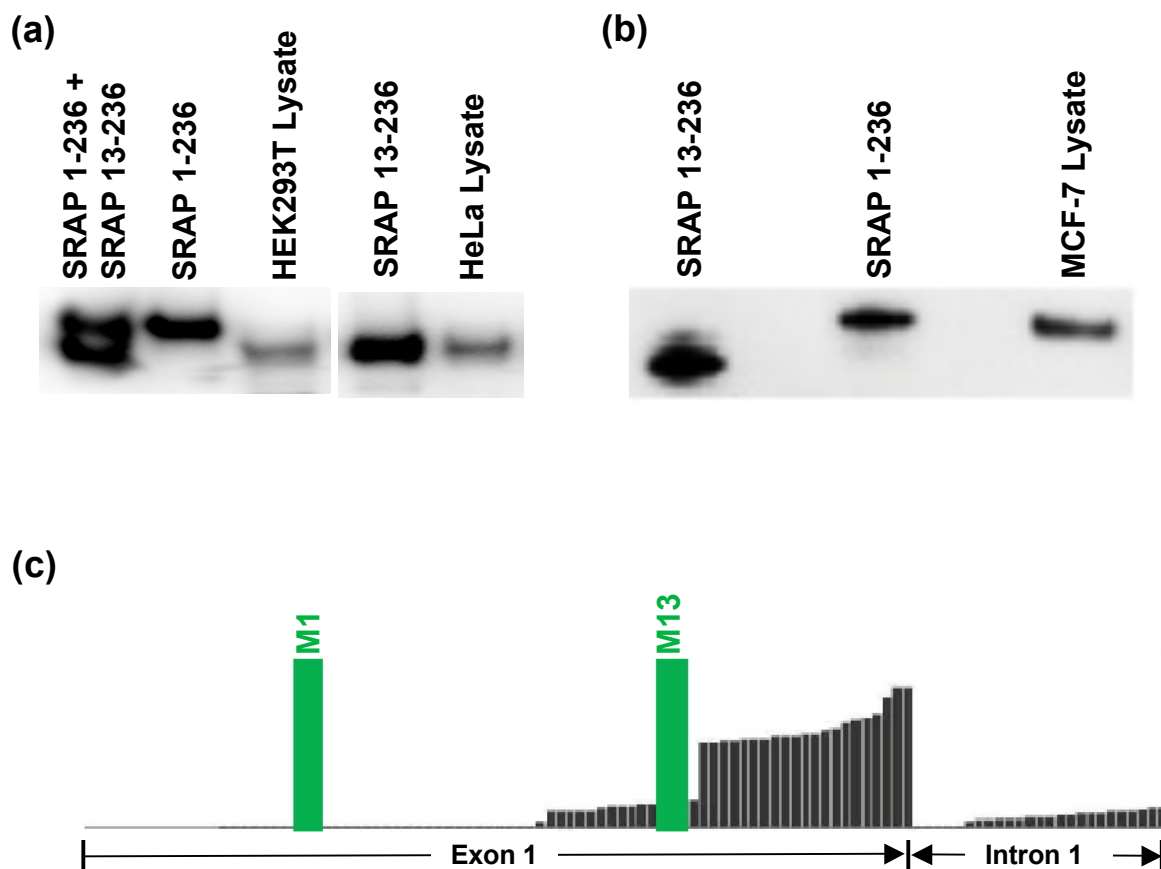
*The GAPDH primer listed was described in [1]. We also used Human GAPD (GAPDH) Endogenous Control (Applied Biosystems, 4326317E) for GAPDH RT-qPCR. The GREB1 primer listed was described in [2].

Table S3. Sequences of SRA cDNA cloning primers.

Gene	Forward (5' to 3')	Reverse (5' to 3')
Primer 1	ggcaatgcaagaggaggctgtagaagtc	gtggaagaaggagccaagtacagaag
Primer 2	gcggaagtggagatggcggag	gcataggagatggtgtccggtgagtc
Primer 3	agctgtacgtgaagccgggcaac	gcataggagatggtgtccggtgagtc

Supplementary References

1. Fleury, L., Gerus, M., Lavigne, A. C., Richard-Foy, H. & Bystricky, K. Eliminating epigenetic barriers induces transient hormone-regulated gene expression in estrogen receptor negative breast cancer cells. *Oncogene* 2008;27:4075-4085.
2. Foulds, C. E., Tsimelzon, A., Long, W., Le, A., Tsai, S. Y., Tsai, M. J. & O'Malley, B. W. Research resource: expression profiling reveals unexpected targets and functions of the human steroid receptor RNA activator (SRA) gene. *Mol Endocrinol* 2010;24:1090-1105.



