### **Supporting Information**

for

# Synthesis of zearalenone-16-β,D-glucoside and zearalenone-16sulfate: A tale of protecting resorcylic acid lactones for regiocontrolled conjugation

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# Experimental details (including remarks and general procedures), characterization data, copies of NMR spectra of new compounds, 2D NMR spectra of glucoside 7.

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#### 1. Experimental

#### **General remarks**

All reactions were performed under an argon atmosphere. The progress of reactions was monitored by thin-layer chromatography (TLC) over silica gel 60 F254 (Merck). The chromatograms were visualized by irradiation with ultraviolet light or by heat staining with ceric ammonium molybdate in ethanol/sulfuric acid. LC-ESI-MS/MS was performed on an HCT ion trap mass spectrometer (Bruker, Germany) in full scan mode. Chromatographic separation was done on a 1200 series HPLC system (Agilent Technologies, Germany) using a Luna RP-C18 column ( $3.0 \times 150$  mm, 3 µm particle size, Phenomenex, Germany) and application of pure substances was achieved using a TLC–MS interface (Camag, Germany). Preparative column chromatography was performed on silica gel 60 (Merck, 40-63 µm) or RP-C18 silica gel (Merck, 40-63 µm) using a Büchi Sepacore<sup>TM</sup> Flash System. NMR spectra were recorded on a Bruker DPX-200 MHz or Avance DRX-400 MHz spectrometer. Data were recorded and evaluated using TOPSPIN 1.3 (Bruker Biospin). All chemical shifts are given in ppm relative to tetramethylsilane. The calibration was done using residual solvent signals. Multiplicities are abbreviated as s (singlet), d (doublet), t (triplet), q (quartet), b (broad signal). Zearalenone was obtained from Fermentek (Israel) and all other chemicals were purchased from ABCR (Germany) or Sigma-Aldrich (Austria/Germany).

#### General procedure A: regioselective acetylation of RAL type compounds

To a solution of the RAL type compound (1.0 mmol) and DMAP (1.2 mg, 10  $\mu$ mol) in dry toluene (5 mL) at 0 °C was slowly added a solution of Ac<sub>2</sub>O (110 mg, 1.1 mmol) in dry toluene (1 mL). The reaction mixture was gradually warmed to room temperature and stirred for further 16 h. The solvent was removed under reduced pressure and the residue was purified by column chromatography (hexanes/EtOAc gradient elution) to yield the desired 4'-O-acetylated product.

#### 4-Acetoxy-2-hydroxybenzoic acid isopropyl ester (10)

Following general procedure A, **10** was obtained as a white solid (203 mg, 85%); R<sub>f</sub> 0.4 (hexanes/EtOAc, 9/1); <sup>1</sup>H NMR (200 MHz, CDCl<sub>3</sub>)  $\delta$  11.07 (s, 1H, 2-OH), 7.84 (d, *J* = 8.8 Hz, 1H, H-6), 6.72 (d, *J* = 2.3 Hz, 1H, H-3), 6.62 (dd, *J* = 8.8, 2.3 Hz, 1H, H-5), 5.26 (sept., *J* = 6.2 Hz, 1H, COOCH), 2.27 (s, 3H, CH<sub>3</sub>COO), 1.36 (d, *J* = 6.2 Hz, 6H, (CH<sub>3</sub>)<sub>2</sub>CH); <sup>13</sup>C NMR (50 MHz, CDCl<sub>3</sub>)  $\delta$  169.2 (s), 168.5 (s), 162.9 (s), 156.1 (s), 131.0 (d), 112.9 (d), 110.7 (s), 110.4 (d), 69.3 (d), 21.8 (q), 21.1 (q); HRMS *m*/*z* calcd for C<sub>12</sub>H<sub>14</sub>NaO<sub>5</sub><sup>+</sup> [M+Na]<sup>+</sup> 261.0733, found 261.0750.

