

The American Journal of Human Genetics, Volume 94

Supplemental Data

**Biallelic Variants in *TTLL5*, Encoding
a Tubulin Glutamylase, Cause Retinal Dystrophy**

Panagiotis I. Sergouniotis, Christina Chakarova, Cian Murphy, Mirjana Becker, Eva Lenassi, Gavin Arno, Monkol Lek, Daniel G. MacArthur, UCL-Exomes Consortium, Shomi S. Bhattacharya, Anthony T. Moore, Graham E. Holder, Anthony G. Robson, Uwe Wolfrum, Andrew R. Webster, and Vincent Plagnol

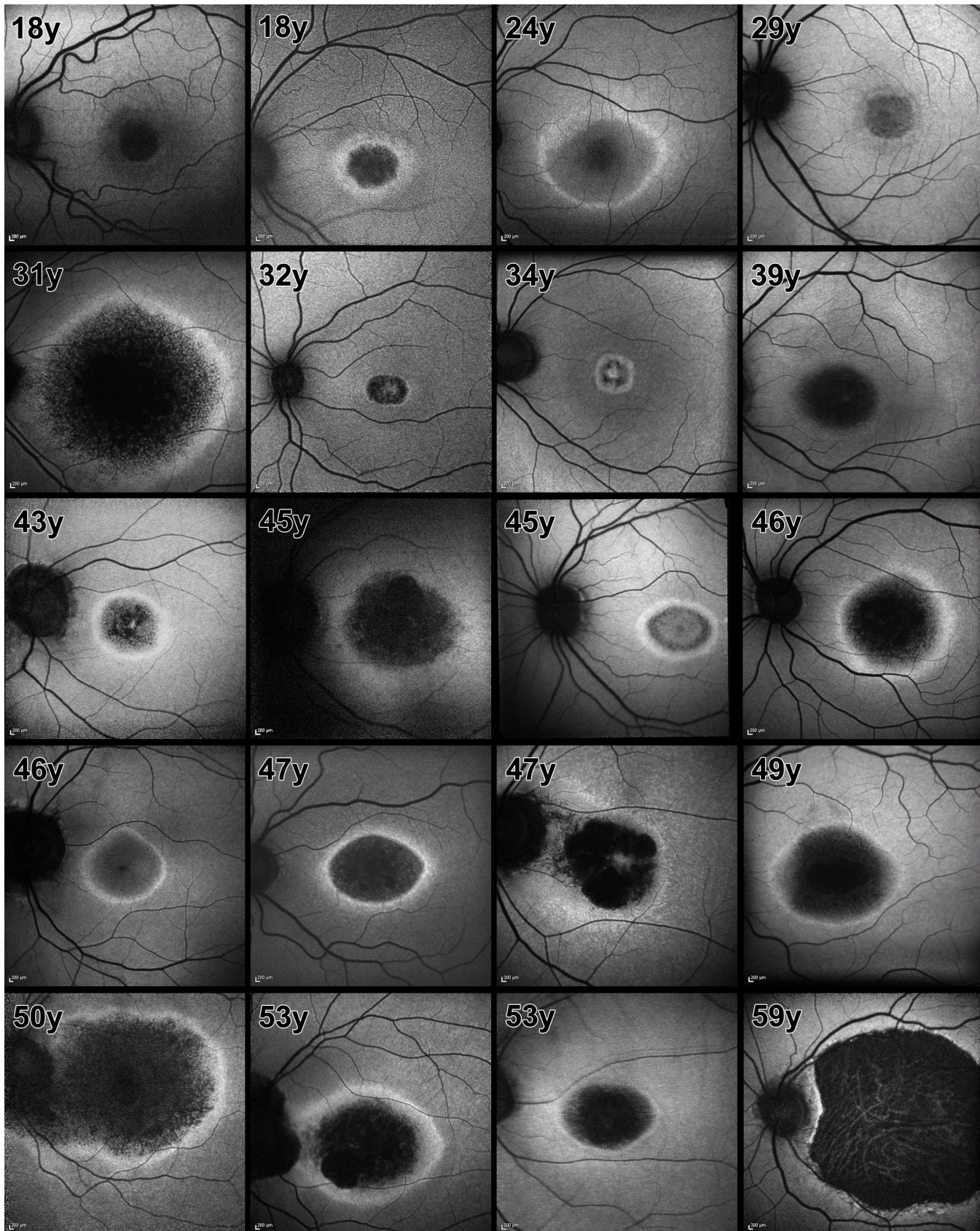


Figure S1. Fundus autofluorescence (FAF) images of 20 individuals with: [i] a retinal dystrophy with early cone photoreceptor involvement, [ii] no previous genetic testing or an unknown molecular diagnosis after previous genetic screening, [iii] absence of fundoscopic features suggestive of *ABCA4*-retinopathy (yellow-white flecks and/or peripapillary sparing). Exome sequencing was performed in all these as well as eight additional cases with a similar phenotype (including subjects CD1 and CD2). FAF imaging uses naturally occurring fluorescence to map metabolic changes at the level of the retinal pigment epithelium. The FAF pattern observed in these 20 individuals is not unlike that seen in retinopathy due to dominant mutations in *GUCY2D* [MIM *600179], *PROM1* [MIM *604365], *RIMS1* [MIM *606629] and *CRX* [MIM *602225], and recessive mutations in *KCNV2* [MIM *607604]. Retinopathy due to mutations in *PRPH2* [MIM *179605], *ABCA4* [MIM *601691] and *RPGR* [MIM *312610] should also be in the differential although the presentation would be atypical for these.

