## **Supporting Material**

## **Micro-structures in the organ of Corti help outer hair cells form traveling waves along the cochlear coil**

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**Figure S1. Finite element model**. (A) The apex model near  $Z = 10$  mm. (B) The base model near  $Z = 2$ mm. Each radial section has 25 nodes (red dots). Three-dimensional coordinates of 8 characteristic nodes were shown next to each model (A0, AM, AX, BB, CC, DD, E0 and E1). Note that three nodes are at different plane from the others  $(Z = Z_0 - 5 \text{ µm}$  for BB, E0 and E1) due to outer hair cell tilt. The sections are 10 µm apart along the Z-axis. These sections are bound by longitudinal elements along the TM, BM, RL (through nodes CC and BB), and through Deiters cellouter hair cell junction (node DD). Additionally, the DCpp connect node DD of a section to node BB of two sections later (i.g., DD at  $Z = Z_0 + 0$  and BB at  $Z = Z_0$  - $20 \mu m$ ). The geometry gradually changes along the  $Z$ direction by interpolating the two coordinates at  $Z=10$ mm  $(A)$  and  $Z=2$  mm  $(B)$ . The shaded areas indicate the primary mass components—TM, BM, organ of Corti (OC) and fluid layer.



## **Table S1. Mechanical properties**

Dimensions in μm, areas in μm<sup>2</sup>, Young's modulus, Y<sub>M</sub>, in MPa, and stiffness in mN/m.

\* Mass thickness represents the thickest part of the basilar membrane (cf. Edge et al., 1998). The mass thickness is used to create mass matrix, while stiffness thickness used to formulate stiffness matrix.

\*\* Tolomeo *et al.*, estimated higher Young's modulus and low shear modulus (anisotropic properties). This work used isotropic elements for pillar cells. Because the pillar cells are much stiffer than any other elements in the organ of Corti, the isotropy affected the results minimally.

\*\*\* Dulon *et al.*, measured the bending stiffness of the Deiters cell phalangeal process (0.015~0.44 mN/m, N=7). We estimated Young's modulus that results in similar bending stiffness as theirs (0.003 mN/m at the apex and 0.15 mN/m at the base). †The basilar membrane is divided into two parts, the arcuate zone (BMA) beneath the tunnel of Corti and the pectinate zone

(BMP), BMA being 1/3 of the total width. The thickness is that of the fiber layer(s) running radially. ‡ A radial section of the tectorial membrane is divided into two parts, the root attached to the spiral limbus and the body

overlying the OHC stereocilia, the root being 1/3 of the total TM length and half the thickness of the body.

Symbol		Apex	Base	Description	Reference
H	$(\mu m)$	6	2	Hair bundle height	(9)
$\gamma$		0.11	0.25	Elongation of the gating spring per unit displacement of hair bundle tip	(7)
$\boldsymbol{A}$	$(ms^{-1})$	10	100	Channel activation rate constant	(15)
$k_{GS}$	(mN/m)	6	6	Gating spring stiffness	(7)
b	(nm)	0.4	0.6	Gating swing	(7)
$\boldsymbol{c}$	(nm)	0.7	0.7	Ca binding modification	(7, 16)
$k_{\rm B}$	$(ms^{-1}\mu M^{-1})$	0.4	0.4	Ca binding coefficient	(7, 16)
	$K_D^C$ (µM)	20	100	Ca dissoc. const. when a channel is closed	(7)
$K_p^O$	$(\mu M)$	1	1	Ca dissoc. const. when a channel is open	(7)
$C_0$	$(\mu M)$	1	1	$[Ca^{2+}]$ near the channel when a channel remains closed	(17, 18)
$C_1$	$(\mu M)$	20	100	$[Ca2+]$ near the channel when a channel remains open	(17, 18)

 **Table S2. Electrical and mechano-transduction properties of the outer hair cell** 

## **Supporting References**

- 1. Nam J-H, Cotton JR, & Grant JW (2005) Effect of Fluid Forcing on Vestibular Hair Bundles. *J Vestib Res* **15**(5-6):263-278.
- 2. Plassmann W, Peetz W, & Schmidt M (1987) The cochlea in gerbilline rodents. *Brain Behav Evol* **30**(1- 2):82-101.
- 3. Schweitzer L, Lutz C, Hobbs M, & Weaver SP (1996) Anatomical correlates of the passive properties underlying the developmental shift in the frequency map of the mammalian cochlea. *Hear Res* **97**(1-2):84- 94.
- 4. Edge RM*, et al.* (1998) Morphology of the unfixed cochlea. *Hear Res* **124**(1-2):1-16.
- 5. He DZ, Evans BN, & Dallos P (1994) First appearance and development of electromotility in neonatal gerbil outer hair cells. *Hear Res* **78**(1):77-90.
- 6. Iwasa KH & Adachi M (1997) Force generation in the outer hair cell of the cochlea. *Biophys J* **73**(1):546- 555.
- 7. Beurg M, Nam JH, Crawford A, & Fettiplace R (2008) The actions of calcium on hair bundle mechanics in mammalian cochlear hair cells. *Biophys J* **94**(7):2639-2653.
- 8. Roth B & Bruns V (1992) Postnatal development of the rat organ of Corti. II. Hair cell receptors and their supporting elements. *Anat Embryol (Berl)* **185**(6):571-581.
- 9. Lim DJ (1986) Functional structure of the organ of Corti: a review. *Hear Res* **22**:117-146.
- 10. Strelioff D & Flock A (1984) Stiffness of sensory-cell hair bundles in the isolated guinea pig cochlea. *Hear Res* **15**(1):19-28.
- 11. Tolomeo JA, Steele CR, & Holley MC (1996) Mechanical properties of the lateral cortex of mammalian auditory outer hair cells. *Biophys J* **71**(1):421-429.
- 12. Dulon D, Blanchet C, & Laffon E (1994) Photo-released intracellular Ca2+ evokes reversible mechanical responses in supporting cells of the guinea-pig organ of Corti. *Biochem Biophys Res Commun* **201**(3):1263- 1269.
- 13. Gueta R, Barlam D, Shneck RZ, & Rousso I (2006) Measurement of the mechanical properties of isolated tectorial membrane using atomic force microscopy. *Proc Natl Acad Sci U S A* **103**(40):14790-14795.
- 14. Gu JW, Hemmert W, Freeman DM, & Aranyosi AJ (2008) Frequency-dependent shear impedance of the tectorial membrane. *Biophys J* **95**(5):2529-2538.
- 15. Ricci AJ, Kennedy HJ, Crawford AC, & Fettiplace R (2005) The transduction channel filter in auditory hair cells. *J Neurosci* **25**(34):7831-7839.
- 16. Nam JH & Fettiplace R (2008) Theoretical conditions for high-frequency hair bundle oscillations in auditory hair cells. *Biophys J* **95**(10):4948-4962.
- 17. Beurg M, Nam JH, Chen Q, & Fettiplace R (2010) Calcium balance and mechanotransduction in rat cochlear hair cells. *J Neurophysiol* **104**(1):18-34.
- 18. Beurg M, Fettiplace R, Nam JH, & Ricci AJ (2009) Localization of inner hair cell mechanotransducer channels using high-speed calcium imaging. *Nat Neurosci* **12**(5):553-558.
- 19. Newmark (1959) A method of computation for structural dynamics, *ASCE J. Eng. Mech. Div.* **85**:67-94