#### 14-O-Acetylzearalenone (14)

Following general procedure A (reduced scale), **14** was obtained starting from ZEN (**1**, 32 mg, 0.1 mmol) as a white solid (27 mg, 75%); R<sub>f</sub> 0.4 (hexanes/EtOAc, 2/1); <sup>1</sup>H NMR (200 MHz, CDCl<sub>3</sub>)  $\delta$  11.88 (s, 1H, 16-OH), 7.00 (d, J = 15.6 Hz, 1H, H-12), 6.67-6.62 (m, 2H, H-13, H-15), 5.71 (ddd, J = 15.4, 10.1, 4.3 Hz, 1H, H-11), 5.11-4.93 (m, 1H, H-3), 2.81 (ddd, J = 18.7, 12.1, 2.5 Hz, 1H, H<sub>a</sub>-8), 2.59 (m, 1H, H<sub>a</sub>-6), 2.37-2.05 (m, 5H, H<sub>a</sub>-5, H<sub>b</sub>-6, H<sub>b</sub>-8, H<sub>ab</sub>-10), 2.29 (s, 3H, CH<sub>3</sub>COO), 1.85-1.54 (m, 5H, H<sub>a</sub>-6), 2.37-2.05 (m, 5H, H<sub>a</sub>-5, H<sub>b</sub>-6, H<sub>b</sub>-8, H<sub>ab</sub>-10), 2.29 (s, 3H, CH<sub>3</sub>COO), 1.85-1.54 (m, 5H, H<sub>a</sub>b-4, H<sub>b</sub>-5, H<sub>ab</sub>-9), 1.39 (d, J = 6.1 Hz, 3H, CH<sub>3</sub>); <sup>13</sup>C NMR (50 MHz, CDCl<sub>3</sub>)  $\delta$  211.0 (s, C-7), 171.1 (s, CH<sub>3</sub>COO), 168.8 (s, C-1), 164.6 (s, C-14), 154.9 (s, C-16), 143.4 (s, C-12a), 133.5 (d, C-11), 132.6 (d, C-12), 113.7 (d, C-13), 109.6 (d, C-15), 108.3 (s, C-16a), 74.1 (d, C-3), 43.0 (t, C-6), 36.7 (t, C-8), 34.7 (t, C-4), 31.1 (t, C-10), 22.2 (t, C-5), 21.2 (q, CH<sub>3</sub>COO), 21.0 (t, C-9), 20.8 (q, CH<sub>3</sub>); NMR data matched that reported previously by Munoz et al.<sup>1</sup> HRMS *m/z* calcd for C<sub>20</sub>H<sub>24</sub>NaO<sub>6</sub><sup>+</sup> [M+Na]<sup>+</sup> 383.1465, found 383.1466.

#### General procedure B: regioselective *p*-methoxybenzylation of RAL type compounds

To a solution of the RAL type compound (1 mmol) and  $Cs_2CO_3$  (433 mg, 1.5 mmol) in dry DMF (5 mL) was slowly added a solution of *p*-methoxybenzyl chloride (PMB-Cl, 172 mg, 1.1 mmol) in dry DMF (1 mL). The reaction mixture was stirred at room temperature for 24 h, diluted with water (20 mL) and extracted with EtOAc (3 × 15 mL). The combined organic layer was washed with brine,

<sup>&</sup>lt;sup>1</sup> Munoz, L.; Castro, J.L.; Cardelle, M.; Castedo, L.; Riguera, R. *Phytochemistry* **1989**, *28*, 83-85.

dried over Na<sub>2</sub>SO<sub>4</sub> and concentrated. Column chromatography (hexane/EtOAc gradient elution) afforded the desired 4'-O-PMB protected compound.

