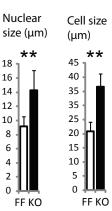
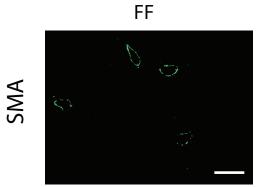


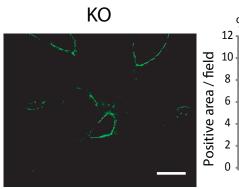
FF KO ^{Nu}siz

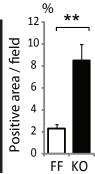


В

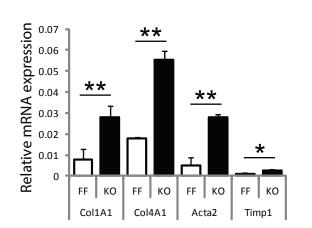
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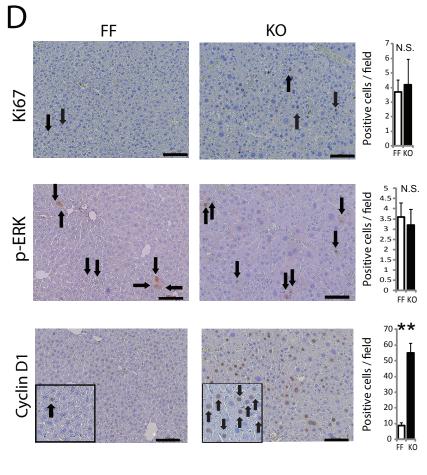






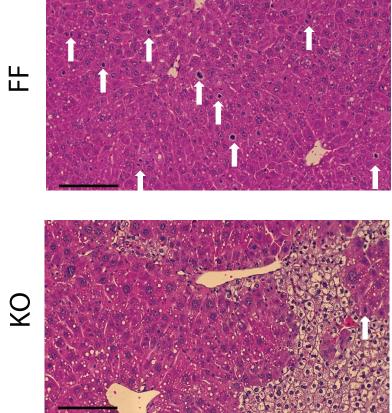
С



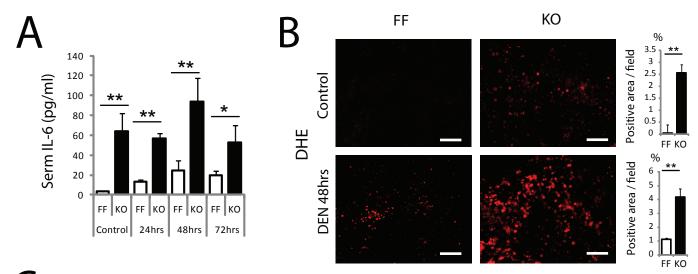


Α

В

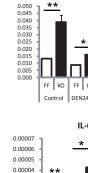


KO FF PAS PASD

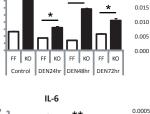


C

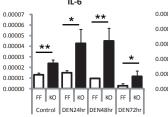
Relative mRNA expression

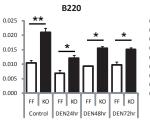


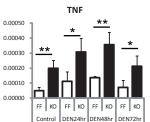
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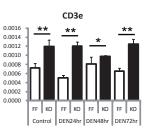


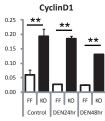
EMR1

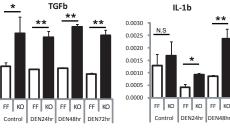


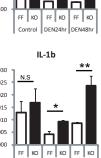




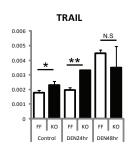


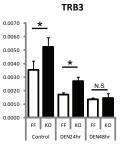


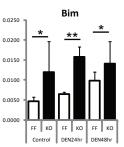




DR5 ** 0.0100 0.0090 0.0080 0.0070 0.0060 0.0050 0.0040 0.0030 0.0020 0.0010 0.0000 FF KO FF KO FF KO Control DEN24hr DEN48hr







