The American Journal of Human Genetics, Volume *94* Supplemental Data

# Convergence of Genes and Cellular Pathways Dysregulated in Autism Spectrum Disorders

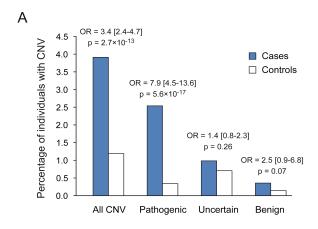
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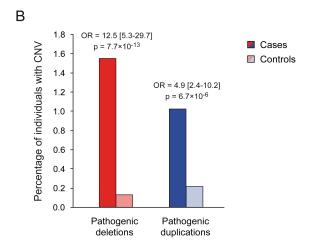
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# **SUPPLEMENTAL FIGURES**





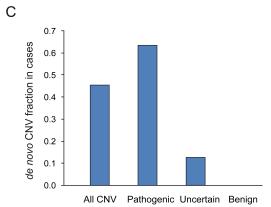
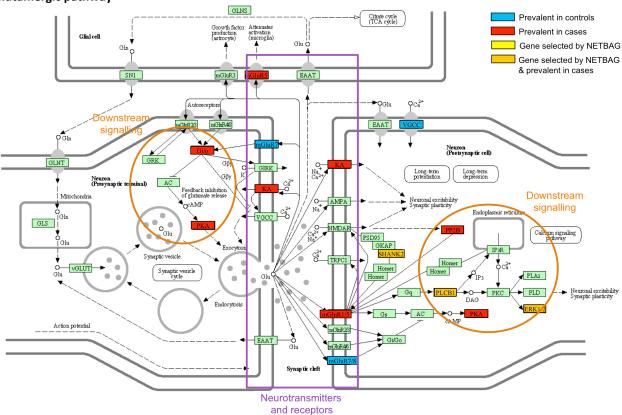


Figure S1. CNV burden in genes and loci implicated in ASD and ID in affected and control subjects of all ancestries

CNV data from 2,446 cases and 4,768 controls irrespective of ancestry were analyzed for overlaps with a list of genes and loci causally implicated in ASD/ID (**Tables S6A-S6D**). Overlap results include 299 non-European cases and 1,843 non-Europeans controls. Only CNVs affecting autosomal dominant and X-linked dominant genes/loci in both genders (132 genes, 56 loci) as well as X-linked recessive genes/loci in males (52 genes, 2 loci) were considered ('all CNV'). Exonic CNVs ≥30 kb affecting an ASD/ID gene and CNVs overlapping at least 50% of the target loci were selected for further analysis. After curation, CNVs were divided into three categories: pathogenic, uncertain clinical significance or benign. (**A**) Percentage of individuals with CNVs overlapping genes and loci implicated in ASD/ID ('all CNV'), pathogenic, uncertain or benign CNVs and odds ratio (OR) in cases and controls. (**B**) Percentage of individuals with pathogenic deletions or duplications and OR in cases and controls. (**C**) *De novo* CNV fraction in each category in cases.

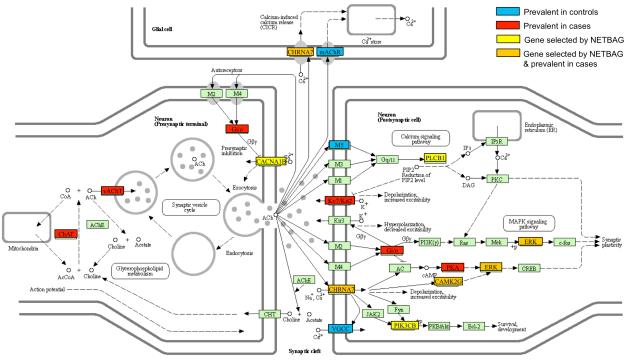
Figure S2. Projection of the glutamergic (A) and cholinergic (B) synapse subclusters enriched in cases over controls, onto the KEGG pathways

#### A. Glutamergic pathway



19 cases and 2 controls in the glutamatergic pathway: 11 deleted genes in 19 cases (average size= 635 kb, median size= 265 kb; average number of genes= 16, median number of genes= 3): GNG13 (1), MAPK3 (5), GNG2 (2), PRKACB (1), SHANK2 (3), PPP3CB (1), SHANK3 (3), GRIK2 (1), GRM5 (1), PLCB1 (1), SHANK1 (1); 2 deleted genes in 2 controls (average/median size= 85 kb; average/median number of genes= 1): CACNA1C, GRM7. Genes selected by NETBAG (main Figure 4B) are highlighted with yellow or orange shaded boxes.

#### B. Cholinergic pathway



17 cases and 2 controls in the cholinergic pathway. Genes selected by NETBAG (main Figure 4B) are highlighted with yellow or orange shaded boxes

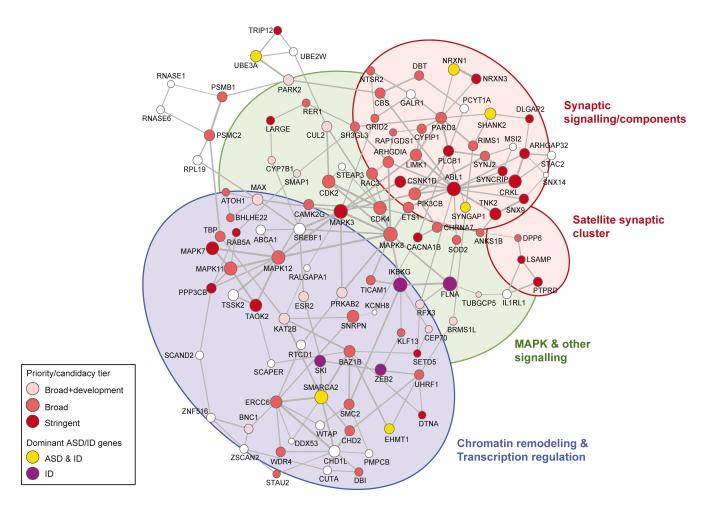


Figure S3. Candidacy tier for de novo CNV genes selected by NETBAG

Genes implicated in dominant forms of ASD and ID or in ID are highlighted in yellow and purple, respectively. Genes implicated in autosomal recessive ID (*ERCC6*, *CBS*, *DBT*, *PLCB1* and *LARGE*) were not given a specific color for ease of comprehension of the figure. Multiple lines of evidence from the literature and dedicated databases (DECIPHER, ISCA and ECARUCA) for the *CHD2* gene (recently involved in other neurodevelopmental disorders), and the candidate genes *SETD5*, *LSAMP*, *TRIP12*, *SYNCRIP*, *DTNA*, and *PIK3CB*, are given in the section 'Higlighted genes'.

NETBAG analysis of *de novo* CNV genes: For this analysis, recurrent events were counted once (or combined in one region) and events that did not intersect any genes were removed; the resulting final set used in the analyses consisted of 75 CNV regions intersecting 874 unique RefSeq genes. Derived gene clusters were scored using the de-weighted method, where the contribution of each individual gene to the overall cluster score is given by the weighted sum of its connections (edges) to the other cluster genes, and significance was obtained by generating random events with the same gene count as observed in the *de novo* CNV dataset with 1,000 randomizations (main **Figure 4B**; **Tables S14A-S14E**). The highest scoring cluster obtained using a searching procedure of up to two genes per CNV (global p-value 0.02) is shown. The list of up to 2 genes selected per CNV is given in **Table S14A**. The majority of the genes identified by NETBAG (92.0%, 104/113) are highly expressed in brain. When using a search procedure of up to one gene per CNV, a list of 69 genes was obtained (global p-value <0.01); this list is given in **Table S14A**. Analysis of established annotation resources, such as PubMed, OMIM, GeneReviews, EntrezGene and iHOP, suggested that a significant fraction of genes in the identified network either play a well-defined functional role in the brain or have been previously implicated in neurological and psychiatric disorders. An overview of functional information about each of the genes forming the cluster can be found in **Table S14B**.

Prenatal, postnatal or unbiased brain expression of genes selected by NETBAG (additional information relevant to main Figure 4B, based on NETBAG data). Nodes were colored according to whether a gene's brain expression is prenatally or postnatally biased, or has no biased expression in an analysis of 12 developmental stages of the BrainSpan RNA-seq dataset. Genes were considered biased if their mean expression in prenatal developmental stages was increased more than two-fold compared to their post-natal expression, or vice versa. Genes with related function according to Gene Ontology (GO)<sup>2</sup> and KEGG pathways<sup>3</sup> were indicated by shaded areas; a complete listing of over-represented GO terms and pathways is given in **Table S14D**. By further examining these genes (**Table S14C**), we found that 77% (87/113) of them have a strong link with neuronal and brain-specific genes. We explored whether affected genes were biased to certain developmental stages by interrogating the temporal expression of genes selected by NETBAG. We found no overall significant

bias to prenatally expressed genes, but we did observe prenatal bias for a cluster of genes participating in chromatin remodeling and transcription regulation (main **Figure 5B**, **Table S14E**). The prenatally-biased expression profile was characterized by high expression during development and sharp decrease after birth. We further compared expression levels (**Table S14B**) and found that most of the genes are highly expressed in the neocortex. Similar results were obtained when looking at LoF *de novo* single SNVs in 122 genes assembled from four ASD exome-sequencing studies.<sup>4-7</sup>

#### Candidacy tier of ASD candidate genes

To identify potential new ASD targets and gain mechanistic insights, we expanded our analysis to several gene lists described below. These gene lists were used to build a candidacy tier, by identifying which genes are more likely to be implicated in ASD based on membership in the gene lists. These gene lists were used for the analyses presented in main **Figure 3**, **Figure S3** and **Table S14C**. Our analyses focused on exonic events overlapping or disrupting exons, and deletions and duplications were analysed separately. P-values associated with odds ratios were calculated using Fisher exact tests.

- 1) Fragile X mental retardation protein (FMRP) targets: the gene encoding FMRP, FMR1, responsible for fragile X syndrome, is mutated in ~2% of ASD cases. Significant overlap between FMRP-targets and ASD candidate genes was reported recently, and lossifov et al. had previously shown that rare de novo LoF SNVs identified in their exome sequencing study were enriched in this class of genes. We used a set of FMRP-RNA interactors identified experimentally (n=842), using crosslinking and immunoprecipitation (CLIP) experiments in mouse. We note that a later paper detected FMPR targets computationally, using human RNA sequence motif analysis (n=939 top genes as provided by the authors), but given the limited overlap between the genes in the two papers (~20%), we decided to use for our enrichment analyses only the list of targets that were experimentally identified by Darnell et al. .
- 2) Post-synaptic density genes: The full list of experimentally identified human post-synaptic density genes was collected from Bayes et al. (n=1,453).<sup>11</sup>
- **3)** Haploinsufficiency index: We used the predicted haploinsufficiency index (HI) by Huang et al. <sup>12</sup> In particular, we used the imputed predictions as these have better gene coverage, and showed a similar performance compared to the original list of predicted genes pre-imputation when looking at dominant and recessive ASD and ID genes (data not shown). We defined four gene-sets: all genes with any value of HI (17,081 genes), the top 55% HI genes (pHI ≥0.15, 8,862 genes), top 26% (pHI ≥0.35, 4,136 genes), and top 14% (pHI ≥0.55, 2,214 genes). We focused our analysis on deletions because deletions, not duplications, are associated with HI, and show that HI is an important contributor to ASD etiology (duplication breakpoints can also disrupt genes or their regulatory elements and lead to haploinsufficiency).
- **4)** *Neurodevelopmental/neuropsychiatric phenotypes*: Genes associated with neurodevelopmental/neuropsychiatric phenotypes were mined from HPO (Human Phenotype Ontology)<sup>13</sup> for human and MGI/MPO for mouse (Mouse Genome Informatics/Mammalian Phenotype Ontology, The Jackson Laboratory, Bar Harbor, Maine, <a href="http://www.informatics.jax.org">http://www.informatics.jax.org</a>; data retrieved on November, 2012).<sup>14</sup>
  - a) For human, we selected all genes annotated for "Behavioural/Psychiatric abnormality" (HP:0000708) and/or "Cognitive impairment" (HP:0100543), as well as children terms in the ontology. We only considered genes with HPO phenotype annotation derived from OMIM, as for these we could more reliably infer the mode of inheritance based on the first digit of the OMIM ID (1 = autosomal dominant, 2 = autosomal recessive, 3 = X-linked).
  - b) For mouse, we selected all genes annotated for "nervous system phenotype" (MP:0003631) and "behavior/neurological phenotype" (MP:0005386), as well as children terms in the ontology; eQTL and complex phenotypes were removed. Autosomal dominant mode of inheritance was inferred by parsing MGI mutation allele tables and retaining only gene-phenotype annotations supported by a heterozygous mutated allele; genes on the human X chromosome were removed to obtain the autosomal dominant subset.
- 5) Neurodevelopmental function: Two neurodevelopmental lists were generated to help evaluate the candidacy of the genes selected by NETBAG: "NeuroF" was purely based on the manual curation of Gene Ontology terms and pathways, and "NeuroF\_EM" was based on gene-sets found associated by the logistic regression test in the enrichment map (EM) (results shown in main Figure 4A) and clustered as: (a) Cell projection/Neural development/Axonogenesis, or (b) Neuronal synapse. The Gene Ontology terms and pathways used to generate NeuroF are listed in Table S13A.
- **6) Developmental genes:** Genes were collected from all gene-sets found associated by the logistic regression test and clustered as Development/Cell Proliferation/Cell Motility in the enrichment map ("Dev\_EM").
- 7) Brain-expressed genes: Three gene lists were constructed based on the human BrainSpan RNA-seq dataset:
  - a) ExprBspan\_BrainAny\_log2rpkm3.0: genes with RPKM ≥3.0 in at least five BrainSpan data-points (a data-point corresponds to any unique combination of donor subject x brain region x developmental time-point).
  - b) ExprBspan BrainAny log2rpkm4.5: genes with RPKM ≥4.5 in at least five BrainSpan data-points.
  - c) ExprBspan\_NotBrainExprF: genes RPKM ≥1.0 not satisfying the above definition, yet annotated for at least one Gene Ontology term or pathway; these are similar in number to ExprBspan\_BrainAny\_log2rpkm4.5 and therefore were used as a negative control list, representing functionally characterized genes not expressed or expressed at very low levels in brain.

Brain-expressed gene lists were also defined based on the Novartis expression atlas (HG-U133A Affymetrix arrays), and used only for the definition of candidate gene lists:

- a) Novartis Brain-expressed genes: for a gene to be considered brain expressed it needed to have the robust multi-array average (RMA) gene expression values in whole-brain or fetal brain: (i) greater than the array median expression value (considering all tissues), and (ii) greater than the median expression in non-nervous tissues.
- b) Novartis Brain-specific genes: for a gene to be considered brain-specific it needed to have the RMA gene expression values in whole brain or fetal brain: (i) greater than the array median expression value (considering all tissues), and (ii) greater than the double of the median expression in non-nervous tissues.

Three lists were compiled based on the results from the 1-gene hit odds ratio analysis of single gene-sets and their combinations (main Figures 3A-B):

- 1) Stringent (1,088 genes): clearly linked to synaptic components and/or autosomal dominant neurodevelopmental/neuropsychiatric phenotypes.
- 2) Broad (5,342 genes): linked to synaptic components, or neurodevelopmental/neuropsychiatric phenotypes, or brain expression.
- 3) Broad development (7,158 genes): extended by gene-sets corresponding to broader developmental functions found by the logistic regression gene-set test and enrichment map clusters.

In particular, the following definitions were used:

- 1) Stringent, union of:
  - a) FMRP targets from Darnell et al.<sup>9</sup>
  - b) Genes found in at least two of these lists:
    - i) NeuroPheno Dom
    - ii) Intersection of: (a) Novartis Brain-expressed and (b) HI ≥ 0.55
    - iii) Postsynaptic density genes, full dataset from Bayes et al. 11
    - iv) FMRP targets from Ascano et al. 10
- 2) Broad, union of:
  - a) Stringent candidates (as defined above, in Stringent)
  - b) Novartis Brain-specific
  - c) Intersection of: (i) ExprBspan\_BrainAny\_log2rpkm4.5 and (ii) HI ≥ 0.35
  - d) Intersection of: (i) NeuroF and (ii) HI ≥ 0.35
  - e) Intersection of: (i) ExprBspan\_BrainAny\_log2rpkm3.0 and (ii) NeuroPheno All
- 3) Broad development, union of:
  - a) Broad candidates (as defined above, in Broad)
  - b) Intersection of: (i) NeuroF\_EM and (ii) NeuroPheno All
  - c) Development cluster in the enrichement map (Figure 4A, main text)

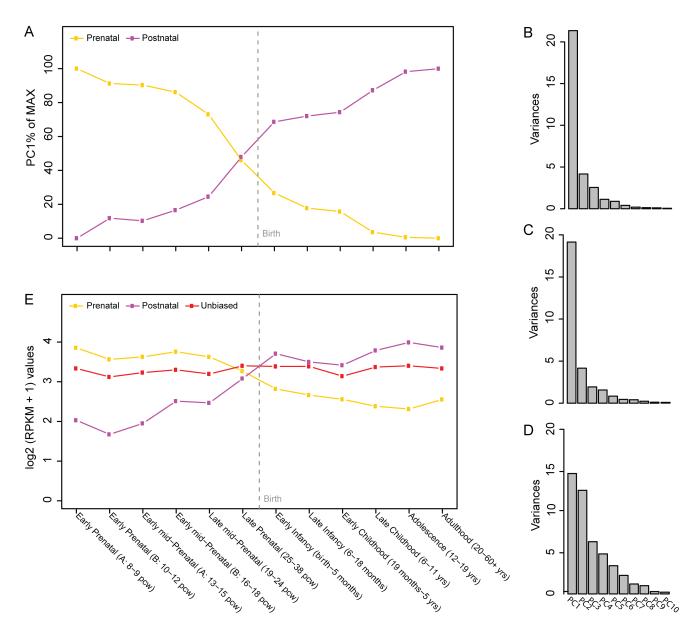


Figure S4. Expression profile for 113 *de novo* CNV genes selected by NETBAG in the neocortex across 12 developmental stages

The time course of BrainSpan RNA-seq gene expression data was quantile normalized and log2 transformed. (A) The first principle components (PC1, as percentage of the maximum, %/MAX) for the 113 CNV genes are shown in 2 expression groups (prenatally biased, postnatally biased). The amount of variance explained by each principle component is shown for (B) Prentatally-biased genes, (C) Postnatally-biased genes, and (D) Unbiased genes. (E) The group centroids (median) of the 3 expression groups (Prenatally biased, Postnatally biased, Unbiased) are plotted at each developmental stage with the Y axis showing log2 transformed expression profiles log2(RPKM + 1). PCW, post-conceptional weeks. Panels B-D compared the variance explained by PC1 for the 3 expression groups. PC1 for Unbiased genes is not representative, i.e., it does not explain more variance than other PCs and it is unable to profile the major direction of the expression group. Therefore we did not include the unbiased group in panel A.

RNA-Seq data preprocessing and normalization: The human brain developmental transcriptome data, consisting of a total of 12 developmental stages (from 8-9 post-conceptional weeks [PCW] to 20-60 years) in 524 samples, was downloaded from BrainSpan resource (http://brainspan.org/; Allen Institute for Brain Science, 2012; analyses presented here used the October 2013 version). In this dataset, gene expression profiles measured by RNA-sequencing experiments were represented as reads per kilobase of transcript per million mapped reads (RPKM), which were pre-summarized to gene levels based on a composite model defining a gene as the union of all exonic nucleotides across all transcripts. A sample level filtering procedure was implemented after comparison of the distribution of each sample to the entire distribution (all 524 samples); the top 2% outliers were removed based on the sum of chisquares with visual assistance on boxplot. Subsequently, a gene level filter was applied so that only genes with a RPKM value > 0 in

more than 80% of the samples in any brain region of any developmental stage were kept. To ensure quality of the data, the samples were further filtered by RNA integrity number (RIN) > 8. As a result, 42,965 gene entries and 416 samples passed QC and entered the downstream analysis where quantile normalization and log2 transformation procedures were performed.

Gene expression level classification: We then sought to classify genes by their expression levels across multiple brain regions and developmental stages. The genes with lower 20% expression level in all 8 brain regions (neocortex, amygdala, hippocampus, striatum, diencephalon, upper rhombic lip, cerebellar cortex, and dorsolateral prefrontal cortex) were classified as "non-expressed" genes. The remaining 80% genes were classified into 3 expressional categories specifically for two brain regions, the neocortex (NCX) and dorsolateral prefrontal cortex (DFC): 1) lowly expressed (bottom 25% averaging across NCX or DFC samples); 2) highly expressed (top 25% averaging across NCX or DFC samples); and 3) the remainder of genes were considered moderately expressed for NCX or DFC regions.

Identification of differentially expressed genes: We analysed the 113 genes resulting from the NETBAG functional network analysis of 102 *de novo* CNVs, and determined their prenatal-biased or postnatal-biased status by a nonparametric algorithm, similar to the method used in Xu et al. Specifically, for each of the 8 brain regions, we used the median of samples to represent the expression level of a certain given gene under a specific developmental stage. Subsequently, two groups were defined: prenatal (periods 2A, 2B, 3A, 3B, 4, 5) and postnatal (periods 6 through 11). Wilcoxon rank sum test with a p value cut-off of 0.05 and group median fold change cut-off of 1 were applied to determine differentially expressed genes between prenatal and postnatal groups. Genes with significantly higher expression in prenatal or postnatal group were considered prenatally or postnatally biased genes, while the remaining genes were classified as unbiased. Interestingly, we observed that some genes (e.g. KCNH8, CRKL) presented a peculiar profile: a "U shape", i.e., high expression in the first developmental stages followed by a decrease in expression and later, towards the last developmental stages, a new increase in expression. If there was a predominance of high expression in the left or in the right branch of the "U" curve, the genes were classified as prenatally-biased or postnatally-biased, respectively. The inverse could be observed for genes with a profile resembling an inverted U (e.g. RAC3).

**Profile of gene expression across 12 developmental stages:** Expression profile across 12 developmental stages of the 113 genes resulting from the NETBAG functional network categorized into 3 groups (prenatal, postnatal and unbiased) was plotted for 8 brain regions after Principle Component Analysis was performed with a custom R script. The first principle component of each expressional group was plotted in **Figure S4** for the neocortex region. Of note, the chromatin/transcription module in **Figures 4B** and **S4** showed a predominance of genes with a prenatally-biased expression profile, suggesting that besides the function of the altered gene, the timing of the effect of the genetic perturbation may also be of critical importance in determining disease risk.

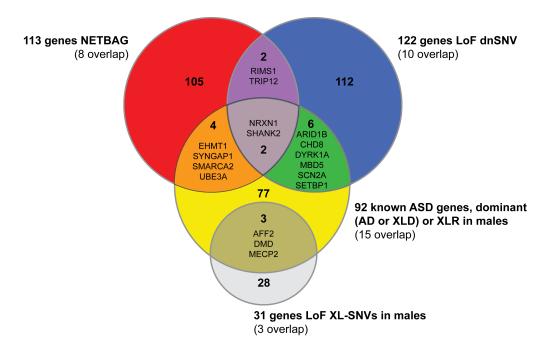


Figure S5. Overlap among gene lists pre-DAPPLE analysis

For the protein-protein interaction (PPI) network analyses presented in **Figures 5A and S5**, we used the Disease Association Protein-Protein Link Evaluator (DAPPLE)<sup>19</sup> under the assumption that causal genetic variants affect a limited number of mechanisms that are detectable by PPIs. DAPPLE takes a list of genes or loci as input and searches for significant physical connectivity among proteins encoded by those genes based on protein-protein interactions available in the InWeb database.<sup>20</sup> DAPPLE will then assess the statistical significance of several network connectivity parameters as well as of the connectivity of individual proteins to other seed proteins using a within-degree node-label permutation method. The Venn diagram shows the overlap between four lists used as input in DAPPLE (i.e. pre-DAPPLE):

- 1) 113 genes affected by *de novo* CNVs in AGP probands selected by NETBAG.
- 2) 122 genes with *de novo* LoF SNVs from four published whole exome sequencing studies comprising 965 ASD cases.<sup>4-7</sup> LoF variants included nonsense, frameshift, and splice site-disrupting mutations. One of the 122 genes is X-linked (*RPS6KA3*), and the remainder are autosomal. Three genes were not present in the InWeb database and were thus not included in the analyses (*DHRS4L1*, *PCDHA13*, and *RAD21L1*).
- 3) 92 genes known to be involved in ASD (list updated from Betancur, 2011)<sup>21</sup> (**Table S6A**); only autosomal dominant (AD) or X-linked (XL) genes were included (XL dominant, XLD, and XL recessive, XLR), since heterozygous CNVs affecting genes involved in autosomal recessive disorders are not deleterious *per se*.
- 4) 31 X-linked genes with hemizygous LoF SNVs in male ASD cases and not observed in male controls. 22

Of the 336 genes analysed (339 unique genes pre-DAPPLE minus 3 not present in the Inweb database), 17 genes overlap between two or more lists. Specifically, the *de novo* CNV genes selected by NETBAG overlap with the list of genes with *de novo* LoF SNVs and with the list of genes known to be involved in ASD; two of the genes (*NRXN1*, *SHANK2*) were shared by the three groups. There is no overlap between the 113 genes selected by NETBAG and the X-linked genes with LoF SNVs. Likewise, the Venn diagram post-DAPPLE depicted in main **Figure 5A** shows the overlap among the 151 genes selected by DAPPLE. It resembles the pre-DAPPLE Venn diagram since the relationship between the four gene lists was maintained, including the sharing of two genes (*NRXN1*, *SHANK2*) by the three main lists of genes. Over-representation analysis identified convergent functional themes related to neuronal development and axon guidance, signalling pathways, and chromatin/transcription regulation. Moreover, 90.7% (137/151) of the gene products reside within the same interconnected cluster.

We then further analysed the 113 genes selected by NETBAG and the DAPPLE output network of 151 proteins in terms of brain expression during development and in terms of functional groups (**Table S14E** and **S15**, respectively). Regarding the 113 gene list, we observed that the functional group composed of chromatin remodelers and transcription factors is significantly enriched for prenatal-biased genes and that the functional group composed of signalling genes is enriched for postnatal-biased genes. Similarly, for the DAPPLE output network, the chromatin remodelers/transcription factors functional group is significantly enriched for prenatal-biased genes, with a smaller p-value than the one estimated for the same functional group on the 113 genes list. Interestingly, in this case the functional group composed of neuronal genes is significantly enriched for postnatal-biased genes.

**Phylop and GERP scores:** Basewise conservation scores (PhyloP) across 45 vertebrate genomes (including the human genome) were downloaded from the UCSC website (http://genome.ucsc.edu/)<sup>23</sup> and processed with reference to the UCSC gene track (known genes). Using only the exome, the average base score was calculated. When different variants were present in a gene, all isoforms were aggregated and the average score was used. Likewise, the Genomic Evolutionary Rate Profiling (GERP++) score was downloaded from

Sidow's Lab website (http://mendel.stanford.edu/SidowLab/downloads/gerp/index.html). GERP score was calculated by taking the average basewise score of a gene. Both Phylop and GERP++ scores of genes were ranked. The ranking percentage of the scores (e.g. X% meaning X% of genes in the entire genome are ranked behind the given gene in terms of GERP or Phylop scores) are calculated to characterize each of the genes. Genes in the DAPPLE network (main **Figure 5B**) are among the top 75% more conserved genes compared to the genome average (based on GERP scores and PhyloP), and are typically long (median 103 kb vs. 10 kb for the genes in whole genome). Functionally significant and highly conserved genes tend to be more central in physical protein-protein and regulatory networks.<sup>24</sup>

## **HIGHLIGHTED GENES**

## Genes involved recently in other disorders

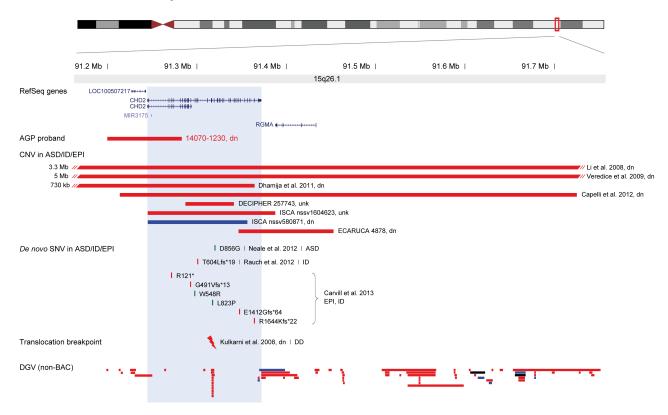


Figure S6. CNV and SNV identified in *CHD2* (chromodomain helicase DNA binding protein 2) in chromosome 15q26.1

Chr15:91,165,000-91,750,000 (hg18). Abbreviations: ASD, autism spectrum disorder; dn, *de novo*; DD, developmental delay; EPI, epilepsy; ID, intellectual disability; unk, unknown inheritance.

#### 1) AGP

- Proband 14070-1230, male, *CHD2* exonic deletion (chr15:91200007-91283004), *de novo*. The deletion also involves *LOC100507217* and *MIR3175*. The affected younger brother also carries the deletion, absent from both parents. SNP genotyping indicated that the deletion occurred in the paternal chromosome, suggesting paternal germline mosaicism. Phenotype: index case, 14 yo, autism (meets criteria both on ADI-R and ADOS), walked at 14 mo, first words 24 mo, phrases 26 mo, currently verbal, ID (Griffiths: language DQ 68, performance DQ 46, global DQ 53); normal growth, normal neurological exam; minor dysmorphic features: micrognatia, protruding ears; no epilepsy; brain MRI: altered angular gyrus (normal variant, unknown pathological significance). Brother, 12 yo, mild autism on ADOS, walked at 15 mo, first words 24 mo, phrases 36 mo, verbal, ID (Griffiths: language DQ 65, performance DQ 61, global DQ 51); normal growth, normal neurological exam; minor dysmorphic features similar to his brother (micrognatia, protruding ears); epilepsy, onset 9 y, partial with secondary generalization, difficult to control with carbamazepine; EEG showed paroxistic activity in the left temporal lobe.
- No CHD2 deletions among 4,768 controls and 4,875 parents.

#### 2) Other human genetic evidence

#### CNV:

• Li et al.<sup>25</sup>: Male with t(15;22)(q26.1;q11.2) translocation and a 3.3 Mb deletion encompassing *CHD2* (chr15:89197342-92489641) at the 15q breakpoint, *de novo*. Phenotype: developmental delay, walked at 20 mo, language delay, developmental motor coordination disorder, height and head circumference consistently ≤3<sup>rd</sup> centile, weight ≤5<sup>th</sup> centile, delayed bone age (6-mo-old bone age at 18 mo of chronological age, and 18-mo-old bone age at 36 mo chronological age), mild dysmorphic features (anteverted nares, unilateral auricular pit, fetal fingertip pads, low posterior hairline, and back hirsutism), left eye amblyopia corrected surgically at age 2. He had two episodes of febrile seizures. *IGF1R* was translocated to chromosome 22 and showed 50%

reduction of expression, which could be responsible for the growth deficiency. Angelman syndrome was ruled out by methylation analysis.

- Veredice et al. <sup>26</sup>: Female, 5 Mb deletion encompassing *CHD2* (chr15:87796000-92700000), *de novo*. Phenotype: severe intractable myoclonic epilepsy with photosensitivity, with onset at 6 mo, ID, growth delay, peculiar facial features and minor physical anomalies. Born at 37 weeks, weight 3<sup>rd</sup> centile, length 10<sup>th</sup> centile, head circumference 2<sup>nd</sup> centile. At 21 mo her weight was 3<sup>rd</sup> centile, with length and head circumference <<3<sup>rd</sup> centile. She had congenital hypothyroidism, bicuspid aortic valve, diffuse hypotonia, ligamentous laxity, and dysmorphic features (upward slanting eyes, epicanthal fold, depressed nasal bridge, full cheeks, prominent lips, downturned corners of the mouth, protruding tongue, large ears with anteverted lobe, single palmar crease, increased 1<sup>st</sup>-2<sup>nd</sup> interdigital space and redundant nuchal skin). Her language was limited to a few words and she had mild developmental delay (Griffith's developmental quotient 67). Brain MRI showed cerebellar vermis hypoplasia with mega-cisterna magna.
- Dhamija et al.<sup>27</sup>: Female, deletion encompassing *CHD2*, with a minimum size of 731 kb (chr15:90633409-91364628) and a maximum size of 936 kb (chr15:90530351-91466733), *de novo*. Phenotype: 9 yo girl with developmental delay, ASD, growth delay, and intractable generalized epilepsy, with onset at 3.5 y. Initially the seizures were partial complex but later became generalized; most prominent were absence seizures, occurring many times a day. She had delayed motor development (walked at 24 mo) and delayed language development with echolalia and mild generalized cognitive delay upon formal testing. On examination, she had mild dysmorphic features (widely set eyes, bilateral pits on the helix of the ears, and crowded teeth with prominent incisors), short stature and head circumference <3<sup>rd</sup> centile. Brain MRI was normal.
- Capelli et al.<sup>28</sup>: Female, 511 kb deletion encompassing 4 genes, *CHD2*, *LOC100507217*, *MIR3175* and *RGMA* (chr15:91213864-91724860), *de novo*. *RGMA* (repulsive guidance molecule, member A) exerts a negative control on axon growth and could also potentially contribute to the phenotype. Phenotype: when examined at the age of 6 y, she presented global developmental delay, epilepsy, autistic behavior, severe speech impairment with minimal use of words, short attention span, facial dysmorphisms (prognathia, wide mouth, short and widely spaced teeth, strabismus), gait ataxia with uplifted arms and hand flapping, slight hypotonia and hyperactive deep tendon reflexes. Weight, height and head circumference were at the 50<sup>th</sup>, 75<sup>th</sup> and 10<sup>th</sup> centiles respectively. Angelman syndrome was suspected but the methylation pattern at *SNRPN* was normal. Seizures started at the age of 24 mo and were partially controlled with valproic acid. (Corresponds to subject 249888 in DECIPHER.)
- DECIPHER<sup>29</sup>: Subject 257743, male, CHD2 intragenic deletion (chr15:91287269-91341234), inheritance unknown. Phenotype: autism, macrocephaly, proportionate short stature, scoliosis. This individual also has a 16p11.2 deletion syndrome, inheritance unknown.
- ISCA<sup>30</sup>: Subject nssv1604623, unknown gender, deletion encompassing 3 genes, *CHD2*, *MIR3175* and *RGMA* (chr15:91245098-91387818), inheritance unknown. Phenotype: seizures.
- ISCA: Subject nssv580871, unknown gender, intragenic *CHD2* duplication (chr15:91245028-91356680), *de novo*. Phenotype: developmental delay and additional significant developmental and morphological phenotypes referred for genetic testing.
- ECARUCA<sup>31</sup>: Subject 4878, male, deletion encompassing only 2 genes, *CHD2* and *RGMA* (chr15:91346585-91452586), *de novo*. Phenotype: ID, truncal obesity, large ear lobules, bulbous nasal tip, low posterior/trident hairline, cryptorchid testes, small hands and feet, tapering fingers, hypotonia, seizures, abnormal EEG.

## **Translocation breakpoint:**

• Kulkarni et al.<sup>32</sup>: Female proband with a balanced *de novo* translocation t(X;15)(p22.2;q26.1); the 15q26.1 breakpoint disrupts *CHD2*, whereas the Xp22.2 breakpoint lies in a gene-free region. Phenotype: developmental delay, learning problems, scoliosis, hirsutism, high arched palate, normal face (including eyes and ears), syndactyly of the toes, height <30<sup>th</sup> centile and head circumference <25<sup>th</sup> centile. No seizures reported at age 17 y.

#### SNV

- Carvill et al.<sup>33</sup> (targeted sequencing in epileptic encephalopathies): 6 individuals (4 males and 2 females) with *de novo* mutations in *CHD2* (3 frameshift, 1 stop, 2 missense). Phenotype: 2 had myoclonic atonic epilepsy, 3 had epileptic encephalopathy not otherwise specified and 1 had Lennox-Gastaut syndrome. Seizure onset occurred between 1-3 y; myoclonic seizures were present in all six, photosensitivity in three. All had ID (2 moderate, 4 severe), 1 had ASD.
- Neale et al.<sup>5</sup> (exome sequencing in ASD): Subject 10C100480, male, *CHD2* missense variant, *de novo*. The variant is predicted to be either benign by PolyPhen2<sup>34</sup> and PANTHER,<sup>35</sup> or damaging by SIFT,<sup>36</sup> SNAP,<sup>37</sup> and Mutation Taster;<sup>38</sup> the affected residue is highly conserved: GERP 5.48, <sup>39</sup> ConSurf 6/9. <sup>40</sup> Phenotype: ASD (no other information provided).
- Rauch et al. (exome sequencing in nonsyndromic sporadic ID): Subject MS134, female, *CHD2* frameshift mutation, *de novo*. Phenotype: severe nonsyndromic sporadic ID (IQ 50-69), absence seizures (onset at 5 y), Duane anomaly, no ASD.

## 3) Functional evidence

CHD2 encodes a member of the chromodomain helicase DNA-binding (CHD) family of proteins, characterized by the presence of
chromodomains (chromatin organization modifier) and SNF2-related helicase/ATPase domains. CHD proteins alter gene
expression by modification of chromatin structure, playing critical roles during development.

- · Expressed in the brain.
- Chd2 knockout in mice results in embryonic lethality; heterozygous mice have variable postnatal lethality and growth retardation, with reduced body fat, pronounced lordokyphosis, renal disease and anemia in adults.<sup>42,43</sup>

#### 4) Other evidence

- % haploinsufficiency (HI) = 15.8 (likely to be haploinsufficient). 12
- Mutations in CHD7 cause CHARGE (coloboma, heart defects, atresia of the choanae, retardation of growth and developmental, genital and/or urinary defects and ear abnormalities) syndrome, associated with syndromic ID and sometimes ASD. 44,45
- Nine de novo SNV in CHD8 reported in subjects with ASD (3 nonsense, 4 frameshift, 1 splice, and 1 single amino acid deletion). 6,46

#### 5) Comment

CHD2 mutations were implicated in epileptic encephalopathies (Carvill et al. 2013) while this manuscript was in preparation. The deletion in the two siblings with ASD identified in our study, together with the review of other cases, also implicates this gene in ASD. CHD2 haploinsufficiency is highly penetrant (all cases with known inheritance are *de novo*), associated with ID and variably with epilepsy and ASD. Although the phenotype description is missing in several cases and is incomplete in others, precluding complete analyses, among 18 cases (the missense variant reported by Neale et al. not included), ID is reported in 16, epilepsy in 13 and autism/ASD in 6 cases. Language delay and/or limited speech are described in 5. Eight individuals are reported with dysmorphic features, usually mild. Features reported in 2 or more individuals include: strabismus or amblyopia (n=3), ear pits (n=2), hirsutism (n=2), and low posterior hairline (n=2). At least 4 had growth delay/short stature, and 2 have scoliosis, two phenotypes reported in heterozygous *Chd2* mice.

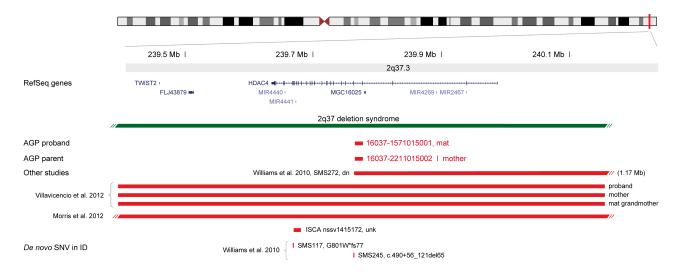


Figure S7. CNV and SNV identified in HDAC4 (histone deacetylase 4) in chromosome 2g37.3

Chr2:239,350,000-240,180,000 (hg18). Abbreviations: dn, de novo; ID, intellectual disability; mat, maternal; unk, unknown inheritance

#### 1) AGP

- Proband 16037-1571015001, male, 12 kb intragenic deletion of HDAC4, removing exon 4 (chr2:239766528-239778481), maternal. Phenotype: autism (ADI-R and ADOS positive), language delay (first words and phrases 42 mo), functional language, WISC-III at 21 y: verbal IQ 72, performance IQ 87, full scale IQ 77. Born at 37 weeks of gestation, no congenital malformations, normal general exam. Had seizures at age 4-5 y, treated with Depakote; treatment stopped for 2-3 y without seizure activity. Unfortunately, we were unable to have the proband re-evaluated by a clinical geneticist to assess the presence of mild dysmorphic features and hand or feet abnormalities. The father has schizophrenia; the mother has depression, no other information is available about her phenotype.
- No HDAC4 deletions among 4,768 controls and 4,874 parents (1 deletion in the mother of proband 16037-1571015001).

#### 2) Other human genetic evidence

Over 120 individuals with 2q37 terminal deletions (2q37 deletion syndrome, also known as brachydactyly mental retardation syndrome) have been reported. Common features include developmental delay/ID, ASD, hypotonia, mild facial dysmorphism (frontal bossing, round face, depressed nasal bridge, abnormal or prominent ears, deep-set eyes, anteverted nares, and thin upper lip), short stature, obesity and brachymetaphalangy of digits 3-5 (>50%).

to result in brachydactyly mental retardation syndrome. Williams et al. 48 described 5 individuals with 2q37 deletion syndrome in whom the smallest region of overlap contained only *HDAC4*. Sequencing of *HDAC4* identified *de novo* loss-of-function mutations in two additional subjects. Although *HDAC4* mutations may be causative for most of the features of the 2q37 microdeletion syndrome, other genes might also be involved, since individuals with distal deletions not including *HDAC4* have been reported with ID, ASD and seizures. Because no *HDAC4* mutations or single gene deletions had been reported previously in ASD, this gene was not yet considered as a cause of ASD (i.e. we had included it in the list of ID genes, not in the ASD list).

• 2q37 deletions as well as the 2 reported *HDAC4* mutations occur *de novo*, so the maternal transmission observed in the AGP proband would appear a highly unusual finding. However, a recent case report<sup>49</sup> described for the first time a deletion overlapping *HDAC4* inherited from a mildly affected parent (see below).

#### CNV:

As noted above, there are many large 2q37 deletions reported in the literature and in the databases. Here we review only cases with small deletions overlapping *HDAC4* and two recently reported familial cases.

