

# Supplementary Information

**A mesophilic autotrophic ammonia-oxidizing archaeon of the thaumarchaeal group I.1a cultivated from a deep oligotrophic soil horizon**

Man-Young Jung<sup>a</sup>, Soo-Je Park<sup>b</sup>, So-Jeong Kim<sup>a</sup>, Jeong-Geol Kim<sup>a</sup>, Jaap S. Sinninghe Damsté<sup>c</sup>, Che Ok Jeon<sup>d\*</sup>, and Sung-Keun Rhee<sup>a\*</sup>

\*Addressed correspondence to Sung-Keun Rhee, [rhees@chungbuk.ac.kr](mailto:rhees@chungbuk.ac.kr); Che Ok Jeon, [cojeon@cau.ac.kr](mailto:cojeon@cau.ac.kr).

This file includes:

Supplementary Table S1 to S3

Supplementary reference

Supplementary Figure Legends

Supplementary Figures S1 to S8

**Table S1.** Primers used for PCR amplification during library construction and real-time quantification.

<i>Target gene</i>	<i>Application</i>	<i>Sequence (5' to 3')</i>	<i>Position</i>	<i>Reference</i>
<i>16S rRNA</i>				
519F 727R	Archaeal quantification	CAGCMGCCGCGGTAA GCTTTCRTCCCTCACCGT	519–533 <sup>a</sup> 712–727 <sup>a</sup>	(1, 2)
Bac518F Bac786R	Bacterial quantification	CCAGCAGCCGCGGTAAT CTACCAGGGTATCTAATC	518–534 <sup>a</sup> 786–803 <sup>a</sup>	(3, 4)
27F	Clone library of bacteria (with1492R)	AGAGTTTGATCMTGGCTCAG	8–27 <sup>a</sup>	(2)
20F 1492R	Phylogenetic analysis and clone library of AOA	TTCCGGTTGATCCYGCCRG TACGGYTACCTTGTTACGACTT	2–20 <sup>a</sup> 1510–1492 <sup>a</sup>	(5) (2)
βAMOF βAMOR	First PCR (β-AOB)	TGGGGRATAACGCAYCGAAAG AGACTCCGATCCGGACTACG	143–163 <sup>a</sup> 1296–1315 <sup>a</sup>	(6)
CTO189F CTO654R	Second PCR (β-AOB)	GGAGRAAAGYAGGGGATCG CTAGCYTTGTAGTTTCAAACGC	189–207 <sup>a</sup> 633–654 <sup>a</sup>	(7)
<i>amoA</i>				
AamoAF AamoAR	Phylogenetic analysis and clone library of AOA	STAATGGTCTGGCTTAGACG ACATACAGATGGATGGCCGC	19–38 <sup>b</sup> 582–601 <sup>b</sup>	(8)
amoA1F amoA2R	PCR (β-AOB)	GGGGTTTCTACTGGTGGT CCCCTCKGSAAAGCCTTCTTC	332–349 <sup>c</sup> 820–822 <sup>c</sup>	(9)
A189F A682R	PCR (γ-AOB)	GGNGACTGGGACTTCTGG GAASGCNGAGAAGAASGC	172–189 <sup>d</sup> 665–682 <sup>d</sup>	(10)

<sup>a</sup> Numbering is based on the 16S rRNA gene of *Escherichia coli*.

<sup>b, c, d</sup> Numbering is based on the *amoA* gene of a metagenomic clone from the Sargasso Sea (AACY01435967), *N. europaea* (L08050), and *Nitrosococcus oceani* (AF047705), respectively.

**Table S2.** Comparison of 16S rRNA and *amoA* gene of strain MY2 with those of other AOA strains and fosmid clones.

Species or clone	% identity <sup>a</sup>	
	Strain MY2	
	16S rRNA	<i>amoA</i>
<i>Nitrosopumilus maritimus</i> (11)	92.9	79.5
“ <i>Candidatus Nitrosoarchaeum koreensis</i> ” (12)	92.9	80.5
“ <i>Candidatus Nitrosoarchaeum limnia</i> ” (13)	93.3	81.7
<i>Cenarchaeum symbiosum</i> (14)	93.2	76.1
“ <i>Candidatus Nitrosotalea devanaterre</i> ” (15)	88.9	75.1
“ <i>Candidatus Nitrososphaera gargensis</i> ” (16)	84.3	71.3
<i>Nitrososphaera viennensis</i> (17)	85.4	70.9
“ <i>Candidatus Nitrosocaldus yellowstonii</i> ” (18)	82.8	70.1
54d9 (19)	85.1	71.8

<sup>a</sup> Comparisons are based on 16S rRNA genes (Ca. 1300 bp) and *amoA* genes (Ca. 600 bp).