Human	MPIVMARDLEETASSEDEE-VISQED----	HPCIMWTGG-CRRIPVLVHADAILTKDN	54
Mouse	MPVVMARDLEETASSEDED-LANQED----	HPCIMWTGG-CRRIPVLVHAEAILTKDN	54
Chicken	-----MARGLEESGSSSEEEEEEDAGDGLLDHPCIRWTGGGCRRIPIFVHADAILTND	56	
Lizard	MPVGMARDLEETDSSSEEEEEVEGPE----	HPCITWTGG-FRRIPIILVHADAIITKDS	55
Pufferfish	-----LCSHRD----	NPCVAWCGL-SRSIPVLLFFPEAAVSKDG	34
Frog	----MVPRGQQDEQSEEDDD--SKKGE----	YSCILWAGG-SRKVPIVMFHAEAVLHKNL	49
	:	.*: * * * :*.:*.:* : ::	
Human	NIRVIGERYHLSYKIVRTDSRLVRSILTAHGFHEVHPSSTDYNLMWTGSHLKPFLLR	114	
Mouse	NIRVIGERYHLSYKIVRTDSRLVRSILTAHGFHEVHPSSTDYNLMWTGSHLKPFLLR	114	
Chicken	YLRLIGERYHLSYKIVRTDSRLVRSILTAHGFHEVHPNSSDYNLMWTGSHLKPFLLR	116	
Lizard	YTRLIGERYRLAFKIVRTDSRLVRSILSAHGFREVHPSNEYNLMWTGSHLKPFLLR	115	
Pufferfish	RISSTGERYHMAFKIVRTESRLVRGILANHGFRVHNSDFNLMWGSGLKPYMLRN	94	
Frog	SLRAVGERYKLSYKIVRTDSRLVRSILSAHGFQEVNANSDFNIMWTGSHVHPYIMR	109	
	****:.:*****:*****.*.: ***:***: .*.:*.:***:***:***:***:***		
Human	EAQKVNHFPRSYELTRKDRLYKNIIRMQHTHGFKAFHILPQTFLLPAEYAEFCNSYSKDR	174	
Mouse	EAQKVNHFPRSYELTRKDRLYKNIIRMQHTHGFKAFHILPQTFLLPAEYAEFCNSYSKDR	174	
Chicken	DIQKVNHFPRSYELTRKDRLYKNVSRMQLSHGFKTFHILPQTFILPAEYQEFCS	176	
Lizard	DIQKVNHFPRSYELTRKDRLYKNIIRMQTYGFKSFHVLVLPQTFILPAEYQEFCS	175	
Pufferfish	DFQKVNHFPRSYELTRKDRLYKNIQRMQQAHGFKDFHIVPQTFVLPYEQEFCS	154	
Frog	NFQKVNHFPRSYELTRKDRLYKNVSRMQLSHGFKNFHLLPQTYLLPAEYQDFCTAF	169	
	: *****:*****: ***: ***:***:***:***:***:***:***:***		
Human	GPWIVKPVASSRGRGVYLINPNQISLEENILVSRYINNPLIIDDFKFDVRLV	234	
Mouse	GPWIVKPVASSRGRGVYLINPNQISLEENILVSRYINNPLIIDDFKFDVRLV	234	
Chicken	GPWIVKPVASSRGRGVYLINPNQIVLEDNIVSRYINNPLIIDDFKFDVRLV	236	
Lizard	GPWIVKPVASSRGRGVYLINSPNQISLEENILVSRYINNPLIIDDFKFDVRLV	235	
Pufferfish	GPWIIKPVASSRGRGIYLVSNPTQISVDDNIVSRYINNPLIIDDFKFDVRLV	214	
Frog	GPWIVKPVASSRGRGVYLINSPSLISMEDNIVSRYIGNPLIIDDFKFDVRLV	229	
	****:*****:***:..*.* :.:*****.****** *****:****		
Human	PLVIYLYEEGLARFATVRYDQGAKNIRNQFMHLTNYSVNKKSGDYVSCDDPEVEDYGN	294	
Mouse	PLVIYLYEEGLARFATVRYDQGSKNIRNQFMHLTNYSVNKKSGDYVSCDDPEVEDYGN	294	
Chicken	PLVIYLYEEGLARFATVRYDQASKNIRNQFMHLTNYSVNKKSGDYVSCDDPEVEDYGN	296	
Lizard	PLLVYLYEEGLARFATVRYDQGAKNIRNQFMHLTNYSVNKKSGDYVSCDDPEVEDYGN	295	
Pufferfish	PLLIYVYEEGLARFATVRYDQTSKNIKNTFMHLTNYSVNKKSSDYVSCDDPEVEDYGN	274	
Frog	PLVIYLYEEGLTRFATAKYDRAAKNIRNQFMHLTNYSVNKKSGDYVSCDDPDVEDYGN	289	
	.:*.:***:***:.*.: ***:.* *****.******.******		
Human	SMSAMRLRYLKQEGRDTTALMAHVEDLI IKTI ISAE LAIATACTFVPHRSSCFELYG	354	
Mouse	SMSAMRLRYLKQEGKDTTALMAHVEDLI IKTI ISAE LAIATACTFVPHRSSCFELYG	354	
Chicken	SMSAMRLRYLKQEGRDTAALMASVEDLI IKTVSAE LAIATACTFVPHRSGCFELYG	356	
Lizard	SMSAMRLRYLKQEGKDTTALMASVEDLI IKTILSAE LAIASACKAFVPHRGVCFELYG	355	
Pufferfish	SMSAVLRYLKQEGKDTTLLMRQVEDLI IKAIMGAEQQIATACTFVPHKTNCFELYG	334	
Frog	SMSAMRLRYLKQDGKDTAALMSQVEDLI IKTIVSAELPIASACKSLITHRGNCFGMR	349	
	****:*****:***: ** *****:..*.* ***:***:***:***:***:***		
Human	LIDSTLKPWLLEVNLS-----PSLACDAPLDLKIKASMISDMFTVVG	409	
Mouse	LIDNTLKPWLLEVNLS-----PSLACDAPLDLKIKASMISDMFTVVG	409	
Chicken	LIDDTLKPWLLEVNLS-----PSLACDAPLDLKIKASMLSDMFTLVGFVCQDP	410	
Lizard	LIDSTLKPWLLEVNLS-----PSLACDAP-DLKIKASMISDMFTLVGFVCQDP	408	
Pufferfish	LIDANLKPWLLEVNLS-----PSLACDAPLDLKIKASMIADMFTLVGFVCQDP	388	
Frog	CLRGVLRPTMLTIFQIFEGIPSLYIDAPLDLKVKASMISDMFTLVGVECQDP	407	
	: **:* :* : :. *** ***:***:***:***:***:***:***:***		
Human	PIYPTFESSRRNPFQKQOR-----CRPLSASDAEMKNLVGSAREKGPGLG	455	
Mouse	SIYPSFESSRRNPFQKQOR-----TRPLSASDAEMKNLVASAREKVPGLG	455	
Chicken	TVYHSSESVRRNPFQKQORPASAQSQPTNTRMTRPLSASDVEMKNLMSSGREKATGR	470	
Lizard	TSFYSS-EARRNPFQKQORPVSAQSRSSTNAKLSRPLSASDAEMKNLMSSAKEKIPGR	467	
Pufferfish	SERVTLPSLKHAAQRTQ-----VLERPLSEPTAAKNGRVAGSKDKLAVKQ-	435	
Frog	---RASSSLYDKRTQKSTH-----QRPLSANDIDT-GLQVGNREK-AVRRT	448	
	: : : : ***** . . :.* . :		