- Villavicencio-Lorini et al. 49: Report of the first three generation familial case of brachydactyly mental retardation syndrome with an interstitial 2q37.3 microdeletion of 758 kb (chr2:239395957-240154599). Subject 1: female index case, only child, motor development and growth delays during the first year of life, with subsequent catch up. When evaluated at 2 y 8 mo, she had midface hypoplasia, deep set eyes, posteriorly rotated and low-set ears, thin upper lip and pointed chin. Speech development was normal; she exhibited aggressive tantrums and sleep difficulties. Brachydactyly was excluded clinically and radiologically. Subject 2: 45-year-old mother of the index case, history of developmental and growth delays during childhood, but she was later able to attend normal school. She reported reduced spatial orientation and memory deficits. Examination revealed coarse facial appearance with broad and depressed nasal bridge, highly arched eyebrows, deep set eyes and narrow palpebral fissures; growth parameters were normal. Her hands and feet appeared normal, and brachydactyly was excluded radiologically. Subject 3: 68-year-old grandmother of the index case, she is the mother of subject 2, her only child. Family history revealed that her sister's daughter had ID with hydrocephalus and paraplegia of unknown cause. Subject 3 had severe osteoarthritis, dysmorphic facial features similar to those of her daughter with highly arched eyebrows, narrow palpebral fissures and everted full lips, with normal growth parameters. She communicated in simple sentences and her intellectual skills appeared to be lower than normal. Clinically her hands were normal.
- Morris et al. <sup>50</sup>: Report of a familial case of brachydactyly mental retardation syndrome, including a parent with mild symptoms and a child exhibiting a more severe phenotype. Cytogenetic testing showed a cryptic balanced translocation in the mother, t(2;10)(q37;q26), which resulted in a 9.84 Mb deletion on chromosome 2q37.1 (chr2:232810566-242654701) and a 10q26.1 duplication (chr10:131931089–135253240) in her son. *HDAC4* was deleted in the child but present and translocated in his mother. Interestingly, *HDAC4* expression in lymphocytes was 67% in the mother and 23% in the son compared to normal controls. Since the predicted expression after loss of one copy of *HDAC4* is ~50%, these findings suggest that there is an additional unknown mechanism decreasing *HDAC4* dosage, both in the mother and the child.
  - Subject 1: male proband, only child; upon examination at the age of 15 y he was overweight (BMI 28), with short stature (–3 SD), facial dysmorphism with round face, prominent forehead, highly arched eyebrows, sparse hair, low-set ears, downslanted palpebral fissures, depressed nasal bridge, thin upper lip, and high palate. He also presented type E brachydactyly of fourth fingers and toes, and syndactyly of the second and third toes. Brain MRI showed abnormal gyration of the frontal lobes. He was schooled in a specialized institute. Subject 2: mother of subject 1, she had one previous termination of pregnancy for occipital meningoencephalocele. Family history showed a maternal aunt and three maternal cousins with ID. She presented with a similar dysmorphic features as her son (round face, prominent forehead, highly arched eyebrows, low-set ears and thin upper lip), short stature, obesity (BMI 45), and brachydactyly of the fourth finger of the right hand, and third and fourth fingers on the left. She had no obvious ID.
- ISCA: Subject nssv1415172, unknown gender, HDAC4 intragenic deletion of 10 kb (chr2:239670063-239680639), inheritance
  unknown. Phenotype: developmental delay and additional significant developmental and morphological phenotypes referred for
  genetic testing.

#### 3) Functional evidence

- Histones play a critical role in transcriptional regulation, cell cycle progression, and development. Histone acetylation/deacetylation alters chromosome structure and affects transcription factor access to DNA. HDAC4 encodes a histone deacetylase that shuttles between the nucleus and cytoplasm in response to calcium-regulated signals. HDAC4 has been shown to regulate a transcriptional program essential for synaptic plasticity and memory; by repressing genes encoding synaptic proteins, it affects the strength and architecture of excitatory synapses.
   Conditional deletion of Hdac4 in mouse forebrain neurons leads to impairments in spatial learning and memory and long-term synaptic plasticity.
- Highly expressed in the brain.

#### • 4) Comment

Although many cases of 2q37 deletion syndrome with ASD have been reported in the literature, no deletion involving only *HDAC4* or mutation in this gene had been reported in individuals with ASD. The identification of an intragenic exonic deletion of the *HDAC4* gene in one AGP proband implicates haploinsufficiency of this gene as a cause for autism.

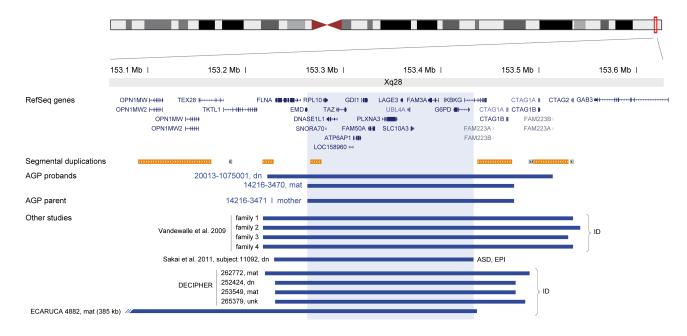


Figure S8. Recurrent duplications at Xq28 including GDI1 (GDP dissociation inhibitor 1)

ChrX:153,085,000-153,640,000 (hg18). Abbreviations: ASD, autism spectrum disorder; dn, *de novo*; EPI, epilepsy; ID, intellectual disability; mat, maternal; unk, unknown inheritance

#### 1) AGP

- Proband 20013-1075001, male, Xq28 duplication of 292 kb (chrX:153222048-153514311), de novo. The duplication encompasses 21 genes, including 3 genes involved in ID (FLNA, GDI1, IKBKG) and is flanked by segmental duplications. Phenotype: sporadic autism (ADI-R and ADOS positive), no language delay, normal IQ (WISC-III at 16 y: verbal IQ 93, performance IQ 94, full scale IQ 93), no dysmorphic features, head circumference -1.4 SD, normal neurological exam, no seizures.
- Proband 14216-3470, male, Xq28 duplication of 211 kb (chrX:153263157-153474401), maternally inherited. The duplication overlaps 19 genes, including 2 genes involved in ID (*GDI1*, *IKBKG*) and is flanked by segmental duplications. Phenotype: autism (ADI-R and ADOS positive), neurodevelopmental delay with onset at 2 y (first words 18 mo, first phrases 48 mo), mild ID (verbal IQ 61, performance IQ 65, full scale IQ 72), no dysmorphic features, sleep problems, no epilepsy. A brother with developmental delay also carries the duplication. He has a confirmed learning disorder and probably mild intellectual disability (he is an adult now and was not evaluated formally for this study).
- No similar duplications among 2,022 male controls and 2,441 fathers.

#### 2) Other human genetic evidence

• Mutations in *GDI1* are a rare cause of nonsyndromic X-linked ID<sup>53,54</sup> and have not yet been reported in ASD.

#### CNV:

- Vandewalle et al. <sup>55</sup>: Four unrelated families with X-linked ID with recurrent 0.3 Mb Xq28 copy number gain (153.20-153.54 Mb) mediated by segmental duplications. Only males are affected, carrier mothers show skewed X chromosome inactivation. The copy-number gain is variable, ranging from 2 to 5 copies, and includes *GDI1*, *FLNA* and *IKBKG*, involved in ID through mutations. The authors suggest *GDI1* is the most likely candidate gene; it is highly expressed in the brain and its expression in blood is correlated with the severity of the phenotype. *FLNA* duplications have been reported in four males with intestinal dysfunction, without ID <sup>56</sup> and in one male control in the Database of Genomic Variants (DGV); a duplication including *FLNA* and *IKBKG* was reported in a healthy male. <sup>55</sup>
  - <u>Family 1</u>: four affected males in two generations, Xq28 copy number gain (chrX:153218000-153535000). Phenotype: non-syndromic moderate ID. <u>Family 2</u>: two affected brothers, Xq28 copy number gain (chrX:153218000-153542000). Phenotype: severe ID, epilepsy in the elder brother. <u>Family 3</u>: three affected males, Xq28 copy number gain (chrX:153218000-153530000). Phenotype: the index case presented with delayed speech, learning disabilities, and mild ID. All affected male subjects had moderate ID, a peculiar face with dysmorphic features, and macrocephaly. Reported previously as case 6 in Madrigal et al. <u>Family 4</u>: male (sporadic case), Xq28 copy number gain (chrX:153218000-153535000). Phenotype: global psychomotor delay, mild dysmorphism.
- Sakai et al.<sup>58</sup>: Subject 11092, male, Xq28 duplication (chrX:153229170-153433332), de novo. Phenotype: ASD, history of seizures, normal IQ (verbal IQ 108, performance IQ 125, full scale IQ 109).

- DECIPHER: Subject 262772, male, Xq28 duplication (chrX:153219789-153490319), maternal. No phenotype information; the subject carries two other duplications (chr3:685234-1166340 and chr17:31862-164024), both inherited.
- DECIPHER: Subject 252424, male, Xq28 duplication (chrX:153230084-153475911), de novo. No phenotype information; the subject carries an additional CNV (chr1:91820025-92502808 duplication, de novo).
- DECIPHER: Subject 253549, male, Xq28 duplication (chrX:153230084-153475911), maternal. No phenotype information; only CNV reported in the subject.
- DECIPHER: Subject 265379, male, Xq28 duplication (chrX:153230109-153485889), unknown inheritance. Phenotype: ID, hypotonia, sleep apnea. Only CNV reported in the subject.
- ECARUCA: Subject 4882, male, Xq28 duplication (chrX:153051459-153436637), maternal. Phenotype: ID, spasticity/increased tendon reflexes, macrocephaly, constipation, short stature.
  - (Three ISCA individuals with duplications of this region —2 with developmental delay, 1 with ASD— were not added because their gender is unknown.)

#### 3) Functional evidence

- *GDI1* encodes the GDP-dissociation inhibitor alpha (αGDI), which regulates the activity of small GTPases of the Rab family involved in intracellular vesicular trafficking. *Gdi1*-deficient mice exhibit membrane accumulation of Rab GTPases and decrease in the reserve pool of synaptic vesicles in the hippocampus, leading to altered synaptic plasticity and short-term memory deficits.<sup>59</sup>
- GDI1 is highly expressed in the brain.

#### • 4) Comment

We identified two AGP probands with a recurrent Xq28 duplication corresponding to a genomic disorder recently described in ID. The identification of these two independent ASD cases, together with an ASD subject reported previously<sup>58</sup>, implicates duplications of this region in ASD. Among the genes contained in the region, aberrant gene dosage of *GDI1* is likely to be responsible for the neurodevelopmental phenotype of subjects carrying this CNV since mutations in this gene are involved in nonsyndromic X-linked ID<sup>53,54</sup> and the expression of *GDI1* in the blood of individuals with the Xq28 duplication correlated with the severity of the phenotype. Finally, *GDI1* is highly expressed in the brain and its function in intracellular vesicular trafficking together with the synaptic phenotype of the *Gdi1*-deficient mice make this gene particularly interesting with regard to the ASD/ID phenotypes.

## Candidate genes affected by de novo CNV in AGP probands

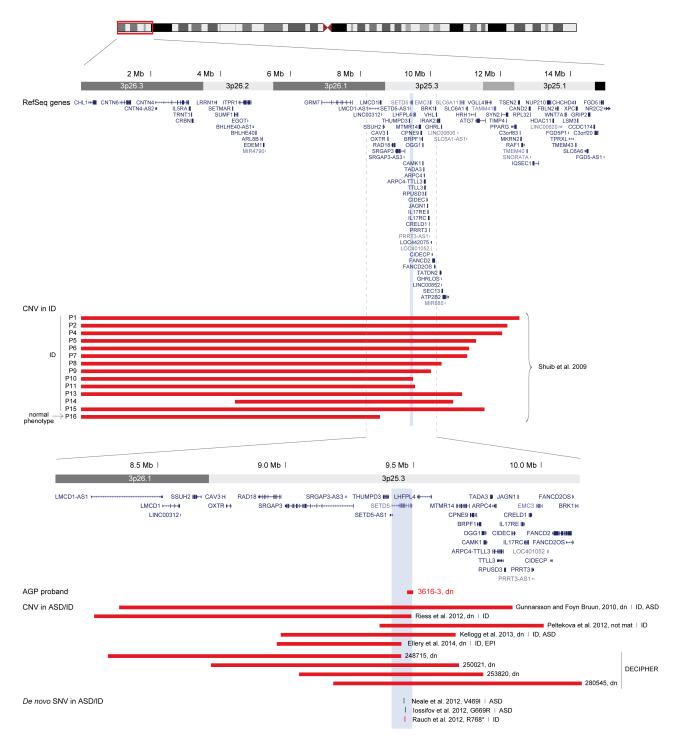


Figure S9. CNV and SNV identified in SETD5 (SET domain containing 5) in chromosome 3p25.3

Chr3:1-15,000,000 (hg18). Abbreviations: ASD, autism spectrum disorder; dn, *de novo*; EPI, epilepsy; ID, intellectual disability; mat, maternal

#### 1) AGP

 Proband 3616\_3, male, 24 kb deletion including only SETD5 (chr3:9474320-9498362), de novo. Phenotype: Born at term after normal pregnancy and delivery. No feeding or sleep problems; body length and head circumference during the first year of life were consistently between 0 to -1 SD; walked at 15 mo, language delay (first words 24 mo), functional language, autism (meets criteria for autism on both ADI-R and ADOS), borderline IQ (WISC-III: verbal IQ 64, performance IQ 82, full scale IQ 70). Concerns about ADHD-like behavior in childhood, but did not fulfill criteria for ADHD and there are no concerns about hyperactivity at present. Normal vision and hearing; had sensory-integration training because of over and undersensitivity; asthma and hay fever, no other medical issues, no seizures. At 16 y, normal height and weight (50<sup>th</sup> centile), no obvious dysmorphic features (no ptosis, normal ears, nose and mouth), normal fingers and toes.

• No SETD5 deletions among 4,768 controls and 4,875 parents.

#### 2) Other human genetic evidence

• 3p deletion syndrome is a rare contiguous gene disorder resulting from terminal and interstitial deletions involving chromosome 3p25-p26. Common features include ID, growth retardation, microcephaly, dysmorphic facial features, and ptosis. <sup>60</sup> Rare individuals with a 3p26-p25 deletion and normal intelligence or only mild abnormalities have been reported, including several cases inherited from seemingly unaffected parents, indicating variable penetrance. There are over 30 individuals with 3p25-p26 deletions reported, the majority with large cytogenetically visible rearrangements, making delineation of the critical region and identification of candidate genes difficult. Shuib et al. <sup>60</sup> reported 14 individuals with cytogenetically visible deletions of 3p25, all with marked ID except one subject (P16) with a normal phenotype, ascertained incidentally because of recurrent miscarriages. Notably, the deletions in subjects with ID all overlapped SETD5 whereas the subject without ID had the smallest terminal deletion, with a breakpoint proximal to SETD5 (Figure S9). Here we review recently described cases with clinical features of distal 3p deletion syndrome carrying smaller interstitial deletions.

#### CNV:

- Gunnarsson and Foyn Bruun<sup>61</sup>: Female, 1.6 Mb 3p26.1-p25.3 deletion of 24 genes including *SETD5* (chr3:8305426-9885334), *de novo*. The girl was born at 37 weeks, with birth measures below –2 SD. She was noted to have dysmorphic facial features (hypertelorism, ptosis, strabism, flat and broad nasal root, long philtrum, downturned corner of the mouth, low set ears), bilateral overlap of the 2<sup>nd</sup> and 4<sup>th</sup> toes over the 3<sup>rd</sup> and 5<sup>th</sup> toes, atrial and ventricular septal defects, hypotonia, and severe developmental delay, with absent language at the age of 4 y. She showed autistic behavior; she smiled and laughed, but had poor eye contact and did not interact with other children, focusing on objects in her close vicinity. Vision, hearing, and growth were normal. At the age of 4, she had transient seizures, documented with EEG; a MRI showed asymmetry of thalamus. (Corresponds to subject 253231 in DECIPHER.)
- Riess et al.<sup>62</sup>: Female, 1.24 Mb 3p26.1-p25.3 deletion of 12 genes including SETD5 (chr3:8250541-9491586), de novo. Normal birth measures, hypotonia during the first year of life, delayed psychomotor development. She started to walk and talk at the age of 2 y. On examination at the age of 3, she had large head (75<sup>th</sup> centile) compared to her height (10<sup>th</sup> centile), bilateral strabismus, large fontanelle, prominent forehead, depressed nasal bridge, thin upper lip and prominent philtrum. Ultrasound examinations of the heart and abdominal organs were normal.
- Peltekova et al.<sup>63</sup>: Female, 643 kb 3p25.3 deletion of 23 genes including *SETD5* (chr3:9367274-10010209), inheritance unknown (not maternal, father's DNA not available). The proband was born via cesarean section for breech presentation at 37 weeks gestation; birth weight 3<sup>rd</sup>–15<sup>th</sup> centile, length 5<sup>th</sup> centile, and head circumference 25<sup>th</sup> centile. She presented polydactyly in all four extremities, cleft palate, atrial septal defect, and bowel malrotation. At 1 year of age she developed tonic–clonic seizures and had episodes of aspiration pneumonia requiring placement of a gastrostomy tube. She was non-verbal as an adult, but smiled, laughed and was interactive. At 22 y her height and weight were <3<sup>rd</sup> centile, and head circumference was <5<sup>th</sup> centile. She had low anterior hairline, short forehead, thick eyebrows with synophrys, microphthalmia, downslanting palpebral fissures, blepharophimosis with mild ptosis, hypotelorism, esotropia, low set and cupped ears, and scoliosis. Hands and feet were small, with bilateral syndactyly of the 2<sup>nd</sup> and 3<sup>rd</sup> toes. Brain MRI showed parenchymal volume loss and atrophy of all structures except the brain stem.
- Kellogg et al.<sup>64</sup>: Female, 684 kb 3p25.3 deletion of 7 genes including *SETD5* (chr3:8980098-9664733), *de novo*. The proband was born at full term, with normal birth weight (50<sup>th</sup> centile) and length (90<sup>th</sup> centile). She had strabismus (corrected surgically) and developmental delay. Obsessive-compulsive disorder, with repetitively smelling of various objects, was noted at 10 y. On examination at age 11, she had dysmorphic features, including prominent ear lobes, right ear pit, depressed nasal bridge, anteverted nares, long philtrum, and proximally placed thumbs. Height, weight and head circumference were 50<sup>th</sup>, 10<sup>th</sup> and 50<sup>th</sup> centile, respectively; the brain MRI was normal. Assessment at 5 y 8 mo placed her in the borderline to intellectually disabled range (Stanford-Binet IV), and she met criteria for autism spectrum on ADOS.

The authors compared this individual with three previously reported cases with interstitial 3p25 deletions with ID and characteristic facial features<sup>61-63</sup> and identified a region of overlap including only three genes: *THUMPD3*, *SETD5* and *SETD5-AS1*, which could play a critical role in the neurocognitive phenotype of the 3p deletion syndrome.

• Ellery et al. 65: Male, 486 kb 3p25.3 deletion of 6 genes including SETD5 (chr3:8965201-9450984), de novo. The proband was born at term with a birth weight of 4600 g, after a pregnancy marked by gestational diabetes. Bilateral postaxial polydactyly, a single palmar crease, right preauricular pit, mild hypertelorism, anteverted nostrils and micrognathia were noted at birth. Hypotonia, feeding difficulties and developmental delay became evident afterwards. A diagnosis of Simpson–Golabi–Behmel syndrome was considered. He walked independently at 4 y but tired easily; at 6 y his vocabulary was limited to two-dozen words and occasional

short sentences. He developed grand mal seizures at 7 y, treated with carbamazepine. He had a disturbed sleep pattern with frequent waking. On examination at 8 y of age, he had epicanthic folds, mild hypertelorism, high palate, myopathic facies with coarse features, and pectus excavatum. He had poor muscle bulk and showed a partial Gower's sign. These features persisted into adulthood.

- DECIPHER: Subject 248715, female, 1.1 Mb 3p26.1-p25.3 deletion of 12 genes including SETD5 (chr3:8330935-9450984), de novo.
   Phenotype: ID/DD, macrocephaly, coarse facial features, synophrys, gum hypertrophy, low posterior/trident hairline, general abnormalities of hair texture. Only CNV reported in the subject.
- DECIPHER: Subject 250021, male, 946 kb 3p25.3 deletion of 10 genes including SETD5 (chr3:8724500-9671040), de novo. No phenotype information; only CNV reported in the subject.
- DECIPHER: Subject 253820, male, 700 kb 3p25.3 deletion of 9 genes including *SETD5* (chr3:9061621-9763580), *de novo*. Phenotype: ID, hypertelorism, micrognathia, low-set ears, hydrocephalus, horseshoe kidney. Only CNV reported in the subject.
- DECIPHER: Subject 280545, male, 1.1 Mb 3p25.3 deletion of 33 genes including *SETD5* (chr3:9186364-10290795), *de novo*. Phenotype: global developmental delay, stereotypic behavior, absent speech, abnormality of the corpus callosum, abnormality of the hypothenar eminence, small thenar eminence, facial asymmetry and blepharophimosis. Only CNV reported in the subject.
- Note that several SETD5 exonic deletions are reported in controls from DGV among non-BAC based studies, all from Shaik et al.;
   these deletions are likely to be a study-specific artifact.

#### SNV:

- Neale et al.<sup>5</sup> (exome sequencing in ASD): Subject 09C98906, male, SETD5 missense variant, de novo. The variant is predicted to be either benign (PolyPhen2, SNAP, Mutation Taster) or damaging (SIFT); the substituted residue is variable according to ConSurf (1/9); GERP score 1.91 (constrained >2). Phenotype: ASD, verbal IQ 93, performance IQ 112, full scale IQ 97.
- lossifov et al. (exome sequencing in ASD): Subject 13576, female, *SETD5* missense variant (gene listed as *KIAA1757*), *de novo*. The variant is predicted to be damaging (PolyPhen2, SNAP and Mutation Taster; not scored by SIFT). The substituted residue is variable according to ConSurf (1/9); GERP score 5.69. Phenotype: ASD (no other information provided).
- Rauch et al.<sup>41</sup> (exome sequencing in nonsyndromic sporadic ID): Subject ER14209, female, *SETD5* nonsense mutation, *de novo*. Phenotype: Born full term, weight –0.95 SD, head circumference –1.46 SD, sitting at 8 mo, walking at 20 mo, first words at 48 mo, IQ 70, mild attention deficit disorder, no ASD, no seizures, strabism, recurrent infections, constipation, prominent finger joints, facial dysmorphisms. When last evaluated at 9 y, height –0.78 SD, head circumference –1.16 SD, spoke in fluent sentences.

#### 3) Functional evidence

- The gene is predicted to be a methyltransferase; several other genes encoding methyltransferases have been shown to be altered
  in ASD/ID (see examples below).
- · Expressed in the brain.

#### 4) Other evidence

- % HI = 21.3 (0%-10%: likely to be haploinsufficient; 90%-100%: not likely to be haploinsufficient)
- Haploinsufficiency of NSD1 (nuclear receptor binding SET domain protein 1), encoding a histone methyltransferase, causes Sotos syndrome, a neurodevelopmental disorder characterized by overgrowth, distinctive craniofacial appearance, and variable ID, sometimes associated with ASD. 67,68
- Another gene encoding a histone methyltransferase, EHMT1 (euchromatic histone-lysine N-methyltransferase), is involved in Kleefstra syndrome through deletions (9q34.3 deletion syndrome) or mutations. EHMT1 haploinsufficiency is associated with ID and sometimes with ASD.<sup>69</sup> An AGP proband with a 9q34.3 deletion encompassing EHMT1 was identified in the present study (proband 6259-3, Table S7B).

#### 5) Comment

The minimal region of overlap between our proband and those of 9 other cases with deletions involving 3p25.3 reviewed here contains only *SETD5*. Remarkably, all deletions are *de novo*. This finding, together with the recent report of a *de novo* loss-of-function mutation in a subject with language delay and borderline IQ (70, like in our proband)<sup>41</sup> suggest that *SETD5* is involved in cognitive, social, and language development. Therefore, *SETD5* could be associated with these features in 3p deletion syndrome. Two other *de novo* variants, identified in whole exome studies in ASD,<sup>4,5</sup> are missense changes and their functional effect is difficult to predict.

Interestingly, the AGP proband with the intragenic SETD5 deletion, as well as the subject with the SETD5 loss-of-function mutation, have a borderline IQ (70), whereas other individuals with larger deletions have ID ranging from severe to moderate, suggesting that other genes in the region contribute to the cognitive deficit associated with 3p deletions. Similarly, our proband lacks some of the characteristic but variable features associated with the 3p deletion syndrome, including growth retardation and dysmorphic features, such as ptosis and depressed nasal bridge.

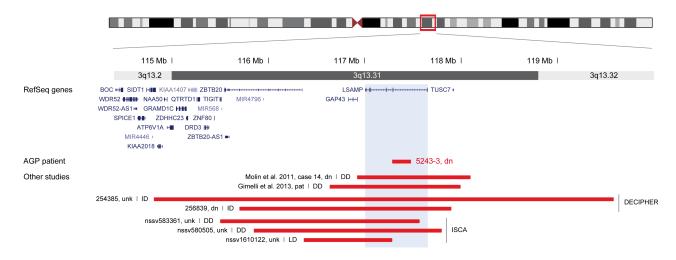


Figure S10. CNV identified in *LSAMP* (limbic system-associated membrane protein) in chromosome 3q13.31

Chr3:114,400,000-120,000,000 (hg18). Abbreviations: dn, de novo; DD, developmental delay; ID, intellectual disability; LD, learning disability; pat, paternal; unk, unknown inheritance

#### 1) AGP

- Proband 5245\_3, male, 192 kb deletion including only LSAMP (chr3:117285007-117477191), de novo. The deletion was shown to be mosaic; the percentage of mosaicism was estimated at 50% of deleted cells using a formula based on the deviation of the B allele frequency distribution.<sup>70</sup> Phenotype: born at 29 wks, intraventricular hemorrhage, mild cerebral palsy; autism (based on ADI-R and ADOS), low non-verbal IQ (<1<sup>st</sup> centile), language delay (1<sup>st</sup> centile), apraxia, abnormal sleep EEG without seizures; alopecia areata, no dysmorphic features. Multiplex family, a sister with ASD doesn't carry the CNV.
- No exonic deletions of LSAMP among 4,768 controls and 4,875 parents (1 control and 1 mother have intronic deletions).

#### 2) Other human genetic evidence

## CNV:

- Molin et al.<sup>71</sup>: Subject 14, male, 1.18 Mb 3q13.31 deletion (chr3:116922662-118098190) containing three genes, *GAP43*, *LSAMP* and *TUSC7*. Uncertain inheritance (absent in the mother and father not tested). Five year old boy with developmental delay; no other phenotype information provided. *GAP43* is found in growth cones of extending axons in the central nervous system. (Corresponds to subject 252520 in DECIPHER.)
- Gimelli et al.<sup>72</sup>: Girl with 1.36 Mb 3q13.31 deletion (chr3:116640577-118002810) containing three genes, *GAP43, LSAMP*, and *TUSC7*. The deletion was inherited from the father, who had slightly delayed psychomotor development but his cognitive level was not tested. The proband had developmental delay, clumsiness and attention deficit, associated with renal, vascular and skeletal anomalies.
- DECIPHER: Subject 254385, female, 4.8 Mb 3q13.2-q13.32 deletion involving LSAMP (chr3:114813585-119579912), inheritance
  unknown. Phenotype: ID, muscular hypotonia, downslanted palpebral fissures, joint laxity, open mouth. Only CNV reported in the
  subject.
- DECIPHER: Subject 256839, female, 2.2 Mb 3q13.31 deletion involving LSAMP (chr3:115701890-117895614), de novo. Phenotype:
   ID, broad nasal tip, short nose, anteverted nares. Only CNV reported in the subject.
- ISCA: Subject nssv583361, unknown gender, 2 Mb 3q13.31 deletion involving LSAMP (chr3:115499855-117567970), inheritance
  unknown. Phenotype: developmental delay and additional significant developmental and morphological phenotypes referred for
  genetic testing.
- ISCA: Subject nssv580505, unknown gender, 1.9 Mb 3q13.31 deletion involving LSAMP (chr3:115850559-117797499), inheritance unknown. Phenotype: global developmental delay.
- ISCA: Subject nssv1610122, unknown gender, 918 kb 3q13.31 deletion involving LSAMP and GAP43 (chr3:116367264-117285252), inheritance unknown. Phenotype: specific learning disability.

#### 3) Functional evidence

LSAMP encodes a neuronal surface glycoprotein belonging to the cell adhesion molecule (CAM) family, found in cortical and subcortical regions of the limbic system. During development of the limbic system, it is found on the surface of axonal membranes and growth cones, where it acts as a selective homophilic adhesion molecule, and guides the development of specific

patterns of neuronal connections. LSAMP mediates selective neuronal growth and axon targeting and also contributes to the guidance of developing axons and remodeling of mature circuits in the limbic system. This protein is essential for normal growth of the hippocampal mossy fiber projection.<sup>73</sup> Lsamp<sup>-/-</sup> mice have normal gross neuroanatomical organization but display heightened reactivity to novelty, reduced anxiety-like behaviors, impaired synaptic plasticity, and spatial memory deficit.<sup>74,75</sup> LSAMP has also been shown to function as a tumor suppressor gene.

• Brain expression: LSAMP is expressed on limbic regions but also, less intensely, in midbrain and hindbrain regions.

#### 4) Other evidence

% HI = 10.5 (likely to be haploinsufficient)

#### 5) Comment

The minimal region of overlap between our proband and seven other cases with 3q13.31 deletions reviewed here contains only the *LSAMP* gene. All cases with known inheritance are *de novo* and would thus support involvement of deletions in this region in neurodevelopmental disorders. Other cases with deletions involving only *LSAMP* or deleterious mutations are needed to implicate this gene in ASD and ID.

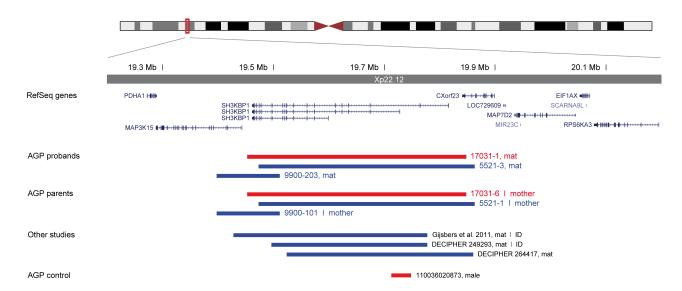


Figure S11. CNV identified in *SH3KBP1* (SH3-domain kinase binding protein 1) in chromosome Xp22.12

ChrX:19,200,000-20,200,000 (hg18). Abbreviations: ID, intellectual disability; mat, maternal

#### 1) AGP

- Proband 17031\_1, male, deletion involving SH3KBP1 and CXorf23 (chrX:19450969-19845766), maternal. CXorf23 encodes a
  protein of unknown function. Phenotype: autism (based on ADI-R and ADOS), language delay, functional language, verbal IQ 79;
  physical examination at 5 y 8 mo revealed no dysmorphic features, neurological examination was normal except for a deficit in
  coordination and gross and fine motor development; no seizures, normal sleep EEG. Sporadic case, mother unaffected, negative
  family history of neuropsychiatric disorders.
- Proband 5521\_3, male, partial duplication of SH3KBP1 and CXorf23 (chrX:19471138-19861338), maternal. Phenotype: autism (based on ADI-R and ADOS), below average IQ (<1<sup>st</sup> centile), nonverbal, seizure disorder, coarse facial features. Sporadic case, mother unaffected.
- Proband 9900\_203, male, partial duplication of SH3KBP1 and MAP3K15 (chrX:19396116-19509785), maternal. Expression study in cell lines showed no increased expression of SH3KBP1. The CNV also involves MAP3K15, encoding a mitogen-activated protein (MAP) kinase expressed in the brain. Phenotype: autism (meets criteria on ADI-R and ADOS), language delay (first words 18 mo, first phrases 42 mo), limited language, moderate ID (Bayley Scales of Infant Development II, mental developmental index 45), strabismus (like his mother), normal physical exam, no dysmorphic features, no epilepsy. Sporadic case, mother unaffected.
- 1 of 2,022 male controls carries a SH3KBP1 deletion overlapping only exon 1 of transcript variant 2, not present in other isoforms (chrX:19710729-19746114); no deletions among 2,441 fathers.

#### 2) Other human genetic evidence

#### CNV:

- Gijsbers et al. <sup>76</sup>: Male, 349 kb duplication overlapping partially 2 genes, *SH3KBP1* and *MAP3K15* (chrX:19425768-19775308), maternal. Phenotype: growth retardation, severe ID, absent or delayed speech, stereotypic movement of head and hands, bitemporal narrowing, narrow palpebral fissures, deep-set eyes, large mouth, widely spaced teeth. The healthy mother and grandmother carried the same Xp22.12 duplication and showed skewed X inactivation. The authors interpreted the CNV as potentially pathogenic.
- DECIPHER: Subject 249293, male, intragenic *SH3KBP1* duplication (chrX:19494754-19775308), maternal. Phenotype: ID, narrow forehead, abnormality of the eyebrow, deep set eyes, abnormality of the mouth, widely spaced teeth, proportionate short stature, abnormal CNS myelination. Only CNV reported in the subject.
- DECIPHER: Subject 264417, male, duplication overlapping partially 2 genes, SH3KBP1 and CXorf23 (chrX:19521667-19858019), maternal. No phenotype information; only CNV reported in the subject.

#### 3) Functional evidence

- The SH3KBP1 gene encodes CIN85, an endocytic scaffold protein that facilitates protein-protein interactions and has been
  implicated in numerous cellular processes including apoptosis, cytoskeletal rearrangement, cell adhesion and clathrin-dependent
  endocytosis. CIN85 plays a role in receptor internalization, including dopamine receptors and epidermal growth factor
  receptor.<sup>77,78</sup> CIN85 localizes at synapses and interacts with the scaffold protein S-SCAM via dendrin.<sup>79</sup>
- Mice lacking CIN85 exon 2 (present in both isoforms expressed in the central nervous system) show hyperactivity and increased exploratory behavior, but no alterations in synaptic plasticity or learning and memory. These mice show increased dopamine and dopamine D2 receptors in the striatum, due to impaired endocytic internalization of D2 receptors.
- · Expressed in the brain.

#### 4) Comment

The deletion in proband 17031\_1 results in a *SH3KBP1* null allele. The intragenic duplication in DECIPHER subject 249293 is also likely to disrupt the gene, acting as a deletion. In contrast, the functional consequence of the partial duplications observed in AGP probands 5521-3 and 9900-203, and in the two other subjects (<sup>76</sup> and DECIPHER 264417) are difficult to predict and thus can not be counted as evidence in favor of the implication of *SH3KBP1* alterations in neurodevelopmental disorders, in the absence of expression studies. In one proband (9900\_203) in whom we evaluated *SH3KBP1* mRNA expression in cell lines, no alteration was observed. Other affected males with deletions, intragenic duplications or deleterious mutations of *SH3KBP1* are required to implicate loss of function of this gene in ASD and ID.

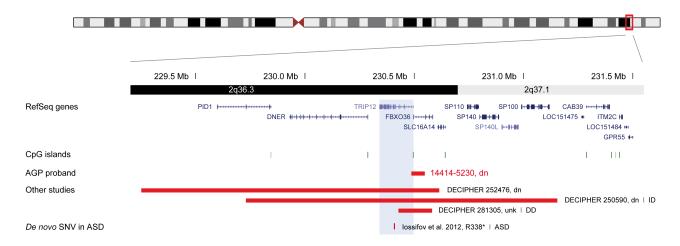


Figure S12. CNV and SNV identified in *TRIP12* (thyroid hormone receptor interactor 12) in chromosome 2q36.3

Chr2:229,200,000-231,550,000 (hg18). Abbreviations: ASD, autism spectrum disorder; DD, developmental delay; dn, de novo; ID, intellectual disability; unk, unknown inheritance

#### 1) AGP

• Proband 14414\_5230, male, 60 kb deletion involving 2 genes, *TRIP12* and *FBXO36* (chr2:230486629-230547253), *de novo*. The deletion only involves the first exon of *TRIP12*, which is non-coding, but seems to be part of the promoter of the gene since a CpG island is located in this region. The other deleted gene, *FBXO36*, encodes a F-box protein that plays a role in ubiquitination and is

not likely to be haploinsufficient (% HI = 52.9). Phenotype: autism (ADI-R and ADOS positive), no language delay (first words 12 mo, first phrases 24 mo), verbal, Griffiths at 5 y 9 mo: language DQ 87, performance DQ 78, global DQ 79.

• No TRIP12 deletions among 4,768 controls and 4,875 parents.

#### 2) Other human genetic evidence

#### CNV:

- DECIPHER: Subject 252476, female, 1.4 Mb deletion involving TRIP12 (chr2:229250832-230614988), de novo. No phenotype information; only CNV reported in the subject.
- DECIPHER: Subject 250590, female, 1.4 Mb deletion involving TRIP12 (chr2:229728861-231153046), de novo. Phenotype: ID, delayed speech and language development, epicanthus, hypermetropia, low hanging columella, palpebral edema, broad philtrum, thin upper lip vermilion, wide mouth. Only CNV reported in the subject.
- DECIPHER: Subject 281305, male, 144 kb deletion involving TRIP12 (chr2:230432282-230576372), inheritance unknown.
   Phenotype: global developmental delay and cystic renal dysplasia. Only CNV reported in the subject.

#### SNV

lossifov et al.<sup>4</sup> (exome sequencing in ASD): Subject 12867, female, TRIP12 de novo nonsense mutation. Phenotype: ASD (no other information provided).

#### 3) Functional evidence

- TRIP12 encodes a HECT-type E3 ubiquitin-ligase, which plays a role in degradation of ubiquitin fusion substrates and can regulate chromatin function to maintain genome integrity.
- Expressed in the brain.

#### 4) Other evidence

- % HI = 5.3 (highly likely to be haploinsufficient)
- HUWE1, mutated in X-linked ID, also encodes a HECT-type E3 ubiquitin-ligase involved in the ubiquitin-fusion degradation (UFD) pathway. Double knock-down of HUWE1 and TRIP12 results in additive stabilization of an UFD substrate, suggesting functional redundancy between both proteins.

#### 5) Comment

The evidence from the AGP subject is not very strong; although the *TRIP12* deletion is *de novo*, only the first non-coding exon is involved and the effect on the protein is difficult to predict. Expression studies are necessary to interpret the clinical significance of this CNV. The support from other human genetic studies is limited, but the *de novo* loss-of function *TRIP12* mutation in a female with ASD identified in an exome sequencing study, together with the function of the protein, make this gene an interesting candidate for ASD.

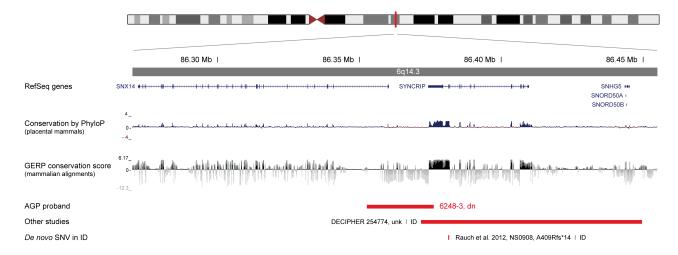


Figure S13. CNV and SNV identified in *SYNCRIP* (synaptotagmin binding, cytoplasmic RNA interacting protein) in chromosome 6q14.3

Chr6:86,270,000-86,455,000 (hg18). Abbreviations: dn, de novo; ID, intellectual disability; unk, unknown inheritance

#### 1) AGP

- Proband 6248\_3, male, 23 kb deletion involving 2 genes, SYNCRIP and SNX14 (chr6:86352577-86376159), de novo. The deletion only involves the 3' untranslated region (UTR) of SYNCRIP; this region might contain regulatory elements that are crucial for gene expression. SNX14 encodes a member of the sorting nexin family that are involved in intracellular trafficking; this gene is not predicted to be haploinsufficient (% HI = 58.6), but its contribution to the phenotype can not be ruled out. Phenotype: autism, severe ID, no language; born by C-section with increased height and head circumference. One absence seizure at 11 y, with abnormal EEG, probably related to medications taken at that time, no other seizures afterwards. At 15 y: height >+3 SD, macrocephaly (head circumference >+3 DS) (his unaffected father and 2 brothers are also very tall and have macrocephaly); long and narrow hands and feet, 2 café-au-lait spots, normal neurological exam.
- No SYNCRIP deletions among 4,768 controls; 1 carries a partial SYNCRIP duplication (chr6:86382351-87814038); no CNV overlapping SYNCRIP in 4,875 parents.

#### 2) Other human genetic evidence

#### CNV:

DECIPHER: Subject 254774, male, 78 kb deletion including SYNCRIP (chr6:86371713-86449627), inheritance unknown. The
deletion also involves SNHG5 (snoRNA host gene) as well as SNORD50A and SNORD50B (snoRNAs). Only CNV reported in the
subject. Phenotype: ID.

Note that there are several large deletions (>5 Mb) overlapping this gene in DECIPHER and ISCA, not reviewed here.

#### SNV

Rauch et al.<sup>41</sup> (exome sequencing in nonsyndromic sporadic ID): Subject NS0908, female, SYNCRIP frameshift mutation, de novo.
Phenotype: severe nonsyndromic sporadic ID (IQ <50), myoclonic astatic seizures (onset at 13 mo), no ASD, MRI at 24 mo: prominent lateral ventricles.</li>

#### 3) Functional evidence

- SYNCRIP encodes a nuclear ribonucleoprotein implicated in mRNA processing mechanisms including mRNA stability and transport, RNA editing and splicing and localized mRNA translation. SYNCRIP is a component of mRNA transport granules in dendrites.
   Selective mRNA transport, local translation and subsequent protein synthesis in neuronal dendrites are part of the fundamental mechanisms involved in synaptic plasticity, learning and memory.
- Other RNA binding proteins have been implicated in ID and ASD, including FMRP, involved in fragile X syndrome, the most frequent monogenic cause of ID and ASD, and ZC3H14, involved in recessive non-syndromic ID.
- Expressed in the brain.

## 4) Other evidence

• % HI = 1.5 (highly likely to be haploinsufficient)

#### 5) Comment

The AGP proband carries a *de novo SYNCRIP* deletion that only affects the highly conserved 3'UTR of the gene. Expression studies are necessary to determine the effects of the deletion. Although this gene does not have support from many other CNV studies, the *de novo* loss-of function *SYNCRIP* mutation reported in an individual with ID and the function of the encoded protein make this gene a strong candidate for involvement in neurodevelopmental disorders. Additional cases with small deletions affecting *SYNCRIP* and/or with deleterious mutations are required to implicate this gene definitely in ASD, ID and other neurodevelopmental phenotypes.