Figure_S1

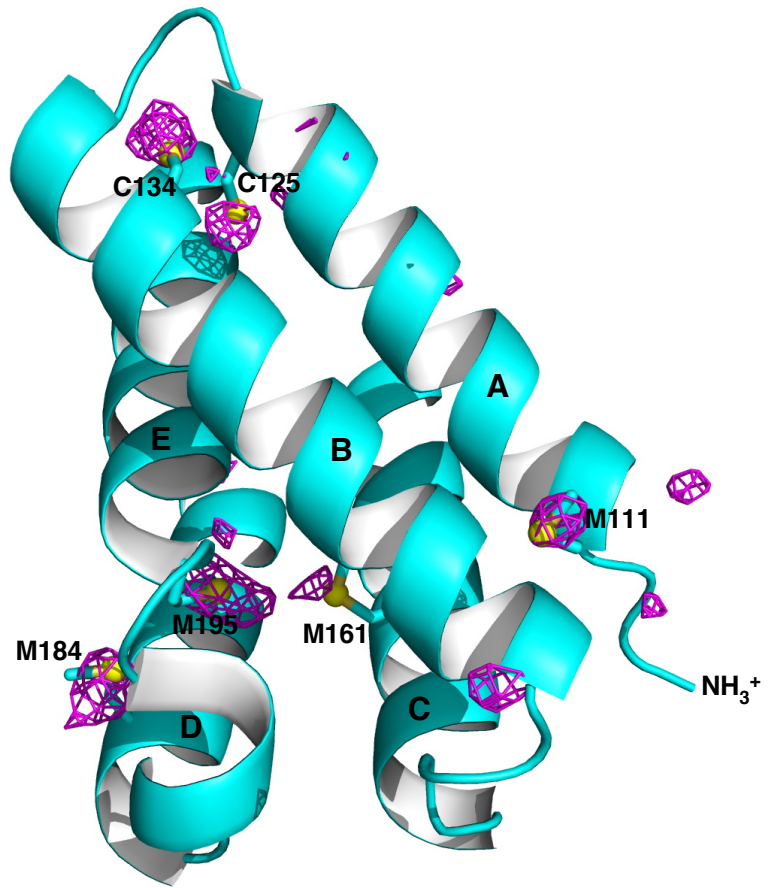
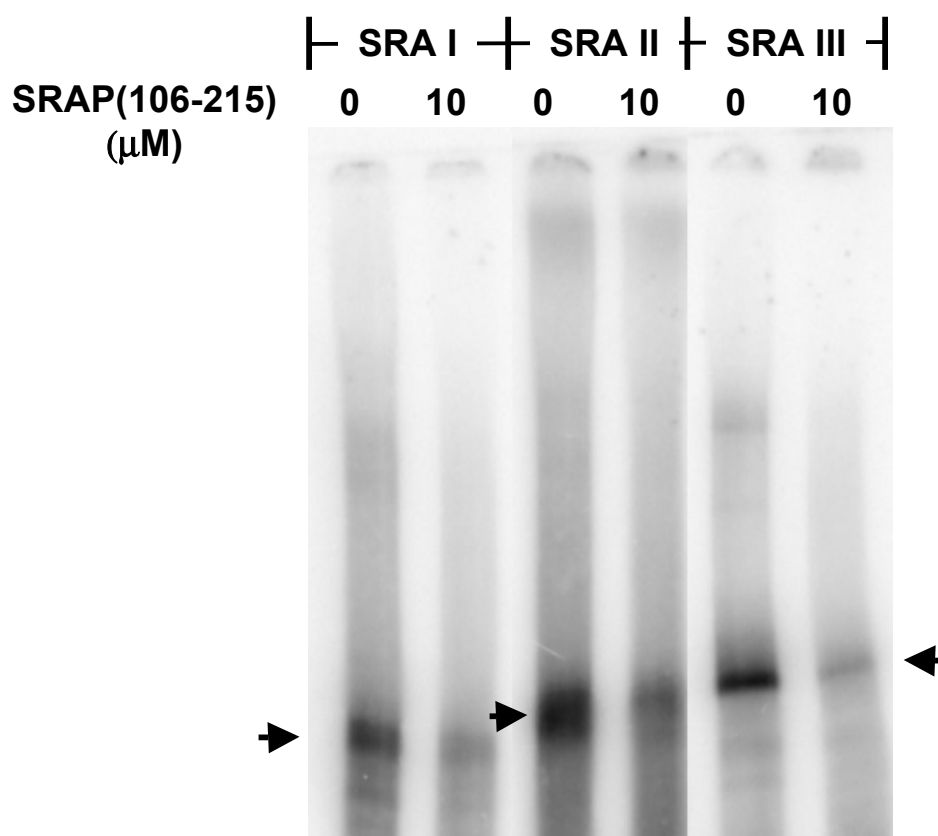


