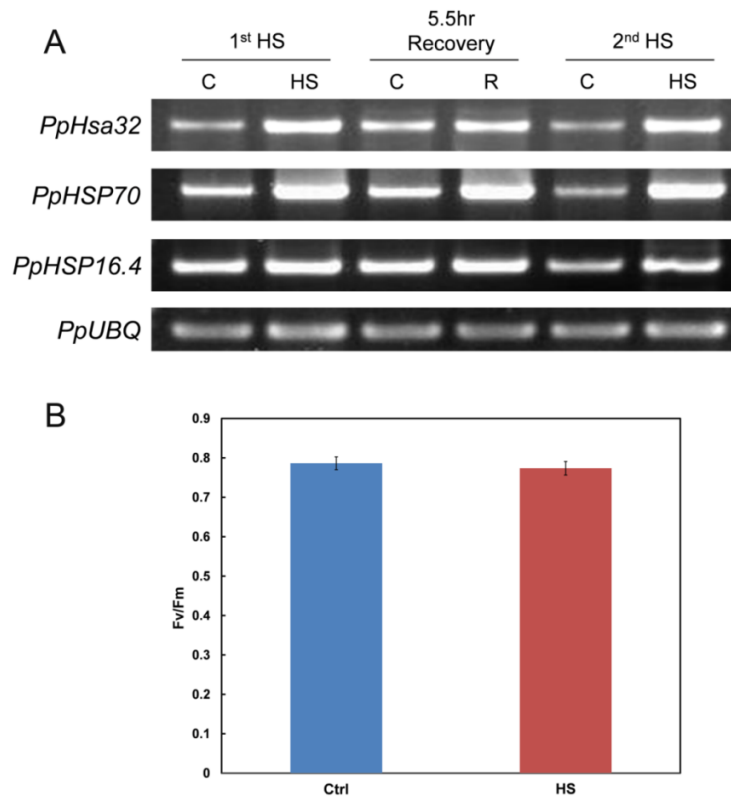


SUPPLEMENTARY FIGURES AND TABLES

Genome-wide analysis of heat-sensitive alternative splicing in *Physcomitrella patens*

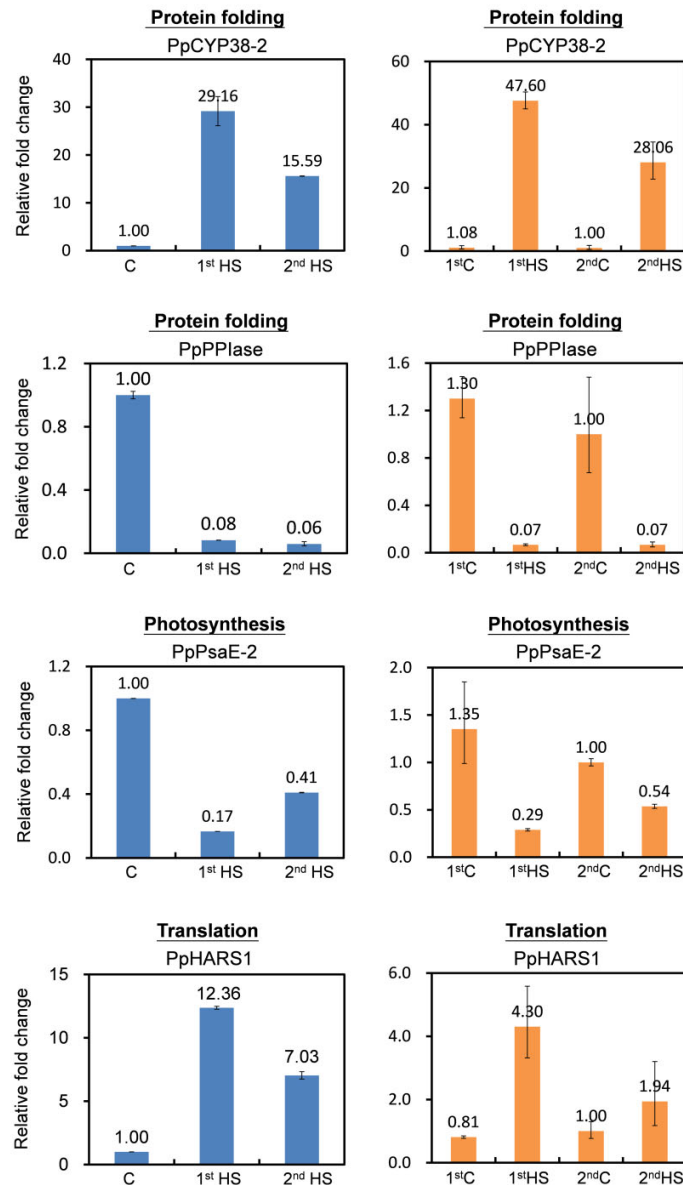
Chiung-Yun Chang, Wen-Dar Lin and Shih-Long Tu