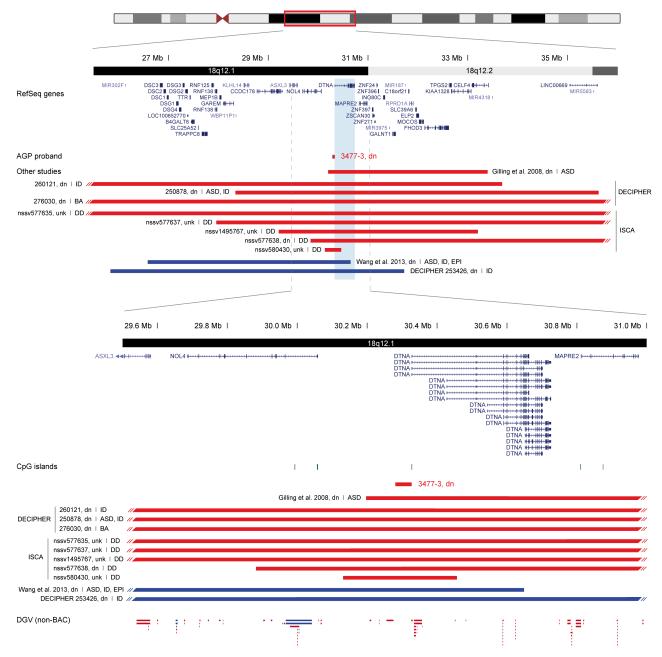


Figure S14. CNV identified in DTNA (dystrobrevin alpha) in chromosome 18q12.1

Chr18:25,500,000-36,000,000 (hg18). Abbreviations: ASD, autism spectrum disorder; BA, behavioral/psychiatric abnormality; dn, *de novo*; DD, developmental delay; EPI, epilepsy; ID, intellectual disability; unk, unknown inheritance

#### 1) AGP

- Proband 3477\_3, male, 47 kb deletion involving DTNA (chr18:30280260-30327512), de novo. Only the first non-coding exon of the longer isoforms of DTNA (2, 5, 7 and 17) is disrupted by the deletion. It may be part of the promoter of the gene since a CpG island is located in the region. Phenotype: ASD (ASD on ADI-R, autism on ADOS), verbal; WISC-R at 5 y 7 mo: verbal IQ 88, performance IQ 88, full scale IQ 86; no cardiovascular or neuromuscular abnormalities.
- No DTNA deletions among 4,768 controls and 4,875 parents.

#### 2) Other human genetic evidence

#### CNV:

• Gilling et al.<sup>84</sup>: Female, de novo translocation t(5;18)(q34;q12.2), with a 3.2 Mb deletion at the 18q breakpoint encompassing 20 genes including DTNA (chr18:30197000-33392000); the breakpoint on 5q did not contain any known genes. She also carried a

- 1.27 Mb deletion on chromosome 4q35, containing two genes (*MTNR1A* and *FAT*); inherited from her father. Phenotype: born at term, prolonged delivery with asphyxia noted at birth. She presented mild cerebral palsy, language delay, autism (met criteria on ADI-R and ADOS), no ID (WAIS-R at 34 y: verbal IQ 78, performance IQ 105, full scale IQ 88), high-grade myopia, no dysmorphism, hyperflexible joints, no seizures.
- DECIPHER: Subject 260121, female, 13.4 Mb 18q12.1 deletion (chr18:20286120-33684898), de novo. Phenotype: ID, abnormality of the face, malformation of the heart and great vessels. Only CNV reported in the subject.
- DECIPHER: Subject 250878, male, 7.3 Mb deletion including DTNA (chr18:28338083-35619727), de novo. Phenotype: ID (full scale IQ 40-50), autism, delayed speech and language development, flat occiput, hypotelorism, narrow nasal bridge, narrow nares, narrow mouth, high palate, wide intermamillary distance, absent nipples, proximal placement of thumb, hypotonia, tall stature, abnormality of the male genitalia. This individual also carries a duplication involving 3 genes, HAO1, TMX4 and PLCB1 (chr20:7022125-8482355), inherited from a normal parent.
- DECIPHER: Subject 276030, male, 14.5 Mb deletion including DTNA (chr18:25269813-39765123), de novo. The deletion is mosaic (percentage of mosaicism not indicated). Phenotype: behavioral/psychiatric abnormality, autoagression and motor delay. Only CNV reported in the subject.
- ISCA: Subject nssv577635, unknown gender, 10.9 Mb 18q12.1 deletion including DTNA (chr18:25278473-36237614), inheritance
  unknown. Phenotype: developmental delay and additional significant developmental and morphological phenotypes referred for
  genetic testing.
- ISCA: Subject nssv577637, unknown gender, 11.9 Mb 18q12.1 deletion including DTNA (chr18:27945491-39904182), inheritance unknown. Phenotype: global developmental delay, muscular hypotonia, short stature.
- ISCA: Subject nssv1495767, unknown gender, 4 Mb deletion including DTNA (chr18:29207760-33198709), inheritance unknown.
   Phenotype: developmental delay and additional significant developmental and morphological phenotypes referred for genetic testing.
- ISCA: Subject nssv577638, unknown gender, 7.3 Mb 18q12.1 deletion including *DTNA* (chr18:29881080-37228316), *de novo*. Phenotype: global developmental delay, strabismus, stridor.
- ISCA: Subject nssv580430, unknown gender, deletion involving only DTNA (chr18:30131834-30456329), inheritance unknown.
   Only the first non-coding exon of the longer isoforms (2, 5, 7 and 17) is disrupted by the deletion, which is very similar to the one observed in the AGP proband 3477-3. Phenotype: developmental delay and additional significant developmental and morphological phenotypes referred for genetic testing.
- Wang et al.<sup>85</sup>: Female, 4.1 Mb duplication including *DTNA* (chr18:26587700-30649280), *de novo*. Phenotype: autistic disorder (met criteria on ADI-R and ADOS), language delay, non-verbal, mild ID, focal epilepsy, short stature (5<sup>th</sup> centile), no dysmorphic features, mild myopia.
- DECIPHER: Subject 253426, male, 5.9 Mb duplication including DTNA (chr18:25834124-31721175), de novo. Phenotype: moderate
  ID (WAIS-IV full scale IQ 51), no ASD, recurrent seizures, facial dysmorphism, short stature, Chiari malformation, cryptorchidism,
  strabismus. Only CNV reported in the subject.

The smallest region of overlap among these 11 cases contains only the DTNA gene.

#### SNV:

 Ichida et al.<sup>86</sup>: Heterozygous missense mutation in DTNA in a large pedigree with left ventricular non compaction (family NVLNC-09), no neurological/behavioral phenotype described. No other DTNA mutations reported in the literature.

#### 3) Functional evidence

- DTNA belongs to the dystrobrevin subfamily of the dystrophin family, like DMD, involved in Duchenne's/Becker muscular dystrophy, sometimes associated with ID/ASD. Bystrobrevin alpha is part of the transmembrane dystrobrevin-associated protein complex (DPC), which participates in synaptic transmission at the neuromuscular junction, long-term memory and synaptic plasticity.
- $\alpha$ -dystrobrevin knockout mice exhibit mild muscular dystrophy but show no obvious CNS defects, likely reflecting coexpression of the homolog  $\beta$ -dystrobrevin, which is predominantly expressed in the brain. Double mutants lacking both  $\alpha$ -dystrobrevin and  $\beta$ -dystrobrevin exhibit synaptic and behavioral defects similar to those seen in dystrophin-deficient mice. Both dystrobrevin isoforms are required for the maturation and function of a subset of inhibitory synapses in the cerebellum and for correct execution of motor behaviors that depend on cerebellar integrity.
- · Expressed in the brain.

#### 4) Other evidence

- % HI = 3.2 (highly likely to be haploinsufficient)
- Upregulation of DTNA has been reported in the temporal cortex of subjects with autism<sup>92</sup> and in the prefrontal cortex of individuals with bipolar disorder.<sup>93</sup>

 Dystrobrevin immunostaining is severely reduced at the sarcolemma of individuals with Duchenne muscular dystrophy and to a lesser extent in individuals with Becker muscular dystrophy.

#### 5) Comment

The gene function and the description of overlapping CNV would make this an excellent candidate gene, potentially pathogenic. However, the deletion in the AGP proband only affects the first non-coding exon of the longer isoforms, so it is difficult to know if it is deleterious. Expression studies are necessary to assess the effect on mRNA in this individual. Furthermore, description of other cases with small CNV overlapping *DTNA* or SNV are required to implicate this gene specifically. Indeed, given that the majority of the overlapping CNV are very large and contain many genes, it is difficult to ascribe pathogenicity to alterations of a single gene in the interval.

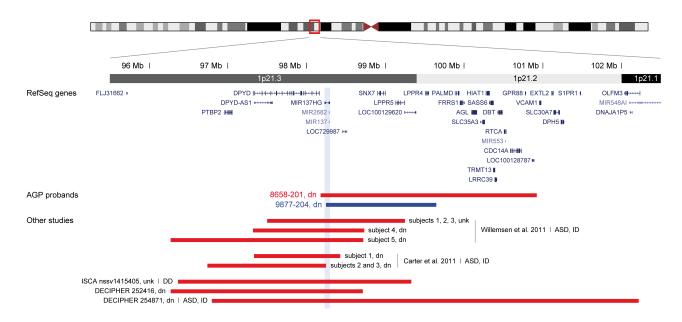


Figure S15. CNV overlapping MIR137 (microRNA 137) in chromosome 1p21.3

Chr1:95,500,000-102,500,000 (hg18). Abbreviations: ASD, autism spectrum disorder; dn, *de novo*; DD, developmental delay; ID, intellectual disability; unk, unknown inheritance

#### 1) AGP

- Proband 8658\_201, female, 2.7 Mb 1p21.3-p21.2 deletion containing 22 genes including MIR137 (chr1:98175622-100923952), de novo.
   Phenotype: autism on ADI and ADOS, comorbid ADHD, no language delay; low average IQ (WASI at 21 y: verbal IQ 78, performance IQ 88, full scale IQ 81); overweight, height and head circumference 50<sup>th</sup> centile, high pain tolerance, no epilepsy.
- Proband 9877\_204, male, 1.4 Mb 1p21.3-p21.2 duplication containing 8 genes including MIR137 (chr1:98247355-99645560), de novo. Phenotype: autism on ADI-R and ADOS, language delay (first words 36 mo, first phrases 60 mo), functional language, mild ID (WISC-R at 11 y: verbal IQ 51, performance IQ 80, full scale IQ 64), normal height and head circumference, weight -1.6 SD, no dysmorphic features, normal physical exam, no epilepsy.

Both CNV in AGP probands also involve the *LPPR4* and *LPPR5* genes (lipid phosphate phosphatase-related proteins, types 4 and 5). *LPPR4* is specifically expressed in neurons and involved in axonal outgrowth during development and regenerative sprouting; <sup>95</sup> no haploinsufficiency score available (no information for *LPPR5*).

• No MIR137 deletions among 4,768 controls; 1 carries a MIR137 duplication (chr1:97673140-98319409); no CNV in 4,875 parents.

#### 2) Other human genetic evidence

#### CNV:

Willemsen et al.<sup>96</sup>: Chromosome 1p21.3 microdeletions comprising DPYD and MIR137 associated with ID in 3 sibs and 2 unrelated subjects; the minimal region of overlap includes only DPYD and MIR137. The individuals displayed decreased expression of both precursor and mature miR-137 levels, as well as increased expression of the downstream targets MITF, EZH2, and KLF4. DPYD is involved in autosomal recessive dihydropyrimidine dehydrogenase deficiency; the significance of a defect in only one allele is uncertain.

<u>Subjects 1, 2 and 3:</u> siblings carrying a 1.75 Mb 1p21.3 deletion (chr1:97500000-99250000), inheritance unknown (parents deceased). <u>Subject 1:</u> male, borderline-mild ID (verbal IQ 65, performance IQ 90, full scale IQ 73), features of ASD, tendency to overeat, remarkably shy and friendly behavior, weight 90<sup>th</sup> centile, deep set eyes, broad nasal tip, long ears, thick lower lip, myopia. <u>Subject 2</u>: male, mild-moderate ID (verbal IQ < performance IQ 70, full scale IQ 52), features of ASD, tendency to overeat, self mutilation, aggressive outbursts, remarkably shy and friendly behavior, speech deficits, weight >98<sup>th</sup> centile, broad nasal tip, long ears, macrostomia, thick lower lip, astigmatism. <u>Subject 3</u>: female, mild-moderate ID (no other information available).

<u>Subject 4</u>: male, 1.41 Mb 1p21.3 deletion (chr1:97320000-98730000), *de novo*. Phenotype: mild ID (verbal IQ 59, performance IQ 71, global IQ 62), features of ASD, tendency to overeat, remarkably shy and friendly behavior, speech deficits, weight 98<sup>th</sup> centile, deep set eyes, astigmatism, myopia, broad nasal tip, full cheeks, thick lower lip, micrognathia and long ears.

<u>Subject 5</u>: female, 2.45 Mb 1p21.3 deletion (chr1:96270000-98720000), *de novo*. Phenotype: mild ID (global IQ 66), aggressive outbursts, remarkably shy and friendly behavior, weight >98<sup>th</sup> centile, full cheeks, long ears, thick lower lip.

- Carter et al.<sup>97</sup>: <u>Subject 1</u>: male, 1.1 Mb 1p21.3 deletion (chr1:97332167-98424667), *de novo*. Phenotype: severely delayed language, ID, autism. The individual also carries a balanced translocation t(9;21)(p13.3;q22.1) and has a *PTCHD1* missense mutation. Both abnormalities are inherited from the mother. The translocation was also transmitted to a healthy sister. <u>Subjects 2 and 3</u>: siblings with a 1.5 Mb 1p21.3 deletion (chr1:96742150-98243813), *de novo*. In their mother, the deleted region from chromosome 1p21.3 was inserted into chromosome 10. <u>Subject 2</u>: female, severe language delay, adaptive skills severely delayed, autism. <u>Subject 3</u>: male, language delay, no ID (full scale IQ 99), ASD. Both sibs had mild dysmorphic features, including upslanting palpebral fissures and small joint hypermobility.
- ISCA: Subject nssv1415405, unknown gender, 3 Mb 1p21.3 deletion (chr1:96362589-99332669), inheritance unknown.
   Phenotype: developmental delay and additional significant developmental and morphological phenotypes referred for genetic testing.
- DECIPHER: Subject 252416, female, 2.4 Mb 1p21.3 deletion (chr1:96274145-98715464), de novo. No phenotype information; only CNV reported in the subject.
- DECIPHER: Subject 254871, male, 5.4 Mb 1p21.3-p21.1 deletion (chr1:96792350-102220124), de novo. Phenotype: autism, ID, spotty hyperpigmentation, precocious puberty. Only CNV reported in the subject.

#### 3) Functional evidence

- The mature microRNA transcript miR-137 regulates neuronal maturation: overexpression of miR-137 inhibits dendritic and spine morphogenesis in newborn cells in the adult hippocampus and in cultured hippocampal neurons, whereas a reduction in miR-137 had opposite effects, miR-137 has also been shown to modulate neurogenesis in adult neural stem cells. Significant enrichment of miR-137 at the synapses of cortical and hippocampal neurons suggests a role in regulating local synaptic protein synthesis machinery.
- Expressed in the brain, enriched in neurons, at the synaptic compartment.

#### 4) Other evidence

• An intronic SNP in *MIR137* was strongly associated with schizophrenia in a mega-analysis combining genome-wide association study data from over 40,000 individuals. <sup>100</sup>

## 5) Comment

1p21.3 deletions of variable sizes overlapping *MIR137* reported in 11 individuals with ASD and/or ID. All cases in which inheritance is known originated *de novo*. Interestingly, both overexpression and inhibition of miR-137 had significant but opposite effects on dendritic development of hippocampal neurons, suggesting that the *MIR137* gene may be dosage sensitive, and that both the deletion and duplication observed in AGP probands could interfere with neuronal maturation.

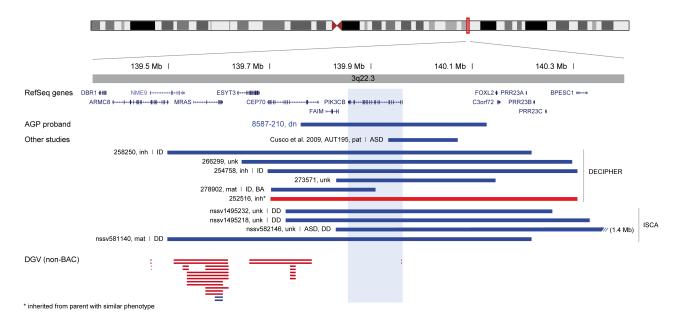


Figure S16. CNV identified in *PIK3CB* (phosphatidylinositol-4,5-bisphosphate 3-kinase, catalytic subunit beta) in chromosome 3q22.3

Chr3:139,350,000-140,400,000 (hg18). Abbreviations: ASD, autism spectrum disorder; BA, behavioral/psychiatric abnormality; dn, *de novo*; DD, developmental delay; inh, inherited; ID, intellectual disability; mat, maternal; pat, paternal; unk, unknown inheritance

#### 1) AGP

- Proband 8587\_210, female, 366 kb duplication involving PIK3CB, CEP70 and FAIM (chr3:139760896-140127703), de novo. The entire PIK3CB gene is duplicated, which could result in increased expression, leading to excessive phosphatidylinositol 3-kinase (PI3K) activity. Phenotype: autism (ADI-R and ADOS positive), no language delay (first words 10 mo, first phrases 12 mo), verbal, WISC-III at 13 y 9 mo: verbal IQ 92, performance IQ 78, full scale IQ 84.
- No deletions or duplications of *PIK3CB* among 4,768 controls and 4,875 parents.

#### 2) Other human genetic evidence

#### CNV:

- Cusco et al.<sup>101</sup>: Subject AUT195, male, 3q22.3 PIK3CB partial duplication (chr3:139934042-140070771), paternal. Phenotype: autism, mild ID, unilateral sensorineural deafness, no dysmorphism, no seizures. The functional consequences of a partial duplication are unknown.
- DECIPHER: Subject 258250, female, 720 kb 3q22.3 duplication involving PIK3CB (chr3:139497567-140217021), inherited from normal parent. Phenotype: cognitive impairment, Dandy-Walker malformation. Only CNV reported in the subject.
- DECIPHER: Subject 266299, male, 650 kb 3q22.3 duplication involving PIK3CB (chr3:139644292-140297350), inheritance unknown. Phenotype: microtia. This individual carries another duplication (chr3:160280710-160878458), inheritance unknown.
- DECIPHER: Subject 254758, male, 610 kb 3q22.3 duplication involving *PIK3CB* (chr3:139695765-140307747), inherited from normal parent. Phenotype: ID, short attention span, microcephaly, long face, high anterior hairline, downslanted palpebral fissures, depressed nasal tip, prominent nasal bridge, macrotia, micrognathia, abnormality of the pinna, prominent ears, slender build, scoliosis, mitral regurgitation, atrial septum defect, cryptorchidism, large hands. Only CNV reported in the subject.
- DECIPHER: Subject 273571, male, 315 kb 3q22.3 duplication involving *PIK3CB* (chr3:139830639-140145866), inheritance unknown. No phenotype information; only CNV reported in the subject.
- DECIPHER: Subject 252516, female, 606 kb 3q22.3 deletion involving *PIK3CB* (chr3:139701603-140307606), inherited from parent with similar phenotype. No phenotype information; only CNV reported in the subject.
- DECIPHER: Subject 278902, female, 207 kb 3q22.3 duplication involving *PIK3CB* (chr3:139701603-139908842), maternal. Phenotype: moderate ID and behavioral/psychiatric abnormality. The phenotype of the mother is unknown. Only CNV reported in the subject.

- ISCA: Subject nssv1495232, unknown gender, 527 kb 3q22.3 duplication involving PIK3CB (chr3:139730880-140257978), inheritance unknown. Phenotype: developmental delay and additional significant developmental and morphological phenotypes referred for genetic testing.
- ISCA: Subject nssv1495218, unknown gender, 600 kb 3q22.3 duplication involving PIK3CB (chr3:139730880-140331637), inheritance unknown. Phenotype: developmental delay and additional significant developmental and morphological phenotypes referred for genetic testing.
- ISCA: Subject nssv582146, unknown gender, 1.4 Mb 3q22.3-q23 duplication involving PIK3CB (chr3:139830647-141256588), inheritance unknown. Phenotype: autism, developmental delay and additional significant developmental and morphological phenotypes referred for genetic testing.
- ISCA: Subject nssv581140, unknown gender, 719 kb 3q22.3 duplication involving PIK3CB (chr3:139497567-140217080), maternal.
   Phenotype: developmental delay and additional significant developmental and morphological phenotypes referred for genetic testing.

#### 3) Functional evidence

- PIK3CB encodes an isoform of the catalytic subunit PI3Kbeta of phosphoinositide 3-kinase (PI3K). These signaling molecules
  activate a wide range of downstream targets that regulate multiple cellular processes, including intracellular trafficking of
  proteins, cell proliferation, migration and survival.
- Expressed in the brain.

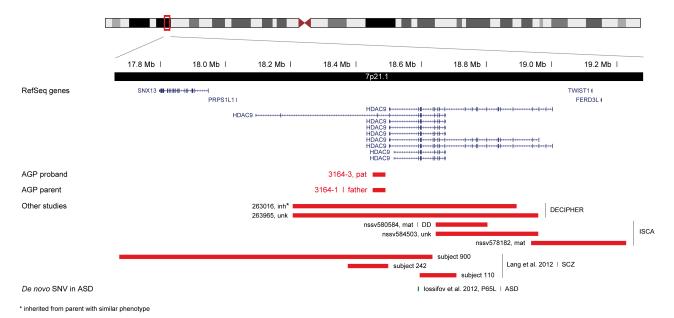
#### 4) Other evidence

- % HI = 0.5 (highly likely to be haploinsufficient)
- PI3K is regulated by the fragile X mental retardation protein (FMRP),<sup>10</sup> and is elevated in fragile X syndrome Fmr1-knockout mice,<sup>102,103</sup> suggesting that dysregulated PI3K signaling may underlie the synaptic impairments in fragile X syndrome. Accordingly, PI3K antagonists rescue fragile X syndrome phenotypes, including dysregulated synaptic protein synthesis, excess AMPA receptor internalization, and increased spine density.<sup>102</sup>

#### 5) Comment

The only report identified in the literature is a partial *PIK3CB* duplication in a subject with ASD, paternal;<sup>101</sup> the functional consequences of a partial duplication are difficult to predict. Eight cases of whole *PIK3CB* duplication are described in DECIPHER and ISCA, 4 are inherited and for the other 4 the inheritance is unknown. Thus, the duplication of *PIK3CB* identified in the AGP proband 8587-210 is the only one *de novo*. No similar duplications overlapping this gene were found among AGP controls, parents or population controls in DGV. Taken together these findings suggest that *PIK3CB* duplications could represent a risk factor for ASD/ID, associated with incomplete penetrance and/or variable expressivity. Further studies in larger samples of cases and controls are necessary to confirm this hypothesis.

## Examples of candidate genes affected by inherited CNV in AGP probands



## Figure S17. CNV and SNV identified in HDAC9 (histone deacetylase 9) in chromosome 7p21.1

Chr7:17,660,000-19,280,000 (hg18). Abbreviations: ASD, autism spectrum disorder; dn, *de novo*; DD, developmental delay; inh, inherited; mat, maternal; pat, paternal; SCZ, schizophrenia; unk, unknown inheritance

#### 1) AGP

- Proband 3164\_3, male, rare 40 kb deletion involving only HDAC9 (chr7:18450792-18490822), paternal. Phenotype: autism (ADI-R and ADOS positive), language delay (first words 54 mo, first sentences 60 mo), verbal, mild ID (PPVT-III at 6 y 10 m: verbal IQ 67); multiplex family, affected sibling not yet tested. Phenotype information about father not available.
- No HDAC9 exonic deletions among 4,768 controls and 4,874 parents (1 deletion in the father of proband 3164-003).

#### 2) Other human genetic evidence

#### CNV:

- DECIPHER: Subject 263016, female, HDAC9 deletion (chr7:18206711-18892382), inherited from parent with similar phenotype.
   No phenotype information; only CNV reported in the subject.
- DECIPHER: Subject 263965, female, HDAC9 deletion (chr7:18206712-18958442), inheritance unknown. No phenotype information; only CNV reported in the subject.
- ISCA: Subject nssv580584, unknown gender, HDAC9 deletion (chr7:18644447-18803445), maternal. Phenotype: global developmental delay, seizures.
- ISCA: Subject nssv584503, unknown gender, *HDAC9* deletion (chr7:18644447-18958471), inheritance unknown. Phenotype: abnormal facial shape, facial asymmetry.
- ISCA: Subject nssv578182, unknown gender, 7p21.1 deletion of 3 genes: HDAC9, TWIST1 and FERD3L (chr7:18937678-19227544), maternal. Phenotype: craniosynostosis (defects in TWIST1 cause autosomal dominant craniosynostosis type 1).
- Lang et al.<sup>104</sup>: Three schizophrenia individuals with exonic HDAC9 deletions among 3391 cases (inheritance unknown); no HDAC9 deletion in 3181 controls.

#### SNV

 lossifov et al.<sup>4</sup> (exome sequencing in ASD): Subject 13076, male, HDAC9 missense variant, de novo. The variant appears to be damaging (PolyPhen2, SIFT, PANTHER), affecting a highly conserved residue (GERP 5.93, ConSurf 9/9). Phenotype: ASD (no other information provided).

#### 3) Functional evidence

- HDAC9 encodes a histone deacetylase, expressed in the brain. Histones play a critical role in transcriptional regulation, cell cycle
  progression, and development.
- · Expressed in the brain.

#### 4) Other evidence

- % HI = 2.9 (highly likely to be haploinsufficient)
- The HDAC family of genes has already been involved in ASD/ID through HDAC4 (involved in brachydactyly-mental retardation syndrome) and HDAC8 (X-linked ID, mutations are responsible for Cornelia de Lange syndrome).

#### 5) Comment

In addition to the *HDAC9* paternal deletion identified in AGP proband 3164-003, we identified 5 other overlapping deletions in subjects in DECIPHER and ISCA, as well as three deletions in schizophrenia. The deletion was inherited in 3 subjects; no information was available for the others. No *HDAC9* exonic deletions were observed among the AGP 4,768 controls and 4,874 parents; in addition, no deletions overlapping this gene were found in DGV. Taken together these findings suggest that *HDAC9* deletions could represent a risk factor for ASD, ID and schizophrenia, associated with incomplete penetrance/variable expressivity. Further studies in larger samples of cases and controls are necessary to confirm this hypothesis.

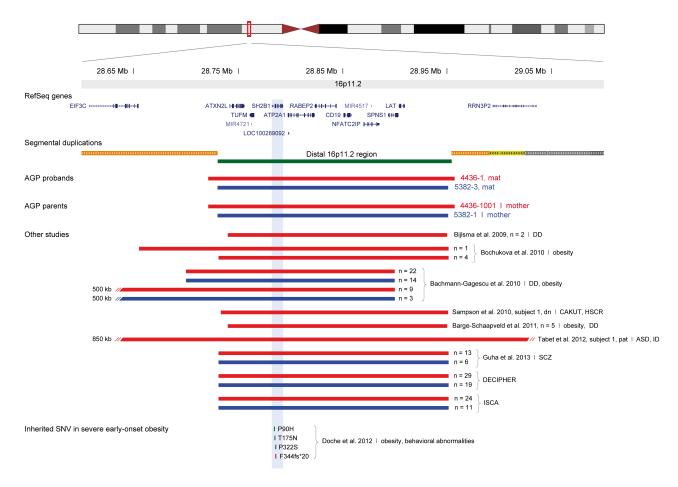


Figure S18. CNV identified in the distal 16p11.2 region containing SH2B1 (SH2B adaptor protein 1)

Chr16:28,600,000-29,100,000 (hg18). Abbreviations: ASD, autism spectrum disorder; CAKUT, congenital anomalies of the kidney and urinary tract; DD, developmental delay; HSCR, Hirschsprung disease; ID, intellectual disability; mat, maternal; pat, paternal; SCZ, schizophrenia.

Longer CNV encompassing the distal 16p11.2 region and the proximal 16p11.2 region (29.5-30.1 Mb) involved in ASD, ID, and regulation of body mass index are not shown here.

#### 1) AGP

• Proband 4436\_1, male, distal 16p11.2 deletion including *SH2B1* (chr16:28721599-28957155), maternal. Phenotype: ASD diagnosis, no language delay (first words and phrases 15 mo), verbal, WISC-IV at 8 y 8 mo: verbal IQ 95, performance IQ 67, full scale IQ 76. Sporadic case.

- Proband 5382\_3, male, distal 16p11.2 duplication including SH2B1 (chr16:28730274-28950951), maternal. Phenotype: autism
  (ADI-R and ADOS positive), language delay (first words 42 mo, first phrases 48 mo), verbal; PPVT-IV verbal IQ 82, Leiter-R Brief
  performance IQ 97 (both at 11 y 10 m). Sporadic case.
- 1 distal 16p11.2 deletion among 4,768 controls and 1 deletion among 4,875 parents; no reciprocal duplications in controls or parents (the mother of proband 5382\_3 was excluded from the microarray analyses after quality checks, her duplication was identified during gPCR validation of the CNV in her son).

#### 2) Other human genetic evidence

#### CNV

- Deletions at distal 16p11.2, with a minimal common region of 220 kb (28.73–28.95 Mb), have been implicated in early-onset obesity and developmental delay, <sup>105-108</sup> and in other variable phenotypes, including behavioral problems such as ASD and ADHD, anomalies of the kidney and urinary tract and Hirschsprung disease. <sup>109,110</sup> Whereas deletions appeared to be significantly enriched in populations with early-onset obesity or with developmental delay, <sup>105,108</sup> reciprocal duplications were not enriched in cases compared to controls. <sup>105</sup>
- A recent meta-analysis in large clinical cohorts with developmental delay, ID, ASD and congenital malformations referred for genetic testing found deletions at distal 16p11.2 in 23 of 31516 cases and in 2 of 13696 controls (OR 5, P = 0.01). Reciprocal duplications were found in 25 of 31516 cases and in 3 of 13696 controls (OR 3.62, P = 0.02). Analysis of three ASD cohorts (AGP, SSC, and AGRE; n = 3955) found 1 deletion (OR 1.73, P = 0.53) and 1 duplication (OR 1.15, P = 1). The lack of a significant effect of these CNV in ASD was suggested to be due to the relatively small sample size. 111
- Tabet et al. 110: Male, 847 kb 16p11.2 distal deletion containing *SH2B1* (chr16:28401454-29249055), paternal. Autism, severely delayed speech, childhood-onset obesity, IQ 47. At age 19, he was tall (+2.5 SD), with troncular obesity (+4 SD). The father was described as being non talkative, introverted and having few social relationships.
- Guha et al. 112: Deletions at distal 16p11.2 were reported in schizophrenia, in 13 of 13850 cases (0.094%) and 3 of 19954 controls (0.015%) (OR 6.25 [95% CI, 1.78-21.93]; *P* = 0.001). The rate of duplications in the region was not significantly different between cases and controls: 6 of 13850 cases (0.043%) vs 13 of 19954 controls (0.065%).

#### SNV:

• Doche et al. <sup>113</sup>: The minimal deleted interval contains nine genes, including *SH2B1*, which plays a role in the regulation of body weight and glucose metabolism in mice (see below). Mutation screening of *SH2B1* in 300 individuals with severe early-onset obesity revealed five mutations, one frameshift and three missense (including one found in two subjects). Mutation carriers exhibited childhood-onset obesity, hyperphagia, insulin resistance and short stature as adults. Neurobehavioral phenotypes included social isolation, speech and language delay and aggression. All mutations were inherited from overweight/obese parents reported to also have variable behavioral abnormalities. The mutations were absent from 500 controls.

## 3) Functional evidence

- SH2B1 encodes an adaptor protein that binds to a large range of receptor tyrosine kinases and is thus involved in multiple biological pathways, including leptin and insulin signaling. The widely expressed scaffold protein SH2B1 binds to the receptors for nerve growth factor, insulin and insulin-growth factor 1, and has been implicated in neuronal differentiation and neurite outgrowth.<sup>114,115</sup>
- Expressed in the brain.
- Sh2b1 deficient mice develop obesity and diabetes, a phenotype rescued by neuron-specific expression of SH2B1. 116

#### 4) Other evidence

• % HI = 18.7 (likely to be haploinsufficient)

#### 5) Comment

SH2B1 haploinsufficiency is clearly implicated in early-onset obesity. Recent evidence suggests that distal 16p11.2 deletions could also be involved in neurodevelopmental phenotypes, associated with incomplete penetrance and variable expressivity. Although a significant enrichment has been reported in samples with developmental delay/ID, the risk effect appears to be weak compared to other recurrent CNV. The implication of deletions at distal 16p11.2 in schizophrenia and the description of maladaptive behaviors in individuals carrying SH2B1 mutations, all lend further support to their role as risk factors. The involvement of distal 16p11.2 duplications in ID and ASD is difficult to assess at present, since they have not been found to be consistently enriched in cases. Further studies, comparing the frequency of distal 16p11.2 deletions and duplications in larger samples of cases and controls, are needed to clarify the impact of these CNV in neurodevelopmental disorders.

# **SUPPLEMENTAL TABLES**

**Table S1A. Autism strict and spectrum classifications** 

ASD diagnostic category -	Phenotype classifications					
ASD diagnostic category	ADI-R	ADOS				
Strict	Autism	Autism				
	Autism	NA				
Consistencias	Autism	ASD				
Spectrum	ASD	Autism				
	NA	Autism				

NA, not available or not administered

Table S1B. Quality control – Family and control sample breakdown

Quality control filters	Initial	Filter 1 Low call rate	Filter 2 Mendelian errors	Filter 3 Gender- mismatch	<b>Filter 4</b> Duplicates	Filter 5 High LRR/ BAF SD	Filter 6 Excess calls	Filter 7 Excess de novos	Filter 8 Peri- centrom.	Filter 9 Large chrom. abnorm.	Filter 10 Incomplete phenotype data		Filter 12 European- only
# Single probands	51	56	55	55	55	90	106	106	106	106	106	106	102
# Proband + mother duo	10	30	30	30	30	100	125	125	125	131	126	126	106
# Proband + father duo	12	31	31	31	31	113	127	127	127	142	137	137	119
# Complete trios*	2772	2677	2620	2613	2606	2268	2161	2158	2155	2126	2077	2077*	1820
Total # families	2845	2794	2736	2729	2722	2571	2519	2516	2513	2505	2446	2446	2147
# Father + mother only (with or without relatives)	258	277	273	273	273	309	324	324	326	332	380	199	196
# Fathers only	12	21	22	22	22	60	69	69	69	68	73	28	28
# Mothers only	16	27	27	27	27	52	60	60	61	62	67	32	32
# Relatives only	0	0	0	0	0	0	0	0	0	0	0	4	4
# Technical controls	9	9	9	8	8	8	8	8	8	8	8	8	8
Control datasets													
SAGE (dbGaP) <sup>a</sup>	1847	1847	-	1847	1829	1815	1793	-	1792	1769	-	1769	1166#
OC (European-only) <sup>b#</sup>	511	509	-	509	501	475	440	-	437	433	-	433	234#
HABC (dbGaP) <sup>c</sup>	2860	2860	-	2857	2809	2658	2571	-	2570	2566	-	2566	1240

**CNV detection and quality control evaluation**: For samples that passed the SNP and intensity QC, genome-wide CNVs were detected using a multiple-algorithm approach to maximize sensitivity and specificity of CNV calling. For a detailed description see the Supplemental information of Pinto et al. Briefly, CNVs were identified by using QuantiSNP, iPattern. Pattern. and PennCNV: the family-based CNV detection option of PennCNV was used to confirm inheritance.

We excluded CNVs when they failed stringent QC criteria: <5 probes and low confidence score (QuantiSNP log Bayes factor <15); if CNVs resided in regions of extreme GC content (>70%); or if they were within centromere proximal cytobands. CNVs detected by QuantiSNP and iPattern in one individual with a minimum of 5 consecutive probes covering at least 5 kb of sequence were merged using outer probe boundaries (i.e., union of the CNVs). All CNVs by any algorithm with size larger than 1 Mb were inspected manually, and all samples that passed all above QC filters were inspected for the presence of large abnormalities in chromosomes X and Y (that is, in addition to the algorithm calling). As a final step, we joined CNVs that appeared to be artificially split by either of the calling algorithms and also removed CNVs that spanned known large assembly gaps in hg18 (greater than 200 kb).

Filter descriptions: A total of 9,050 individuals from 2,845 ASD families were genotyped as part of Stages 1(117) and 2 and those passing QC filters were used in the rare CNV analysis. Incomplete families, where proband-father/mother duos passed QC filters were also analysed for CNVs. The number and the composition of families remaining after each filtering step is indicated. Counts may increase or decrease after each step, as removal of individuals in some instances will break complete trios into proband+parent duos or single probands. Filter 1: Low call rate or high missingness. Filter 2: High Mendelian error rate (with or without proband gender mismatch); families with unresolved gender mismatches were excluded; families where both parents had a gender mismatch without high Mendelian error rate were kept and the parents' gender swapped. Families with high Mendelian rate for one parent only were excluded. Filter 3: Proband with gender mismatch but no Mendelian error (i.e., another sib of the same family was genotyped instead) resulted in exclusion of the whole family. Technical controls with gender mismatches were excluded. Filter 4: Any duplicate samples/families had one sample/family excluded. Filter 5: Samples with high standard deviation (SD) of log R ratio of intensities (LRR) and/or B allele frequency (BAF), or showing extreme/wide intensities. Filter 6: Samples with excess of CNV calls by at least one of the algorithms, except those with fragmented calls due to large chromosome abnormalities. Filter 7: Samples with excess of de novos (which were confirmed to be false positives with experimental validation). Filter 8: Lack of CNV calls after three filters (removing pericentromeric calls, <30 kb size, 50% overlap with segmental duplication

blocks). Filter 9: Large chromosomal abnormalities >7.5 Mb. A list of chromosome abnormalities detected in probands can be found in **Table S1C**, and for parents and controls in **Table S17B**. Subjects were removed if one or more CNVs were found to be cell-line artifacts after experimental validation. If a parent failed QC at this step, his/her family was not excluded. Filter 10: No phenotype data in database or proband did not meet full criteria for ASD. Filter 11: Parents that passed QC but are parents of probands that failed QC. Filter 12: European ancestry only.

Ancestry: Ancestry for each of the four samples (AGP cases, and SAGE, HABC, and OC controls) was inferred by eigenvector decomposition and clustering. To identify European subjects from the Ontario controls, we used the multidimensional scaling (MDS) function of PLINK to cluster the OC subjects with HapMap-CEU. The remaining three samples had been described and analyzed for ancestry in previous studies. <sup>117,121,122</sup> We used the results from those published studies to identify subjects of inferred European ancestry for this study. In each of the published studies, ancestry was inferred by using SpectralGem<sup>123</sup> to analyze thousands of high quality SNPs genotyped for all subjects. After the clustering step from SpectralGem, it was simple to identify groups of European ancestry because a substantial fraction of the contributing AGP sites were European.

Data from 2,446 families passed all QC steps (13% of subjects excluded), adding 1,359 new families to the combined analysis. Of the new families, 1,168 families were European and 191 were of other ancestries. As described in detail in Pinto et al. 117, to avoid confounding by ancestry, all downstream CNV analyses used European-only cases (n=2,147) and controls (n=2,640). For the analyses presented in **Figure S1**, we extended the number of controls to include 1,843 subjects from other ancestries —517 SAGE and 1,326 HABC non-Europeans controls that passed QC— giving a total of 4,768 control subjects (2,022 males and 2,746 females) from all ancestries to be compared to ASD cases of all ancestries.

- \* de novo CNVs were detected and confirmed in a total of 2077 complete trios passing array QC plus 19 families in which at least one of the parents failed initial array QC but additional experimental validation in both parents confirmed the presence of de novo CNVs, giving a total of 2096 complete trios of all ancestries (1838 European trios = 1820 + 18) studied.
- <sup>a</sup> **Study of Addiction: Genetics and Environment (SAGE) cohort**: Both raw intensities and genotypes were obtained from 1,847 SAGE control subjects from NHGRI-dbGaP (accession: phs000092.v1.p1) as part of the larger SAGE case-control study<sup>124</sup>. The consented sample included 31% males and 69% females, with a mean age of 39.2 y (SD 9.1); 73% of subjects self-identified as European-American, 26% as African-American and 1% as other (http://zork.wustl.edu/gei /study\_description.html). Subjects may have had exposure to alcohol (and possibly to other drugs), but did not meet criteria for alcohol or other drug dependence. The subset of control dataset used in the specific CNV analyses in this paper is composed of 1,166 unrelated European control samples that passed all quality control filters (75% had DNA extracted from whole blood and 25% from cell lines), composed of 370 males and 796 females.
- Ontario Colorectal Cancer case-control study cohort (OC): 433 unrelated European control subjects from the population-based Colorectal Cancer case-control study, recruited randomly from the province of Ontario in Canada (Ontario Familial Colorectal Cancer Registry, OFCCR) as described elsewhere array. The OC control sample consisted of 199 females and 234 males with mean age of 61.8 y (range: 27-78); all subjects were self-identified as non-Hispanic whites and estimated to be of European ancestry from their genotypes. All DNA samples were extracted from whole blood. The GenomeStudio v. 2010.3, with the clustering algorithm GenTrain2 and a GenCall cutoff of 0.15 was used to generate genotypes. The same quality control procedures applied to the ASD family samples and SAGE controls were used here. For the main CNV analysis, we used only the 234 OC males.
- HealthABC (HABC): 1,240 unrelated European control subjects from the whole-genome-association study of visceral adiposity in the Health, Aging, and Body Composition (HealthABC or HABC)<sup>127</sup> were used in the main analyses. The HABC cohort studied the factors that contribute to disability and the decline in function of healthier older persons, with a particular emphasis on changes in body composition. The HealthABC study recruited 3,075 70-79 year-old community-dwelling adults (41% African-American, remainder were white-European), who were initially free of mobility and activities of daily living disability. Genotyping was performed by the Center for Inherited Disease Research (CIDR) using the Illumina Human1Mv3 (duo) BeadChip system like most of the AGP data, providing excellent comparability with the case dataset. Both raw intensities and genotypes were obtained from JAAMH-dbGaP (accession: phs000169.v1.p1) for 2,860 samples, resulting in 1,240 European (637 males and 603 females) and 1,326 non-European samples after QC. Samples were excluded from the dataset in case of sample failure, genotypic sex mismatch, or first-degree relative of an included individual based on genotype data.
- To try to balance the number of male and female controls used in the various analyses (by stage and by platform), we only included male samples from the OC dataset (n=234), and excluded 86 female SAGE samples of European ancestry (i.e., all males [n=370] and 796 females were included).

Abbreviations: chrom. abnorm., chromosomal abnormalitities; pericentrom., pericentromeric; M, males.