Figure S2



Fig_S3: SRA frag binding by SRAP C-terminal domain

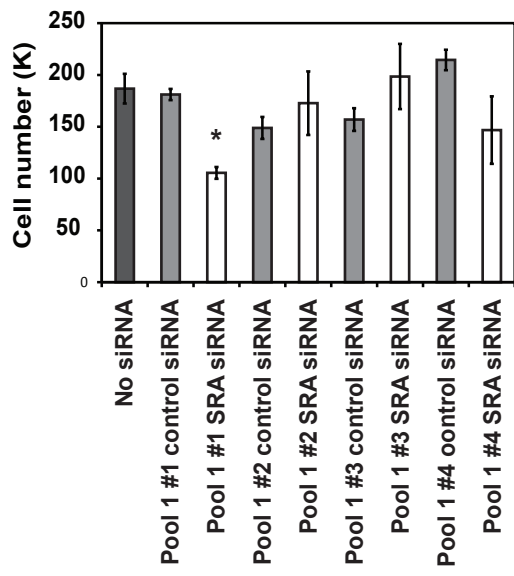


Figure S4

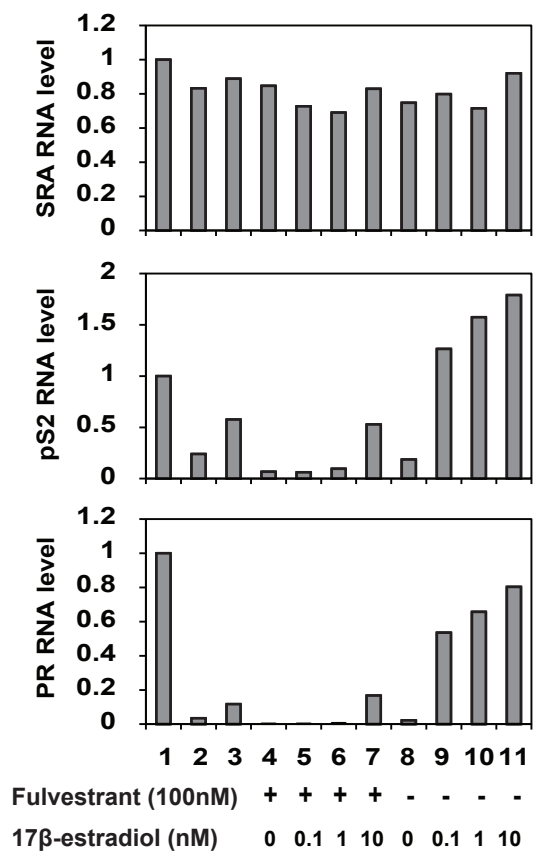


Figure S5

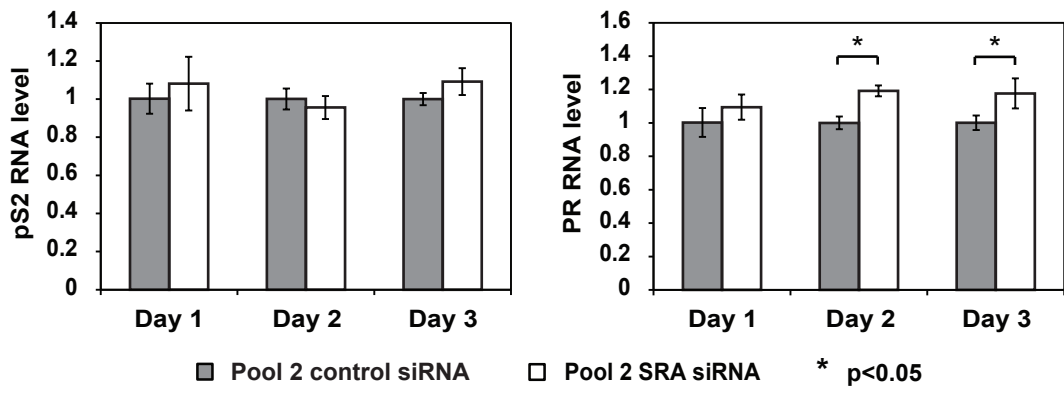


Figure S6