**Table S3.** Clusters of orthologous groups (COG) automated classification.

Functional category	MY2		<i>N. maritimus</i>		<i>N. koreensis</i>		<i>N. gargensis</i>		
	ORF (unique ORF)	% of ORF	ORF	% of ORF	ORF	% of ORF	ORF	% of ORF	
<b>CELLULAR PROCESSES AND SIGNALING</b>									
D	Cell cycle control and mitosis	14 (11)	0.66	11	0.55	5	0.26	9	0.25
M	Cell wall/membrane/envelop biogenesis	52 (34)	2.44	49	2.45	46	2.37	50	1.40
N	Cell motility	32 (26)	1.50	6	0.30	9	0.46	23	0.64
O	Post-translational modification, protein turnover, chaperone functions	77 (36)	3.61	74	3.71	60	3.08	90	2.52
T	Signal Transduction	76 (70)	3.56	47	2.35	42	2.16	65	1.82
U	Intracellular trafficking and secretion	17 (8)	0.80	11	0.55	8	0.41	10	0.28
V	Defense mechanisms	4 (1)	0.19	13	0.65	14	0.72	15	0.42
Z	Cytoskeleton	1 (0)	0.05	0	0.00	1	0.05	0	0.00
<b>INFORMATION STORAGE AND PROCESSING</b>									
A	RNA processing and modification	2 (2)	0.09	0	0.00	1	0.05	1	0.03
B	Chromatin Structure and dynamics	1 (0)	0.05	0	0.00	1	0.05	2	0.06
J	Tranlsation	134 (29)	6.28	136	6.81	129	6.63	145	4.07
K	Transcription	98 (37)	4.59	76	3.81	89	4.58	118	3.31
L	Replication and repair	69 (35)	3.23	56	2.80	54	2.78	163	4.57
<b>METABOLISM</b>									
C	Energy production and conversion	92 (29)	4.31	100	5.01	86	4.42	118	3.31
E	Amino Acid metabolis and transport	105 (32)	4.92	109	5.46	98	5.04	125	3.51
F	Nucleotide metabolism and transport	45 (10)	2.11	45	2.25	42	2.16	46	1.29
G	Carbohydrate metabolism and transport	44 (16)	2.06	46	2.30	41	2.11	58	1.63
H	Coenzyme metabolis	87 (33)	4.08	95	4.76	75	3.86	82	2.30
I	Lipid metabolism	30 (6)	1.41	30	1.50	27	1.39	37	1.04

P	Inorganic ion transport and metabolism	58 (24)	2.72	63	3.15	52	2.67	86	2.41
Q	Secondary Structure	12 (7)	0.56	15	0.75	17	0.87	29	0.81
<b>POORLY CHARACTERIZED</b>									
R	General Functional Prediction only	156 (86)	7.31	152	7.61	116	5.96	176	4.94
S	Function Unknown	112 (60)	5.25	86	4.31	72	3.70	121	3.39
<b>COG gene</b>		<b>1318 (592)</b>	<b>62.0</b>	<b>1220</b>	<b>61.1</b>	<b>1085</b>	<b>55.8</b>	<b>1569</b>	<b>44.0</b>
<b>Total ORFs</b>		<b>2126</b>	<b>100</b>	<b>1997</b>	<b>100</b>	<b>1945</b>	<b>100</b>	<b>3566</b>	<b>100</b>

## Supplementary reference

1. **Park BJ, Park SJ, Yoon DN, Schouten S, Sinninghe Damsté JS, Rhee SK.** 2010. Cultivation of autotrophic ammonia-oxidizing archaea from marine sediments in co-culture with sulfur-oxidizing bacteria. *Appl Environ Microbiol* **76**:7575-7587.
2. **Weisburg WG, Barns SM, Pelletier DA, Lane DJ.** 1991. 16S ribosomal DNA amplification for phylogenetic study. *J Bacteriol* **173**:697-703.
3. **Baker GC, Smith JJ, Cowan DA.** 2003. Review and re-analysis of domain-specific 16S primers. *J Microbiol Methods* **55**:541-555.
4. **Muyzer G, de Waal EC, Uitterlinden AG.** 1993. Profiling of complex microbial populations by denaturing gradient gel electrophoresis analysis of polymerase chain reaction-amplified genes coding for 16S rRNA. *Appl Environ Microbiol* **59**:695-700.
5. **DeLong EF.** 1992. Archaea in coastal marine environments. *Proc Natl Acad Sci U S A* **89**:5685-5689.
6. **McCaig AE, Embley TM, Prosser JI.** 1994. Molecular analysis of enrichment cultures of marine ammonia oxidisers. *FEMS Microbiol Lett* **120**:363-367.
7. **Kowalchuk GA, Stephen JR, De Boer W, Prosser JI, Embley TM, Woldendorp JW.** 1997. Analysis of ammonia-oxidizing bacteria of the beta subdivision of the class Proteobacteria in coastal sand dunes by denaturing gradient gel electrophoresis and sequencing of PCR-amplified 16S ribosomal DNA fragments. *Appl Environ Microbiol* **63**:1489-1497.
8. **Francis CA, Roberts KJ, Beman JM, Santoro AE, Oakley BB.** 2005. Ubiquity and diversity of ammonia-oxidizing archaea in water columns and sediments of the ocean. *Proc Natl Acad Sci U S A* **102**:14683-14688.
9. **Rotthauwe JH, Witzel KP, Liesack W.** 1997. The ammonia monooxygenase