#### 2-Hydroxy-4-((p-methoxybenzyl)oxy)benzoic acid isopropyl ester (15)

Following general procedure B, **15** was obtained as a white solid (256 mg, 81%); R<sub>f</sub> 0.8 (hexanes/EtOAc, 3/1); <sup>1</sup>H NMR (200 MHz, CDCl<sub>3</sub>)  $\delta$  11.07 (s, 1H, 2-OH), 7.66 (d, *J* = 8.5 Hz, 1H, H-6), 7.26 (d, *J* = 8.5 Hz, 2H, PMB), 6.84 (d, *J* = 8.5 Hz, 2H, PMB), 6.46-6.36 (m, 2H, H-3, H-5), 5.17 (sept., *J* = 6.5 Hz, 1H, COOCH), 4.92 (s, 2H, OCH<sub>2</sub>), 3.74 (s, 3H, OCH<sub>3</sub>), 1.29 (d, *J* = 6.4 Hz, 6H, (CH<sub>3</sub>)<sub>2</sub>CH); <sup>13</sup>C NMR (50 MHz, CDCl<sub>3</sub>)  $\delta$  169.6 (s), 164.6 (s), 163.8 (s), 159.6 (s), 131.2 (d), 129.3 (d), 128.1 (s), 114.1 (d), 108.0 (d), 106.1 (s), 101.6 (d), 69.9 (t), 68.7 (q), 55.3 (q), 21.9 (q); HRMS *m/z* calcd for C<sub>18</sub>H<sub>20</sub>NaO<sub>5</sub><sup>+</sup> [M+Na]<sup>+</sup> 339.1203, found 339.1195.

#### 14-O-(p-Methoxybenzyl)zearalenone (18)

Following general procedure B (reduced scale), **18** was obtained starting from ZEN (**1**, 32 mg, 0.1 mmol) as a white solid (31 mg, 71%);  $R_f$  0.6 (hexanes/EtOAc, 2/1); <sup>1</sup>H NMR (200 MHz, CDCl<sub>3</sub>)  $\delta$  12.01 (s, 1H, 16-OH), 7.27 (d, J = 8.6 Hz, 2H, PMB), 6.94 (d, J = 14.9 Hz, 1H, H-12), 6.84 (d, J = 8.8 Hz, 2H, PMB), 6.46 (d, J = 2.6 Hz, 1H, H-13), 6.39 (d, J = 2.6 Hz, 1H, H-15), 5.60 (ddd, J = 15.5, 10.3, 4.0 Hz, 1H, H-11), 5.03-4.83 (m, 3H, H-3, OCH<sub>2</sub>), 3.74 (s, 3H, OCH<sub>3</sub>), 2.77 (ddd, J = 18.2, 12.2, 2.9 Hz, 1H, H<sub>a</sub>-8), 2.60-2.45 (m, 1H, H<sub>a</sub>-6), 2.37-1.96 (m, 5H, H<sub>a</sub>-5, H<sub>b</sub>-6, H<sub>b</sub>-8, H<sub>ab</sub>-10), 1.78-1.45 (m, 5H, H<sub>a</sub>-4, H<sub>b</sub>-5, H<sub>ab</sub>-9), 1.31 (d, J = 6.1 Hz, 3H, CH<sub>3</sub>); <sup>13</sup>C NMR (50 MHz, CDCl<sub>3</sub>)  $\delta$  211.0 (s, C-7), 171.4 (s, C-1), 165.6 (s, C-14), 163.2 (s, C-16), 159.6 (s, Ar-OCH<sub>3</sub>), 143.3 (s, C-12a), 133.3 (d, C-11), 132.4 (d, C-12), 129.3 (d, PMB), 128.1 (s, PMB), 114.0 (d, PMB), 108.9 (d, C-13), 103.7 (s, C-16a), 100.8 (d, C-15), 73.4 (d, C-3), 69.9 (t, OCH<sub>2</sub>), 55.4 (q, OCH<sub>3</sub>), 42.9 (t, C-6), 36.6 (t, C-8), 34.7 (t, C-4), 31.0 (t, C-10), 22.3 (t, C-5), 21.0 (t, C-9), 20.9 (q, CH<sub>3</sub>); HRMS *m/z* calcd for  $C_{26}H_{30}NaO_6^+$  [M+Na]<sup>+</sup> 461.1935, found 461.1957.