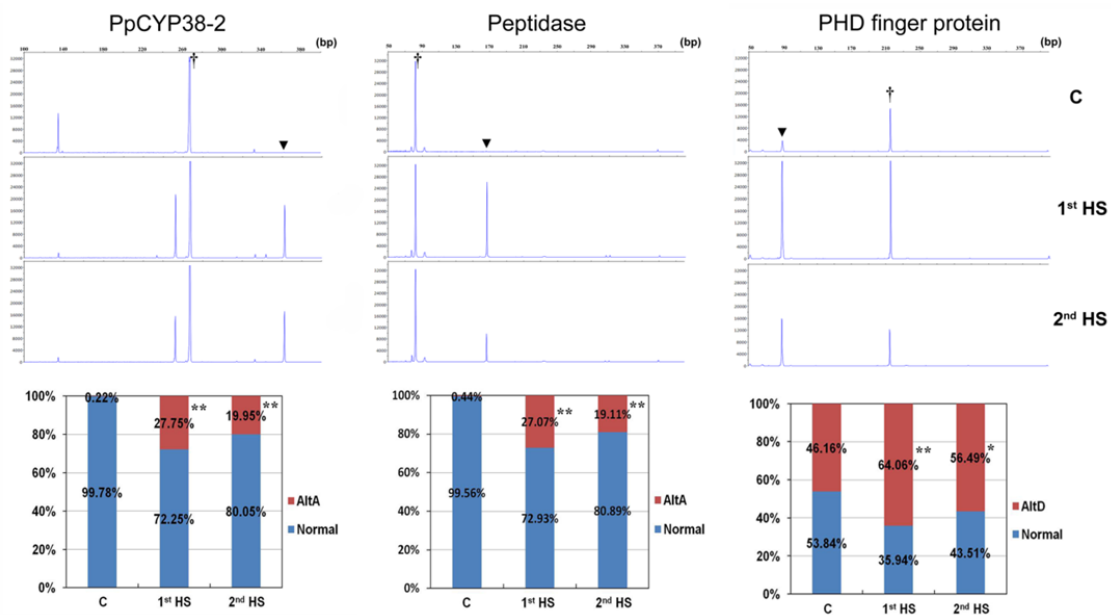
Supplemental Figure S1. Confirmation of heat shock (HS) response in *Physcomitrella*. (A) Expression of HS marker genes. Pooled RNA from control (C), 1st HS, 5.5 h recovery, and 2nd HS samples were used for RT-PCR reaction. Primer sets specific for *PpHsa32*, *PpHSP70*, *PpHSP16.4*, and *PpUBQ* transcripts were used. *PpUBQ* was used as a control. C, control; HS, HS-treated sample; R, 5.5 h recovered sample. (B) Photosynthetic efficiency of HS-treated samples. Chlorophyll *a* fluorescence of protonema was measured immediately after 2nd HS treatment. Protonema grown under normal temperature was used as the control (Ctrl).



Supplemental Figure S2. Comparison of genes differentially expressed and alternatively spliced in response to 1st HS and 2nd HS. Venn diagrams show the comparison of differential expressed genes (DEGs) and heat-sensitive IR (HS-IR), AltDA (HS-AltDA) and ES (HS-ES) genes identified in 1st HS and 2nd HS samples. Numbers of DEGs and heat-sensitive AS genes are listed under each set. Numbers of specific and overlapped genes in each diagram are indicated.



Supplemental Figure S3. Validation of HS-IR events. qRT-PCR data from pooled RNA (also in the manuscript) for 4 genes were shown in the left panels (blue bars), where “C” indicates the untreated control of 2nd HS (2nd C in the right panels). qRT-PCR data from unpooled RNAs were shown in the right panels (orange bars). *PpACT2* was first used as an internal control for normalizing each qPCR reaction. Level of the IR isoforms from three biological replicates were further normalized for overall expression level of each gene and then compared with data from the control of 2nd HS (2nd C) to generate the relative IR level. Corresponding gene products of selected IR events and representative processes are shown above each graph.



Supplemental Figure S4. Validation of heat-regulated AltD and AltA sites. High-resolution RT-PCR was used for determining the expression level of selected AltD and AltA isoforms. Pooled RNA from control (C), 1st HS and 2nd HS samples were used in triplicate for RT-PCR. Primer sets designed for amplifying the AltD and AltA regions were used. DNA fragments were separated on an ABI3730 DNA Analyzer with 3 technical repeats. Integrated peak areas of RT-PCR products identified with expected sizes were used as the relative expression level. Percentage of each AS isoform was calculated by dividing by the sum of all transcripts. Significance: ** $P < 0.01$; * $0.01 > P > 0.05$. †, PCR products from constitutive spliced isoforms; ▼, PCR products from alternatively spliced isoforms. Gene IDs and annotations are in Supplemental Dataset S9.

