

Essential Parameters for Structural Analysis and Dereplication by ^1H NMR Spectroscopy

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■ SUPPORTING INFORMATION

■ TABLE OF CONTENTS

	Table of Contents	page(s)
S1	HNMR and HiFSA Data for Compound 1 (Case Study 1)	S-3 to S-6
S2	HNMR and HiFSA Data for Compound 2 (Case Study 2)	S-7 to S-10
S3	HNMR and HiFSA Data for Compound 3 (Case Study 3)	S-11 to S13
S4	HNMR and HiFSA Data for Compound 4 (Case Study 4)	S-14 to S17
S5	Supporting Tables for Case Study 4 (Agnuside, Compound 4)	S-18
S6	HNMR and HiFSA Data for Compound 5 (Case Study 5)	S-19 to S-22
S7	HNMR and HiFSA Data for Compound 6 (Case Study 6)	S-23 to S-25
S8	HNMR ^{and} HiFSA Data for Compound 7 (Case Study 7)	S-26 to S-29
S9	HNMR Spectral Simulation for Micromolide (9) (Case Study 9)	S-30
S10	HNMR and HiFSA Data for Compound 10 (Case Study10)	S-31 to S-32
S11	Raw NMR Data & Software-based NMR Analysis	S-33 to S-34

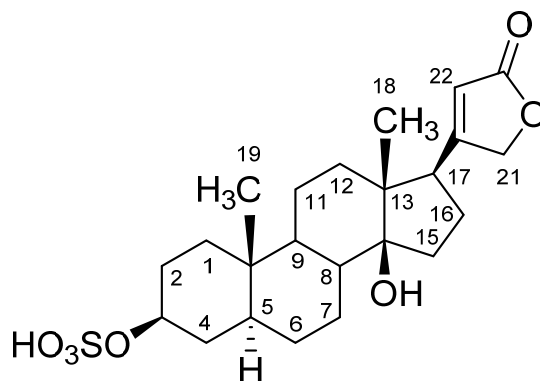
S1: HNMR and HiFSA Data for Compound 1 (Case Study 1)

S1a: HiFSA Profile of Uzarigenin-3-sulfate (1)

In CD₃OD (600 MHz, 298 K)

Proton	δ (ppm)	$\Delta\nu_{1/2}$ (Hz)	Coupling	J(Hz)	Coupling	J(Hz)
H1A	1.0478	1.91	H1A / H1B	-13.33	H8A / H7A	3.61
H1B	1.7740	2.19	H1A / H18	-0.64	H8A / H7B	12.28
H2A	2.0134	1.79	H1A / H2A	3.83	H9A / H11A	12.51
H2B	1.5307	1.76	H1A / H2B	13.99	H9A / H11B	3.60
H3A	4.2470	1.80	H1B / H2A	3.31	H11A / H11B	-13.55
H4A	1.4155	1.66	H1B / H2B	3.96	H12A / H12B	-13.69
H4B	1.8255	1.77	H2A / H2B	-12.99	H12A / H11A	13.78
H5A	1.1579	1.96	H2A / H4B	1.99	H12A / H11B	3.83
H6A	1.2731	1.99	H2A / H3A	4.91	H12A / H19	-0.66
H6B	1.3669	2.14	H2B / H3A	11.51	H12B / H11A	3.65
H7A	2.0330	1.96	H4A / H5A	12.88	H12B / H11B	3.03
H7B	1.1116	1.93	H4A / H4B	-12.63	H15A / H15B	-13.71
H8A	1.5839	2.02	H4A / H3A	11.32	H15A / H16A	10.26
H9A	0.9959	2.15	H4B / H3A	4.92	H15A / H16B	10.12
H11A	1.2830	2.27	H5A / H4B	3.06	H15B / H16A	9.13
H11B	1.5246	2.18	H6A / H6B	-13.25	H15B / H16B	1.77
H12A	1.4525	2.18	H6A / H5A	12.37	H16A / H16B	-13.87
H12B	1.4938	2.06	H6B / H5A	2.98	H17A / H16A	5.53
H15A	2.1079	1.71	H7A / H7B	-12.93	H17A / H16B	9.69
H15B	1.6954	1.81	H7A / H6A	3.71	H17A / H22A	-0.69
H16A	1.8529	1.73	H7A / H6B	2.77	H21A / H21B	-18.32
H16B	2.1511	1.82	H7B / H6A	13.20	H22A / H21A	-1.74
H17A	2.8217	1.67	H7B / H6B	4.14	H22A / H21B	-1.82
H18*	0.8364	1.19	H8A / H9A	11.57		
H19*	0.8761	1.44				
H21A	5.0209	1.49				
H21B	4.9105	0.93				
H22A	5.8883	0.89				

* The Me groups were assigned based on the ¹H-prediction.



S1b. Complete HiFSA Profile of Uzarigenin-3-sulfate (1) in CD₃OD (600 MHz, 298 K) using PERCH's parameters (.pms) format

* NEW: the lines beginning by * are comment lines !
 * To keep all the chemical shifts fixed during iteration
 * replace "CHEMICAL SHIFTS(HZ):" by "..SHIFTS(HZ): fixed"
 * The couplings can be fixed in the same way

NMR-data: [path]

#Date 6. 3.2014; Time 14:10: 4 perch.bst

CHEMICAL SHIFTS (PPM) :

PROTON	2*SPIN=	1 SPECIES=1H	POPULATION(Y)=	0.09225
H8A	/ 1	1.583875 1*1*1	STAT=Y	PRED= 1.728 RANGE= 0.417 WIDTH(Y)= 2.028 RESP(Y)= 0.9067 HSQC= C8
H9A	/ 1	0.995865 1*1*1	STAT=Y	PRED= 0.965 RANGE= 0.232 WIDTH(Y)= 2.162 RESP(Y)= 0.9030 HSQC= C9
H17A	/ 1	2.821666 1*1*1	STAT=Y	PRED= 2.489 RANGE= 0.322 WIDTH(Y)= 2.059 RESP(Y)= 0.9087 HSQC= C17
H12A	/ 1	1.452478 1*1*1	STAT=Y	PRED= 1.034 RANGE= 0.250 WIDTH(Y)= 2.247 RESP(Y)= 0.8674 HSQC= C12
H12B	/ 1	1.493789 1*1*1	STAT=Y	PRED= 1.649 RANGE= 0.260 WIDTH(Y)= 2.064 RESP(Y)= 0.8989 HSQC= C12
H11A	/ 1	1.282972 1*1*1	STAT=Y	PRED= 1.330 RANGE= 0.212 WIDTH(Y)= 2.293 RESP(Y)= 0.9246 HSQC= C11
H11B	/ 1	1.524588 1*1*1	STAT=Y	PRED= 1.565 RANGE= 0.252 WIDTH(Y)= 2.200 RESP(Y)= 0.8857 HSQC= C11
H15A	/ 1	2.107928 1*1*1	STAT=Y	PRED= 1.809 RANGE= 0.462 WIDTH(Y)= 1.718 RESP(Y)= 0.8625 HSQC= C15
H15B	/ 1	1.695405 1*1*1	STAT=Y	PRED= 1.723 RANGE= 0.265 WIDTH(Y)= 1.854 RESP(Y)= 0.8999 HSQC= C15
H7A	/ 1	2.032966 1*1*1	STAT=Y	PRED= 1.948 RANGE= 0.277 WIDTH(Y)= 1.968 RESP(Y)= 0.8894 HSQC= C7
H7B	/ 1	1.111591 1*1*1	STAT=Y	PRED= 1.167 RANGE= 0.217 WIDTH(Y)= 1.938 RESP(Y)= 0.8698 HSQC= C7
H16A	/ 1	1.852888 1*1*1	STAT=Y	PRED= 1.912 RANGE= 0.437 WIDTH(Y)= 1.743 RESP(Y)= 0.8865 HSQC= C16
H16B	/ 1	2.151134 1*1*1	STAT=Y	PRED= 1.712 RANGE= 0.430 WIDTH(Y)= 1.933 RESP(Y)= 0.9059 HSQC= C16
H18	/ 1	0.876058 1*1*3	STAT=Y	PRED= 0.838 RANGE= 0.225 WIDTH(Y)= 1.465 RESP(Y)= 0.9045 HSQC= C19
H6A	/ 1	1.273053 1*1*1	STAT=Y	PRED= 1.320 RANGE= 0.155 WIDTH(Y)= 1.984 RESP(Y)= 0.9488 HSQC= C6
H6B	/ 1	1.366855 1*1*1	STAT=Y	PRED= 1.342 RANGE= 0.207 WIDTH(Y)= 2.155 RESP(Y)= 0.8995 HSQC= C6
H1A	/ 1	1.047752 1*1*1	STAT=Y	PRED= 0.887 RANGE= 0.152 WIDTH(Y)= 1.991 RESP(Y)= 0.8741 HSQC= C1
H1B	/ 1	1.773988 1*1*1	STAT=Y	PRED= 1.606 RANGE= 0.230 WIDTH(Y)= 2.211 RESP(Y)= 0.9089 HSQC= C1
H19	/ 1	0.836429 1*1*3	STAT=Y	PRED= 0.833 RANGE= 0.150 WIDTH(Y)= 1.254 RESP(Y)= 0.8876 HSQC= C18
H2A	/ 1	2.013384 1*1*1	STAT=Y	PRED= 2.107 RANGE= 0.212 WIDTH(Y)= 1.949 RESP(Y)= 0.8985 HSQC= C2
H2B	/ 1	1.530692 1*1*1	STAT=Y	PRED= 1.849 RANGE= 0.300 WIDTH(Y)= 1.799 RESP(Y)= 0.8772 HSQC= C2
H4A	/ 1	1.415541 1*1*1	STAT=Y	PRED= 1.857 RANGE= 0.372 WIDTH(Y)= 1.665 RESP(Y)= 0.8898 HSQC= C4
H22A	/ 1	5.888342 1*1*1	STAT=Y	PRED= 6.160 RANGE= 0.547 WIDTH(Y)= 0.940 RESP(Y)= 0.9176 HSQC= C22
H5A	/ 1	1.157920 1*1*1	STAT=Y	PRED= 1.135 RANGE= 0.160 WIDTH(Y)= 1.968 RESP(Y)= 0.9201 HSQC= C5
H4B	/ 1	1.825476 1*1*1	STAT=Y	PRED= 1.852 RANGE= 0.260 WIDTH(Y)= 1.773 RESP(Y)= 0.8853 HSQC= C4
H3A	/ 1	4.247007 1*1*1	STAT=Y	PRED= 4.365 RANGE= 0.367 WIDTH(Y)= 1.814 RESP(Y)= 0.8898 HSQC= C3
H21A	/ 1	5.020935 1*1*1	STAT=Y	PRED= 4.903 RANGE= 0.222 WIDTH(Y)= 1.516 RESP(Y)= 0.9583 HSQC= C21
H21B	/ 1	4.910519 1*1*1	STAT=Y	PRED= 4.866 RANGE= 0.202 WIDTH(Y)= 1.310 RESP(Y)= 1.0000 HSQC= C21
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MeOH	/ 2	3.300116 1*1*1	STAT=Y	PRED= 3.300 RANGE= 0.050 WIDTH(Y)= 1.056 RESP(Y)= 1.0000
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MET_D1/	2	-99.999998 1*1*1	STAT=Y	PRED= -99.999 RANGE= 0.050 WIDTH(Y)= 1.500 RESP(Y)= 1.0000
MET_D2/	2	-99.999998 1*1*1	STAT=Y	PRED= -99.999 RANGE= 0.050 WIDTH(Y)= 1.500 RESP(Y)= 1.0000
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WATER	/ 3	4.888587 1*1*1	STAT=Y	PRED= 4.891 RANGE= 0.050 WIDTH(Y)= 4.723 RESP(Y)= 1.0000

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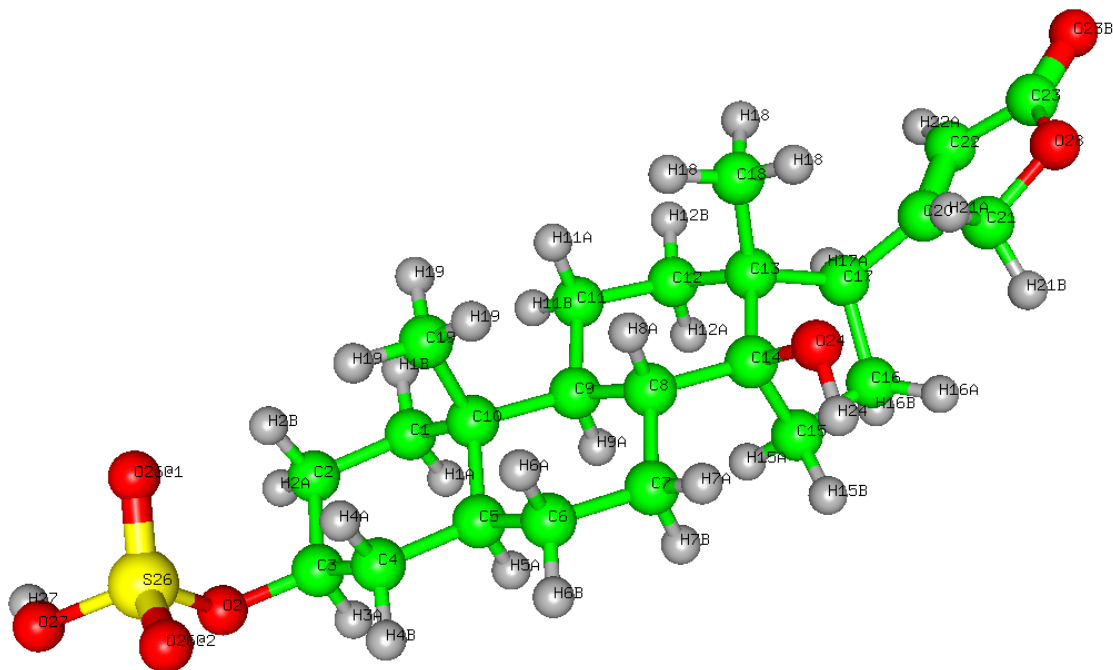
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J25_34	3.6052	J H8A	H7A	STAT=Y	PRED= 3.590 RANGE= 3.800
J25_35	12.2813	J H8A	H7B	STAT=Y	PRED= 12.780 RANGE= 2.560
J26_30	12.5121	J H9A	H11A	STAT=Y	PRED= 12.590 RANGE= 2.560
J26_31	3.6044	J H9A	H11B	STAT=Y	PRED= 2.980 RANGE= 2.800
J27_36	5.5309	J H17A	H16A	STAT=Y	PRED= 3.960 RANGE= 9.200
J27_37	9.6920	J H17A	H16B	STAT=Y	PRED= 10.720 RANGE= 6.400
J27_51	-0.6894	J H17A	H22A	STAT=Y	PRED= -2.500 RANGE= 1.200
J28_29	-13.6932	J H12A	H12B	STAT=Y	PRED= -13.060 RANGE= 1.500
J28_30	13.7807	J H12A	H11A	STAT=Y	PRED= 12.850 RANGE= 2.560
J28_31	3.8316	J H12A	H11B	STAT=Y	PRED= 3.800 RANGE= 3.800
J28_38	-0.6643	J H12A	H19	STAT=Y	PRED= -0.780 RANGE= 0.400
J29_30	3.6546	J H12B	H11A	STAT=Y	PRED= 3.520 RANGE= 3.800
J29_31	3.0316	J H12B	H11B	STAT=Y	PRED= 3.040 RANGE= 3.800
J30_31	-13.5488	J H11A	H11B	STAT=Y	PRED= -13.170 RANGE= 1.500
J32_33	-13.7061	J H15A	H15B	STAT=Y	PRED= -12.450 RANGE= 2.000
J32_36	10.2649	J H15A	H16A	STAT=Y	PRED= 6.700 RANGE= 6.000
J32_37	10.1202	J H15A	H16B	STAT=Y	PRED= 10.340 RANGE= 4.400
J33_36	9.1272	J H15B	H16A	STAT=Y	PRED= 10.440 RANGE= 4.400
J33_37	1.7733	J H15B	H16B	STAT=Y	PRED= 1.430 RANGE= 4.800
J34_35	-12.9331	J H7A	H7B	STAT=Y	PRED= -13.170 RANGE= 1.500
J34_41	3.7097	J H7A	H6A	STAT=Y	PRED= 4.030 RANGE= 3.800
J34_42	2.7684	J H7A	H6B	STAT=Y	PRED= 2.410 RANGE= 2.800
J35_41	13.2040	J H7B	H6A	STAT=Y	PRED= 12.580 RANGE= 2.560
J35_42	4.1397	J H7B	H6B	STAT=Y	PRED= 4.240 RANGE= 3.800
J36_37	-13.8660	J H16A	H16B	STAT=Y	PRED= -12.770 RANGE= 2.000
J41_42	-13.2526	J H6A	H6B	STAT=Y	PRED= -13.210 RANGE= 1.500
J41_57	12.3742	J H6A	H5A	STAT=Y	PRED= 12.460 RANGE= 2.560
J42_57	2.9837	J H6B	H5A	STAT=Y	PRED= 2.790 RANGE= 2.800