Human GSVLGLSMEEIKVLRVKEENDRRGGFIRIFPTSETWEIYGSYLEHKTSMNYMLATRLFQ 515
Mouse GSVLGLSMEEIKVLRVKEENDRRGGFIRIFPTSETWEIYGSYLEHKTSMNYMLATRLFQ 515
Chicken SSVLGLSMEEIKVLRVVDENERRGGFIRIFPTPLTWDLYGSFLEYKTSMNYMLATRLFQ 530
Lizard GSMLGLSMEEIKVLRVKEDEYERRGGFIRIFPTPIITWDYGSFLEHKTTMNYMLATRLFQ 527
Pufferfish ESTLSLTAAEIKVLRRIQEEYERRGGFIRIFPTAETWELYGEYLESKTSMNYTVANRLFH 495
Frog SCLLGLSIEELKILRRVQDEYERRGGFVRIFFRHNTWQLYGSFLEYKTSLNMYMLVTHLFP 508
. *.*: **:*:***::* :*****:**** **: **.:** **::** :...:**

Human DR-----MTADGAPELKIESLNSKAKLHAALYERKLLSLEVRKRRRRSSRLR 562
Mouse DRGNPRRSLLTGRARVSTEGAPELKVESMNSKAKLHAALYERKLLSLEVRKRRRRSGRLR 575
Chicken DRDKMKGDLITG----RSRELDLGRDLTNLEAVDSSHSLFYERKLVSLERKRRRCRTKAR 586
Lizard DPCNAE-----PSRE-LG--LDVVCNAQLHAALYERKLLSLEVRKRRRRRHGKLR 574
Pufferfish GRLGMGNKSLHK--FMERGNVSGNVQLQVESFHDCHVIQYERKLLTLETHKRRRRHRLTSR 553
Frog NR-----AAGNDHCEKNWDPRMHAAFYERKLVSLHLR--RARHRLTR 549
. * *****:*. : * * *

Human AMRPKYPVITQPAEMNVKTETESEEEEEVALDNEDEEQEASQEEASAGFLRENQAKYTPSL 622
Mouse AMRPKYPVIAQPAEMNIKTETESEEEEEVGLDNDDEEQEASQEEASAGSLGENQAKYTPSL 635
Chicken AAQTRSSGTSQPTKLSLT-DTEGEEEEEAAD--EDEE----QDGTGLSLSNSQLKSKPKL 639
Lizard PRRSRLSGALQSTDFALKSEMECEEEEEETTE--EDEEPEIPQNETADCLKNMKVKSKPQL 632
Pufferfish SAAGKRK-SGSSQNLFOKCLSESKTSLTSLG-----SQEAEECAQEEREVEKAVL 602
Frog KTGLSHAPQCSDEHQSSKEQEEEEEELDEN-----HEL 584
. . . * : *

Human TALVENTPKENSM-KVREWNNKGGHCCKLETQELEPK-----FNLMQIQLDNGNLSKMQA 676
Mouse TVIVENSPRDNAM-KVAEWTNKGEPCCKIEAQEPESK-----FNLMQIQLDNGNLSKVQA 689
Chicken SELVKTASKERLT-EKLDKKTTRNGGEPFLEKSDSKSQ-----FNLLQILKKGDNLSKVQA 693
Lizard SEQVEPSHQGKLTKNQLEQKPKSEELPCESKLDTVPKSVLFPNLLQVLHENKNLSKVQA 692
Pufferfish EPLSKRALEAELS-KQMAASLKRCQAEALSSSEAAAHNGGHKVSLLDVLQQGWDLSKVQA 661
Frog EVVQEKVSSPDSS-KI I I P P P R-----ISLMDILRKGADLSKVQA 623
: : : ..*::*... :***:**

