

SUPPLEMENTAL INFORMATION

**Type II fatty acid biosynthesis is essential for *Plasmodium falciparum* sporozoite
development in the midgut of *Anopheles* mosquitoes**

Ben C. L. van Schaijk, T. R. Santha Kumar, Martijn W. Vos, Adam Richman,
Geert-Jan van Gemert, Tao Li, Abraham G. Eappen, Kim C. Williamson,
Belinda J. Morahan, Matt Fishbaugher, Mark Kennedy, Nelly Camargo,
Shahid M. Khan, Chris J. Janse, Kim Lee Sim, Stephen L. Hoffman, Stefan H. I. Kappe,
Robert W. Sauerwein, David A. Fidock and Ashley M. Vaughan

Table S1. Primers used in this study

Primers	Sequence (5'-3')	Restriction site ^a	Target ^b
P1	ATTAACCCTCACTAAAGGGA	N/A	T3
P2	GATAGCGATTTTTTTTACTGTCTG	N/A	<i>hrp2-3'</i>
P3	AGAAAGAAGAACAAGAT	N/A	<i>fabI-F</i>
P4	TTGATTTAAGTGGTCCTGCACTG	N/A	<i>fabI -R</i>
P5	GCGGCGGCCGCTAATGATTTGTTTTATTGCTGGTATAG	NotI	<i>fabI -F^c</i>
P6	CGGGATCCTTAGGCTAAACTCTGGTATCAG	BamHI	<i>fabI -R^d</i>
P7	ATTGTGTATAATAAACATTTACGC	N/A	U <i>fabB/F^e</i>
P8	CTCTAAATTATTATCGTCCCTTG	N/A	D <i>fabB/F^e</i>
P9	CACTGCGCGCGTGTACACCTGCATGAGC	BssHII	<i>fabB/F -F</i>
P10	GATCGTACGCAGGAGCATTATTTAAGGAACAACC	BsiWI	<i>fabB/F -R</i>
P11	ATGCCCGGGATTTGGAGGCCATAACACAGC	XmaI	<i>fabB/F -F</i>
P12	GATGCTAGCGAGTGTTATGTAAAAGGGATAATGATC	NheI	<i>fabB/F -R</i>

^a Restriction sites are underlined. N/A, not applicable

^b F= forward primer, R= reverse primer

^c The NotI site is followed by a TAA stop codon and TG to change the frame of the disrupted downstream sequence

^d The BamHI site is followed by a TAG stop codon and a G to change the frame of the disrupted downstream sequence

^e Upstream (U) and Downstream (D) primers used for diagnostic long-range PCR

Fig. S1. Genetic deletion of *fabB/F* in *P. falciparum* NF54 parasites

(A) Schematic representation of the genetic deletion of *P. falciparum* (*Pf*) *fabB/F* by double crossover homologous recombination on chromosome 6. The targeting construct p-*PffabB/F* carries the resistance marker cassette human *dhfr* fused to *gfp* (*hdhfr-gfp*). This fusion is flanked by two FRT sites (white triangles), which serve as the recognition sites for the enhanced recombinase enzyme FLPe. The schematic shows the disrupted locus (*Pf* Δ *fabB/F*), as well as the locus from which the *dhfr-gfp* marker cassette was removed (*Pf* Δ *fabB/F* *FLPe), following transfection with the plasmid pMV-FLPe. p7, p8: primer pairs for long range PCR analysis. HindIII (H) was used for further characterization of the long range PCR products by restriction enzyme analysis; EcoRI (E) restriction sites were used for Southern blot analysis. *cam*: calmodulin; *hrp2*: histidine rich protein 2; *hsp86*: heatshock protein 86; *fcu*: cytosine deaminase/uracil phosphoribosyl-transferase; *pbd*: *P. berghei dhfr-ts* 3' untranslated region. (B) Long range PCR analysis of genomic DNA from WT and *Pf* Δ *fabB/F* before and after pMV-FLPe transfection (*Pf* Δ *fabB/F* *FLPe), confirmed the integration and removal of the *hdhfr-gfp* marker, respectively. (C) Southern analysis of genomic DNA from WT and mutant asexual blood stage parasites confirmed double crossover-based deletion of the *fabB/F* gene and the subsequent excision of the *hdhfr-gfp* marker in the FLPe-transfected *Pf* Δ *fabB/F* parasites. DNA was digested with EcoRI and probed with the *fabB/F* 3' untranslated region (shown as D-*fab b/f*). The expected product sizes were 10.6 kb for the plasmid Δ *fab b/f*, 6.0 kb for the wild type NF54 locus; 1.4 kb for the *Pf* Δ *fabB/F* locus (containing *dhfr-gfp* in the place of the original *fab B/F* gene); and 4.3 kb for the *Pf* Δ *fabB/F* *FLPe locus (no longer carrying *dhfr-gfp*).