J43_44	-13.3260	J H1A	H1B	STAT=Y	PRED=	-13.110	RANGE=	1.500
J43_45	-0.6363	J H1A	H18	STAT=Y	PRED=	-0.790	RANGE=	0.400
J43_48	3.8301	J H1A	H2A	STAT=Y	PRED=	4.360	RANGE=	3.800
J43_49	13.9914	J H1A	H2B	STAT=Y	PRED=	12.910	RANGE=	2.560
J44_48	3.3128	J H1B	H2A	STAT=Y	PRED=	2.510	RANGE=	2.800
J44_49	3.9635	J H1B	H2B	STAT=Y	PRED=	4.180	RANGE=	3.800
J48_49	-12.9878	J H2A	H2B	STAT=Y	PRED=	-12.860	RANGE=	1.500
J48_58	1.9948	J H2A	H4B	STAT=Y	PRED=	2.710	RANGE=	1.280
J48_59	4.9081	J H2A	H3A	STAT=Y	PRED=	4.400	RANGE=	3.800
J49_59	11.5103	J H2B	H3A	STAT=Y	PRED=	10.140	RANGE=	2.560
J50_57	12.8753	J H4A	H5A	STAT=Y	PRED=	12.860	RANGE=	2.560
J50_58	-12.6342	J H4A	H4B	STAT=Y	PRED=	-12.830	RANGE=	1.500
J50_59	11.3207	J H4A	H3A	STAT=Y	PRED=	10.390	RANGE=	2.560
J51_64	-1.7403	J H22A	H21A	STAT=Y	PRED=	-2.060	RANGE=	1.200
J51_65	-1.8208	J H22A	H21B	STAT=Y	PRED=	-2.070	RANGE=	1.200
J57_58	3.0575	J H5A	H4B	STAT=Y	PRED=	2.960	RANGE=	2.800
J58_59	4.9156	J H4B	H3A	STAT=Y	PRED=	4.200	RANGE=	3.800
J64_65	-18.3240	J H21A	H21B	STAT=Y	PRED=	-10.080	RANGE=	4.000
JMETH1	1.6500	J MeOH	MET_D1	STAT=Y	PRED=	1.650	RANGE=	0.150
JMETH1	1.6500	J MeOH	MET_D2	STAT=Y	PRED=	1.650	RANGE=	0.150

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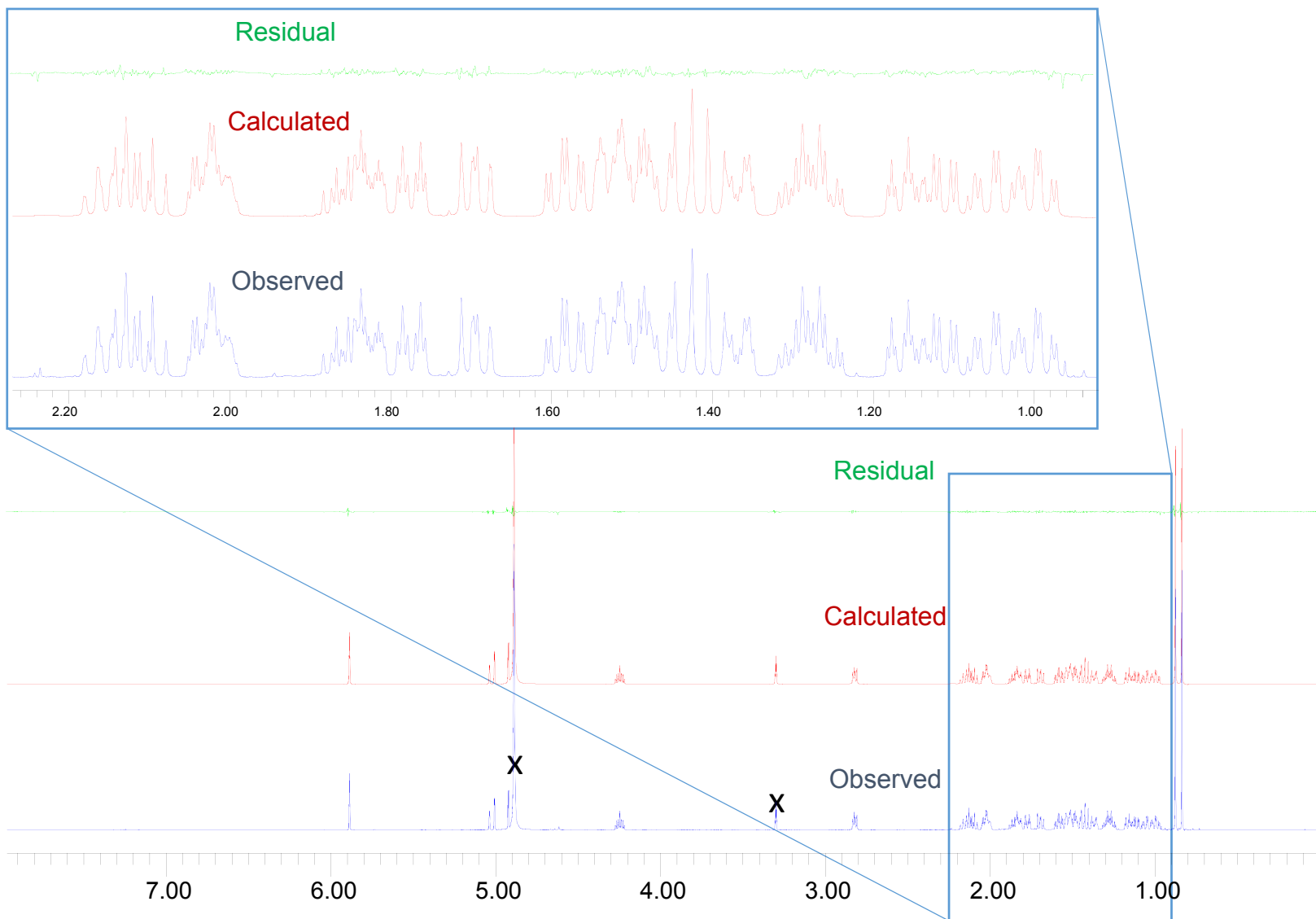
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 -0.03494010 = Right frequency (ppm)
 0.000 = Acquisition time (s, for QMTLS)
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 52.020 = GAUSSIAN (% , 0=use default from INF)
 0.000 = Dispersion contribution (% , 0=use default from INF)
 0.00000000 = Decoupling frequency (for DORES)

END of FILE



S1c. The ^1H NMR Fingerprint of Uzarigenin-3-sulfate (1) in CD_3OD (600 MHz, 298 K)

The X marks on the observed spectrum indicate the solvent signals, CHD_2OD and HOD , which were fitted as well.



S2: HNMR and HiFSA Data for Compound 2 (Case Study 2)

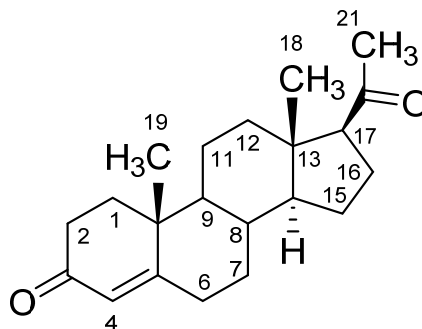
S2a: HiFSA Profile of Progesterone (2)

In CD₃OD (900 MHz, 298 K)

Proton	δ (ppm)	$\Delta\nu_{1/2}$ (Hz)
H1A	1.7420	1.78
H1B	2.1171	1.64
H2A	2.3177	1.42
H2B	2.5099	1.49
H4A	5.7374	1.22
H6A	2.5161	1.57
H6B	2.3365	1.58
H7A	1.9281	1.64
H7B	1.1016	1.65
H8A	1.6661	1.81
H9A	1.0521	1.85
H11A	1.5394	1.60
H11B	1.6990	1.77
H12A	1.5239	1.79
H12B	2.1126	1.71
H14A	1.2640	1.86
H15A	1.7631	1.61
H15B	1.3136	1.56
H16A	2.1740	1.42
H16B	1.7026	1.42
H17A	2.6701	2.01
H18*	0.6963	1.43
H19*	1.2593	1.42
H21	2.1449	1.15

Coupling	J(Hz)	Coupling	J(Hz)
H1A / H1B	-13.40	H11A / H11B	-13.90
H1A / H19	-0.60	H12A / H12B	-12.48
H1A / H2A	4.42	H12A / H11A	13.26
H1A / H2B	14.86	H12A / H11B	4.26
H1B / H2A	3.05	H12A / H18	-0.55
H1B / H2B	5.21	H12B / H11A	4.10
H2A / H2B	-17.04	H12B / H11B	2.86
H2A / H4A	1.00	H14A / H8A	10.83
H6A / H6B	-14.65	H14A / H15A	7.31
H6A / H4A	-1.90	H14A / H15B	12.66
H6B / H4A	-0.56	H15A / H15B	-12.37
H7A / H7B	-12.83	H15A / H16A	3.03
H7A / H6A	5.46	H15A / H16B	9.60
H7A / H6B	2.39	H15B / H16A	11.61
H7B / H6A	13.97	H15B / H16B	6.69
H7B / H6B	4.18	H16A / H16B	-13.83
H8A / H9A	10.82	H17A / H16A	9.19
H8A / H7A	3.54	<u>H17A / H16B</u>	<u>9.32</u>
H8A / H7B	11.70		
H9A / H11A	12.44		
<u>H9A / H11B</u>	<u>4.07</u>		

* The Me groups were assigned based on the ¹H-prediction.



S2b. Complete HiFSA Profile of Progesterone (2) in CD₃OD (900 MHz, 298 K) using PERCH's parameters (.pms) format

* NEW: the lines beginning by * are comment lines !
 * To keep all the chemical shifts fixed during iteration
 * replace "CHEMICAL SHIFTS(HZ):" by "...SHIFTS(HZ): fixed"
 * The couplings can be fixed in the same way

NMR-data: [path]

#Date 19.12.2013; Time 17:10:13 perch.2ND

```
CHEMICAL SHIFTS (PPM): Similarity% = 100.000
PROTON  2*SPIN= 1 SPECIES=1H      POPULATION(Y)= 0.11964
H14A / 1 1.263993 1*1*1 STAT=Y PRED= 1.264 RANGE= 0.072 WIDTH(Y)= 1.859 RESP(Y)= 1.0000
H8A / 1 1.666058 1*1*1 STAT=Y PRED= 1.666 RANGE= 0.072 WIDTH(Y)= 1.806 RESP(Y)= 1.0000
H9A / 1 1.052063 1*1*1 STAT=Y PRED= 1.052 RANGE= 0.072 WIDTH(Y)= 1.849 RESP(Y)= 1.0000
H17A / 1 2.670139 1*1*1 STAT=Y PRED= 2.670 RANGE= 0.072 WIDTH(Y)= 2.007 RESP(Y)= 1.0000
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H11A / 1 1.539393 1*1*1 STAT=Y PRED= 1.539 RANGE= 0.072 WIDTH(Y)= 1.597 RESP(Y)= 1.0000
H11B / 1 1.699024 1*1*1 STAT=Y PRED= 1.699 RANGE= 0.072 WIDTH(Y)= 1.772 RESP(Y)= 1.0000
H15A / 1 1.763103 1*1*1 STAT=Y PRED= 1.763 RANGE= 0.072 WIDTH(Y)= 1.611 RESP(Y)= 1.0000
H15B / 1 1.313565 1*1*1 STAT=Y PRED= 1.314 RANGE= 0.072 WIDTH(Y)= 1.555 RESP(Y)= 1.0000
H7A / 1 1.928077 1*1*1 STAT=Y PRED= 1.928 RANGE= 0.072 WIDTH(Y)= 1.640 RESP(Y)= 1.0000
H7B / 1 1.101633 1*1*1 STAT=Y PRED= 1.102 RANGE= 0.072 WIDTH(Y)= 1.645 RESP(Y)= 1.0000
H16A / 1 2.174043 1*1*1 STAT=Y PRED= 2.174 RANGE= 0.072 WIDTH(Y)= 1.415 RESP(Y)= 1.0000
H16B / 1 1.702611 1*1*1 STAT=Y PRED= 1.703 RANGE= 0.072 WIDTH(Y)= 1.417 RESP(Y)= 1.0000
H18 / 1 0.696299 1*1*3 STAT=Y PRED= 0.696 RANGE= 0.072 WIDTH(Y)= 1.425 RESP(Y)= 1.0000
H6A / 1 2.516118 1*1*1 STAT=Y PRED= 2.516 RANGE= 0.072 WIDTH(Y)= 1.567 RESP(Y)= 1.0000
H6B / 1 2.336540 1*1*1 STAT=Y PRED= 2.337 RANGE= 0.072 WIDTH(Y)= 1.580 RESP(Y)= 1.0000
H1A / 1 1.742003 1*1*1 STAT=Y PRED= 1.742 RANGE= 0.072 WIDTH(Y)= 1.779 RESP(Y)= 1.0000
H1B / 1 2.117062 1*1*1 STAT=Y PRED= 2.117 RANGE= 0.072 WIDTH(Y)= 1.638 RESP(Y)= 1.0000
H19 / 1 1.259275 1*1*3 STAT=Y PRED= 1.259 RANGE= 0.072 WIDTH(Y)= 1.422 RESP(Y)= 1.0000
H2A / 1 2.317691 1*1*1 STAT=Y PRED= 2.318 RANGE= 0.072 WIDTH(Y)= 1.416 RESP(Y)= 1.0000
H2B / 1 2.509924 1*1*1 STAT=Y PRED= 2.510 RANGE= 0.072 WIDTH(Y)= 1.494 RESP(Y)= 1.0000
H4A / 1 5.737395 1*1*1 STAT=Y PRED= 5.737 RANGE= 0.072 WIDTH(Y)= 1.220 RESP(Y)= 1.0000
H21 / 1 2.144930 1*1*3 STAT=Y PRED= 2.145 RANGE= 0.072 WIDTH(Y)= 1.152 RESP(Y)= 1.0000
MET_H  2*SPIN= 1 SPECIES=1H      POPULATION(Y)= 0.69264
MeOH / 2 3.329850 1*1*1 STAT=Y PRED= 3.330 RANGE= 0.049 WIDTH(Y)= 1.308 RESP(Y)= 1.0000
MET_D  2*SPIN= 2 SPECIES=2D      POPULATION(Y)= 0.69264
MET_D1/ 2 -100.000002 1*1*1 STAT=Y PRED= -99.999 RANGE= 0.050 WIDTH(Y)= 1.500 RESP(Y)= 1.0000
MET_D2/ 2 -100.000002 1*1*1 STAT=Y PRED= -99.999 RANGE= 0.050 WIDTH(Y)= 1.500 RESP(Y)= 1.0000
MET_D3/ 2 -100.000002 1*1*1 STAT=Y PRED= -99.999 RANGE= 0.050 WIDTH(Y)= 1.500 RESP(Y)= 1.0000
MET_OH 2*SPIN= 1 SPECIES=1H      POPULATION(Y)= 0.17011
WATER / 3 4.899922 1*1*1 STAT=Y PRED= 4.900 RANGE= 0.049 WIDTH(Y)= 2.934 RESP(Y)= 1.0000
```

