

Supporting Information (SI)

**Accumulation of heptaprenyl diphosphate sensitizes *Bacillus subtilis* to bacitracin:
Implications for the mechanism of resistance mediated by the BceAB transporter**

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short title: heptaprenyl diphosphate sensitizes *Bacillus subtilis* to bacitracin

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Table S1: Bacterial strains, plasmids, and oligos used in this study.^a

Strain	Genotype or description	Source, reference, or construction ¹
<i>B. subtilis</i> strains		
168	<i>trpC2</i>	Lab Stock
CU1065	<i>trpC2 attSPβ</i>	Lab Stock
HB0800	NCBI3610 (<i>trp</i> ⁺)	Lab Stock
HB0106	CU1065 <i>bcrC-pMUTIN</i> (MLS)	(Cao and Helmann, 2002)
HB0928	168 <i>bceAB::kan</i>	(Mascher <i>et al.</i> , 2003)
HB13129	168 <i>ytpA::MLS</i>	LFH PCR --> 168
HB13320	168 <i>amyE::P_{spac(hy)}-ytpB (cat)</i>	pTK083 --> 168
HB13321	168 <i>ytpB::MLS</i>	LFH PCR --> 168
HB13346	168 <i>ytpB::MLS amyE::P_{spac(hy)}-ytpB (cat)</i>	chrDNA HB13320 --> HB13321
HB13348	168 <i>bcrC-pMUTIN</i> (MLS)	chrDNA HB0106 --> 168
HB13350	168 <i>ytpB::spec</i>	ECE79 --> HB13321
HB13351	168 <i>ytpB::spec bcrC-pMUTIN</i> (MLS)	chrDNA HB0106 --> HB13350
HB13354	168 <i>ytpB::spec bceAB::kan</i>	chrDNA HB0928 --> HB13350
HB13355	168 <i>bcrC-pMUTIN</i> (MLS) <i>bceAB::kan</i>	chrDNA HB0928 --> HB13348
HB13356	168 <i>ytpB::spec bcrC-pMUTIN</i> (MLS) <i>bceAB::kan</i>	chrDNA HB0928 --> HB13351
HB13358	168 <i>sqhC::MLS</i>	LFH PCR --> 168
HB13360	168 <i>sqhC::MLS ytpB::spec</i>	HB13358 chrDNA --> HB13350
HB13367	168 <i>amyE::P_{bceA}-lacZ (cat)</i>	pTK099 --> 168
HB13369	168 <i>amyE::P_{bceA}-lacZ (cat) ytpB::spec</i>	pTK099 --> HB13350
HB13387	168 <i>trp</i> ⁺	chrDNA HB800 --> 168
HB13388	168 <i>bceAB::kan trp</i> ⁺	chrDNA HB800 --> HB0928
HB13389	168 <i>ytpB::spec trp</i> ⁺	chrDNA HB800 --> HB13350
HB13390	168 <i>ytpB::spec bceAB::kan trp</i> ⁺	chrDNA HB800 --> HB13354
HB13397	168 <i>amyE::P_{spac(hy)}-bceAB (cat)</i>	pTK102 --> 168
HB13398	168 <i>amyE::P_{spac(hy)}-hepSmenHhepT (cat)</i>	pTK103 --> 168
HB13399	168 <i>amyE::P_{spac(hy)}-menA (cat)</i>	pTK104 --> 168
HB13401	168 <i>amyE::P_{spac(hy)}-bceAB (cat) ytpB::spec</i>	chrDNA HB13397 --> HB13350
HB13402	168 <i>amyE::P_{spac(hy)}-hepSmenHhepT (cat) ytpB::spec</i>	chrDNA HB13398 --> HB13350
HB13403	168 <i>amyE::P_{spac(hy)}-menA (cat) ytpB::spec</i>	chrDNA HB13399 --> HB13350
HB13438	<i>menA::kan</i>	LFH-PCR --> 168
HB13439	<i>ytpB::MLS menA::kan</i>	HB 13438 chrDNA --> HB13321
HB13443	168 <i>uppSI (kan-SD*-uppS) ytpB::MLS</i>	HB13321 chrDNA --> HB13648
HB13447	168 <i>hepT::kan</i>	LFH-PCR --> 168
HB13448	168 <i>ytpB::MLS hepT::kan</i>	HB13446 chrDNA --> HB13447
HB13648	168 <i>uppSI (kan-SD*-uppS)</i>	(Lee and Helmann, 2013)
HB13649	168 <i>uppSI (kan-SD*-uppS) amyE::P_{spac(hy)}.uppS</i>	(Lee and Helmann, 2013)
HB17031	168 <i>bcrC-pMUTIN</i> (MLS) <i>trp</i> ⁺	chrDNA HB800 --> HB13348