Human RIAFSAYLQHVQIRLMKDSGGQTFASWAAKEDEQMELVVRFLKRASNNLQHSRLMVLPS 736
Mouse RLAFSAYLQHVQIRLTKDSGGQTLSPSWAAKEDEQMELVVRFLKRASNNLQHSRLMVLPS 749
Chicken RRAFSAYLQHVQLRLMKDVGDQFQNAAWAAKEDEQMELVVHFLKRAASNLQQLSLRMLLPS 753
Lizard RKAFSAYLHRVQLRLMKEAGDQVHNPAAWAAKEDEQMELVVRFLKRAASNLQQLSLRMLLPS 752
Pufferfish RKAFSSYLQRVQQRLLAESR-TDAIPAWPDKDNDQMDLVIRFLKRAASNLQQDIQVAFPS 720
Frog RNAFSCYLQRVQNRQLQTERNPERVQP----KEEQIELVMRFLQRGANLKRSLPLNLP 679
* **.***::** ** : *::*::**::**:*.: **::: : : *.

Human RRLALLERRRILAHQLGDFIIVYNKETEQMAEKKS KKKVE-----EEEEEDGVNMFNFQ 789
Mouse RRLALLERRRILAHQLGDFIGVYNKETEQMAEKKS KKKLE-----EEEEEDGVNAESFQ 802
Chicken RHLGLNDRRRILAHQLGDFIICYNKETEQMIQKRS KKKQE-----EEEE-GVNPEGFQ 805
Lizard RRLALFDRRRILAHQLGDFIICYNRETQMAQKKLKQKQE-----EEEEGVDPPEGFH 805
Pufferfish RQLPLQDRRRILSHQLGDFIHCYDQVFALPAKQKQKQETENIVKKQVSDGGLCVNAGVVF 780
Frog QSVPYLERRHLLAKLLGDFVALYNQETQQMQNSEETQSNE-----ECGVNPDDFE 729
: : :***:**: **:*: *:: :... :.. * * : *

Human EFIRQASEAELEEVLTFFYTQKNKSASVFLGTHSKISKNNNN--YSDSGAKGDHPETIMEE 847
Mouse EFIRQASEAELEEVLTFFYTQKNKSASVFLGTHSKSKNSSS--YSDSGAKGDHPET-IQE 859
Chicken NFITRASERDLEEVLTFFYTQKNKSASVFLGTNSGTTKPRNTSNQSENQPQGDHPVEMKNT 865
Lizard EFVIKASESDLEEVLTFFYTHKNKSASVFLGTNPGTSKHSNS--QLENRGK----EVVKEN 859
Pufferfish EYISAASEAELEEVLTFFYTQKNKSSTVFLGAQGKSVRPKSCR-FSDAEAAVRQPSDRQEA 839
Frog AFVADASENELEEVLTFFYTQKNKSASVFLGTPNADRRETG---KPPGGPQNRLTCERSV 786
:: *** :*****:****:****: : .

Human VKIKPP-----KQQQTTEIHSKLSRFTTSAEKEAKLVYSNSS----SGPTA 890
Mouse VKIKQP-----KQQQATEIHADKLSRFTTSSGKEAKLVYTNCS SFC-SPA AV 905
Chicken KGDQAKSSVADLPVEGRVVRVCKARVKSTLPEKS--TFSLNAEVKLPRCCPPSAASSASGA 923
Lizard VAEELK-----DDQLKETRLSSNIHFQKQAGVTSSPEDENAPQLSFLSATSSSVPDA 911
Pufferfish VTTQPEIQAP-----QHRSSFPATIDSADIQQHRSASSATLPLYCLPP----PPPPP 888
Frog VMSSAP-----SQPSVSKPGYSQGCNVSDGPIIITNSAVN----- 821
. .

Human TLQKIPNTHLSS-VTTSDLSPGPGCHHSSLSQIP--SAIPSMHPQP-TILLNTVSASASPC 946
Mouse LLQRLPSSHLSSVITTSALSAGPGHHASLSQIP--PAVPSLPHQP-ALLSPVPDNAPPS 962
Chicken TFRRSTSSQLPSQPTASGNPQVPGHCSLPTPPSGLRLIHSSSSLPSSQSQSTATDCSSVF 983
Lizard ILPQSI SFPQLPSHTAASDNAQLPDHLGFLSSGSR LIPSSS-----FQNAAMDSWSST 965
Pufferfish QLPSYAQSLAKSQFCYSERPPDPAYASSAVVS-----QPLGVSWTP 930
Frog GLPESCSQRNSSTHVLSNDASLCSTIVGSNGIH-----VSPAKLT 861
: . * * .

Human LHPGAQNI P SPTGLPRCRSGSHTIGPFSSFQSAAH IYSQKLSRPSSAKAG--SCYLNKHH 1004
Mouse IHSGTQNV S-PAGLPRCRSGSYTIGPFSSFQSAAH IYSQKLSRPSSAKAAG-SCHPHKHH 1020
Chicken TNPVSSEASSLAGLHRC-SGSYTIGPLSSFQRAAQ IYSQRLSRSPSAKAGLRHRSPSGQR 1042
Lizard TKNVPQLSSNAGLHRCQSGSFTASPFSSFQSA M Q IYSQRLTRPSSAKAGSRSHSPSRQR 1025
Pufferfish VSTGKNPPNQVLRRIQSFTSSMSCGGASSLPR TMQLYSQKLSRPTSTIHS-FSCSPHESP 989
Frog VTPGSWAKS-----GSRPHSSSLGTFSSFQSAAQ IYSQKLRPSSTRSECNHVSVHCNY 915
. . * : . ** : : : ** : * * . . *