#### General procedure C: regioselective TIPS-protection of RAL type compounds

To a solution of the RAL type compound (1 mmol) and imidazole (156 mg, 2.3 mmol) in dry  $CH_2Cl_2$  (10 mL) was slowly added TIPS-Cl (216 mg, 1.1 mmol). The reaction mixture was stirred at room temperature for 16 h, diluted with  $CH_2Cl_2$  (20 mL), washed with brine, dried over Na<sub>2</sub>SO<sub>4</sub> and concentrated. The residue was purified by column chromatography (hexanes/EtOAc gradient elution).

#### 2-Hydroxy-4-(triisopropylsilyloxy)benzoic acid isopropyl ester (21)

Following general procedure C, **21** was obtained as a colorless viscous oil (350 mg, 99%);  $R_f 0.9$  (hexanes/EtOAc, 12/1); <sup>1</sup>H NMR (200 MHz, CDCl<sub>3</sub>)  $\delta$  11.0 (s, 1H, 2-OH), 7.70 (d, J = 8.7 Hz, 1H, H-6), 6.44 (d, J = 2.3 Hz, 1H, H-3), 6.39 (dd, J = 8.6, 2.3 Hz, 1H, H-5), 5.24 (sept., J = 6.2 Hz, 1H, COOCH), 1.35 (d, J = 6.3 Hz, 6H, (CH<sub>3</sub>)<sub>2</sub>CH), 1.31-1.16 (m, 3H, SiCH), 1.12-1.04 (m, 18H, SiCH(CH<sub>3</sub>)<sub>2</sub>); <sup>13</sup>C NMR (50 MHz, CDCl<sub>3</sub>)  $\delta$  169.6 (s), 163.5 (s), 162.5 (s), 131.2 (d), 112.1 (d), 107.5 (d), 106.6 (s), 68.6 (d), 21.9 (q), 17.8 (q), 12.9 (d); HRMS *m/z* calcd for C<sub>19</sub>H<sub>32</sub>NaO<sub>4</sub>Si<sup>+</sup> [M+Na]<sup>+</sup> 375.1962, found 375.1960.

#### 14-O-(Triisopropylsilyl)zearalenone (22)

Following general procedure C (reduced scale), **22** was obtained starting from ZEN (**1**, 100 mg, 0.31 mmol) as a white solid (145 mg, 99%); R<sub>f</sub> 0.8 (hexanes/EtOAc, 2/1); <sup>1</sup>H NMR (200 MHz, CDCl<sub>3</sub>)  $\delta$  11.85 (s, 1H, 16-OH), 6.90 (d, J = 15.2 Hz, 1H, H-12), 6.30 (d, J = 2.5 Hz, 1H, H-13), 6.25 (d, J = 2.5 Hz, 1H, H-15), 5.51 (ddd, J = 15.3, 10.2, 3.8 Hz, 1H, H-11), 4.97-4.79 (m, 1H, H-3), 2.75 (ddd, J = 18.4, 12.2, 3.0 Hz, 1H, H<sub>a</sub>-8), 2.55-2.41 (m, 1H, H<sub>a</sub>-6), 2.34-1.93 (m, 5H, H<sub>a</sub>-5, H<sub>b</sub>-6, H<sub>b</sub>-8, H<sub>a</sub>b-10), 1.76-1.35 (m, 5H, H<sub>a</sub>b-4, H<sub>b</sub>-5, H<sub>a</sub>b-9), 1.26 (d, J = 6.1 Hz, 3H, CH<sub>3</sub>), 1.21-1.09 (m, 3H, SiCH), 1.02-0.93 (m, 18H, SiCH(CH<sub>3</sub>)<sub>2</sub>); 13C NMR (50 MHz, CDCl<sub>3</sub>)  $\delta$  211.1 (s, C-7), 171.4 (s, C-1), 165.3 (s, C-16), 161.2 (s, C-14), 143.4 (s, C-12a), 133.4 (d, C-11), 132.1 (d, C-12), 113.0 (s, C-13), 106.7 (d, C-15), 104.1 (d, C-16a), 73.3 (d, C-3), 43.0 (t, C-6), 36.7 (t, C-8), 34.8 (t, C-6), 31.1 (t, C-16), 200.12 (t, C-16),