# Table S1C. Quality control – Chromosome abnormalities detected in probands

(Chromosome abnormalities in parents and controls are listed in Table S17B)

Stage	Sample ID	Sex	Family type	DNA source	Cytoband	Anomaly type	Karyotype	Comment
Confir	med chromo	soma	l abnorn	nalities				
1	5467_3	М	SPX	Blood	1q42.3-q44	High intensity, BAF split	N/A	13.7 Mb 1q42-q44 duplication (1:233476547- 247165725), <i>de novo</i> ; confirmed by qPCR
1	14270_3930	F	SPX	Blood	6q25.3-q27	High intensity, BAF split	46,XX.ish der(22)t(6;22) (6q25.3;p11.2) pat(6qtel+)	10 Mb 6q25.3-q27 duplication (6:160773919- 170761395); confirmed by FISH, resulting from a balanced translocation in the father
1	13137_1543	F	UNK	Blood	8p12-8q12.1	8p-q duplication	47,XX,+r[10]/4 6,XX[70]	27 Mb 8p12-8q12.1 duplication (8:31928590- 58996070); karyotype: mosaic supernumerary ring chromosome
1	5420_3	М	UNK	CL	Whole chr 21	High chr 21 intensity, BAF split	47,XY+21	Down syndrome, confirmed by karyotype
1	5266_4	F	UNK	Blood	Whole chr 2	Run of homozygosity entire chr 2	N/A	<i>De novo</i> uniparental disomy chr 2, confirmed in blood DNA
2	21020_1	M	SPX	CL (array), blood (qPCR & FISH)	4p16	4p16.3p16.1 DUP	46,XY, der(8)t(4;8)(4p 16.1→ter; 8p23.1→ter)	De novo unbalanced translocation t(4p16;8p23) leading to 4p16.3p16.1 duplication (4:53403-9016339) and 8p23.3p23.1 deletion; maternal origin by microsatellite markers. Normal karyotype, translocation validated by FISH and qPCR; the 4p terminal duplication (8.9 Mb) includes the Wolf-Hirschhorn syndrome region
2	21020_1	М	SPX	(see above)	8p23	8p23.3p23.1 DEL	(see above)	De novo unbalanced translocation t(4p16;8p23) (see above). The 6.8 Mb 8p23.3p23.1 terminal deletion (8:154984-6994825) overlaps numerous deletions described in individuals with ID
2	2175_1	М	SPX	Blood	22q13.2-qter	22q13.2-qter BAF split (no visible LRR deflection)	N/A	De novo 10 Mb 22q13.2-qter uniparental disomy (22:39729380-49582267), mosaicism in approximately 24% of cells; normal MLPA
2	4316_1	F	UNK	Blood	Whole chr X	Whole chr BAF split (no visible LRR deflection)	N/A	Triple X syndrome, <i>de novo</i> , mosaic
1	5257_3	М	SPX	Blood	Whole chr Y	Male, high chr Y intensity	47,XYY	XYY syndrome, de novo, confirmed by karyotype
1	5515_3	M	SPX	Blood	Whole chr Y	Male, high chr Y intensity	47,XYY	XYY syndrome, <i>de novo</i> , confirmed by karyotype
Other	abnormalitie	es resi	ulting fro	om cell line	artifacts or not	validated in blood		
1	5010_3	M	MPX	CL	1q43-qter DEL, 9q13-qter DUP	1q43-q44 low intensity & BAF split; 9q arm high intensity & little BAF split	47,XY	1q43-qter 9.4 Mb DEL, mosaic (1:237816283- 247249719), 9q13-qter 69.8 Mb DUP, mosaic (9:70400178-140273252); found in the same DNA batch by Affy 500K-EA; normal karyotype; parents normal; AffyCytoScanHD on blood DNA showed normal results: cell line artifact
1	5321_3	М	SPX	CL	Whole chr 4	Run of homozygosity entire chr 4	N/A	Uniparental disomy chr 4, confirmed in cell line; no blood DNA available so we can't exclude a cell line artifact
1	6379_4	М	SPX	CL	Whole chr 14	High intensity, BAF split	46,XY	Whole chr 14 duplication by one algorithm only; karyotype excluded a chr14 trisomy: cell line artifact

Samples that passed QC filters but showed CNVs by any algorithm larger than 7.5 Mb, long runs of homozygosity, or CNVs encompassing/or close to the centromere on any of the autosomes or chromosome X were further inspected manually by plotting their log2 ratio intensities as well as allelic genotype ratios. A cutoff of >7.5 Mb was selected to be consistent with large cytogenetically visible chromosome abnormalities. Samples containing such alterations were excluded from the main burden analyses, except for two AGP probands with XYY syndrome that were retained since chromosome Y markers were not used in the CNV analysis, and two probands with uniparental disomy (proband 2175\_1 with a segmental uniparental disomy of chr 22q was excluded during QC because of excessive number of calls).

Abbreviations: BAF, B allele frequency; CL, cell line; DEL, deletion; DUP, duplication; EA: early access (Affy500K-EA vs. Affy500K); F, female; LRR; log R ratio; M, male; MPX, multiplex; N/A, not available; SPX, simplex; UNK, unknown family type (extended family not evaluated for ASD).

**Table S2A. Sample characteristics** 

Classification		Sex	<b>Stage 1</b> (n=1087)			<b>Stage 2</b> (new cases, n=1359)		Combined (n=2446)	
			European n=979 <sup>a</sup>	Other n=108	European n=1168	Other n=191	European n=2147	Other n=299	
ASD <sup>b</sup>		М	826	90	1027	171	1853	261	
		F	153	18	141	20	294	38	
Developmental	Without DI	М	433	51	569	65	1002	116	
impairment <sup>c</sup>		F	67	11	63	9	130	20	
	With DI	М	361	35	413	103	774	138	
		F	84	7	72	11	156	18	
Family type <sup>d</sup>	Simplex	М	366	33	660	132	1026	165	
		F	62	3	78	14	140	17	
	Multiplex M	М	330	53	208	30	538	83	
		F	59	14	43	5	102	19	
	Unknown	М	131	4	159	9	290	13	
		F	31	1	20	1	51	2	

<sup>&</sup>lt;sup>a</sup> 17 of the 996 (1.7%) European cases used in Pinto et al. <sup>117</sup> were excluded from the combined sample after applying additional OC steps.

The ratio of simplex to multiplex families was 1.82 (1348:742), with 356 (14.5%) families of unknown status. The ratio of males to females was 6.4:1; 46% of cases (n=1,086 out of 2,354) showed developmental impairment (DI). There were 1.43 times more females than males with DI compared to no-DI (M:F for DI is 5.2 vs. 7.45 for no-DI, 95% CI 1.12-1.82, chi square p=0.003). When considering family type, there were 1.47 times more males than females in simplex compared to multiplex families (95% CI 1.13-1.91, chi square p=0.0039), and simplex cases had 1.57 fold more DI compared to multiplex cases (95% CI 1.30-1.90, chi-square p=0.004).

 Simplex males: 1026+165=1191
 Multiplex males: 538+83=621

 Simplex females: 140+17=157
 Multiplex females: 102+19=121

 Simplex M:F ratio: 1191/157= 7.59:1
 Multiplex M:F ratio: 621/121= 5.13:1

b Subjects met criteria for strict autism or autism spectrum according to the ADI-R and/or ADOS (see Methods for detailed description).

<sup>&</sup>lt;sup>c</sup> Developmental impairment is a hierarchical classification based on scores on full-scale IQ, performance IQ, verbal IQ and the Vineland Adaptive Behavior Scales composite score (see Methods for detailed description). A cut-off score of 70 was applied on all measures. Some data are missing on this item, ranging from 2%-4%.

d Family-history reports were taken to inform on the family type. Multiplex families had at least two individuals receiving validated ASD diagnoses who were first to third degree relatives (for third degree, only cousins were considered). Simplex families had only one known individual with ASD in first to third (cousin) degree relatives. Families that did not fall into the multiplex or simplex criteria above were classified as unknown.

# Table S2B. Sample characteristics (continued)

### Cases

		European	European		
	European	Males	Females	Other	Total
Stage 1	979	826	153	108	1,087
Stage 2	1,168	1,027	141	191	1,359
Stage 1 + 2 (all cases)	2,147	1,853	294	299	2,446

## Cases, breakdowns

	Stage 1+2 Males		Stage 1+2 Females		Stage 1+2 All	
	All ancestries	European	All ancestries	European	All ancestries	European
Single probands	92	88	14	14	106	102
Proband+mother duo	101	83	25	23	126	106
Proband+father duo	120	104	17	15	137	119
Complete trios	1,801	1,578	276	242	2,077	1,820
Total probands	2,114	1,853	332	294	2,446	2,147

### **Controls**

		European	European
PRIMARY controls	European	Males	Females
Stage 1: 1166 SAGE	1,166	370	796
Stage 2: 1240 HABC + 234 OC	1,474	871	603
Totals	2,640	1,241	1,399

## **Complete Trios**

		European	European			
	European	Males	Females	Other	Total (1)	Total (2)
Stage 1	862	733	129	91	953	n/a
Stage 2	958	845	113	166	1,124	n/a
Stage 1 + 2 (all trios)	1,820	1,578	242	257	2,077	2,096

<sup>(1)</sup> trios with array data after QC available from both parents

## Cases with rare CNVs >30 kb

European	European	European
(all)	(genic)*	(exonic)
868	747	691
1,046	871	795
1,914	1,618	1,486
2,359	1,971	1,820
4,273	3,589	3,306
	(all) 868 1,046 1,914 2,359	(all) (genic)* 868 747 1,046 871 1,914 1,618 2,359 1,971

<sup>\*</sup> with 10 kb-flanking

### **Additional controls**

		Other	Other
	Other	ancestries	ancestries
SECONDARY controls	ancestries	Males	Females
Stage 1: SAGE	517	185	332
Stage 2: HABC	1,326	596	730
Totals	1,843	781	1,062
Controls All ancestries - totals	4,483	2,022	2,461

<sup>(2)</sup> trios in (1) plus additional trios with laboratory validation data from both parents; n/a: not applicable

### Tables S3A-S3C. CNV burden

# **Table S3A. Platform comparison**

### **CNV** rate

Туре

All DEL DUP

DEL

DUP

Туре

All

DEL

DUP

All

DEL

DUP

#### 1M single: 1382 cases / 1400 controls

			1382 case	es / 140	0 controls
Group		# rare	Baseline	Case/	P value
		CNVs	CNV rate	Ctrl	
			(Ctrl)	ratio	
All	•	6,506	2.32	1.02	0.27621
All		3,353	1.17	1.07	0.03401
All		3,153	1.15	0.96	0.83521
30 – 500 kb		6,088	2.18	1.01	0.39300
≥ 500 kb		418	0.14	1.15	0.09388
≥ 1 Mb		127	0.04	1.24	0.13127
30 – 500 kb		3,235	1.12	1.07	0.03287
≥ 500 kb		118	0.04	1.01	0.50979
≥1 Mb		42	0.01	1.65	0.08491
30 – 500 kb		2,853	1.06	0.94	0.93625
≥ 500 kb		300	0.10	1.20	0.06567
≥ 1 Mb		85	0.03	1.09	0.39456

### 1M duo: 765 cases / 1240 controls

# rare	Baseline	Case/	P value
CNVs	CNV rate	Ctrl	
	(Ctrl)	ratio	
4,407	2.16	1.05	0.06357
2,248	1.10	1.05	0.13466
2,159	1.06	1.05	0.14956
4,201	2.08	1.02	0.25742
206	0.08	1.79	0.00005
76	0.03	2.00	0.00214
2,187	1.08	1.02	0.34081
61	0.02	2.87	0.00005
24	0.00	4.86	0.00026
2,014	1.00	1.02	0.31434
145	0.06	1.47	0.01390
52	0.02	1.39	0.15035

### Total CNV size (kb)

Group

ΑII

ΑII

Αll

30 – 500 kb ≥ 500 kb ≥ 1 Mb

30 – 500 kb ≥ 500 kb ≥ 1 Mb

30 – 500 kb ≥ 500 kb ≥ 1 Mb

1M single: 1382 cases / 1400 controls

1302 Case	3 / 140	o controls
Baseline	Case/	P value
size (Ctrl)	Ctrl	
	ratio	
442.7	1.10	0.03171
222.2	1.12	0.06035
362.8	1.08	0.10991
298.9	0.99	0.56721
995.1	1.24	0.00201
1,621.0	1.27	0.00651
167.9	1.02	0.28813
948.0	1.45	0.00841
1,551.0	1.51	0.02177
233.9	0.98	0.72926
985.7	1.12	0.08485
1,649.0	1.11	0.15156

### 1M duo: 765 cases / 1240 controls

Baseline	Case/	P value
size (Ctrl)	Ctrl	
	ratio	
365.1	1.39	0.00001
190.1	1.35	0.00011
305.1	1.41	0.00005
285.8	1.06	0.07551
998.7	1.44	0.00534
1,519.0	1.59	0.00514
168.7	0.97	0.70923
964.5	1.40	0.08070
1,411.0	1.42	0.12728
223.3	1.11	0.01821
980.7	1.45	0.01585
1,487.0	1.77	0.00600

#### **Gene count**

### 1M single: 1382 cases / 1400 controls

1M duo:
765 cases / 1240 controls

Туре	Group		# Rare	# Genes	Baseline	Case/	P value	Pcorr	•	# Rare	# Genes	Baseline	Case/	P value	Pcorr
			genic	inters.	gene rate	Ctrl				genic	inters.	gene rate	Ctrl		
			CNVs	by rare	(Ctrl)	gene				CNVs	by rare	(Ctrl)	gene		
				CNVs		ratio					CNVs		ratio		
All	All	_	4,180	5,210	4.11	1.22	0.00140	0.00120		2,681	3,587	3.12	1.64	0.00001	0.00001
DEL	All		1,806	2,040	1.37	1.33	0.00122	0.01068		1,164	1,469	1.16	1.40	0.00146	0.05123
DUP	All	_	2,374	4,002	2.74	1.17	0.03831	0.00824		1,517	2,597	1.95	1.78	0.00001	0.00004
All	30 – 500 kb		3,800	3,795	3.12	1.03	0.30338	0.33000		2,496	2,780	2.71	1.12	0.02734	0.02645
	≥ 500 kb		380	2,011	0.99	1.84	0.00034	0.00234		185	1,039	0.41	5.07	0.00001	0.00019
	≥ 1 Mb	_	122	1,075	0.37	2.65	0.00096	0.00231		71	641	0.22	7.10	0.00001	0.00329
DEL	30 – 500 kb		1,700	1,407	1.09	1.15	0.02952	0.10020		1,113	1,028	1.08	0.92	0.84269	0.86370
	≥ 500 kb		106	732	0.28	2.05	0.00992	0.01546		51	485	0.08	7.80	0.00001	0.00589
	≥ 1 Mb	_	41	504	0.13	2.80	0.01122	0.11560		23	339	0.04	10.00	0.00005	0.05489
DUP	30 – 500 kb		2,100	2,849	2.03	0.96	0.70648	0.43300		1,383	2,045	1.63	1.26	0.00239	0.00397
	≥ 500 kb		274	1,561	0.72	1.76	0.00545	0.01727		134	697	0.33	4.38	0.00008	0.01109
	≥ 1 Mb	_	81	721	0.24	2.57	0.01518	0.00677		48	391	0.17	6.38	0.00086	0.08588

Baseline CNV rate (Ctrl): average number of CNVs per control subject

Baseline size (Ctrl): average total size per control subject in kb

Baseline gene rate (Ctrl): average number of genes intersected by CNVs per control subject

Pcorr: corrected for global differences in CNV size and rate

100,000 permutations

Global burden analyses for rare CNVs were performed using PLINK v1.0730, R stats and custom scripts, as previously described in detail. 117 We tested for global increased burden in a combined set of 2,147 European ASD cases compared to 2,640 European controls for three measures: CNV rate, CNV size (Tables S3A-S3B) and the average number of genes affected by CNVs (gene-count) (Table 1, main text). We observed a significantly increased burden in the number of genes affected by rare CNVs in cases over controls (Table 1). This enrichment for rare genic CNVs was apparent for both deletions and duplications, and remained after we further controlled for potential case-control differences that could be present due to biological differences or technical biases (Table 1). Similar findings were obtained when data were broken down by array type (Table S3A), or when each stage was considered separately (Table S3B).

## Table S3B. Stage 1 (Pinto et al.) versus Stage 2 (new cases) versus Combined (all 2147 **European cases**)

#### **CNV** rate

Group

ΑII 30 – 500 kb

≥ 500 kb ≥ 1 Mb

30 – 500 kb ≥ 500 kb ≥1Mb

30 - 500 kb ≥ 500 kb ≥1 Mb

Туре

All DUP

All

DEL

DUP

# Stage 1:

979 cases / 1166 controls						
# rare	Baseline	Case/	P value			
CNVs	CNV rate	Ctrl				
	(Ctrl)	ratio				
5,153	2.44	0.97	0.85304			
2,664	1.23	1.02	0.29184			
2,489	1.21	0.91	0.98261			
4,830	2.29	0.96	0.90681			
323	0.14	1.11	0.19004			
91	0.04	1.02	0.50026			
2,575	1.19	1.03	0.27934			
89	0.04	0.97	0.59513			
27	0.01	1.28	0.32376			
2,255	1.11	0.89	0.99523			
234	0.10	1.17	0.13809			
64	0.03	0.93	0.66292			

#### Stage 2: 1168 cases / 1474 controls

		•	
# rare	Baseline	Case/	P value
CNVs	CNV rate	Ctrl	
	(Ctrl)	ratio	
5,923	2.16	1.09	0.00066
3,063	1.11	1.09	0.00554
2,860	1.05	1.08	0.02640
5,634	2.08	1.06	0.01827
289	0.08	1.88	0.00002
106	0.02	2.45	0.00003
2,978	1.09	1.07	0.03504
85	0.02	2.71	0.00002
38	0.01	4.73	0.00005
2,656	0.99	1.05	0.13460
204	0.06	1.63	0.00037
68	0.02	1.80	0.01192

#### Combined stage 1 + stage 2: 2147 cases / 2640 controls

۷.	LT/ Cases /	2040	COILLIOIS
# rare	Baseline	Case/	P value
CNVs	CNV rate	Ctrl	
	(Ctrl)	ratio	
11,076	2.28	1.03	0.05804
5,727	1.16	1.06	0.01238
5,349	1.12	1.00	0.49532
10,464	2.17	1.01	0.27705
612	0.11	1.43	0.00003
197	0.03	1.62	0.00066
5,553	1.14	1.05	0.04012
174	0.03	1.59	0.00173
65	0.01	2.58	0.00024
4,911	1.04	0.97	0.82515
438	0.08	1.37	0.00084
132	0.02	1.31	0.07407

#### Total CNV size (kb)

Туре	Group
All	All
DEL	All
DUP	All
All	30 – 500 kb
	≥ 500 kb
	≥ 1 Mb
DEL	30 – 500 kb
	≥ 500 kb
	≥ 1 Mb
DUP	30 – 500 kb
	≥ 500 kb

Stage 1:						
979 cases	/ 1166 control					

Baseline	Case/	P value
size (Ctrl)	Ctrl	
	ratio	
453.1	1.03	0.26520
222.5	1.02	0.40872
365.2	1.06	0.20712
304.4	0.97	0.78403
1009.0	1.14	0.04687
1632.0	1.18	0.04903
168.3	0.99	0.56454
952.3	1.12	0.19717
1560.0	1.17	0.16605
233.9	0.97	0.75751
997.0	1.11	0.13966
1658.0	1.15	0.14467

Stage 2: 1168 cases / 1474 controls

Case/	P value
Ctrl	
ratio	
1.41	0.00001
1.38	0.00006
1.36	0.00001
1.06	0.04013
1.49	0.00027
1.54	0.00144
0.99	0.63485
1.64	0.01571
1.61	0.03369
1.08	0.03083
1.37	0.00872
1.52	0.00919
	1.41 1.38 1.36 1.06 1.49 1.54 0.99 1.64 1.61

Combined stage 1 + stage 2: 2147 cases / 2640 controls

Baseline	Case/	P value
size (Ctrl)	Ctrl	
	ratio	
406	1.22	0.00001
207.9	1.21	0.00028
333.3	1.21	0.00007
296.8	1.02	0.22737
990.1	1.32	0.00005
1,591.0	1.38	0.00013
171.2	0.99	0.64394
957.4	1.42	0.00371
1,531.0	1.44	0.01435
229.7	1.03	0.18388
972.4	1.24	0.00429
1,586.0	1.34	0.00380

### **Gene count**

Туре

All

DEL

DUP

≥ 1 Mb

Group

ΑII

ΑII

ΑII

30 – 500 kb

30 - 500 kb

30 - 500 kb

≥ 500 kb ≥1Mb

≥ 500 kb

 $\geq 1 \, \text{Mb}$ 

≥ 500 kb

≥ 1 Mb

Stage 1: 979 cases / 1166 controls Case/

Ctrl

gene

ratio

1.17

1.25

0.98

2.39

1.16

1.63

0.9

P value

0.02076

0.01603

0.60840

0.01245

0.03994 0.09459

0.08343

0.92581

0.00462

0.01603

0.00648

0.08104

0.13570

0.62180

189

Stage 2:							
1168 cases /	1474 controls						

# Rare	# Genes	Baseline	Case/	P value	Pcorr			
genic	inters.	gene rate	Ctrl					
CNVs	by rare	(Ctrl)	gene					
	CNVs		ratio					
3,607	4,514	3.08	1.66	0.00001	0.00001			
1,568	1,924	1.16	1.55	0.00001	0.01498			
2,039	3,244	1.92	1.73	0.00001	0.00001			
3,345	3,414	2.70	1.15	0.00384	0.02582			
262	1,485	0.38	5.29	0.00001	0.00001			
101	865	0.17	8.12	0.00001	0.00001			
1,495	1,296	1.06	0.99	0.55163	0.79270			
73	696	0.09	7.91	0.00001	0.00051			
37	492	0.07	7.84	0.00002	0.10500			
1,850	2,491	1.63	1.26	0.00094	0.00364			

8.30

0.00001 0.00026

### Combined stage 1 + stage 2: 2147 cases / 2640 controls

	# Rare	# Genes	Baseline	Case/	P value	Pcorr
	genic	inters.	gene rate	Ctrl		
	CNVs	by rare	(Ctrl)	gene		
		CNVs		ratio		
	6,859	6,745	3.55	1.41	0.00001	0.00001
	2,946	2,804	1.23	1.40	0.00001	0.00049
_	3,913	5,217	2.32	1.41	0.00001	0.00001
	6,307	5,163	2.89	1.07	0.03752	0.03628
	552	2,491	0.66	2.88	0.00001	0.00001
_	187	1,337	0.26	4.48	0.00001	0.00001
	2,795	2,014	1.07	1.07	0.11389	0.20110
	151	947	0.16	3.60	0.00001	0.00051
	63	647	0.08	4.58	0.00004	0.02289
_	3,512	3,934	1.83	1.08	0.08690	0.03750
	401	1,896	0.50	2.64	0.00001	0.00026
_	124	890	0.18	4.43	0.00004	0.00036

Baseline CNV rate (Ctrl): average number of CNVs per control subject

Baseline size (Ctrl): average total size per control subject in kb

# Rare

genic

CNVs

3,252

1,378

1.874

2,962

1.300

1,662

212

86

78

26

Baseline gene rate (Ctrl): average number of genes intersected by CNVs per control subject

# Genes Baseline

inters, gene rate

(Ctrl)

4.16

1.32

3.14

0.38

0.25

0.11

2.07

by rare

CNVs

4,319

1,505

3.417

3,211

805

454

278

2,411

1,325

632

Pcorr: corrected for global differences in CNV size and rate

100,000 permutations

979

Table S3C. Characteristics of rare CNVs in 2,147 European ASD probands and 2,640 European controls

		ASD probands,	European			Controls, European			
	(n = 2,147 probands; 1,838 trios)					(n = 2,640)			
-	Stringent CNVs <sup>a</sup> (all sizes)	Stringent CNVs <sup>b</sup> (≥30 kb size, no pericentromeric + no segdup)	Rare CNVs <sup>c</sup>	Rare <i>de novo</i> CNVs <sup>d</sup>	Stringent CNVs <sup>a</sup> (all sizes)	Stringent CNV <sup>b</sup> (≥30 kb size, no pericentromeric + no segdup)	Rare CNVs <sup>c</sup>		
Samples	2,147	2,147	1,941	86	2,640	2,640	2,359		
# CNVs	36,034	15,423	5,054	89	46,101	20,341	6,022		
Mean / median CNVs per genome <sup>e</sup>	16.8 / 16	7.18 / 7	2.64 / 2	1.03 / 1	17.5 / 17	7.70 / 8	2.55 / 2		
Mean / median CNV size (kb)	107.0 / 44.8	162.5 / 94.5	187.9 / 88.4	1,244.5 / 477.4	91.0 / 39.3	150.6 / 89.9	159.1 / 87.0		
% Gain / loss	26.4 / 73.6	39.6 / 60.4	47.9 / 52.1	33.7 / 66.3	24.8 / 75.2	36.9 / 63.1	49.3 / 50.7		
# Recurrent/overlapping CNVs (%) / # loci f	33,803 (93.8%) / 1,536	13,812 (89.5%) / 912	3,431 (67.9%) / 822	34 (38.2%) / 11	43,905 (95.2%) / 1,613	18,641 (91.6%) / 1,019	4,321 (71.8%) / 936		
# CNVs >1 Mb (%)	365 (1.0%)	277 (1.8%)	112 (2.2%)	28 (31.5%)	264 (0.6%)	226 (1.1%)	85 (1.4%)		
# CNVs >100-999 kb (%)	9,782 (27.1%)	7,019 (45.5%)	2,445 (48.4%)	42 (47.2%)	11,945 (25.9%)	9,341 (45.9%)	2,537 (42.1%)		

<sup>&</sup>lt;sup>a</sup> Stringent CNVs are CNV called by two or more algorithms. CNVs detected in the same individual by at least two algorithms were merged with the outside probes used as boundaries. All sizes, no filter applied.

b Stringent CNVs ≥30 kb and filtered for pericentromeric calls as well as calls overlapped by segmental duplications (segdup) for >50% of their length.

<sup>&</sup>lt;sup>c</sup> Rare stringent CNVs ≥30 kb (filtered for pericentromeric and segdup calls) present at a frequency <1% in the total sample of 2,147 European cases and 2,640 European controls.

Inheritance state was estimated for CNVs detected in 1,838 European probands from complete trios, including 1,820 complete trios with array data after QC available from both parents plus 18 families in which at least one of the parents failed initial array QC but additional laboratory validation was obtained for both parents. Laboratory validation confirmed that at least 4.7% (86/1,838) of European families carried at least one *de novo* CNV (average of 1 verified *de novo* CNV/sample). Similar rates were obtained for families of all ancestries, with 4.7% (98/2,096) carrying at least one *de novo* event.

e Probands with CNVs larger than 7.5 Mb are listed in **Table S1C**; they were excluded from the main burden analyses.

f Number and percentage of recurrent and/or overlapping CNVs in the dataset (%), and corresponding number of CNV loci.

Table S4. Rare *de novo* CNVs in probands confirmed experimentally

14310_4270   M	Sample ID	Sex	Ancestry	CNV type	Chr#:start-end (hg18)	Size	Cytoband	RefSeq gene(s)	Location
1003; 1571033001 M Eur Gain 11444809343-146252557 1,842,025 1,921.1 1 5; genes [industre COPU], GAS, GAIS] genes [industre COPU], GAIS, GAIS [industre COPU], GAIS, GAI	14310_4270	М	Eur		1:105623065-105795181	172,117	1p21.1	_	intergenic
1992 301 M OBER GAIN 1.1485008F3.46833872 1.288.2594 12.11 1.55 genes (Includes CPUL, CAS, GAMS) genes (Includes STORE, CAS, GAMS) (Includes S	14310_4270	М	Eur	Gain	1:105896243-106480692	584,450	1p21.1	_	intergenic
1835   201   M	16035_1571013001	М	Eur	Gain	1:144482933-146325557	1,842,625	1q21.1	15 genes (includes CHD1L, GJA5, GJA8)	gene/exonic
1315.15123   F	1952_301	М	Other	Gain	1:144500467-146336720	1,836,254	1q21.1	15 genes (includes CHD1L, GJA5, GJA8)	gene/exonic
978.05. M Eur	3635_201	М	Eur	Loss	1:144500467-146377870	1,877,404	1q21.1	17 genes (includes CHD1L, GJA5, GJA8)	gene/exonic
See 201   F	13135_1523	F	Eur	Gain	1:144838594-146308287	1,469,694	1q21.1	14 genes (includes CHD1L, GJA5, GJA8)	gene/exonic
### 1887_20.0 M Other Gain 1.98247385-99645550 1.988_206 1.921.39_21.2 LOCIOLIZEGO, IPPRA, IPPRS, MIRLST, SNAT	6356_5		Eur	Loss	1:2118508-2325536	207,029		2 MORN1, SKI, LOC100129534, RER1, C1orf86	gene/exonic
1.000   1.00	8658_201	F	Eur	Loss	1:98175622-100923952	2,748,331	1p21.3-p21.2	18 genes (includes CDC14A, MIR137, RTCD1, SNX7)	gene/exonic
13082_963	9877_204	М	Other	Gain	1:98247355-99645560	1,398,206	1p21.3-p21.2	LOC100129620, LPPR4, LPPR5, MIR137, SNX7	gene/exonic
1220_1	5089_5	М	Other	Loss	2:102292943-102345460	52,518	2q12.1	IL1RL1	gene/exonic
13017_353	13082_963	М	Eur	Gain	2:11712589-11741036	28,448	2p25.1	NTSR2	gene/exonic
14414_5230	4260_1	М	Eur	Gain	2:119603876-119882278	278,403	2q14.2	C1QL2, C2orf76, DBI, STEAP3	gene/exonic
14068_1180	13027_353	F	Eur	Gain	2:144837809-145315383	477,575	2q22.3	DKFZp686O1327, ZEB2	gene/exonic
13017_223	14414_5230	М	Eur	Loss	2:230486629-230547253	60,625	2q36.3	FBXO36, TRIP12	gene/exonic
1321E_2883	14068_1180	М	Eur	Gain	2:50493827-50677835	184,009	2p16.3	NRXN1	gene/exonic
13153_1703 M Eur Loss 2-50990306-5122043 231,738 2p16.3 NRXM1 gene/ex   13037_463 M Eur Loss 2-5100276-51157742 155.167 2p16.3 NRXM1 gene/ex   13037_463 M Eur Loss 3-93474320-9498562 24,043 3p25.3 SFTD5 gene/ex   13037_463 M Eur Loss 3-194720-9498562 24,043 3p25.3 SFTD5 gene/ex   13174.3 M Eur Loss 3-19212098-1960299 1512,302 3p24.3 CAPR   13174.3 M Eur Loss 3-19212098-1960299 1712,302 3p24.3 CAPR   13174.3 M Eur Loss 3-19212061-20096322 175,772 3p24.3 CAPR   13174.5 M Eur Loss 3-1192780699-140127703 366.808 3p22.3 CEPPO, FAIM, PRIXCB   13174.5 M Eur Loss 3-1192780699-140127703 366.808 3p22.3 CEPPO, FAIM, PRIXCB   13174.5 M Eur Loss 3-1195971510-197675831 1,704,322 3p29 20 genes (includes PCYT1A, TFRC, TNK2) gene/ex   13587_210 M Eur Loss 3-1195971510-197675831 1,704,322 3p29 20 genes (includes PCYT1A, TFRC, TNK2) gene/ex   13587_210 M Other Gain 4-94659049-95220967 630,619 4q22.2 ATOH1, GRID2   13588817-019675821 1,704,322 3p29 20 genes (includes PCYT1A, TFRC, TNK2) gene/ex   13587_210 M Eur Loss 5-88888170-98505590 917,340 5q14.3 MR3660   13588612 DIN M Eur Loss 6-1656785155-1584989874 1,704,720 6q25.3 AMB3660   1358612 DIN M Eur Loss 6-1656783155-1584989874 1,704,720 6q25.3 AMB3660   13587_210 M Eur Loss 6-16567831575-1584989874 1,704,720 6q25.3 AMB3660   13587_210 M Eur Loss 6-165023074-160081618 58,545 6q25.3 SOD2, WTAP   13583_3	13017_223	F	Eur	Loss	2:50539877-50730546	190,670	2p16.3	NRXN1	gene/exonic
13037_463   M   Eur   Loss   2-51002576-51157742   155,167   2p16.3   NRXWI   gene/ew   Sene/ew   Sene/e	13216_2383	М	Eur	Loss	2:50968208-51214171	245,964	2p16.3	NRXN1	gene/exonic
1816_3	13153_1703	M	Eur	Loss	2:50990306-51222043	231,738	2p16.3	NRXN1	gene/exonic
F	13037_463	M	Eur	Loss	2:51002576-51157742	155,167	2p16.3	NRXN1	gene/exonic
STATE   STAT	3616_3	М	Eur	Loss	3:9474320-9498362	24,043	3p25.3	SETD5	gene/exonic
SEAST_210   F   Eur	5220_3	F	Eur	Gain	3:19127998-19640299	512,302	3p24.3	KCNH8	gene/exonic
\$587_210	3174_3	М	Eur	Loss	3:19921061-20096832	175,772	3p24.3	C3orf48, EFHB, KAT2B, RAB5A	gene/exonic
8588_201         M         Eur         Loss         3:195971510-197675831         1,704,322         3q.29         20 genes (includes PCYTIA, TRRC, TNK2)         gene/exe           8677_201         M         Other         Gain         4:94659049-95289667         630,619         4g.22.2         ATOHI, GRID2         Gene/exe           8709_201         F         Eur         Loss         5:88588170-89505509         917,340         5q14.3         MIR3660         gene/exe           8709_201         F         Eur         Gain         6:160738751505158489874         1,704,720         6q25.3         AMIDIB, MIR3692, SERACI, SWX9, SYNU2, ZDHHC14         gene/exe           8614_201         M         Eur         Loss         6:1629388879-162629938         41,060         6q25.3         AMIDIB, MIR3692, SERACI, SWX9, SYNU2, ZDHHC14         gene/exe           8604_201         M         Eur         Loss         6:16936788-170761395         1,624,603         6q27         14 genes (includes PSMBI, TBP, TCTE3, THBS2, gene/exe         gene/exe           8353_3         F         Eur         Loss         6:3635297-7863715199         1,624,603         6q27         14 genes (includes PSMBI, TBP, TCTE3, THBS2, gene/exe         gene/exe           818133_302         M         Eur         Loss         6:86352	5245_3	M	Eur	Loss	3:117285007-117477191	192,185	3q13.31	LSAMP (50% mosaic)	gene/exonic
March   Color   Colo	8587_210	F	Eur	Gain	3:139760896-140127703	366,808	3q22.3	CEP70, FAIM, PIK3CB	gene/exonic
Separage	8588_201	М	Eur	Loss	3:195971510-197675831	1,704,322	3q29	20 genes (includes PCYT1A, TFRC, TNK2)	gene/exonic
S709_201   F	8677_201	М	Other	Gain	4:94659049-95289667	630,619	4q22.2	ATOH1, GRID2	gene/exonic
Side   3	6402_3	М	Eur	Gain	4:98781694-99627129	845,436	4q22.3-q23	C4orf37, RAP1GDS1, TSPAN5	gene/exonic
Side	8709_201	F	Eur	Loss	5:88588170-89505509	917,340	5q14.3	MIR3660	gene/exonic
8612_201 M Eur Loss 6:162588879-162629938 41,060 6q26 PARK2 gene/ex 8404_201 M Eur Loss 6:169136788-170761395 1,624,608 6q27 14 genes (includes PSMB1, TBP, TCTE3, THBS2, gene/ex 8404_201 M Eur Loss 6:33399849-33512042 112,194 6p21.32 CUTA, KIFC1, PHF1, SYNGAP1 gene/ex 8418133_302 M Eur Gain 6:69540429-73375020 3,834,592 6q12-q13 B3GA712_BA13_CGOrf155_CGOrf57_COL19A1, COL19A1_AM135A_LMBR01_MIR30A_MIR30C2_OFR11, RIMS1_SALMBR01_MIR30A_MIR30C2_OFR11, RIMS1_DAUGA_	5386_3	М	Other	Loss	6:156785155-158489874	1,704,720	6q25.3	ARID1B, MIR3692, SERAC1, SNX9, SYNJ2, ZDHHC14	gene/exonic
Separage	6164_3	F	Eur	Gain	6:160023074-160081618	58,545	6q25.3	SOD2, WTAP	gene/exonic
WORZP    WORZP    Sene/ex   WORZP    Sene/ex	8612_201	М	Eur	Loss	6:162588879-162629938	41,060	6q26	PARK2	gene/exonic
18133   302	8404_201	М	Eur	Loss	6:169136788-170761395	1,624,608	6q27	· ·	gene/exonic
COL9A1, FAMI35A, LMBRD1, MIR30A, MIR30C2, OGRILI, RIMS1, SMAP1   GORRIL, RIMS1, SMAP1   gene/ext	5353_3	F	Eur	Loss	6:33399849-33512042	112,194	6p21.32	CUTA, KIFC1, PHF1, SYNGAP1	gene/exonic
F	18133_302	М	Eur	Gain	6:69540429-73375020	3,834,592	6q12-q13	COL9A1, FAM135A, LMBRD1, MIR30A, MIR30C2,	gene/exonic
Say	6248_3	M	Eur	Loss	6:86352577-86376159	23,583	6q14.3	SNX14, SYNCRIP	gene/exonic
8446_201	1960_301	F	Eur	Loss	7:102699832-102798745	98,914	7q22.1	<u> </u>	gene/exonic
STX1A    S	5370_3	М	Eur	Loss	7:153775586-153844747	69,162	7q36.2	DPP6	gene/exonic
Section								STX1A)	gene/exonic
MIR124-2									gene/exonic
8500_201 M Eur Loss 8:74692898-74967280 274,383 8q21.11 STAU2, UBE2W gene/ext 14237_2650 M Eur Loss 9:103456776-107140273 3,683,498 9q31.1 ABCA1, CYLC2, GRIN3A, LOC286367, NIPSNAP3A, NIPSNAP3B, OR13C2, OR13C3, OR13C4, OR13C5, OR13C8, OR13C9, OR13D1, OR13F1, SLC44A1, SMC2 14417_5260 M Eur Loss 9:117311405-117727764 416,360 9q33.1 C9orf27 (or LINC00474) gene/ext 16259_3 M Eur Loss 9:139516033-140208462 692,430 9q34.12 ABL1 gene/ext 16259_3 M Eur Loss 9:3999606-9631169 231,564 9p23 ARRDC1, C9orf37, CACNA1B, EHMT1, FLI40292 (or EHMT1-IT1), MIRG02, MRPL41, PNPLA7, TUBBP5, WDR85, ZMYND19 16246_4 M Eur Loss 9:98998-334508 235,511 9p24.3 C9orf66, CBWD1, DOCK8, FOXD4 gene/ext 13123_1403 F Eur Loss 9:98998-3682923 3,583,926 9p24.3-p24.2 C9orf66, CBWD1, DMRT1, DMRT2, DMRT3, DOCK8, FOXD4 18172_302 M Other Gain 10:34963905-35536952 573,048 10p11.21 CREM, CUL2, PARD3 gene/ext 1853_2_101 M Eur Loss 10:45633089-51564756 5,931,668 10q11.21- 56 genes (includes CHAT, ERCC6, MAPK8, SLC18A3) gene/ext								MIR124-2	gene/exonic
14237_2650 M Eur Loss 9:103456776-107140273 3,683,498 9q31.1 ABCA1, CYLC2, GRIN3A, LOC286367, NIPSNAP3A, NIPSNAP3B, OR13C2, OR13C3, OR13C4, OR13C5, OR13C8, OR13C9, OR13D1, OR13F1, SLC44A1, SMC2 14417_5260 M Eur Loss 9:132572435-132597937 25,503 9q34.12 ABL1 gene/ext 16259_3 M Eur Loss 9:139516033-140208462 692,430 9q34.3 ARRDC1, C9orf37, CACNA1B, EHMT1, FLJ40292 (or EHMT1-IT1), MIR602, MRPL41, PNPLA7, TUBBP5, WDR85, ZMYND19 gene/ext 163246_4 M Eur Loss 9:9399606-9631169 231,564 9p23 PTPRD gene/ext 16324_4 M Eur Loss 9:98998-334508 235,511 9p24.3 C9orf66, CBWD1, DOCK8, FOXD4 gene/ext 16323_1403 F Eur Loss 9:98998-3682923 3,583,926 9p24.3-p24.2 C9orf66, CBWD1, DMRT1, DMRT2, DMRT3, DOCK8, FOXD4 SMARCA2, VLDLR 18172_302 M Other Gain 10:34963905-35536952 573,048 10p11.21 CREM, CUL2, PARD3 gene/ext 18172_302 M Eur Loss 10:45633089-51564756 5,931,668 10q11.21- 56 genes (includes CHAT, ERCC6, MAPK8, SLC18A3) gene/ext							•		gene/exonic
14417_5260         M         Eur         Loss         9:117311405-117727764         416,360         9q33.1         C9orf27 (or LINC00474)         gene/exc           9756_201         M         Other         Loss         9:132572435-132597937         25,503         9q34.12         ABL1         gene/exc           6259_3         M         Eur         Loss         9:139516033-140208462         692,430         9q34.3         ARRDC1, C9orf37, CACNA1B, EHMT1, FLI40292 (or EHMT1-IT1), MIR602, MRPL41, PNPLA7, TUBBP5, WDR85, ZMYND19         gene/exc           6246_4         M         Eur         Loss         9:9399606-9631169         231,564         9p23         PTPRD         gene/exc           5032_4         M         Eur         Loss         9:98998-334508         235,511         9p24.3         C9orf66, CBWD1, DOCK8, FOXD4         gene/exc           13123_1403         F         Eur         Loss         9:98998-3682923         3,583,926         9p24.3-p24.2         C9orf66, CBWD1, DMR71, DMR72, DMR73, DOCK8, FOXD4, KANK1, KCNV2, KIAA0020, RFX3, SMARCA2, VLDLR           18172_302         M         Other         Gain         10:34963905-35536952         573,048         10p11.21         CREM, CUL2, PARD3         gene/exc           8854_201         M         Eur         Loss         10:45633089-51564756								ABCA1, CYLC2, GRIN3A, LOC286367, NIPSNAP3A,	gene/exonic
9756_201 M Other Loss 9:132572435-132597937 25,503 9q34.12 ABL1 gene/ext 6259_3 M Eur Loss 9:139516033-140208462 692,430 9q34.3 ARRDC1, C9orf37, CACNA1B, EHMT1, FLJ40292 (or EHMT1-IT1), MIR602, MRPL41, PNPLA7, TUBBP5, WDR85, ZMYND19 6246_4 M Eur Loss 9:9399606-9631169 231,564 9p23 PTPRD gene/ext 6303_4 M Eur Loss 9:98998-334508 235,511 9p24.3 C9orf66, CBWD1, DOCK8, FOXD4 gene/ext 63123_1403 F Eur Loss 9:98998-3682923 3,583,926 9p24.3-p24.2 C9orf66, CBWD1, DMRT1, DMRT2, DMRT3, DOCK8, FLJ35024, FOXD4, KANK1, KCNV2, KIAA0020, RFX3, SMARCA2, VLDLR 63172_302 M Other Gain 10:34963905-35536952 573,048 10p11.21 CREM, CUL2, PARD3 gene/ext 6426_4 M Eur Loss 10:45633089-51564756 5,931,668 10q11.21- 56 genes (includes CHAT, ERCC6, MAPK8, SLC18A3) gene/ext	14417 5260	M	Fur	loss	9.117311405-117727764	416 360	9n33 1		2 gene/exonic
6259_3 M Eur Loss 9:139516033-140208462 692,430 9q34.3 ARRDC1, C9orf37, CACNA1B, EHMT1, FLJ40292 (or EHMT1-IT1), MIR602, MRPL41, PNPLA7, TUBBP5, WDR85, ZMYND19 6246_4 M Eur Loss 9:9399606-9631169 231,564 9p23 PTPRD gene/exc 65032_4 M Eur Loss 9:98998-334508 235,511 9p24.3 C9orf66, CBWD1, DOCK8, FOXD4 gene/exc 13123_1403 F Eur Loss 9:98998-3682923 3,583,926 9p24.3-p24.2 C9orf66, CBWD1, DMRT1, DMRT2, DMRT3, DOCK8, FLJ35024, FOXD4, KANK1, KCNV2, KIAA0020, RFX3, SMARCA2, VLDLR 18172_302 M Other Gain 10:34963905-35536952 573,048 10p11.21 CREM, CUL2, PARD3 gene/exc 8534_201 M Eur Loss 10:45633089-51564756 5,931,668 10q11.21- 56 genes (includes CHAT, ERCC6, MAPK8, SLC18A3) gene/exc									gene/exonic
6246_4 M Eur Loss 9:9399606-9631169 231,564 9p23 PTPRD gene/ext 5032_4 M Eur Loss 9:98998-334508 235,511 9p24.3 C9orf66, CBWD1, DOCK8, FOXD4 gene/ext 13123_1403 F Eur Loss 9:98998-3682923 3,583,926 9p24.3-p24.2 C9orf66, CBWD1, DMRT1, DMRT2, DMRT3, DOCK8, gene/ext 18172_302 M Other Gain 10:34963905-35536952 573,048 10p11.21 CREM, CUL2, PARD3 gene/ext 18172_302 M Eur Loss 10:45633089-51564756 5,931,668 10q11.21- 56 genes (includes CHAT, ERCC6, MAPK8, SLC18A3) gene/ext								ARRDC1, C9orf37, CACNA1B, EHMT1, FLJ40292 (or EHMT1-IT1), MIR602, MRPL41, PNPLA7, TUBBP5,	gene/exonic
5032_4 M Eur Loss 9:98998-334508 235,511 9p24.3 <i>C9orf66, CBWD1, DOCK8, FOXD4</i> gene/ext 13123_1403 F Eur Loss 9:98998-3682923 3,583,926 9p24.3-p24.2 <i>C9orf66, CBWD1, DMRT1, DMRT2, DMRT3, DOCK8, gene/ext FLJ35024, FOXD4, KANK1, KCNV2, KIAA0020, RFX3, SMARCA2, VLDLR</i> 18172_302 M Other Gain 10:34963905-35536952 573,048 10p11.21 <i>CREM, CUL2, PARD3</i> gene/ext 8534_201 M Eur Loss 10:45633089-51564756 5,931,668 10q11.21- 56 genes (includes <i>CHAT, ERCC6, MAPK8, SLC18A3</i> ) gene/ext	6246_4	М	Eur	Loss	9:9399606-9631169	231,564	9p23		gene/exonic
13123_1403 F Eur Loss 9:98998-3682923 3,583,926 9p24.3-p24.2 C90rf66, CBWD1, DMRT1, DMRT2, DMRT3, DOCK8, gene/exc FLJ35024, FOXD4, KANK1, KCNV2, KIAA0020, RFX3, SMARCA2, VLDLR 18172_302 M Other Gain 10:34963905-35536952 573,048 10p11.21 CREM, CUL2, PARD3 gene/exc 8534_201 M Eur Loss 10:45633089-51564756 5,931,668 10q11.21- 56 genes (includes CHAT, ERCC6, MAPK8, SLC18A3) gene/exc					9:98998-334508				gene/exonic
18172_302 M Other Gain 10:34963905-35536952 573,048 10p11.21 <i>CREM, CUL2, PARD3</i> gene/exc 8534_201 M Eur Loss 10:45633089-51564756 5,931,668 10q11.21- 56 genes (includes <i>CHAT, ERCC6, MAPK8, SLC18A3</i> ) gene/exc								C9orf66, CBWD1, DMRT1, DMRT2, DMRT3, DOCK8, FLJ35024, FOXD4, KANK1, KCNV2, KIAA0020, RFX3,	
	18172_302	М	Other	Gain	10:34963905-35536952	573,048	10p11.21		gene/exonic
	8534_201	М	Eur	Loss	10:45633089-51564756	5,931,668	10q11.21-	56 genes (includes CHAT, ERCC6, MAPK8, SLC18A3)	gene/exonic
							q11.23		