Human SGI AKTQKEGEDASLYSKRYNQSMVTAELQRLAEKQAARQYSPSSHINLLTQQVTNLNLA 1064
Mouse SGI AKTQKEGEDVSLN-RRYNQSLVTAELQRLAEKQAARQYSPASHISLLTQQVTNLNLA 1079
Chicken VSSIMMNKGTEDAPSLGKRYSPSMVAEELQQLAEKQAACQYSPSHISLLTQQLTSLNLA 1102
Lizard SAFARVTKDGEEC----KRF SHGVIAEELHRLAEKQATRQYCPSSHINLLTQQLTNLNLM 1081
Pufferfish RGATPTFKELHPRPEP-TQSNQQAFLSALQK LADKQAARRYASSSHINLLTHHLTQMNLA 1048
Frog PSLCANCTALNIP-----EARNAFSCYLQRVQNR LQTERNPERVQPEEEQILTSMN I K 969
. . . . * : : : : : : : : * : * :

Human TGIINRSSASAPPTLRPIISP SG--PTWSTQSDPQAPENHSSSPGSRSLQTGGFAWEGEV 1122
Mouse SSVINRSSASTPPTLRPVISP SG--PTWSIQPDLHASETHSSPPGSRSLQTGGFAWEGEV 1137
Chicken SGAVSKGNAAVPPSYRSALNRKG--PLCTVQSDTLTDDRRCISSAVRAPESDRFAWEGEM 1160
Lizard NGAVSRVNTTS--SYRPSLNPGG--SFWAFQNTV I I SNHDKPMQEMALETDRFAWEGDA 1137
Pufferfish NRMLSRDGFALNPPVQRTAAPAAQRPEWAGQLMLY GDRVHVCLPTNRPQKDRDDAFKQGT 1108
Frog DGAFGSGSFRH----CSAKSFCG-----RAVHAGTETVESITRDIQRRRS AWESDQ 1016
. . . . : * : : :

Human ENNVYSQATGVVPQHKYHP-TAGSYQLQFALQQLQLEQQKLSRQLLDQSRARHQAI FGSQT 1181
Mouse ENNAYSKT TGVPVQHKYHP-TAGSYQLHFALQQLQLEQQKLSRQLLDQSRARHQAI FGSQT 1196
Chicken ENNVYGK VTRSPLAHP-----NYQLNLAVQQLQQLQKLSRQLLEQS QARHQALFASYS 1213
Lizard ENSLHSKLIGSQPLHPKASSSTGSYQLHFALQQLQQLQKLSRQLLDQSRARHQALFANFP 1197
Pufferfish QS-PYSLLTPMTPQQIKPP-APGSDQLQSAIKK LQQQLRSRQFLDQSHRQQALF---- 1162
Frog ESGTFSFSSDVPLQHQ-----PDQM QYSAKGGQHPDSAI I SLPNQTCTLLPTPPVSHK 1069
: . . : * : : : : . : : * : :

Human LPNSNLWTMNGAGCRISSATASGQKPTTLPQKV VPPPSSCASLVPKPPPNEHQ-VLRR A 1240
Mouse LPNSSLWTMNGPGCRISSAT TGGQKPN TLPQKV VAPPNS-STLVSKPASNHKQ-VLRKP 1254
Chicken QSSTSHVPMSPGSGAHTTSSATSSIQKAASLHKVMPSQCTPSQLVPKPPANHRQAVVRKT 1273
Lizard TSSISSITLSSGSGARRTSSAISSSQKASTLHKVMSSQSASSHLIPKPPASHRQTVIRKV 1257
Pufferfish -----
Frog QSAARTLSATR-----LVRVAPVEQHG----TP 1093

Human TSQKAS-KGSSAEGQLNGLQSSLN-PAAFVPIT SSTDPAHTKI----- 1281
Mouse ASQRAS-KGSSAEGQLNGLQSSLN-PAAFMPITNSTGSLEAPQVIFARSKPLPTQSGALA 1312
Chicken AAQRIS-KVSSVERQLNGFQNSLRGAASCELGSNSTASACREGLALNTRRNPE SCFQVWG 1332
Lizard ASQRISNRAISMEGQMNGFQNSLDSATSCEPLTNSTGEAKIKK----- 1300
Pufferfish -----
Frog TSTIVSDFGTPSQGSMEATQI IFARARPSAPKIDIKGQRK----- 1133

Human -----
Mouse TVIGQRKSKSVKSGTI 1328
Chicken KGKKQQ----- 1338
Lizard -----
Pufferfish -----
Frog -----

Figure S2. Amino acid sequence alignment of TTLL5 orthologs from all branches of the vertebrate animal kingdom. Conservation of the p.Glu543 residue (highlighted in purple) that is mutated in subjects CD3, CD4 and CD5 is observed despite some sequence divergence elsewhere. The alignment was performed with ClustalW2 (EMBL-EBI, Hinxton, UK) using appropriate Ensembl transcripts

Table S1. Clinical and genetic findings from the 28 cases that were selected for exome sequencing

