SUPPORTING INFORMATION

Synthetic and Immunological Studies of sTn Derivatives Carrying Substituted Phenylacetylsialic Acids as Cancer Vaccine Candidates

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Synthetic Procedures

General Methods. NMR spectra were recorded on a 400 or 500 MHz instrument with chemical shifts reported in ppm (δ) in reference to Me₄Si if not specified otherwise. Coupling constants (*J*) are reported in hertz (Hz). High resolution electron spray ionization mass spectra (HR ESI MS) were obtained with a Waters Micromass-LCTPremier-XE instrument, and matrix-assisted laser desorption ionization-time of flight (MALDI-TOF) MS were recorded with a Bruker Ultraflex mass spectrometer. Thin layer chromatography (TLC) was performed on silica gel GF254 plates with detection by phosphomolybdic acid in EtOH or by 1% H₂SO₄ in EtOH. Molecular sieves were dried under high vacuum at 170-180 °C for 6-10 h immediately before use. Commercial anhydrous solvents and other reagents were used without further purification. The sTn-HSA, sTnNPhAc-KLH, and sTnNPhAc-HSA conjugates were previously prepared.¹

2-Azidoethyl O-[methyl 4,7,8,9-tetra-O-acetyl-3,5-dideoxy-5-trifluoroacetamido-D-glycero- α -D-galacto-2-nonulopyranosylonate]-(2 \rightarrow 6)-2-acetamido-2-deoxy- α -D-galactopyranoside (5). A mixture of 3 (1.4 g, 2.37 mmol), 4 (0.71 g, 2.15 mmol), and activated molecular sieves (3 Å, 3.0 g) in anhydrous acetonitrile (10.0 mL) was stirred at rt for 2 h under an atmosphere of argon. After the mixture was cooled to -35 °C, N-iodosuccinamide (NIS) (1.9 g, 8.60 mmol) and triflic acid (TfOH) (76 µL, 0.86 mmol) were added with stirring and the mixture was kept at -35 °C for 0.5 h. The solid material was filtered off and then washed with DCM. The filtrate and washings were combined and extracted with aqueous Na₂S₂O₃ and water, dried over anhydrous Na₂SO₄ and concentrated under vacuum. The residue was dissolved in 65% HOAc/H₂O (v/v, 20 mL) and heated at 65 °C for 1 h while stirring. The mixture was concentrated and co-evaporated with toluene. The residue was purified by flash column chromatography (CH₂Cl₂/MeOH 25:1) to afford the desired product 5 as a white foamy solid (0.91 g, 52%), R_f 0.12 (CH₂Cl₂/MeOH 16:1) and its β -anomer (0.25 g, 14%), R_f 0.15 (CH₂Cl₂/MeOH 16:1). ¹H NMR (CDCl₃, 400 MHz): δ 7.43 (d, J 9.6 Hz, 1H, NHCOCF₃), 6.06 (d, J 8.8 Hz, 1H, NHCOCH₃), 5.34 (td, J 6.4, 2.4 Hz, 1H, H-8'), 5.30 (dd, J 7.6, 2.4 Hz, 1H, H-7'), 4.98 (td, J 11.2, 4.8 Hz, 1H, H-4'), 4.86 (d, J 4.0 Hz, 1H, H-1), 4.40-4.30 (m, 3H, H-2, H-6' and H-9'a), 4.12-3.99 (m, 2H, H-9'b and H-5'), 3.98-3.92 (m, 2H, 3-OH and OCH₂CH₂N₃), 3.89-3.83 (m, 3H, H-4 and H-6), 3.80 (s, 3H, COOCH₃), 3.76-3.69 (m, 1H, H-3), 3.67-3.59 (m, 2H, 4-OH and OCH₂CH₂N₃), 3.53-3.46 (m, 1H, OCH₂CH₂N₃), 3.37-3.30 (m, 2H, H-5 and OCH₂CH₂N₃), 2.61 (dd, J 13.2, 4.8 Hz, 1H, H-3'e), 2.11 (s, 2 x 3H, Ac), 2.01, 1.99 (s, 3 x 3H, Ac), 1.93 (t, J 12.8 Hz, 1H, H-3'a). ¹³C NMR (CDCl₃, 100 MHz): δ 172.7, 171.2, 171.1, 170.8, 170.1, 168.2, 158.1, 157.7, 117.1, 114.6, 99.0, 98.0, 72.3, 70.6, 69.7, 69.1, 68.7, 68.4, 67.7, 67.5, 63.8, 62.7, 53.3, 50.8, 50.5, 50.0, 37.7, 23.4, 21.2, 20.9, 20.8, 20.7. HR ESI MS (m/z) calcd. for $C_{30}H_{42}F_3N_5NaO_{18}$ $(M + Na)^+$ 840.2375, found 840.2365.

 $N-\{2-O-\{O-\{Methyl\ 4,7,8,9-tetra-O-acetyl-3,5-dideoxy-5-trifluoroacetamido-D-glycero-\alpha-D-acetyl-3,5-dideoxy-5-trifluoroacetamido-D-glycero-\alpha-D-acetyl-3,5-dideoxy-5-trifluoroacetamido-D-glycero-acetyl-3,5-dideoxy-5-trifluoroacetamido-D-glycero-acetyl-3,5-dideoxy-5-trifluoroacetamido-D-glycero-acetyl-3,5-dideoxy-5-trifluoroacetamido-D-glycero-acetyl-3,5-dideoxy-5-trifluoroacetamido-D-glycero-acetyl-3,5-dideoxy-5-trifluoroacetamido-D-glycero-acetyl-3,5-dideoxy-5-trifluoroacetamido-D-glycero-acetyl-3,5-dideoxy-5-trifluoroacetamido-D-glycero-acetyl-3,5-dideoxy-5-trifluoroacetamido-D-glycero-acetyl-3,5-dideoxy-5-trifluoroacetamido-D-glycero-acetyl-3,5-dideoxy-5-trifluoroacetamido-D-glycero-acetyl-3,5-dideoxy-5-trifluoroacetamido-D-glycero-acetyl-3,5-dideoxy-5-trifluoroacetamido-D-glycero-acetyl-3,5-dideoxy-5-trifluoroacetamido-D-glycero-acetyl-3,5-dideoxy-5-trifluoroacetyl-3,5-dideoxy-5-trifluoroacetyl-3,5-dideoxy-5-trifluoroacetyl-3,5-dideoxy-5-trifluoroacetyl-3,5-dideoxy-5-trifluoroacetyl-3,5-dideoxy-5-trifluoroacetyl-3,5-dideoxy-5-trifluoroacetyl-3,5-dideoxy-5-trifluoroacetyl-3,5-dideoxy-5-trifluoroacetyl-3,5-dideoxy-5-trifluoroacetyl-3,5-dideoxy-5-trifluoroacetyl-3,5-dideoxy-5-trifluoroacetyl-3,5-dideoxy-5-trifluoroacetyl-3,5-dideoxy-5-trifluoroacetyl-3,5-dideoxy-5-trifluoroacetyl-3,5-dideoxy-5-trifluoroacetyl-3,5-dideoxy-5-trifluoroacetyl-3,5-dideoxy-5-trifluoroacetyl-3,5-dideoxy-5-dideoxy-5-trifluoroacetyl-3,5-dideoxy-5-trifluoroacetyl-3,5-dideoxy-5-trifluoroacetyl-3,5-dideoxy-5-trifluoroacetyl-3,5-dideoxy-5-dideoxy-5-trifluoroacetyl-3,5-dideoxy-5-dideoxy-5-trifluoroacetyl-3,5-dideoxy-5$ galacto-2-nonulopyranosylonate]- $(2\rightarrow 6)$ -2-acetamido-2-deoxy- α -D-galactopyranosyl}-ethyl **4-pentenamide (6).** After a solution of **5** (0.4 g, 0.489 mmol) in MeOH (10 mL) was stirred with 10% Pd/C (90 mg) under a H₂ atmosphere at rt overnight, the catalyst was filtered off, and then 4-pentenoic anhydride (0.2 mL, 0.979 mmol) was added at 0 °C to the filtrate. The mixture was stirred at rt overnight. The solution was concentrated under vacuum and the residue was purified by flash chromatography (DCM/MeOH 15:1) to afford 6 as a white foamy solid (0.28 g, 68%). $R_f 0.25$ (DCM/MeOH 10:1). ¹H NMR (CDCl₃, 400 MHz): δ 6.95 (d, J 6.4 Hz, 1H, NHCOCH₃), 6.58 (d, J 9.6 Hz, 1H, NHCOCF₃), 6.07 (t, J 5.2 Hz, 1H, OCH₂CH₂NHCO), 5.87-5.76 (m, 1H, CH=CH₂), 5.41-5.35 (m, 1H, H-8'), 5.29 (dd, J 8.0, 1.6 Hz, 1H, H-7'), 5.10-4.96 (m, 3H, CH=CH₂ and H-4'), 4.87 (d, J 4.0 Hz, 1H, H-1), 4.77 (br, 1H, 3-OH), 4.34 (dd, J 12.0, 2.4 Hz, 1H, H-9'a), 4.29 (dd, J 10.8, 2.4 Hz, 1H, H-6'), 4.23-4.16 (m, 1H, H-2), 4.08 (dd, J 12.8, 6.0 Hz, 1H, H-9'b), 4.00 (q, J 9.6 Hz, 1H, H-5'), 3.91-3.83 (m, 3H, H-4, H-5 and H-6a), 3.82 (s, 3H, COOCH₃), 3.77-3.62 (m, 5H, H-3, H-6b and OCH₂CH₂N₃), 3.31-3.23 (m, 1H, OCH₂CH₂N₃), 2.95 (br, 1H, 4-OH), 2.64 (dd, J 12.8, 4.8 Hz, 1H, H-3'e), 2.43-2.36 (m, 2H, CH₂CH₂CH=CH₂), 2.34-2.28 (m, 2H, CH₂CH₂CH=CH₂), 2.15, 2.14, 2.13 (s, 3 x 3H, Ac), 2.03, 2.02 (s, 2 x 3H, Ac), 1.95 (t, J 12.8 Hz, 1H, H-3'a). ¹³C NMR (CDCl₃, 100 MHz): δ 173.8, 173.6, 171.3, 171.0, 170.7, 170.0, 168.2, 158.1, 157.8, 137.0, 116.9, 116.0, 114.6, 99.1, 97.8, 72.2, 71.0, 69.5, 69.0, 68.8, 68.2, 67.9, 67.7, 63.8, 62.7, 53.3, 50.8, 49.9, 39.3, 37.7, 35.9, 29.7, 23.0, 21.2, 21.0, 20.9, 20.8. HR ESI MS (m/z) calcd. for $C_{35}H_{50}F_3N_3NaO_{19}$ $(M + Na)^+$ 896.2888, found 896.2861.