Sample ID	Sex	Ancestry	CNV type	Chr#:start-end (hg18)	Size	Cytoband	RefSeq gene(s)	Location
14393_5020	М	Eur	Loss	10:74801551-75278951	477,401	10q22.2	AGAP5, ANXA7, BMS1P4, CAMK2G, CHCHD1, FUT11, KIAA0913, MYOZ1, NDST2, PPP3CB, SEC24C, SYNPO2L, USP54, ZMYND17	gene/exonic
6240_4	М	Eur	Loss	11:126633939-132060374	5,426,436	11q24.2-q25	20 genes (includes ARHGAP32, SNX19)	gene/exonic
6325_3	М	Eur	Loss	11:70077507-70506315	428,809	11q13.3	MIR3664, SHANK2	gene/exonic
6319_3	М	Eur	Loss	11:70119917-70187872	67,956	11q13.3	SHANK2	gene/exonic
5237_3	М	Eur	Loss	11:70154458-70220632	66,175	11q13.3	SHANK2	gene/exonic
6053_3	М	Eur	Gain	12:54218922-58779615	4,560,694	12q13.2-q14.1	101 genes (includes CDK2, CDK4, SMARCC2)	gene/exonic
5272_3	М	Other	Loss	12:98445422-98540678	95,257	12q23.1	ANKS1B	gene/intronic
1050_3	F	Eur	Gain	14:20279711-20345174	65,464	14q11.2	EDDM3A, EDDM3B, RNASE1, RNASE6	gene/exonic
8638_201	М	Eur	Loss	14:35203374-35369853	166,480	14q13.2	BRMS1L, RALGAPA1	gene/exonic
4272_1	М	Eur	Loss	14:63824114-65347410	1,523,297	14q23.2-q23.3	18 genes (includes ESR2, MAX, MTHFD1, SPTB)	gene/exonic
17009_1	М	Eur	Loss	14:78575296-78596793	21,498	14q31.1	NRXN3	gene/intronic
20187_1464001	М	Eur	Gain	15:18811937-26209270	7,397,334	15q11.2-q13.1	125 genes (includes CYFIP1, UBE3A, HERC2)	gene/exonic
8630_201	М	Eur	Gain	15:19800798-26209270	6,408,473	15q11.2-q13.1		gene/exonic
20069_1328001	М	Eur	Gain	15:20203578-26209270	6,005,693		112 genes (includes CYFIP1, UBE3A, HERC2)	gene/exonic
17035 1	F	Eur	Gain	15:20274130-26120360	5,846,231		111 genes (includes CYFIP1, UBE3A, HERC2)	gene/exonic
8430_204	M	Other	Loss	15:20301669-20777695	476,027	15q11.2	CYFIP1, NIPA1, NIPA2, TUBGCP5, WHAMML1	gene/exonic
13050 593	M	Eur	Gain	15:21190624-26203954	5,013,331	•	101 genes (includes <i>UBE3A</i> , <i>HERC2</i> )	gene/exonic
16040_1571029001		Eur	Loss	15:28450423-30303265	1,852,843		ARHGAP11B, CHRFAM7A, CHRNA7, FAM7A1, FAM7A2, FAN1, KLF13, LOC100288637, MIR211, MTMR10, OTUD7A, TRPM1	gene/exonic
6101 4	М	Other	Loss	15:74735339-74929817	194,479	15q24.3	SCAPER	gene/exonic
8695_201	М	Eur	Loss	15:81728085-82623936	895,852	15q25.2	ADAMTSL3, BNC1, LOC648809, SH3GL3	gene/exonic
14181 2940	М	Eur	Gain	15:82906265-82985247	78,983	15q25.2	LOC100506874, SCAND2, UBE2Q2P1, ZSCAN2	gene/exonic
14070_1230	М	Eur	Loss	15:91200007-91283004	82,998	15q26.1	CHD2, LOC100507217, MIR3175	gene/exonic
2204_1	М	Eur	Loss	16:29466569-30147029	680,461	16p11.2	42 genes (includes KCTD13, MAPK3, SEZ6L2)	gene/exonic
20089_1391001	M	Eur	Loss	16:29502984-30107306	604,323	16p11.2	29 genes (includes KCTD13, MAPK3, SEZ6L2)	gene/exonic
5068_3	F	Eur	Loss	16:29502984-30127026	624,043	16p11.2	40 genes (includes <i>KCTD13, MAPK3, SEZ6L2</i> )	gene/exonic
5262_4	M	Eur	Gain	16:29502984-30210849	707,866	16p11.2	43 genes (includes KCTD13, MAPK3, SEZ6L2)	gene/exonic
4030_1	M	Eur	Gain	16:29554843-30107306	552,464	16p11.2	28 genes (includes KCTD13, MAPK3, SEZ6L2)	gene/exonic
5359_4	M	Eur	Loss	16:29554843-30195224	640,382	16p11.2	42 genes (includes <i>KCTD13</i> , <i>MAPK3</i> , <i>SEZ6L2</i> )	gene/exonic
	M	Eur		17:17156307-18262979				gene/exonic
3439_3 2211_1	F	Eur	Loss	17:17150307-18202979	1,106,673 2,932,260	17p11.2 17p11.2	22 genes (includes RAI1, SREBF1, LLGL1, TOP3A) 53 genes (includes MAPK7, RAI1, SREBF1, FAM83G, LLGL1, TOP3A)	
5056 4	М	Eur	Gain	17:34612208-34732327	120,120	17q12	FBXL20, RPL19, STAC2	gene/exonic
8463 202	M	Eur	Loss	17:52774693-52895975	121,283	17q22	MSI2	gene/exonic
5444_3	M	Eur	Gain	17:76953064-77782267	829,204	17q25.3	38 genes (includes ACTG1, ARHGDIA, FASN, RAC3, HGS, DUS1L)	gene/exonic
5444_3	М	Eur	Loss	17:77785939-77849717	63,779	17q25.3	CSNK1D, SLC16A3	gene/exonic
3477_3	М	Eur	Loss	18:30280260-30327512	47,253	18q12.1	DTNA	gene/exonic
14331 4450	М	Eur	Loss	18:72085223-73670156	1,584,934	18q23	GALR1, LOC284276, MBP, ZNF236, ZNF516	gene/exonic
6358 <u>6</u>	М	Eur	Loss	19:4548413-5287389	738,977	19p13.3	ARRDCS, C19orf10, DPP9, FEM1A, KDM4B, MIR7-3, NCRNA00306, PLIN3, PTPRS, TICAM1, TNFAIP8L1, UHRF1	gene/exonic
5335_3	М	Eur	Loss	20:14545734-14948785	403,052	20p12.1	MACROD2, MACROD2-AS1	gene/exonic
5046_3	М	Other	Loss	20:8607242-8637441	30,200	20p12.3	PLCB1	gene/exonic
20180_1704001	М	Eur	Gain	21:43018846-43444308	425,463	21q22.3	CBS, NDUFV3, PDE9A, PKNOX1, U2AF1, WDR4	gene/exonic
3183_7	M	Eur	Loss	22:17241748-19819918	2,578,171	22q11.21	63 genes (includes TBX1, CRKL, TSSK2)	gene/exonic
8627_201	М	Eur	Gain	22:17257787-19793730	2,535,944	22q11.21	63 genes (includes TBX1, CRKL, TSSK2)	gene/exonic
17015_1	М	Eur	Loss	22:17257787-19795780	2,537,994	22q11.21	63 genes (includes TBX1, CRKL, TSSK2)	gene/exonic
4271_1	M	Eur	Gain	22:17257787-19795780	2,537,994	22q11.21	63 genes (includes TBX1, CRKL, TSSK2)	gene/exonic
8630_201	M	Eur	Loss	22:32346124-32413987	67,864	22q12.3	LARGE	gene/exonic
2072_1	М	Eur	Loss	22:45159185-49582267	4,423,083	22q13.31- q13.33	47 genes (includes SHANK3)	gene/exonic
6130_4	F	Eur	Loss	22:47996161-49512530	1,516,370	22q13.32- q13.33	37 genes (includes SHANK3)	gene/exonic
14291_4120	F	Eur	Loss	22:49468716-49485255	16,540	22q13.33	SHANK3	gene/exonic
16079_1571066001	М	Eur	Loss	22:49470371-49567383	97,013	22q13.33	ACR, RABL2B, RPL23AP82, SHANK3	gene/exonic
20013_1075001	М	Eur	Gain	X:153239048-153521797	282,750	Xq28	20 genes (includes EMD, FLNA, GDI1, IKBKG, PLXNA3, RPL10)	gene/exonic

We identified 102 rare *de novo* CNVs in 99 cases; three individuals (5444\_3, 8630\_201 and 14310\_4270) have 2 *de novo* CNV each. Of the 99 subjects with *de novo* events, 60 (60%) are simplex, 26 (27%) multiplex, and 13 (13%) of unknown status. Eleven *de novo* chromosomal abnormalities >7.5 Mb identified in probands are listed in **Table S1C**. Abbreviations: Eur, European; F, female; M, male.

Table S5A. Parent of origin for rare *de novo* validated CNVs in probands

Sample ID	Chr#:start-end (hg18)	CNV type	Family type	Parent of origin	SNPs:  #paternal #maternal total
14310_4270	1:105623065-105795181	Gain	SPX	Paternal	SNPs:  20 0 41
14310_4270	1:105896243-106480692	Gain	SPX	Paternal	Custom-designed microsatellite (14310b)
16035_1571013001	1:144482933-146325557	Gain	MPX	Unknown	No informative SNPs or microsatellites
1952_301	1:144500467-146336720	Gain	MPX	Paternal	SNPs:  24 0 534
8635_201	1:144500467-146377870	Loss	MPX	Paternal	SNPs:  73 0 617
13135_1523	1:144838594-146308287	Gain	UNK	Paternal	SNPs:  121 56 667
6356_5	1:2118508-2325536	Loss	SPX	Maternal	SNPs:  0 11 92
8658_201	1:98175622-100923952	Loss	SPX	Maternal	SNPs:  0 199 1018
9877_204	1:98247355-99645560	Gain	SPX	Unknown	Ambiguous, SNPs:  74 27 488; microsat ?D1S2739
5089 5	2:102292943-102345460	Loss	MPX	Unknown	No informative SNPs or microsatellites
13082_963	2:11712589-11741036	Gain	UNK	Maternal	Custom-designed microsatellite (13082b)
4260_1	2:119603876-119882278	Gain	SPX	Maternal	SNPs:  1 26 118 (Griswold et al. 2012) <sup>128</sup>
13027_353	2:144837809-145315383	Gain	UNK	Maternal	SNPs:  0 40 124
14414 5230	2:230486629-230547253	Loss	SPX	Unknown	No informative SNPs
14068 1180	2:50493827-50677835	Gain	SPX	Maternal	SNPs:  0 15 75
13017 223	2:50539877-50730546	Loss	UNK	Paternal	SNPs:  19 0 89
13216_2383	2:50968208-51214171	Loss	UNK	Maternal	SNPs:  0 10 88
13153_1703	2:50990306-51222043	Loss	UNK	Paternal	Custom-designed microsatellite (13153a)
13037 463	2:51002576-51157742	Loss	SPX	Paternal	SNPs:  10 0 47
			MPX	Unknown	No informative SNPs
5245_3 8587_210	3:117285007-117477191 3:139760896-140127703	Loss	UNK	Paternal	
					SNPs:  5 0 105
5220_3	3:19127998-19640299	Gain	SPX	Paternal	SNPs:  38 0 205
8588_201	3:195971510-197675831	Loss	UNK	Maternal	SNPs:  0 35 633
3174_3	3:19921061-20096832	Loss	MPX	Maternal	SNPs:  3 0 72
3616_3	3:9474320-9498362	Loss	UNK	Unknown	No informative SNPs
8677_201	4:94659049-95289667	Gain	SPX	Paternal	SNPs:  28 1 195
6402_3	4:98781694-99627129	Gain	SPX	Maternal	SNPs:  0 28 230
8709_201	5:88588170-89505509	Loss	SPX	Paternal	SNPs:  48 0 249
5386_3	6:156785155-158489874	Loss	MPX	Paternal	SNPs:  115 0 605
6164_3	6:160023074-160081618	Gain	SPX	Unknown	Ambiguous, SNPs:  2 5 23 *
8612_201	6:162588879-162629938	Loss	UNK	Paternal	Custom-designed microsatellite (8612a)
8404_201	6:169136788-170761395	Loss	MPX	Maternal	SNPs:  0 104 648
5353_3	6:33399849-33512042	Loss	SPX	Paternal	SNPs:  1 0 63 **
18133_302	6:69540429-73375020	Gain	SPX	Maternal	SNPs:  0 182 1516
6248_3	6:86352577-86376159	Loss	SPX	Unknown	No informative SNPs
1960_301	7:102699832-102798745	Loss	MPX	Paternal	D7S2509
5370_3	7:153775586-153844747	Loss	SPX	Maternal	SNPs:  0 6 43
8446_201	7:72344426-73782113	Loss	SPX	Maternal	SNPs:  0 66 615
1142_4	8:48631388-48802529	Gain	MPX	Paternal	SNPs:  3 0 27
6321_3	8:65354366-66254869	Loss	SPX	Unknown	No informative SNPs or microsatellites
5290_3	8:704383-1521910	Gain	SPX	Paternal	SNPs:  85 37 361
8500_201	8:74692898-74967280	Loss	MPX	Maternal	SNPs:  0 22 91
14417_5260	9:117311405-117727764	Loss	SPX	Paternal	SNPs:  30 0 141
9756_201	9:132572435-132597937	Loss	SPX	Unknown	No informative SNPs
6259_3	9:139516033-140208462	Loss	SPX	Paternal	SNPs:  23 0 316
6246_4	9:9399606-9631169	Loss	SPX	Unknown	No informative SNPs; no informative microsatellites***
5032_4	9:98998-334508	Loss	MPX	Paternal	SNPs:  15 0 128
13123_1403	9:98998-3682923	Loss	UNK	Paternal	SNPs:  204 0 1992
18172_302	10:34963905-35536952	Gain	SPX	Paternal	SNPs:  61 0 179
8534_201	10:45633089-51564756	Loss	SPX	Maternal	SNPs:  0 161 1197
14393 5020	10:74801551-75278951	Loss	SPX	Paternal	SNPs:  1 0 225 and patD10S188
6240_4	11:126633939-132060374	Loss	SPX	Paternal	SNPs:  351 0 2144
6325_3	11:70077507-70506315	Loss	SPX	Maternal	SNPs:  0 15 138 (Leblond et al 2012) <sup>129</sup>
6319_3	11:70119917-70187872	Loss	SPX	Paternal	SNPs:  3 0 24
UJ1J J	TT.10TTJJT1 .10T01017	LUJJ	JI /\	raterial	JIN J.  J U 47

Sample ID	Chr#:start-end (hg18)	CNV type	Family type	Parent of origin	SNPs:  #paternal #maternal total
6053_3	12:54218922-58779615	Gain	MPX	Maternal	SNPs:  0 241 1766
5272_3	12:98445422-98540678	Loss	MPX	Maternal	SNPs:  0 5 32
1050_3	14:20279711-20345174	Gain	MPX	Paternal	SNPs:  14 0 49
8638_201	14:35203374-35369853	Loss	UNK	Paternal	Custom designed microsatellite (8638b)
4272_1	14:63824114-65347410	Loss	SPX	Maternal	SNPs:  0 47 563 (Griswold et al. 2012) <sup>128</sup>
17009_1	14:78575296-78596793	Loss	SPX	Unknown	No informative SNPs or microsatellites
20187_1464001	15:18811937-26209270	Gain	SPX	Maternal	D15S1002, D15S128
8630_201	15:19800798-26209270	Gain	MPX	Paternal	SNPs:  328 9 2265
20069_1328001	15:20203578-26209270	Gain	SPX	Maternal	SNPs:  0 467 2230
17035_1	15:20274130-26120360	Gain	SPX	Maternal	D15S1002, D15S128
8430_204	15:20301669-20777695	Loss	SPX	Paternal	SNPs:  16 0 187
13050_593	15:21190624-26203954	Gain	UNK	Maternal	D15S128
16040_1571029001	15:28450423-30303265	Loss	MPX	Paternal	SNPs:  56 0 481
6101_4	15:74735339-74929817	Loss	MPX	Paternal	SNPs:  5 0 36
8695_201	15:81728085-82623936	Loss	SPX	Maternal	SNPs:  0 27 299
14181_2940	15:82906265-82985247	Gain	SPX	Maternal	SNPs:  0 4 18
14070_1230	15:91200007-91283004	Loss	MPX	Paternal	SNPs:  1 0 22
2204_1	16:29466569-30147029	Loss	SPX	Maternal	SNPs:  0 28 248
20089_1391001	16:29502984-30107306	Loss	SPX	Maternal	SNPs:  0 21 306
5068_3	16:29502984-30127026	Loss	MPX	Paternal	SNPs:  5 0 249
5262_4	16:29502984-30210849	Gain	SPX	Paternal	SNPs:  69 4 249
4030_1	16:29554843-30107306	Gain	MPX	Unknown	Ambiguous, SNPs:  22 22 253
5359_4	16:29554843-30195224	Loss	SPX	Maternal	SNPs:  0 1 248; no informative microsatellites
3439_3	17:17156307-18262979	Loss	SPX	Paternal	SNPs: 184 0 439
2211_1	17:17169258-20101517	Loss	SPX	Paternal	SNPs:  79 0 927
5056_4	17:34612208-34732327	Gain	MPX	Maternal	SNPs:  0 1 57; no informative microsatellites
8463_202	17:52774693-52895975	Loss	MPX	Paternal	SNPs:  5 0 66
5444_3	17:76953064-77782267	Gain	SPX	Paternal	SNPs:  20 5 276
5444_3	17:77785939-77849717	Loss	SPX	Paternal	Custom-designed microsatellite
3477_3	18:30280260-30327512	Loss	SPX	Maternal	Custom-designed microsatellite (20xAC, 22xAC)
14331_4450	18:72085223-73670156	Loss	SPX	Paternal	SNPs:  168 0 900
6358_6	19:4548413-5287389	Loss	SPX	Maternal	SNPs:  0 57 310
5335_3	20:14545734-14948785	Loss	SPX	Maternal	SNPs:  0 7 151
5046_3	20:8607242-8637441	Loss	MPX	Maternal	Custom-designed microsatellite (5046)
20180_1704001	21:43018846-43444308	Gain	SPX	Maternal	SNPs:  7 33 225
3183_7	22:17241748-19819918	Loss	MPX	Maternal	SNPs:  0 184 1251
8627_201	22:17257787-19793730	Gain	SPX	Paternal	SNPs: 292 94 1350
17015_1	22:17257787-19795780	Loss	SPX	Maternal	SNPs:  0 190 1444
4271_1	22:17257787-19795780	Gain	SPX	Maternal	SNPs:  105 249 1311
8630_201	22:32346124-32413987	Loss	MPX	Unknown	Ambiguous, SNPs:  21 25 291; no informative microsatellites
2072_1	22:45159185-49582267	Loss	MPX	Maternal	Gonadal mosaicism, two affected sibs (Moessner et al. 2008) <sup>130</sup>
6130_4	22:47996161-49512530	Loss	SPX	Paternal	SNPs:  170 0 875
14291_4120	22:49468716-49485255	Loss	SPX	Paternal	SNPs:  1 0 5
16079_1571066001	22:49470371-49567383	Loss	SPX	Paternal	SNPs:  7 0 39
20013_1075001	X:153239048-153521797	Gain	SPX	Unknown	_
16076_1571045001	X:22768481-23133948	Loss	SPX	Unknown	_

For 85 of 102 *de novo* events it was possible to determine the parent-of-origin from SNPs or microsatellite genotypes. Of the 85 *de novo* CNVs where parental origin could be assigned, 45 (53%) were paternally-derived and 40 (47%) were maternally-derived. These findings do not confirm the results from Hehir-Kwa et al. <sup>131</sup> of an increased rate of paternally-derived *de novo* events. Furthermore, paternal age was not found to be increased in fathers of ASD probands (with or without *de novo* CNVs) when compared to fathers of controls from that study (**Table S5H**). Parental bias was also not observed when we separately considered *de novo* events by type of CNV (pathogenic, uncertain significance, intergenic/intronic), mechanism of causality (mediated/flanked by segmental duplications or not) and type of family (simplex or multiplex).

 $Abbreviations: microsat, microsatellite\ marker;\ MPX,\ multiplex;\ SPX,\ simplex;\ UNK,\ unknown\ family\ type.$ 

<sup>\*</sup> Similar results when using a larger 500 kb-flanking SNP window: ambiguous (SNPs: |8|17|203)

<sup>\*\*</sup> Similar results when using a larger 500 kb-flanking SNP window: pat (SNPs: |1|0|369)

<sup>\*\*\*</sup> When using a 500 kb-flanking window: ambiguous (SNPs: |0|1|208)

# Tables S5B-S5H. Parent of origin of *de novo* CNVs – breakdown by family type and CNV characteristics

Table S5B. Breakdown by type of CNV

	Paternal	Maternal	Total
Pathogenic	23	20	43
Uncertain	20	19	39
Intergenic/ intronic	2	1	3
Total	45	40	85

Classification according to Tables S5A and S7A

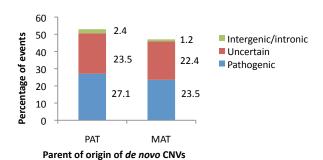


Table S5C. Breakdown by type of CNV, including large chromosomal abnormalities

	Paternal	Maternal	Total
Pathogenic	25	22	47
Uncertain	20	19	39
Intergenic/ intronic	2	1	3
Total	47	42	88

Classification according to Tables S5A and S7A

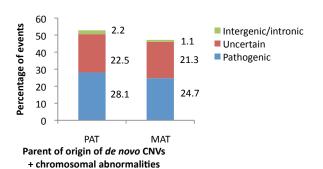


Table S5D. Breakdown by mechanism (CNV flanked or not by segmental duplications)

	Pa	Paternal		aternal	Total
	SD	no SD	SD	no SD	_
Pathogenic	10	13	13	7	43
Uncertain	1	19	1	18	39
Intergenic/ intronic	0	2	0	1	3
Total	11	34	14	26	85

Classification according to **Tables S5A** and **S7A** SD, segmental duplication

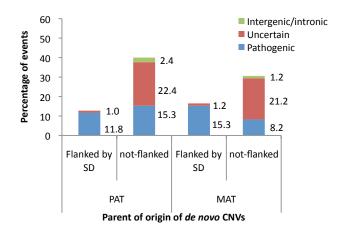


Table S5E. Breakdown of CNVs by type of family

	# Probands without de novo events	# Probands with de novo events	Total	%
AGP SPX	1142	59	1201	4.9
AGP MPX	600	26	626	4.2
AGP Unknown	256	13	269	4.8
Total	1998	98	2096	4.7
SSC	1059	65	1124	5.8

Number of simplex (SPX) and multiplex (MPX) families after QC is 1201 and 626, respectively. SSC, Simons Simplex Collection  $^{132}$ 

Table S5F. Breakdown of *de novo* events by type of family

	Paternal	Maternal	Total
AGP SPX	23	26	49
AGP MPX	13	9	22
SSC	27	34	61
Total	63	69	132

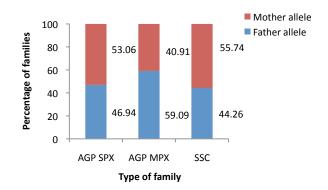


Table S5G. Breakdown of *de novo* CNVs by size in different family types

	<30 kb	30 kb- 500 kb	500 kb- 1 Mb	>1 Mb
AGP SPX	4	23	15	17
AGP MPX	0	14	2	10
Total	4	37	17	27

Average CNV size is 1,199,751 bp for SPX, 1,180,129 bp for MPX, and 1,041,851 bp for families classified as unknown.

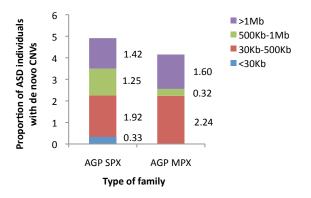


Table S5H. Breakdown of de novo CNVs by paternal age

	# Fathers in group*	Median paternal age ± SD
ASD without de novo CNVs	1659	32.33 ± 5.74
ASD with de novo CNVs	76	32.17 ± 6.57
ASD with de novo CNVs, paternal in origin, with flanking SD	6	33.75 ± 6.03
ASD with de novo CNVs, paternal in origin, without flanking SD	24	33.71 ± 6.08

\*45 *de novo* CNVs were paternally derived from 43 fathers (2 affected individuals had 2 paternally-derived *de novo* CNVs each), and parental age was available for 30 of them. The average maternal and paternal age at childbirth in our group of rare *de novo* CNVs was 29.6 and 32.2 years, respectively (76/99 had information on parental age). These age ranges were similar to parental ages at childbirth in the control cohort used by Hehir-Kwa et al. (31.4 and 32.1 years, respectively).

# Tables S6A-S6D. List of genes and loci implicated in ASD and ID

## Table S6A. Genes implicated in ASD

	Gene	Chr. band	Disorder	Inheritance patterr
1	POMGNT1	1p34.1	Muscle-eye-brain disease	AR
2	RPE65	1p31.3	Leber congenital amaurosis	AR
3	DPYD	1p21.3	Dihydropyrimidine dehydrogenase deficiency	AR
4	NRXN1	2p16.3	Disrupted in ASD, ID, schizophrenia (dominant); Pitt-Hopkins-like syndrome 2 (recessive)	AD/AR
5	NPHP1	2q13	Joubert syndrome type 4, nephronophthisis	AR
6	MBD5	2q23.1	Autosomal dominant ID, 2q23.1 microdeletion syndrome	AD
7	SCN1A	2q24.3	Severe myoclonic epilepsy of infancy (Dravet syndrome)	AD
8	SCN2A	2q24.3	Benign familial neonatal-infantile seizures, intractable childhood epilepsy	AD
9	SATB2	2q33.1	Cleft palate and ID; implicated in the 2q33.1 microdeletion syndrome	AD
10	BTD	3p24.3	Biotinidase deficiency	AR
	FOXP1	3p14.1	Non-syndromic ID with language impairment and ASD	AD
11			, , ,	
12	PRSS12	4q26	Autosomal recessive non-syndromic ID	AR
13	NIPBL	5p13.2	Cornelia de Lange syndrome	AD
14	MEF2C	5q14.3	ID, stereotypic movements, epilepsy, and/or cerebral malformations; 5q14.3 microdeletion syndrome	AD
15	ALDH7A1	5q23.2	Pyridoxine-dependent epilepsy	AR
16	NSD1	5q35.2-q35.3	Sotos syndrome	AD
17	ALDH5A1	6p22.2	Succinic semialdehyde dehydrogenase deficiency	AR
18	SYNGAP1	6p21.32	Autosomal dominant ID	AD
19	AHI1	6q23.3	Joubert syndrome 3	AR
20	PEX7	6q23.3	Refsum disease, Rhizomelic chondrodysplasia punctata, type 1	AR
21	ARID1B	6q25.3	ID, speech impairment, minor anomalies and variable corpus callosum abnormalities; Coffin-Siris syndrome	AD
22	HOXA1	7p15.2	HOXA1 syndrome, Bosley-Salih-Alorainy variant	AR
23	BRAF	7g34	Cardio-facio-cutaneous syndrome	AD
			,	
24	CNTNAP2	7q35-q36.1	Cortical dysplasia-focal epilepsy syndrome, Pitt-Hopkins-like syndrome 1 (recessive); the clinical	AR
			significance of the disruption of 1 allele is unknown	
25	HGSNAT	8p11.21	Mucopolysaccharidosis type IIIC (Sanfilippo syndrome C)	AR
26	TUSC3	8p22	Autosomal recessive non-syndromic ID	AR
27	CHD7	8q12.2	CHARGE syndrome	AD
28	VPS13B	8q22.2	Cohen syndrome	AR
29	SMARCA2	9p24.3	Nicolaides-Baraitser syndrome; Coffin-Siris syndrome	AD
30	STXBP1	9q34.11	Non-syndromic epilepsy, ID and autism; early infantile epileptic encephalopathy	AD
31	POMT1	9q34.13	Limb-girdle muscular dystrophy with ID; Walker-Warburg syndrome	AR
32	TSC1	9q34.13	Tuberous sclerosis	AD
		•		
33	EHMT1	9q34.3	Kleefstra syndrome (9q subtelomeric deletion syndrome)	AD
34	PTEN	10q23.31	PTEN hamartoma-tumor syndrome, ID and ASD with macrocephaly	AD
35	FGFR2	10q26.13	Apert syndrome	AD
36	HRAS	11p15.5	Costello syndrome	AD
37	IGF2	11p15.5	Aberrant imprinting of <i>IGF2</i> is associated with Beckwith–Wiedermann syndrome and Silver–Russell syndrome	AD
38	KCNJ11	11p15.1	DEND syndrome (developmental delay, epilepsy, and neonatal diabetes)	AD
39	SHANK2	11q13.3	Non-syndromic ID and ASD	AD
40	DHCR7	11q13.4	Smith-Lemli-Opitz syndrome	AR
41	FOLR1	11q13.4	Cerebral folate transport deficiency	AR
12	HEPACAM	11q24.2	Megalencephalic leukoencephalopathy with subcortical cysts (recessive); leukodystrophy and macrocephaly (dominant)	AR/AD
43	CACNA1C	12p13.33	Timothy syndrome	AD
44	GRIN2B	12p13.3	Autosomal dominant ID	AD
<del>14</del> 15	KRAS	12p13.1	Cardio-facio-cutaneous syndrome	AD
	SCN8A			
16 17		12q13.13	Early infantile epileptic encephalopathy	AD
47 40	GNS	12q14.3	Mucopolysaccharidosis type IIID (Sanfilippo disease D)	AR
48	BBS10	12q21.2	Bardet-Biedl syndrome	AR
49	CEP290	12q21.32	Joubert syndrome 5, Leber congenital amaurosis, Bardet-Biedl syndrome, Meckel syndrome	AR
50	PAH	12q23.2	Phenylketonuria	AR
51	PTPN11	12q24.13	Noonan syndrome	AD
52	CHD8	14q11.2	Autosomal dominant ASD and ID	AD
53	FOXG1	14q12	Congenital variant of Rett syndrome	AD
54	L2HGDH	14q22.1	L-2-hydroxyglutaric aciduria	AR
55	UBE3A	15q11.2	Angelman syndrome	AD
56	SPRED1	15q14	Neurofibromatosis type 1-like syndrome (Legius syndrome)	AD
			,, , , , , , , , , , , , , , , , , , , ,	
57	GATM	15q21.1	Arginine:glycine amidinotransferase (AGAT) deficiency	AR
58	MAP2K1	15q22.31	Cardio-facio-cutaneous syndrome	AD
59	TSC2	16p13.3	Tuberous sclerosis	AD
60	CREBBP	16p13.3	Rubinstein-Taybi syndrome	AD
51	SRCAP	16p11.2	Floating-Harbor syndrome	AD
52	BCKDK	16p11.2	Autosomal recessive autism, ID & epilepsy/abnormal EEG	AR
			· · · · · · · · · · · · · · · · · · ·	

	Gene	Chr. band	Disorder	Inheritance pattern
64	ANKRD11	16q24.3	KBG syndrome; 16q24.3 microdeletion syndrome	AD
65	YWHAE	17p13.3	Miller-Dieker syndrome	AD
66	PAFAH1B1	17p13.3	Isolated lissencephaly, Miller-Dieker syndrome	AD
67	GUCY2D	17p13.1	Leber congenital amaurosis	AR
68	RAI1	17p11.2	Smith-Magenis syndrome (deletion, mutation), Potocki-Lupski syndrome (duplication)	AD
69	RNF135	17q11.2	Overgrowth syndrome; haploinsufficiency of RNF135 contributes to the phenotype of the NF1 microdeletion syndrome	AD
70	NF1	17q11.2	Neurofibromatosis type 1	AD
71	NAGLU	17q21.31	Mucopolysaccharidosis type IIIB (Sanfilippo syndrome B)	AR
72	SGSH	17q25.3	Sanfilippo syndrome A (mucopolysaccharidosis III A)	AR
73	SETBP1	18q12.3	Haploinsufficiency of SETBP1 causes the core clinical features of the del(18)(q12.2q21.1) syndrome	AD
74	SMAD4	18q21.2	Myhre syndrome	AD
75	TCF4	18q21.2	Pitt-Hopkins syndrome	AD
76	NFIX	19p13.13	Sotos-like overgrowth syndrome, Marshall-Smith syndrome	AD
77	GAMT	19p13.3	Guanidine acetate methyltransferase (GAMT) deficiency	AR
78	DMPK	19q13.32	Myotonic dystrophy type 1 (Steinert disease)	AD
79	MKKS	20p12.2	Bardet-Biedl syndrome	AR
80	DYRK1A	21q22.13	Autosomal dominant ID	AD
81	ADSL	22q13.1	Adenylosuccinate lyase deficiency	AR
82	SHANK3	22q13.33	Phelan-mcdermid syndrome (22q13 deletion syndrome)	AD
83	NLGN4X	Xp22.31-p22.32	Non-syndromic X-linked ID and ASD	XLR
84	MID1	Xp22.2	Opitz syndrome (Opitz/BBB syndrome)	XLR
85	AP1S2	Xp22.2	Syndromic X-linked ID, Fried type; non-syndromic X-linked ID	XLR
86	NHS	Xp22.13	Nance-Horan syndrome	XLD
87	CDKL5	Xp22.13	Early infantile epileptic encephalopathy	XLD
88	PTCHD1	Xp22.11	Non-syndromic X-linked ID and ASD	XLR
89	ARX	Xp21.3	X-linked lissencephaly and abnormal genitalia, West syndrome, Partington syndrome, non-syndromic X-linked ID	XLR
90	IL1RAPL1	Xp21.2-p21.3	Non-syndromic X-linked ID and ASD	XLR
91	DMD	Xp21.1-21.2	Muscular dystrophy, Duchenne and Becker types	XLR
92	ОТС	Xp11.4	Ornithine transcarbamylase deficiency	XLD/XLR
93	CASK	Xp11.4	Syndromic and non-syndromic X-linked ID	XLD
94	NDP	Xp11.3	Norrie disease	XLR
95	KDM6A	Xp11.3	Kabuki syndrome	XLD
96	SYN1	Xp11.23	X-linked epilepsy and ID	XLR
97	FTSJ1	Xp11.23	Non-syndromic X-linked ID	XLR
98	PQBP1	Xq11.23	Renpenning syndrome, non-syndromic ID	XLR
99	CACNA1F	Xp11.23	X-linked incomplete congenital stationary night blindness	XLR
100	KDM5C	Xp11.22	Syndromic X-linked ID, Claes-Jensen type; non-syndromic X-linked ID	XLR
101	IQSEC2	Xp11.22	Non-syndromic X-linked ID	XLD
102	SMC1A	Xp11.22	Cornelia de Lange syndrome	XLD
103	HSD17B10	Xp11.22	17-beta-hydroxysteroid dehydrogenase X deficiency, X-linked syndromic ID with choreoathetosis and abnormal behavior	XLD
104	PHF8	Xp11.22	Siderius-Hamel syndrome	XLR
105	FGD1	Xp11.22	Aarskog-Scott syndrome, non-syndromic X-linked ID	XLR
106	OPHN1	Xq12	X-linked ID with cerebellar hypoplasia and distinctive facial appearance	XLR
107	MED12	Xq13.1	Lujan-Fryns syndrome, Opitz-Kaveggia syndrome	XLR
108	NLGN3	Xq13.1	Non-syndromic X-linked ID and ASD	XLR
109	ATRX	Xq21.1	Alpha-thalassemia/mental retardation syndrome, non-syndromic X-linked ID	XLD
110		Xq22.1	Female-limited epilepsy with ID, early infantile epileptic encephalopathy	XL - affected female
111		Xq22.3	Non-syndromic X-linked ID	XLD
112		Xq22.3	Type 1 lissencephaly	XLD
113	1	Xq24	Non-syndromic X-linked ID, Opitz-Kaveggia/Lujan-Fryns phenotype	XLR
114		Xq24	Danon disease	XLD
115		Xq25	Syndromic X-linked ID, Wu type; non-syndromic X-linked ID	XLR
116		Xq25	Lowe syndrome	XLR
117		Xq26.2	Borjeson-Forssman-Lehmann syndrome	XLR
118		Xq26.3	Syndromic X-linked ID, Christianson type	XLD?
119		Xq27.3	Fragile X syndrome	XLD:
120	1	Xq28	Fragile X mental retardation 2	XLR
120	SLC6A8		Creatine deficiency syndrome, non-syndromic X-linked ID	XLR
		Xq28	MASA (mental retardation, aphasia, shuffling gait, and adducted thumbs) syndrome	XLR
122		Xq28		
123	MECP2	Xq28	Rett syndrome, non-syndromic X-linked ID (mutation, deletion; XL dominant); MECP2 duplication syndrome (XL recessive)	XLD/XLR

Abbreviations: AD, autosomal dominant; AR, autosomal recessive; ASD, autism spectrum disorder; ID, intellectual disability; XL, X-linked; XLD, X-linked dominant; XLR, X-linked recessive