D

0.040

0.035

0.030

0.025

0.020

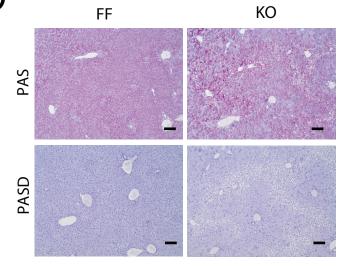
0.015

0.010

0.005

0.000

FF KO

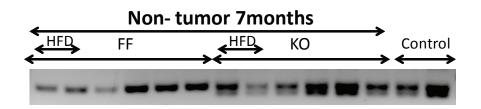


Α

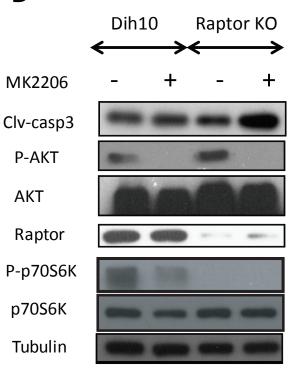
 Tumor 7 months

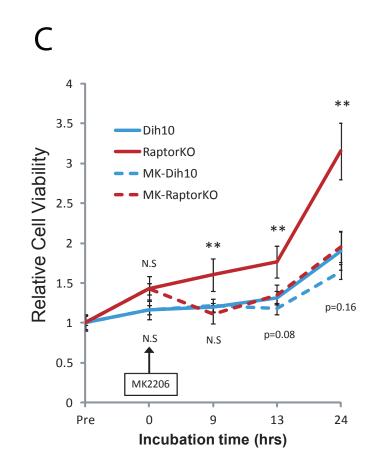
 FF
 KO
 Floxed allele
 228bp

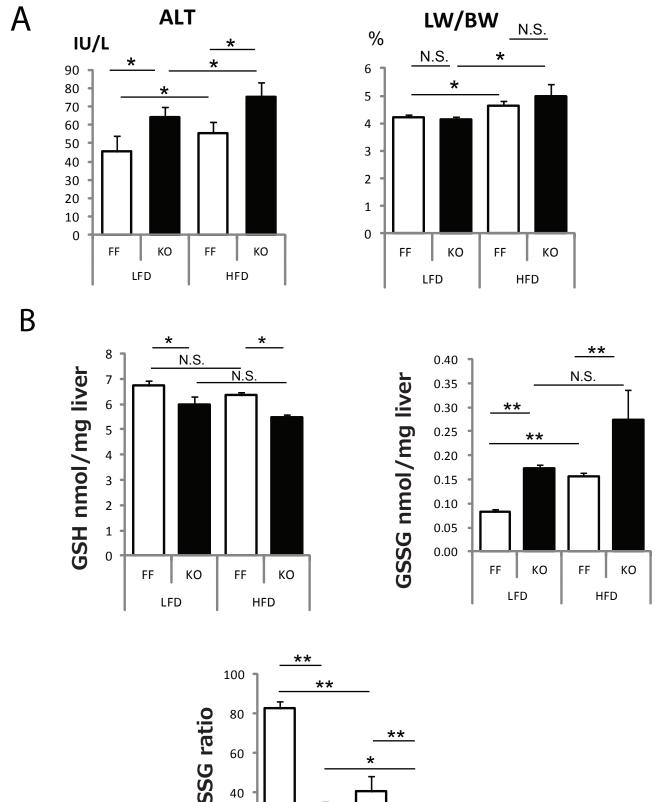
 Deleted allele
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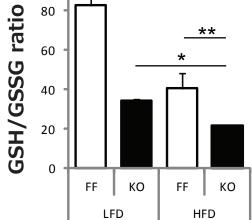












Supplemental Figure legends

Figure S1, related to Figure 1. Rapamycin treatment induces autophagy but also inhibits IL-10 expression. (A) Livers from mice kept on LFD or HFD with or without rapamycin (Rapa or R) treatment were collected and homogenized, and p62 and LC3 expression were analyzed by immunoblotting. The loading control tubulin blot is the same as in Figure 1D. (B) Relative amounts of IL-10 and SREBP1c mRNAs in above livers were determined by real time qPCR and normalized to cyclophilin mRNA. (C) AML12 cells were incubated with indicated concentration of rapamycin for 16hrs, and then relative hepatocyte viability/proliferation was analyzed. (D) Immunoblot analysis of hepatocyte lysates collected 16hrs after rapamycin incubation. (E) The morphorogical change of hepatocytes after rapamycin incubation. All the bar graphs in Figure S1 represent means +/- S.D.

