

Supplementary Material for

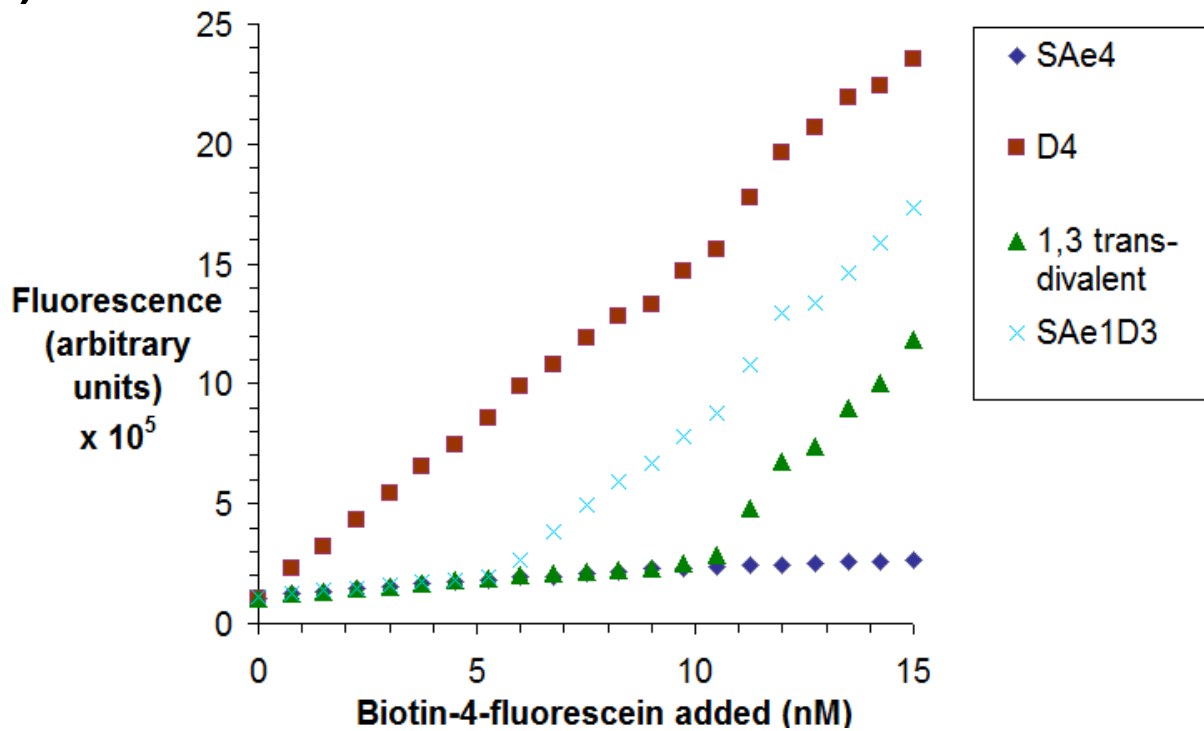
Plug-and-Play Pairing via Defined Divalent Streptavidins