- structural gene *amoA* as a functional marker: molecular fine-scale analysis of natural ammonia-oxidizing populations. *Appl Environ Microbiol* **63**:4704-4712.
10. **Nold SC, Zhou J, Devol AH, Tiedje JM.** 2000. Pacific Northwest marine sediments contain ammonia-oxidizing bacteria in the beta subdivision of the *Proteobacteria*. *Appl Environ Microbiol* **66**:4532-4535.
  11. **Könneke M, Bernhard AE, de la Torre JR, Walker CB, Waterbury JB, Stahl DA.** 2005. Isolation of an autotrophic ammonia-oxidizing marine archaeon. *Nature* **437**:543-546.
  12. **Jung MY, Park SJ, Min D, Kim JS, Rijpstra WI, Sinninghe Damste JS, Kim GJ, Madsen EL, Rhee SK.** 2011. Enrichment and characterization of an autotrophic ammonia-oxidizing archaeon of mesophilic crenarchaeal group I.1a from an agricultural soil. *Appl Environ Microbiol* **77**:8635-8647.
  13. **Mosier AC, Allen EE, Kim M, Ferriera S, Francis CA.** 2012. Genome sequence of "Candidatus Nitrosopumilus salaria" BD31, an ammonia-oxidizing archaeon from the San Francisco Bay estuary. *J Bacteriol* **194**:2121-2122.
  14. **Hallam SJ, Konstantinidis KT, Putnam N, Schleper C, Watanabe Y-i, Sugahara J, Preston C, de la Torre J, Richardson PM, DeLong EF.** 2006. Genomic analysis of the uncultivated marine crenarchaeote *Cenarchaeum symbiosum*. *Proc Natl Acad Sci U S A* **103**:18296-18301.
  15. **Lehtovirta-Morley LE, Stoecker K, Vilcinskas A, Prosser JI, Nicol GW.** 2011. Cultivation of an obligate acidophilic ammonia oxidizer from a nitrifying acid soil. *Proc Natl Acad Sci U S A* **108**:15892-15897.
  16. **Hatzenpichler R, Lebedeva EV, Spieck E, Stoecker K, Richter A, Daims H, Wagner M.** 2008. A moderately thermophilic ammonia-oxidizing crenarchaeote from a hot spring. *Proc Natl Acad Sci U S A* **105**:2134-2139.



17. **Tourna M, Stieglmeier M, Spang A, Konneke M, Schintlmeister A, Urich T, Engel M, Schloter M, Wagner M, Richter A, Schleper C.** 2011. *Nitrososphaera viennensis*, an ammonia oxidizing archaeon from soil. Proc Natl Acad Sci U S A **108**:8420-8425.
18. **de la Torre JR, Walker CB, Ingalls AE, Konneke M, Stahl DA.** 2008. Cultivation of a thermophilic ammonia oxidizing archaeon synthesizing crenarchaeol. Environ Microbiol **10**:810-818.
19. **Treusch AH, Leininger S, Kletzin A, Schuster SC, Klenk HP, Schleper C.** 2005. Novel genes for nitrite reductase and Amo-related proteins indicate a role of uncultivated mesophilic crenarchaeota in nitrogen cycling. Environ Microbiol **7**:1985-1995.

## Supplementary Figure Legends

**Fig. S1.** Effect of various antibiotics on the ammonia oxidation activity of strain MY2. Activity of ammonia oxidation was determined by nitrite accumulation. Initial inoculum cell density was ca.  $2.5 \times 10^6$  cells ml<sup>-1</sup>. Concentration of streptomycin (100 µg ml<sup>-1</sup>), kanamycin (50 µg ml<sup>-1</sup>), ampicillin (50 µg ml<sup>-1</sup>), penicillin-G (50 µg ml<sup>-1</sup>), gentamycin (20 µg ml<sup>-1</sup>), mitomycin-C (20 µg ml<sup>-1</sup>), tetracycline (15 µg ml<sup>-1</sup>). Control culture was set without antibiotics. The error bars represent the standard deviations from triplicate experiments.

**Fig. S2.** Phylogenetic analysis of the archaeal 16S rRNA gene sequence (ca. 1.3 kbp) obtained from strain MY2. The archaeal 16S rRNA gene was amplified using primers 20F and 1492R. Branching patterns supported by > 50% bootstrap values (1,000 iterations) according to the neighbor-joining method are denoted by their respective bootstrap values. The cluster groups are shown at the right of the figure based on the origins of the reference sequences. “ThAOA” indicates a thermophilic AOA lineage. The scale bar represents 2% estimated sequence divergence. Enriched or isolated AOAs among the reference sequences are indicated in boldface.

**Fig. S3.** Effect of different concentrations of allylthiourea (ATU) on the ammonia oxidation by strain MY2. Strain MY2 was grown in AFM with 1 mM ammonia. Ammonia oxidation activity was indicated by nitrite accumulated. The error bars represent the standard deviations from three replicates.

**Fig. S4.** Circular representation of the genome of strain MY2. The outermost circle shows RNA (circle 1), and the gene content is predicted on the reverse and forward strands (circles 2 and 3). The colors of predicted ORFs are based on COG functional categories (see the key for the color designations). Circle 4 shows the GC skew and circle 5 shows the G + C ratio

(values greater or smaller than the average percentage in the overall chromosome are shown in red and green, respectively)

**Fig. S5.** Dot plot representation of the pairwise alignments of the strain MY2, “*Ca. N. gargensis*”, “*Ca. N. koreensis*” and *N. maritimus* genomes. Alignments were performed on the six-frame amino acid translation of the genome sequences using the program in the MUMmer 3.23 package. In all plots, a dot indicates a gene compared, with forward or reverse matches shown in red and blue, respectively.