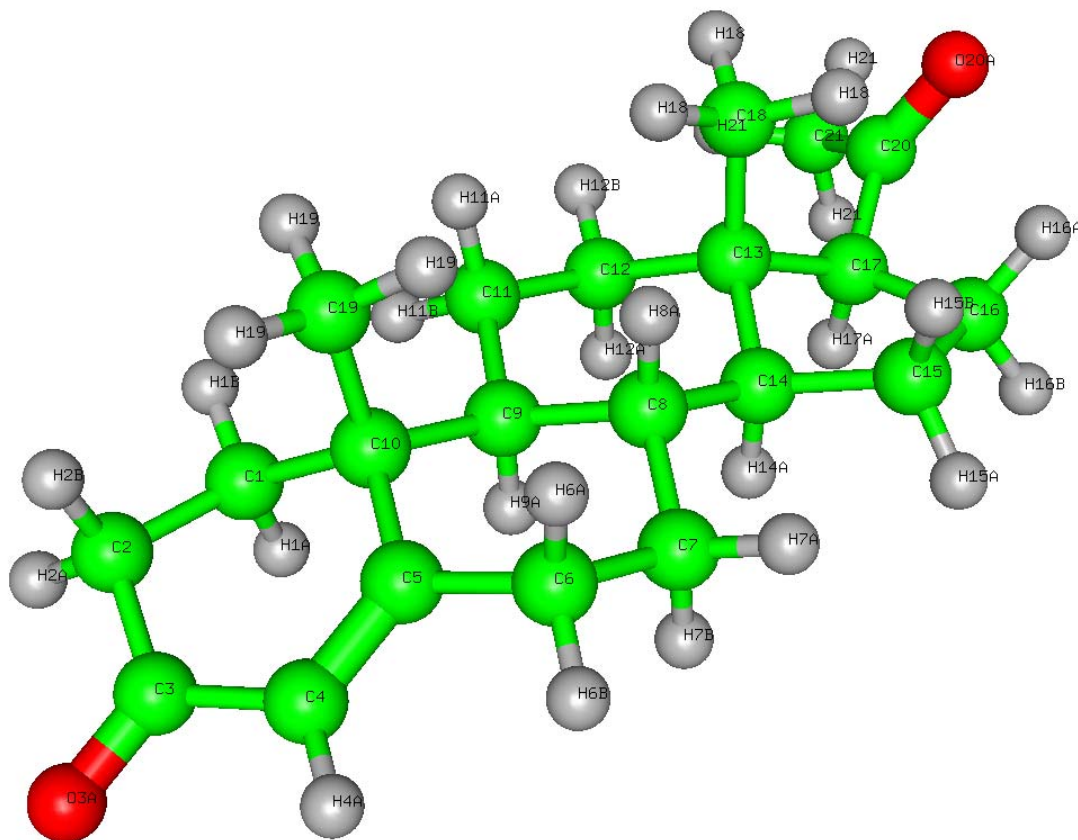
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COUPLING CONSTANTS (HZ): Similarity% = 26.218
J24_25 10.8287 J H14A H8A STAT=Y PRED= 10.685 RANGE= 0.100
J24_32 7.3130 J H14A H15A STAT=Y PRED= 7.255 RANGE= 0.100
J24_33 12.6579 J H14A H15B STAT=Y PRED= 12.752 RANGE= 0.100
J25_26 10.8204 J H8A H9A STAT=Y PRED= 10.854 RANGE= 0.100
J25_34 3.5446 J H8A H7A STAT=Y PRED= 3.549 RANGE= 0.100
J25_35 11.7014 J H8A H7B STAT=Y PRED= 11.711 RANGE= 0.100
J26_30 12.4418 J H9A H11A STAT=Y PRED= 12.420 RANGE= 2.570
J26_31 4.0651 J H9A H11B STAT=Y PRED= 4.084 RANGE= 0.100
J27_36 9.1884 J H17A H16A STAT=Y PRED= 9.199 RANGE= 0.100
J27_37 9.3186 J H17A H16B STAT=Y PRED= 9.303 RANGE= 0.100
J28_29 -12.4787 J H12A H12B STAT=Y PRED= -12.470 RANGE= 0.100
J28_30 13.2626 J H12A H11A STAT=Y PRED= 13.258 RANGE= 0.100
J28_31 4.2615 J H12A H11B STAT=Y PRED= 4.283 RANGE= 0.100
J28_38 -0.5499 J H12A H18 STAT=Y PRED= -0.482 RANGE= 0.100
J29_30 4.1038 J H12B H11A STAT=Y PRED= 4.113 RANGE= 0.100
J29_31 2.8562 J H12B H11B STAT=Y PRED= 2.860 RANGE= 0.100
J30_31 -13.8953 J H11A H11B STAT=Y PRED= -13.919 RANGE= 0.100
J32_33 -12.3670 J H15A H15B STAT=Y PRED= -12.362 RANGE= 0.100
J32_36 3.0332 J H15A H16A STAT=Y PRED= 3.020 RANGE= 0.100
J32_37 9.5984 J H15A H16B STAT=Y PRED= 9.611 RANGE= 0.100
J33_36 11.6147 J H15B H16A STAT=Y PRED= 11.608 RANGE= 0.100
J33_37 6.6874 J H15B H16B STAT=Y PRED= 6.691 RANGE= 0.100
J34_35 -12.8262 J H7A H7B STAT=Y PRED= -12.830 RANGE= 0.100
J34_41 5.4583 J H7A H6A STAT=Y PRED= 5.454 RANGE= 0.100
J34_42 2.3909 J H7A H6B STAT=Y PRED= 2.385 RANGE= 0.100
J35_41 13.9716 J H7B H6A STAT=Y PRED= 13.974 RANGE= 0.100
J35_42 4.1782 J H7B H6B STAT=Y PRED= 4.187 RANGE= 0.100
J36_37 -13.8262 J H16A H16B STAT=Y PRED= -13.829 RANGE= 0.100
J41_42 -14.6493 J H6A H6B STAT=Y PRED= -14.654 RANGE= 0.100
J41_50 -1.9039 J H6A H4A STAT=Y PRED= -1.899 RANGE= 0.100
J42_50 -0.5646 J H6B H4A STAT=Y PRED= -0.049 RANGE= 0.100
J43_44 -13.3969 J H1A H1B STAT=Y PRED= -13.386 RANGE= 0.100
```


J43_45	-0.6045	J H1A	H19	STAT=Y	PRED= -0.570	RANGE= 0.100
J43_48	4.4215	J H1A	H2A	STAT=Y	PRED= 4.417	RANGE= 0.100
J43_49	14.8637	J H1A	H2B	STAT=Y	PRED= 14.859	RANGE= 0.100
J44_48	3.0490	J H1B	H2A	STAT=Y	PRED= 3.039	RANGE= 0.100
J44_49	5.2053	J H1B	H2B	STAT=Y	PRED= 5.206	RANGE= 0.100
J48_49	-17.0409	J H2A	H2B	STAT=Y	PRED= -17.060	RANGE= 3.400
J48_50	1.0013	J H2A	H4A	STAT=Y	PRED= 0.976	RANGE= 0.100
JMETH1	1.6457	J MeOH	MET_D1	STAT=Y	PRED= 1.650	RANGE= 0.300
JMETH1	1.6457	J MeOH	MET_D2	STAT=Y	PRED= 1.650	RANGE= 0.300

CONTROL PARAMETERS:

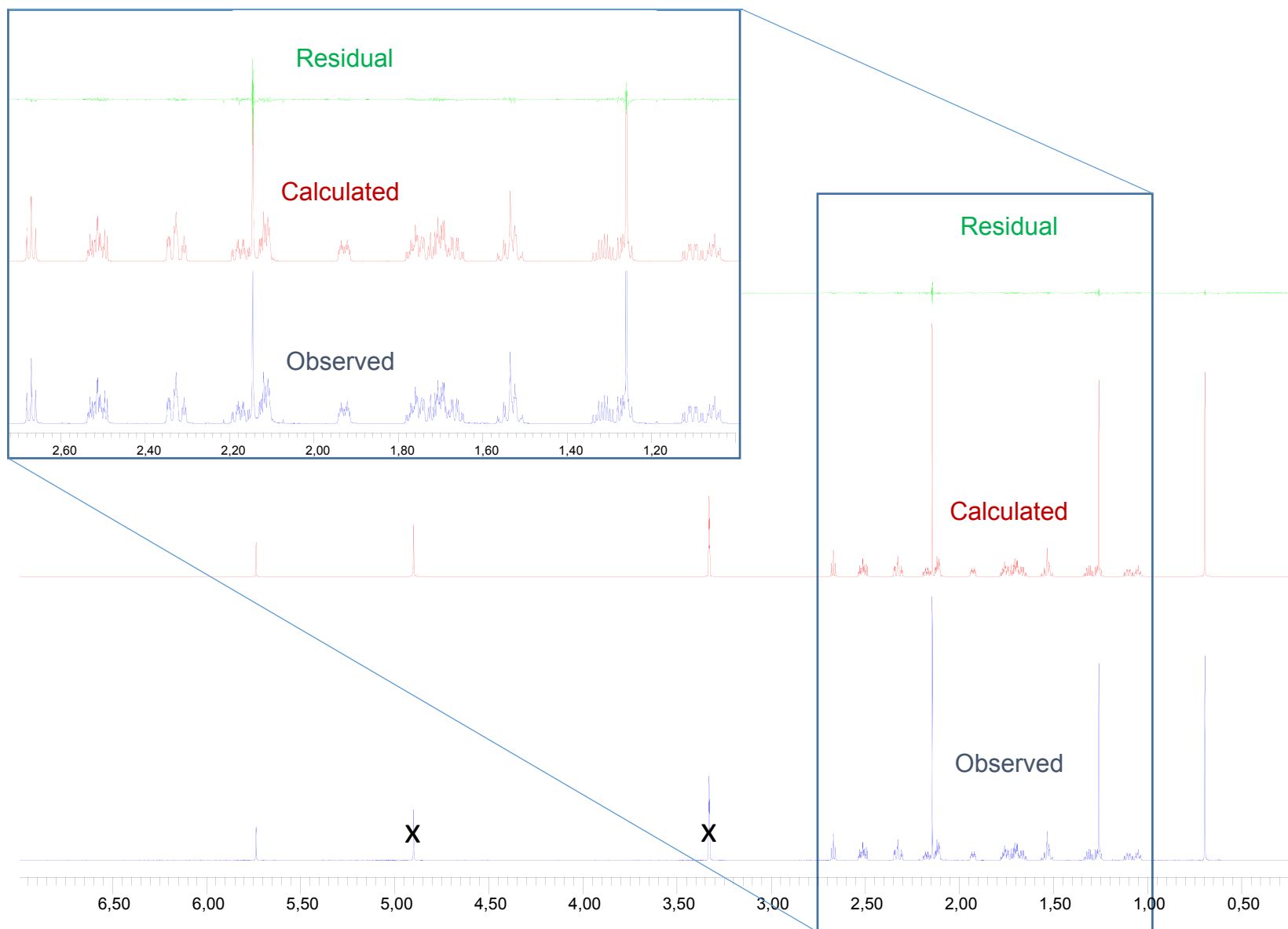
Solvent = none (def. 99% enriched)
 1.000 = Concentration (vol%, def=1.0%)
 0.00100000 = Minimum line-intensity
 0.00100000 = Diagonalization criterium (not in use)
 899.93200102 = FIELD(1H,MHz), used to transform shifts to ppms
 10.98591033 = Left frequency (ppm)
 -0.98817477 = Right frequency (ppm)
 10.000 = Acquisition time (s, for QMFLS)
 0.302 = Line-width (for modes D, P & T, 0=use defaults)
 0.050000000 = Data-point resolution (Hz)
 43.160 = GAUSSIAN (% , 0=use default from INF)
 0.000 = Dispersion contribution (% , 0=use default from INF)
 0.00000000 = Decoupling frequency (for DORES)

END of FILE



S2c. The ^1H NMR Fingerprint of Progesterone (2) in CD_3OD (900 MHz, 298 K)

The X marks on the observed spectrum indicate the solvent signals, CHD_2OD and HOD , which in this case were fitted as well.

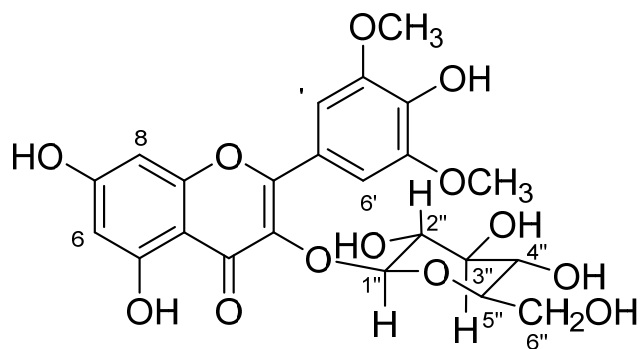


S3: HNMR and HiFSA Data for Compound 3 (Case Study 3)

S3a: HiFSA Profile of Compound 3

In CD₃OD (600 MHz, 298 K)

Proton	δ (ppm)	$\Delta\nu_{1/2}$ (Hz)
H1''	5.4663	0.85
H2''	3.4617	0.99
H3''	3.4473	0.89
H5''	3.2482	1.23
H4''	3.2983	1.66
H6''A	3.5653	1.52
H6''B	3.7467	1.54
H8	6.4193	0.69
H6	6.2088	0.71
H2'_6'	7.5234	0.87
H7'_8'	3.9314	0.92



Coupling	J(Hz)
H1'' / H2''	7.73
H2'' / H3''	9.28
H3'' / H4''	8.80
H5'' / H4''	9.75
H5'' / H6''A	5.64
H5'' / H6''B	2.32
H6''A / H6''B	-11.94
H8 / H6	2.10
H2'_6' / H2'_6'	1.82

S3b. Complete HiFSA Profile of Syringetin-3-O-β-D-glucoside (3) in CD₃OD (600 MHz, 298 K) using PERCH's parameters (.pms) format

* NEW: the lines beginning by * are comment lines !
* To keep all the chemical shifts fixed during iteration
* replace "CHEMICAL SHIFTS(HZ):" by "..SHIFTS(HZ): fixed"
* The couplings can be fixed in the same way

NMR-data: [path]

#Date 7. 3.2014; Time 12:46:15 perch.new

CHEMICAL SHIFTS (PPM):