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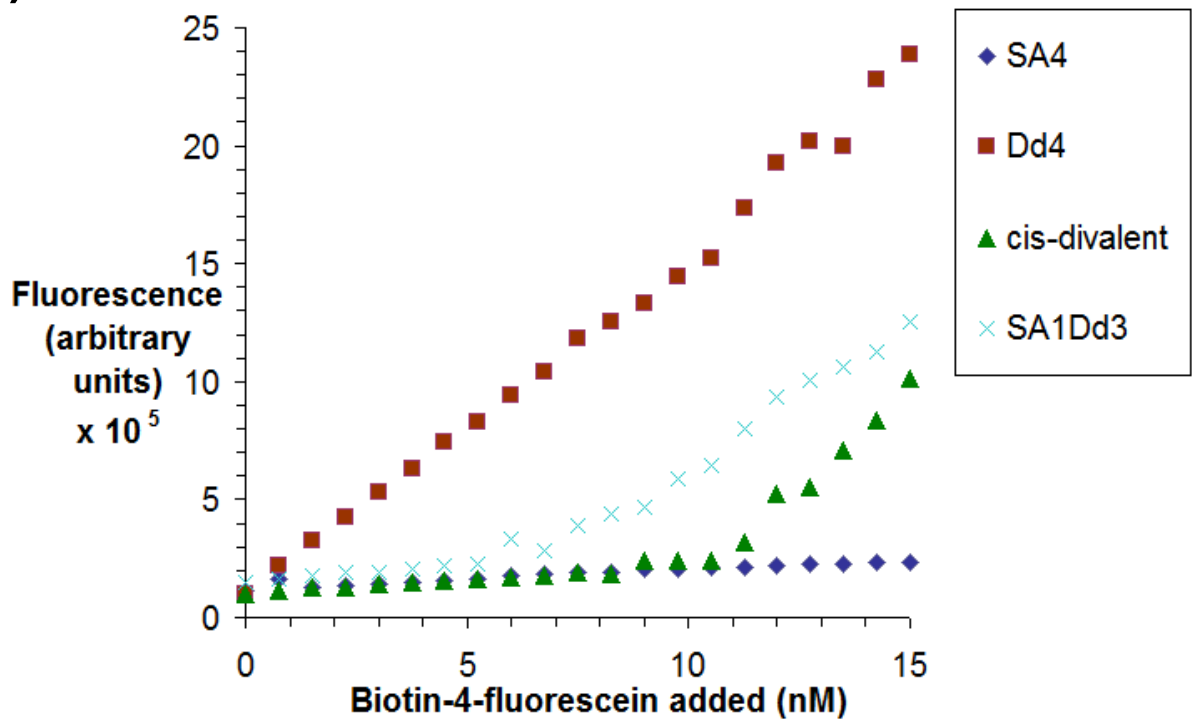
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(a)

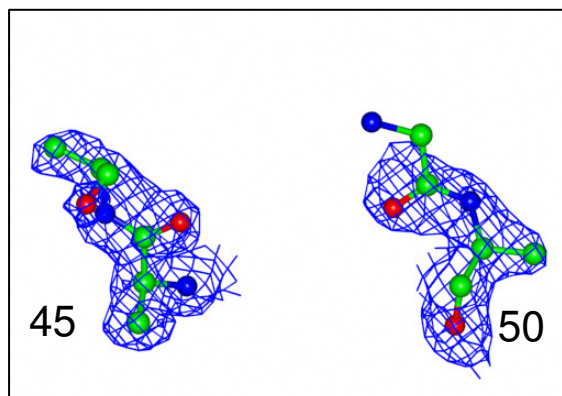


(b)

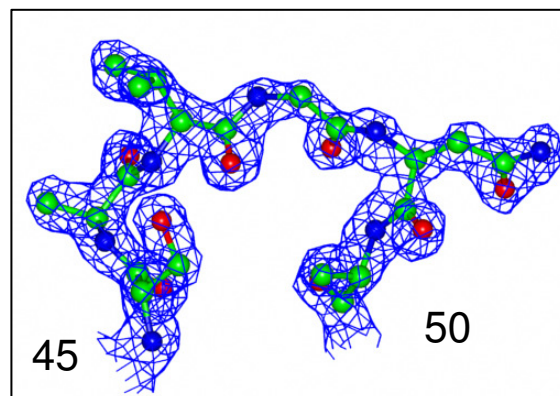


Supplementary Figure S1. Titration of biotin-binding sites. 20 nM of each streptavidin variant (monomer concentration) was incubated with the indicated concentration of biotin-4-fluorescein for 1 h and then fluorescence was measured. As [biotin-4-fluorescein] increases, fluorescence increases slowly at concentrations where all is being bound by streptavidin, but once the biotin-binding sites are saturated, fluorescence increases rapidly.