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                *           24           *           48           *
Pp1s19_153V6 : -----MSLSFNN-----SEAGCMLFY-----
Pp1s85_27V6 : -----MSLAFDN-----SETGCMLFY-----
Pp1s12_41V6 : -----MSLSFDN-----SETGCMLLF-----
Pp1s207_20V6 : -----MTLSLDTSC-----TEGASLLTY-----
AT-HSFB4 : -----MAMMVENSYGGYGG-----GGGERIQLMVE-----
HSFB1 : -----MTAVT-----
AT-HSFB2a : -----MNSPPVDAMITGESS-----
AT-HSFB3 : -----MEDAGEHLRCNDNVNDEERLPLEF-----MIGNSTS-----
AT-HSFB2b : ---MPGEQTGETPTVAGVGGGAGCSAGN-----SGSSGCGAGGGGGSGGGGGGGG-----
ATHSFA1d : ---MDV-----SKVTTSDGG-----GDSMETKPSQPQPAAI-----
HSFA1a : ---MFVNFKYFSFFIRTKMGDVTGGGTN-----IGEAVTAPPPrNPHPATL-----
ATHSFA1e : -----MGTVCESVATA-----KSSTAVMS-----
AT-HSFA7b : -----MDPS-----SSSRARSMPPVPME-----
ATHSFA2 : -----MEELKVEEETVFTG-----SVAASSSVGSSSSPRPME-----
AT-HSFA6b : ---MDPSFRFIKEEFPAGFSDSPSPSSSSYLYSSMAEAAINDPTTLYSQPLE-----
AT-HSFA6a : -----MDYNLPIPLE-----
AT-HSFA7a : -----MMNPF-----LPEGCDPPPPQPM-----
HSFA1b : -----MESVPEVSP-----NSNTPSIPPPVN-----
Pp1s249_84V6 : ---MVSQVLKTDPHWPPLEESMGQERYA-----LNTDVI PSSAVGSLNSVQPMD-----
Pp1s84_95V6 : ---MVSQVVKTDPHWPPLEESMGQERYA-----LNSDVVPSVPGFHNLSAQPM-----
Pp1s42_157V6 : ---MGSETPWPSVETHDNAGKLA-----LAAGIASANPAP---QMDAPP-----
Pp1s31_388V6 : ---MGGETQWPSAEAKEKNGIPP-----PAGGTASGNPTS---QMDAP-----
AT-HSFA8 : -----MVKSTD-----GGGSSSS-----
AT-HSFA4c : -----MD-----ENNGG-----
AT-HSFA5 : -----MNGALGNSSA-----SVSGGEG-----
AT-HSFC1 : -----MEDDNSNNNN-----
AT-HSFA3 : ---MSPKKDAVSKPTPIVPPVSRSDIPG-----SLYVDTDMGFSGSPMPPLD-----
AT-HSFA9 : MTAIPNVVDIESSSSSLCQETATETVTV-----RGSDDSSSKPDDVLLIKEEDDAVNLSLGF-----
AT-HSFA4a : -----MD-----ENNHGVS-----
AT1G77570.1 : -----MLKSRPNYSK-----LLICEVCEVLGM-----
AT4G18870.1 : -----MSKNEGSLTS-----