C-10), 22.3 (t, C-5), 21.0 (t, C-9), 20.9 (q, CH<sub>3</sub>), 17.9 (q, SiCH( $CH_3$ )<sub>2</sub>), 12.7 (d, SiCH); HRMS m/z calcd for C<sub>27</sub>H<sub>42</sub>NaO<sub>5</sub>Si<sup>+</sup> [M+Na]<sup>+</sup> 497.6294, found 497.6299.

#### General procedure D: Königs-Knorr glucosylation

To a solution of the glucosyl acceptor (0.5 mmol) and bromoacetoglucose **13** (617 mg, 1.5 mmol) in dry MeCN (10 mL) was added molecular sieve 3 Å (1 g) and the resulting suspension was stirred at room temperature for 1 h. After addition of Ag<sub>2</sub>O (174 mg, 0.75 mmol), the reaction mixture was stirred in the dark for 48 h. Since TLC analysis indicated remaining glucosyl acceptor, **13** (411 mg, 1 mmol) and Ag<sub>2</sub>O (116 mg, 0.5 mmol) were added and stirring was continued in the dark for 24 h. The reaction mixture was diluted with CH<sub>2</sub>Cl<sub>2</sub>, filtered through Celite and concentrated under reduced pressure. Silica gel filtration (hexane/EtOAc gradient elution) was performed to remove most the carbohydrate byproducts. Since we were not able to fully purify the crude glucosylated products, these compounds were directly subjected to deprotection within subsequent steps.

#### 5-Hydroxy-2-(isopropoxycarbonyl)phenyl-β,D-glucoside (17)

Following general procedure D, crude **16** was obtained starting from the PMB protected ZEN mimic **15** (95 mg, 0.3 mmol). This material was dissolved in  $CH_2Cl_2/H_2O$  (10/1, 6 mL) and cooled to 0 °C. After addition of DDQ (102 mg, 0.45 mmol) the reaction mixture was gradually warmed to room temperature. TLC/MS analysis indicated full conversion after stirring for 24 h. The reaction mixture was diluted with  $CH_2Cl_2$  (20 mL), washed with satd. aq. NaHCO<sub>3</sub>, dried over Na<sub>2</sub>SO<sub>4</sub> and concentrated. The crude product was dissolved in THF/H<sub>2</sub>O (4/1, 7 mL) and KOH (168 mg, 3 mmol) was added. After stirring for 2 h at room temperature, the solution was diluted with water (15 mL) and acidified to pH 6.5 by slow addition of 0.2 M aq. HCl followed by extraction with EtOAc (4 × 15 mL). The combined organic layer was dried over Na<sub>2</sub>SO<sub>4</sub> and concentrated under reduced pressure. The residue was purified by column chromatography (MPLC, RP-C18, MeCN/H<sub>2</sub>O, gradient elution) to yield the desired product **17** as a white solid (32 mg, 30%); R<sub>f</sub> 0.5 (CH<sub>2</sub>Cl<sub>2</sub>/MeOH, 5/1); <sup>1</sup>H NMR