Family ID	Age tested, sex	Likely disease-causing gene (exome sequencing result)	Electrophysiological diagnosis	Genetic testing prior exome sequencing	Family history
gc18728 (CD1)	38,M	<i>TLL5</i> p.[(Glu529fs)];[(Glu529fs)].	CD	<i>ABCA4</i> microarray.	No other affected; consanguinity.
gc19552 (CD2)	45,M	<i>TLL5</i> p.[(Leu134fs);(Trp1118*)].	CD	<i>ABCA4</i> microarray.	No other affected.
gc17090	18,F	Not known.	Not available (clinically MD).	None.	Sister affected.
gc15017	18,M	<i>ABCA4</i> p.[(Gly1961Glu);(Asp295fs)].	MD	None.	Brother affected.
gc19458	24,F	<i>CRX</i> p.[(Arg43Cys)];[=].	MD	None.	No other affected.
gc17004	29,F	Not known.	MD	<i>ABCA4</i> microarray.	No other affected.
gc17898	31,F	<i>PROM1</i> p.[(Arg373Cys)];[=].	CRD	<i>ABCA4</i> microarray; <i>PRPH2</i> all exons.	Children & paternal uncle affected; consanguinity.
gc15235	32,F	Not known.	MD	<i>ABCA4</i> microarray; <i>PRPH2</i> all exons.	Two affected siblings.
gc19146	34,M	Not known.	CRD	<i>ABCA4</i> microarray.	No other affected.
gc17967	39,M	Not known.	CRD	None.	No other affected; consanguinity.
gc17988	43,M	<i>RPGR</i> p.[(Glu1060fs)];[0].	CRD	<i>GUCA1A</i> all exons.	No other affected.
gc4728	45,M	Not known.	MD	<i>ABCA4</i> microarray.	No other affected.
gc16362	45,M	<i>ABCA4</i> p.[(Arg1843Gly)];[?].	MD	None.	No other affected.
gc19964	46,M	Not known.	MD	<i>PRPH2</i> all exons.	No other affected.
gc19080	46,M	<i>RPGR</i> p.[(Lys1106fs)];[0].	CD	None.	No other affected.
gc16258	47,M	Not known.	MD	<i>ABCA4</i> microarray.	No other affected.
gc5342	47,F	Not known.	MD	None.	No other affected.
gc17836	49,M	Not known.	CD	None.	Sister affected; consanguinity.
gc19457	50,M	Not known.	RCD	None.	No other affected.
gc18729	53,M	<i>ABCA4</i> p.[(Gly1961Glu)];[?].	CD	<i>ADAM9</i> all exons; <i>RPGR</i> exon ORF15.	No other affected.
gc16711	53,M	<i>CRX</i> p.[(Arg91Lys)];[=].	CRD	<i>RS1</i> all exons.	No other affected.
gc16174	59,M	Not known.	MD	<i>ABCA4</i> microarray; <i>PRPH2</i> all exons.	No other affected.
gc18250	18,M	<i>CDH3</i> p.[(Asp523fs)];[(Asp523fs)].	MD	None.	No other affected; consanguinity.
gc17784	19,M	<i>ABCA4</i> p.[(Asp1734Thr)];[?].	RCD	<i>RS1</i> all exons; genotyping array.	No other affected; consanguinity.
gc19018	50,F	Not known.	CRD	<i>RIMS1</i> exons 14-15; <i>GUCY2D</i> exon 13; <i>PRPH2</i> all exons.	Affected sister, father & paternal grandfather.
gc18280	56,F	<i>CRX</i> p.[(Tyr258*)];[=].	MD	<i>PRPH2</i> all exons.	Affected sister & son.
gc16966	63,M	Not known.	MD	<i>ABCA4</i> microarray; <i>PRPH2</i> all exons.	No other affected.
gc4055	74,M	Not known.	CD	All exons of <i>PRPH2</i> , <i>PROM1</i> & <i>RS1</i> .	Father affected.

CD, cone dystrophy; CRD, cone-rod dystrophy; RCD, rod-cone dystrophy; MD, macular dystrophy; *ABCA4* microarray, *ABCA4* APEX microarray (ABCR400 or ABCR600 chip, Asper Ophthalmics, Tartu, Estonia). All genes except *ABCA4* were screened using Sanger sequencing. Individuals with family IDs gc17898, gc19457, gc16711, gc19018 and gc18280 were excluded from the case-control analysis based on ancestry.

Table S2. Top five most significant autosomal genes: the total count of non-synonymous and splice altering rare variants was compared between probands with retinal dystrophy and internal controls

	Chromosome	Number of non-synonymous and splice altering variants cases ^a (n = 23)	Number of non-synonymous and splice altering variants in UCL-exomes controls ^b (n = 1,465)	Sequence Kernel Association Test (SKAT) P-value	Binomial P-value
<i>TLL5</i>	14	9	50	0.00069	4.46e-7
<i>KRTAP10-8</i>	21	5	11	0.0036	1.54e-6
<i>TPR</i>	1	9	67	2.12e-4	2.73e-5
<i>RTTN</i>	18	7	37	1.21e-4	3.08e-5
<i>MUC16</i>	19	21	351	6.25e-4	4.09e-5

^aCase group: 28 probands with [i] a retinal dystrophy with early cone photoreceptor involvement, [ii] an unknown molecular diagnosis after previous genetic screening or no previous genetic testing, [iii] absence of fundoscopic and fundus autofluorescence imaging features suggestive of *ABCA4*-retinopathy. Five of these 28 cases were excluded based on ancestry (Figure 2A).

^bUCL-exomes control group: 1,750 individuals analyzed with using the same sequence variant calling strategy as the 28 retinal dystrophy cases. After [i] inferring ancestry based on the exome sequencing data and using a principal component analysis to exclude samples that did not cluster with the bulk of the UCL-exomes samples, which are predominantly of European origin, [ii] removing all samples with a history of retinal disease and [iii] excluding related control samples, we were left with 1,465 unrelated controls.

Genes are ranked based on the binomial P-value test which tests for equal proportion of non-synonymous and splice altering rare variants between cases and controls, against the alternative of an excess of the same class of variants in cases. To define “rare” variants we utilized two cohorts: a subset of 25% of UCL-exomes controls (366 unrelated control samples, randomly sampled and not included directly in the case-control analysis; minor allele frequency of <0.3%) as well as the NHLBI Exome Sequencing Project dataset (minor allele frequency of <0.1% was used).