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Pp1s12_41V6 : DPSTNEIVSWGE-TNTFFVVRPPEFARDLLEKYFKH--NMFSSFVROLNTYGFRKIV--PERWEFASDF
Pp1s207_20V6 : DPATDDIVSWGE-DETFVVRPPEFARDLLENYFKH--NMFSSFVROLNTYGFRKIV--PDHWEFANKF
AT-HSFB4 : DPATDHVSWGD-DDTFFVVRPPEFARDLLENYFKH--NMFSSFVROLNTYGFRKIV--PDRWEFANEF
HSFB1 : DHSDDVSWNE-EGTFFVVKTAFAKDLLEKYFKH--NMFSSFVROLNTYGFRKIV--PDKWEFANDY
AT-HSFB2a : DSSDDVSWNE-DGSSFIVVNPTDFAKDLLEKHEKH--NMFSSFVROLNTYGFRKVV--PDRWEFSNDF
AT-HSFB3 : DPTDGVISWNE-YGTGFVVVQPAEFARDLLETLFKH--CNFSSFVROLNTYGFRKVT--TIWWEFSNEM
AT-HSFB2b : DPVYDELISWNE-DGTFIVVRPAEFARDLLEKYFKH--NMFSSFVROLNTYGFRKVV--PDRWEFSNDC
ATHSFA1d : DHNTDSIVSWSA-NNNSFIVVKPPEFARDLLEKNEKH--NMFSSFVROLNTYGFRKVD--PDRWEFANEH
HSFA1a : DPATDAIVSWSP-TNNSFIVVDPPEFSRDLEKYFKH--NMFSSFVROLNTYGFRKVD--PDRWEFANEH
ATHSFA1e : DPLTDDVSWSS-GNNSFVVVNPPEFAQELLEKYFKH--NMFSSFVROLNTYGFRKVD--PDRWEFANEH
AT-HSFA7b : DPNTDHIVSWNR-GGTSFVVVDLHFSFSTILLRYFKH--NMFSSFVROLNTYGFRKIE--AERWEFANEH
ATHSFA2 : DPATDVVSWSN-GRNSFVVVDSHKFSTILLRYFKH--NMFSSFVROLNTYGFRKID--PDRWEFANEH
AT-HSFA6b : DSRNHHVSWSK-SNNSFIVVDPQAFSVTLLEPRFFKH--NMFSSFVROLNTYGFRKVN--PDRWEFANEH
AT-HSFA6a : DSSNNIVSWSR-DNNSFIVVEPETFALICLERCFKH--NMFSSFVROLNTYGFRKID--TERWEFANEH
AT-HSFA7a : DPNTDHIVSWNR-GGTSFVVVDLHFSFSTILLRYFKH--NMFSSFVROLNTYGFRKIE--AERWEFANEH
HSFA1b : DPLTNEVSWSS-GNNSFVVVSAPEFSKVLLEKYFKH--NMFSSFVROLNTYGFRKVD--PDRWEFANEH
Pp1s249_84V6 : DPATDPVSWSA-GNNSFIVVNPPEFAQELLEKYFKH--NMFSSFVROLNTYGFRKVD--PDRWEFANEH
Pp1s84_95V6 : DPATDPVSWST-GHNSFIVVNPPEFAQELLEKYFKH--NMFSSFVROLNTYGFRKVD--PDRWEFANEH
Pp1s42_157V6 : VSATDAIVSWSE-VGNSFVVVNPPEFAQELLEKYFKH--NMFSSFVROLNTYGFRKVD--PDRWEFANEH
Pp1s31_388V6 : DPATDAIVSWSE-VGNSFVVVNPPEFAQELLEKYFKH--NMFSSFVROLNTYGFRKVD--PDRWEFANEH
AT-HSFA8 : DSTDSIIWSPSADNSFVILDTHVSVQLLEKYFKH--NMFSSFVROLNTYGFRKVD--ADRWEFANDG
AT-HSFA4c : DSSSDSVWVWSE-NNKSFIVKNEAFAFRDLEPRFFKH--NMFSSFVROLNTYGFRKVD--PEKWEFLNDD
AT-HSFA5 : DSSDQIVSWSA-NNNSFIVVNHAEFSRDLEPRFFKH--NMFSSFVROLNTYGFRKID--PERWEFLNDD
AT-HSFC1 : DPTDVLIIWGP-AHNSFIVVDPDLFQSRILEAYFKH--NMFSSFVROLNTYGFRKVD--PDRWEFANEH
AT-HSFA3 : DPTLDPVSWGL-TGASFVVVDPLEFARILLRNEKH--NMFSSFVROLNTYGFRKID--TDKWEFANEH
AT-HSFA9 : TKVTDPVVSWSP-TRKSFIINDSYESENLEKYFKH--NMFSSFVROLNTYGFRKVD--SDRWEFANEH
AT-HSFA4a : DSSDSIVSWSQ-SNKSFIVVNPPEFSRDLEPRFFKH--NMFSSFVROLNTYGFRKID--PEQWEFANDD
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AT4G18870.1 : DLSLDSIIWWSQ-SGKSFIINNPPEFYNNLLORFCFQ--RINIEFSFLFSHGFRKID--SGKWEFANDN