HB17032	168 <i>bcrC-pMUTIN(MLS) bceAB::kan trp⁺</i>	chrDNA HB800 --> HB13355
HB17033	168 <i>ytpB::spec bcrC-pMUTIN (MLS) trp⁺</i>	chrDNA HB800 --> HB13351
HB17034	168 <i>ytpB::spec bcrC-pMUTIN (MLS) bceAB::kan trp⁺</i>	chrDNA HB800 --> HB13356
HB17055	168 <i>amyE::P_{spac(hy)}.uppS (cat)</i>	chrDNA HB13649 --> 168
HB17057	168 <i>ytpB::spec amyE::P_{spac(hy)}.uppS (cat)</i>	chrDNA HB17055 --> HB13350

Plasmids	Genotype or description	Source
ECE76	Vector for integration of a <i>cat</i> resistance cassette into a MLS resistance cassette	(Steinmetz and Richter, 1994)
ECE79	Vector for integration of a <i>spec</i> resistance cassette into a MLS resistance cassette	(Steinmetz and Richter, 1994)
pdg1661	Vector for integration of <i>lacZ</i> fusions at <i>amyE</i>	(Guerout-Fleury <i>et al.</i> , 1996)
pPL82	Vector for IPTG-inducible control of gene expression with integration at the <i>amyE</i> locus	(Quisel <i>et al.</i> , 2001)
pTK083	pPL82-ytpB	This work
pTK099	pDG1661- <i>P_{bceA}</i> (<i>cat</i>)	This work
pTK102	pPL82- <i>bceAB</i> (<i>cat</i>)	This work
pTK103	pPL82- <i>hepSmenHhepT</i> (<i>cat</i>)	This work
pTK104	pPL82- <i>menA</i> (<i>cat</i>)	This work

Oligos	Name	Sequence
5170	ytpA-up-fwd	GAGAACAGGAAAGAATAGGT
5171	ytpA-up-rev (MLS)	GAGGGTTGCCAGAGTTAAAGGATCACAATCACGGCAACAGGTCT
5172	ytpA-do-fwd (MLS)	CGATTATGTCTTTTGCAGTCGGCAATGAACCCGAGAGAGAAGAT
5173	ytpA-do-rev	AGGCTGGAACGAATACGCAA
5856	ytpB-up-fwd	TCATGTGGACCTGGAAAGCA
5857	ytpB-up-rev (MLS)	GAGGGTTGCCAGAGTTAAAGGATCGTCCAGCTCTTGATGAACGA
5858	ytpB-do-fwd (MLS)	CGATTATGTCTTTTGCAGTCGGCGGGCTTGCTTGGCGTTTA
5859	ytpB-do-rev	TGATCGTCCACCGCATTACA
5860	ytpB up XmaI	TCAGCCCGGGGAACCCGAGAGAGAAGATGT
5861	ytpB do XbaI	AGTCTCTAGATCAATTACGACAAACGGCAGT
5944	PbceA Forward EcoRI	GATCGAATTCGAACATGTCATAAGCGTGTGACG
5945	PbceA Reverse BamHI	GATCGGATCCTATCGATGCCCTCAGCACTCC
5958	sqhC upstream F	GCCTTTACCGGTTCCACTGA
5959	sqhC upstream R	GAGGGTTGCCAGAGTTAAAGGATCCGCCTCACTTTCTCCTGAAGT
5960	sqhC downstream F MLS	CGATTATGTCTTTTGCAGTCGGCGGAGACAGCAAATGAAACGTGA
5961	sqhC downstream R	CCACGACATTCCACCAGTGA
5977	hepST UbiE F XmaI	GATCCCGGGTGTGAATTTGGGGACAAGGGT
5978	hepST UbiE R XbaI	GATCTCTAGATTAATCAATCCGGGGCGGAG