General procedure for the synthesis of compounds 7a-d. To a stirred solution of 6 (180.0 mg, 0.206 mmol) in methanol (4.0 mL) was added a sodium methoxide solution in methanol (0.1 N, 0.4 mL). After the mixture was stirred at rt overnight, it was neutralized with Amberlite 15 (H⁺) resin. The resin was filtered off and washed with methanol. The filtrate and the washings were combined and concentrated to a small volume (ca. 3.0 mL), and then mixed with an aqueous NaOH solution (1.0 N, 1.0 mL). After the solution was stirred at rt overnight, it was neutralized with Amberlite 15 (H⁺) resin. The filtrate and the washings were combined and concentrated under vacuum to give the fully deprotected disaccharide 110 mg [MALDI TOF MS (m/z) calcd. for $C_{24}H_{40}N_3Na_2O_{14}$ (M - H + 2Na)⁺, 640.23, found 640.10], which was used directly in the next step without further purification. After the resultant disaccharide (28.0 mg, 0.047 mmol) was dissolved in MeOH (4.0 mL), a NaOH aqueous solution (1 N) was added to adjust the pH value of the solution to slightly basic, and then an individual acyl anhydride, i.e. (p-methylphenylacetic, p-methoxyphenylacetic, p-acetylphenylacetic or p-chlorophenylacetic anhydrides, 0.19 mmol), was added at 0 °C. After the reaction finished as shown by TLC, the mixture was concentrated under vacuum. The residue was dissolved in a small amount of H₂O, extracted with AcOEt to

remove any less polar materials and the product was then purified on a Biogel P-2 column using distilled water as the eluent to give **7a-d** as a white solid (30-33 mg) after lyophilization.

N-{2-*O*-{*O*-[3,5-Dideoxy-5-(*p*-methylphenylacetamido)-D-*glycero*-α-D-*galacto*-2-nonulopyranosylonic acid]-(2→6)-2-acetamido-2-deoxy-α-D-galactopyranosyl}-ethyl} 4-pentenamide (7a). ¹H NMR (D₂O, 400 MHz): δ 7.22 (s, 4H, aromatic H), 5.89-5.77 (m, 1H, CH=CH₂), 5.11-4.99 (m, 2H, CH=CH₂), 4.13 (dd, *J* 11.6, 3.6 Hz, 1H, H-2), 3.95 (br, 2H), 3.92-3.64 (m, 8H), 3.64-3.44 (m, 6H), 3.33-3.24 (m, 2H), 2.70 (dd, *J* 12.0, 4.0 Hz, 1H, H-3'e), 2.38-2.32 (m, 4H, COCH₂CH₂CH=CH₂), 2.31 (s, 3H, PhCH₃), 2.02 (s, 3H, NHCOCH₃), 1.66 (t, *J* 12.4 Hz, 1H, H-3'a). ¹³C NMR (D₂O, 100 MHz): δ 176.6, 176.1, 174.8, 173.4, 137.8, 137.3,

52.1, 50.0, 42.5, 40.5, 39.2, 35.3, 29.7, 22.3, 20.4. HR ESI MS (m/z) calcd. for $C_{33}H_{48}N_3Na_2O_{15}$ (M - H + 2Na)⁺ 772.2881, found 772.2878.

132.2, 129.8, 129.3, 115.9, 100.4, 97.4, 72.8, 72.0, 69.8, 68.7, 68.6, 68.1, 68.0, 66.9, 63.9, 63.1,

$N-\{2-O-\{O-[3,5-Dideoxy-5-(p-methoxyphenylacetamido)-D-glycero-\alpha-D-galacto-2-p-methoxyphenylacetamido\}$

nonulopyranosylonic acid]-(2→6)-2-acetamido-2-deoxy-α-D-galactopyranosyl}-ethyl} 4-pentenamide (7b). 1 H NMR (D₂O, 400 MHz): δ 7.26 (d, J 8.8 Hz, 2H, aromatic H), 6.98 (d, J 8.8 Hz, 2H, aromatic H), 5.89-5.78 (m, 1H, CH=CH₂), 5.11-4.99 (m, 2H, CH=CH₂), 4.82 (d, J 3.2 Hz, 1H, H-1), 4.13 (dd, J 11.6, 4.0 Hz, 1H, H-2), 3.98-3.94 (m, 2H), 3.91-3.84 (m, 1H), 3.82 (s, 3H, PhOCH₃), 3.83-3.77 (m, 3H), 3.76-3.58 (m, 5H), 3.57-3.45 (m, 5H), 3.33-3.25 (m, 2H), 2.71 (dd, J 12.0, 4.4 Hz, 1H, H-3'e), 2.38-2.30 (m, 4H, COCH₂CH₂CH=CH₂), 2.02 (s, 3H, NHCOCH₃), 1.65 (t, J 12.4 Hz, 1H, H-3'a). 13 C NMR (D₂O, 100 MHz): δ 176.6, 176.2, 174.8, 173.6, 158.3, 137.2, 130.6, 127.9, 115.9, 114.7, 100.6, 97.4, 72.8, 72.1, 69.8, 68.7, 68.6, 68.2, 68.0, 66.8, 63.9, 63.0, 55.7, 52.2, 50.0, 42.0, 40.7, 39.2, 35.3, 29.7, 22.3. HR ESI MS (m/z) calcd. for C₃₃H₄₈N₃Na₂O₁₆ (M - H + 2Na)⁺ 788.2830, found 788.2827.