Figure S2, related to Figure 2. Increased nuclear and cell size of Raptor-deficient hepatocytes. (A) Nuclear and cell sizes of hepatocytes in H&E stained liver sections from *Raptor^{F/F}* (FF), and *Raptor^{Δhep}* (KO) mice were determined. (n= 4) (scale bar: 100 µm). (B) Smooth muscle actin (SMA) staining of liver tissue sections from FF and KO mice (scale bar: 100 µm). SMA positive areas were quantified with Image J software and the results are depicted in the bar graphs. (n= 4). (C) The mRNA expression of fibrogenic markers, collagen α 1(I) (Col1A1), collagen α 1(IV) (Col4A1), actin α (Acta), and TIMP1 (Timp1) were analyzed by real time qPCR. (D) Liver cell proliferation was analyzed by immunostaining with antibodies against Ki67, p-ERK, and cyclin D1. The results were quantified and are depicted as above (n= 4). All the bar graphs in Figure S2 represent means +/- S.D.

Figure S3, related to Figure 3. Loss of Raptor inhibits hepatocyte mitosis, but enhances ERK and STAT3 activation after partial hepatectomy. $Raptor^{F/F}$ and $Raptor^{dhep}$ mice were subjected to PH as described in Figure. 3. (A) High magnification images of H&E stained liver sections 48 hrs after PH (scale bar: 100 µm). White arrows indicate representative mitotic hepatocytes. (B) PAS and PAS diastase (PASD) staining of liver tissues showing glycogen accumulation (scale bar: 100 µm).

Figure S4, related to Figure 4. Loss of Raptor enhances cyclin D1 expression, IL-6 production and ROS accumulation after DEN challenge. $Raptor^{F/F}$ or $Raptor^{1hep}$ were treated with 100 mg/Kg DEN as in Figure. 4. Livers were removed for analysis after 24 or 48 or 72 hrs. (A) Serum IL-6 was determined at the indicated times (n= 3). (B) Liver sections were prepared 48 hrs after DEN injection and analyzed for ROS accumulation by DHE staining. DHE positive areas were quantified and the results are exhibited in the bar graphs to the right (scale bar: 100 µm, n= 3). (C) mRNA expression of immune cell markers, cyclin D1, pro-inflammatory cytokines, and cell death mediators was quantitated by real-time qPCR at the indicated time-points after DEN injection (n= 3). (D) PAS and PAS diastase (PASD) staining of liver tissues showing glycogen accumulation (scale bar: 100 µm). All the bar graphs in Figure S4 represent means +/- S.D.

Figure S5, related to Figure 5. Genotyping of DEN-induced HCC and non-tumor tissue, and the effect of AKT inhibiton on HCC cells. (A) *Raptor^{F/F}* and *Raptor^{Δhep}* males were injected with 25 mg/Kg. Non-tumor and tumor liver tissues were isolated and subjected to *Raptor* genotyping by PCR. (B) Dih10 cells and HCC cells isolated from a *Raptor*^{Δhep} mouse were cultured and then incubated with the AKT inhibitor MK2206. Immunoblot analysis of HCC cell lysates collected 13hrs after MK2206 incubation. (C) Relative cell viability/proliferation in HCC cells incubated with or without MK2206 was analyzed at the indicated times (MK-Dih10; Dih10 cells incubated with MK2206, MK-Raptor; HCC cells isolated from a *Raptor*^{Δhep} mouse and incubated with MK2206). Results are means +/- S.D.

Figure S6, related to Figure 6. Inhibition of Raptor enhances liver damage. (A) Serum ALT and liver weight as percentage of total body weight (LW/BW) in LFD-fed and HFD-fed 7-months old *Raptor^{F/F}* (FF) and *Raptor^{\Deltahep}* (KO) mice. Results are means +/- S.E.M. (n= 4). (B) GSH, GSSG, and GSH/GSSG ratio in LFD-fed and HFD-fed 7 months old *Raptor^{F/F}* (FF) and *Raptor^{\Deltahep}* (KO) mice. Results are means +/- S.E.M. (n= 4). (B) GSH, GSSG, and GSH/GSSG ratio S.E.M. (n=4).

Supplemental Experimental Procedures

GSH and GSSG quantification.

GSH and GSSG were measured with a GSSG/GSH Quantification kit (Dojindo).