Table S6B. Loci implicated in ASD

	Disorder	Chr. band	Start (hg18)	End (hg18)	Genes involved	Inheritanc pattern
1	1p36 deletion syndrome	1p36.32-p36.33	823,946	5,308,621	contiguous gene syndrome	AD
2	1q21.1 deletion/duplication syndrome	1q21.1	145,044,110	145,861,130	?	AD
_	1q41q42 microdeletion syndrome	1q41q42	219,500,000	223,000,000	?	AD
	1g43g44 microdeletion syndrome	1q43q44	238,000,000	247,249,718	?	AD
_	2p16.1p15 microdeletion syndrome	2p16.1p15	57,595,300	61,591,838	?	AD
_	2q13 deletion/duplication	2q13	111,158,601	112,782,250	?	AD
	2q23.1 microdeletion syndrome	2q23.1	148,932,242	148,987,514	MBD5	AD
_	2q33.1 deletion syndrome	2q32.3-q33.2	196,633,334	204,915,185	SATB2 (maybe other gene(s) contribute too)	AD
	2q37 deletion syndrome (brachydactyly-mental retardation syndrome)	2q37.3	239,619,630	242,579,273	HDAC4 (other distal gene(s) contribute too)	AD
)	3q13.31 microdeletion syndrome	3q12.3q21.3	115,335,356	115,916,848	?	AD
	3q29 microdeletion/microduplication syndrome	3q29	197,240,451	198,829,062	? (candidates: PAK2 and DLG1)	AD
2	Wolf-Hirschhorn syndrome	4p16.3	62,448	2,297,002	contiguous gene syndrome	AD
3	4q21 microdeletion syndrome	4q21.21-q21.22	82,228,875	83,182,488	?	AD
l l	Cri du Chat syndrome (5p deletion)	5p15.2-p15.33	90,693	11,400,262	?	AD
	5q14.3 microdeletion syndrome	5q14.3	88,049,814	88,235,678	MEF2C	AD
_	Sotos syndrome (deletion), 5q35.2q35.3	5q35.2-q35.3	175,661,584	176,946,567	NSD1	AD
	duplication					
	6p subtelomere deletion syndrome	6p25	100,000	3,000,000	? (FOXC1 involved in ophthalmologic findings)	AD
	Williams syndrome (deletion), 7q11.23 duplication syndrome	7q11.23	72,382,390	73,780,449	contiguous gene syndrome	AD
	8p23.1 deletion/duplication syndrome	8p23.1	8,156,705	11,803,128	? (GATA4 involved in heart defects)	AD
	8q21.11 microdeletion syndrome	8q21.11	77,389,019	77,928,794	?	AD
l	Kleefstra syndrome (9q subtelomeric deletion syndrome)	9q34.3	139,523,184	140,273,252	EHMT1	AD
	10p14p15 deletion syndrome	10p14-p15.1	4,700,001	10,600,000	? (GATA3 involved in hypoparathyroidism, deafness, renal disease)	AD
3	10n12n11 microdolation	10n12 21n11 21	20 022 105	29,138,742		۸D
_	10p12p11 microdeletion	10p12.31p11.21	28,833,195		?	AD
_	10q22-q23 deletion	10q22.3-q23.2	81,682,644	88,931,994	?	AD
_	Distal 10q deletion syndrome	10q26.2-q26.3	128,000,000	135,374,737	?	AD
	11p15.5 duplication: Beckwith- Wiedemann/Silver-Russell syndromes	11p15.4-p15.5	1,970,000	2,870,000	H19, IGF2	AD
7	WAGR syndrome (11p13 deletion syndrome)	11p13	31,760,085	32,467,564	?	AD
	Potocki-Shaffer syndrome (11p11.2 deletion syndrome)	11p11.2	43,941,853	46,021,136	PHF21A (EXT2 and ALX4 involved in bone defects)	AD
	Jacobsen syndrome (11q deletion syndrome)	11q23.3-qter	115,400,001	134,452,384	?	AD
	12q14 microdeletion syndrome	12q14	63,358,186	66,931,792	? (HMGA2 involved in short stature)	AD
	Terminal deletion 14q syndrome	14q32.31-q32.33	101,500,000	105,000,000	?	AD
	Angelman syndrome (maternal deletion), Prader- Willi syndrome (paternal deletion), 15q11-q13	15q11.2-q13.1	21,309,483	26,230,781	Angelman: maternal <i>UBE3A</i> ; Prader-Willi: paternally expressed genes (HBII-85 snoRNA	AD
3	duplication syndrome 15q13.3 deletion syndrome	15q13.2-q13.3	28,924,396	30,232,700	cluster; SNURF-SNRPN, NDN and MAGEL2?) CHRNA7	AD
	(duplication = uncertain significance)	45 04 1 5 1 5	ma . a	70.045	2	
	15q24 microdeletion syndrome	15q24.1-q24.2	72,164,227	73,949,332	?	AD
	Distal 15q25.2q25.3 microdeletion	15q25.2q25.3	82,944,098	83,484,862	?	AD
	15q26 overgrowth syndrome	15q26.3	97,175,493	100,218,756	IGF1R	AD
	Rubinstein-Taybi syndrome (deletion), 16p13.3 duplication syndrome	16p13.3	3,715,057	3,870,122	CREBBP	AD
	16p13.11 microdeletion syndrome (duplication = uncertain significance)	16p13.11	15,411,955	16,199,769	?	AD
9	16p11.2-p12.2 microdeletion/microduplication syndrome	16p11.2-p12.2	21,521,457	28,949,693	?	AD
0	16p11.2 microdeletion/microduplication syndrome	16p11.2	29,557,497	30,107,356	?	AD
1	Miller-Dieker syndrome/isolated lissencephaly (deletion), 17p13.3 microduplication	17p13.3	1,129,706	2,535,659	PAFAH1B1, YWHAE	AD
2	Smith-Magenis syndrome (deletion), Potocki- Lupski syndrome (duplication)	17p11.2	16,697,836	20,160,243	RAI1	AD
	NF1 microdeletion/microduplication syndrome	17q11.2	26,186,948	27,242,780	NF1 (RNF135 contributes to the overgrowth,	AD
	17q12 deletion syndrome (renal cysts and	17q12	31,930,169	33,323,031	facial dysmorphism and ID) ? (HNF1B involved in renal cysts and diabetes	AD
5	diabetes syndrome), 17q12 duplication syndrome Koolen-De Vries syndrome (17q21.31 microdeletion syndrome), 17q21.31	17q21.31	41,060,949	41,544,225	syndrome) KANSL1	AD
	microduplication syndrome					
6	del(18)(q12.2q21.1) syndrome	18q12.2q21.1	39,890,000	41,700,000	SETBP1	AD
	Down syndrome (trisomy 21)	whole chr 21	1	46,944,323	contiguous gene syndrome	AD

	Disorder	Chr. band	Start (hg18)	End (hg18)	Genes involved	Inheritance pattern
48	22q11.2 deletion syndrome (velocardiofacial/DiGeorge syndrome), 22q11.2 duplication syndrome	22q11.21-q11.22	17,041,725	18,691,904	contiguous gene syndrome, <i>TBX1</i> is responsible for most of the physical malformations	AD
49	Phelan-McDermid syndrome (22q13 deletion syndrome), 22q13 duplication	22q13.33	49,392,382	49,525,811	SHANK3	AD
50	Xq28 duplication syndrome ( <i>MECP2</i> duplication syndrome)	Xq28	152,403,094	153,044,193	MECP2	XLR
51	Turner syndrome (X0)	whole chr X	1	154,913,754	contiguous gene syndrome	
52	Klinefelter syndrome (XXY)	whole chr X	1	154,913,754	contiguous gene syndrome	
53	XYY syndrome	whole chr Y	1	57,772,954	contiguous gene syndrome	
54	XXYY syndrome	whole chr X-Y			contiguous gene syndrome	
55	45,X/46,XY mosaicism	whole chr X	1	154,913,754	contiguous gene syndrome	

Abbreviations: AD, autosomal dominant; AR, autosomal recessive; ASD, autism spectrum disorder; ID, intellectual disability; XLR, X-linked recessive

**Table S6C. Genes implicated in ID** 

	Gene	Chr. band	Disorder	Inheritance pattern
1	SKI	1p36.33	Shprintzen-Goldberg syndrome	AD
2	GALE	1p36.11	Galactose epimerase deficiency (galactosemia III)	AR
3	FUCA1	1p36.11	Fucosidosis	AR
4	ARID1A	1p36.11	Coffin-Siris syndrome	AD
5	PIGV	1p36.11	Hyperphosphatasia mental retardation syndrome	AR
6	SLC2A1	1p34.2	Glucose transport defect	AD
7	ST3GAL3	1p34.3	Autosomal recessive non-syndromic ID	AR
8	STIL	1p33	Primary microcephaly	AR
9	ALG6	1p31.3	Congenital disorder of glycosylation, type Ic	AR
LO	DBT	1p21.2	Maple syrup urine disease, type II	AR
11	AP4B1	1p13.2	Autosomal recessive ID with spastic paraplegia	AR
12	NRAS	1p13.2	Noonan syndrome	AD
13	GATAD2B	1q21.3	Autosomal dominant ID	AD
L4	KCNJ10	1q23.2	SESAME syndrome (seizures, sensorineural deafness, ataxia, mental retardation, and electrolyte	AR
•		1920.2	imbalance)	
15	ASPM	1q31	Microcephaly and ID	AR
.6	SYT14	1q32.2	ID with adult-onset spinocerebellar ataxia	AR
.7	RAB3GAP2	1q41	Martsolf syndrome (congenital cataracts, hypogonadism, and ID)	AR
18	TBCE	1q42.3	Hypoparathyroidism-retardation-dysmorphism syndrome	AR
.9	FH	1q43	Fumarase deficiency	AR
20	MYCN	2p24.3	Feingold syndrome (microcephaly-oculo-digito-esophageal-duodenal syndrome), Microcephaly and digital	AD
.0	WITCH	2024.5	abnormalities with normal intelligence	7.0
21	SOS1	2p22.1	Noonan syndrome	AD
2	ERCC3	2q14.3	Trichothiodystrophy; Xeroderma pigmentosum, group B	AR
23	RAB3GAP1	2q21.3	Warburg Micro syndrome 1	AR
.3	ZEB2	2q21.3 2q22.3	Mowat-Wilson syndrome (Hirschsprung disease-mental retardation syndrome)	AD
25	BBS5	2q31.1	Bardet-Biedl syndrome	AR
26	GAD1	2q31.1 2q31.1	Spastic quadriplegic cerebral palsy	AR
27	TMEM237	2q33.1	Joubert syndrome 14	AR
28	HDAC4	2q33.1 2q37.3	Brachydactyly mental retardation syndrome (2q37 deletion syndrome)	AD
29	D2HGDH	2q37.3 2q37.3	D-2-hydroxyglutaric aciduria	AR
30	CRBN	3p26.2	Autosomal recessive non-syndromic ID	AR
30 31	SUMF1		•	AR
		3p26.2	Multiple sulfatase deficiency	
32	TSEN2	3p25.1	Pontocerebellar hypoplasia type 2B	AR
3	RAF1	3p25.1	Noonan syndrome, LEOPARD syndrome	AD
34	TGFBR2	3p24.1	Loeys-Dietz syndrome	AD
35	GLB1	3p22.3	GM1-gangliosidosis, Mucopolysaccharidosis IVB	AR
36	CTNNB1	3p22.1	Autosomal dominant ID	AD
37	GTDC2	3p22.1	Walker-Warburg syndrome	AR
38	LZTFL1	3p21.31	Bardet-Biedl syndrome with situs inversus and polydactyly	AR
39	DAG1	3p21.31	Limb-girdle muscular dystrophy	AR
10	ARL13B	3q11.2	Joubert syndrome 8	AR
11	ARL6	3q11.2	Bardet-Biedl syndrome	AR
12	CEP63	3q22.2	Primary microcephaly	AR
13	ATR	3q23	Seckel syndrome	AR
14	ALG3	3q27.1	Congenital disorder of glycosylation, type Id	AR
15	KIAA0226	3q29	Syndromic ID with ataxia, dysarthria and epilepsy	AR
16	IDUA	4p16.3	Mucopolysaccharidosis Ih (Hurler syndrome); mucopolysaccharidosis Is (Scheie syndrome)	AR
17	CC2D2A	4p15.3	Joubert syndrome 9, Meckel syndrome, COACH syndrome	AR
18	QDPR	4p15.32	Hyperphenylalaninemia due to dihydropteridine reductase deficiency	AR
19	SRD5A3	4q12	Kahrizi syndrome; congenital disorder of glycosylation, type Iq	AR
50	TMEM165	4q12	Congenital disorder of glycosylation, type iik	AR
51	CEP135	4q12	Autosomal-recessive primary microcephaly	AR

	Gene	Chr. band	Disorder	Inheritance pattern
52	SLC4A4	4q13.3	Renal tubular acidosis, proximal, with ocular abnormalities	AR
53	BBS7	4q27	Bardet-Biedl syndrome	AR
54	BBS12	4q27	Bardet-Biedl syndrome	AR
55	AGA	4q34.3	Aspartylglucosaminuria	AR
56 57	NSUN2 ANKH	5p15.31 5p15.2	Autosomal-recessive syndromic ID, Dubowitz syndrome Chondrocalcinosis 2, Craniometaphyseal dysplasia	AR AR
58	C5orf42	5p13.2	Joubert syndrome 17	AR
59	MOCS2	5q11.2	Molybdenum cofactor deficiency, type B	AR
60	ERCC8	5q11.2 5q12.1	Cockayne syndrome type A	AR
51	SIL1	5q31.2	Marinesco-Sjogren syndrome	AR
62	TUBB2B	6p25.2	Asymmetric polymicrogyria	AD
63	NEU1	6p21.3	Sialidosis type I, Sialidosis type II	AR
64	MOCS1	6p21.2	Molybdenum cofactor deficiency, type A	AR
65	SLC17A5	6q13	Salla disease; Sialic acid storage disorder, infantile	AR
66	ELOVL4	6q14.1	Ichthyosis, spastic quadriplegia, and mental retardation (recessive); Macular dystrophy (dominant)	AR/AD
67	BCKDHB	6q14.1	Maple syrup urine disease, type Ib	AR
68	RARS2	6q15	Pontocerebellar hypoplasia	AR
69	GRIK2	6q16.3	Autosomal recessive non-syndromic ID	AR
70	SOBP	6q21	Autosomal recessive syndromic and non-syndromic ID	AR
71	LAMA2	6q22.33	Merosin-deficient congenital muscular dystrophy type 1A	AR
72	ARG1	6q23.2	Argininemia	AR
73	MED23	6q23.2	Autosomal recessive non-syndromic ID	AR
74	GTF2H5	6q25.3	Trichothiodystrophy	AR
75	ACTB	7p22.1	Baraitser-Winter syndrome	AD
76	FAM126A	7p15.3	Hypomyelinating leukodystrophy	AR
77	BBS9	7p14.3	Bardet-Biedl syndrome	AR
78	C7orf11	7p14.1	Trichothiodystrophy	AR
79	GUSB	7q11.21	Mucopolysaccharidosis VII	AR
80	AP4M1	7q22.1	Autosomal recessive tetraplegic cerebral palsy with ID	AR
81	RELN	7q22	Lissencephaly	AR
82	DLD CEP41	7q31.1	Maple syrup urine disease, type III	AR
83	TPK1	7q32.2	Joubert syndrome 15  This miss match blism dust unction syndrome (onice discussed by acceptable part by type)	AR
84 85	EZH2	7q35	Thiamine metabolism dysfunction syndrome (episodic encephalopathy type)	AD AD
86	MCPH1	7q36.1 8p23	Weaver syndrome Microcephaly and ID	AR
87	ERLIN2	8p12	Autosomal recessive ID, motor dysfunction and multiple joint contractures	AR
88	CA8	8q12.1	Cerebellar ataxia, quadrupedal locomotion and ID	AR
89	TMEM67	8q21	Joubert syndrome 6, Meckel-Gruber syndrome	AR
90	RAD21	8q24.11	Cornelia de Lange syndrome	AD
91	KCNK9	8q24.3	Birk-Barel mental retardation dysmorphism syndrome (imprinting defect)	AD
92	TRAPPC9	8q24.3	Autosomal recessive non-syndromic ID	AR
93	RECQL4	8q24.3	Baller-Gerold syndrome, Rothmund-Thomson syndrome and RAPADILINO syndrome	AR
94	VLDLR	9p24.2	Cerebellar ataxia and ID	AR
95	SLC1A1	9p24.2	Dicarboxylic aminoaciduria	AR
96	PIGO	9p13.3	Hyperphosphatasia with mental retardation syndrome	AR
97	EXOSC3	9p13.2	Pontocerebellar hypoplasia and spinal motor neuron degeneration	AR
98	TGFBR1	9q22.33	Loeys–Dietz syndrome	AD
99	FKTN	9q31.2	Fukuyama congenital muscular dystrophy with type 2 lissencephaly, Walker-Warburg syndrome	AR
L00	TRIM32	9q33.1	Bardet-Biedl syndrome	AR
101	CDK5RAP2	9q33.2	Microcephaly vera	AR
102	SPTAN1	9q34.11	West syndrome with severe cerebral hypomyelination, spastic quadriplegia and ID	AD
103	KCNT1	9q34.3	Malignant migrating partial seizures of infancy; nocturnal frontal lobe epilepsy, ID	AD
104	INPP5E	9q34.3	Joubert syndrome 1	AR
105	MAN1B1	9q34.3	Autosomal recessive non-syndromic ID	AR
106	RAB18	10p12.1	Warburg micro syndrome  Cooking a windrome time B. Cooking only foois skaletal syndrome	AR
107	ERCC6	10q11.23	Cockayne syndrome type B, Cerebro-oculo-facio-skeletal syndrome	AR
801	KIAA1279	10q21.3	Goldberg-Shprintzen megacolon syndrome  Hypermethioninemia due to adenosine kinase deficiency	AR
L09 L10	ADK KAT6B	10q22.2	, ,	AR AD
.10	POLR3A	10q22.2 10q22.3	Say-Barber-Biesecker-Young-Simpson syndrome (SBBYSS syndrome)  Hypomyelinating leukodystrophy with or without oligodontia and/or hypogonadotropic hypogonadism	AR
112	KIF11	10q22.3 10q23.33	Microcephaly variably associated with congenital lymphedema, chorioretinopathy and learning difficulties	AD
.12	TCTN3	10q23.33	Joubert syndrome 18, orofaciodigital syndrome IV	AR
.14	SMC3	10q25.2	Cornelia de Lange syndrome	AD
115	SHOC2	10q25.2	Noonan syndrome	AD
116	SLC25A22	11p15.5	Autosomal recessive neonatal epileptic encephalopathy	AR
17	PAX6	11p13.3	Isolated and syndromic aniridia, including Gillespie syndrome (aniridia, cerebellar ataxia and ID)	AD
118	SLC35C1	11p11.2	Congenital disorder of glycosylation, type iic	AR
119	PHF21A	11p11.2	Potocki-Shaffer syndrome (11p11.2 deletion)	AD
20	TMEM138	11q12.2	Joubert syndrome 16	AR
	TMEM216	11q12.2	Joubert syndrome 2	AR
21				

123         ALCS         11q14.1         Congenital disorder of glycosylation type II           124         MEDI 7         11q23.1         Primary microcephaly of postnatal onset, spasticity, epilepsy, and ID           125         ALGS         11q23.3         Congenital disorder of glycosylation, type II           126         GR         11q23.3         Congenital disorder of glycosylation, type II           127         PVPLI         11q23.3         Cleft lip/palate ectodermal dysplasia syndrome           128         MLL2         12q13.12         Lissencephaly           130         DiPZB         12q13.13         Mental retardation, FRA12A type           131         SUOX         12q13.2         Suffice oxidase deficiency           132         GNPFAB         12q23.2         Growth retardation with deafness and mental retardation due to IGF1 deficient design of the subspace o	•
125         ALGS         11q23.1         Congenital disorder of glycosylation, type II           126         CBL         11q23.3         Noonan syndrome-like disorder           127         PVRLI         11q23.3         Cleft lip/palate ectodermal dysplasia syndrome           128         MuL2         12q13.12         Lisencephaly           130         DiP28         12q13.13         Mental retardation, FRA12A type           131         SUX         12q13.2         Suffice oxidase deficiency           132         GNPTAB         12q23.2         Mucolipidosis III alpha/beta           137         JCRAB         12q23.3         Leukodystrophy, hypomyelinating, 8, with or without oligodontia and/or hypog           133         JGPT         13q12.12         Microcephaly vera, Seckel syndrome           136         CENPI         13q12.12         Microcephaly vera, Seckel syndrome           137         SLC2SAL5         13q13.1         Cockayne syndrome           138         MiR17HG         13q33.1         Cockayne syndrome           140         COL4A1         13q34         Porencephaly           141         APAST         14q21.2         Autosomal recessive ID with spastic paraplegia           142         MAGT2         14q22.1         Cockayne syndro	AR AD AR AD AD AD AR AD AR AR AC AR
126         GBI         11q23.3         Noonan syndrome-like disorder           7PPRI         11q23.3         Cleft lip/palate ectodermal dysplasia syndrome           128         ML2         12q13.12         Kabuki syndrome           129         TUBALA         12q13.12         Lissencephaly           130         DIP2B         12q13.13         Mental retardation, FRA12A type           131         SUOX         12q13.2         Suffite oxidase deficiency           132         GNPTAB         12q23.2         Mucolipidosis III alpha/beta           133         GFI         12q23.2         Growth retardation with deafness and mental retardation due to IGF1 deficiency           134         POLR3B         12q23.2         Growth retardation with deafness and mental retardation on without oligodontia and/or hypog           135         CRPC         13q12.1         Cutsis laxa with pilepsy and mental retardation           136         CRPD         13q12.1         Microcephaly vera, Seckel syndrome           137         SLC25A15         13q14.11         Orinital trial t	AD AR AD AD AD AD AR AR AR CY AR Ronadotropic hypogonadism AR
127         WPLI         11q23.3         Cleft lip/patate ectodermal dysplasia syndrome           128         MLZ         12q13.12         Kabuki syndrome           129         TUBALA         12q13.12         Lissencephaly           130         DIPZB         12q13.13         Mental retardation, FRA12A type           131         SUOX         12q13.2         Suffice oxidase deficiency           132         GRPTAB         12q23.2         Mucolipidosis III alpha/beta           133         GFI         12q23.2         Growth retardation with deafness and mental retardation due to IGF1 deficient           134         POLR3B         12q23.3         Leukodystrophy, hypomyelinating, 8, with or without oligodontia and/or hypog           135         AFF6VOA2         12q24.31         Cutis laxa with epilepsy and mental retardation           136         CEND         13q12.12         Microcephaly vera, Seckel syndrome           137         SICZ5A15         13q14.11         Orinithine translocase deficiency           138         MRITHG         13q31.3         Feingold Syndrome           140         COL4A1         13q34         Porencephaly           141         APASI         14q12         Autosomal recessive ID with spastic paraplegia           142         MAGAT2	AR AD AD AD AR AR AR COMMENT OF THE MERCEN C
128         MLLZ         12q13.12         Lissencephaly           130         DIP2B         12q13.13         Mental retardation, FRA12A type           131         SUOX         12q13.2         Sulfite oxidase deficiency           132         GNPTAB         12q23.2         Mucolipidosis III alpa/peta           133         GFI         12q33.2         Growth retardation with deafness and mental retardation due to IGF1 deficiency           134         POLR3B         12q23.3         Leukodystrophy, hypomyelinating, 8, with or without oligodontia and/or hypog           135         ATP6V0A2         12q24.31         Cuts Isaxa with epilepsy and mental retardation           136         CENPJ         13q12.12         Microcephaly vera, Seckel syndrome           137         SLC25A15         13q14.11         Orinithine translocase deficiency           138         MiR17HG         13q31.3         Feingold syndrome           140         COL4A1         13q34         Porencephaly           141         AP6L5         14q31.3         Personation of Syndrome           142         MGAT2         14q22.1         Congenital disorder of glycosylation, type iia           143         GCH1         14q22.1         Congenital disorder of glycosylation, type iia           144	AD AD AD AR AR AR cy AR cy AR A
129         IVBATAL         12q13.12         Lissencephaly           30         DIP2B         12q13.13         Mental retardation, FRA12A type           131         SUOX         12q13.2         Sulfite oxidase deficiency           132         GNPTAB         12q23.2         Mucolipidosis III alpha/beta           134         POLR3B         12q33.3         Leukodystrophy, hypomyelinating, 8, with or without oligodontia and/or hypog           135         APP600A2         12q24.31         Cutis laxa with epilepsy and mental retardation           136         CENDP         13q12.12         Microcephaly vera, Seckel syndrome           137         SLCZ5A15         13q14.11         Orinithine translocase deficiency           138         MIRITAR         13q33.1         Cockayne syndrome, Cerebro-oculo-facio-skeletal syndrome           140         COLAI         13q33.1         Cockayne syndrome, Cerebro-oculo-facio-skeletal syndrome           141         APA51         14q12         Autosomal recessive ID with spastic paraplegia           142         MAGAT2         14q21.2         Cockayne syndrome or facio-skeletal syndrome           143         GGH         14q22.3         Walker-Warburg syndrome           144         POMT2         14q22.3         Walker-Warburg syndrome	AD AD AR AR CY AR AR Sonadotropic hypogonadism AR AR AR AR AR AR AR AD AR AD AR AR AD AR AR AD AR AR AD
130   10/128   12q13.13   Mental retardation, FRA12A type	AD AR AR CY AR CY AR CONTROL AR AR AR AR AR AR AR AR AD AR AR AD AR
SUOK   12q13.2   Suffite oxidase deficiency	AR AR cy AR cy AR gonadotropic hypogonadism AR AR AR AR AR AR AD
132   GNPTAB   12q23.2   Mucolipidosis III alpha/beta   13q13.1   13q23.2   Growth retardation with deafness and mental retardation due to IGF1 deficient   13q12.12   Growth retardation with deafness and mental retardation due to IGF1 deficient   13q12.12   Cutis laxa with epilepsy and mental retardation   13q12.12   Microcephaly vera, Seckel syndrome   13q12.12   Microcephaly vera, Seckel syndrome   13q12.13   13q12.12   Microcephaly vera, Seckel syndrome   13q12.13   Section   13q12.13   Feingold syndrome   13q14.11   Orinithine translocase deficiency   13q12.13   Feingold syndrome   13q14.11   Orinithine translocase deficiency   13q12.13   Feingold syndrome   14q12.14   Congenital disorder of glycosylation, type iia   14q12   Autosomal recessive ID with spastic paraplegia   14q12   Microcephaly   Autosomal recessive ID with spastic paraplegia   14q24.14   Macrocephaly   Mac	AR  Cy AR  Gonadotropic hypogonadism AR  AR  AR  AR  AR  AR  AD  AR  AR
133   GF1   12q23.2   Growth retardation with deafness and mental retardation due to IGF1 deficient	onadotropic hypogonadism AR AR AR AR AR AD AR AD AR AD AR AD AR AD AR AD AR
135ATP6V0A212q24.31Cutis laxa with epilepsy and mental retardation136CENPI13q12.12Microcephaly vera, Seckel syndrome137SLC25A1513q14.11Orinithine translocase deficiency138MR17HG13q31.3Feingold syndrome140COL4A113q34Porencephaly141AP4S114q12Autosomal recessive ID with spastic paraplegia142MGAT214q22.1Congenital disorder of glycosylation, type iia143GCH114q22.2-q22.3Tetrahydrobiopterin-deficient hyperphenylalaninemia B144POMT214q31.3Krabbe disease145GALC14q31.3Autosomal recessive non-syndrome146ZC3H1414q31.3Autosomal recessive non-syndromic ID147TTC814q31.3Bardet-Biedl syndrome148VRK114q32.2Pontocerebellar hypoplasia type 1149DYNC1H114q32.31Severe ID with neuronal migration defects; Charcot-Marie-Tooth disease, axon150SLC12A615q14Andermann syndrome151CEP12515q21.1Primary microcephaly152AP4E115q21.2Spastic paraplegia 51, autosomal recessive153BBS415q24.1Bardet-Biedl syndrome154KIF715q26.1Joubert syndrome 12155IOH215q15.1D-2-hydroxyglutaric aciduria 2156GNPTG16p13.3Mucolipidosis II gamma157TBCL02416p13.3Autosomal recessive syndrome of focal epile	AR AR AR AD AR AD AR AD AR AD AR
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137         SLC25A15         13q14.11         Orinithine translocase deficiency           138         FRCCS         13q31.3         Felingold syndrome           140         COL4A1         13q34         Porencephaly           141         AP4S1         14q12         Autosomal recessive ID with spastic paraplegia           142         MGAT2         14q22.1         Congenital disorder of glycosylation, type iia           143         GCH1         14q22.2         Congenital disorder of glycosylation, type iia           144         POMT2         14q22.3         Tetrahydrobiopterin-deficient hyperphenylalaninemia B           144         POMT2         14q23.3         Krabbe disease           145         GALC         14q31.3         Autosomal recessive non-syndrome           145         GALC         14q31.3         Bardet-Bied syndrome           148         VRK1         14q32.3         Severe ID with neuronal migration defects; Charcot-Marie-Tooth disease, axon: St. Clarcot.           149         DYNCLH1         14q32.31         Severe ID with neuronal migration defects; Charcot-Marie-Tooth disease, axon: St. Clarcot.           150         SLC12A6         15q14         Andermann syndrome           151         CEP152         15q21.1         Primary microcephaly	AR AD AR AD AR AR
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146         ZC3H14         14q31.3         Autosomal recessive non-syndromic ID           147         TTC8         14q31.3         Bardet-Biedl syndrome           148         VRK1         14q32.2         Pontocerebellar hypoplasia type 1           149         DYNC1H1         14q32.31         Severe ID with neuronal migration defects; Charcot-Marie-Tooth disease, axons           150         SLC12A6         15q14         Andermann syndrome           151         CEP152         15q21.1         Primary microcephaly           152         AP4E1         15q21.2         Spatic paraplegia 51, autosomal recessive           153         BBS4         15q24.1         Bardet-Biedl syndrome           154         KIF7         15q26.1         Joubert syndrome 12           155         IDH2         15q26.1         D-2-hydroxyglutaric aciduria 2           156         GNPTG         16p13.3         Mucolipidosis III gamma           157         TBCID24         16p13.3         Autosomal recessive syndrome of focal epilepsy, dysarthria, and ID           158         ZNF423         16q12.1         Joubert syndrome 19 (dominant); nephronophthisis (recessive)           159         TUBB3         16q34.3         Cortical dysplasia, complex, with other brain malformations           160	AR
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171PIGL17p11.2CHIME syndrome (colobomas, heart defects, ichthyosiform dermatosis, mental anomalies, including conductive hearing loss)172ALDH3A217p11.2Sjogren-Larsson syndrome173SLC46A117q11.2Folate malabsorption174SMARCE117q21.2Coffin-Siris syndrome175EFTUD217q21.31Mandibulofacial dysostosis with microcephaly176GFAP17q21.31Alexander disease	AR
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176 GFAP 17q21.31 Alexander disease	AD
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177 Minor 17922 Datact Dieut Syndrollie, Wiecker Syndrollie	AD
178 COG1 17q25.1 Congenital disorder of glycosylation, type iig	AD AD
179 TSEN54 17q25.1 Pontocerebellar hypoplasia type 2A	AD AD AR
180 ACTG1 17q25.3 Baraitser-Winter syndrome	AD AD AR AR
181 RBBP8 18q11.2 Seckel syndrome, Jawad syndrome	AD AD AR
182   IER3IP1 18q21.1 Microcephaly with simplified gyration, epilepsy, and infantile diabetes	AD AD AR AR AR
183 RTTN 18q22.2 Polymicrogyria, ID	AD AD AR AR AR AD
184 MAP2K2 19p13.3 Cardio-facio-cutaneous syndrome	AD AD AR AR AR AR AR AD
185 MCOLN1 19p13.2 Mucolipidosis IV	AD AD AR AR AR AR AD AD AR AD AR
186 SMARCA4 19p13.2 Coffin-Siris syndrome	AD AD AR AR AR AR AD AR AD AR AD AR AR AR AR
187 CC2D1A 19p13.12 Autosomal recessive non-syndromic ID	AD AD AR AR AR AR AD AR AD AR AD AR AR AD AR AR AD AR AR AR
188 GPSN2 19p13.12 Autosomal recessive non-syndromic ID	AD AD AR AR AR AR AD AR AD AR AR AD AR
189 WDR62 19q13.12 Severe brain malformations, including microcephaly, pachygyria and hypoplasia	AD AD AR AR AR AR AD AR AD AR AD AR AR AR AD AR AR AD AR AR AD AR AD AR AD AR AD AR AD AR AD
190 BCKDHA 19q13.2 Maple syrup urine disease, type la	AD  AD  AR  AR  AR  AR  AD  AR  AD  AR  AR
191 ERCC2 19q13.32 Cockayne syndrome, Trichothiodystrophy, Cerebro-oculo-facio-skeletal syndrome	AD AD AR AR AR AR AR AD AR AR AD AR AR AR AD AR AR AD AR AR AR AR AD AR
192 ERCC1 19q13.32 Cerebro-oculo-facio-skeletal syndrome	AD AD AR AR AR AR AR AD AR AR AD AR AR AR AD AR AR AD AR AR AR AR AD AR

	Gene	Chr. band	Disorder	Inheritance patter
193	FKRP	19q13.32	Congenital muscular dystrophy 1C, limb-girdle muscular dystrophy type 2I, muscle-eye-brain disease, Walker-Warburg syndrome	AR
.94	PNKP	19q13.33	Microcephaly, seizures and defects in DNA repair	AR
95	ASXL1	20q11.21	Bohring-Opitz syndrome	AD
96	DNMT3B	20q11.2	Immunodeficiency-centromeric instability-facial anomalies syndrome 1	AR
97	CTSA	20q13.12	Galactosialidosis	AR
98	ARFGEF2	20q13.13	Autosomal recessive periventricular heterotopia with microcephaly	AR
99	DPM1	20q13.13	Congenital disorder of glycosylation, type le	AR
00	CBS	21q22.3	Homocystinuria	AR
01	PCNT	21q22.3	Seckel syndrome, Majewski osteodysplastic primordial dwarfism type II	AR
02	SNAP29	22q11.21	Cerebral dysgenesis, neuropathy, ichthyosis, and palmoplantar keratoderma syndrome	AR
.03	SMARCB1	22q11.23	Coffin-Siris syndrome	AD
04		22q12.3	Congenital muscular dystrophy	AR
205	EP300	22q13.2	Rubinstein-Taybi syndrome	AD
06	CYB5R3	22q13.2	Methemoglobinemia type II	AR
.07	ALG12	22q13.33	Congenital disorder of glycosylation, type Ig	AR
80	СНКВ	22q13.33	Congenital muscular dystrophy, mitochondrial structural abnormalities and ID	AR
.09	HCCS	Xp22.2	Microphthalmia with linear skin defects syndrome	XLD
10	OFD1	Xp22.2	Oral-facial-digital syndrome type I (XL dominant), Simpson-Golabi-Behmel syndrome, type 2 (XL recessive), Joubert syndrome 10	XLD/XLR
11	FANCB	Xp22.2	VACTERL with hydrocephalus, Fanconi anemia of complementation group B	XLR
12	PDHA1	Xp22.12	Pyruvate decarboxylase deficiency	XLD
213	RPS6KA3	Xp22.12	Coffin-Lowry syndrome, non-syndromic ID	XLD
214	MBTPS2	Xp22.12	Ichthyosis follicularis, atrichia and photophobia syndrome	XLR
215	SMS	Xp22.11	X-linked ID, Snyder-Robinson type	XLR
216	GK	Xp21.2	Glycerol kinase deficiency	XLD
217	TSPAN7	Xp11.4	Non-syndromic X-linked ID	XLR
218	BCOR	Xp11.4	Syndromic Lenz microphthalmia-2, oculofaciocardiodental syndrome	XLD
219	ATP6AP2	Xp11.4	X-linked ID with epilepsy	XLR
220	MAOA	Xp11.3	Brunner syndrome (Monoamine oxidase A deficiency)	XLR
221	PORCN	Xp11.23	Focal dermal hypoplasia	XLD
222	SYP	Xp11.23	Non-syndromic X-linked ID	XLD
223	SHROOM4	Xp11.22	Stocco dos Santos X-linked ID syndrome, non-syndromic X-linked ID	XLR
224	HUWE1	Xp11.22	Non-syndromic X-linked ID (duplications, XL recessive); syndromic X-linked ID, Turner type (point mutations, XL dominant)	XLD/XLR
225	ARHGEF9	Xq11.1	Syndromic X-linked ID, hyperekplexia and epilepsy	XLR
226	DLG3	Xq13.1	Non-syndromic X-linked ID	XLR
227	HDAC8	Xq13.1	Cornelia de Lange syndrome; Wilson-Turner syndrome	XLD
228	SLC16A2	Xq13.2	T3 transporter deficiency; syndromic and non-syndromic ID	XLD
229	ATP7A	Xq21.1	Menkes disease, occipital horn syndrome	XLR
230	PGK1	Xq21.1	Phosphoglycerate kinase-1 deficiency	XLR
231	BRWD3	Xq21.1	Non-syndromic X-linked ID	XLR
232		Xq21.1	Non-syndromic X-linked ID	XLR
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233	TIMM8A	Xq22.1	Mohr-Tranebjaerg syndrome, Jensen syndrome	XLR
234	RAB40AL	Xq22.1	Syndromic X-linked ID, Martin-Probst type	XLR
235	PLP1	Xq22.2	Pelizaeus-Merzbacher disease	XLR
236	PRPS1	Xq22.3	Phosphoribosylpyrophosphate synthetase superactivity, Arts syndrome	XLR
237	PAK3	Xq22.3	Non-syndromic X-linked ID	XLR
238	UBE2A	Xq24	Syndromic X-linked ID, Nascimento-type	XLR
239	NDUFA1	Xq24	Mitochondrial complex I deficiency (syndromic X-linked ID)	XLD
240	CUL4B	Xq24	Syndromic X-linked ID, Cabezas type	XLR
241	ZDHHC9	Xq25	Syndromic X-linked ID, Raymond type; non-syndromic X-linked ID	XLD
	GPC3			XLR
242		Xq26.2	Simpson-Golabi-Behmel syndrome type 1	
243	HPRT1	Xq26.2	Lesch-Nyhan syndrome	XLR
244	SOX3	Xq27.1	Isolated GH deficiency, short stature and ID	XLR
245	IDS	Xq28	Mucopolysaccharidosis II (Hunter syndrome)	XLR
246	NSDHL	Xq28	CK syndrome (XL recessive); CHILD syndrome (Congenital Hemidysplasia with Ichthyosiform Nevus and Limb Defects) (XL dominant)	XLR/XLD
247	ABCD1	Xq28	Adrenoleukodystrophy	XLD
248	HCFC1	Xq28	Non-syndromic X-linked ID	XLR
249	FLNA		Bilateral periventricular nodular heterotopia, otopalatodigital syndrome, frontometaphyseal dysplasia	XLD
		Xq28		
250	GDI1	Xq28	Non-syndromic X-linked ID	XLD
251	IKBKG	Xq28	Incontinentia pigmenti	XLD
252	DKC1	Xq28	Dyskeratosis congenita	XLR

Note that all the ASD genes listed in **Table S6A** are also involved in ID.