$N-\{2-O-\{O-[5-(p-Acetylphenylacetamido)-3,5-dideoxy-D-glycero-\alpha-D-galacto-2-$

nonulopyranosylonic acid]-(2→6)-2-acetamido-2-deoxy-α-D-galactopyranosyl}-ethyl} 4-pentenamide (7c). 1 H NMR (D₂O, 400 MHz): δ 7.96 (d, J 8.8 Hz, 2H, aromatic H), 7.45 (d, J 8.0 Hz, 2H, aromatic H), 5.89-5.76 (m, 1H, CH=CH₂), 5.10-4.99 (m, 2H, CH=CH₂), 4.82 (d, J 3.2 Hz, 1H, H-1), 4.14 (dd, J 12.0, 3.6 Hz, 1H, H-2), 3.98-3.93 (m, 2H), 3.92-3.64 (m, 10H), 3.63-3.45 (m, 4H), 3.34-3.25 (m, 2H), 2.72 (dd, J 12.8, 4.8 Hz, 1H, H-3'e), 2.65 (s, 3H, CH₃COPh), 2.39-2.32 (m, 4H, COCH₂CH₂CH=CH₂), 2.02 (s, 3H, NHCOCH₃), 1.65 (t, J 12.4 Hz, 1H, H-3'a). 13 C NMR (D₂O, 100 MHz): δ 203.9, 176.6, 174.9, 174.7, 173.7, 141.6, 138.6, 137.2, 135.7, 129.7, 129.4, 128.9, 115.9, 100.6, 97.4, 72.8, 72.1, 69.8, 68.7, 68.6, 68.3, 68.0, 66.8, 63.9, 62.9, 52.2, 50.0, 42.8, 40.7, 39.2, 35.3, 29.7, 26.5, 22.3. HR ESI MS (m/z) calcd. for C₃₄H₄₈N₃Na₂O₁₆ (M - H + 2Na)⁺ 800.2830, found 800.2807.

N-{2-*O*-{*O*-[5-(*p*-Chlorophenylacetamido)-3,5-dideoxy-D-*glycero*-α-D-*galacto*-2-nonulopyranosylonic acid]-(2→6)-2-acetamido-2-deoxy-α-D-galactopyranosyl}-ethyl} 4-pentenamide (7d). ¹H NMR (D₂O, 400 MHz): δ 7.37 (d, *J* 8.0 Hz, 2H, aromatic H), 7.27 (d, *J* 8.0 Hz, 2H, aromatic H), 5.88-5.77 (m, 1H, CH=CH₂), 5.10-4.98 (m, 2H, CH=CH₂), 4.82 (d, *J* 4.0 Hz, 1H, H-1), 4.13 (dd, *J* 11.2, 4.0 Hz, 1H, H-2), 3.98-3.93 (m, 2H), 3.92-3.74 (m, 5H), 3.74-3.63 (m, 3H), 3.62-3.43 (m, 6H), 3.32-3.24 (m, 2H), 2.71 (dd, *J* 12.8, 4.8 Hz, 1H, H-3'e), 2.38-2.29 (m, 4H, COCH₂CH₂CH=CH₂), 2.01 (s, 3H, NHCOCH₃), 1.64 (t, *J* 12.0 Hz, 1H, H-3'a). ¹³C NMR (D₂O, 100 MHz): δ 176.5, 175.5, 174.7, 173.6, 137.2, 133.9, 132.8, 130.9, 129.1, 128.8, 115.9, 100.6, 97.4, 72.8, 72.1, 69.8, 68.7, 68.6, 68.3, 68.0, 66.8, 63.9, 63.0, 52.2, 50.0, 42.2, 40.7, 39.2, 35.3, 29.7, 22.3. HR ESI MS (*m*/*z*) calcd. for C₃₂H₄₅ClN₃Na₂O₁₅ (M - H + 2Na)⁺ 792.2335, found 792.2335.

General procedure for the synthesis of compounds 8a-d. To the stirred solution of 7a-d (18 mg) in MeOH (5 mL) at -78 °C, ozone was bubbled until a blue color appeared and remained at -78 °C for 0.5 h. After introducing nitrogen to remove the remaining ozone, Me₂S (0.5 mL) was added at -78 °C. The resultant solutions were allowed to warm to rt over a period of 1 h and stand for another 1 h before it was condensed in vacuum. The crude products were purified by a Biogel P-2 column using distilled water as the eluent to give the aldehydes after lyophilization as white solids, which were used in the following conjugation reactions without further purification.

N-{2-O-{O-[3,5-Dideoxy-5-(p-methylphenylacetamido)-D-glycero-α-D-galacto-2-nonulopyranosylonic acid]-(2 \rightarrow 6)-2-acetamido-2-deoxy-α-D-galactopyranosyl}-ethyl} 4-oxo-butanamide (8a). 1 H NMR (D₂O, 500 MHz): δ 7.12 (s, 4H, aromatic H), 5.34-5.23 (m, 1H, -CH(OH)₂), 4.74 (d, J 3.5 Hz, 1H, H-1), 4.05-3.99 (m, 1H), 3.29-3.14 (m, 2H), 2.21 (s, 3H, PhCH₃), 1.91 (s, 3H, NHCOCH₃). HR ESI MS (m/z) calcd. for C₃₂H₄₆N₃O₁₆ (M – H)⁺ 728.2878, found 728.2861.

N-{2-*O*-{*O*-[3,5-Dideoxy-5-(*p*-methoxyphenylacetamido)-D-*glycero*-α-D-*galacto*-2-nonulopyranosylonic acid]-(2→6)-2-acetamido-2-deoxy-α-D-galactopyranosyl}-ethyl} 4-oxo-butanamide (8b). ¹H NMR (D₂O, 500 MHz): δ 7.18 (d, *J* 8.0 Hz, 2H, aromatic H), 6.89 (d, *J* 8.0 Hz, 2H, aromatic H), 5.37-5.23 (m, 1H, -<u>CH</u>(OH)₂), 4.74 (d, *J* 3.5 Hz, 1H, H-1), 4.07-3.98 (m, 1H), 3.70 (s, 3H, PhO<u>CH₃</u>), 1.91 (s, 3H, NHCO<u>CH₃</u>). HR ESI MS (*m/z*) calcd. For C₃₂H₄₆N₃O₁₇ (M – H)⁺ 744.2827, found 744.2822.

N-{2-O-{O-[5-(p-Acetylphenylacetamido)-3,5-dideoxy-D-glycero- α -D-galacto-2-nonulopyranosylonic acid]-(2 \rightarrow 6)-2-acetamido-2-deoxy- α -D-galactopyranosyl}-ethyl} 4-oxo-butanamide (8c). 1 H NMR (D₂O, 500 MHz): δ 7.85 (d, J 8.5 Hz, 2H, aromatic H), 7.34 (d,

J 8.5 Hz, 2H, aromatic H), 5.34-5.23 (m, 1H, -CH(OH)₂), 4.73 (d, J 3.5 Hz, 1H, H-1), 4.05-3.98 (m, 1H), 3.90-3.54 (m, 12H), 3.54-3.43 (m, 2H), 3.42-3.33 (m, 2H), 3.26-3.18 (m, 2H), 2.53 (s, 3H, PhCOCH₃), 1.91 (s, 3H, NHCOCH₃). HR ESI MS (m/z) calcd. for C₃₃H₄₆N₃Na₂O₁₇ (M - H + 2Na)⁺ 802.2623, found 802.2612.

N-{2-O-{O-[5-(p-Chlorophenylacetamido)-3,5-dideoxy-D-glycero-α-D-galacto-2-nonulopyranosylonic acid]-(2 \rightarrow 6)-2-acetamido-2-deoxy-α-D-galactopyranosyl}-ethyl} 4-oxo-butanamide (8d). 1 H NMR (D₂O, 500 MHz): δ 7.28 (d, J 8.5 Hz, 2H, aromatic H), 7.17 (d, J 8.5 Hz, 2H, aromatic H), 5.34-5.23 (m, 1H, -CH(OH)₂), 4.73 (d, J 3.5 Hz, 1H, H-1), 4.06-3.99 (m, 1H), 3.90-3.35 (m, 16H), 3.27-3.16 (m, 2H), 1.91 (s, 3H, NHCOCH₃). HR ESI MS (m/z) calcd. for C₃₁H₄₃ClN₃Na₂O₁₆ (M - H + 2Na)⁺ 794.2127, found 794.2136.

General procedure for the coupling 8a-d to KLH and HSA. A solution of 8a-d (7 mg), KLH or HSA (7 mg), and NaBH₃CN (7 mg) in 0.1 M NaHCO₃ solution (0.1 mL, pH 7.5-8.0) was allowed to stand at rt in the dark for 4 days with occasional shaking. The reaction mixture was then purified with a Biogel A 0.5 column using 0.1 M PBS buffer (I = 0.1, pH =7.8) as the eluent. Fractions containing the glycoconjugate, as characterized by the bicinchoninic acid (BCA) assay for proteins and the resorcinol assay for sialic acid, were combined and dialyzed against distilled water for 2 days. The solution was then lyophilized to afford white solids of the desirable glycoconjugates 1a-d and 2a-d (ca. 6-7 mg).