**Fig. S6.** Venn diagrams showing the numbers of ORFs shared between the predicted proteins of strain MY2, *N. koreensis* and *N. maritimus*. Shared ORFs from the query genome were considered to be conserved if they had a BLAST match of  $\geq 60\%$  of the overall sequence identity and 70% of the length of the query ORF. Numbers in brackets below species names indicate total number of ORFs. The bar graph presents the percentage of COGs of strain MY2.

**Fig. S7.** Ammonia monooxygenase (*amo*) gene order of strain MY2 compared with ammonia oxidizing bacteria (AOB), close relatives of thaumarchaeal group I.1a (name in red) and I.1b (name in blue).

**Fig. S8.** Organization of flagella and chemotaxis genes in ammonia oxidizing archaea. Flagellum-associated genes (*flaB*, *flaG*, *flaF*, *flaH*, *flaI*, *flaJ*) are shown in light blue. Chemotaxis genes (MCP, methyl-accepting chemotaxis protein; *cheA*, histidine kinase; *cheY*, response regulator; *cheW*, adapter protein; *cheC*, *cheD*, *cheR*, *cheB*, adaptation proteins) are shown in light green. The gene for the transcriptional regulator *TrmB* is shown in white and unknown proteins are shown in grey. Color bars indicate the percentage of identity with MY2.