Table S3. Prioritization of variants identified by exome sequencing in three probands with *TTLL5*-retinopathy

	subject CD1	subject CD2	subject CD3
All exonic variants	21,111	21,742	22,783
Total non-synonymous and splice altering rare ^a variants	450	485	716
Homozygous non-synonymous and splice altering rare ^a variants	11	9	47
Homozygous presumed loss-of-function rare ^a variants	3 ^b	0	3 ^c
Genes with two heterozygous presumed loss-of-function rare ^a variants	0	1 ^d	0

^aRare variants: variants with: [i] minor allele frequency of <0.3% in 366 randomly sampled internal UCL-exomes controls and [ii] minor allele frequency of <0.1% in the ~6500 samples in the NHLBI Exome Sequencing Project dataset.

^bc.[202G>T];[(202G>T)], p.[(Glu68*);[(Glu68*)] in ENST00000449873-*TBX15* [MIM *604127]; c.[1628_1631del];[(1628_1631del)], p.[(543_544del)];[(543_544del)] in ENST00000464606-*ZC3HAV1* [MIM *607312]; c.[1586_1589delAGAG];[(1586_1589delAGAG)], p.[(Glu529fs)];[(Glu529fs)] in ENST00000298832-*TTLL5* [MIM *612268].

^cc.[321_322insAC];[(321_322insAC)], p.[(Thr107fs)];[(Thr107fs)] in ENST00000408995-*FHL2* [MIM 602633]; c.[91G>T];[(91G>T)], p.[(Glu31*);[(Glu31*)] in ENST00000377294-*ZKSCAN4* [MIM *611643] and c.[1627G>T];[(1627G>T)], p.[(Glu543*);[(Glu543*)] in ENST00000298832-*TTLL5* [MIM *612268].

^dc.[401delT(;);3354G>A], p.[(Leu134fs)(;)(Trp1118*)] in ENST00000298832-*TTLL5* [MIM *612268].

Exome sequencing was performed using a solution-phase exome capture (SureSelectXT Human All Exon V5, Agilent, CA, USA) and the Illumina HiSeq2000 sequencer (Illumina, CA, USA). Reads were aligned to the hg19 human reference sequence using Novoalign version 2.07.19 (Novocraft, Selangor, Malaysia). Genome Analysis Tool Kit (GATK) version 2.7.4 and ANNOVAR (2013Nov17 version; Open Bioinformatics, MA, USA) were used for variant calling and annotation of single nucleotide polymorphisms and small insertions/deletions. Filtering of variants and case-control analysis were carried out using R scripts.

Each of these three individuals was born to consanguineous parents. Prior exome sequencing of DNA from subject CD3, homozygosity mapping in samples from subject CD3 and his affected brother (subject CD4) was performed (Human Mapping 50K Array Xba 240, Affymetrix, CA, USA). This had yielded four regions of shared homozygosity that were over 10 cM; *TTLL5* was in the largest shared chromosomal segment.

Table S4. Primer sequences and conditions used for *TLL5* mutation screening

Primer name	Primer sequence	Primer name	Primer sequence
TLL5_ex2F	tgtggcatattgaggacat	TLL5_ex18F	tgtcttttccttggcactt
TLL5_ex2R	ggcccagaaagagagcctta	TLL5_ex18R	cccctccacttttccaatc
TLL5_ex3F	gggagatgtgattcccaca	TLL5_ex19F	ggtgtgggtggcactttat
TLL5_ex3R	gggctggggatctgctta	TLL5_ex19R	aagagcaaaggccaaaatgt
TLL5_ex4F	ggtgtaattttccccatc	TLL5_ex20F	gagagtgcacatgggtgct
TLL5_ex4R	ctggtaaagccactccaaaa	TLL5_ex20R	aaatgccaaccaatgagac
TLL5_ex5F	aaccctcccattcctgaac	TLL5_ex21F	cataatagaagcatcctcaaaggcc
TLL5_ex5R	ggtgcagtgcagcaagaatca	TLL5_ex21R	caaagattgcttcacattgaag
TLL5_ex6F	cactacagggggacttgagg	TLL5_ex22F	ccttttgtctgggtctg
TLL5_ex6R	tgccagtgtgcccttacata	TLL5_ex22R	ccactggccttcagaagta
TLL5_ex7F	cctcctccctcgtctatt	TLL5_ex23F	cattctgcaactttactggg
TLL5_ex7R	ttcctgccagtaaggcaaac	TLL5_ex23R	catgaaaatagcaacataattggc
TLL5_ex8F	tggttacctggaggaact	TLL5_ex24F	agaaaatcactgcgggatg
TLL5_ex8R	aaggaacctgctcctttct	TLL5_ex24R	tactgtccccattctccac
TLL5_ex9F	cctccgaagtcaagggtgtg	TLL5_ex25F	ggctgtgggtgcttcatct
TLL5_ex9R	agcacagcagttgaggaggt	TLL5_ex25R	cccctctttcacccttct
TLL5_ex10F	gtccatgggtttggagtg	TLL5_ex26F	gacatgcctgctctgttca
TLL5_ex10R	aatggagaagcagcaggaga	TLL5_ex26R	gctactggatgcaatgcaa
TLL5_ex11F	agaaagaattgccgcctc	TLL5_ex27F	ggattctaggttatggaacc
TLL5_ex11R	cagctgtcaactgcaggaa	TLL5_ex27R	cttcaaatgcctgtaacag
TLL5_ex12F	tccctggcacctacattct	TLL5_ex28F	tcctcctgagtgctttgt
TLL5_ex12R	ctcaggggacttctgaccaa	TLL5_ex28R	ctagttaggtgcccagagga
TLL5_ex13F	gcccataagcacagcagaat	TLL5_ex29F	ggtttagtgggggagtgaga
TLL5_ex13R	atggccctagatccaggtt	TLL5_ex29R	actccccatgagctgtccaa
TLL5_ex14F	ttttgccaggattttcc	TLL5_ex30F	gctgcactggcaacattaga
TLL5_ex14R	ggagccaagtgtcgtagaaa	TLL5_ex30R	aattttagcccacgctgag
TLL5_ex15F	gaggggtgtgtgggagagt	TLL5_ex31F	aggccccatgctttctgata
TLL5_ex15R	ctgtgcctgtttctgagca	TLL5_ex31R	atgccattgccaatgttt
TLL5_ex16F	gaattgagctataaatcttag	TLL5_ex32F	gagcttccacttagaggtgaac
TLL5_ex16R	gatagttatgcccaagaatatg	TLL5_ex32R	ctttatatcatctctgtgcagcag
TLL5_ex17F	gacaaactcatgtcttacattg		
TLL5_ex17R	cacaaagtttaggacagtcccc		