Analysis of the carbohydrate loading levels of glycoconjugates 1a-d and 2a-d.² The solution of an exactly weighed glycoconjugate (0.35-0.6 mg) in distilled water (1.0 mL) was mixed with the resorcinol reagent (2.0 mL), and the mixture was heated in a boiling water bath for 30 min. It was cooled to rt, and to the solution was then added an extraction solution (1-butanol acetate and 1-butanol, 85:15 v/v, 3.0 mL). The mixture was shaken vigorously before it was allowed to stand still for *ca.* 10 min to allow the organic and inorganic layers to be separated well. The organic layer was transferred to a 1.0-cm cuvette, and its absorbance at 580 nm was determined with an UV-Vis spectrometer, utilizing a blank extraction solution as the control. The sialic acid contents of the glycoconjugates were determined against a calibration curve created with the solution of individual standard NeuNPhAc derivatives analyzed under the same condition. The carbohydrate loading of each glycoconjugate was calculated according to the following equation, and the results are shown in Table 1.

Carbohydrate loading (%) = $\frac{\text{carbohydrate content (mg) in the sample}}{\text{weight of the gly coconjugate sample (mg)}} \times 100\%$

Table 1. Antigen Loading Levels of the Conjugates

sample .		KLH co	njugates		HSA conjugates			
	1a	1b	1c	1d	2a	2b	2c	2d
Loading (%)	3.6	4.3	2.9	5.4	5.8	7.6	5.7	6.9

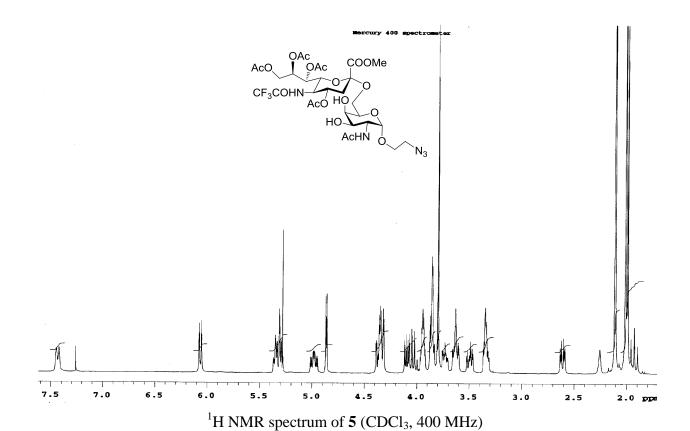
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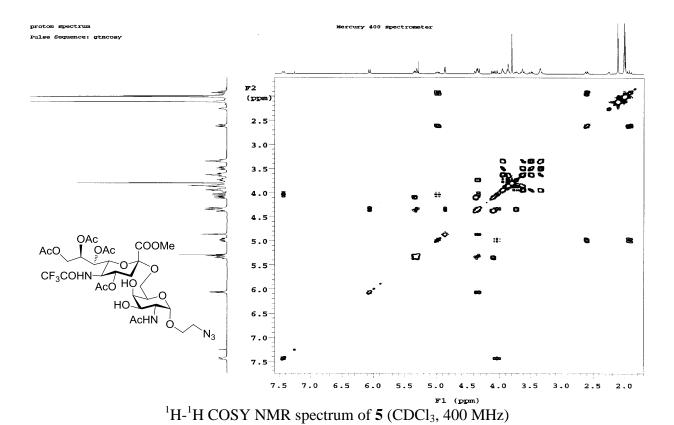
- 1. Wang, Q.; Ekanayaka, S. A.; Wu, J.; Zhang, J.; Guo, Z. Synthetic and Immunological Studies of 5'-N-Phenylacetyl sTn to Develop Carbohydrate-Based Cancer Vaccines and to Explore the Impacts of Linkage between Carbohydrate Antigens and Carrier Proteins. *Bioconjugate Chemistry* **2008**, 19, 2060-2067.
- 2. Svennerholm, L. Quantitative estimation of sialic acids. II. A colorimetric resorcinol-hydrochloric acid method. *Biochim Biophys Acta* **1957**, 24, 604-11.

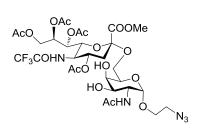
Immunological Studies

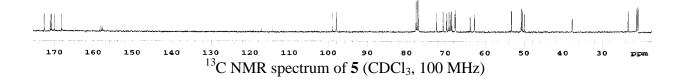
Immunization of Mice. Groups of female C57BL/6 mice (5 per group) at the age of 6-8 weeks (Jackson Laboratories, Bar Harbor, ME) were immunized into 3 intramuscular sites with a total of 0.1 mL of the emulsion of **1a-d** (containing 2 μg of carbohydrate antigen) and Titermax Gold adjuvant (Sigma Chemical, St. Louis, MO) on day 0, 14, 21 and 28, respectively. The mice were bled on day -1 prior to the initial immunization and after immunization on day 27 and day 37. Blood samples collected at each time point were clotted to obtain antisera and stored at -80 °C.

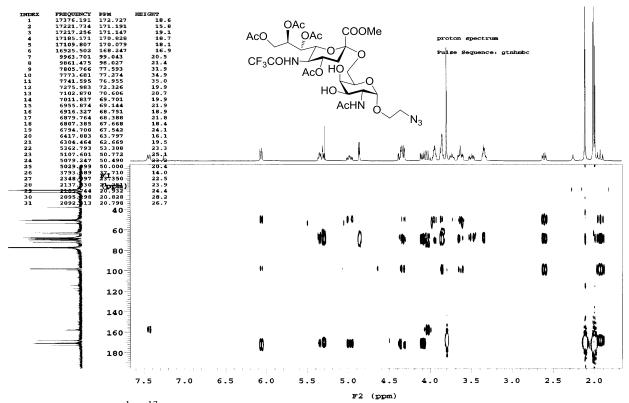
Enzyme-Linked Immunosorbent Assay (ELISA). ELISA plates were first treated respectively with 100 μL of sTn-HSA, sTnNPhAc-HSA or conjugate **2a-d** solution (2 μg/mL) in the coating buffer (0.1 M bicarbonate, pH 9.6) overnight at 4 °C, followed by washing 3 times with PBS containing 0.05% Tween-20 (PBST). Individual or pooled antisera from **1a-d** inoculated mice were diluted 1:300 to 1:72900 in serial half-log dilutions in PBS and incubated for 2 h at 37 °C in the coated ELISA plates (100 μL/well). The plates were then washed and incubated with 1:1000 dilution of alkaline phosphatase linked goat anti-mouse kappa, IgM or IgG2a antibody or with a 1:2000 dilution of alkaline phosphatase linked goat anti-mouse IgG1 or IgG3 antibody for 1 h at rt. Finally, the plates were washed and developed with 100 μL of PNPP solution (1.67 mg/mL in PNPP buffer) for 30 min at rt for colorimetric readout using a BioRad 550 plate reader at 405 nm wavelength. For titer analysis, optical density (OD) values were plotted against dilution values, and a best-fit line was obtained. The equation of this line was employed to calculate the dilution value at which an OD of 0.5 was achieved, and the antibody titer was calculated at the inverse of this dilution value.