```
PROTON 2*SPIN= 1 SPECIES=1H POPULATION(Y)= 0.02288
H8 / 1 6.419266 1*1*1 STAT=Y PRED= 6.419 RANGE= 0.150 WIDTH(Y)= 0.691 RESP(Y)= 1.0000 HSQC= C8
H6 / 1 6.208833 1*1*1 STAT=Y PRED= 6.208 RANGE= 0.150 WIDTH(Y)= 0.705 RESP(Y)= 1.0000 HSQC= C6
H2'_6' / 1 7.523360 1*2*1 STAT=Y PRED= 7.577 RANGE= 0.150 WIDTH(Y)= 0.867 RESP(Y)= 1.0000 HSQC= C2_6
H7'_8' / 1 3.931432 2*1*3 STAT=Y PRED= 3.938 RANGE= 0.150 WIDTH(Y)= 0.923 RESP(Y)= 1.0000 HSQC= C7_8
H1" / 1 5.466307 1*1*1 STAT=Y PRED= 5.454 RANGE= 0.150 WIDTH(Y)= 0.851 RESP(Y)= 1.0000 HSQC= C1"
H2" / 1 3.461699 1*1*1 STAT=Y PRED= 3.504 RANGE= 0.150 WIDTH(Y)= 0.988 RESP(Y)= 1.0000 HSQC= C2"
H3" / 1 3.447291 1*1*1 STAT=Y PRED= 3.457 RANGE= 0.150 WIDTH(Y)= 0.893 RESP(Y)= 1.0000 HSQC= C3"
H5" / 1 3.248218 1*1*1 STAT=Y PRED= 3.258 RANGE= 0.150 WIDTH(Y)= 1.229 RESP(Y)= 1.0000 HSQC= C5"
H4" / 1 3.298299 1*1*1 STAT=Y PRED= 3.341 RANGE= 0.150 WIDTH(Y)= 1.663 RESP(Y)= 1.0000 HSQC= C4"
H6"A / 1 3.565335 1*1*1 STAT=Y PRED= 3.715 RANGE= 0.150 WIDTH(Y)= 1.521 RESP(Y)= 1.0000 HSQC= C6"
H6"B / 1 3.746671 1*1*1 STAT=Y PRED= 3.657 RANGE= 0.150 WIDTH(Y)= 1.536 RESP(Y)= 1.0000 HSQC= C6"
MET_H 2*SPIN= 1 SPECIES=1H POPULATION(Y)= 0.64187
MeOH / 2 3.306004 1*1*1 STAT=Y PRED= 3.306 RANGE= 0.050 WIDTH(Y)= 0.805 RESP(Y)= 1.0000
MET_D 2*SPIN= 2 SPECIES=2D POPULATION(Y)= 0.64187
MET_D1/ 2 -100.000003 1*1*1 STAT=Y PRED= -99.999 RANGE= 0.050 WIDTH(Y)= 1.500 RESP(Y)= 1.0000
MET_D2/ 2 -100.000003 1*1*1 STAT=Y PRED= -99.999 RANGE= 0.050 WIDTH(Y)= 1.500 RESP(Y)= 1.0000
MET_OH 2*SPIN= 1 SPECIES=1H POPULATION(Y)= 0.27192
WATER / 3 4.839322 1*1*1 STAT=Y PRED= 4.839 RANGE= 0.050 WIDTH(Y)= 1.982 RESP(Y)= 1.0000
TMS 2*SPIN= 1 SPECIES=1H POPULATION(Y)= 0.06333
TMS / 4 0.000143 1*1*1 STAT=Y PRED= 0.000 RANGE= 0.050 WIDTH(Y)= 0.397 RESP(Y)= 1.0000
```

COUPLING CONSTANTS (HZ):

```
J26_27 2.0990 J H8 H6 STAT=Y PRED= 2.110 RANGE= 1.790
J30_31 1.8166 J H2'_6' H2'_6' STAT=Y PRED= 1.780 RANGE= 1.790
J49_50 7.7336 J H1" H2" STAT=Y PRED= 7.310 RANGE= 3.590
J50_51 9.2783 J H2" H3" STAT=Y PRED= 9.040 RANGE= 3.590
J51_53 8.7986 J H3" H4" STAT=Y PRED= 9.040 RANGE= 3.590
J52_53 9.7451 J H5" H4" STAT=Y PRED= 10.500 RANGE= 2.590
J52_55 5.6420 J H5" H6"A STAT=Y PRED= 5.990 RANGE= 6.400
J52_56 2.3179 J H5" H6"B STAT=Y PRED= 1.980 RANGE= 3.000
J55_56 -11.9398 J H6"A H6"B STAT=Y PRED= -12.300 RANGE= 0.600
JMETH1 1.6450 J MeOH MET_D1 STAT=Y PRED= 1.650 RANGE= 0.150
JMETH1 1.6450 J MeOH MET_D2 STAT=Y PRED= 1.650 RANGE= 0.150
```

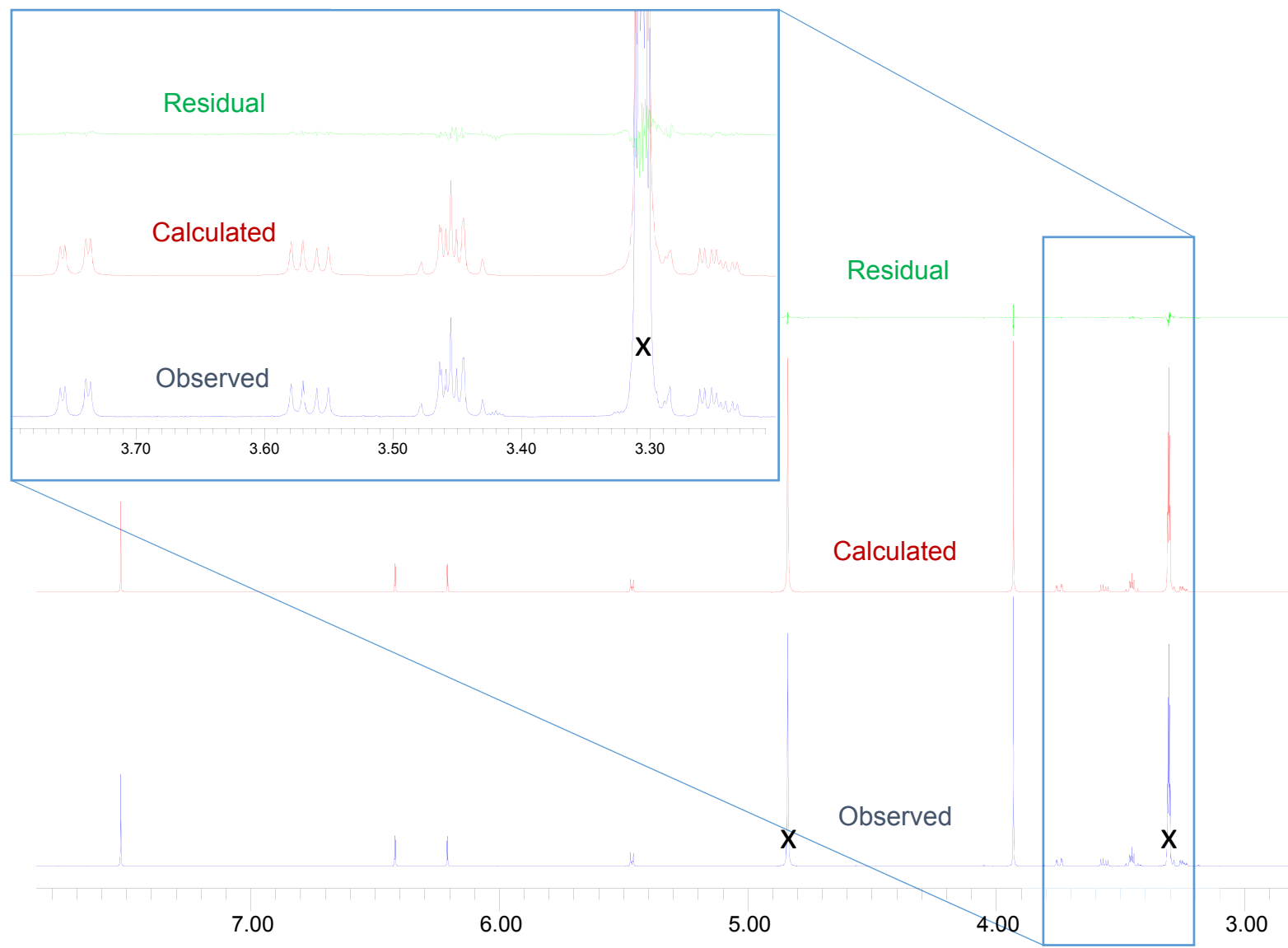
CONTROL PARAMETERS:

```
Solvent = none (def. 99% enriched)
1.000 = Concentration (vol%, def=1.0%)
0.00100000 = Minimum line-intensity
0.00100000 = Diagonalization criterium (not in use)
600.23000000 = FIELD(1H,MHz), used to transform shifts to ppms
14.88688909 = Left frequency (ppm)
-5.13748487 = Right frequency (ppm)
0.000 = Acquisition time (s, for QMtls)
0.371 = Line-width (for modes D, P & T, 0=use defaults)
0.183401705 = Data-point resolution (Hz)
0.000 = GAUSSIAN (% , 0=use default from INF)
0.000 = Dispersion contribution (% , 0=use default from INF)
0.00000000 = Decoupling frequency (for DORES)
```

END of FILE

S3c. The ^1H NMR Fingerprint of Syringetin-3-O- β -D-glucoside (3) in CD_3OD (600 MHz, 298 K)

The X marks on the observed spectrum indicate the solvent signals, CHD_2OD and HOD , which in this case were fitted as well.



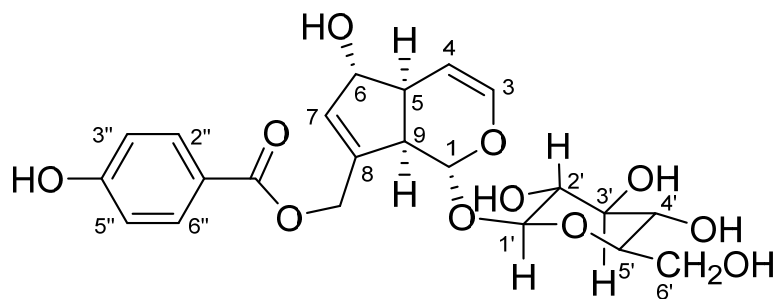
S4: HNMR and HiFSA Data for Compound 4 (Case Study 4)

S4a: HiFSA Profile of Agnuside (4)

In CD₃OD (300 MHz, 298 K)

Proton	δ (ppm)	$\Delta\nu_{1/2}$ (Hz)				
H1	4.9930	1.01				
H1'	4.6958	0.85				
H11A	5.0876	1.17				
H11B	4.9144	1.23				
H2'	3.2422	1.04				
H2_6	7.9182	0.73				
H3	6.3373	0.98				
H3'	3.3760	1.02				
H3_5	6.8410	0.74				
H4	5.1166	0.87				
H4'	3.3178	0.96				
H5	2.7012	1.02				
H5'	3.2748	1.25				
H6	4.4671	1.20				
H6'A	3.8465	1.30				
H6'B	3.6567	1.40				
H7	5.8304	1.05				
H9	2.9923	1.18				

Coupling	J(Hz)	Coupling	J(Hz)
H1' / H2'	7.91	H7 / H11A	-1.61
H2' / H3'	9.28	H7 / H11B	-1.87
H3' / H4'	8.85	H9 / H5	7.84
H3_5 / H3_5	2.25	H9 / H1	7.34
H4 / H3	6.10	H9 / H6	-1.93
H4' / H5'	9.68	H9 / H7	-1.23
H5 / H6	5.46	H9 / H11A	-0.74
H5 / H4	3.90	H9 / H11B	-0.86
H5 / H3	-1.99	H11A / H11B	-14.75
H5' / H6'A	2.25	H2_6 / H2_6	2.58
H5' / H6'B	5.74	H2_6 / H3_5	8.61
H6 / H7	1.50	H3_5 / H2_6	0.35
H6 / H11A	-2.21		
H6 / H11B	-1.71		
H6'A / H6'B	-11.97		



S4b. Complete HiFSA Profile of Agnuside (4) in CD₃OD (300 MHz, 298 K) using PERCH's parameters (.pms) format

* NEW: the lines beginning by * are comment lines !
 * To keep all the chemical shifts fixed during iteration
 * replace "CHEMICAL SHIFTS(HZ):" by "..SHIFTS(HZ): fixed"
 * The couplings can be fixed in the same way

NMR-data: [path]

#\$œ Date 8. 3.2014; Time 13:30: 6 perch.new

CHEMICAL SHIFTS (PPM) :

```

PROTON  2*SPIN= 1 SPECIES=1H  POPULATION(Y)= 0.07698
H9 / 1 2.992252 1*1*1 STAT=Y WIDTH(Y)= 1.182 RESP(Y)= 1.0000 HSQC= C9
H5 / 1 2.701207 1*1*1 STAT=Y WIDTH(Y)= 1.020 RESP(Y)= 1.0000 HSQC= C5
H1 / 1 4.992977 1*1*1 STAT=Y WIDTH(Y)= 1.006 RESP(Y)= 1.0000 HSQC= C1
H6 / 1 4.467097 1*1*1 STAT=Y WIDTH(Y)= 1.198 RESP(Y)= 1.0000 HSQC= C6
H7 / 1 5.830424 1*1*1 STAT=Y WIDTH(Y)= 1.045 RESP(Y)= 1.0000 HSQC= C7
H1' / 1 4.695760 1*1*1 STAT=Y WIDTH(Y)= 0.849 RESP(Y)= 1.0000 HSQC= C1'
H2' / 1 3.242201 1*1*1 STAT=Y WIDTH(Y)= 1.036 RESP(Y)= 1.0000 HSQC= C2'
H4 / 1 5.116609 1*1*1 STAT=Y WIDTH(Y)= 0.868 RESP(Y)= 1.0000 HSQC= C4
H3' / 1 3.376035 1*1*1 STAT=Y WIDTH(Y)= 1.018 RESP(Y)= 1.0000 HSQC= C3'
H4' / 1 3.317849 1*1*1 STAT=Y WIDTH(Y)= 0.955 RESP(Y)= 1.0000 HSQC= C4'
H5' / 1 3.274789 1*1*1 STAT=Y WIDTH(Y)= 1.248 RESP(Y)= 1.0000 HSQC= C5'
H11A / 1 5.087647 1*1*1 STAT=Y WIDTH(Y)= 1.167 RESP(Y)= 1.0000 HSQC= C11
H11B / 1 4.914419 1*1*1 STAT=Y WIDTH(Y)= 1.231 RESP(Y)= 1.0000 HSQC= C11
H3 / 1 6.337333 1*1*1 STAT=Y WIDTH(Y)= 0.979 RESP(Y)= 1.0000 HSQC= C3
H6'A / 1 3.846523 1*1*1 STAT=Y WIDTH(Y)= 1.295 RESP(Y)= 1.0000 HSQC= C6'
H6'B / 1 3.656731 1*1*1 STAT=Y WIDTH(Y)= 1.399 RESP(Y)= 1.0000 HSQC= C6'
H2_6 / 1 7.918190 1*2*1 STAT=Y WIDTH(Y)= 0.734 RESP(Y)= 1.0000 HSQC= C2_6
H3_5 / 1 6.841045 1*2*1 STAT=Y WIDTH(Y)= 0.735 RESP(Y)= 1.0000 HSQC= C3_5
MET_H  2*SPIN= 1 SPECIES=1H  POPULATION(Y)= 0.07520
MeOH / 2 3.308142 1*1*1 STAT=Y PRED= 3.306 RANGE= 0.050 WIDTH(Y)= 0.898 RESP(Y)= 1.0000
MET_D  2*SPIN= 2 SPECIES=2D  POPULATION(Y)= 0.07520
MET_D1/ 2 -99.999990 1*1*1 STAT=Y PRED= -99.999 RANGE= 0.050 WIDTH(Y)= 1.500 RESP(Y)= 1.0000
MET_D2/ 2 -99.999990 1*1*1 STAT=Y PRED= -99.999 RANGE= 0.050 WIDTH(Y)= 1.500 RESP(Y)= 1.0000
TMS 2*SPIN= 1 SPECIES=1H  POPULATION(Y)= 0.01287
TMS / 3 -0.000333 1*1*1 STAT=Y PRED= -0.010 RANGE= 0.050 WIDTH(Y)= 0.486 RESP(Y)= 1.0000
D2O 2*SPIN= 1 SPECIES=1H  POPULATION(Y)= 0.83495
D2O / 4 4.745296 1*1*1 STAT=Y PRED= 4.780 RANGE= 0.050 WIDTH(Y)= 2.208 RESP(Y)= 1.0000
  
```

COUPLING CONSTANTS (HZ) :

