

**TABLE S1** Observed diversity and estimated sample coverage in 16S rRNA sequencing analysis of Canestrato Pugliese cheese during manufacture and ripening

<b>Days of ripening</b>	<b>No. of reads</b>	<b>No. of OTUs</b>	<b>Chao1 richness</b>	<b>Shannon diversity index</b>	<b>ESC (%)</b>
Milk	5727	14	17.33	1.43	99%
Curd <sup>a</sup>	3555	44	46.57	3.05	99%
1 <sup>b</sup>	5001	34	38.66	1.15	99%
3	2206	23	26.33	0.75	99%
7	6334	36	49.75	0.92	99%
15	9357	39	43.20	1.75	99%
30	8945	40	42.50	1.27	99%
45	3687	41	44.00	2.06	99%
60	7843	45	52.33	2.08	99%
75	7965	43	51.25	1.65	99%
90	6304	43	47.00	1.99	99%

Abbreviations: OTU, operational taxonomic unit; ESC, estimated sample coverage. Chao1 richness, Shannon diversity, and ESC were calculated with Qiime at the 3% distance level.

<sup>a</sup>Curd after moulding.

<sup>b</sup>Curd after dry salting.

**TABLE S2** Community Level Catabolic Profiles (CLCPs) of Canestrato Pugliese cheese during manufacture and ripening

Days of ripening	<i>H'</i>	<i>S</i>	<i>E</i>
Milk	2.68 ± 0.2 D	12.33 ± 1.5D	2.64 ± 0.2BC
Curd <sup>a</sup>	3.00 ± 0.1C	16.67 ± 2.5 B	2.46 ± 0.1A
1 <sup>b</sup>	2.71 ± 0.1 E	12.68 ± 1.5 D	2.47 ± 0.2A
3	3.25 ± 0.1 A	20.33 ± 0.2 A	2.49 ± 0.1A
7	3.09 ± 0.3 BC	15.67 ± 1.1 B	2.59 ± 0.2B
15	3.15 ± 0.2 B	16.00 ± 1.5 B	2.61 ± 0.3B
30	3.03 ± 0.2 C	16.67 ± 0.6B	2.48 ± 0.1A
45	3.00 ± 0.3 C	15.23 ± 2.1B	2.53 ± 0.3AB
60	3.03 ± 0.1C	13.67 ± 1.5C	2.68 ± 0.2 BC
75	3.09 ± 0.2 BC	13.67 ± 1.1C	2.72 ± 0.1C
90	2.84 ± 0.1D	13.33 ± 0.6C	2.53 ± 0.2AB

Each value was expressed as the mean ± standard deviations for two batches of each type of cheese, analysed in triplicate.

Means within the column with different letters (A-D) are significantly different ( $P < 0.05$ ).

*H'*, Shannon's diversity; *S*, substrate richness; *E*, substrate evenness.

<sup>a</sup>Curd after moulding.

<sup>b</sup>Curd after dry salting

**TABLE S3** Correlations<sup>a</sup> between the abundance of operational taxonomic units (OTUs) and total free amino acid (FFA), concentration of Asp, Glu, Leu, Phe, and Val, area of hydrophilic and hydrophobic peaks<sup>b</sup>, and carbon substrates<sup>c</sup> (polimers, amines, carboxylic acids, carbohydrates, and amino acids)

Descriptors	Significance	Abundance of OTUs							
		<i>L. plantarum</i> group	<i>L. casei</i> group	<i>Lactobacillus</i> sp.	<i>L. sakei</i> group	<i>Lactobacillaceae</i> family	<i>L. brevis</i>	<i>Lc. lactis</i>	<i>St. thermophilus</i>
FAA	P	3.92E-08	7.90E-05	0.00013	0.00028	0.0012	0.0019		
	FDR	8.23E-07	0.0008	0.0009	0.0015	0.0049	0.0067		
	r	0.89	0.74	0.72	0.70	0.64	0.62		
Asp	P	1.49E-07	6.12E-05	0.0003	0.0004				
	FDR	3.14E-06	0.0006	0.002	0.0025				
	r	0.87	0.75	0.69	0.69				
Glu	P	4.05E-07	0.0001	0.00064	0.0004				
	FDR	8.50E-06	0.0012	0.0034	0.0029				
	r	0.85	0.73	0.67	0.68				
Leu	P	5.40E-08	2.61E-05	0.00013	0.0002		0.0027		
	FDR	1.13E-06	0.0003	0.0009	0.0009		0.011		
	r	0.88	0.77	0.72	0.72		0.61		
Phe	P	3.36E-08	3.52E-05	0.0002	0.0002		0.0024		
	FDR	7.07E-07	0.0004	0.0011	0.0013		0.01		
	r	0.89	0.76	0.71	0.71		0.61		
Val	P	9.21E-08	5.29E-05	0.0002	0.0002				
	FDR	1.93E-06	0.0005	0.0013	0.0011				
	r	0.88	0.75	0.71	0.71				