Fig. S2

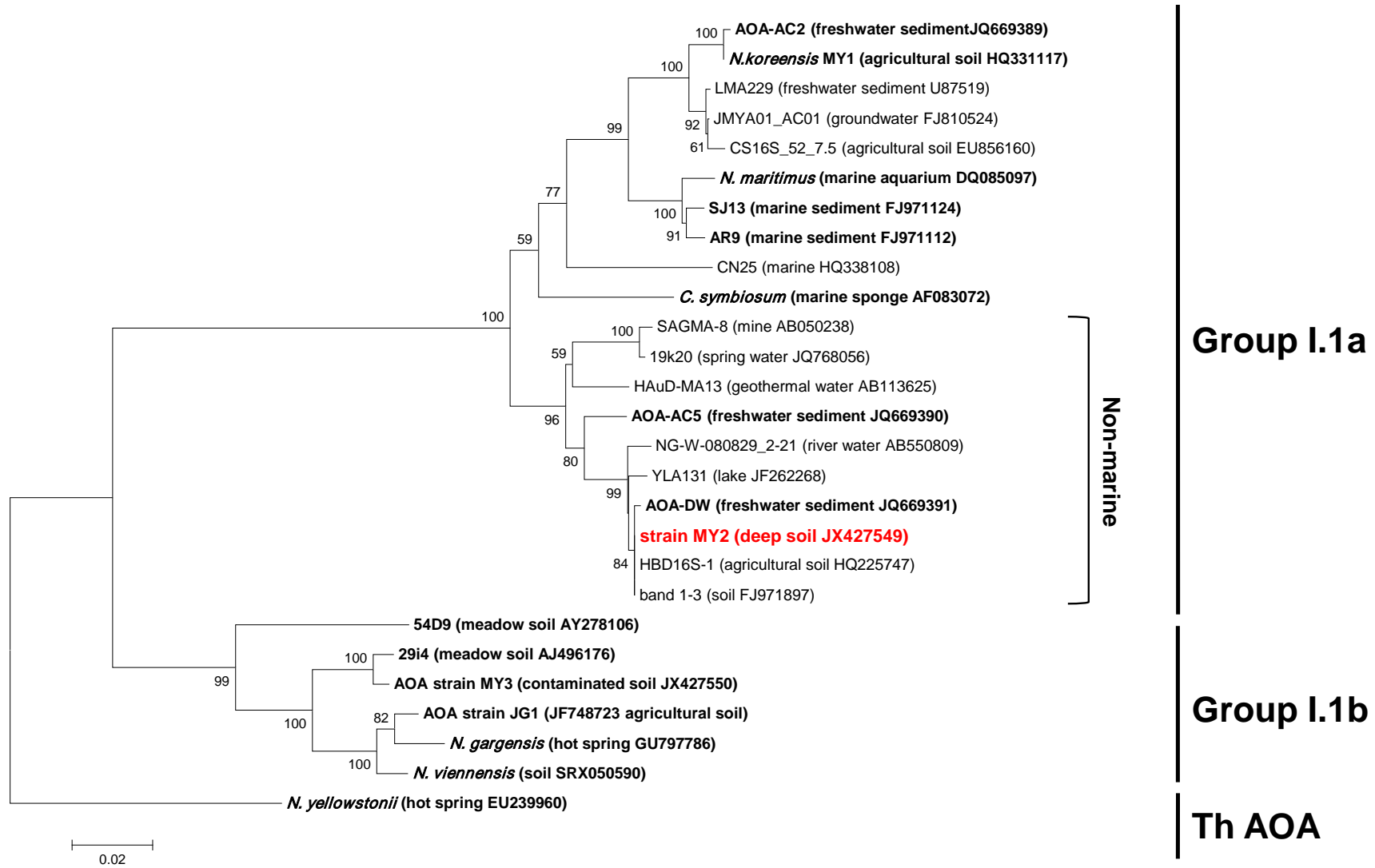


Fig. S3

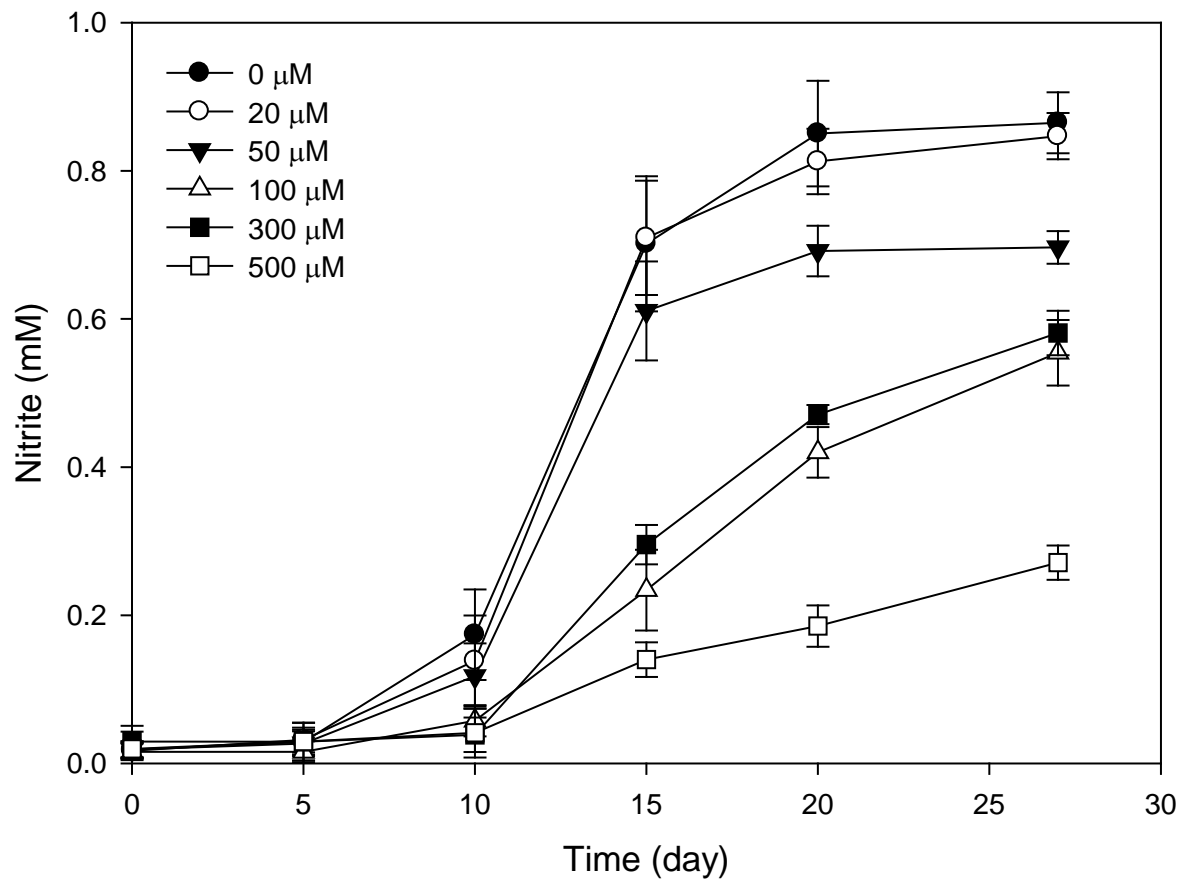