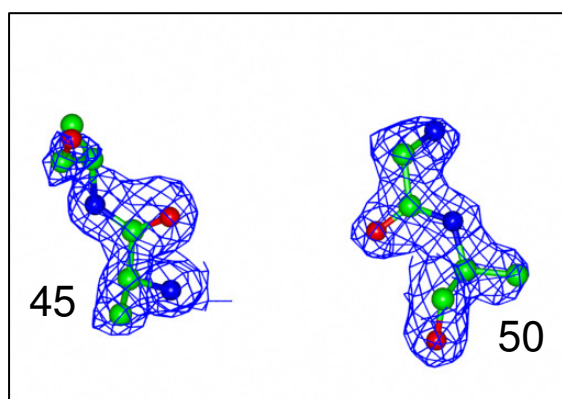
Chain a



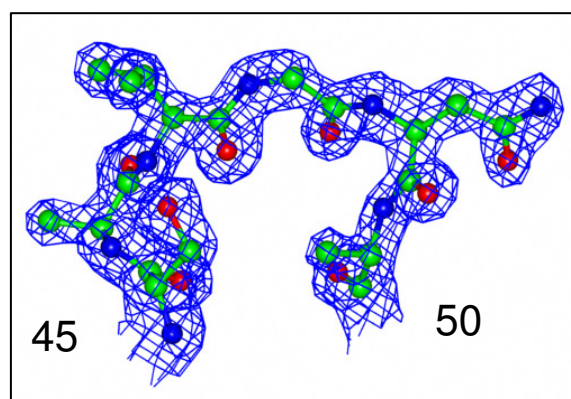
Chain b



Chain c

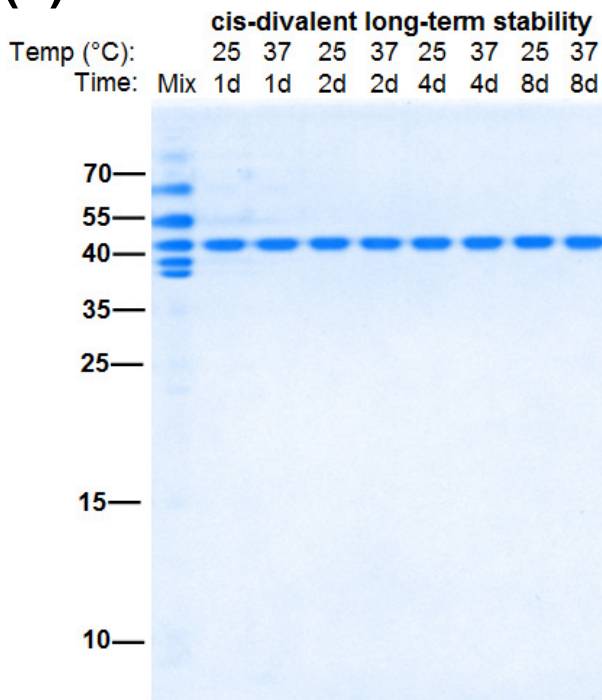


Chain d

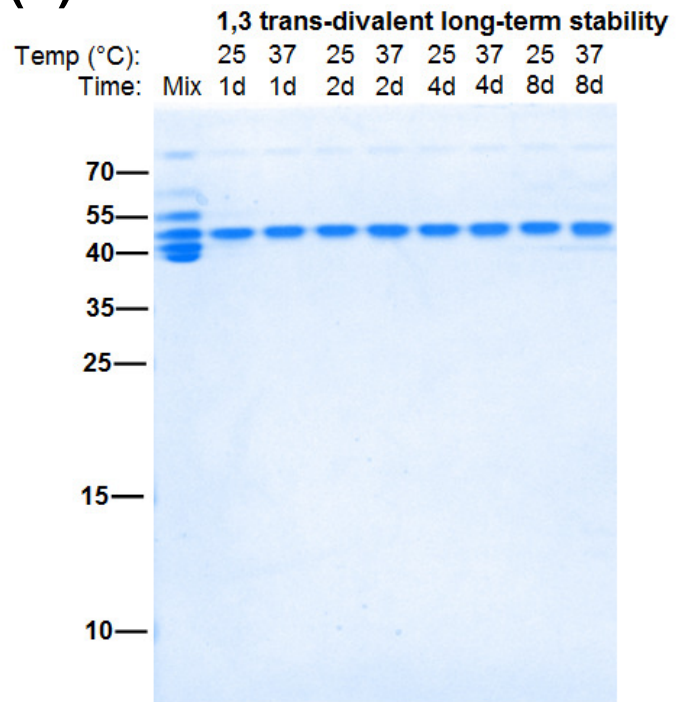


Supplementary Figure S2. Electron density around residues 45-50 in chains a-d of cis-divalent streptavidin ($2F_o - F_c$ contoured at 1 rmsd). Residues in stick format are shown surrounded by the observed electron density (blue mesh).