Abbreviations: AD, autosomal dominant; AR, autosomal recessive; ID, intellectual disability; XL, X-linked; XLD, X-linked dominant; XLR, X-linked recessive

Table S6D. Loci implicated in ID

	Disorder	Chr. band	Start (hg18)	End (hg18)	Genes involved	Inheritance pattern
1	Thrombocytopenia-absent radius (TAR) syndrome	1q21.1	144,110,432	144,305,571	RBM8A	AR
2	3p deletion syndrome	3p26.3p25.3	1	9,000,000	contiguous gene syndrome?	AD
3	Proximal 15q25.2 microdeletion	15q25	80,451,495	82,719,635	?	AD
4	ATR-16 syndrome (alpha-thalassemia/mental retardation syndrome)	16p13.3	30,843	774,373	?	AD
5	17p13.1 microdeletion syndrome	17p13.1	7,429,371	7,937,620	?	AD
6	19p13.13 microdeletion/microduplication syndrome	19p13.13	12,793,474	13,104,643	?	AD
7	Cat-Eye syndrome	22p13-22q11.21	15,772,953	16,971,860	?	AD
8	22q11.2 distal deletion syndrome	22q11.2	20,445,848	22,026,229	?	AD
9	Pelizaeus-Merzbacher disease	Xq22.2	102,918,095	102,934,203	PLP1	XLR

Note that all the ASD loci listed in **Table S6B** are also involved in ID.
Abbreviations: AD, autosomal dominant; AR, autosomal recessive; ID, intellectual disability; XLR, X-linked recessive

Table S7A. CNVs overlapping ASD or ID genes and loci in affected and control subjects (all ancestries)

Cases

AGP ID	Sex	Chr#:start-end (hg18)	Size	CNV type	Inheri- tance	Classification	Validation	CNV description
16035_1571013001	М	1:144482933-146325557	1,842,625	Gain	dn	Pathogenic	qPCR	1q21.1 duplication syndrome
8635_201	М	1:144500467-146377870	1,877,404	Loss	dn	Pathogenic	qPCR	1q21.1 deletion syndrome
1952_301	М	1:144500467-146336720	1,836,254	Gain	dn	Pathogenic	qPCR	1q21.1 duplication syndrome
13135_1523	F	1:144838594-146308287	1,469,694	Gain	dn	Pathogenic	qPCR	1q21.1 duplication syndrome
4291_1	М	1:144967972-146317915	1,349,944	Gain	mat	Pathogenic	qPCR	1q21.1 duplication syndrome
16074_1571042001	М	2:50037898-50407550	369,653	Loss	mat	Pathogenic	qPCR	NRXN1 exonic deletion
14068_1180	М	2:50493827-50677835	184,009	Gain	dn	Pathogenic	qPCR	NRXN1 intragenic duplication, predicted to result in a premature truncated protein
13017_223	F	2:50539877-50730546	190,670	Loss	dn	Pathogenic	qPCR	NRXN1 exonic deletion
13216_2383	М	2:50968208-51214171	245,964	Loss	dn	Pathogenic	qPCR	NRXN1 exonic deletion
13153_1703	М	2:50990306-51222043	231,738	Loss	dn	Pathogenic	qPCR	NRXN1 exonic deletion
13037_463	М	2:51002576-51157742	155,167	Loss	dn	Pathogenic	qPCR	NRXN1 exonic deletion
17027_1	М	2:51076611-51147600	70,990	Loss	pat	Pathogenic	qPCR	NRXN1 exonic deletion
5353_3	F	6:33399849-33512042	112,194	Loss	dn	Pathogenic	qPCR	SYNGAP1 exonic deletion
5386_3	М	6:156785155-158489874	1,704,720	Loss	dn	Pathogenic	qPCR, Illumina 1M	ARID1B exonic deletion
8446_201	М	7:72344426-73782113	1,437,688	Loss	dn	Pathogenic	qPCR	Williams syndrome (7q11.23 deletion)
13123_1403	F	9:98998-3682923	3,583,926	Loss	dn	Pathogenic	qPCR	Terminal 9p deletion, 3.58 Mb (14 genes)
6259_3	М	9:139516033-140208462	692,430	Loss	dn	Pathogenic	qPCR	Kleefstra syndrome (9q34.3 deletion including EHMT1)
6325_3	М	11:70077507-70506315	428,809	Loss	dn	Pathogenic	qPCR	SHANK2 exonic deletion
6319_3	М	11:70119917-70187872	67,956	Loss	dn	Pathogenic	qPCR	SHANK2 exonic deletion
5237_3	М	11:70154458-70220632	66,175	Loss	dn	Pathogenic	qPCR, Agilent 1M	SHANK2 exonic deletion
6240_4	М	11:126633939-132060374	5,426,436	Loss	dn	Pathogenic	qPCR	Chromosome 11q deletion syndrome (Jacobsen syndrome)
20187_1464001	М	15:18811937-26209270	7,397,334	Gain	dn	Pathogenic	qPCR	15q11-q13 duplication syndrome, maternally derived
8630_201	М	15:19800798-26209270	6,408,473	Gain	dn	Pathogenic	qPCR	15q11-q13 duplication syndrome, paternally derived
20069_1328001	М	15:20203578-26209270	6,005,693	Gain	dn	Pathogenic	qPCR	15q11-q13 duplication syndrome, maternally derived
17035_1	F	15:20274130-26120360	5,846,231	Gain	dn	Pathogenic	qPCR	15q11-q13 duplication syndrome, maternally derived
8117_202	М	15:21168391-26217954	5,049,564	Gain	mat	Pathogenic	qPCR	15q11-q13 duplication syndrome, maternally derived
8741_201	F	15:21168391-26315093	5,146,703	Gain	mat	Pathogenic	qPCR	15q11-q13 duplication syndrome, maternally derived
13050_593	М	15:21190624-26203954	5,013,331	Gain	dn	Pathogenic	qPCR	15q11-q13 duplication syndrome, maternally derived
1950_301	M	15:26762141-30436163	3,674,023	Loss	mat	Pathogenic	Illumina 550K	15q13.3 deletion syndrome
16040_1571029001	M	15:28450423-30303265	1,852,843	Loss	dn	Pathogenic	qPCR	15q13.3 deletion syndrome
14167_2720	M	15:28705540-30436163	1,730,624	Loss	pat	Pathogenic	qPCR	15q13.3 deletion syndrome
18100_302	M	15:28714502-30303265	1,588,764	Loss	pat	Pathogenic	qPCR	15q13.3 deletion syndrome
5537_3	М	15:82722026-83529838	807,813	Loss	mat	Pathogenic	Affy 500K, Illumina 2.5M, qPCR	Distal 15q25.2 deletion syndrome
14283_4060	М	16:14771033-16307313	1,536,281	Loss	mat	Pathogenic	qPCR	16p13.11 deletion syndrome
3441_3	M	16:14808871-16215852	1,406,982	Loss	pat	Pathogenic	qPCR	16p13.11 deletion syndrome
14412_5210	M	16:14960247-16307313	1,347,067	Loss	mat	Pathogenic	qPCR, LR-PCR	16p13.11 deletion syndrome
2204_1	M	16:29466569-30147029	680,461	Loss	dn	Pathogenic	qPCR	16p11.2 deletion syndrome
3544_3	M	16:29499858-30107306	607,449	Gain	mat	Pathogenic	_	16p11.2 duplication syndrome
5068_3	F	16:29502984-30127026	624,043	Loss	dn	Pathogenic	qPCR, Affy 500K, Agilent 1M	16p11.2 deletion syndrome, 50% mosaicism
20089_1391001	М	16:29502984-30107306	604,323	Loss	dn	Pathogenic	qPCR	16p11.2 deletion syndrome
3211_3	М	16:29502984-30127026	624,043	Gain	mat	Pathogenic	qPCR	16p11.2 duplication syndrome
5262_4	М	16:29502984-30210849	707,866	Gain	dn	Pathogenic	qPCR, Affy 500K	16p11.2 duplication syndrome
5359_4	М	16:29554843-30195224	640,382	Loss	dn	Pathogenic	qPCR, Affy 500K, Agilent 1M	16p11.2 deletion syndrome
20127_4014001	M	16:29554843-30130862	576,020	Loss	pat	Pathogenic	qPCR	16p11.2 deletion syndrome
4030_1	M	16:29554843-30107306	552,464	Gain	dn	Pathogenic	qPCR	16p11.2 duplication syndrome
3439_3	М	17:17156307-18262979	1,106,673	Loss	dn	Pathogenic	qPCR	Smith-Magenis syndrome (17p11.2 deletion including RA/1)
2211_1	۲.	17:17169258-20101517	2,932,260	Loss	dn	Pathogenic	qPCR	Smith-Magenis syndrome (17p11.2 deletion including RAI1)
14315_4320	M	17:31621634-33323919	1,702,286	Gain	mat	Pathogenic	qPCR	17q12 duplication syndrome
3183_7	M	22:17241748-19819918	2,578,171	Loss	dn	Pathogenic	qPCR	22q11.2 deletion syndrome
17015_1	M	22:17257787-19795780	2,537,994	Loss	dn	Pathogenic	qPCR	22q11.2 deletion syndrome
3127_4	M	22:17257787-19793730	2,535,944	Gain	pat	Pathogenic	qPCR	22q11.2 duplication syndrome
4271_1	М	22:17257787-19795780	2,537,994	Gain	dn	Pathogenic	qPCR	22q11.2 duplication syndrome
5261_4	F	22:17257787-19795780	2,537,994	Gain	pat	Pathogenic	qPCR, Illumina 1M	22q11.2 duplication syndrome
16074_1571042001	М	22:17257787-19793730	2,535,944	Gain	pat	Pathogenic	qPCR	22q11.2 duplication syndrome
8627_201	M	22:17257787-19793730	2,535,944	Gain	dn	Pathogenic	qPCR	22q11.2 duplication syndrome
2072_1	М	22:45159185-49582267	4,423,083	Loss	dn	Pathogenic	qPCR	Phelan-McDermid syndrome (22q13 deletion including SHANK3)
6130_4	F	22:47996161-49512530	1,516,370	Loss	dn	Pathogenic	qPCR, MLPA	Phelan-McDermid syndrome (22q13 deletion including SHANK3)
16079_1571066001	М	22:49470371-49567383	97,013	Loss	dn	Pathogenic	qPCR	Phelan-McDermid syndrome (22q13 deletion including SHANK3)
5240_4	М	X:23116188-23280628	164,441	Loss	mat	Pathogenic	qPCR, Illumina 1M	PTCHD1 exonic deletion
5126_4	М	X:28931559-29478966	547,408	Gain	mat	Pathogenic	qPCR, Agilent 1M	IL1RAPL1 intragenic duplication of exons 3-5
8597_201	М	X:31303978-32025062	721,085	Loss	mat	Pathogenic	qPCR	DMD deletion of exons 45-60
3019_3	М	X:32100618-32315937	215,320	Gain	mat	Pathogenic	qPCR	DMD duplication of exons 31-44
20013_1075001	М	X:153239048-153521797	282,750	Gain	dn	Pathogenic	qPCR, LR-PCR	Xq28 duplication encompassing 20 genes, including 3 involved in ID: FLNA, GDI1, IKBKG. Corresponds to the recurrent Xq28 duplication reported in XLID families; 55 GDI1 is the most
14216_3470	M	X:153263157-153474401	211,245	Gain	mat	Pathogenic	qPCR	likely candidate gene (sequencing coordinates: chrX:153322048-153514311, size 292 kb) Xq98 duplication encompassing 18 genes, including 2 involved in ID: GDI1 and IKBKG. Recurrent Xq28 duplication involved in XLID (see above)
	М	1:2118508-2325536	207,029	Loss	dn	Uncertain	qPCR	Deletion encompassing 5 genes, including SKI, involved in Shprintzen-Goldberg syndrome
6356_5	IVI							through dominant negative mutations
6356_5 6317_5	M	1:26767083-26941756	174,674	Gain	pat	Uncertain	qPCR	through dominant negative mutations  ARID1A partial duplication; ARID1A mutations reported recently in Coffin-Siris syndrome

AGP ID	Sex	Chr#:start-end (hg18)	Size	CNV type	Inheri- tance	Classification	Validation	CNV description
3424_3	М	2:148881443-149078468	197,026	Gain	mat	Uncertain	qPCR	MBD5 partial duplication; MBD5 is implicated in autosomal dominant ID through deletions (2q23.1 deletion syndrome) and mutations
3599_3	F	6:155906594-157336808	1,430,215	Gain	pat	Uncertain-likely benign	qPCR	ARID1B partial duplication; haploinsufficiency of ARID1B causes ID and Coffin-Siris syndrome
13037_463	М	9:137682721-137840339	157,619	Loss	mat	Uncertain	_	Deletion encompassing 4 genes, including KCNT1; gain-of-function KCNT1 mutations cause epilepsy
16072_1571036001	М	11:381049-1019320	638,272	Gain	pat	Uncertain	qPCR	Duplication encompassing 35 genes, including HRAS (involved in Costello syndrome through activating mutations); although duplication of HRAS is not expected to be pathogenic, the contribution of other genes in the interval is unknown
20070_1331001	M	16:14808871-15935225	1,126,355	Gain	mat	Uncertain	qPCR	16p13.11 microduplication, significance unknown. Recurrent 16p13.11 deletions are associated with variable phenotype and incomplete penetrance, and have been reported in subjects with diverse neuropsychiatric disorders, including ID, ASD, schizophrenia and epilepsy, sometimes inherited from unaffected parents. Duplications have been described in neurodevelopmental disorders and in controls, with studies reporting either no enrichment or a small but significant enrichment in cases. Further studies in larger case populations and controls are needed to clarify their role as risk factors
9766_202	М	16:15032942-16199484	1,166,543	Gain	mat	Uncertain	qPCR	16p13.11 microduplication (see above)
14142_2400	М	16:15387380-16256106	868,727	Gain	pat	Uncertain	qPCR	16p13.11 microduplication (see above)
5258 3	М	16:15387380-16270740	883,361	Gain	pat	Uncertain	qPCR, Illumina 1M	16p13.11 microduplication (see above)
4182 1	М	16:15387380-16199484	812,105	Gain	mat	Uncertain	_	16p13.11 microduplication (see above)
2265 1	М	16:15387380-18176669	2,789,290	Gain	mat	Uncertain	qPCR	16p13.11 microduplication (see above)
8703_201	М	17:1092256-1249222	156,967	Gain	pat	Uncertain-likely benign	•	Duplication of 3 genes, partially overlapping YWHAE; whole duplications are pathogenic (17p13.3 duplication syndrome); there is an AGP control with a YWHAE partial duplication; several partial duplications reported in DGV
5444_3	М	17:76953064-77782267	829,204	Gain	dn	Uncertain	qPCR	829 kb duplication encompassing 38 genes, including ACTG1 (involved in Baraitser-Winter syndrome through dominant-negative or gain-of-function missense mutations); although duplication of ACTG1 is not expected to be pathogenic, the contribution of the other genes in the interval is unknown
14242_3660	F	22:21995356-22598120	602,765	Gain	mat	Uncertain	_	602 kb duplication encompassing 15 genes, including SMARCB1; SMARCB1 mutations reported recently in Coffin-Siris syndrome
6381_3	М	X:30521738-30789831	268,094	Gain	mat	Uncertain	qPCR	Duplication of 2 genes, partially overlapping GK; GK mutations and deletions cause XLID
5126_4	М	X:32948977-33330592	381,616	Gain	mat	Uncertain	Agilent 1M	DMD duplication of exon 1 of the Dp427 transcripts; the effect of this duplication is difficult to predict; experimental evidence at the RNA level is required to interpret the significance
4356_1	М	X:38001148-38346471	345,324	Gain	mat	Uncertain-likely benign	_	Duplication of RPGR, OTC, and TSPAN7; similar duplication found in an AGP male control.  OTC mutations and deletions cause ornithine transcarbamylase deficiency; TSPAN7 duplications are found in healthy controls
4152_1	М	X:40260354-40372806	112,453	Gain	mat	Uncertain	_	Duplication encompassing 3 genes, including ATP6AP2; ATP6AP2 silent mutation affecting splicing described in XLID with epilepsy
3240_3	М	X:44706702-44919064	212,363	Gain	mat	Uncertain	_	Duplication of 2 genes, partially overlapping KDM6A partial duplication; KDM6A mutations and deletions cause X-linked Kabuki syndrome
14314_4310	М	X:70865247-71509736	644,490	Gain	mat	Uncertain	qPCR	644 kb duplication encompassing 13 genes, partially overlapping HDAC8; HDAC8 mutations described recently in Cornelia de Lange syndrome and XLID resembling Wilson-Turner syndrome; similar partial duplication of HDAC8 observed in a male control
1348 301	М	X:147163528-147758700	595,173	Gain	mat	Uncertain	qPCR	AFF2 partial duplication; involved in XLID through trinucleotide expansion or deletion
5036_4	М	X:148075334-148617551	542,218	Gain	mat	Uncertain-likely benign		542 kb duplication encompassing 9 genes, including IDS; mutations and deletions cause mucopolysaccharidosis type II; duplication reported in unaffected males. Although duplication of IDS is not expected to be pathogenic, the contribution of the other genes in the interval is unknown. The duplication is also present in the proband's unaffected
4354_1	М	X:148344051-148707925	363,875	Gain	mat	Uncertain-likely	_	brother, suggesting this CNV is unlikely to be pathogenic 363 kb duplication encompassing 9 genes, including [DS; mutations and deletions cause
4166_1	М	X:151620401-151805387	184,987	Gain	mat	benign Uncertain	_	mucopolysaccharidosis type II; duplication reported in unaffected males (see above) Xq28 duplication of 10 genes, including NSDHL; mutations and deletions cause syndromic XLID (CK syndrome and CHILD syndrome); similar duplication including NSDHL in a male AGP control
9901_201	М	3:12608293-12786824	178,532	Gain	pat	Benign	_	Duplication of 2 genes, partially overlapping RAF1; RAF1 is involved in Noonan syndrome through activating mutations; similar duplication present in 2 AGP controls
20087_1386001	М	3:12610706-12786824	176,119	Gain	mat	Benign	-	Duplication of 2 genes, partially overlapping RAF1; RAF1 is involved in Noonan syndrome through activating mutations; similar duplication present in 2 AGP controls
4457_1	М	X:38375788-38515190	139,403	Gain	mat	Benign	_	TSPANT partial duplication; mutations and deletions cause XLID, duplications are found in healthy controls
5524_3	М	X:38375788-38515190	139,403	Gain	mat	Benign	_	TSPAN7 partial duplication (see above)
6034_3	М	X:38375788-38515190	139,403	Gain	mat	Benign	_	TSPAN7 partial duplication (see above)
20033_1227001	М	X:38375788-38515190	139,403	Gain	mat	Benign	_	TSPAN7 partial duplication (see above)
20141_1396001	М	X:38375788-38515190	139,403	Gain	mat	Benign	qPCR	TSPAN7 partial duplication (see above)
5089_5	M	X:53555568-53640902	85,335	Gain	mat	Benign	_	HUWE1 partial duplication. Whole gene duplications cause non-syndromic XLID; similar recurrent partial duplications of HUWE1 reported recently, considered polymorphic 133
17018_1	М	X:53568262-53640902	72,641	Gain	mat	Benign	qPCR	HUWE1 partial duplication (see above)
Controls								

ID	Sex	Chr#:start-end (hg18)	Size	CNV type	Classification	CNV description
HABC_902399_902399	F	1:144614719-146470277	1,855,559	Loss	"Pathogenic"	1q21.1 deletion syndrome
B436528_1007852654	M	1:144627859-146546371	1,918,513	Loss	"Pathogenic"	1q21.1 deletion syndrome
B984152_1007842480	F	1:144800611-145863421	1,062,811	Gain	"Pathogenic"	1q21.1 duplication syndrome
B618929_1007875266	M	1:144933825-145518117	584,293	Gain	"Pathogenic"	1q21.1 duplication syndrome
B666224_1007871687	М	1:144967972-145863421	895,450	Gain	"Pathogenic"	1q21.1 duplication syndrome
HABC_902895_902895	F	2:50829989-51064129	234,141	Loss	"Pathogenic"	NRXN1 exonic deletion
B964957_1007872180	F	7:72344426-73782113	1,437,688	Gain	"Pathogenic"	7q11.23 duplication syndrome (Williams syndrome region)
B914224_1007874975	М	10:89620404-89723400	102,997	Loss	"Pathogenic"	PTEN exonic deletion
HABC_902475_902475	М	15:36401817-36434987	33,171	Loss	"Pathogenic"	SPRED1 deletion; gene involved in Legius syndrome (phenotypic overlap with neurofibromatosis 1); most individuals don't have ID and only present dermatologic findings
B675955_1007841005	F	16:29415871-30239704	823,834	Gain	"Pathogenic"	16p11.2 duplication syndrome
B416484_1007875540	M	16:29502984-30127026	624,043	Loss	"Pathogenic"	16p11.2 deletion syndrome
B879700_1007854073	F	16:29502984-30127026	624,043	Gain	"Pathogenic"	16p11.2 duplication syndrome
HABC_900681_900681	М	16:29554843-30180288	625,446	Gain	"Pathogenic"	16p11.2 duplication syndrome

ID	Sex	Chr#:start-end (hg18)	Size	CNV type	Classification	CNV description
B121881_1007874637	М	17:31621634-33323919	1,702,286	Gain	"Pathogenic"	17q12 duplication syndrome
HABC_901636_901636	М	22:17248170-19795780	2,547,611	Gain	"Pathogenic"	22q11.2 duplication syndrome
B928258_1007854097	F	22:17257787-18693299	1,435,513	Gain	"Pathogenic"	22q11.2 duplication syndrome
110036016178_	М	2:148753482-148819104	65,623	Loss	Uncertain	MBD5 exonic deletion; deletions of MBD5 cause autosomal dominant ID, but this one overlaps only the long isoform, which has not been fully characterized and contains 5 additional non-coding exons (this deletion overlaps 1 of the non-coding exons)
B183736_1007853714	F	8:117910382-117948637	38,256	Gain	Uncertain-likely benign	1 of the non-coding exons)  RAD21 partial duplication; mutations and deletions cause Cornelia de Lange syndrome
HABC_900405_900405	М	8:117910403-117948935	38,533	Gain		RAD21 partial duplication; mutations and deletions cause Cornelia de Lange syndrome
HABC_900854_900854	M	9:2018757-2080718	61,962	Loss		SMARCA2 deletion of exons 2-19, removes the translation start site in exon 2; mutations resulting in Nicolaides- Baraitser syndrome are thought to act through a dominant—negative or gain-of-function manner and cluster in exons 15–25; deletions encompassing SMARCA2 do not cause this syndrome, except for one reported in-frame deletion overlapping the mutation-clustering region. The deletion in the AGP control involves exons 2-19 and removes the translation start site in exon 2.
B116679_1007853952	М	9:129475725-129875601	399,877	Gain	Uncertain	Duplication encompassing 19 genes, partially overlapping STXBP1; mutations and deletions of STXBP1 cause nonsyndromic ID with epilepsy and infantile epileptic encephalopathy
B246752_1007872634	F	9:139616009-139725155	109,147	Gain	Uncertain	Duplication encompassing 3 genes, partially overlapping EHMT1; EHMT1 mutations and deletions cause Kleefstra syndrome
B252606_1007874475	М	12:1889583-2538831	649,249	Gain	Uncertain	Duplication encompassing 4 genes, partially overlapping CACNA1C, involved in Timothy syndrome through activating mutations
B936611_1007853579	F	12:25087733-25286046	198,314	Gain	Uncertain	Duplication encompassing 4 genes, partially overlapping KRAS; KRAS is involved in cardio-facio-cutaneous syndrome through activating mutations
B978305_1007874920	М	13:109744730-110078003	333,274	Gain	Uncertain	$\label{eq:condition} Duplication encompassing 4 genes, partially overlapping \textit{COL4A1}; only \textit{COL4A1} missense mutations reported thus far, no deletions or duplications$
B777599_1007853701	F	14:101564897-101608061	43165	Gain	Uncertain	DYNC1H1 partial duplication; all mutations identified so far (ID with neuronal migration defects and motor neuropathies) are heterozygous missense mutations, suggesting a dominant-negative effect
HABC_900744_900744	F	15:28723577-30232287	1,508,711	Gain	Uncertain	15q13.3 microduplication
HABC_901557_901557	F	15:28730804-30389965	1,659,162	Gain	Uncertain	15q13.3 microduplication
B833125_0057060983		16:1995854-2052977	57,124	Gain	Uncertain	Duplication encompassing 5 genes, partially overlapping TSC2; TSC2 mutations and deletions cause tuberous sclerosis  1.0.1.3.1. microdunication
HABC_902940_902940 HABC_901197_901197	M F	16:14831165-16199484	1,368,320	Gain	Uncertain Uncertain	16p13.11 microduplication
		16:14882793-16199484 16:14977368-16190572	1,316,692	Gain		16p13.11 microduplication
HABC_902864_902864	M	16:14977368-16190572	1,213,205 2,688,545	Gain Gain	Uncertain Uncertain	16p13.11 microduplication 16p13.11 microduplication
HABC_902897_902897 HABC_901863_901863	F	16:87658525-87896679	238,155	Gain	Uncertain	CDH15 duplication and ANKRD11 partial duplication; CDH15 is involved in non syndromic ID through mutations
HABC 901829 901829	F	16:87729306-87962759	233,454	Gain	Uncertain	and ANKRD11 in KBG syndrome through mutations and deletions  CDH15 duplication and ANKRD11 partial duplication (see above)
HABC_902145_902145	F	17:1136077-1211814	75,738	Gain		Duplication of 2 genes, partially overlapping YWHAE; YWHAE whole gene duplications are pathogenic; several
HABC 902046 902046	М	17:2437871-2476338	38,468	Gain	benign Uncertain	partial duplications in DGV PAFAH1B1 partial duplication; whole gene duplications are pathogenic
HABC_901640_901640	М	22:21320275-23360745	2,040,471	Gain	Uncertain	2 Mb duplication encompassing 44 genes, including <i>SMARCB1</i> ; <i>SMARCB1</i> mutations reported in Coffin-Siris syndrome
B278753_1007874641	М	22:21995356-22676385	681,030	Gain	Uncertain	681 kb duplication encompassing 20 genes, including SMARCB1; SMARCB1 mutations reported in Coffin-Siris syndrome
HABC_900402_900402	F	X:11038333-11069582	31,250	Gain	benign	Duplication of 2 genes, including HCCS; HCCS mutations and deletions cause a syndromic form of XLID
HABC_902971_902971	F	X:13633067-13700254	67,188	Gain	benign	Duplication encompassing 4 genes, including OFD1; mutations and deletions cause syndromic XLID
HABC_901634_901634	М	X:17239813-17435795	195,983	Gain	benign	NHS partial duplication; mutations and deletions cause Nance-Horan syndrome
HABC_902647_902647	М	X:18574793-18780863 X:38013482-38643203	206,071	Gain	benign	Xp22.13 duplication encompassing 3 genes, partially overlapping CDKL5; CDKL5 mutations and deletions cause epileptic encephalopathy  Duplication encompassing 4 genes, including OTC and TSPAN7. OTC mutations and deletions cause ornithine
HABC_900416_900416	М	A.36013462-36043203	629,722	Gain	Uncertain-likely benign	transcarbamylase deficiency; no cases have been reported with a duplication. <i>TSPANT</i> mutations and deletions cause XLID, duplications are found in healthy controls
B345605_1007844543	М	X:71239825-71490721	250,897	Gain	Uncertain-likely benign	Duplication encompassing 8 genes, partially overlapping HDAC8; HDAC8 mutations described recently in Cornelia de Lange syndrome and XLID
HABC_900366_900366	F	X:76924341-77030430	106,090	Gain	benign	Partial duplication of ATRX and MAGT1; ATRX is involved in XLID through mutations or deletions
HABC_900333_900333	F	X:134903813-135264655	360,843	Gain	Uncertain	Xq26.3 duplication encompassing 4 genes, partially overlapping <i>SLC9A6</i> ; mutations and deletions cause syndromic XLID (Christianson syndrome)
B418695_1007840289	М	X:151644548-151839695	195,148	Gain	Uncertain	Xq28 duplication encompassing 10 genes, including NSDHL; NSDHL mutations and deletions cause XLID (CK syndrome and CHILD syndrome); several overlapping duplications reported in DGV
HABC_902725_902725	F	X:153340432-153435070	94,639	Gain	benign	95 kb duplication encompassing 7 genes, partially overlapping <i>IKBKG</i> ; only mutations, deletions and intragenic duplications described
HABC_902313_902313 HABC_901807_901807	M	X:153440007-153622054 3:12608293-12781123	182,048	Gain	benign Benign	182 kb duplication encompassing 7 genes, partially overlapping <i>IKBKG</i> ; only mutations, deletions and intragenic duplications described Duplication encompassing 2 genes, partially overlapping <i>RAF1</i> ; <i>RAF1</i> is involved in Noonan syndrome through
B260038_1007841400	F	3:12610706-12781123	170,418	Gain	Benign	Duplication encompassing 2 genes, partially overlapping NAF1; NAF1 is involved in Noonan syndrome through Duplication encompassing 2 genes, partially overlapping RAF1; RAF1 is involved in Noonan syndrome through
B630497 1007872229	F	12:2085709-2127756	42,048	Loss	Benign	activating mutations  CACNA1C deletion; CACNA1C causes Timothy syndrome through activating mutations
B131548_1007842659	F	12:2663790-2714138	50,349	Gain	Benign	CACNA1C partial duplication; CACNA1C causes Timothy syndrome through activating mutations
HABC_902939_902939	F	12:2675893-2717980	42,088	Gain	Benign	CACNA1C partial duplication; CACNA1C causes Timothy syndrome through activating mutations
110036016517_	М	X:38375788-38515190	139,403	Gain	Benign	TSPAN7 partial duplication; mutations and deletions cause XLID, duplications are found in healthy controls
B818627_1007854359	М	X:38375788-38515190	139,403	Gain	Benign	TSPAN7 partial duplication; mutations and deletions cause XLID, duplications are found in healthy controls

This table shows the CNVs overlapping ASD/ID genes and loci interpreted as pathogenic, uncertain or benign in cases and controls of all ancestries. Phenotype information and CNV segregation in siblings can be found in **Table S8**. In order to compare the burden of CNVs overlapping ASD/ID genes and loci in cases and controls (**Figures 1** and **S1**), CNVs were interpreted irrespective of affected status. A small number of CNVs that would have been considered pathogenic in an affected individual were identified among controls; all were CNVs known to be associated with incomplete penetrance/variable expressivity.

Abbreviations: DGV, Database of Genomic Variants; dn, *de novo*; F, female; ID, intellectual disability; LR-PCR, long range PCR; M, male; mat, maternal; MLPA, multiplex ligation-dependent probe amplification; pat, paternal; qPCR, quantitative PCR; XLID, X-linked intellectual disability. —, no validation attempted, CNV confirmed by visual inspection.

Table S7B. Pathogenic CNVs in affected subjects (all ancestries)
Pathogenic CNVs overlapping ASD/ID genes or loci (stringent CNV, >30 kb) n = 64

AGP ID	Sex	Family type	Cytoband	Chr#:start-end (hg18)	Size	CNV type	Inheritance	CNV description	Classification penetrance/ expressivity §
16035_1571013001	М	Familial	1q21.1	1:144482933-146325557	1,842,625	Gain	De novo	1q21.1 duplication syndrome	VE/IP
1952_301	М	Familial	1q21.1	1:144500467-146336720	1,836,254	Gain	De novo	1q21.1 duplication syndrome	VE/IP
8635_201	М	Familial	1q21.1	1:144500467-146377870	1,877,404	Loss	De novo	1q21.1 deletion syndrome	VE/IP
13135_1523	F	Unknown	1q21.1	1:144838594-146308287	1,469,694	Gain	De novo	1q21.1 duplication syndrome	VE/IP
4291_1	М	Familial	1q21.1	1:144967972-146317915	1,349,944	Gain	Maternal	1q21.1 duplication syndrome	VE/IP
16074_1571042001†	М	Sporadic	2p16.3	2:50037898-50407550	369,653	Loss	Maternal	NRXN1 exonic deletion	VE/IP
14068_1180	М	Sporadic	2p16.3	2:50493827-50677835	184,009	Gain	De novo	NRXN1 intragenic duplication, predicted to result in a premature truncated protein	VE/IP
13017_223	F	Sporadic	2p16.3	2:50539877-50730546	190,670	Loss	De novo	NRXN1 exonic deletion	VE/IP
13216_2383	M	Unknown	2p16.3	2:50968208-51214171	245,964	Loss	De novo	NRXN1 exonic deletion	VE/IP
13153_1703	M	Sporadic	2p16.3	2:50990306-51222043	231,738	Loss	De novo	NRXN1 exonic deletion	VE/IP
13037_463	M	Sporadic	2p16.3	2:51002576-51157742	155,167	Loss	De novo	NRXN1 exonic deletion	VE/IP
17027_1	M	Familial	2p16.3	2:51076611-51147600	70,990	Loss	Paternal	NRXN1 exonic deletion	VE/IP
5353_3	F	Sporadic	6p21.32	6:33399849-33512042	112,194	Loss	De novo	SYNGAP1 exonic deletion	HP
5386_3	M	Familial	6q25.3	6:156785155-158489874	1,704,720	Loss	De novo	ARID1B exonic deletion	HP
8446_201	М	Sporadic	7q11.23	7:72344426-73782113	1,437,688	Loss	De novo	Williams syndrome (7q11.23 deletion)	HP
13123_1403	F	Sporadic	9p24.3-p24.2	9:98998-3682923	3,583,926	Loss	De novo	Terminal 9p deletion, 3.58 Mb (14 genes)	HP
6259_3	М	Sporadic	9q34.3	9:139516033-140208462	692,430	Loss	De novo	Kleefstra syndrome (9q34.3 deletion including EHMT1)	HP
6325_3	М	Sporadic	11q13.3	11:70077507-70506315	428,809	Loss	De novo	SHANK2 exonic deletion <sup>129</sup>	HP
6319 3	М	Sporadic	11q13.3	11:70119917-70187872	67,956	Loss	De novo	SHANK2 exonic deletion 117,129	HP
5237 3	М	Sporadic	11q13.3-q13.4	11:70154458-70220632	66,175	Loss	De novo	SHANK2 exonic deletion 117,134	HP
6240 4	M	Sporadic	11q24.2-q25	11:126633939-132060374	5,426,436	Loss	De novo	Chromosome 11q deletion syndrome (Jacobsen syndrome)	HP
20187_1464001	M	Sporadic	15q11.2-q13.1	15:18811937-26209270	7,397,334	Gain	De novo	15q11-q13 duplication syndrome, maternally derived	VE/IP
8630 201	M	Familial	15q11.2-q13.1 15q11.2-q13.1	15:19800798-26209270	6,408,473	Gain	De novo	15q11-q13 duplication syndrome, maternally derived	VE/IP VE/IP
20069_1328001	M	Sporadic	15q11.2-q13.1	15:20203578-26209270	6,005,693	Gain	De novo	15q11-q13 duplication syndrome, maternally derived	VE/IP
17035_1	F	Sporadic	15q11.2-q13.1	15:20274130-26120360	5,846,231	Gain	De novo	15q11-q13 duplication syndrome, maternally derived	VE/IP
8117_202	М	Familial	15q11.2-q13.1	15:21168391-26217954	5,049,564	Gain	Maternal	15q11-q13 duplication syndrome, maternally derived	VE/IP
8741_201	F	Familial	15q11.2-q13.1	15:21168391-26315093	5,146,703	Gain	Maternal	15q11-q13 duplication syndrome, maternally derived	VE/IP
13050_593	М	Sporadic	15q11.2-q13.1	15:21190624-26203954	5,013,331	Gain	De novo	15q11-q13 duplication syndrome, maternally derived	VE/IP
1950_301	M	Familial	15q13.1-q13.3	15:26762141-30436163	3,674,023	Loss	Maternal	15q13.3 deletion syndrome	VE/IP
16040_1571029001	M	Familial	15q13.2-q13.3	15:28450423-30303265	1,852,843	Loss	De novo	15q13.3 deletion syndrome	VE/IP
14167_2720	М	Sporadic	15q13.2-q13.3	15:28705540-30436163	1,730,624	Loss	Paternal	15q13.3 deletion syndrome	VE/IP
18100_302	М	Familial	15q13.2-q13.3	15:28714502-30303265	1,588,764	Loss	Paternal	15q13.3 deletion syndrome	VE/IP
5537 3	М	Familial	15q13.2-q13.3	15:82722026-83529838	807,813	Loss	Maternal	Distal 15q25.2 deletion syndrome	VE/IP
14283_4060	М	Sporadic	16p13.11	16:14771033-16307313	1,536,281	Loss	Maternal	16p13.11 deletion syndrome	VE/IP
3441 3	М	Sporadic	16p13.11	16:14808871-16215852	1,406,982	Loss	Paternal	16p13.11 deletion syndrome	VE/IP
14412_5210	M	Sporadic	16p13.11	16:14960247-16307313	1,347,067	Loss	Maternal	16p13.11 deletion syndrome	VE/IP
2204 1	M	Sporadic	16p11.2	16:29466569-30147029		Loss	De novo	16p11.2 deletion syndrome	VE/IP
		· ·	· ·		680,461			•	
3544_3	M	Familial	16p11.2	16:29499858-30107306	607,449	Gain	Maternal	16p11.2 duplication syndrome	VE/IP
20089_1391001	M	Sporadic	16p11.2	16:29502984-30107306	604,323	Loss	De novo	16p11.2 deletion syndrome	VE/IP
5068_3	F	Familial	16p11.2	16:29502984-30127026	624,043	Loss	De novo	16p11.2 deletion syndrome, 50% mosaicism	VE/IP
5262_4	М	Sporadic	16p11.2	16:29502984-30210849	707,866	Gain	De novo	16p11.2 duplication syndrome	VE/IP
3211_3	M	Familial	16p11.2	16:29502984-30127026	624,043	Gain	Maternal	16p11.2 duplication syndrome	VE/IP
5359_4	M	Sporadic	16p11.2	16:29554843-30195224	640,382	Loss	De novo	16p11.2 deletion syndrome	VE/IP
20127_4014001	M	Sporadic	16p11.2	16:29554843-30130862	576,020	Loss	Paternal	16p11.2 deletion syndrome	VE/IP
4030_1	M	Familial	16p11.2	16:29554843-30107306	552,464	Gain	De novo	16p11.2 duplication syndrome	VE/IP
3439_3	М	Sporadic	17p11.2	17:17156307-18262979	1,106,673	Loss	De novo	Smith-Magenis syndrome (17p11.2 deletion including RAI1)	HP
2211_1	F	Sporadic	17p11.2	17:17169258-20101517	2,932,260	Loss	De novo	Smith-Magenis syndrome (17p11.2 deletion including RAI1)	HP
14315_4320	М	Sporadic	17q12	17:31621634-33323919	1,702,286	Gain	Maternal	17q12 duplication syndrome	VE/IP
3183_7	М	Familial	22q11.21	22:17241748-19819918	2,578,171	Loss	De novo	22q11.2 deletion syndrome	VE/IP
17015 1	М	Sporadic	22q11.21	22:17257787-19795780	2,537,994	Loss	De novo	22q11.2 deletion syndrome	VE/IP
8627 201	М	Sporadic	22q11.21	22:17257787-19793730	2,535,944	Gain	De novo	22q11.2 duplication syndrome	VE/IP
4271 1	M	Sporadic	22q11.21	22:17257787-19795780	2,537,994	Gain	De novo	22q11.2 duplication syndrome	VE/IP
16074 1571042001†	M	Sporadic	22q11.21 22q11.21	22:17257787-19793780	2,535,944	Gain	Paternal	22q11.2 duplication syndrome	VE/IP
		· ·							
3127_4	М	Familial	22q11.21	22:17257787-19793730	2,535,944	Gain	Paternal	22q11.2 duplication syndrome	VE/IP
5261_4	F	Familial	22q11.21	22:17257787-19795780	2,537,994	Gain	Paternal	22q11.2 duplication syndrome	VE/IP
2072_1	M	Familial	22q13.31-q13.33	22:45159185-49582267	4,423,083	Loss	De novo	Phelan-McDermid syndrome (22q13 deletion including SHANK3) <sup>130,135</sup>	HP
6130_4	F	Sporadic	22q13.32-q13.33	22:47996161-49512530	1,516,370	Loss	De novo	Phelan-McDermid syndrome (22q13 deletion including SHANK3)	HP
16079_1571066001	М	Sporadic	22q13.33	22:49470371-49567383	97,013	Loss	De novo	Phelan-McDermid syndrome (22q13 deletion including SHANK3)	HP
5240_4	М	Familial	Xp22.11	X:23116188-23280628	164,441	Loss	Maternal	PTCHD1 exonic deletion 136,137	— (sex chr)
5126_4	М	Familial	Xp21.3	X:28931559-29478966	547,408	Gain	Maternal	IL1RAPL1 intragenic duplication of exons 3-5	— (sex chr)
8597_201	М	Sporadic	Xp21.2-p21.1	X:31303978-32025062	721,085	Loss	Maternal	DMD deletion of exons 45-60	— (sex chr)
3019_3	М	Familial	Xp21.1	X:32100618-32315937	215,320	Gain	Maternal	DMD duplication of exons 31-44 (sequencing coordinates: chrX:32097213-32321945, size: 224733 bp) <sup>138</sup>	— (sex chr)
20013_1075001	M	Sporadic	Xq28	X:153239048-153521797	282,750	Gain	De novo	Xq28 duplication encompassing 20 genes, including 3 involved in ID: FLNA, GDI1, IKBKG. Recurrent Xq28 duplication implicated in XLID; GDI1 is the most likely candidate gene <sup>55</sup> (sequencing coordinates: chrX:153,222,048-153,514,311, size 292 kb)	— (sex chr)
14216_3470	М	Sporadic	Xq28	X:153263157-153474401	211,245	Gain	Maternal	Xq28 duplication encompassing 18 genes, including 2 involved in ID: GDI1 & IKBKG. Recurrent Xq28 duplication implicated in XLID; GDI1 is the most likely candidate gene	— (sex chr)

Other pathogenic CNVs, including large *de novo* CNVs as well as CNVs not included in the main analyses (chromosomal abnormalities, CNVs <30 kb, non stringent CNVs or CNVs identified with 1 algorithm only and validated), n = 20

AGP ID	Sex	Family type	Cytoband	Chr#:start-end (hg18)	Size	CNV type	Inheritance	CNV description	Classification penetrance/ expressivity §
8658_201	F	Sporadic	1p21.3-p21.2	1:98175622-100923952	2,748,331	Loss	De novo	2.7 Mb 1p21.3-p21.2 deletion, <i>de novo</i> (18 genes, including <i>DPYD</i> and <i>MIR137</i> ). 1p21.3 microdeletions comprising <i>DPYD</i> and <i>MIR137</i> have been reported in subjects with ID	— (novel region)
5467_3	М	Sporadic	1q42.3-q44	1:233476547-247165725	13,689,179	Gain	De novo	13.7 Mb 1q42.3-q44 duplication, <i>de novo</i> (114 genes). This CNV overlaps the critical region of the 1q43-q44 deletion syndrome. Distal duplications of the long arm of chromosome 1 are rare, reported in few individuals in its pure form (chromosome abnormality)	НР
5236_3	F	Familial	2p16.3	2:50705521-50719594	14,074	Loss	Maternal	NRXN1 exonic deletion (CNV is exonic according to RefSeq, but not to UCSC) (<30 kb, non stringent: QSNP PCNV)	VE/IP
5328_3	М	Sporadic	2p16.3	2:51044181-51120644	76,464	Loss	Paternal	NRXN1 exonic deletion (non stringent: QSNP PCNV)	VE/IP
16037_1571015001	M	Sporadic	2q37.3	2:239765200-239777909	12,710	Loss	Maternal	HDAC4 small intragenic deletion. HDAC4 is responsible for some of the features of the 2q37 deletion syndrome (brachydactyly mental retardation syndrome). There are many 2q37 deletions reported in individuals with ASD, but no mutations or single gene deletions of HDAC4 had been reported in ASD. (sequencing coordinates: chr2:239766528-239778481, size 11954 bp) (<30 kb)	VE/IP
21020_1‡	М	Sporadic	4p16.3-p16.1	4:53403-9016339	8,962,937	Gain	De novo		
21020_1‡	М	Sporadic	8p23.3-p23.1	8:154984-6994825	6,839,842	Loss	De novo	De novo unbalanced translocation leading to 4p16.3-p16.1 duplication and 8p23.3-p23.1 deletion (see above). The 8p terminal deletion spans 6.8 Mb; numerous terminal deletions of 8p have been described in the literature in subjects with ID, behavioral issues and mild dysmorphic features (chromosome abnormality)	НР
14270_3930	F	Sporadic	6q25.3-q27	6:160773919-170761395	9,987,477	Gain	De novo	10 Mb duplication 6q25.3-q27, de novo, 46,XX.ish der(22)t(6;22)(6q25.3;p11.2)pat(6qtel+). "6q duplication syndrome" is a rare cytogenetic abnormality reported in few individuals in its pure form; the duplications can affect any part of the 6q arm and few overlap. Most of the reported cases are the result of abnormal segregation of a balanced translocation carried by a parent, like in this case (chromosome abnormality)	НР
8404_201	М	Familial	6q27	6:169136788-170761395	1,624,608	Loss	De novo	1.6 Mb <i>de novo</i> deletion in 6q27 region (14 genes). Terminal 6q deletion syndrome. The CNV contains <i>DLL1</i> , an ASD candidate gene: <i>de novo</i> frameshift variant identified in ASD exome study <sup>4</sup>	НР
13137_1543	F	Sporadic	8p12-8q12.1	8:31928590-58996070	27,067,481	Gain	De novo	27 Mb pericentromeric duplication 8p12-8q12.1, <i>de novo</i> . The karyotype revealed a mosaic supernumerary ring chromosome (47, XX, +r[10]/46, XX[70]) of unknown origin, shown to involve chr 8 by SNP array. Trisomy 8 syndrome is characterized by mild to severe mental and growth deficiency, facial dysmorphisms, and limb abnormalities. In cases of supernumerary marker chromosomes or supernumerary ring chromosomes derived from chr 8, mosaic or non-mosaic, the clinical presentation varies from normal phenotype to features overlapping the trisomy 8 syndrome. (chromosome abnormality)	НР
8534_201	М	Sporadic	10q11.21-q11.23	10:45633089-51564756	5,931,668	Loss	De novo	5.9 Mb 10q11.22-q11.23 deletion (56 genes), de novo. Recurrent deletions in this region are associated with variable clinical features, with ID as the only feature present in the majority of individuals; most deletions are inherited from apparently normal parents, indicating variable expressivity/incomplete penetrance <sup>139</sup>	VE/IP
4312_1	М	Sporadic	10q11.21-q11.23	10:45550419-51496386	5,945,968	Loss	Paternal	5.9 Mb 10q11.22-q11.23 deletion (56 genes), paternal. Recurrent deletion (see above)	VE/IP
6053_3	М	Familial	12q13.3-q14.1	12:54218922-58779615	4,560,694	Gain	De novo	4.5 Mb <i>de novo</i> duplication in 12q13 (101 genes); no similar CNV reported	— (novel region)
14070_1230	М	Familial	15q26.1	15:91200007-91283004	82,998	Loss	De novo	CHD2 exonic deletion. CHD2 de novo mutations reported recently in epileptic encephalopathy and ID; several deletions reported in individuals with ID, ASD and epilepsy	НР
5420_3*	М	Sporadic	Whole chr 21	21:1-247249719	247,249,719	Gain	De novo	Down syndrome (47,XY+21) (chromosome abnormality)	НР
14291_4120	F	Sporadic	22q13.33	22:49470371-49480446	10,076	Loss	De novo	SHANK3 deletion of exons 9-13 (<30 kb, called with 1 algorithm only: PCNV) (sequencing coordinates: 49468716-49485255, size 16540 bp)	НР
5241_3	М	Familial	Xp21.1	X:31793278-31822704	29,427	Loss	Maternal	DMD deletion of exon 48 (predicted to lead to an in-frame deletion) (<30 kb)	— (sex chr)
9861_202	М	Sporadic	Xp11.4	X: 41248675-41259467	10,793	Gain	Maternal	CASK partial duplication of the 5' UTR of exon 1, decreased mRNA expression in cell line (<30 kb, called with 1 algorithm only: PCNV)	— (sex chr)
5257_3*	М	Sporadic	Whole chr Y	Y:1-57772954	57,772,954	Gain	De novo	XYY syndrome (chromosome abnormality)	— (sex chr)
5515_3*	М	Sporadic	Whole chr Y	Y:1-57772954	57,772,954	Gain	De novo	XYY syndrome (chromosome abnormality)	— (sex chr)

This table includes pathogenic CNVs overlapping ASD/ID genes or loci as well as other pathogenic CNVs in cases of all ancestries. The latter included chromosome abnormalities (>7.5 Mb) that had been excluded from the main analyses, selected large rare *de novo* events, as well as experimentally validated smaller CNVs (<30 kb) or CNVs not considered stringent (called by at least one algotihm only).