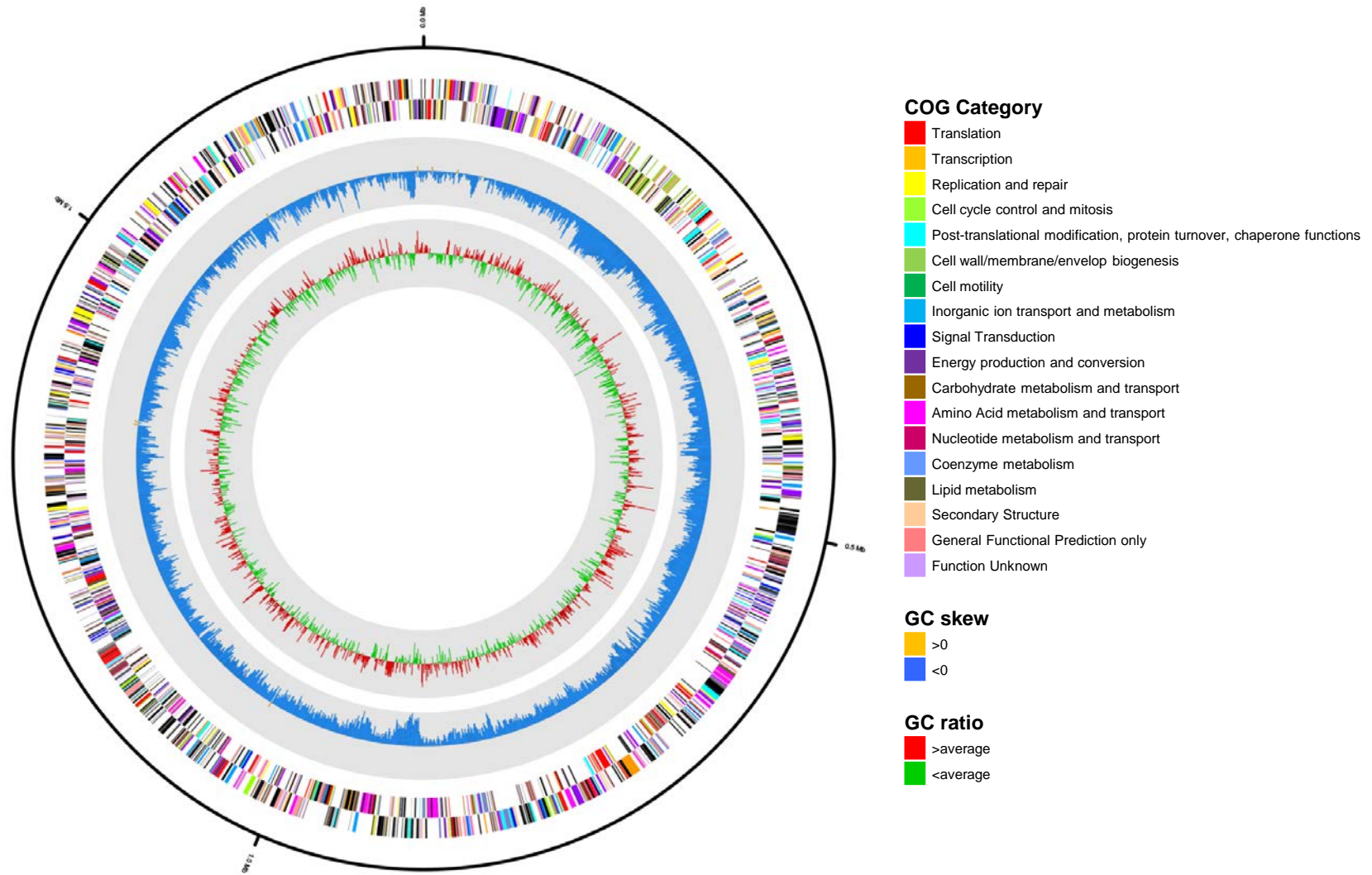
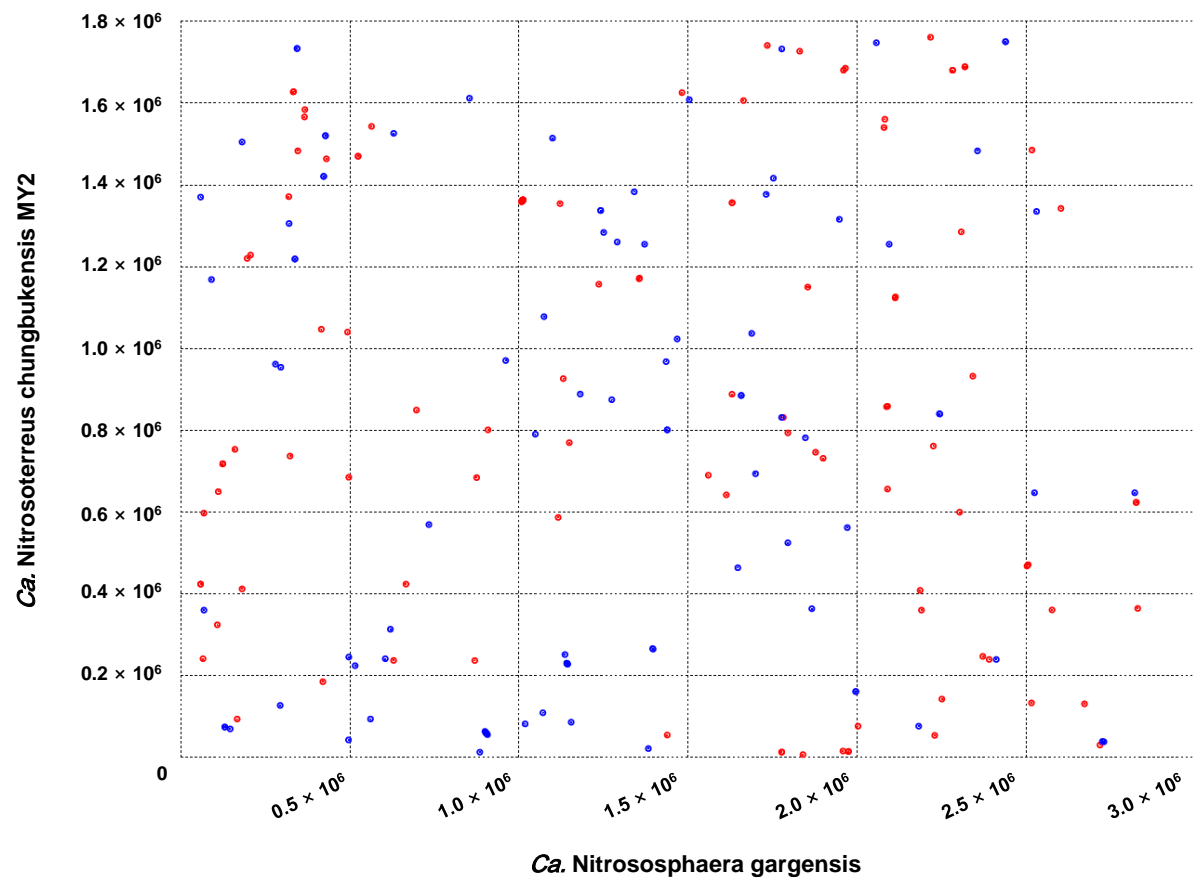