5979	bceAB F XmaI	GATCCCCGGGTATCCAATATAAAGGAGACTGCGT
5980	bceAB R XbaI	GATCTCTAGAACAACCGATCCGGCTGAAAT
5983	menA F XmaI	GATCCCCGGGTTTTAATAGAAAAGGAAGAGGAA
5984	menA R XbaI	GATCTCTAGACAAAATGCGGTTCGATTGTAA
6132	menA up fwd	CCGTACACAAGGATAGGAGA
6133	menA up rev (kan)	CCTATCACCTCAAATGGTTCGCTGGCCAAAGGATCTGCCCCAT
6134	menA do fwd (kan)	CGAGCGCCTACGAGGAATTTGTATCGCTGGGTTGTCGGCTTGGTT
6135	menA do rev	GAAGGCGAAAGCATCTGACA
6136	hepT up fwd	CGGCTTATGAACGGAAATGAT
6137	hepT up rev (kan)	CCTATCACCTCAAATGGTTCGCTGCATCTGTGTTTTTCATTCCATC
6138	hepT do fwd (kan)	CGAGCGCCTACGAGGAATTTGTATCGTATGGCAGTAAGCGAAATGTAT
6139	hepT do rev	CGTTGGGAAAGGCAGAGGT

^aAbbreviations used: chrDNA indicates chromosomal DNA, LFH-PCR indicates long-flanking homology PCR, --> indicates genetic transformation.