¹H-¹³C HMBC NMR spectrum of **5** (CDCl₃, 400/100 MHz)

Elemental Composition Report

ÇOOMe CF₃COHN-AcO HO

Single Mass AnalysisTolerance = 5.0 PPM / DBE: min = -1.5, max = 50.0 Element prediction: Off

Number of isotope peaks used for i-FIT = 3

Monoisotopic Mass, Even Electron Ions 3063 formula(e) evaluated with 24 results within limits (up to 50 best isotopic matches for each mass) N_3

Elements Used:
C: 0-500 H: 0-1000 N: 3-6 O: 10-20 23Na: 0-1 F: 0-4

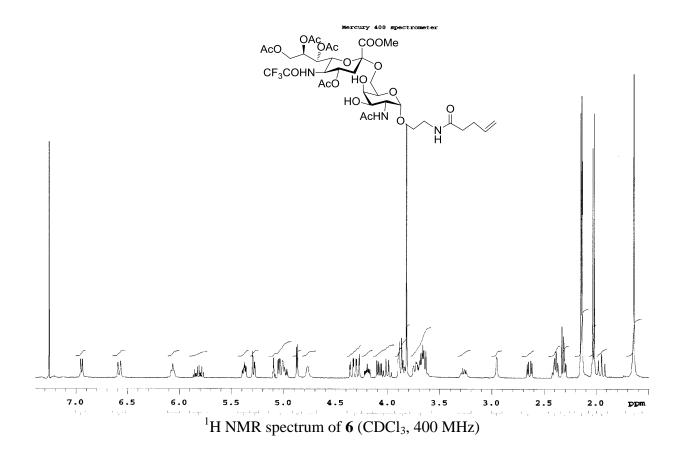
Q. WANG 2008-07b.pro 2009_0710_0487 13 (0.283) Cm (12:16-(1:7+28:34)x4.000)

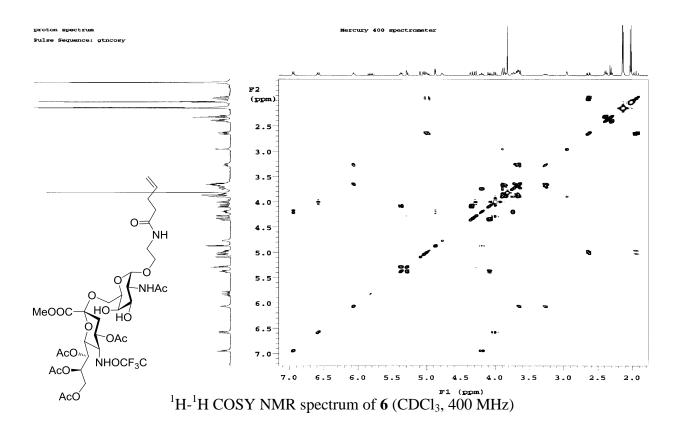
LCT Premier 10-Jul-2009 14:48:52 1: TOF MS ES+ 3.50e+003

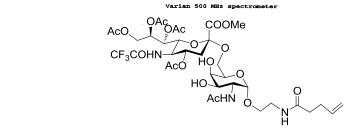
Page 1

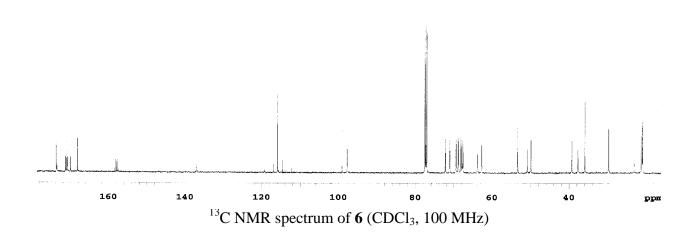
1						1						
% -	944 0246	000 000				8	41.2413 842.2406				0.5	
0		2584 820.232		830.9451 8			نىزدىدىدىدار	9.2956	بيتنين	56.22		9.0329 m/z
81	10.0 815.0	820.0	825.0	830.0	835.0	840.0	845.0	850.0	85	55.0	860	0.0
Minimum: Maximum:		5.0	5.0	-1.5 50.0								
Mass (Calc. Mass	mDa	PPM	DBE	i-FIT	i	-FIT (Norm)	Form	ula			
840.2365	340.2375	-1.0	-1.2	10.5	54.5	C	0.3	C30 23Na		N5	018	e garb
8	340.2386	-2.1	-2.5	6.5	56.5	2	2.2	C27 23Na	H43	N 5	019	
8	340.2363	0.2	0.2	14.5	57.3	3	3.1	C33 23Na	H41	N5	017	
	340.2399 340.2399	-3.4 -3.4	-4.0 -4.0	13.5 13.5	57.7 58.1		3.4 3.9	C32 C32	H41 H43	N5 N5	018 020	F3
E	340.2387	-2.2	-2.6	17.5	59.6		5.4	23Na C35	H40	N5	017	F2
٤	340.2351	1.4	1.7	18.5	59.9		.7	C36	н38	N5	014	F4
	340.2352	1.3	1.5	18.5	60.0		5.8	C36 23Na	H40	N5	016	1.1
	340.2327	3.8	4.5	15.5	60.2	ϵ	5.0	C34 23Na	H39 F4	N5	014	
	340.2368	-0.3	-0.4	19.5	60.6	6	5.3	C39 23Na		И3	012	
	340.2376	-1.1	-1.3	21.5	60.8	6	5.5	C38	H39	N5	016	F
8	340.2364	0.1	0.1	25.5	61.8	7	.5	C41	H38	N5	015	
	340.2356	0.9	1.1	23.5	61.9		.7	C42 23Na		И3	011	
	340.2403	-3.8	-4.5	18.5	62.1		.9	C38 23Na		и3	015	
	340.2340	2.5	3.0	22.5	62.2		.0	C39	H37	N5	013	F3
	340.2340	2.5	3.0	22.5	62.3		.0	C39 23Na	Н39	N5	015	
	340.2392	-2.7	-3.2	22.5	62.4		.1	C41	н38	NЗ	012	F4
	340.2392	-2.7	-3.2	22.5	62.5		.3	C41 23Na		ИЗ	014	
	340.2380 340.2381	-1.5 -1.6	-1.8 -1.9	26.5 26.5	62.7 62.7		. 4	C44	Н37 Н39	N3 N3	011 013	F3
ç	340.2369	-0.4	-0.5	30.5	63.1	0	٥	23Na	1126	17.2	010	770
	340.2345	2.0	2.4	27.5	63.5		.9	C47 C45 23Na	H36 H37 F2	N3 N3	010 010	F2
8	340.2329	3.6	4.3	26.5	64.1	9	. 8	C42	H36	N5	012	F2
8	340.2405	-4.0	-4.8	29.5	64.9		0.7	C46	Н38	N3	013	

HR ESI MS spectrum of 5









Elemental Composition Report

COOMe AcHŃ

Single Mass Analysis

Tolerance = 5.0 PPM / DBE: min = -1.5, max = 50.0 Element prediction: Off

Number of isotope peaks used for i-FIT = 3

Monoisotopic Mass, Even Electron Ions 6232 formula(e) evaluated with 39 results within limits (up to 50 best isotopic matches for each mass) Elements Used:

C: 0-50 H: 0-100 N: 0-5 O: 0-21 23Na: 0-1 F: 0-4

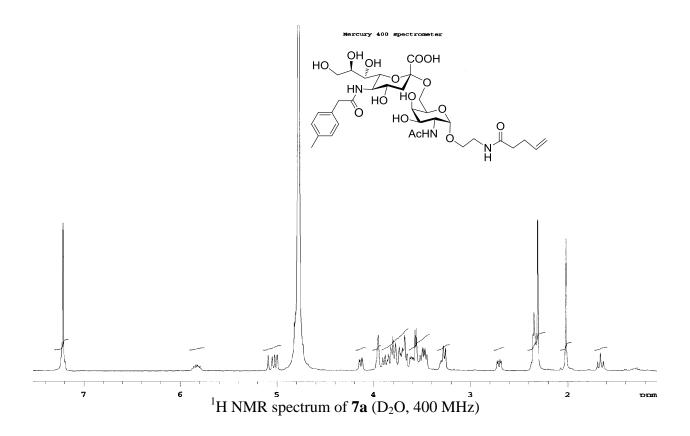
2009_0716_0497 13 (0.283) Cm (10:18-(1:7+40:45)x2.000) LCT Premier 16-Jul-2009 13:33:52 1: TOF MS ES+