Fig. S4



1 **Fig. S5 (A)**

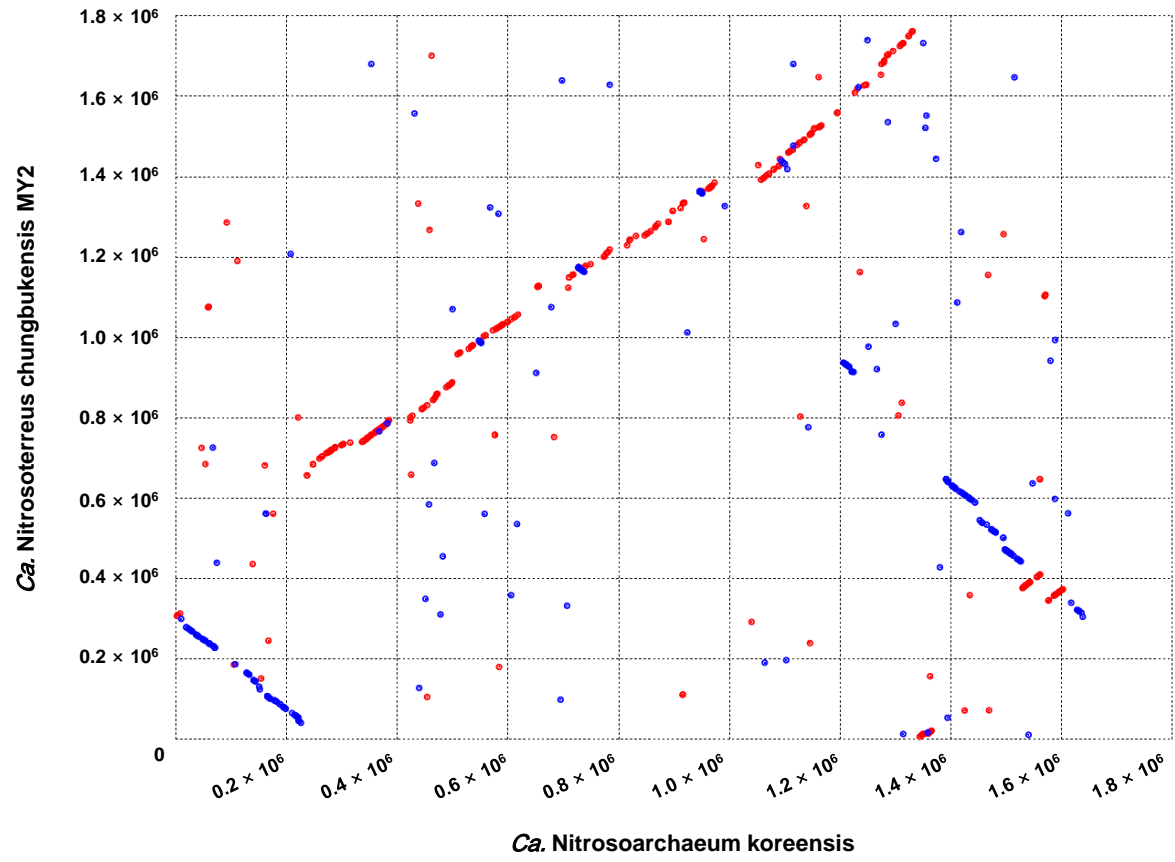


2

3



4 Fig. S5 (B)



5

Fig. S5 (C)