```

J34_35 7.8354 J H9 H5 STAT=Y PRED= 7.835 RANGE= 0.001
J34_36 7.3381 J H9 H1 STAT=Y PRED= 7.338 RANGE= 0.001
J34_37 -1.9273 J H9 H6 STAT=Y PRED= -1.937 RANGE= 0.001
J34_38 -1.2314 J H9 H7 STAT=Y PRED= -1.236 RANGE= 0.001
J34_45 -0.7417 J H9 H11A STAT=Y PRED= -0.746 RANGE= 0.001
J34_46 -0.8637 J H9 H11B STAT=Y PRED= -0.866 RANGE= 0.001
J35_37 5.4602 J H5 H6 STAT=Y PRED= 5.460 RANGE= 0.001
J35_41 3.9005 J H5 H4 STAT=Y PRED= 3.901 RANGE= 0.001
J35_47 -1.9899 J H5 H3 STAT=Y PRED= -1.990 RANGE= 0.001
J37_38 1.4969 J H6 H7 STAT=Y PRED= 1.481 RANGE= 0.001
J37_45 -2.2051 J H6 H11A STAT=Y PRED= -2.215 RANGE= 0.001
J37_46 -1.7083 J H6 H11B STAT=Y PRED= -1.702 RANGE= 0.001
J38_45 -1.6078 J H7 H11A STAT=Y PRED= -1.603 RANGE= 0.001
J38_46 -1.8711 J H7 H11B STAT=Y PRED= -1.880 RANGE= 0.001
J39_40 7.9095 J H1' H2' STAT=Y PRED= 7.909 RANGE= 0.001
J40_42 9.2763 J H2' H3' STAT=Y PRED= 9.276 RANGE= 0.001
J41_47 6.0985 J H4 H3 STAT=Y PRED= 6.098 RANGE= 0.001
J42_43 8.8534 J H3' H4' STAT=Y PRED= 8.852 RANGE= 0.001
J43_44 9.6820 J H4' H5' STAT=Y PRED= 9.682 RANGE= 0.001
J44_49 2.2547 J H5' H6'A STAT=Y PRED= 2.254 RANGE= 0.001
J44_50 5.7399 J H5' H6'B STAT=Y PRED= 5.740 RANGE= 0.001
J45_46 -14.7461 J H11A H11B STAT=Y PRED= -14.746 RANGE= 0.001
J49_50 -11.9672 J H6'A H6'B STAT=Y PRED= -11.967 RANGE= 0.001
J55_56 2.5785 J H2_6 H2_6 STAT=Y PRED= 2.572 RANGE= 0.001
J55_57 8.6090 J H2_6 H3_5 STAT=Y PRED= 8.603 RANGE= 0.001
J55_58 0.3474 J H3_5 H2_6 STAT=Y PRED= 0.361 RANGE= 0.001
J57_58 2.2481 J H3_5 H3_5 STAT=Y PRED= 2.247 RANGE= 0.001
JMETH1 1.6500 J MeOH MET_D1 STAT=Y PRED= 1.650 RANGE= 0.150
JMETH1 1.6500 J MeOH MET_D2 STAT=Y PRED= 1.650 RANGE= 0.150
  
```

CONTROL PARAMETERS:

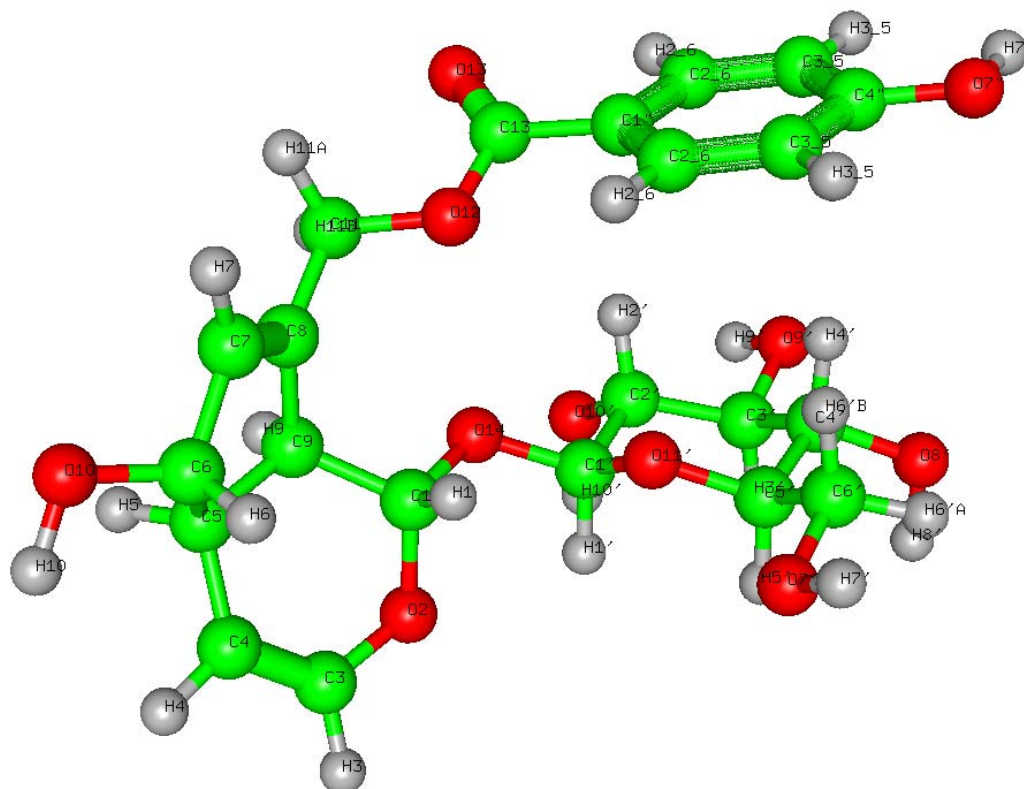
```

Solvent = none (def. 99% enriched)
1.000 = Concentration (vol%, def=1.0%)
0.00100000 = Minimum line-intensity
0.00100000 = Diagonalization criterium (not in use)
300.13460000 = FIELD(1H,MHz), used to transform shifts to ppm
12.12926954 = Left frequency (ppm)
  
```

-0.88577735 = Right frequency (ppm)
0.000 = Acquisition time (s, for QMPLS)
0.484 = Line-width (for modes D, P & T, 0=use defaults)
0.119213397 = Data-point resolution (Hz)
25.974 = GAUSSIAN (% , 0=use default from INF)
0.000 = Dispersion contribution (% , 0=use default from INF)
0.00000000 = Decoupling frequency (for DORES)

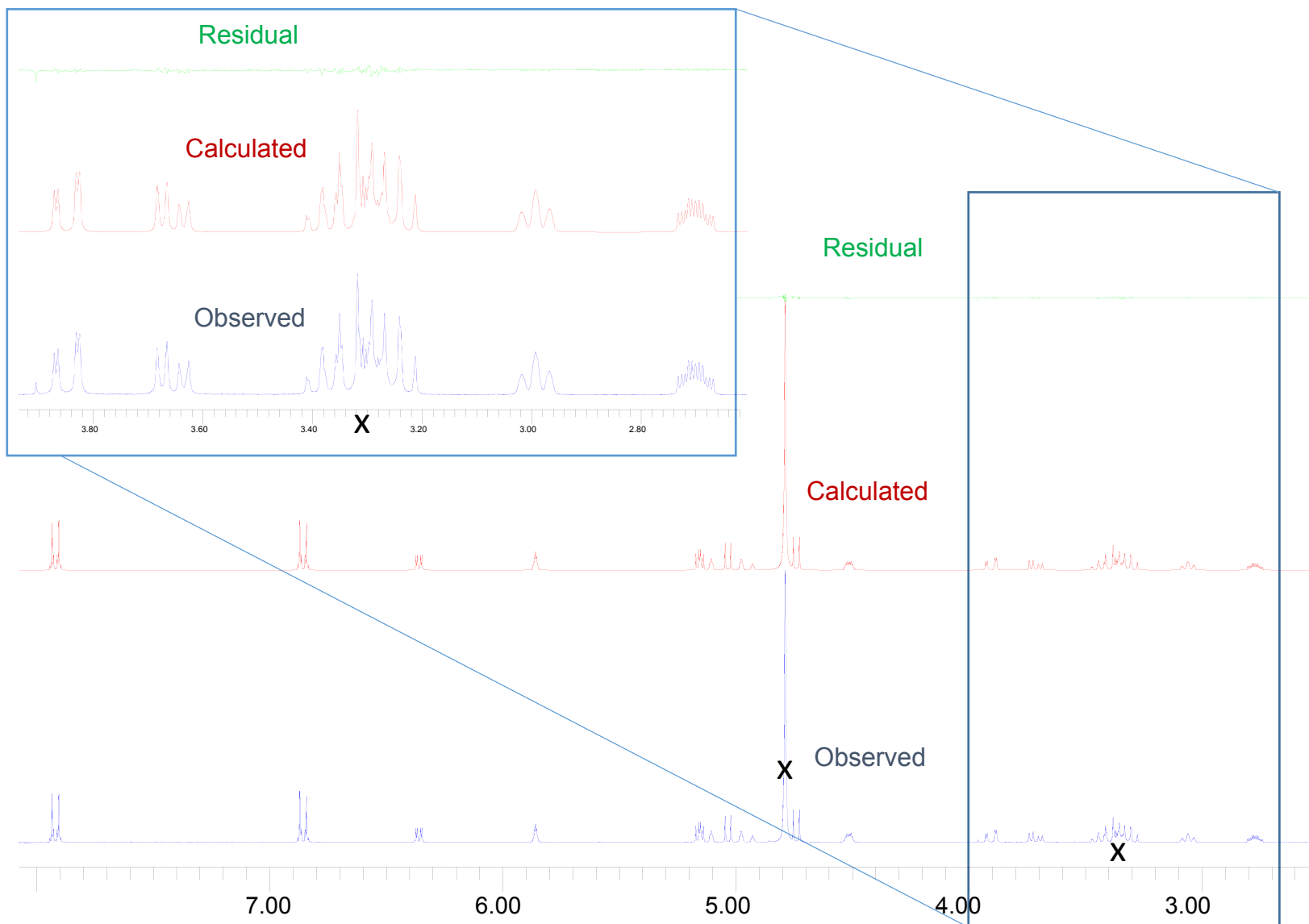
CONSTRAINTS (in equations X0 = 1.0)...use no empty lines
EQUAL MET_D1 = MET_D2

END of FILE



S4c. The ^1H NMR Fingerprint of Agnuside (4) in CD_3OD (300 MHz, 298 K)

The X marks on the observed spectrum indicate the solvent signals, CHD_2OD and HOD , which in this case were fitted as well.



S5: Supporting Tables for Case Study 4 (Agnuside, Compound 4)

NMR-parameters derived from different starting values by iterative spectral analysis. All *J*-couplings and chemical shifts were changed randomly and subsequently optimized using iterative full spin analysis. Subsequent D/T-Mode, Linear baseline, Threshold 0.001, Gaussian-contribution 50% (fixed), Dispersion Contribution 0% (fixed)

Starting Values

Starting Values					
$\delta(A/A')$	$\delta(X/X')$	$J(X,X')$	$J(A/X)$	$J(A,X')$	$J(A,A')$
7.9182	6.8411	2.6329	8.6055	0.3121	2.1721
7.9182	6.8411	4.0000	8.0000	2.0000	3.0000
8.0182	6.7411	4.0000	8.0000	2.0000	3.0000
7.8182	6.9411	4.0000	8.0000	2.0000	3.0000
7.8182	6.9411	6.0000	10.0000	3.0000	5.0000
7.9182	6.8411	2.0000	5.0000	0.1000	1.0000
7.9182	6.8411	8.0000	12.0000	4.0000	6.0000
7.9182	6.8411	8.0000	12.0000	0.1000	1.0000
7.9182	6.8411	2.0000	3.0000	1.0000	1.5000

Optimized by HiFSA, Fitted Spectral Parameters

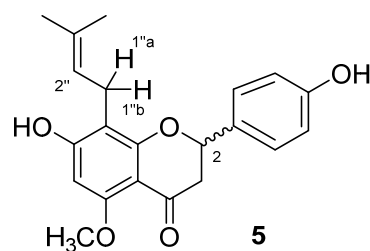
	FIT						
	$\delta(A/A')$	$\delta(X/X')$	$J(X,X')$	$J(A/X)$	$J(A,X')$	$J(A,A')$	RMS
	7.918255	6.841127	2.6354	8.6030	0.3144	2.1687	0.06550
	7.918257	6.841126	2.6321	8.6037	0.3155	2.1690	0.06540
	7.918255	6.841127	2.6365	8.6012	0.3167	2.1633	0.06510
	7.918255	6.841127	2.6365	8.6012	0.3167	2.1633	0.06510
	7.918255	6.841127	2.6349	8.6034	0.3139	2.1698	0.06560
	7.918255	6.841126	2.6348	8.6031	0.3144	2.1684	0.06510
	7.918256	6.841125	2.6341	8.6016	0.3167	2.1674	0.06500
	7.918256	6.841127	2.6349	8.6032	0.3139	2.1695	0.06560
	7.918256	6.841125	2.6352	8.6017	0.3163	2.1667	0.06500
Average	7.918256	6.841126	2.6349	8.6025	0.3154	2.1673	0.06527
STDEVP	0.000001	0.000001	0.0012	0.0009	0.0011	0.0022	0.00023

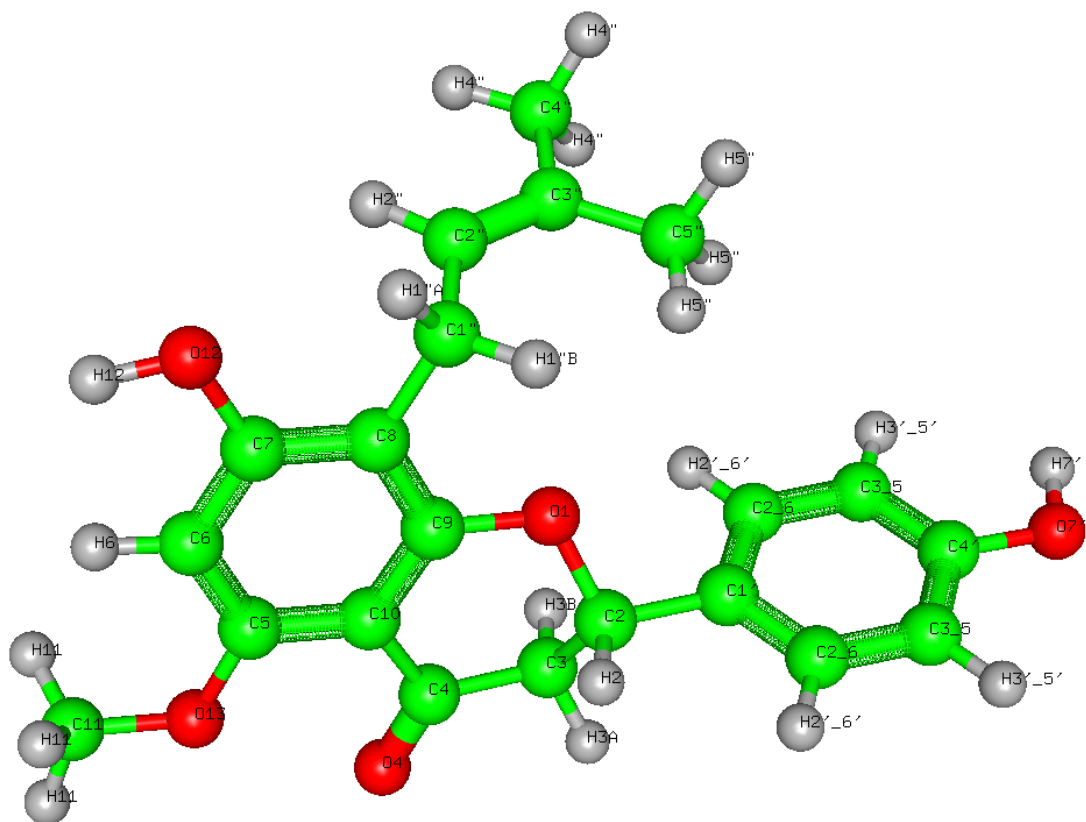
S6: HNMR and HiFSA Data for Compound 5 (Case Study 5)

S6a: HiFSA Profile of Isoxanthohumol (5)

In CD₃OD (500 MHz, 298 K)

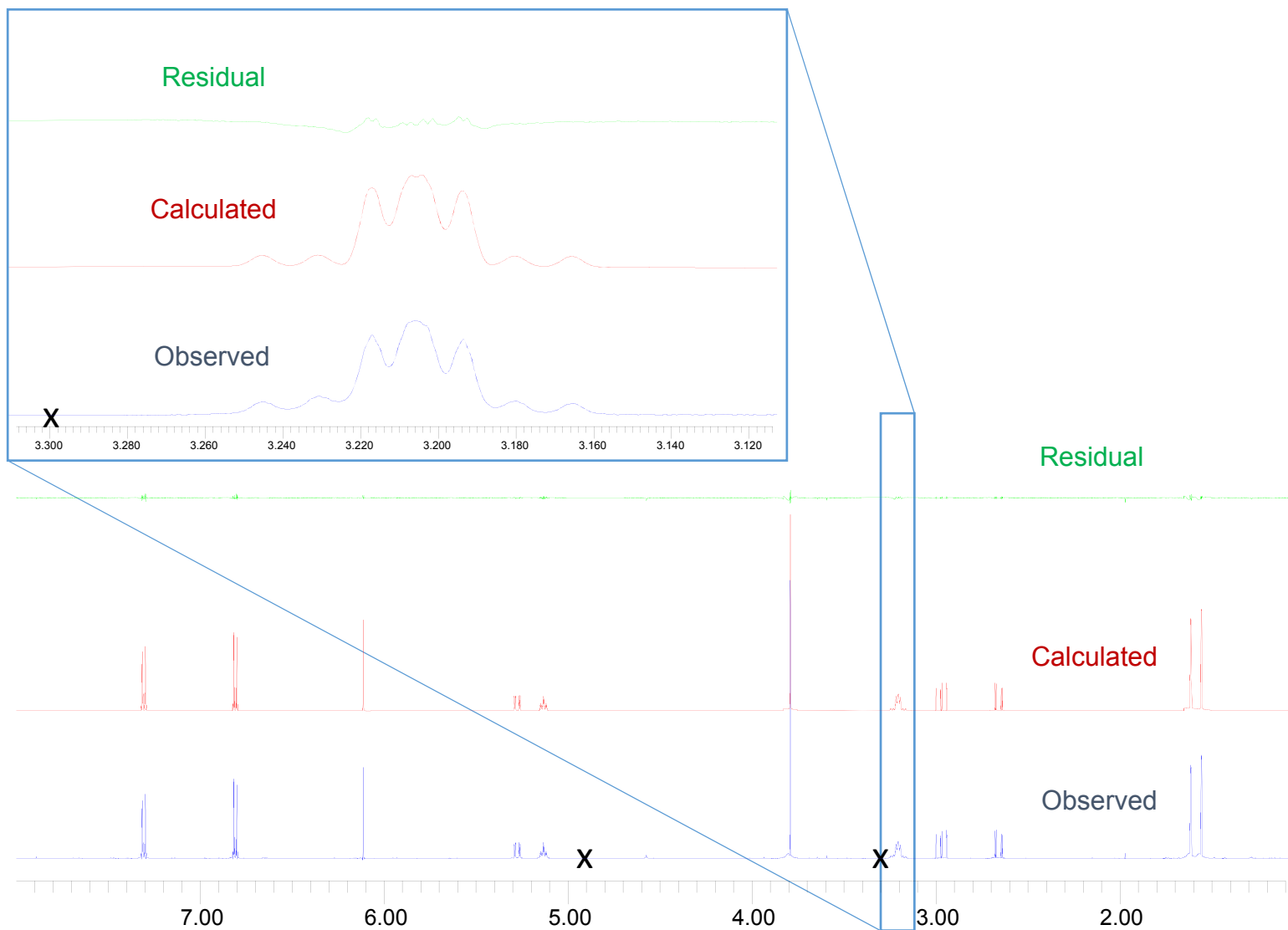
Proton	δ (ppm)	$\Delta\nu_{1/2}$ (Hz)	Coupling	J(Hz)
H2	5.2762	1.48	H2 / H3A	3.00
H3A	2.6598	0.89	H2 / H3B	12.83
H3B	2.9699	0.88	H3A / H3B	-16.65
H6	6.1127	0.99	H1''A / H1''B	-14.08
H11	3.7924	0.96	H1''A / H2''	7.37
H2'_6'	7.3057	1.04	H1''A / H4''	1.17
H3'_5'	6.8093	0.77	H1''B / H2''	7.14
H1''A	3.1935	1.55	H1''B / H4''	1.21
H1''B	3.2177	1.55	H2'_ε / H2'_6	2.39
H2''	5.1338	0.92	H2'_ε / H3'_5	8.44
H4''	1.6147	0.79	H3'_ε / H2'_6	0.34
H5''	1.5572	1.23	H2'' / H4''	-1.29
			H2'' / H5''	-1.39
			H3'_ε / H3'_5	2.68
			H4'' / H5''	-0.41





S6c. The ^1H NMR Fingerprint of Isoxanthohumol (5) in CD_3OD (500 MHz, 298 K)

The X marks on the observed spectrum indicate that solvent signals, CHD_2OD and HOD , were removed artificially in order to allow the residual to be scaled such that it reflects the differences arising from the compound and non-solvent impurities of the sample, rather than the relatively large solvent signals.

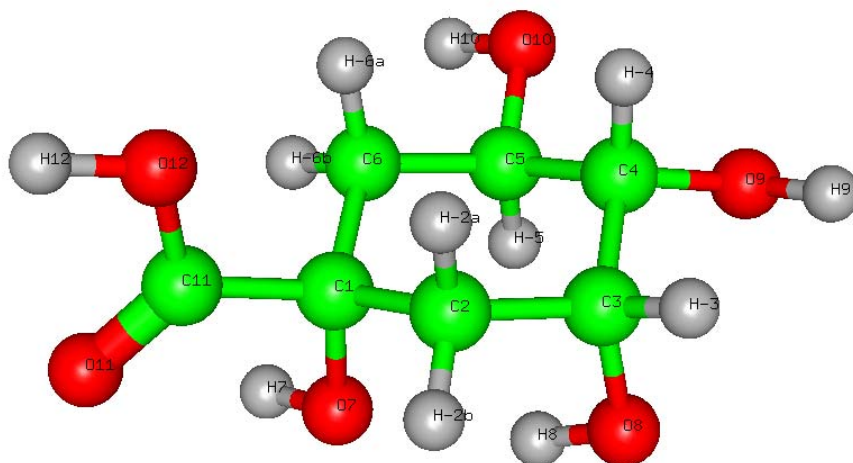
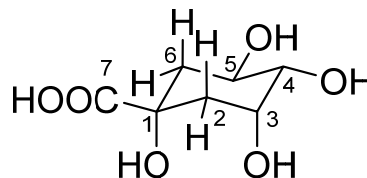


S7: HNMR and HiFSA Data for Compound 6 (Case Study 6)

S7a: HiFSA Profile of Quinic Acid (6)

In CD₃OD (360 MHz, 298 K)

Proton	δ (ppm)	$\Delta\nu_{1/2}$ (Hz)	Coupling	J(Hz)
H-2a	2.0457	1.90	H-2a / H-3	3.07
H-2b	2.0298	1.13	H-2b / H-2a	-14.51
H-3	4.0830	1.34	H-2b / H-3	4.04
H-4	3.3829	1.13	H-3 / H-4	3.17
H-5	3.9896	1.21	H-5 / H-4	9.07
H-6a	1.8464	1.31	H-6a / H-6b	-13.28
H-6b	2.1190	1.73	H-6a / H-5	10.91
			H-6b / H-2b	2.83
			H-6b / H-5	4.69



S7b. Complete HiFSA Profile of Quinic Acid (6) in CD₃OD (360 MHz, 298 K) using PERCH's Parameters (.pms) Format

* NEW: the lines beginning by * are comment lines !
* To keep all the chemical shifts fixed during iteration
* replace "CHEMICAL SHIFTS(HZ)" by "..SHIFTS(HZ,fix)"
* The couplings can be fixed in the same way