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Pp1s12_41V6 : LQPVSVTG---SVQQSRs-----LSP-----TSIED---QAWSSITSA
Pp1s207_20V6 : SGPFSLSC---DAPGSQS-----TSPT-----NSGED---QAWSPFPSP
AT-HSFB4 : PQQHSPFM---SHHAPP-----QIPFSGGSFFLPPPRVTTPEE---DHYWCDDSP
HSFB1 : IASTAGKC---VVVGSFS-----ESNS-----GGDD---HGSSTSSP
AT-HSFB2a : QTVVAPSS---EQRNQT-----MVSFS-----NSGEDNNNNQVMS--SP
AT-HSFB3 : -QHWSHNK---SNHQVPT-----TTMVN---QEGHQRIIGI
AT-HSFB2b : MAAAAAAA---AAVAASAVTVAA--VPVVAHIVSFS-----NSGEE---QVLISSNSP
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HSFA1a : GSSSS---NPQSQQLSQGGs-----MAALSSCVEVGKFGLEEEVERLKRDKNVL
ATHSFA1e : P-----PQQPQVQH-----SSVGACVEVGKFGLEEEVERLKRDKNVL
AT-HSFA7b : PSL-----NYSQSQPE-----AHDPGVELPQLREERHVL
ATHSFA2 : -----LQNVNQOG-----SGMSCVEVGQYGFGEVERLKRDKHVL
AT-HSFA6b : NQM-----QQQSSEQQS-----LDNFCIEVGRYGLDGENDSLRRDKQVL
AT-HSFA6a : -----QTQS-----LEGEIHELRRDMAL
AT-HSFA7a : -----FTPSSSP-----SHD---ACNELRREKQVL
HSFA1b : QN-----QQQTQVQS-----SSVGACVEVGKFGIEEEVERLKRDKNVL
Pp1s249_84V6 : -----QSVQQQQQQHQQSEQGP-----VGPCVEVGKFGLEGEIERLKRDKNVL
Pp1s84_95V6 : -----QSAQQQQQ--HQQTDQGS-----VGPCVEVGKFGLEGEIERLKRDKNVL
Pp1s42_157V6 : -----QQQQ-----GSCVEVGKLGLEGEIERLKRDKNVL
Pp1s31_388V6 : -----QQQQ-----GAYVEGKSGLEAEIERLKTDKNVL
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AT-HSFA4c : SLVNLQAO---NPLTESER-----RSMEDQIERLKNEKEGL
AT-HSFA5 : SHP-----PASSTDQER-----AVLQEQMDKLSREKAAI
AT-HSFC1 : YG-----QLEDGEIVR-----EIERLKEEQREL
AT-HSFA3 : QTCCSSTS---SQSGSPTEVG-----GETEKLKERRAL
AT-HSFA9 : CN-----KEASTTT-----ETEVELKKEEQSPM
AT-HSFA4a : SLPNLQAO---LNPLTDSER-----VRMNNQIERLTKEKEGL
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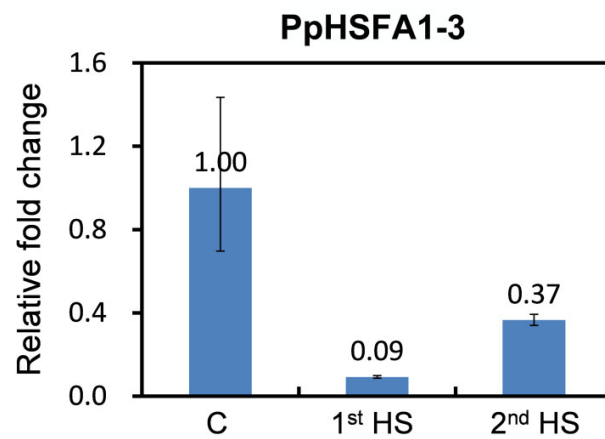
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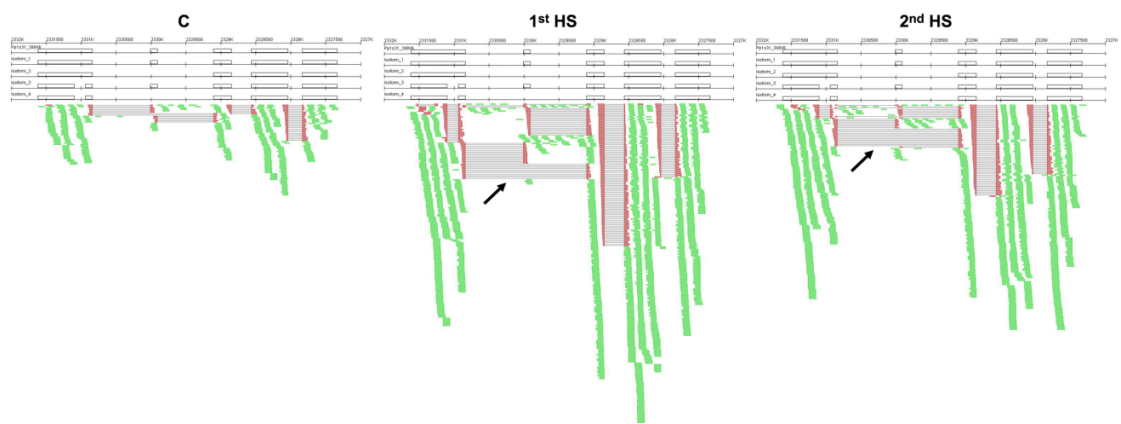
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Pp1s12_41V6 : ---TIGI SDEMEKLRKDNMLI SEI SRLRRLYEEAMAIHQHNFK : 207
Pp1s207_20V6 : ----VSDENERLRHDNRMLI LEI SRLRRLYDDLIVVILQQQ--- : 209
AT-HSFB4 : ---VTAI SEDNERLRRSNTVLM SEI AHMKKLYNDIIYFVQNH--- : 231
HSFB1 : ---VADI SGENEKLKRENNNI SSEI AAAAKQRDELVTFLTGH--- : 191
AT-HSFB2a : ---VTEI LEENEKLRSONIQ NREI TOMKSIDCNITYSLMSNY--- : 210
AT-HSFB3 : ---YTAI LDENKCLKNENELI SCEI GKTKKCKGLMELVERY--- : 217
AT-HSFB2b : CTTAPEI VEENERLRKDNERI RKEI TKLKGLYANIYTLMANF--- : 264
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HSFA1a : ---LQVI VKHLQVMEQRQQQIM SFI AKAVQNPTFLSQFIQKQT-- : 252
ATHSFA1e : ---LQNI VGKVVHMEQRQQQMM SFI AKAVQSPGFLNQFSQQSN-- : 213
AT-HSFA7b : ---VQAI EQRINGAEKKQRHMM SFI RRAVENPSSLQQIFEQK--- : 209
ATHSFA2 : ---VAAI EQRLLVTEKRQQQMM TFI AKALNNPNFVQQFAVMSKE- : 231
AT-HSFA6b : ---LTL EEEKLKKTESKQKQMM SFI ARAMQNPDFIQQIIVEQKEK- : 255
AT-HSFA6a : ---LHL EEEKLVTEVKQEMMMNFI LKKIKKPSFLQSLRKRNLQ- : 192
AT-HSFA7a : ---IKAI EQRIEGTERKQRQMM SFI ARAMQSPSFLHQLLKQ--- : 198
HSFA1b : ---LQNI VGKVVQVMEQRQQQMM SFI AKAVQSPGFLNLVQQNNN- : 219
Pp1s249_84V6 : ---LQAI GORLLT TENRQQHMM SFI AKAMQNP SFLAQLMQQSEN- : 257
Pp1s84_95V6 : ---LQAI GORLLT TENRQQHMM SFI AKAMQNP SFLAQLMQQSEN- : 255
Pp1s42_157V6 : ---LQVI TORFHVSEHRQRMT SFI TKAMQNP SFFAQFVQQQNE- : 228
Pp1s31_388V6 : ---LQNI AQRLHVSESRQQRMTI TFI AKAMANPSLFAQFVQQQNE- : 227
AT-HSFA8 : ---MLHI EDRVQGMEESSQEM SFI VMVMKNPSLLVQLLQPKKE- : 207
AT-HSFA4c : ---VTTI KDRLOHMEQHQSIVAYI SQVLGPK---GLSLNLEN- : 196
AT-HSFA5 : ---FEETI EHVDMMENRQKLLNFI ETATRNPTFVKNF GKKEQL : 207
AT-HSFC1 : ---IQRI NRRTEATEKRPEQMMAFI YKVVEDPDLPRMMLEKERT : 182
AT-HSFA3 : ---VDTI NQRKAABEQKQLL SFI AKLFQNRGFLERLKNFKGKE : 241
AT-HSFA9 : ---MVTI QEKIHGVDTEQQHMI SFI FAKLAKDQRFVERLVKRRKMK : 250
AT-HSFA4a : ---VKEI KERLOHMEKROKTMV SFI SQVLEKP---GLALNLSPC : 200
AT1G77570.1 : ----KARKAASKARKARVQVEFI FOHLQI----- : 147
AT4G18870.1 : ----SSDAI ISWSQSGKSFIIWNPOEFCKDHLRRLFNFLHIHF : 197