(200 MHz, CD<sub>3</sub>OD)  $\delta$  7.68 (d, J = 8.8 Hz, 1H, H-6), 6.80 (d, J = 2.3 Hz, 1H, H-3), 6.52 (dd, J = 8.8, 2.3 Hz, 1H, H-5), 5.16 (sept., J = 6.3 Hz, 1H, COOCH), 4.82 (d, J = 7.2 Hz, 1H, H-1'), 3.94 (dd, J = 11.8, 1.5 Hz, 1H, H<sub>a</sub>-6'), 3.80-3.68 (m, 1H, H<sub>b</sub>-6'), 3.56-3.39 (m, 4H, H-2', H-3', H-4', H-5'), 1.34 (d, J = 6.3 Hz, 6H, (CH<sub>3</sub>)<sub>2</sub>CH); <sup>13</sup>C NMR (50 MHz, CD<sub>3</sub>OD)  $\delta$  167.3 (s), 164.4 (s), 161.4 (s), 134.1 (d), 113.2 (s), 111.0 (d), 105.9 (d), 104.3 (d), 78.5 (d), 77.5 (d), 74.9 (d), 71.2 (d), 69.4 (d), 62.5 (t), 22.2 (q), 22.1 (q); HRMS *m/z* calcd for C<sub>16</sub>H<sub>22</sub>NaO<sub>9</sub><sup>+</sup> [M+Na]<sup>+</sup> 381.1156, found 381.1145.

#### Zearalenone-16-β,D-glucoside (7)

Following general procedure D, crude protected ZEN-16-glucoside was obtained starting from **22** (47 mg, 0.1 mmol). This material was dissolved in dry THF (2 mL) and the resulting solution was cooled to -10 °C. After addition of AcOH (6 mg, 0.1 mmol) and TBAF (0.1 mL, 0.1 mmol, 1 M in THF), the reaction was stirred at -10 °C for 3 h. TLC/MS analysis indicated full conversion of the starting material to the desired product. The solvent was evaporated and the residue was dissolved in THF/H<sub>2</sub>O (4/1, 6 mL). KOH (56 mg, 1 mmol) was added and the reaction mixture was stirred at room temperature for 2 h. The solution was diluted with water (15 mL) and acidified to pH 6.5 by slow addition of 0.2 M aq. HCl followed by extraction with EtOAc (4 × 15 mL). The combined organic layer was dried over Na<sub>2</sub>SO<sub>4</sub> and concentrated under reduced pressure. The residue was purified by column chromatography (MPLC, RP-C18, MeCN/H<sub>2</sub>O, gradient elution) to yield the desired product **7** as a white solid (16 mg, 34%, 3 steps); R<sub>f</sub> 0.4 (CH<sub>2</sub>Cl<sub>2</sub>/MeOH, 6/1); <sup>1</sup>H and <sup>13</sup>C NMR data matched that reported<sup>2</sup> (for additional 2D NMR spectra see Figures S1, S2 and S3); HRMS *m/z* calcd for C<sub>24</sub>H<sub>32</sub>NaO<sub>10</sub><sup>+</sup> [M+Na]<sup>+</sup> 503.1888, found 503.1905.

#### 16-O-((2,2,2,-Trichloroethoxy)sulfonyl)-14-O-(triisopropylsilyl)zearalenone (24)

To a solution of **22** (47.5 mg, 0.1 mmol) in dry  $CH_2Cl_2$  (2 mL) cooled to 0 °C was added 1,2dimethylimidazole (28 mg, 0.5 mmol) and **23** (184 mg, 0.4 mmol). The reaction mixture was warmed

<sup>&</sup>lt;sup>2</sup> Kovalsky Paris, M. P.; Schweiger, W.; Hametner, C.; Stückler, R.; Muehlbauer, G. J.; Varga, E.; Krska, R.; Berthiller,

F.; Adam, G. J. Agric. Food Chem. 2014, 62, 1181-1189.