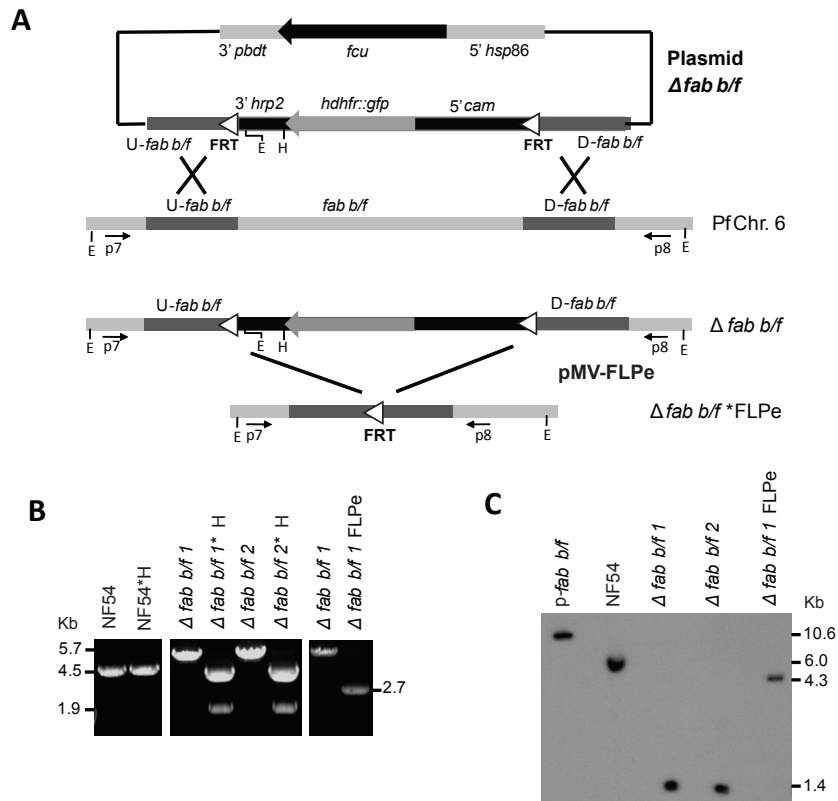


Fig. S2. Genetic disruption of *fabI* in *P. falciparum* NF54 parasites

(A) Representation of genetic disruption of *P. falciparum* (*Pf*) *fabI* by homologous recombination and single site crossover between the endogenous full-length 1.3 kb *PffabI* coding sequence and a truncated 0.6 kb *PffabI* fragment present on the transfection plasmid *pcam-bsd-PfΔfabI*. This resulted in an upstream truncated fragment lacking the 3' end of the coding sequence corresponding to the C-terminal 136 amino acids, followed by one plasmid copy, then a downstream truncated fragment lacking the *PffabI* promoter and 5' coding sequence corresponding to the N-terminal 98 amino acids. The schematic indicates the locations of the primers used to detect integration, as well as the HindIII (H) restriction sites and the *PffabI* probe (black bar) used for Southern blot analysis. (B) PCR results showing *PffabI* disruption via homologous recombination in the *PfΔfabI* B1 and C10 knockout clones. Lanes 1-4: *PfΔfabI* B1; Lanes 5-8: *PfΔfabI* C10; Lanes 9-12: wild type NF54. p1+p2 were specific for plasmid sequences and detected a 0.9 kb band in the knockout clones but not the wild type. p2+p3 produced a 1.1 kb band specific for the upstream truncated *PffabI* fragment only in the knockout clones. Similarly, p1+p4 produced a 0.7 kb band specific for the downstream-truncated *PffabI* fragment only in the knockout clones. p3+p4 detected a 0.8 kb band only in parasites harboring the endogenous full-length *PffabI* allele. The absence of p3+p4 bands in the knockout clones was consistent with complete genetic disruption of *PffabI* in these lines. Sizes of markers are indicated beside the gel. M, Gene Ruler (1 kb plus) marker. (C) Southern blot hybridization of HindIII-digested genomic DNA hybridized with a *PffabI* probe. Lanes are the same as for panel B. For the HindIII digests, the 8.4 kb and 2.1 kb bands in *PfΔfabI* B1 and *PfΔfabI* C10 were consistent with genetic disruption of the endogenous *PffabI* allele with a single copy insertion of the targeting plasmid. This contrasts with the unique 5.4 kb band in wild type NF54. Linearization of the plasmid with HindIII yielded a 5.1 kb band.