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Supplemental Figure S5. Multiple sequence alignment of *Arabidopsis* and *Physcomitrella* heat shock transcription factors (HSFs). DBD and HR-A/B domains of HSF proteins derived from *Arabidopsis thaliana* (AT) and *Physcomitrella patens* (Pp) were used. The genes annotated as *PpHSFA1-1* (Pp1s31_388V6), *PpHSFA1-2* (Pp1s42_157V6) and *PpHSFA1-3* (Pp1s84_95V6) in this study is indicated by arrows.



Supplemental Figure S6. Validation of heat-regulated IR on *PpHSFA1-3* transcripts. Pooled RNA from control (C), 1st HS and 2nd HS samples was analyzed in triplicate for quantitative RT-PCR (qRT-PCR). Primer sets designed for *PpHSFA1-3* IR isoforms, total transcripts of the corresponding gene, and the *PpACT2* were used. *PpACT2* was first used as an internal control for normalizing each qPCR reaction. Level of the IR isoform were further normalized for overall expression level and then compared with data from the HS control to generate the relative IR level. Annotations and gene IDs are in Supplemental Dataset S9.



Supplemental Figure S7. Mapping results for *PpHSFA1-1* from control (C), 1st HS and 2nd HS samples. Arrows indicate reads supporting exon 3 (E3) skipping.