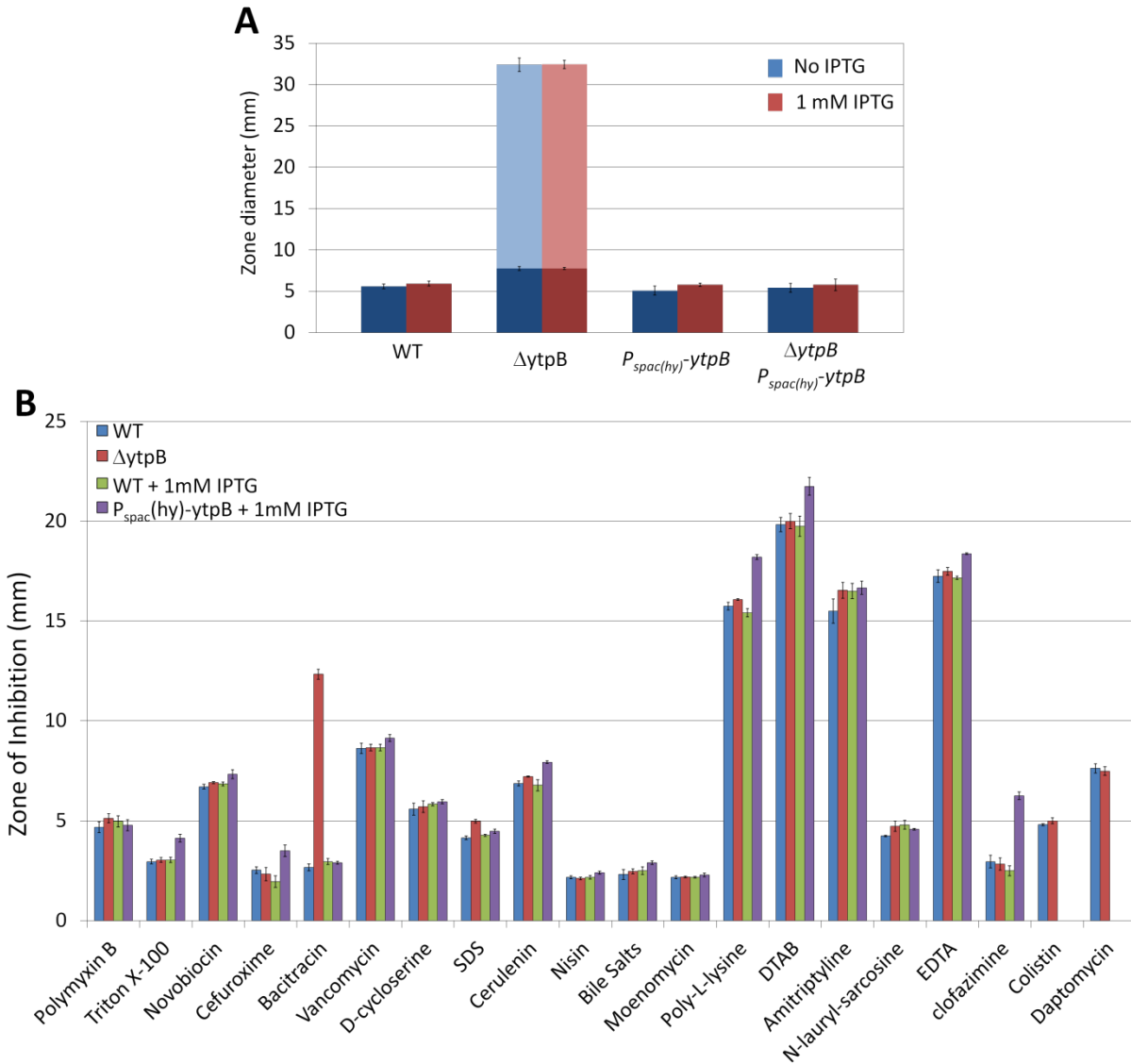


Figure S1: *ytpB* primarily confers resistance against bacitracin. (A) Bacitracin sensitivity of the WT (168), $\Delta ytpB$ (HB13321), $P_{spac(hy)}-ytpB$ (HB13320), and $P_{spac(hy)}-ytpB \Delta ytpB$ (HB13346) strains as measured with a disk diffusion assay. Bacitracin sensitivity in the $\Delta ytpB$ background reverts to wild type upon the introduction of the $P_{spac(hy)}-ytpB$ fusion even in the absence of IPTG induction. (B) The WT (168), $\Delta ytpB$ (HB13129), and $P_{spac(hy)}-ytpB$ (HB13320) strains were assessed for sensitivity to the indicated stresses on MH media supplemented with or without 1 mM IPTG. The $\Delta ytpB$ strain exhibited a large sensitivity phenotype to bacitracin (seen as a zone of early lysis; see text), but was not significantly more sensitive to the other stresses tested. Small but significant sensitivity phenotypes were observed for the $P_{spac(hy)}-ytpB$ strain to triton X-100, cefuroxime, cerulenin, poly-L-lysine, dodecyltrimethylammonium bromide (DTAB), EDTA, and clofazimine.