All primers work at 60°C. Ensembl transcript ID ENST00000298832 was used.

Table S5. Presumed loss-of-function variants in *TLL5* identified in the Broad 26K dataset (26,000 exomes) and the internal UCL-exomes control cohort (1,465 unrelated exomes)

<i>TLL5</i> presumed loss-of-function variants			Broad 26K dataset	UCL-exomes control	Cases ^a
Genomic build position (hg19)	Nucleotide	Protein	allele count (52,000 alleles)	cohort allele count (2,930 alleles)	
14:76156564	c.401delT	p.Leu134Argfs*45	0	0	heterozygous state in CD2
14:76165584	c.556delA	p.Arg186Glyfs*7	1	0	-
14:76173403	c.629dupA	p.Tyr210*	1	0	-
14:76184249	c.789_793delGTTCA	p.Gln263Hisfs*19	1	0	-
14:76200373	c.1166C>G	p.Ser389*	2	0	-
14:76201609	c.1258C>T	p.Ala420*	1	0	-
14:76211845	c.1408C>T	p.Arg470*	2	0	-
14:76211872	c.1435C>T	p.Arg479*	1	0	-
14:76219296	c.1548delC	p.Asp516fs*3	0	1	-
14:76230991	c.1586_1589delAGAG	p.Glu529Valfs*2	0	0	homozygous state in CD1
14:76231034	c.1627G>T	p.Glu543*	0	1	homozygous state in CD3 & CD4
14:76231034	c.1627G>A	p.Glu543Leu	14	0	homozygous state in CD5
14:76231061	c.1654C>T	p.Arg552*	1	1	-
14:76232616	c.1920G>A	p.Trp640*	3	0	-
14:76238090	c.2029C>T	p.Arg677*	4	0	-
14:76238192	c.2132_2133insGATA	p.Met712Ilefs*15	1	2	-
14:76241952	c.2264_2265dupTT	p.Ile756Leufs*29	1	0	-
14:76243171	c.2365C>T	p.Gln789*	1	0	-
14:76245995	c.2466dupT	p.Lys823*	3	0	-
14:76249626	c.2739C>A	p.Cys913*	1	0	-
14:76249741	c.2854C>T	p.Gln952*	1	0	-
14:76249777	c.2890C>T	p.Arg964*	1	0	-
14:76330011	c.3329delG	p.Ser1110Thrfs*13	1	0	-
14:76330037	c.3354G>A	p.Trp1118*	0	0	heterozygous state in CD2
14:76330140	c.3457C>T	p.Gln1153*	1	0	-
14:76330187	c.3504_3517delGAGTCGAGCCCGGC	p.Ser1169Profs*11	2	0	-
14:76368485	c.3744dupG	p.Ser1249Valfs*15	1	0	-
14:76368504	c.3760C>T	p.Gln1254*	1	0	-
14:76420778	c.3835delA	p.Thr1279Leufs*20	1	0	-
Combined frequency of presumed loss-of-function variants			0.0903% (47 total)	0.17% (5 total)	-

^aCase group: 28 probands with [i] a retinal dystrophy with early cone photoreceptor involvement, [ii] an unknown molecular diagnosis after previous genetic screening or no previous genetic testing, [iii] no features of *ABCA4*-retinopathy on fundus autofluorescence imaging.

The cDNA is numbered according to Ensembl transcript ID ENST00000298832. See text for more details on the Broad 26K and UCL-exomes control cohorts.

All exome sequences (UCL-exomes and Broad 26K) were generated using the Illumina technology (HiSeq or GAIIX instruments). UCL-exomes FASTQ files were aligned against the human reference genome hg19 using Novoalign (version 2.07.19). Variants were called using the GATK version 2.7.4. All UCL-exomes samples (cases and controls) and all Broad 26K samples (~28,000 samples overall) were called jointly using the GATK UnifiedGenotyper module, following BAM file reduction as implemented by GATK using default options (same 2.7.4 release). We used the Illumina TruSeq target region for variant calling, with +/- 100 base-pairs on the side of each target region. We followed the GATK best practices and implemented variant recalibration, with separate models for SNPs and insertions-deletions. We excluded read depth from our recalibration model owing to the large read depth variability generated by the heterogeneous capture kits used in the multiple studies that form UCL-exomes. Variants with PASS filter and the highest level recalibration tranche (VQSRTancheSNP99.00to99.90) were retained. We used a variant Phred quality threshold of 30 and a genotype (i.e. sample based) Phred quality threshold of 20, with the exception of heterozygous call for which we found the error model overly permissive and for which we used a more stringent genotype Phred quality threshold of 40.