Supplemental Table S1. Mapping statistics of RNA-sequencing

	C	1st HS	2nd HS	Total
Sequence reads	32,458,315	42,737,188	42,727,285	117,922,788
Mapped reads	22,455,889	23,692,513	21,633,044	67,781,446
Mapping percentage (%)	69.18	55.44	50.63	57.48
Intragenic reads	20,568,144	21,327,252	19,778,224	61,673,620
Exonic reads	15,606,490	16,654,203	15,144,324	47,405,017
Intronic reads	288,738	214,236	209,328	712,302
Splice junction reads	4,127,768	3,972,194	3,949,534	12,049,496
Exon-intron bridge reads	545,148	486,619	475,038	1,506,805
Intergenic reads	1,887,745	2,365,261	1,854,820	6,107,826

Supplemental Table S2. Splicing junctions (SJs) and alternative splicing events identified from BLAT and TopHat mapping tools.

	BLAT				TopHat			
	All	C	1st HS	2nd HS	All	C	1st HS	2nd HS
SJs	124,713	89,866	87,339	81,730	110,438	83,346	85,354	81,750
IR	37,559	24,711	16,941	17,645	41,007	23,786	17,857	19,265
AltDA	31,953	10,590	13,451	11,276	20,051	7,547	11,483	9,927
ES	1,347	671	730	707	1,414	704	864	857

SJs, Splicing junctions. IR, intron retention. AltD/A, alternative donor/acceptor site. ES, exon skipping.

Supplemental Table S3. Premature termination codon (PTC) prediction of IR events

	HS-IR events
IR events	1279
IR transcripts	2020
IR transcripts with retained intron in protein coding region	1556
PTC in the retained intron	1529
PTC in the downstream region	18
IR transcripts without PTC	9
Percentage of IR transcripts with PTC	98.26%

Supplemental Table S4. Primers used in this study

Gene product	Gene ID	Primer name	Sequence (5' to 3')
RT-PCR: HS marker genes			
PpHsa32	Pp1s251_75V6	Hsa32-F	CCATGGAGGAAAAGCTTATTC
		Hsa32-R	TCACGCAAAGTGGATGCTTCG
sHSP16.4	Pp1s27_332V6	sHsp-F	GCTTCTCGCGATCTTATTCGCAG
		sHsp-R	CCAGTTGTACCTTCACGTCGGCTG
PpHSP70	Pp1s115_168V6	Hsp70-2-F	CCAAGCTGTAGTGAACCTG
		Hsp70-2-R	CCTCCGCAGTCGCTGTAATA
PpUBQ	Pp1s46_189V6	PpUBQ-RT-F	CAGCGTCTGATTTTCGCTGG
		PpUBQ-RT-R	CGTCCACTATCAGTACGAAC
qRT-PCR: Intron retention			
KH domain protein	Pp1s87_162V6	87_162 Int2--qF	CCATGTCGTGAGAAAGTGCATT
		87_162 Int2--qR	ACCACCGTGCTGTTGGTTGTAG
		87_162 con--qF	ATCATCAACCTCAACATGGTGG
		87_162 con--qR	CTCCTTTTCCAATAACAAGCCC
PpPTB2	Pp1s326_1V6	Q_02-11_Pp1s326_1V6 (F)	TGACCTGAAGCGTTAGCAACCT
		Q_02-11_Pp1s326_1V6 (R)	CAAAGCCAAAGGCAGAAAACAC
		QC_02-11_Pp1s326_1V6 (F)	ATATTTGCTGCCTGACCACGTT
		QC_02-11_Pp1s326_1V6 (R)	ATGGAGCCACCGGAAGATATG
PpSRP34A	Pp1s28_193V6	28_193Int1-qF	CATGCCCCGTTAGATTTGG
		28_193Int1-qR	AAAGACGTAGGAGCAGTTCGG
		28_193V6-con-qF	TACTCGAAGCCGTTACGTAGC
		28_193V6-con-qR	AGATCGGTTACGCGGAGACTTG
PpS-ACP-DES	Pp1s354_37V6	354_37 Int3--qF	TTCCAATTTCTTGCAGGTCG
		354_37 Int3--qR	TCTTGCCGTCGACAAATCC
		354_37 con--qF	GGCCGCTTGGAAGTAGTTATG
		354_37 con--qR	TGTGTCTCCGTTGGTCGTTTA
PpRPL37aB	Pp1s76_74V6	Pp1s76_74V6-q2F	GGCAGGGTGTGTATGTGTTG
		Pp1s76_74V6-q2R	CAATTCGGCTTTCTTCGTG
		Pp1s76_74V6-qCF	TCACTATGACTAAGCGCACGAA
		Pp1s76_74V6-qCR	CCTCAAACCTGGCACCATAACG