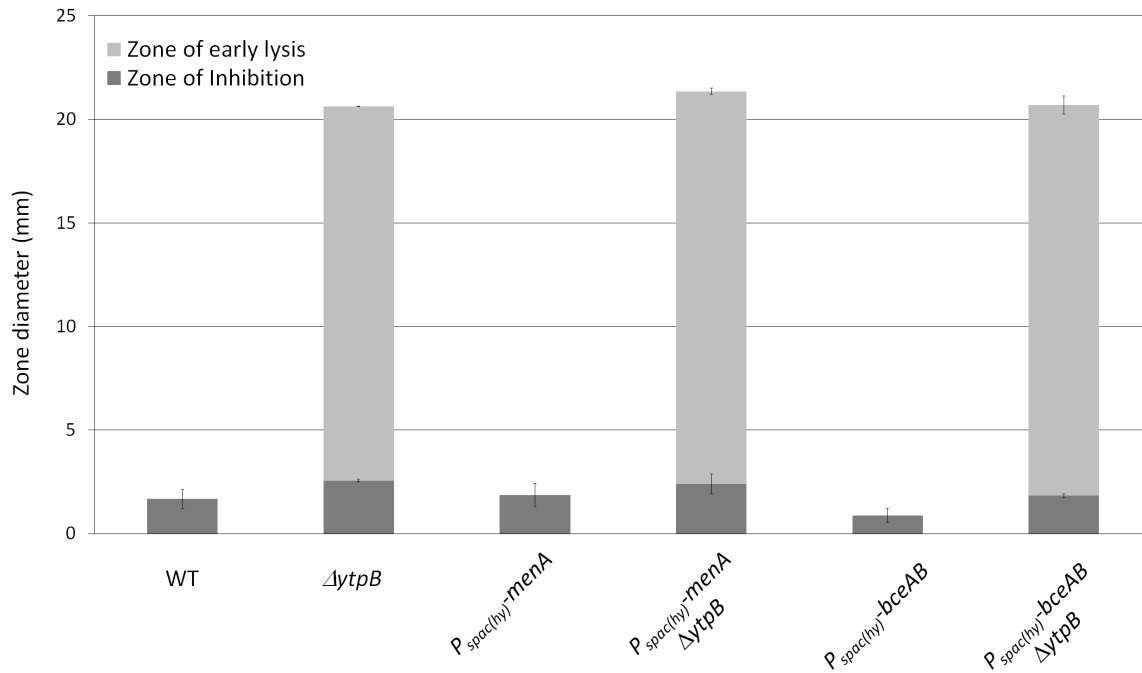


Figure S2: Bacitracin disk diffusion assays of WT (168), $\Delta ytpB$ (HB13350), $P_{spac(hy)}-menA$ (HB13399), $P_{spac(hy)}-menA \Delta ytpB$ (HB13403), $P_{spac(hy)}-bceAB$ (HB13397), and $P_{spac(hy)}-bceAB \Delta ytpB$ (HB13401) strains on MH medium supplemented with 1 mM IPTG. For this assay, bacitracin was used instead of bacitracin zinc. Overexpressing *menA* has no significant effect on bacitracin sensitivity. Overexpressing *bceAB* slightly decreased the bacitracin zone of inhibition in WT cells but did not affect sensitivity in $\Delta ytpB$ cells.

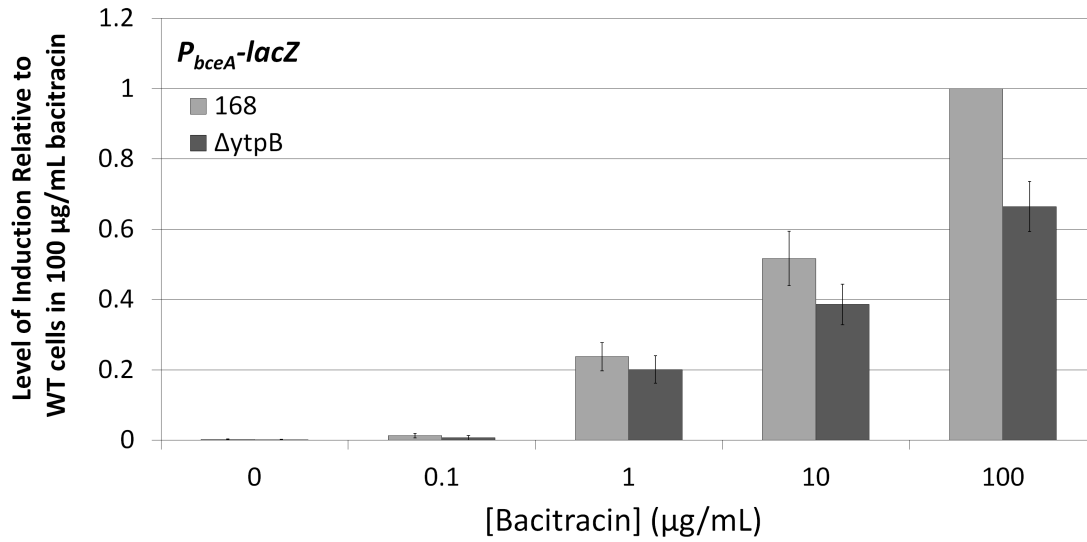


Figure S3. Induction of expression of *bceAB* by bacitracin in WT and $\Delta ytpB$ backgrounds. β -galactosidase assays were used to measure the activity of a $P_{bceA-lacZ}$ construct in WT (HB13367) and $\Delta ytpB$ (HB13369) cells with bacitracin concentrations ranging from 0.1-100 $\mu\text{g ml}^{-1}$.

References

- Cao, M. and Helmann, J.D. (2002) Regulation of the *Bacillus subtilis* *bcrC* bacitracin resistance gene by two extracytoplasmic function σ factors. *J Bacteriol* **184**: 6123-6129.
- Guerout-Fleury, A.-M., Frandsen, N., and Stragier, P. (1996) Plasmids for ectopic integration in *Bacillus subtilis*. *Gene* **180**: 57-61.
- Lee, Y.H. and Helmann, J.D. (2013) Reducing the level of undecaprenyl pyrophosphate synthase has complex effects on susceptibility to cell wall antibiotics. *Antimicrob Agents Ch* **57**: 4267-4275.
- Mascher, T., Margulis, N.G., Wang, T., Ye, R.W., and Helmann, J.D. (2003) Cell wall stress responses in *Bacillus subtilis*: the regulatory network of the bacitracin stimulon. *Mol Microbiol* **50**: 1591-1604.
- Quisel, J.D., Burkholder, W.F., and Grossman, A.D. (2001) *In vivo* effects of sporulation kinases on mutant Spo0A proteins in *Bacillus subtilis*. *J Bacteriol* **183**: 6573-6578.
- Steinmetz, M. and Richter, R. (1994) Plasmids designed to alter the antibiotic resistance expressed by insertion mutations in *Bacillus subtilis*, through *in vivo* recombination. *Gene* **142**: 79-83.