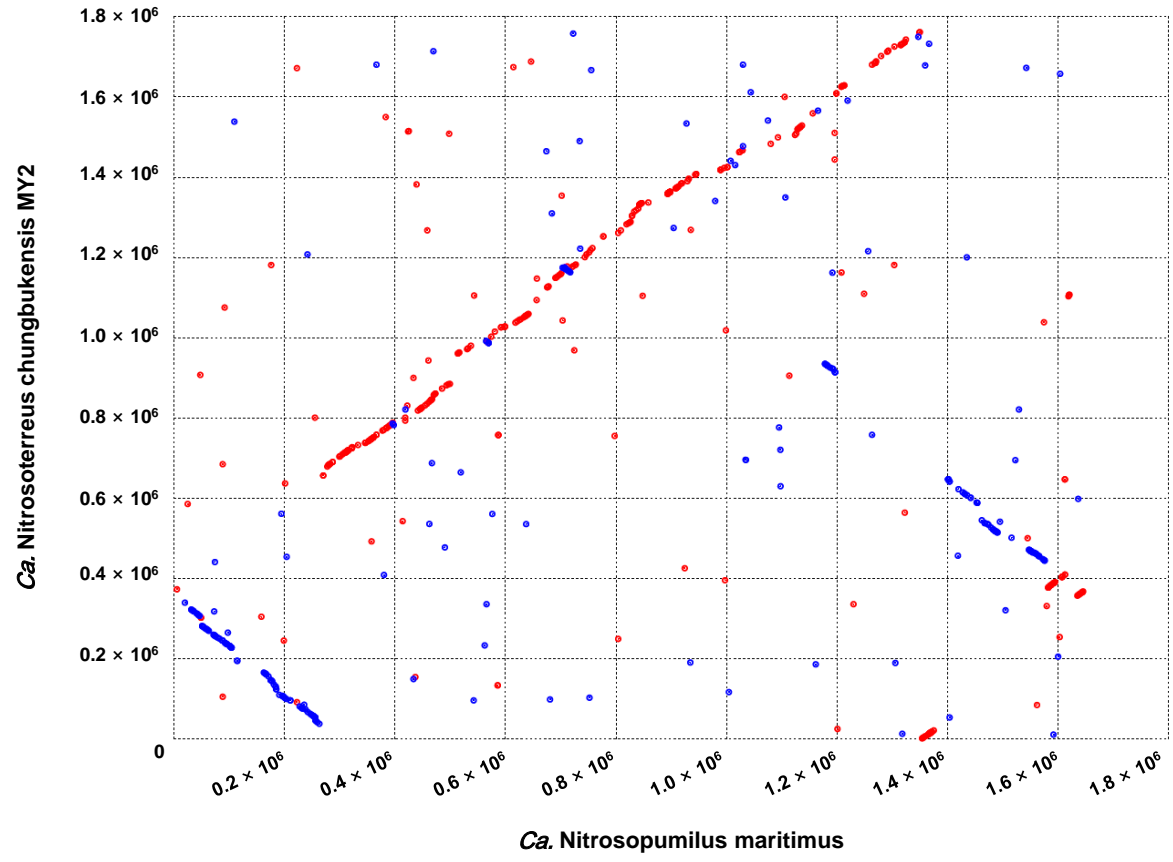


Fig. S6

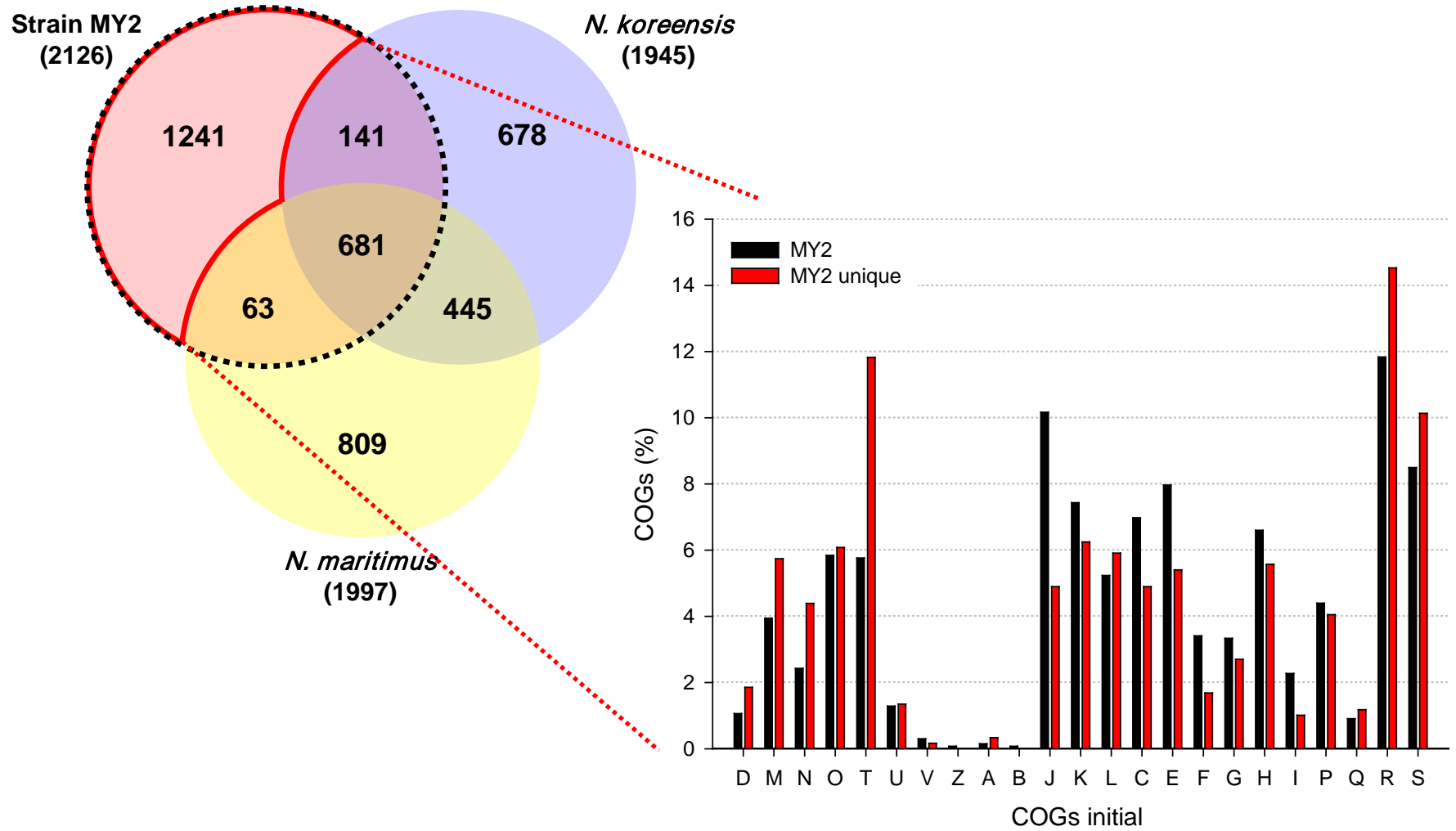




Fig. S8