With the exception of the 16p11.2 duplication found in proband 3544\_3 (note tested), all the CNVs reported in this table were experimentally validated (see details in Table S8).

<sup>§</sup> For the analysis shown in Figure 2E, autosomal pathogenic CNVs were classified as highly penetrant (HP) or associated with variable expressivity and/or incomplete penetrance (VE/IP). CNVs on sex chromosomes or affecting novel regions were not classified for this analysis.

<sup>†</sup> Proband 16074\_1571042001 carries two pathogenic CNV: a NRXN1 exonic deletion inherited from his mother and a 22q11.2 duplication inherited from his father.

<sup>‡</sup> Proband 21020\_1 has a de novo unbalanced translocation leading to 4p duplication and 8p deletion, both considered pathogenic.

<sup>\*</sup> Three cases showed whole chromosome aneuploidies: one case with Down syndrome and two with XYY syndrome.

Abbreviations: F, female; ID, intellectual disability; M, male; UTR, untranslated region; XLID, X-linked intellectual disability

# Table S8. Phenotypes in ASD subjects with pathogenic CNVs or with selected CNVs of uncertain significance

mmc2.xlsx (Excel workbook)

This file also contains information on CNV validation and segregation in siblings, when available.

Table S9. Meta-analysis of loci and genes affected by rare CNVs in large ASD cohorts

Locus/gene	Cas	. <b>GP, Sta</b> .es (185 .ols (124	3 M, 29	94 F)		Ss ases (96 ntrols (4			Case	AGRE ° s (n=1835)* =837; 755 M)**	Case	nbined AGP + es (2821 M, 45 or 837; 755 ontrols (1644 I	50 F + 1 M**)	.835*	AGP+	mbined SSC+AGRE Del	Combined AGP+SSC+AGRE Dup	Del P-value†	Dup P-value†	Del Freq %	Dup Freq %
	Cas	ses	Con	trols	Ca	ses	Con	trols		Cases	(	Cases	Cor	ntrols	Cases	/Controls	Cases/Controls				
	Del	Dup	Del	Dup	Del	Dup	Del	Dup	Del	Dup	Del	Dup	Del	Dup							
16p11.2	5 (4)	4 (2)	-	2	8 (7)	6 (4)	-	-	3 (3)*	2 (0) *	16 (14)	12 (6)	-	2	<b>16</b> /5106;	0/3512	<b>12</b> /5106; 2/3512	2.28E-04	3.52E-02	0.313	0.235
15q13.3 (BP3-BP5, BP4-BP5)#	4 (1)	-	-	1	2 (1)	1 (1)	-	-	2 (0)*	1 (0)*	8 (2)	2 (1)	-	1	<mark>8</mark> /5106;	0/3512	2/5106; 1/3512	1.52E-02	0.637	0.157	0.039
16p13.11#	3 (0)	5 (0)	-	2	1 (0)	2 (1)	1	3	3 (0)*	4 (0; 1 unk)**	7 (0)	11 (1; 1 unk)	1	5	<mark>7</mark> /5106;	1/3512	11/4108; 5/3512	9.86E-02	0.174	0.137	0.268
15q11q13 (BP1-BP3, BP2-BP3)\$	-	6 (5)	-	-	-	1 (1)	-	-	_	6 (2)*	-	13 (8)	-	-	0		13/5106; 0/3512	_	1.10E-03	_	0.255
22q11.2 (DiGeorge syndrome)	2 (2)	5 (2)	-	1	1 (1)	-	-	-	_	3 (1)*	3 (3)	8 (3)	-	1	<mark>3</mark> /5106;	0/3512	8/5106; 1/3512	0.208	6.46E-02	0.059	0.157
17p11.2 (Smith-Magenis syndrome)	2 (2)	-	-	-	-	-	-	-	_	_	2 (2)	_	-	-	2/4108**	'; 0/3512	0	0.291	_	0.049	_
9q34.3 (Kleefstra syndrome)	1 (1)	-	-	-	1 (1)	-	-	-	_	_	2 (2)	_	-	-	2/4108**	'; 0/3512	0	0.291	_	0.049	_
1q21.1	1 (1)	3 (2)	-	1	-	3 (2)	-	-	1 (1)*	2 (0)*	2 (2)	8 (4)	-	1	<mark>2</mark> /5106;	0/3512	8/5106; 1/3512	0.351	6.46E-02	0.039	0.157
17q12	-	1 (0)	-	-	2 (1)	-	-	-	_	_	2 (1)	1 (0)	-	-	<mark>2</mark> /5106;	0/3512	1/5106; 0/3512	0.351	0.593	0.039	0.020
7q11.23 (Williams syndrome)	1 (1)	-	-	-	_	4 (4)	-	-	_	_	1(1)	4 (4)	-	-	<b>1</b> /4108;	0/3512	4/4108; 0/3512	0.539	8.44E-02	0.024	0.097
																		_	_	_	_
PTCHD1/PTCHD1AS (Xp22.11)‡+	8 (1)	-	1	-	3 (1)	-	1	-	1 (0)**	_	12 (2)	_	2	-	12/3576**	; 2/1644 M	0	0.133	_	0.336	_
NRXN1 (2p16.3)	6 (4)	1 (1)§	1	-	3 (1)	-	1	-	4 (0)**	_	13 (5)	1 (1)	2	-	13/4108**	*; 2/3512	1/4108; 0/3512	8.50E-03	0.539	0.316	0.024
SHANK3 (22q13.33)&	4 (4)	-	-	-	-	-	-	-	-	_	4 (4)	-	-	-	4/4108**	°; 0/3512	0	8.44E-02	-	0.097	_
SHANK2 (11q13.3)	3 (3)	-	-	-	-	-	-	-	_	1 (0)**	3 (3)	1 (0)	-	-	3/4108**	'; 0/3512	1/4108; 0/3512	0.157	0.539	0.073	0.024
NLGN3 (Xq13.1) ‡	-	-	-	-	1 (0)	-	-	-	_	-	1 (0)	_	-	-	1/3576**	; 0/1644 M	0	0.685	_	0.028	_
NLGN4X (Xp22.3) ‡	_	-	-	-	-	1 (0)	-	-	_	1 (1 unk)**	-	2 (1 unk)	-	-	0		2/3576; 0/1644 M	_	0.469	_	0.056

CNVs are ordered according to frequency. Numbers in parentheses indicate *de novo* events. For *NRXN1*, *SHANK2*, *SHANK3*, *NLGN3* and *NLGN4X* CNVs are only counted in cases and controls if they affect one or more exons; chromosome X events are only counted for males.

\*Number of AGRE cases in Moreno-de-Luca et al.<sup>111</sup> (n=1835 cases). A few of the regions in this table were not listed in Moreno-de-Luca et al.'s paper; we obtained the counts of subjects with deletions/duplications in these additional regions directly from the authors - the total number of inspected cases after quality control for the additional regions was 837 unrelated probands from 1105 families (or a total of 1472 all affected): 161 families with multiple males and females affected children, 594 families with only one male affected child and 82 families with only one female affected child. The total number of AGRE families with at least one male affected child was 161+82=243. The combined number of cases used to get estimates for this table was: i) 5106 = (AGP+SSC) + AGRE= 3271+1835 or ii) 4108 (\*\*) = 3271+837.

Abbreviations: AGP, Autism Genome Project; AGRE, Autism Genetic Resource Exchange; BP, breakpoint; Del, deletion; Dup, duplication; F, female; Freq, frequency; M, male; SSC, Simons Simplex Collection; unk. unknown.

<sup>†</sup> Fisher exact test, one-sided P-value

<sup>#</sup> Duplications of the 15q13.3 and 16p13.11 regions are of uncertain clinical significance, because they have not been found to be consistently enriched in cases compared to controls.

Sof the 13 duplications of the 15q11q13 region, 5 were maternally inherited (1 AGP, 4 AGRE), and 8 were de novo. Of the 8 de novo duplications, 5 were of maternal origin (4 AGP, 1 SSC), two were of paternal origin (AGP: 8630\_201 and AGRE: AU1135202), and one without information (1 AGRE: AU1042303).

<sup>§</sup> NRXN1 intragenic duplication, predicted to result in a premature truncated protein.

<sup>‡</sup> Only males counted. The combined number of males used to get estimates for this table was 3576 = (AGP+SSC) + AGRE= 2821+755

<sup>+</sup> The two de novo deletions in this locus were found in male probands from an SSC family (12561) and AGP family (subject 5240 4)

<sup>&</sup>lt;sup>8</sup> Includes SHANK3 gene exonic deletions as well as 22q13 deletion syndrome (Phelan-McDermid syndrome).

<sup>&</sup>lt;sup>a</sup> Ref <sup>117</sup> + Stage 2 (this study) (only subjects of European ancestry included)

<sup>&</sup>lt;sup>b</sup> Ref <sup>132</sup>

c Ref 111

Table S10. FMRP targets affected by deletions in probands and not yet implicated in ASD or ID

FMRP targets not yet implicated in ASD or ID						
ABR	MAPK8IP3					
AGTPBP1	MBP					
ARHGAP32	NBEA					
BAI1	NRXN3					
BRSK1	PCDH9					
CAMSAP1	PLCB1					
DTNA	PLEC					
EML2	PRKACB					
EP400	PTPRD					
FAM115A	PTPRT					
FAM21A	R3HDM1					
FAM91A1	RALGAPA1					
FAT4	RALGDS					
GRM5	RPRD2					
KCNH1	SHANK1					
KIAA0430	SPTB					
KIAA0913	TCF25					
LLGL1	TRIP12					
LPHN3	TRPM3					
LPPR4	ULK2					
LYNX1	UNC13C					
MADD	a					

Of the FMRP targets<sup>9</sup> (n=842, one without correspondence in our annotation file) affected by exonic deletions in ASD subjects, 58 genes correspond to known ASD/ID genes (**Tables S6A-S6D**). This table lists the remaining 43 FMRP targets not yet implicated in ASD or ID, considered ASD candidate genes.

## Tables S11A-S11E. Multigene analyses – various models

Table S11A. Primary analysis

	Estimate	Std. Error	z value	Pr(> z )					
Bandal tonation CBN/s with in				(-  - /					
Model treating CNVs within									
(Intercept)	-0.195271	0.027722	-7.044	1.87e-12***					
factor(CNV)Dup	-0.120601	0.039779	-3.032	0.00243**					
ngene	0.030912	0.005768	5.359	8.37e-08***					
RPKM	0.057893	0.024207	2.392	0.01677*					
Model treating CNVs within the same individual as independent, and analyzing deletions only (pseudo R <sup>2</sup> 0.007)									
(Intercept)	-0.21920	0.02979	-7.359	1.85e-13***					
ngene	0.02857	0.01030	2.773	0.005549**					
RPKM	0.14595	0.04153	3.515	0.000441***					
Model treating CNVs within	the same individual	as independent, ar	d analyzing dup	lications only (pseudo R <sup>2</sup> 0.008)					
(Intercept)	-0.282964	0.036126	-7.833	4.78e-15***					
ngene	0.031361	0.006981	4.492	7.04e-06***					
RPKM	0.011110	0.029966	0.371	0.711					
Model with the number of g	genes and the averag	e RPKM nested wit	hin deletion or c	<b>Juplication status</b> (pseudo R <sup>2</sup> 0.008)					
(Intercept)	-0.219196	0.029785	-7.359	1.85e-13***					
factor(CNV)Dup	-0.063768	0.046821	-1.362	0.173217					
factor(CNV)Del:ngene	0.028566	0.010301	2.773	0.005549**					
factor(CNV)Dup:ngene	0.031361	0.006981	4.492	7.04e-06***					
factor(CNV)Del:RPKM	0.145950	0.041528	3.515	0.000441***					
factor(CNV)Dup:RPKM	0.011110	0.029966	0.371	0.710813					

Abbreviations: ngene, number of genes; RPKM, reads per kb per million reads

In our primary analysis we treated CNVs falling within the same individual as independent (main **Figures 3C-D** and **Table S11A**). **Figures 3C-D** show the pattern of increased burden with the increased number of brain-expressed genes affected (1-hit, 2-hit, 3-hit, 4- to 10-hit) by deletions or duplications. The percentage of cases and controls with CNVs overlapping genes is shown for deletions and duplications separately. To account for the fact that some individuals carry a multiplicity of rare CNVs, we also performed analyses counting all genes in all rare CNVs per individual, as well as the brain expression of those genes, and entered those in the model (**Table S11B**); and we evaluated the impact of the largest CNV and its gene expression. Results from these alternative models were similar, showing that the dependence due to CNVs occurring in the same individual has only a minor impact on inference. For each model, Nagelkerke's pseudo-R<sup>2</sup> was calculated to provide a description of how well the model fitted the data.

In **Figure 3C-D** (main text), the expected odds ratios (depicted by stars) were estimated by fitting a logit model of the case status (case/control) with CNV type and three covariates, the deletion/duplication status, the number of genes covered by each CNV and their average brain expression value for the genes covered by the CNV. Gene level expression values for the neocortex were obtained from the BrainSpan RNA sequencing resource, transformed to log(1+RPKM), and the average of the transformed RPKM value for the genes covered by the CNV was used in the model (i.e. total RPKM of all genes covered by the CNV divided by the total number of genes within the CNV). Differences between the effect of the number of genes and expression values between deletion and duplication CNVs were further evaluated by analyzing each subset of the data separately.

In **Table S11A**, we fit a logit model with the following covariates: deletion/duplication status of the CNV, the number of genes covered by each CNV and the average of the transformed RPKM value (i.e. RPKM value transformed to log(1+RPKM)) for the genes covered by the CNV (total RPKM of all genes covered by the CNV divided by the total number of genes). We ignored the fact that individuals could have multiple CNVs. All effects in the model are significant. There is higher risk for autism when the number of genes covered by the CNV is larger and when the average (transformed) RPKM value is higher. Also notice that the risk for autism is slightly lower for duplications as opposed to deletions.

We also checked whether there were differences between the effect of the number of genes and RPKM between deletions and duplications by analyzing each subset of the data separately. Notice that the increased risk due to the number of covered genes for autism is fairly similar for duplications and deletions, while the increased risk due to an increase of the average RPKM is significant for deletions but not for duplications.

Another way to assess the differences between duplications and deletions would be to fit the effect of the number of genes and the average RPKM nested within deletion and duplication CNV status. The results are virtually the same for this model as the previous split data analysis.

Table S11B. Models accounting for the fact that some individuals carry a multiplicity of rare CNVs

	Estimate	Std. Error	z value	Pr(> z )	
Model considering the to	otal number of genes cov	ered by all deletio	ns or duplicatio	ns in an individual (ngeneDel	,
	age RPKM value for the	genes in deleted or	duplicated regi	ons (mRPKMDel, mRPKMDup	) (pseudo
R <sup>2</sup> 0.016)					
(Intercept)	-0.37032	0.04326	-8.560	<2e-16***	
ngeneDel	0.03292	0.01006	3.273	0.001066**	
ngeneDup	0.03506	0.01010	3.471	0.000519***	
mRPKMDel	0.17159	0.05014	3.422	0.000621***	
mRPKMDup	0.01307	0.03796	0.344	0.730597	
Model when selecting C	NVs within individuals ba	ased on the maxim	um number of g	enes covered by the CNVs (p	seudo R <sup>2</sup>
0.018)				,	
(Intercept)	-0.365583	0.042510	-8.600	<2e-16***	
maxGeneDel	0.035244	0.010939	3.222	0.001273**	
maxGeneDup	0.027772	0.006957	3.992	6.55e-05***	
RPKMDel	0.171068	0.048600	3.520	0.000432***	
RPKMDup	-0.000702	0.036458	-0.019	0.984637	
Model when selecting th	ne CNVs based on the ma	ximum average sta	ndardized RPKI	<b>VI</b> (pseudo R <sup>2</sup> 0.015)	
(Intercept)	-0.36725	0.04326	-8.490	<2e-16***	
nGeneDel	0.03289	0.01173	2.805	0.00503**	
nGeneDup	0.02543	0.00795	3.199	0.00138**	
maxRPKMDel	0.17640	0.04392	4.016	5.91e-05***	
maxRPKMDup	0.01741	0.03205	0.543	0.58705	

To take into account the effect of multiple CNVs per individual we decided to re-analyze the data using a model in which we fit the total number of genes covered by all deletions in an individual (ngeneDel), the total number of genes covered by duplications in an individual (ngeneDup) as well as the average RPKM value for these genes in a deletion region (mRPKMDel) and the average RPKM for those in a duplication region (mRPKMDup) (upper panel). All effects, except the one for the average RPKM of genes covered by duplications (mRPKMDup) were significant. Notice the similarity of the results of this model with the one in which we analyzed the duplications and deletions separately. In both analyses the risk due to the number of genes is similar in duplications and deletions. However, only higher mean RPKM values for deletions increase risk for ASD.

Instead of looking at the average number of genes and average RPKM covered by deletions and duplications, one can also identify the CNVs for which the maximum value is obtained. This table shows the results of the generalized linear model when CNVs are selected within individuals based on the maximum number of genes covered by the CNVs (middle panel), as well as the results obtained when selecting the CNVs based on the maximum average standardized RPKM (lower panel). Notice the great similarity between these two analyses as well as the analysis in which we took the average of multiple CNVs per individual. In all three cases the effect of the number of genes covered by the CNVs is significant and the magnitude of the risk is very similar. Also, for all three analyses, the value of the average RPKM is only significant for deletions.

Table S11C. Removing cases with validated de novo CNVs

	Estimate	Std. Error	z value	Pr(> z )					
Model considering the average	number of gene	s and the average RF	KM of genes co	overed by the CNVs in the remaining					
individuals (pseudo R <sup>2</sup> 0.005)									
(Intercept)	-0.343950	0.043740	-7.863	3.74e-15***					
ngeneDel	0.010159	0.011975	0.848	0.39625					
ngeneDup	0.024238	0.010586	2.290	0.02204*					
mRPKMDel	0.137438	0.051433	2.672	0.00754**					
mRPKMDup	0.006056	0.038418	0.158	0.87474					
Model considering the CNVs with the maximum number of genes covered, in the remaining individuals (pseudo R <sup>2</sup> 0.006)									
(Intercept)	-0.343072	0.043122	-7.956	1.78e-15***					
maxGeneDel	0.010173	0.013176	0.772	0.44005					
maxGeneDup	0.020914	0.007687	2.721	0.00651**					
RPKMDel	0.139397	0.049723	2.803	0.00506**					
RPKMDup	-0.004947	0.036953	-0.134	0.89351					
Model considering the CNVs wi	th the maximum	RPKM, in the remai	ning individuals	s (pseudo R <sup>2</sup> 0.006)					
(Intercept)	-0.342025	0.043733	-7.821	5.25e-15***					
nGeneDel	0.009579	0.014296	0.670	0.50285					
nGeneDup	0.020248	0.008438	2.399	0.01642*					
maxRPKMDel	0.131945	0.045620	2.892	0.00382**					
maxRPKMDup	0.001850	0.032626	0.057	0.95477					

To assess the impact of validated *de novo* CNVs we removed 77 cases that had at least one validated *de novo* CNV. (Note that the total dataset contained 90 validated *de novo* CNVs in 87 cases of European ancestry; of these, 85 *de novo* CNVs in 82 unique European cases had sizes ≥30 kb; individuals with chromosomal abnormalities were excluded from the main analyses). Because of the nature of the control data, *de novo* CNV status was not available in the controls.

Results for the analysis of the average number of genes and the average RPKM of genes covered by the CNVs in the remaining individuals showed that the average number of genes covered by deletions is no longer significant (upper panel). In this analysis, only the average number of genes covered by duplications and the average RPKM of genes covered by deletions are significant. A similar pattern of results is found when analyzing the CNVs with the maximum number of genes covered (middle panel), and the CNVs selected based on the maximum RPKM (lower panel). The results in this table show that removing only 77 out of 1,914 cases, or 4% of the case sample, results in a large decrease in signal, suggesting that most of the risk traces to de novo CNVs.

Table S11D. Removing subjects with CNVs considered pathogenic

	Estimate	Std. Error	z value	Pr(> z )					
	ge number of gene	s and the average	RPKM of genes	covered by the CNVs in the remaining					
individuals (pseudo R <sup>2</sup> 0.004)									
(Intercept)	-0.329996	0.043973	-7.505	6.17e-14***					
ngeneDel	0.006021	0.012452	0.484	0.62871					
ngeneDup	0.015398	0.011366	1.355	0.17551					
mRPKMDel	0.159530	0.051078	3.123	0.00179**					
mRPKMDup	0.013352	0.038353	0.348	0.72774					
Model considering the CNVs with the maximum number of genes covered, in the remaining individuals (pseudo R <sup>2</sup> 0.005)									
(Intercept)	-0.326948	0.043399	-7.533	4.94e-14***					
maxGeneDel	0.003602	0.013877	0.260	0.795200					
maxGeneDup	0.015129	0.008447	1.791	0.073290 .					
RPKMDel	0.164659	0.049298	3.340	0.000838***					
RPKMDup	-0.001081	0.037003	-0.029	0.976694					
Model considering the CNVs	with the maximum	RPKM, in the remai	ning individuals	(pseudo R <sup>2</sup> 0.005)					
(Intercept)	-0.330966	0.044004	-7.521	5.43e-14***					
nGeneDel	0.004337	0.014825	0.293	0.769840					
nGeneDup	0.011546	0.009555	1.208	0.226890					
maxRPKMDel	0.156664	0.045120	3.472	0.000516***					
maxRPKMDup	0.011904	0.032612	0.365	0.715084					

The AGP list of CNVs was curated to identify a subset that could be considered pathogenic (**Table S7B**). These CNVs were carried by 82 unique individuals, and of these 69 were of European ancestry and thus included in these analyses (missing individuals include other ancestries as well as individuals with chromosomal abnormalities [**Table S1C**], excluded from the main analyses).

After removing the individuals with pathogenic CNVs we reran the same three analyses as before. In general, these analyses show good agreement with the results from the comparable models obtained by removing the validated *de novo* CNVs only. This is not surprising because the two sets of events overlap fairly substantially. Here again it is shown that the value of RPKM for deletion CNVs is the most important irrespective of CNV status (i.e., even after removing both *de novo* and inherited pathogenic CNVs), significant in all three analyses. The magnitude of the risk associated with the deletion RPKMs is again very consistent to what was found in the previous analysis. Altogether these results are consistent with a genetic interaction model where imbalance of multiple genes intersected by rare *de novo* and inherited pathogenic CNVs contributes to risk. Furthermore, our findings are also in line with a previous report suggesting that deletions have larger effects on transcriptional level and contained more genes with altered expression compared to duplications.

Table S11E. Model including sex

	Estimate	Std. Error	z value	Pr(> z )					
Males (pseudo R <sup>2</sup> 0.011)									
(Intercept)	0.23934	0.05445	4.396	1.1e-05***					
nGeneDel	0.03734	0.01417	2.635	0.00842**					
nGeneDup	0.03118	0.01300	2.398	0.01649*					
maxRPKMDel	0.12490	0.06376	1.959	0.05010 .					
maxRPKMDup	0.03293	0.04898	0.672	0.50133					
Females (pseudo R <sup>2</sup> 0.010)									
(Intercept)	-1.69525	0.09428	-17.980	<2e-16***					
nGeneDel	0.04850	0.01853	2.617	0.00886**					
nGeneDup	0.02393	0.01777	1.347	0.17803					
maxRPKMDel	0.18177	0.10508	1.730	0.08366 .					
maxRPKMDup	0.01009	0.07995	0.126	0.89953					
Model selecting the CNV covering the largest number of genes in males (pseudo R <sup>2</sup> 0.013)									
(Intercept)	0.237196	0.053568	4.428	9.51e-06***					
maxGeneDel	0.033687	0.014671	2.296	0.02167*					
maxGeneDup	0.027932	0.009389	2.975	0.00293**					
RPKMDel	0.132569	0.061994	2.138	0.03248*					
RPKMDup	0.027329	0.047174	0.579	0.56238					
Model selecting the CNV cov	ering the largest nu	mber of genes in fe	males (pseudo I	R <sup>2</sup> 0.012)					
(Intercept)	-1.68903	0.09260	-18.240	<2e-16***					
maxGeneDel	0.05820	0.02053	2.835	0.00458**					
maxGeneDup	0.01797	0.01216	1.477	0.13955					
RPKMDel	0.19642	0.10003	1.964	0.04957*					
RPKMDup	-0.01833	0.07737	-0.237	0.81267					
Model selecting the CNV wit	th the largest average	e standardized RPF	(M in males (pse	eudo R <sup>2</sup> 0.011)					
(Intercept)	0.23799	0.05448	4.369	1.25e-05***					
maxGeneDel	0.02668	0.01536	1.737	0.08239 .					
maxGeneDup	0.02680	0.01086	2.467	0.01361*					
maxRPKMDel	0.16081	0.05596	2.874	0.00406**					
maxRPKMDup	0.03207	0.04131	0.776	0.43749					
Model selecting the CNV wit	th the largest average	e standardized RPR	(M in females (p	seudo R <sup>2</sup> 0.011)					
(Intercept)	-1.684587	0.094701	-17.788	<2e-16***					
maxGeneDel	0.065273	0.021786	2.996	0.00273**					
maxGeneDup	0.008598	0.017921	0.480	0.63137					
maxRPKMDel	0.147739	0.094098	1.570	0.11640					
maxRPKMDup	0.019179	0.068047	0.282	0.77806					

We ran the same three models, except this time the data were separated by sex: 1,641 cases and 1,102 controls were male, while 273 cases and 1,257 controls were female. There is general agreement in the parameter estimates between males and females for all three sets of analyses. We also ran tests for significant differences in predictors when they interacted with sex, but none of these interactions were significant (data not shown).

# Tables S12A-S12D. GO terms, pathways, and MPO enrichment in affecteds versus control subjects

mmc3.xlsx (Excel workbook)

# Table S13A. Gene Ontology terms and pathways used to generate a list of neurodevelopmental functions

Gs ID	Gs Name
GO:0007399	Nervous system development
GO:0050877	Neurological system process
GO:0043025	Neuronal cell body
GO:0043005	Neuron projection
KEGG:04725	Cholinergic synapse
KEGG:04724	Glutamatergic synapse
KEGG:04728	Dopaminergic synapse
KEGG:04727	GABAergic synapse
KEGG:04726	Serotonergic synapse
KEGG:04721	Synaptic vesicle cycle
KEGG:04723	Retrograde endocannabinoid signaling
KEGG:05030	Cocaine addiction
KEGG:05031	Amphetamine addiction
KEGG:05032	Morphine addiction
KEGG:05033	Nicotine addiction
KEGG:04722	Neurotrophin signaling pathway
REACT:708	REACT: Neuronal system
REACT:675	REACT: NCAM signaling for neurite out-growth
REACT:138	REACT: Axon guidance
NCI:142	NCI: netrin pathway
NCI:180	NCI: reelin pathway
KEGG:04360	Axon guidance
KEGG:04720	Long-term potentiation
KEGG:04730	Long-term depression

## Table S13B. Effect size for neurobiological-related clusters

#### **B1. FDR 15%**

Logistic regression-FDR 15% <sup>a</sup>	Combined (% all subjects)	Stage 1	Stage 2
Cases	89 (4.70%)	57 (4.65%)	32 (4.79%)
Controls	35 (1.49%)	17 (1.37%)	18 (1.63%)
OR	3.15	3.39	2.94
Cases (max 10 genes/CNV)	56 (2.96%)	37 (3.02%)	19 (2.84%)
Controls (max 10 genes/CNV)	31 (1.32%)	14 (1.13%)	17 (1.54%)
OR (max 10 genes/CNV)	2.24	2.67	1.85

<sup>&</sup>lt;sup>a</sup>REACT: Neuronal system; REACT: Transmission across chemical synapses; KEGG: Glutamatergic synapse; KEGG: Cholinergic synapse; GO: Generation of neurons; GO: Neuron projection morphogenesis; GO: Neuron differentiation; GO: Neuron projection development; GO: Cell morphogenesis involved in neuron differentiation; GO: Axonogenesis; GO: Neuron projection development; GO: Axon guidance.

FDR, false-discovery rate; OR, odds ratio; max-10 genes-CNVs, estimates considering CNVs with maximum 10 genes.

### **B2. FDR 20%**

Logistic regression-FDR 20% <sup>a</sup>	Combined (% all subjects)	Stage 1	Stage 2		
Cases	99 (5.23%)	64 (5.23%)	35 (5.24%)		
Controls	49 (2.09%)	26 (2.10%)	23 (2.08%)		
OR	2.50	3.39	2.51		
Cases (max 10 genes/CNV)	66 (3.49%)	44 (3.59%)	22 (3.29%)		
Controls (max 10 genes/CNV)	45 (1.92%)	23 (1.86%)	22 (1.99%)		
OR (max 10 genes/CNV)	1.82	1.93	1.65		

<sup>&</sup>lt;sup>a</sup>REACT: Neuronal system; REACT: Transmission across chemical synapses; KEGG: Glutamatergic synapse; KEGG: Cholinergic synapse; GO: Generation of neurons; GO: Neuron projection morphogenesis; GO: Neuron differentiation; GO: Neuron development; GO: Cell morphogenesis involved in neuron differentiation; GO: Axonogenesis; GO: Neuron projection development; GO: Axon guidance; REACT: Neurotransmitter receptor binding and downstream transmission in the postsynaptic cell; KEGG: Retrograde endocannabinoid signaling; REACT: Axon guidance; KEGG: Dopaminergic synapse; KEGG: Neurotrophin signaling pathway; GO: Learning or memory.

FDR, false-discovery rate; OR, odds ratio; max-10 genes-CNVs, estimates considering CNVs with maximum 10 genes.

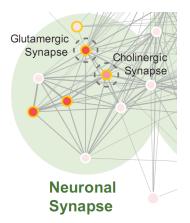
# Table S13C. Neuronal synapse main cluster

# C1. Statistics for all subjects with exonic CNVs (deletions and duplications) at FDR 15%

	Subject N (% all subjects)	Stage 1	Stage 2	
Cases	36 (1.96%)	20 (1.63%)	16 (2.40%)	
Controls	9 (0.38%)	5 (0.40%)	4 (0.36%)	
OR [95% CI]	4.93 [2.31-11.72]	4.04	6.62	

36 cases (25 supporting genes) vs. 9 controls (9 supporting genes). Stage 2 is more enriched, but Stage 1 also displays a respectable signal.

FDR, false-discovery rate; CI, confidence interval; OR: odds ratio



### C2. Detailed statistics for pathways included within the Neuronal synapse cluster at FDR 15%

	All Cases	All Controls	Stage 1 Cases	Stage 1 Controls	Stage 2 Cases	Stage 2 Controls
KEGG Glutamatergic	19 (1.00%)	2 (0.09%)	10 (0.82%)	1 (0.08%)	9 (1.35%)	1 (0.09%)
KEGG Cholinergic	17 (0.90%)	2 (0.09%)	9 (0.74%)	2 (0.16%)	8 (1.20%)	0 (0.00%)
REACT Neuronal System	26 (1.37%)	6 (0.26%)	14 (1.14%)	3 (0.24%)	12 (1.80%)	3 (0.27%)
REACT Synaptic Transmission	21 (1.11%)	3 (0.13%)	11 (0.90%)	2 (0.16%)	10 (1.50%)	1 (0.09%)

Statistics for all subjects with exonic CNVs (deletions and duplications included). Calculation of % of subjects is based on subjects with at least one genic exonic loss; for max-10 this is limited to subjects with a maximum of 10 deleted genes.

# Table S13D. Genes and subjects represented in the enriched cholinergic and glutamergic synapse subclusters

#### D1. Genes

	Cholinergic		ı	Both		Glutamergic	
	Gene name	# cases; # controls	Gene name	# cases; # controls	Gene name	# cases; # controls	
	CHAT*	2; 0 (2*)	_	_	GRIK2	1; 0	
Neurotransmitters (receptors	CHRNA7	4; 0	_	-	GRM5	1; 0	
and metabolism)	KCNJ12	1; 0	_	-	_	_	
	SLC18A3 (VAChT)*	2; 0 (2*)	_	_	_	_	
	-	-	_	-	SHANK1	1; 0	
Scaffolds (synaptic organizers)	-	_	· –	-	SHANK2	3; 0	
	-	_	_	_	SHANK3	3; 0	
	CAMK2G**	1; 0 (1**)	GNG2***	2; 0 (1***)	PPP3CB**	1; 0 (1**)	
Signalling (downstream neurotransmitter receptors and ion channels)	-	_	GNG13	1; 0	_	_	
	-	_	MAPK3***	5; 0 (1***)	_	_	
	-	_	PRKACB	1; 0	-	_	
	_	_	PLCB1	1; 0	_	_	

<sup>26</sup> cases (16 supporting genes) vs. 3 controls (3 genes). Numbers between parentheses represent the number of de novo events.

### D2. ASD subjects

Sample ID	Gene(s) in CNV	CNV inheritance	Biological function	Functional sub-cluster
8534_201	CHAT*; SLC18A3*	dn (5.9 Mb del)		Cholinergic
4312_1	CHAT*; SLC18A3*	pat (5.9 Mb del)		Cholinergic
8465_202	GRIK2	pat (39 kb del)		Glutamergic
8549_201	GRM5	pat (1.98 Mb del)	Neurotransmitters	Glutamergic
3567_4	KCNJ12	pat (324 kb del)	(receptors and	Cholinergic
18100_302	CHRNA7	pat (1.6 Mb del)	metabolism)	Cholinergic
1950_301	CHRNA7	mat (1.7 Mb del)		Cholinergic
14167_2720	CHRNA7	pat (1.7 Mb del)		Cholinergic
16040_157102900	CHRNA7	dn (1.8 Mb del)		Cholinergic
14393_5020	CAMK2G**; PPP3CB**	dn (477 kb del)		Cholinergic; Glutamergic
13204_883	GNG13	unk <sup>\$</sup> (81 kb del)		Both
2204 1	GNG2***	mat (101 kb del)		Both
2204_1	MAPK3***	dn (680 kb del) Signalling (downstream		Both
20057_1290002	GNG2	pat (102 kb del)	— signalling (downstream — neurotransmitter	Both
20089_1391001	МАРКЗ	dn (680 kb del)	- receptors and ion	Both
20127_4014001	МАРКЗ	pat (680 kb del)	– channels)	Both
5359_4	МАРКЗ	dn (680 kb del)	— Citatilleis)	Both
5068_3	МАРКЗ	dn (680 kb del)		Both
5451_3	PRKACB	dn (80 kb del)		Both
5046_3	PLCB1	dn (30 kb del)		Both
5237_3	SHANK2*	dn del		Glutamergic
6319_3	SHANK2*	dn del		Glutamergic
6325_3	SHANK2*	dn del	Caaffalala (aaantia	Glutamergic
16079_1571066001	SHANK3*	dn del	Scaffolds (synaptic	Glutamergic
2072_1	SHANK3*	dn del	<ul><li>organizers)</li></ul>	Glutamergic
6130_4	SHANK3*	dn del		Glutamergic
5340_3	SHANK1	mat del		Glutamergic

<sup>\*</sup> Same gene in two or more samples (recurrent gene); \*\* two genes within the same deletion/same sample (multigene); \*\*\* two genes in two different events in same sample (double-CNV-hit).

<sup>\*\*</sup> Two genes within the same event/same sample (multigene); \*\*\* two genes in two different events in same sample (double-CNV-hit).

<sup>&</sup>lt;sup>\$</sup> Both parental samples failed array QC.

del, deletion; dn, de novo; mat; maternal; pat, paternal; unk, inheritance unknown.

## Tables S14A-S14E. Characterization of genes selected by NETBAG

mmc4.xlsx (Excel workbook)

## Table S15. Functional-group enrichment for DAPPLE results

mmc5.xlsx (Excel workbook)

Table S16. List of 97 high-confidence CNV/SNV genes

Gene symbol	Туре	Gene symbol	Туре	Gene symbol	Туре	Gene symbol	Туре
ABCA1	CNV	DTNA*	CNV	MDM2	LoF-SNV	SKI	CNV
ABL1	CNV	DRP2	LoF XL-SNV in males	MED13L	LoF-SNV	SMARCC2	LoF-SNV
ANK2	LoF-SNV	DST	LoF-SNV	NCKAP1	LoF-SNV	SMC2	CNV
ARHGAP32	CNV	ЕРНВ2	LoF-SNV	NFIA	LoF-SNV	SNRPN	CNV
ARHGDIA	CNV	ERCC6	CNV	NRXN3	CNV	SNX9	CNV
ATOH1	CNV	ESR2	CNV	PARD3	CNV	SOD2	CNV
ATP1B1	LoF-SNV	ETS1	CNV	PARK2	CNV	SPAST	LoF-SNV
BAZ1B	CNV	FLNA	CNV	PAX5	LoF-SNV	SPATA13	LoF-SNV
BCL11A	LoF-SNV	GARNL1	CNV	PIK3CB*	CNV	SREBF1	CNV
BRWD1	LoF-SNV	HAUS7	LoF XL-SNV in males	PIR	LoF XL-SNV in males	STAU2	CNV
CACNA1B	CNV	IKBKG	CNV	PLCB1	CNV	SVIL	LoF-SNV
CAMK2G	CNV	IQGAP2	LoF-SNV	PPM1D	LoF-SNV	SYNCRIP*	CNV
CBX4	LoF-SNV	ITGA5	LoF-SNV	PPP3CB	CNV	SYNJ2	CNV
CDK2	CNV	KAT2B	CNV	PRKAB2	CNV	TAOK2	CNV
CDK4	CNV	KIAA0232	LoF-SNV	PSMB1	CNV	TBC1D23	LoF-SNV
CHD2*	CNV	KLF13	CNV	PSMC2	CNV	TBP	CNV
CNOT3	LoF-SNV	LIMK1	CNV	PTPRD	CNV	TBR1	LoF-SNV
CRKL	CNV	LRP2	LoF-SNV	RAB5A	CNV	TCF3	LoF-SNV
CSDE1	LoF-SNV	MAPK11	CNV	RAC3	CNV	TNK2	CNV
CUBN	LoF-SNV	MAPK12	CNV	RAP1GDS1	CNV	VCP	LoF-SNV
CUL2	CNV	МАРКЗ	CNV	RER1	CNV	ZMYND11	LoF-SNV
CUL3	LoF-SNV	MAPK7	CNV	RIMS1	CNV, LoF-SNV	ZNF292	LoF-SNV
CYFIP1	CNV	МАРК8	CNV	RPS6KA3	LoF-SNV		
DLGAP2	CNV	MAX	CNV	SETD2	LoF-SNV		
DLL1	LoF-SNV	MCF2	LoF XL-SNV in males	SH3GL3	CNV		

This table lists 97 CNV or SNV genes present in the DAPPLE network, but not yet implicated in ASD or ID (i.e. not yet in Tables S6A-S6D), considered high-confidence ASD candidate genes. \*Genes in section 'Highlighted genes'.

## Table S17A. Listing of CNV calls in affected subjects

mmc6.xlsx (Excel workbook)

### Table S17B. Chromosome abnormalities in parents and control subjects

mmc7.xlsx (Excel workbook)

Chromosome abnormalities in probands are listed in Table S1C.

## **Table S17C. Experimentally validated CNVs**

mmc8.xlsx (Excel workbook)

The 456 validated CNVs in **Table S17C** comprise all CNVs validated experimentally across the 2446 ASD cases, including 315 CNVs confirmed in stage 1 samples and 141 CNVs confirmed in stage 2 samples.

### **SUPPLEMENTAL DATAFILES**

Listing of separate Excel workbooks.

Table S8. Phenotypes in ASD subjects with pathogenic CNVs or with selected CNVs of uncertain significance

mmc2.xlsx

Tables S12A-S12D. GO terms, pathways, and MPO enrichment in affecteds versus control subjects

mmc3.xlsx

Tables S14A-S14E. Characterization of genes selected by NETBAG mmc4.xlsx

Table S15. Functional-group enrichment for DAPPLE results mmc5.xlsx

Table S17A. Listing of CNV calls in affected subjects mmc6.xlsx

Table S17B. Chromosome abnormalities in parents and control subjects mmc7.xlsx

Table S17C. Experimentally validated CNVs mmc8.xlsx

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