Pp EF1B/RPS6	Pp1s359_29V6	Pp1s359_29V6-qF	TTTGTGGATGTGTAGGCCATCA
		Pp1s359_29V6-qR	CCCCTGTTGAACATCTCAACGT
		Pp1s359_29V6-qCF	ACCAAGCCAGAGAGCCAAAAG
		Pp1s359_29V6-qCR	ATCATCGTCCCGGTTTCAGTCT
PpHARS1	Pp1s38_401V6	38_401 Int11--qF	TGCTTT CACAGTGTCTTCCGT
		38_401 Int11--qR	ATCGCAAGAGATCGTCGAACAG
		38_401 con--qF	TTTATACCCGCAAAGCTGGTGA
		38_401 con--qR	TTCTCCCACCTTTGTCTCTCAA
PpATG8	Pp1s209_115V6	209_115 Int4--qF	ATATGAGGCGCAGAAACTGG
		209_115 Int4--qR	AAACCCTACAAGCGCCCAT
		209_115 con--qF	TCAAGCAAGAGCATCATCTGGA
		209_115 con--qR	TGTCGCTTTTCTCCGCCTT
PpRPS27	Pp1s63_161V6	Q_03-1_Pp1s63_161V6 (F)	GCTTATGTCCGTCTTGTGTTGC
		Q_03-1_Pp1s63_161V6 (R)	AAGCATCCTTGACACTTCACATC
		QC_03-1_Pp1s63_161V6 (F)	GAAGTGTC AAGGATGCTTCAGC
		QC_03-1_Pp1s63_161V6 (R)	AAGCACTGTGGAGCAACTTCC
PpCYP38-2	Pp1s90_50V6	90_50 Int1--qF	CGCCTCCTCGGATATGAATTC
		90_50 Int1--qR	GGTGCGCCATAGAGATTGAAGT
		90_50 con--qF	CTGGCAAAGTGCGAACTGTTC
		90_50 con--qR	TGTTGCCTTGTACTTGCCAAG
PpPPIase	Pp1s46_106V6	46_106 Int9--qF	GGAGAAGAAACAGCGCAGTGTG
		46_106 Int9--qR	GCACTCCACACTACTTCTTGCA
		46_106 con--qF	CCGTGCTTTTGGGAATGAG
		46_106 con--qR	CAAGCTCCTCTTTCTCCCAACA
PpRING/U-box	Pp1s157_62V6	62 Ubox Int1--qF	TGATGTGAAGTGTGCATTGCTG
		62 Ubox Int1--qR	GACAGAAACCGCATTCCAATTC
		62 Ubox con--qF	CAAACGCCGATTTCTGAAGAG
		62 Ubox con--qR	CATACGCCTTTTGTGAGCAGCAT
PpPsaE-2	Pp1s334_17V6	Pp1s334_17V6-qF	TTTAACGACACCCGGCAAGGTT
		Pp1s334_17V6-qR	AACGTGAGGAGGAGATGGAAC
		Pp1s334_17V6-qCF	TGTCAAGGAAGAGGCTAAGCCA
		Pp1s334_17V6-qCR	CCTCAGAACCTTAACGACGCTG
PpUROD	Pp1s114_123V6	Pp1s114_123V6-qF	GCTGCGATGGTTCTGTTGTACA
		Pp1s114_123V6-qR	GCCAAATCCACATTCTCAGAGC
		Pp1s114_123V6-qCF	TTGCGTGAGGAGGTCGGTAAT

PpTBP1	Pp1s246_34V6	Pp1s114_123V6-qCR	TGATCCGCCCTCGACTATGTAG
		TBP Int7--qF	CTGTGGACCTTGCCAAACATC
		TBP Int7--qR	CAAGAACCAACCCCAACTTC
		TBP con--qF	CGCATCATTCAAAAGCTTGG
PpRBCS2B	Pp1s251_44V6	TBP con--qR	GGAACTTCACGTCACATGATCC
		Pp1s251_44V6-qF	CTCTTACTTGCCCCATTGTCC
		Pp1s251_44V6-qR	AGAGACAGGGGACTTACCACGT
		Pp1s251_44V6-qCF	GAGTTCCACCTTGTTCTCCAGC
PpHSFA1-3	Pp1s84_95V6	Pp1s251_44V6-qCR	GAACTTCGTCATGCCAATGG
		84_95 Int2---qF	GATTACGGCAACAGCAGCAGA
		84_95 Int2---qR	GCGGAAAATTCAAGTACCTGCA
		84_95 Con---qF	AACITTTCCAGCTTCGTTCCGG
		84_95 Con---qR	CCGGATCAACCTTACGGAATC
High-resolution			
RT-PCR			
Peptidase	Pp1s47_94V6	47_94FAM---F	CATTTACGGCACACGCTCTCC
		47_94---R	CGCCAATACTGGTGAATCAGC
PpCYP38-2	Pp1s90_50V6	90_50---F	GACAACCCGAATGTGAAAGATG
		90_50 FAM---R	CTCAAGCGTGGCACCGTAG
PHD finger protein	Pp1s384_37V6	384_37FAM---F	ACCACTATGACGTGCAGAGC
		384_37---R	CTGTTGCATTCCAGCATCAC
PpHSFA1-1	Pp1s31_388V6	31_388E2--F	CATCGGCTGAAGCGAAAGAG
		31_388E2--FAM_R	CTCAATCTCCGCCTCAAGTC