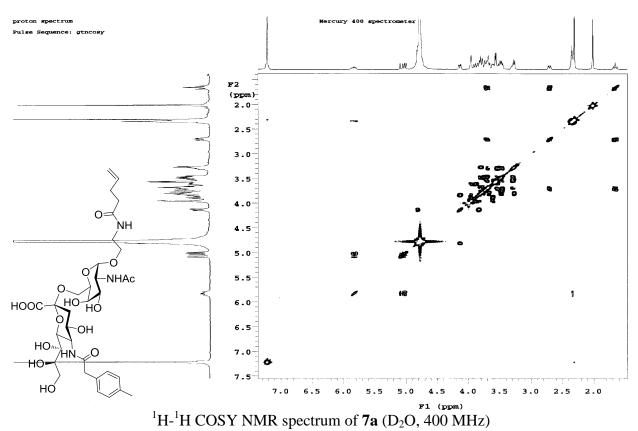
Q. WANG

Page 1

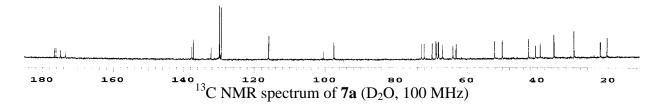
100				896	.2861				1	.37e+005
%- 810.21:	⁵⁹ 829.2466		874.30 368.2935		897.2939 898.2972 91	8.2719 9	47.3972	972.;	²⁷⁵³ 978	3.2919 _
810 8	20 830 840	850 860	870	880 890	900 910	920 930 940	950 96	50	970	980
Minimum: Maximum:		5.0	5.0	-1.5 50.0						
Mass	Calc. Mass	mDa	PPM	DBE	i-FIT	i-FIT (Norm)	Formula			
874.3073	874.3074	-0.1	-0.1	15.5	17.2	7.9	C41 H51 23Na F2		016	
	874.3075	-0.2	-0.2	26.5	20.5	11.2	C49 H48	N	014	
	874.3075	-0.2	-0.2	24.5	19.2	9.9	C45 H44	N5	09	F4
	874.3076	-0.3	-0.3	24.5	19.3	10.0	C45 H46	N5	011	
	874.3069	0.4	0.5	10.5	12.4	3.1	23Na F C35 H53 23Na	N3	021	
	874.3069	0.4	0.5	10.5	11.1	1.8	C35 H51	N3	3 019	F3 ←1
	874.3080	-0.7	-0.8	6.5	11.7	2.4	C32 H52			
	874.3064	0.9	1.0	28.5	20.3	11.1	C48 H43			F3
	874.3064	0.9	1.0	28.5	20.4	11.1	C48 H45 23Na			
	874.3062	1.1	1.3	19.5	18.9	9.6	C44 H48	N	013	F4
	874.3062	1.1	1.3	19.5	18.9	9.7	C44 H50 23Na F		015	1.1
	874.3085	-1.2	-1.4	11.5	15.1	5.8	C38 H52 23Na F3	N	017	
	874.3086	-1.3	-1.5	22.5	20.0	10.7	C46 H49	N	015	F
	874.3087	-1.4	-1.6	20.5	18.5	9.3	C42 H47 23Na F2	N5		-
	874.3088	-1.5	-1.7	31.5	21.3	12.0	C50 H44	N5	010	
	874.3057	1.6	1.8	14.5	16.1	6.8	C38 H50	И3	018	F2
	874.3056	1.7	1.9	3.5	15.6	6.3	C30 H53 23Na F4	и3		
	874.3091	-1.8	-2.1	25.5	20.8	11.6	C48 H45 23Na F4	из	07	
	874.3093	-2.0	-2.3	13.5	16.1	6.8	C37 H52	и3	021	
	874.3051	2.2	2.5	21.5	19.4	10.1	C43 H45	N5	09	
	874.3051	2.2	2.5	23.5	20.8	11.5	23Na F4 C47 H49	N	014	
	874.3050	2.3	2.6	23.5	20.7	11.4	23Na C47 H47	NT.	010	D 2
	874.3096	-2.3	-2.6	7.5	10.0	0.7	C47 H47 C35 H53 23Na F4	N N	012 018	F3
	874.3098	-2.5	-2.9	18.5	19.7	10.4	C43 H50	N	016	E.O.
	874.3098	-2.5	-2.9	16.5	17.6	8.4	C39 H48 23Na F3	N N5	016 013	F2
	874.3100	-2.7	-3.1	27.5	21.4	12.1	C47 H45	N5	011	F
	874.3046	2.7	3.1	18.5	19.1	9.8	C41 H49	N3	017	F
	874.3045	2.8	3.2	7.5	10.8	1.5	C33 H52	N3	019	-

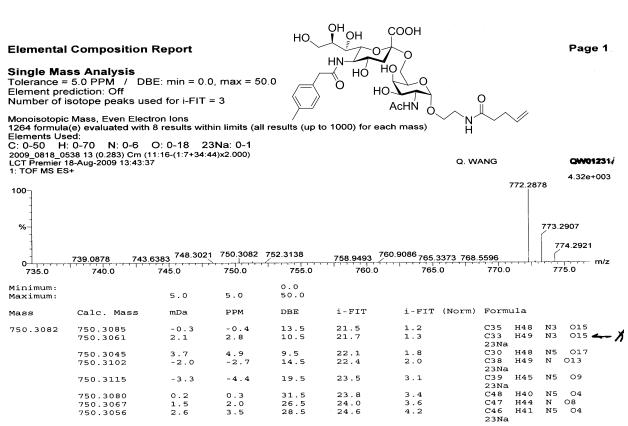
HR ESI MS spectrum of 6



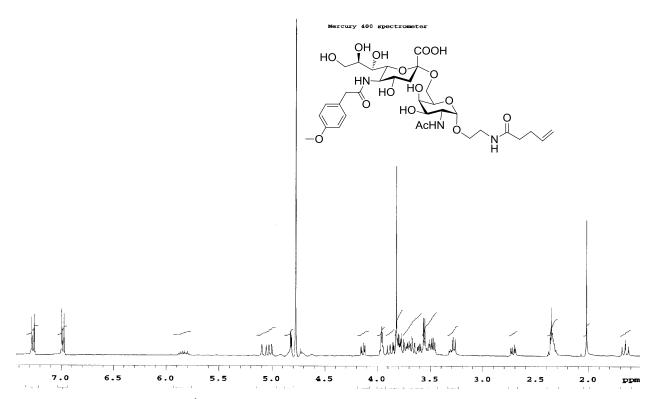


Mercury 400 spectrometer

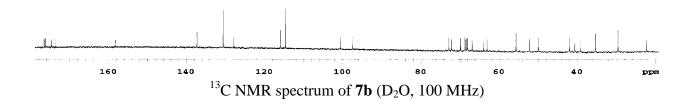


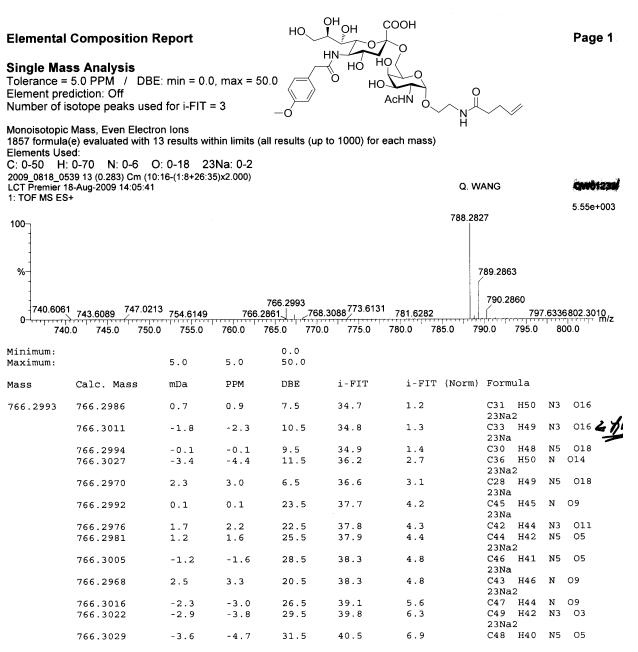


HR ESI MS spectrum of 7a

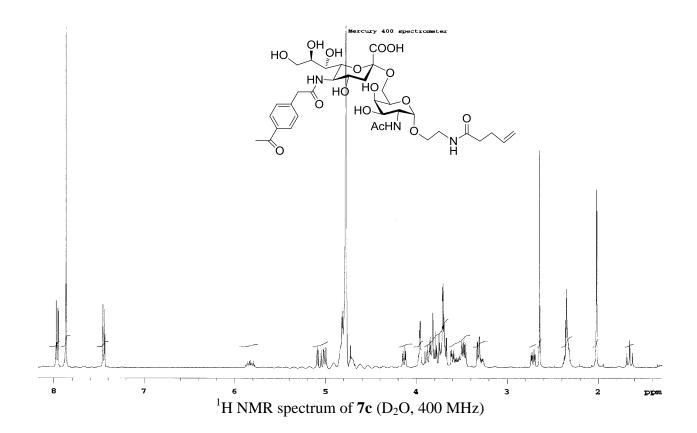


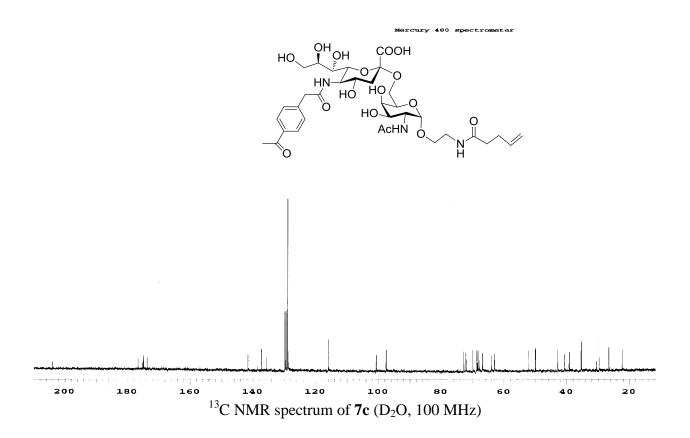
 ^{1}H NMR spectrum of **7b** (D₂O, 400 MHz)