```
NMR-data: [path]
#%e Date 00.00.0000; Time 00:00:00 filename
CHEMICAL SHIFTS (PPM): FIT-INDEX= 100.000
PROTON 2*SPIN=1 SPECIES=1H POPULATION= 1.0000
H-6a/1 1.846350 1*1*1 STAT=Y PRED= 1.894 RANGE= 0.318 WIDTH= 1.311 RESP= 1.0000 HSQC=C6
H-6b/1 2.118980 1*1*1 STAT=Y PRED= 2.047 RANGE= 0.332 WIDTH= 1.730 RESP= 1.0000 HSQC=C6
H-2b/1 2.029790 1*1*1 STAT=Y PRED= 1.979 RANGE= 0.307 WIDTH= 1.127 RESP= 1.0000 HSQC=C2
H-2a/1 2.045740 1*1*1 STAT=Y PRED= 2.076 RANGE= 0.232 WIDTH= 1.902 RESP= 1.0000 HSQC=C2
H-5 /1 3.989590 1*1*1 STAT=Y PRED= 3.900 RANGE= 0.501 WIDTH= 1.205 RESP= 1.0000 HSQC=C5
H-3 /1 4.083040 1*1*1 STAT=Y PRED= 4.076 RANGE= 0.235 WIDTH= 1.339 RESP= 1.0000 HSQC=C3
H-4 /1 3.382940 1*1*1 STAT=Y PRED= 3.526 RANGE= 0.298 WIDTH= 1.133 RESP= 1.0000 HSQC=C4
```

```
COUPLING CONSTANTS (HZ) :
J14_15 -13.2822 J H-6a H-6b STAT=Y PRED= -12.530 RANGE= 1.500
J14_18 10.9067 J H-6a H-5 STAT=Y PRED= 10.880 RANGE= 2.600
J15_16 2.8301 J H-6b H-2b STAT=Y PRED= 2.670 RANGE= 1.300
J15_18 4.6930 J H-6b H-5 STAT=Y PRED= 4.680 RANGE= 3.800
J16_17 -14.5071 J H-2b H-2a STAT=Y PRED= -12.630 RANGE= 1.500
J16_19 4.0377 J H-2b H-3 STAT=Y PRED= 2.800 RANGE= 2.800
J17_19 3.0704 J H-2a H-3 STAT=Y PRED= 3.030 RANGE= 3.800
J18_20 9.0735 J H-5 H-4 STAT=Y PRED= 9.330 RANGE= 3.600
J19_20 3.1721 J H-3 H-4 STAT=Y PRED= 3.320 RANGE= 3.800
```

```
CONTROL PARAMETERS:
SOLVENT = None
REFERENCE = TMS
0.00000000 = CONCENTRATION (vol%)
298.000 = TEMPERATURE (vol%, def=1.0%)
360.13490000 = FIELD(1H,MHz), used to transform shifts to ppm
0.00100000 = Minimum line-intensity
0.00100000 = Diagonalization criterium (not used)
0.00000000 = Left frequency limit (ppm)
0.00000000 = Right frequency limit (ppm)
1.127 = Default line-width (Hz)
0.0000000000 = Data-point resolution (Hz)
0.000 = Gaussian contribution%
0.000 = Dispersion contribution%
0.00000000 = Decoupling frequency (for DORES only)
```

CONSTRAINTS (in equations X0 = 1.0)...use no empty lines:

```
CARBON/DEC 2*SPIN=1 SPECIES=13C POPULATION= 1.0000 OBSERVED equal to PREDICTED
C1 /1 not observed 1*1*1 STAT=DS PRED= 78.190 RANGE= 2.789 WIDTH= 0.500 RESP= 1.0000
C6 /1 not observed 1*1*1 STAT=DS PRED= 41.450 RANGE= 1.753 WIDTH= 0.500 RESP= 1.0000
C2 /1 not observed 1*1*1 STAT=DS PRED= 40.030 RANGE= 2.928 WIDTH= 0.500 RESP= 1.0000
C5 /1 not observed 1*1*1 STAT=DS PRED= 71.380 RANGE= 6.052 WIDTH= 0.500 RESP= 1.0000
C3 /1 not observed 1*1*1 STAT=DS PRED= 72.010 RANGE= 1.893 WIDTH= 0.500 RESP= 1.0000
C4 /1 not observed 1*1*1 STAT=DS PRED= 77.680 RANGE= 9.375 WIDTH= 0.500 RESP= 1.0000
C11 /1 not observed 1*1*1 STAT=DS PRED= 178.960 RANGE= 4.835 WIDTH= 0.500 RESP= 1.0000
```

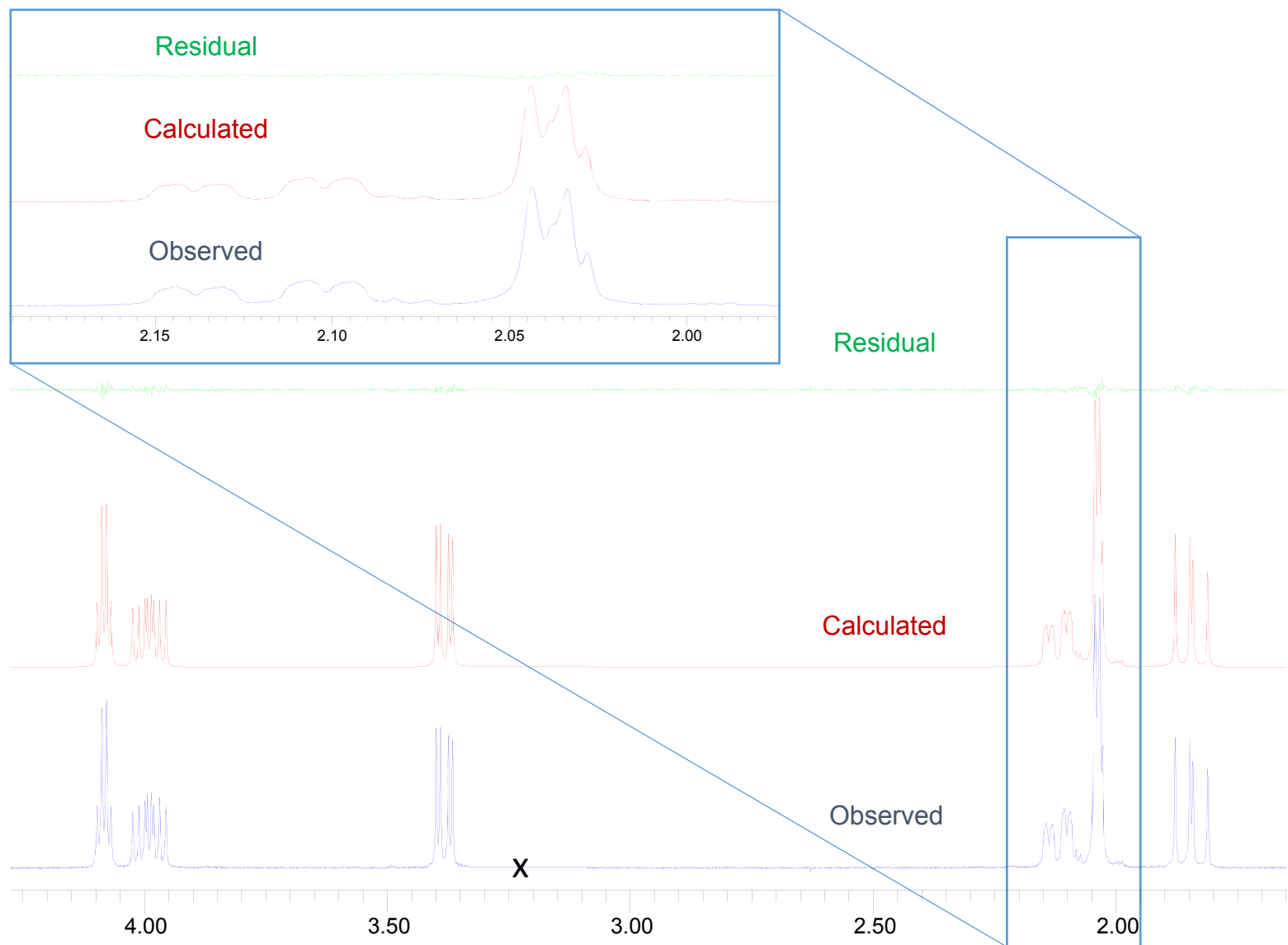
NOE CORRELATIONS() :