to room temperature and stirred overnight. After dilution with CH<sub>2</sub>Cl<sub>2</sub> (10 mL), the solution was washed with water, dried over Na<sub>2</sub>SO<sub>4</sub> and concentrated. Column chromatography over silica gel (hexanes/EtOAc gradient elution) gave the desired product **24** as a colorless viscous oil (62 mg, 91%); R<sub>f</sub> 0.4 (hexanes/EtOAc, 5/1); <sup>1</sup>H NMR (200 MHz, CDCl<sub>3</sub>)  $\delta$  6.96 (d, *J* = 2.2 Hz, 1H, H-13), 6.92 (d, *J* = 2.2 Hz, 1H, H-15), 6.45 (d, *J* = 15.6 Hz, 1H, H-12), 5.93 (ddd, *J* = 15.5, 9.5, 4.7 Hz, 1H, H-11), 5.42-5.25 (m, 1H, H-3), 4.92 (d, *J* = 11.0 Hz, 1H, CH<sub>2a</sub>), 4.84 (d, *J* = 11.0 Hz, 1H, CH<sub>2b</sub>), 2.65 (ddd, *J* = 17.6, 10.9, 3.2 Hz, 1H, H<sub>a</sub>-8), 2.45-2.18 (m, 4H, H<sub>ab</sub>-6, H<sub>b</sub>-8, H<sub>a</sub>-10), 2.15-1.93 (m, 2H, H<sub>a</sub>-5, H<sub>b</sub>-10), 1.80-1.51 (m, 5H, H<sub>ab</sub>-4, H<sub>b</sub>-5, H<sub>ab</sub>-9), 1.37 (d, *J* = 6.3 Hz, 3H, CH<sub>3</sub>), 1.31-1.18 (m, 3H, SiCH), 1.14-1.05 (m, 18H, SiCH(CH<sub>3</sub>)<sub>2</sub>); <sup>13</sup>C NMR (50 MHz, CDCl<sub>3</sub>)  $\delta$  211.1 (s, C-7), 165.2 (s, C-1), 158.0 (s, C-16), 147.8 (s, C-14), 138.7 (s, C-12a), 134.8 (d, C-11), 128.4 (d, C-12), 118.6 (s, C-16a), 116.1 (d, C-13), 110.8 (d, C-15), 92.4 (s, CCl<sub>3</sub>), 80.6 (t, OCH<sub>2</sub>), 72.2 (d, C-3), 43.7 (t, C-6), 37.6 (t, C-8), 35.1 (t, C-4), 31.4 (t, C-10), 21.5 (t, C-5), 21.2 (t, C-9), 19.8 (q, CH<sub>3</sub>), 17.9 (q, SiCH(CH<sub>3</sub>)<sub>2</sub>), 12.60 (d, SiCH); HRMS *m*/z calcd for C<sub>29</sub>H<sub>43</sub>Cl<sub>3</sub>NaO<sub>8</sub>SSi<sup>+</sup> [M+Na]<sup>+</sup> 707.1406, found 707.1433.

#### Zearalenone-16-sulfate, tetrabutylammonium salt (NBu<sub>4</sub>-8)

To a solution of the **24** (55 mg, 0.08 mmol) in dry THF (1 mL), cooled to -10 °C, was added AcOH (5 mg, 0.083 mmol) and TBAF (83 µL, 0.083 mmol, 1 M in THF). The reaction mixture was stirred at -10 °C for 3 h and subsequently concentrated under reduced pressure. The residue was dissolved in dry MeOH (1 mL) and after addition of HCOONH<sub>4</sub> (44 mg, 0.7 mmol) and zinc dust (16 mg, 0.25 mmol), the reaction mixture was stirred for 16 h at room temperature. Filtration through Celite and evaporation of the solvent afforded the crude product, which was purified by column chromatography over silica gel (CH<sub>2</sub>Cl<sub>2</sub>/MeOH/NH<sub>4</sub>OH = 10/4/1). **8** was obtained as tetrabutylammonium salt starting from **24** as a white solid (33 mg, 65%); R<sub>f</sub> 0.5 (CH<sub>2</sub>Cl<sub>2</sub>/MeOH/NH<sub>4</sub>OH, 20/5/1); <sup>1</sup>H NMR (400 MHz, CD<sub>3</sub>OD)  $\delta$  7.01 (d, *J* = 1.9 Hz, 1H, Ar-H), 6.75 (d, *J* = 1.9 Hz, 1H, Ar-H), 6.39 (d, *J* = 15.7 Hz, 1H, H-12), 6.01 (ddd, *J* = 15.7, 10.0, 4.3 Hz,