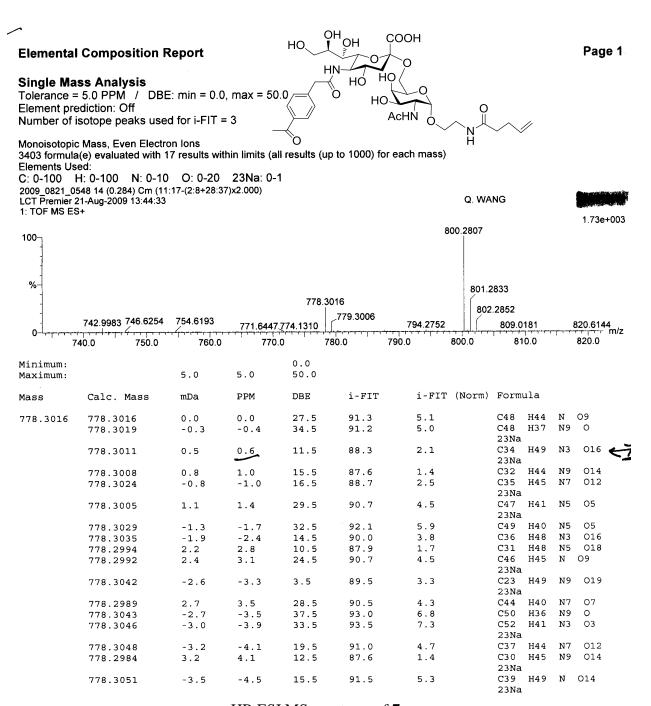




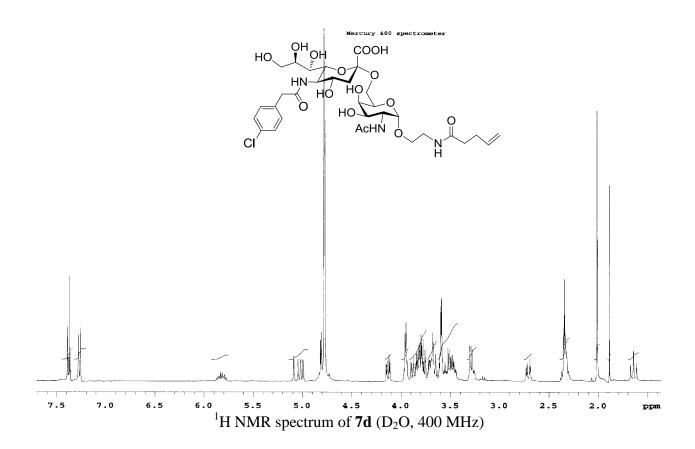
HR ESI MS spectrum of 7b

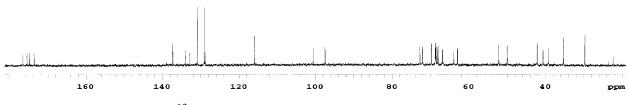






HR ESI MS spectrum of 7c





 13 C NMR spectrum of **7d** (D₂O, 100 MHz)

COOH Page 1 **Elemental Composition Report Single Mass Analysis**Tolerance = 5.0 PPM / DBE: min = 0.0, max = 50.0 ΗÓ Element prediction: Off Number of isotope peaks used for i-FIT = 3 AcHŃ Monoisotopic Mass, Even Electron lons 8127 formula(e) evaluated with 45 results within limits (all results (up to 1000) for each mass)

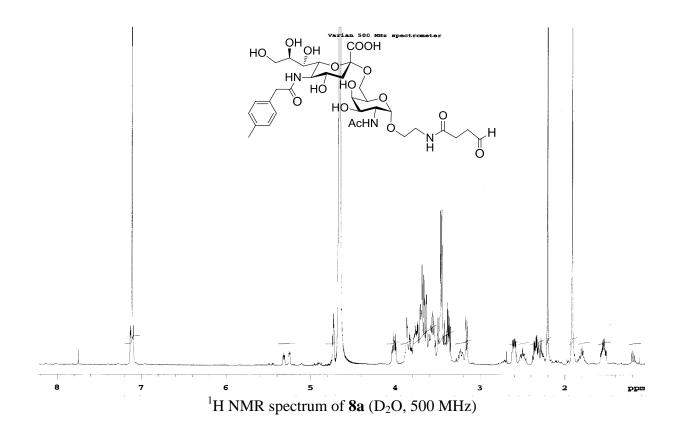
Elements Used:

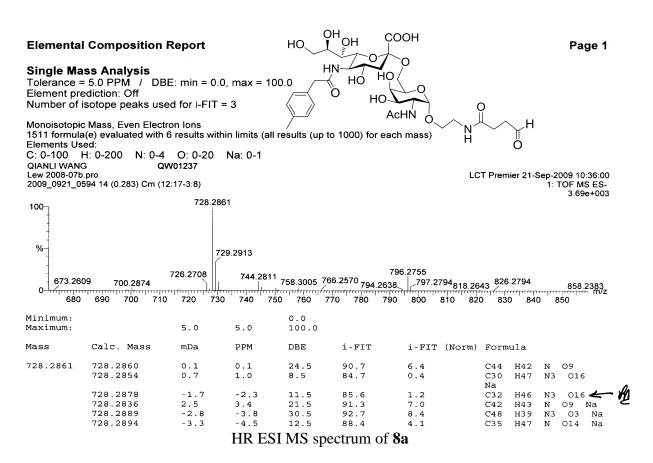
C: 0-100 H: 0-100 N: 0-5 O: 0-20 23Na: 0-2 CI: 0-2 2009_0825_0555a 12 (0.263) Cm (10:15-(1:6+38:45)x2.000) LCT Premier 25-Aug-2009_16:07:38 1: TOF MS ES+