```
SCALAR CORRELATIONS() :
COSY 1 105.16 H-6a H-6b STAT=N PRED= 99.21 RANGE= 11.88
COSY 2 86.36 H-6a H-5 STAT=N PRED= 86.14 RANGE= 20.59
COSY 3 22.41 H-6b H-2b STAT=N PRED= 21.14 RANGE= 10.29
COSY 4 37.16 H-6b H-5 STAT=N PRED= 37.05 RANGE= 30.09
COSY 5 114.86 H-2b H-2a STAT=N PRED= 100.00 RANGE= 11.88
COSY 6 31.97 H-2b H-3 STAT=N PRED= 22.17 RANGE= 22.17
COSY 7 24.31 H-2a H-3 STAT=N PRED= 23.99 RANGE= 30.09
COSY 8 71.84 H-5 H-4 STAT=N PRED= 73.87 RANGE= 28.50
COSY 9 25.12 H-3 H-4 STAT=N PRED= 26.29 RANGE= 30.09
```

END of FILE

S7c. The ^1H NMR Fingerprint of Quinic Acid (6) in CD_3OD (360 MHz, 298 K)

The X mark on the observed spectrum indicate that solvent signal, CHD_2OD , was removed artificially in order to allow the residual to be scaled such that it reflects the differences arising from the compound and non-solvent impurities of the sample, rather than the relatively large solvent signals.

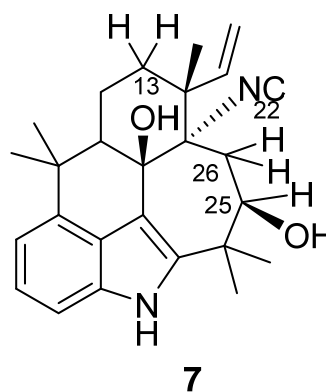


S8: HNMR and HiFSA Data for Compound 7 (Case Study 7)

S8a. HiFSA Profile of Ambiguine N Isonitrile (7)

In CD₃OD (900 MHz, 298 K)

Proton	δ (ppm)	$\Delta\nu_{1/2}$ (Hz)	Coupling	J (Hz)	Coupling	J (Hz)
H5	6.953	1.08	H5 / H6	7.28	H14B / H15	2.36
H6	7.030	1.07	H5 / H7A	0.67	H19 / H20	0.51
H7A	7.091	1.05	H6 / H7A	7.94	H20 / H21A	10.70
H13A	1.520	1.76	H13A / H13B	13.53	H20 / H21B	17.59
H13B	1.991	2.66	H13A / H14A	3.17	H21A / H21B	1.52
H14A	2.143	1.89	H13A / H14B	3.47	H25 / H26A	10.97
H14B	1.811	1.86	H13B / H14A	13.91	H25 / H26B	1.82
H15	2.273	1.58	H13B / H14B	3.49	H26A / H26B	14.13
H17	1.476	1.66	H14A / H14B	13.11	H26A / N	3.37
H18	1.452	2.42	H14A / H15	12.58	H26B / N	1.67
H19	1.581	1.74				
H20	6.183	0.83				
H21A	5.218	1.24				
H21B	5.213	1.28				
H25	3.948	1.39				
H26A	2.980	2.68				
H26B	1.967	2.30				
H27	1.265	2.51				
H28	1.264	1.92				



S8b. Complete HIFSA Profile of Ambiguine N Isonitrile (7) in CD₃OD (900 MHz, 298 K) using PERCH's Parameters (.pms) Format

NMR-data: [path]

#\$œ Date 21. 2.2014; Time 23:38: 6 perch.pms

CHEMICAL SHIFTS (PPM) :

```

PROTON      2*SPIN= 1 SPECIES=1H      POPULATION(Y)= 0.72608
H5 / 1      6.953274 1*1*1 STAT=Y PRED= 7.015 RANGE= 0.170 WIDTH(Y)= 1.087 RESP(Y)= 1.0000 HSQC= C5
H6 / 1      7.029999 1*1*1 STAT=Y PRED= 6.979 RANGE= 0.170 WIDTH(Y)= 1.066 RESP(Y)= 1.0000 HSQC= C6
H7A / 1     7.090594 1*1*1 STAT=Y PRED= 7.178 RANGE= 0.350 WIDTH(Y)= 1.046 RESP(Y)= 1.0000 HSQC= C7
H27 / 1     1.265065 1*1*3 STAT=Y PRED= 1.359 RANGE= 0.366 WIDTH(Y)= 2.508 RESP(Y)= 1.0000 HSQC= C27
H28 / 1     1.263563 1*1*3 STAT=Y PRED= 1.273 RANGE= 0.330 WIDTH(Y)= 1.916 RESP(Y)= 1.0000 HSQC= C28
H25 / 1     3.948152 1*1*1 STAT=Y PRED= 3.609 RANGE= 0.457 WIDTH(Y)= 1.390 RESP(Y)= 1.0000 HSQC= C25
H26A / 1    2.979615 1*1*1 STAT=Y PRED= 2.014 RANGE= 0.510 WIDTH(Y)= 2.678 RESP(Y)= 1.0000 HSQC= C26
H26B / 1    1.966973 1*1*1 STAT=Y PRED= 1.988 RANGE= 0.458 WIDTH(Y)= 2.300 RESP(Y)= 1.0000 HSQC= C26
H19 / 1     1.580856 1*1*3 STAT=Y PRED= 1.441 RANGE= 0.240 WIDTH(Y)= 1.736 RESP(Y)= 1.0000 HSQC= C19
H20 / 1     6.182991 1*1*1 STAT=Y PRED= 6.342 RANGE= 0.907 WIDTH(Y)= 0.826 RESP(Y)= 1.0000 HSQC= C20
H21A / 1    5.218151 1*1*1 STAT=Y PRED= 5.082 RANGE= 0.469 WIDTH(Y)= 1.244 RESP(Y)= 1.0000 HSQC= C21
H21B / 1    5.213064 1*1*1 STAT=Y PRED= 5.225 RANGE= 0.400 WIDTH(Y)= 1.280 RESP(Y)= 1.0000 HSQC= C21
H13A / 1    1.520034 1*1*1 STAT=Y PRED= 1.552 RANGE= 0.350 WIDTH(Y)= 1.764 RESP(Y)= 1.0000 HSQC= C13
H13B / 1    1.990770 1*1*1 STAT=Y PRED= 1.308 RANGE= 0.350 WIDTH(Y)= 2.662 RESP(Y)= 1.0000 HSQC= C13
H14A / 1    2.142576 1*1*1 STAT=Y PRED= 1.689 RANGE= 0.370 WIDTH(Y)= 1.887 RESP(Y)= 1.0000 HSQC= C14
H14B / 1    1.810806 1*1*1 STAT=Y PRED= 1.701 RANGE= 0.340 WIDTH(Y)= 1.862 RESP(Y)= 1.0000 HSQC= C14
H17 / 1     1.476341 1*1*3 STAT=Y PRED= 1.528 RANGE= 0.270 WIDTH(Y)= 1.661 RESP(Y)= 1.0000 HSQC= C17
H18 / 1     1.451820 1*1*3 STAT=Y PRED= 1.364 RANGE= 0.280 WIDTH(Y)= 2.419 RESP(Y)= 1.0000 HSQC= C18
H15 / 1     2.272871 1*1*1 STAT=Y PRED= 2.460 RANGE= 0.450 WIDTH(Y)= 1.584 RESP(Y)= 1.0000 HSQC= C15
14N      2*SPIN= 2 SPECIES=1H      POPULATION(Y)= 0.72608
virtN2/ 1   -20.001645 1*1*1 STAT=N WIDTH(Y)= 1.497 RESP(Y)= 1.0000 HSQC= C26
GHOST 2*SPIN= 1 SPECIES=1H      POPULATION(Y)= 0.27392
G1 / 2      1.505416 1*1*1 STAT=Y PRED= 1.505 RANGE= 0.170 WIDTH(Y)= 1.865 RESP(Y)= 1.0000
G2 / 2      1.575734 1*1*1 STAT=Y WIDTH(Y)= 2.908 RESP(Y)= 1.0000

```

COUPLING CONSTANTS (HZ) :

```

J31_32      7.2822 J H5 H6 STAT=Y PRED= 7.310 RANGE= 0.300
J31_33      0.6742 J H5 H7A STAT=Y PRED= 1.190 RANGE= 0.700
J32_33      7.9400 J H6 H7A STAT=Y PRED= 7.890 RANGE= 0.410
J40_42     10.9709 J H25 H26A STAT=Y PRED= 11.680 RANGE= 1.280
J40_43      1.8191 J H25 H26B STAT=Y PRED= 1.560 RANGE= 1.280
J42_43     14.1325 J H26A H26B STAT=Y PRED= -12.570 RANGE= 0.750
VN2        3.3696 J H26A virtN2 STAT=Y
34         1.6743 J H26B virtN2 STAT=Y
33         0.5123 J H19 H20 STAT=Y
32         10.7021 J H20 H21A STAT=Y
28         17.5941 J H20 H21B STAT=Y
24         1.5157 J H21A H21B STAT=Y
21         13.5306 J H13A H13B STAT=Y
23         3.1672 J H13A H14A STAT=Y
25         3.4737 J H13A H14B STAT=Y
26         13.9069 J H13B H14A STAT=Y
22         3.4920 J H13B H14B STAT=Y
29         13.1064 J H14A H14B STAT=Y
31         12.5775 J H14A H15 STAT=Y
30         2.4700 J H14B H15 STAT=Y
27         2.3635 J H14B H15 STAT=Y

```

CONTROL PARAMETERS:

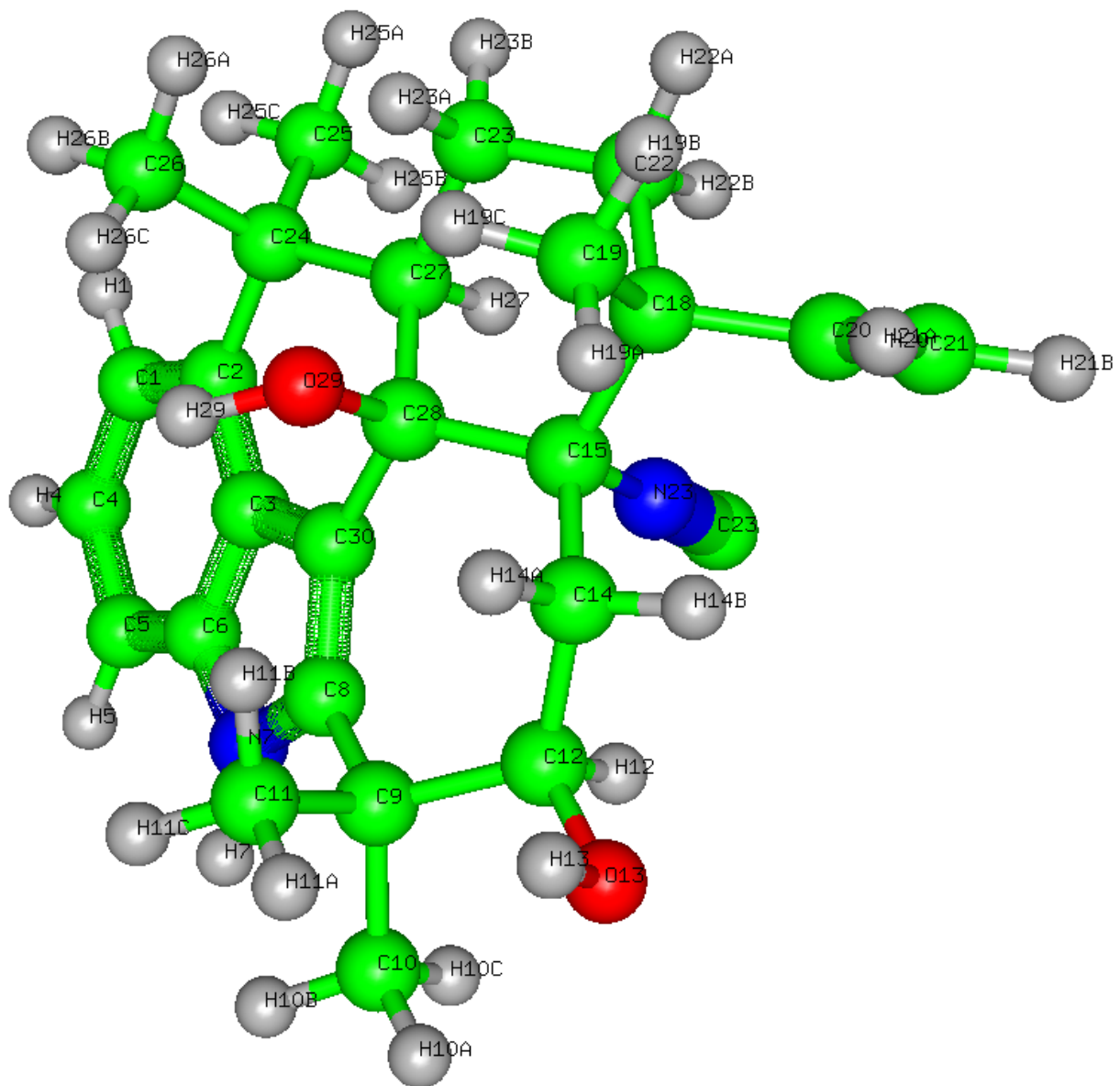
```