1H, H-11), 5.33-5.24 (m, 1H, H-3), 3.26-3.19 (m, 8H, N<sup>+</sup>CH<sub>2</sub>), 2.72 (ddd, J = 17.1, 11.1, 3.1 Hz, 1H, H<sub>a</sub>-8), 2.50 (ddd, J = 13.2, 8.5, 4.5 Hz, 1H, H<sub>a</sub>-6), 2.37-2.21 (m, 3H, H<sub>b</sub>-6, H<sub>b</sub>-8, H<sub>a</sub>-10), 2.10-1.97 (m, 2H, H<sub>a</sub>-5, H<sub>b</sub>-10), 1.85-1.70 (m, 2H, H<sub>a</sub>-4, H<sub>a</sub>-9), 1.69-1.54 (m, 11H, N<sup>+</sup>CH<sub>2</sub>CH<sub>2</sub>, H<sub>b</sub>-4, H<sub>b</sub>-5, H<sub>b</sub>-9), 1.47-1.35 (m, 11H, N<sup>+</sup>CH<sub>2</sub>CH<sub>2</sub>CH<sub>2</sub>, CH<sub>3</sub>), 1.02 (t, J = 7.4 Hz, 12H, N<sup>+</sup>CH<sub>2</sub>CH<sub>2</sub>CH<sub>2</sub>CH<sub>2</sub>CH<sub>3</sub>); <sup>13</sup>C NMR (100 MHz, CD<sub>3</sub>OD)  $\delta$  212.7 (s, C-7), 167.6 (s, C-1), 159.0 (s, C-14), 150.8 (s, C-16), 136.7 (s, C-12a), 132.7 (d, C-11), 128.7 (d, C-12), 117.8 (s, C-16a), 107.5 (d, C-13), 106.8 (d, C-15), 71.2 (d, C-3), 58.05 (q<sub>N</sub>t<sub>H</sub>, <sup>2</sup>J<sub>CN</sub> = 2.1 Hz, N<sup>+</sup>CH<sub>2</sub>, 4C), 43.4 (t, C-6), 37.1 (t, C-8), 34.8 (t, C-4), 31.0 (t, C-10), 23.4 (t, N<sup>+</sup>CH<sub>2</sub>CH<sub>2</sub>CH<sub>2</sub>), 21.2 (t, C-5), 21.0 (t, C-9), 19.3 (t, N<sup>+</sup>CH<sub>2</sub>CH<sub>2</sub>CH<sub>2</sub>), 18.9 (q, CH<sub>3</sub>), 12.6 (q, N<sup>+</sup>CH<sub>2</sub>CH<sub>2</sub>CH<sub>2</sub>CH<sub>3</sub>); HRMS *m/z* calcd for C<sub>18</sub>H<sub>21</sub>O<sub>8</sub>S<sup>-</sup> [M-NBu<sub>4</sub>]<sup>-</sup> 397.0963, found 397.0961.

## 2. NMR spectra

Figure S1: COSY spectrum of 7



Figure S2: HSQC spectrum of 7



Figure S4: <sup>1</sup>H NMR spectrum of 8



Figure S5: <sup>13</sup>C NMR spectrum of 8











Figure S9: <sup>13</sup>C NMR spectrum of 14



Figure S10: <sup>1</sup>H NMR spectrum of 15





Figure S12: <sup>1</sup>H NMR spectrum of 17



Figure S14: <sup>1</sup>H NMR spectrum of 18



Figure S15: <sup>13</sup>C NMR spectrum of 18



Figure S16: <sup>1</sup>H NMR spectrum of 21



Figure S17: <sup>13</sup>C NMR spectrum of 21



Figure S18: <sup>1</sup>H NMR spectrum of 22



Figure S20: <sup>1</sup>H NMR spectrum of 24