**TABLE S3** (continued)

Descriptors	Significance	Abundance of OTUs							
		<i>L. plantarum</i> group	<i>L. casei</i> group	<i>Lactobacillus</i> sp.	<i>L. sakei</i> group	<i>Lactobacillaceae</i> family	<i>L. brevis</i>	<i>Lc. lactis</i>	<i>St. thermophilus</i>
Area hydrophilic peaks	P	3.56E-07	0.0001	0.0002	8.80E-05	0.0009	0.0015		
	FDR	7.47E-06	0.0009	0.0010	0.0009	0.003	0.005		
	r	0.86	0.72	0.71	0.74	0.65	0.63		
Area hydrophobic peaks	P								0.002
	FDR								0.04
	r								0.62
Polymers	P							5.69E-09	
	FDR							1.19E-07	
	r							0.91	
Amines	P							0.0061	
	FDR							0.043	
	r							0.60	
Carbohydrates	P							0.0003	
	FDR							0.007	
	r							0.69	
Amino acids	P							3.77E-07	
	FDR							7.92E-06	
	r							0.85	

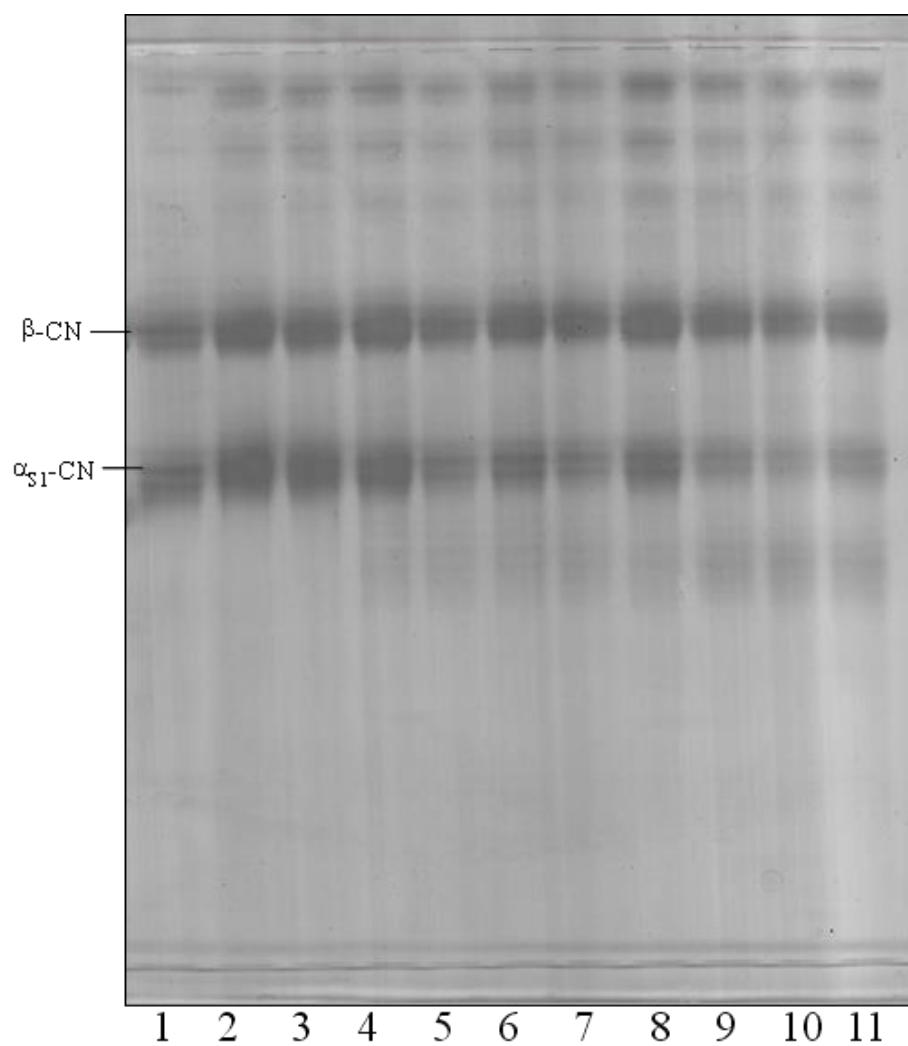
<sup>a</sup>Only the positive correlations with a P<0.05, FDR<0.05 and r>0.06 are reported.

<sup>b</sup>Area of hydrophilic and hydrophobic peaks of the pH 4.6-soluble nitrogen fractions analyzed by reverse phase high pressure liquid chromatography (RP-HPLC) were recognized and matched visually with the Unicorn program (Amersham Biosciences).

<sup>c</sup>Utilization of carbon substrates was carried out by using Biolog Eco-Microplates containing 31 carbon substrates grouped for chemical class (polimers, amines, carbohydrates, and amino acids).

**FIGURE S1** Urea-polyacrylamide gel electrophoresis (PAGE) of the pH 4.6-insoluble (A) and -soluble (B) nitrogen fractions during manufacture and ripening of Canestrato Pugliese cheese. Lanes: 1, ovine casein (CN) standard; 2, curd after moulding; 3 - 11, cheese (post dry salting) after 1, 3, 7, 15, 30, 45, 60, 75 and 90 days of ripening, respectively.

(A)



(B)