Solvent = none (def. 99% enriched)
1.000 = Concentration (vol%, def=1.0%)
0.00100000 = Minimum line-intensity
0.00100000 = Diagonalization criterium (not in use)
900.00000000 = FIELD(1H,MHz), used to transform shifts to ppms
9.50000040 = Left frequency (ppm)
-0.75000000 = Right frequency (ppm)
10.000 = Acquisition time (s, for QMtls)
0.000 = Line-width (for modes D, P & T, 0=use defaults)
0.041308717 = Data-point resolution (Hz)
43.149 = GAUSSIAN (% , 0=use default from INF)
-9.097 = Dispersion contribution (% , 0=use default from INF)
0.00000000 = Decoupling frequency (for DOES)

```

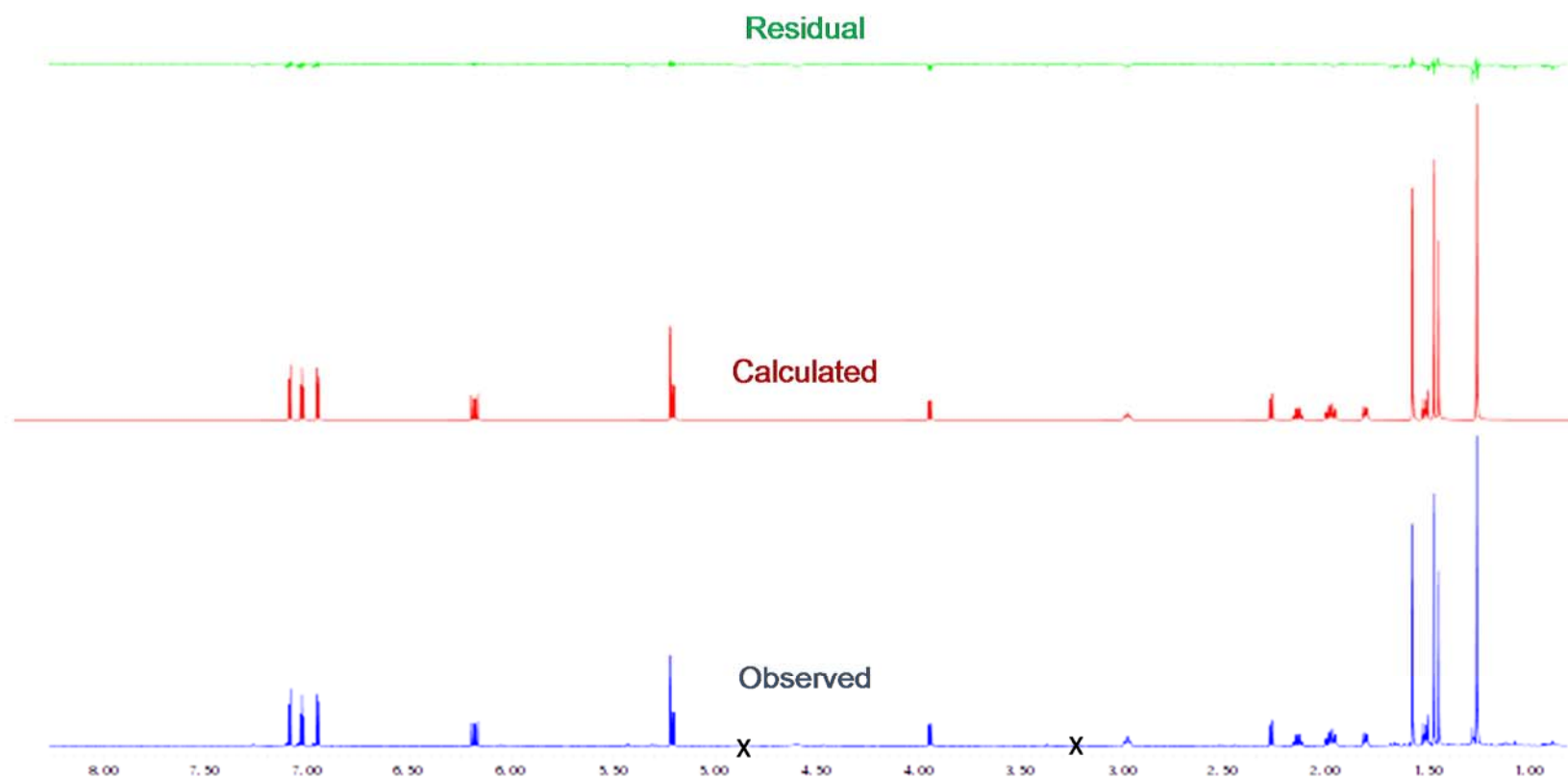
END of FILE

S8c. Optimized Structure of 7 for the Initial Prediction of Parameters in PERCH



S8d. The ^1H NMR Fingerprint of Ambiguine N Isonitrile (7) in CD_3OD (900 MHz, 298 K)

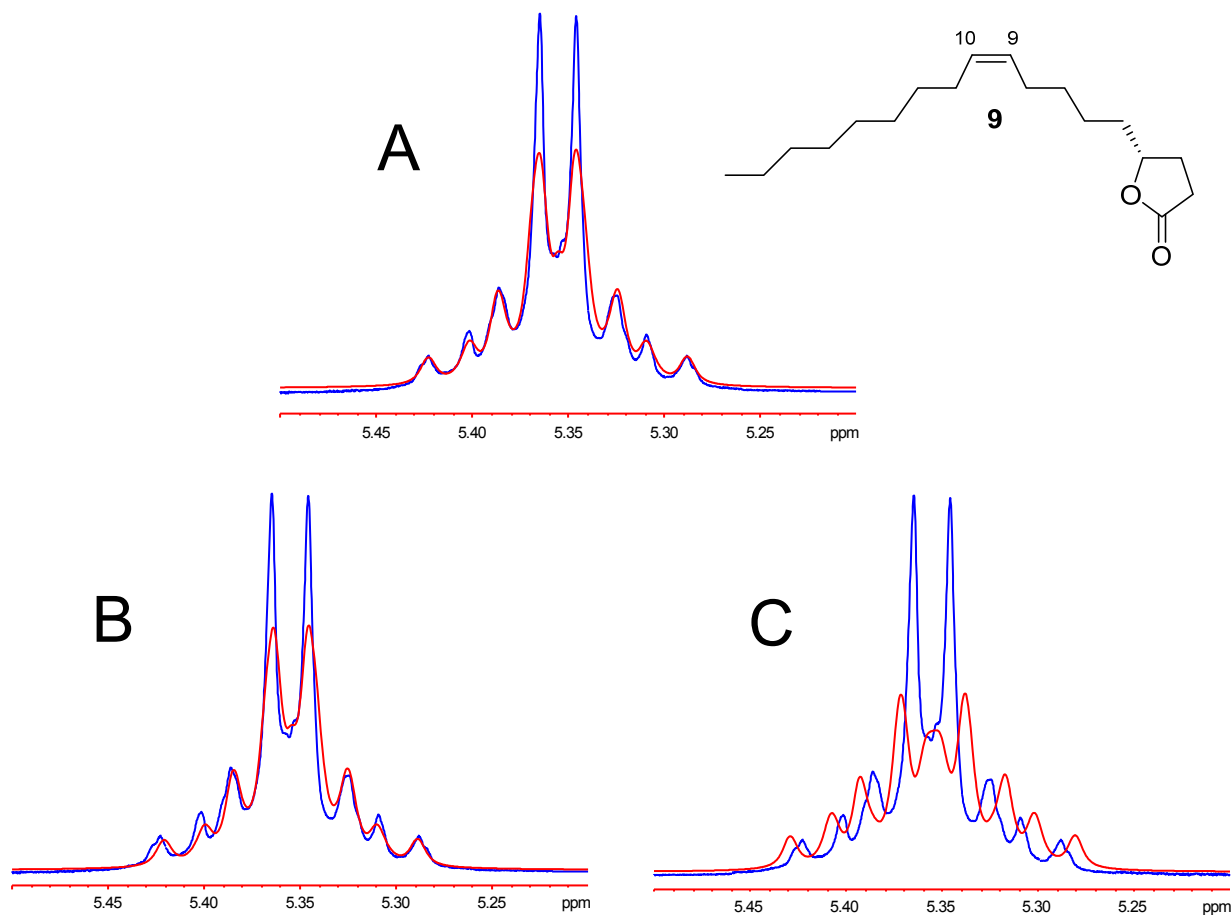
The X marks on the observed spectrum indicate that solvent signals, CHD_2OD and HOD , were removed artificially in order to allow the residual to be scaled such that it reflects the differences arising from the compound and non-solvent impurities of the sample, rather than the relatively large solvent signals.



S9: HNMR Spectral Simulation for Micromolide (9) (Case Study 9) (300 MHz, CDCl₃, 298K)

Spectral Simulation using rounded values for the iterated (A) chemical shifts of the trans olefinic protons, H-9 and H-10, of **9** demonstrate that even subtle differences in δ values produce notable effects in the simulated spectra (B and C). The rounding scenarios reflect a minimal change (1-2 ppb, B) and the maximum possible deviation (7-8 ppb, C) which would still produce the same δ value if two decimal reporting was chosen.

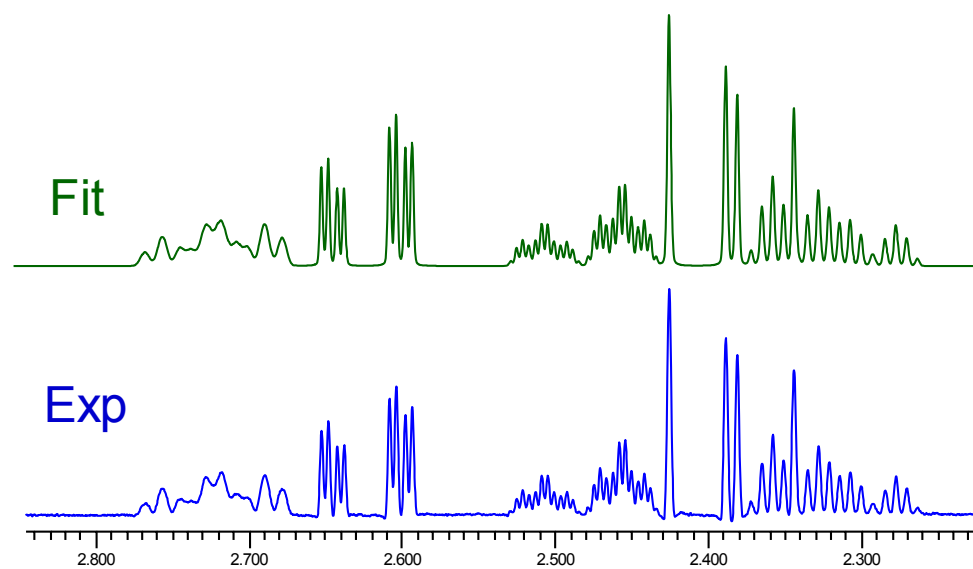
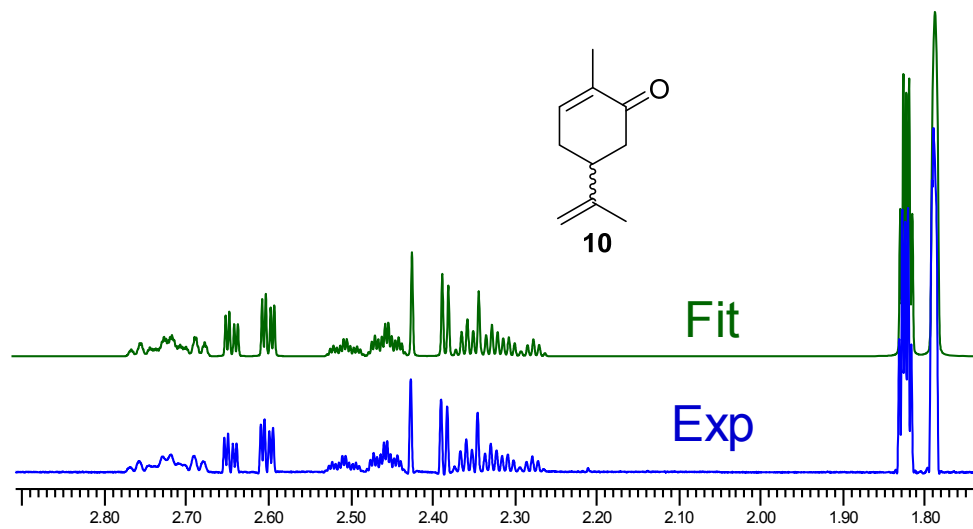
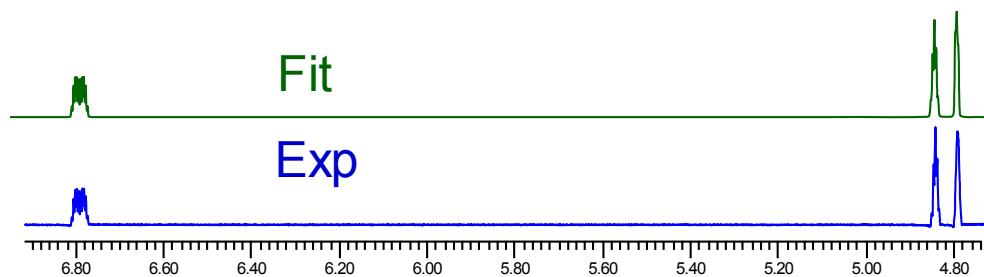
	δ (H-9)	δ (H-10)
A Iteration Result	5.377	5.333
B Sim1 (rounded $\Delta\delta$ =1-2 ppb)	5.375	5.334
C Sim2 (rounded $\Delta\delta$ =7-8 ppb)	5.384	5.325



S10. HNMR and HiFSA Data for Compound 10 (Case Study10)

10a. HiFSA Fingerprint of Carvone (10) (360 MHz, CDCl₃, 298K)

Experimental (Exp, in blue) and fitted iteration (Fit, in green).



S10b. Complete HiFSA Profile of Carvone (10) (360 MHz, CDCl₃, 298K) using PERCH's Parameters (.pms) Format

CHEMICAL SHIFTS (Hz):

```
protons  2*spin = 1
 6 /1 2446.2529 1*1 Width 0.870 Hz
5a /1 892.4883 1*1 Width 0.833 Hz
5b /1 836.5308 1*1 Width 1.009 Hz
 4 /1 980.1106 1*1 Width 1.050 Hz
3a /1 944.1661 1*1 Width 0.766 Hz
3b /1 860.1990 1*1 Width 0.897 Hz
10 /1 644.0833 1*3 Width 0.917 Hz
9b /1 1725.8264 1*1 Width 0.770 Hz
9a /1 1744.1284 1*1 Width 1.058 Hz
 7 /1 656.6725 1*3 Width 0.756 Hz
```

COUPLING CONSTANTS:

```
20 6.0233 J 6 5a
21 2.5568 J 6 5b
11 1.4505 J 6 7
23 -18.2493 J 5a 5b
24 4.4084 J 5a 4
25 1.6351 J 5a 3a
19 1.2954 J 5a 7
26 10.9149 J 5b 4
18 2.5431 J 5b 7
28 3.7618 J 4 3a
29 13.3877 J 4 3b
15 0.7804 J 4 10
30 0.9848 J 4 9b
31 -0.2144 J 4 9a
32 -16.0705 J 3a 3b
16 0.3376 J 10 9b
17 1.4145 J 10 9a
33 1.6579 J 9b 9a
```

CONTROL PARAMETERS:

```
0.0010 = The minimum intensity of a line
0.0010 = Diagonalization criterium (not in use !)
3350.347 = Left frequency limit
-364.127 = Right frequency
0.0000 = Line-width (for modes D, P & T)
0.000000 = Data-point resolution (for mode P)
0.0 = GAUSSIAN %
0.00 = Dispersion contribution %
0.0000 = Decoupling frequency (for DORES)
```


S11: Raw NMR Data & Software-based NMR Analysis

Raw NMR Data. The evaluation of NMR spectra is no longer limited by on-line processing requirements or instrument access, nor is the output format restricted to paper hardcopies. Storing NMR spectral data in digital form is common practice today. In addition to various proprietary data formats from instrument and software vendors, there also have been attempts to standardize the storage of the actual digital spectrum and related information.⁴¹ However, currently there is no standardized open-access format for FID data, regardless of dimension (n-D), in particular not for raw FID data in which the quadrature and/or phase detection schemes are documented and preserved. Analogous to digital photography, digital FID data represents "RAW" file formats, many of which are not fully documented and require reverse engineering approaches in order to be readable. Until appropriate open formats have been defined, the instrument-centered "RAW FID" formats provided by the instrument manufacturers can be widely regarded as the most future proof form of storage. However, adequate precision in HNMR reporting is already achievable with widely available NMR processing software and is a notable means of overcoming this archival limitation, at least for interpreted HNMR spectra. At the same time, the enormous value of FID data to peer review and readership needs to be emphasized.

NMR Software. Tools for off-line post-acquisition processing including Free software (i.e., software source code is available; various sharing models) and other free-of-charge solutions suitable for standard personal computers are widely available (see http://nmrwiki.org/wiki/index.php?title=NMR_Software for a compilation). These tools provide numerous functions for the spectroscopic evaluation, support for the interpretation of NMR spectra, and the extraction of ¹H NMR parameters with adequate precision. In some instances, software supports multiplet analysis, extraction of δ/J values, and adaptation to optional journal output formats. Notably, these offerings typically utilize line fitting mechanisms which make first order assumptions and, therefore, are unable to capture the full characteristics of overlapping signals and the widely observed higher order HNMR spectra. It is also noteworthy, that software-

based evaluation of NMR spectra is a routine practice in qHNMR, which adds quantitation as another valuable dimension to the routine ^1H NMR analysis of natural products (see ^{1,2} and references therein). Overall, the determination of chemical shift (δ) values for ^1H resonances with an adequate number of decimal places requires no, or only little additional effort when using contemporary NMR analysis software. Providing the original experimental data (raw FID), in addition to the tabulation of the extracted NMR information is an effective way of increasing the reproducibility of HNMR spectra even further and enables others to fully reproduce the spectral analysis.

References

- (1) Pauli, G. F.; Jaki, B. U.; Gödecke, T.; Lankin, D. C. *J. Nat. Prod.* **2012**, *75*, 834-851.
- (2) Pauli, G. F.; Jaki, B. U.; Lankin, D. C. *J. Nat. Prod.* **2005**, *68*, 133-149.