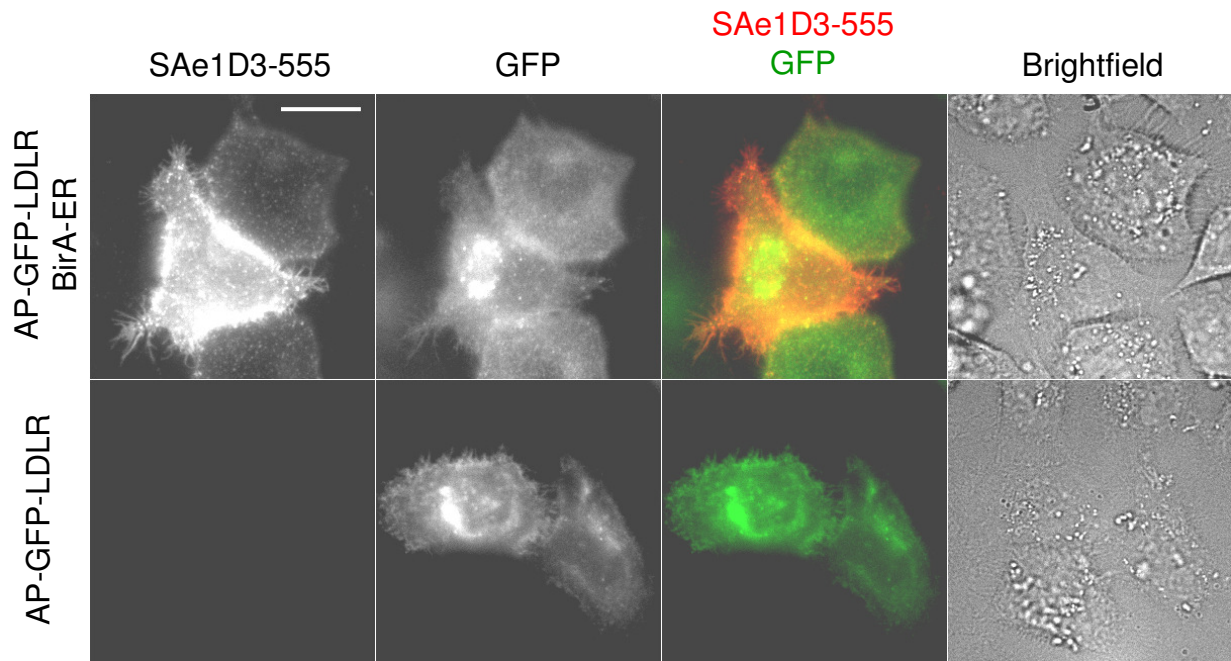
(a)



(b)



Supplementary Figure S3. Stability over time of cis- and 1,3 trans-divalent streptavidins. 10 μ M cis-divalent or 1,3 trans-divalent streptavidin was incubated in PBS at 25°C or 37°C for 1, 2, 4 or 8 days (d). The samples without boiling were then analyzed by SDS-PAGE with Coomassie staining, to check for tetramer rearrangement or dissociation to monomers. The original mixture of unpurified tetramer forms (Mix) provided a reference as to the mobility of possible rearranged products.



Supplementary Figure S4. Labeling of LDL receptor with SAe1D3. HeLa cells were either co-transfected with AP-GFP-LDLR and BirA-ER (top row) or AP-GFP-LDLR alone (bottom row) and stained with SAe1D3-AlexaFluor 555 before live cell wide-field fluorescence microscopy. Images are shown of SAe1D3-AlexaFluor 555 (left, grayscale), GFP (center left, grayscale), a merge (center right) of SAe1D3-555 (red) and GFP (green), and the brightfield image (right, grayscale). Scale-bar 20 μ m.