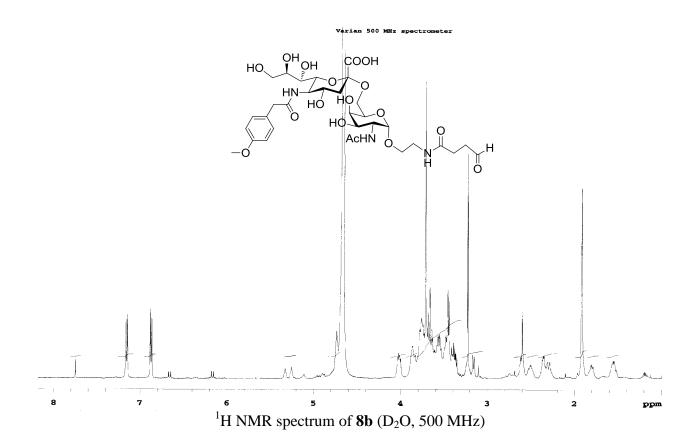
QIANLI WANG	(Charles
	1 590+00

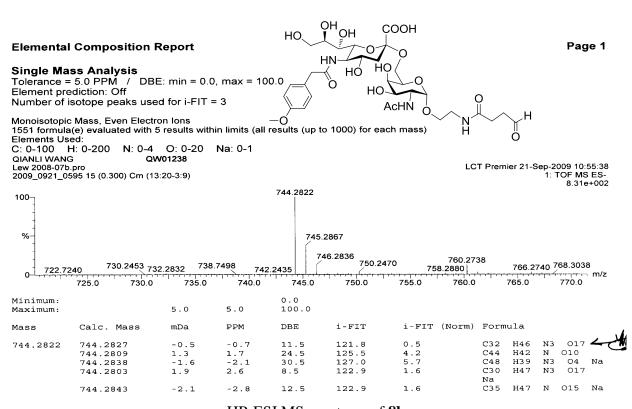
100-			792.23	335				7.000 - 000
100								
%-				793.2385				
1								
774.	9966_776.6085	784.0867	791.6050	795.2372	799.5995	810.5921814.22	\ 010.0130 /	839 822.4267
0 - 1 - 1 - 1 - 1 - 1 - 1		785.0	790.0		800.0 80	05.0 810.0	815.0 820.	111/2
Minimum: Maximum:		5.0	5.0	0.0 50.0				
Mass	Calc. Mass	mDa	PPM	DBE	i-FIT	i-FIT (Norm) Formula	
792.2335	792.2335	0.0	0.0	10.5	115.8	1.7	C32 H45 N 23Na2 Cl	3 015 - XII
	792.2337	-0.2	-0.3	4.5	123.2	9.1	C28 H49 N 23Na Cl2	3 017
	792.2338	-0.3	-0.4	32.5	121.7	7.6	C50 H36 N 23Na2	06
	792.2332	0.3	0.4	22.5	122.8	8.6	C41 H41 N 23Na Cl2	5 06
	792.2340	-0.5	-0.6	18.5	121.4	7.2	C35 H39 N 23Na	5 015
	792.2340	-0.5	-0.6	26.5	117.2	3.0	C46 H40 N 23Na Cl	08
	792.2329	0.6	0.8	28.5	117.8	3.6	C45 H37 N 23Na2 Cl	5 04
	792.2342	-0.7	-0.9	20.5	122.4	8.3	C42 H44 N	010
	792,2342	-0.7	-0.9	12.5	115.9	1.8	C31 H43 N	5 017
	792.2327	0.8	1.0	47.5	122.0	7.8	C61 H30 N	0
	792.2327	0.8	1.0	13.5	121.4	7.2	C34 H43 N 23Na	019
	792.2346	-1.1	-1.4	34.5	121.8	7.6	C49 H34 N	3 08
	792.2324	1.1	1.4	25.5	117.5	3.3	C43 H39 N Cl	3 010
	792.2322	1.3	1.6	31.5	121.7	7.5	C47 H35 N 23Na	3 08
	792.2348	-1.3	-1.6	23.5	122.2	8.1	C44 H42 N 23Na2 Cl2	3 04
	792.2321	1.4	1.8	3.5	124.1	9.9	C25 H48 N	5 019
	792.2351	-1.6	-2.0	16.5	121.7	7.6	C36 H42 N	019
	792.2351	-1.6	-2.0	37.5	122.2	8.0	C51 H32 N 23Na2	5 02
	792.2318	1.7	2.1	9.5	117.8	3.6	C29 H44 N 23Na Cl	5 017
	792.2318	1.7	2.1	17.5	123.4	9.2	C40 H45 N 23Na Cl2	010
	792.2353	-1.8	-2.3	5.5	122.8	8.7	C31 H50 N 23Na2 Cl2	015
	792.2316	1.9	2.4	15.5	122.0	7.9	C33 H40 N 23Na2	5 015
	792.2354	-1.9	-2.4	31.5	117.6	3.5	C47 H36 N	5 04
			IID D	a				

HR ESI MS spectrum of 7d

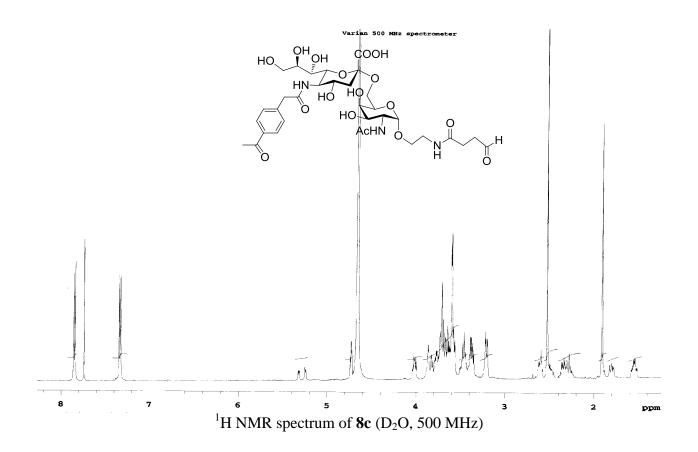


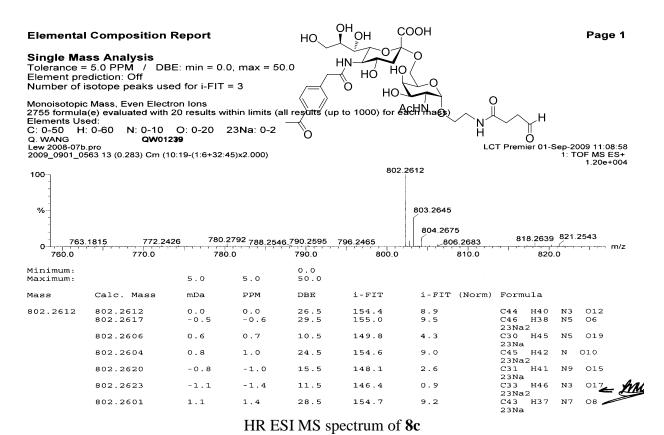


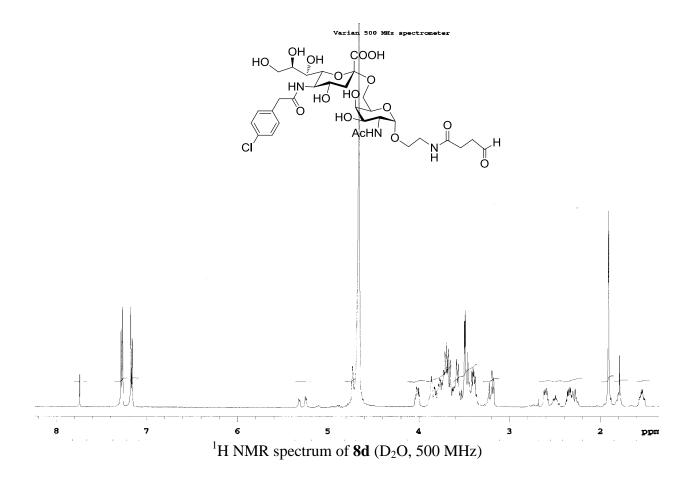


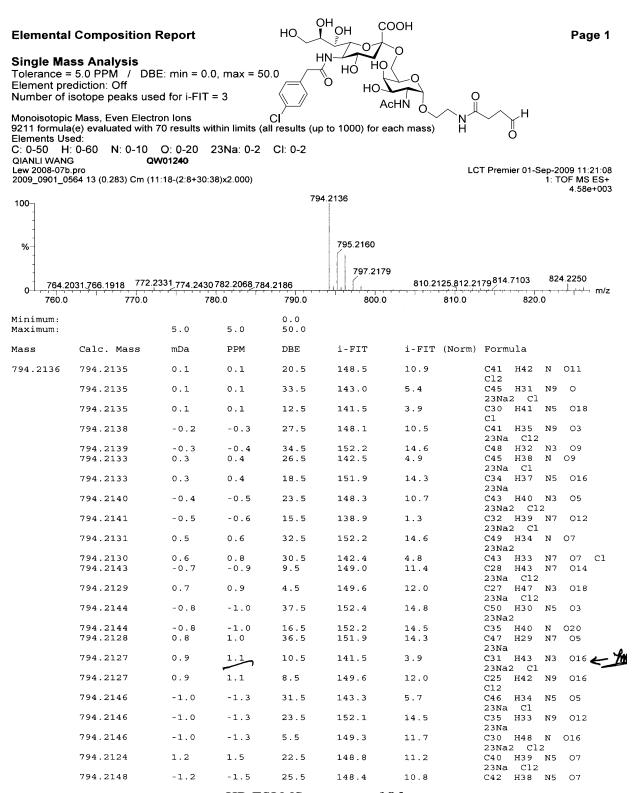


HR ESI MS spectrum of 8b









HR ESI MS spectrum of 8d