Supplementary Materials for

Vampire bats exhibit evolutionary reduction of bitter taste receptor genes common to other bats

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Supplementary Text 1. Description of selection tests on Calhm1 in bats.

To test whether functional relaxation of taste signaling pathway happened along with substantial reduction of taste sensation in vampire bats, we conducted three selection tests for *Calhm1* that are similar to those for *T2R*s. Briefly, we found that, 1) the average ω ratio of *Calhm1* for all branches was significantly smaller than 1 ($P = 1.9 \times 10^{-154}$) after comparing model A with model B (table 1); 2) the ω ratio estimated for the common ancestor of vampire bats was not significantly different from that of the rest of tree (P = 0.985) (see the comparsion between model C and model D in table 1); 3) a model (model F in table 1) allowing ω variation between the ancestral branch of vampire bats and the four branches linking the three vampire bats was not significantly better fit to the data than the null model (P = 0.119) (model E in table 1). For details of parameter estimates for selection tests on bat *Calhm1*, see Table S4. Together, these results suggest that *Calhm1* is under strong purifying selection in bats, and that functional relaxation on the taste signaling pathway was not observed in vampire bats.

Classification		Common name	Scientific name
Suborder	Family Pteropodidae	Geoffroy's rousette fruit bat	Rousettus amplex
YINPTEROCHIROPTERA		greenish naked-backed fruit bat	Dobsonia inermis
		large flying fox	Pteropus vampyrus
		black flying fox	Pteropus alecto
	Family Rhinolophidae	cyclops leaf-nosed bat	Hipposideros cyclops
		Pearson's horseshoe bat	Rhinolophus pearsonii
Suborder	Family Phyllostomidae	Gervais's fruit-eating bat	Artibeus cinereus
YANGOCHIROPTERA		Seba's short-tailed bat	Carollia perspicillata
		common vampire bat	Desmodus rotundus
		white-winged vampire bat	Diaemus youngi
		hairy-legged vampire bat	Diphylla ecaudata
	Family Mormoopidae	Parnell's mustached bat	Pteronotus parnellii
	Family Vespertilionidae	little brown bat	Myotis lucifugus
		David's myotis	Myotis davidii

 Table S1. Species examined in this study.

Amplified gene	Primer name	Primer sequence (5'-3')	Primer pair [*]
T2R1	M R41 1621119		forward
1 41/1	M R41 10/11/20	TGATCAGGCAGAGAAAGATG	forward
	M_R41_104020 M_R41_827I_23	CTTAGGATTTCCTAAAATTAAGA	reverse
	M_R41_027E23		reverse
T7R3	$T_{2R20} 7118$	GGACTCACAGAGTGGGTG	forward
1285	$T2R20_7010$ T2R20_48U20	GTTCTTTCTGGGAATGCTGG	forward
	T2R20_40020	CACAAATGTCTGCTTCAGCT	reverse
	T2R20_005L19	GACTTCAGACGACCAGACT	reverse
T2R4	$T_{2R19} = 56U_{20}$	CAGGACTCATTGTGAATCTG	forward
121()	T_{2R19}_{86U20}	TGGTCAACTACAAGACTTGG	forward
	T2R19_824L22	AGAATAATGAGAACAGAATGTC	reverse
	T2R19_871L21	GAAACAGAGAATCTTCTTTGC	reverse
T2R5	$T_{2R17} = 1112$	ACTGCTGATGGTGGTGGCA	forward
	$T_{2R17} 45U21$	TGAATTTCTCATTGGCCTGGT	forward
	$T_{2R17} = 13.021$ T2R17_818L20	TTTCATCCTGGGATTCCCCA	reverse
	T2R17_839L20	CAGGATTCTCTGACAAGCCT	reverse
T2R7	$T_{2R9} = 16U_{20}$	AGCAACACCTTAATGATCAT	forward
121()	$T_{2R9} = 55U_{20}$	ATGGGAATCTTAGGAAATGC	forward
	T2R9_879L20	GCACCTTTAGAAATGCTTGT	reverse
	$T_{2R9} = 0.0000000000000000000000000000000000$	TCTTTTCAGGATATATGTTACT	reverse
T2R38	M R26 87U21	GATTCTGGTCAATGTCTTCAT	forward
12130	$T_{2R31} 17U18$	CCGTCGTCACTGTGTCCT	forward
	$T_2R_{31} - 76U19$	TTCGTGGTGGGGGATTCTGG	forward
	M R26 712L18	GATGTGGGCCTCCAGGCT	reverse
	T2R31_919L18	CAGGATGGCGTCCACAGC	reverse
T2R39	$T_{2R10} 57U_{21}$	AACTTTCACAATTATAGGCAC	forward
12107	M R28 158U19	CCACAAGTGGCAGGATCCT	forward
	T2R10_880L19	GATTGCCCAAGATCAGTAG	reverse
	T2R10_914L18	GTTGAAGCCGCTTCCAGG	reverse
	M R28 798L20	TGGCATTGAAGATGTTGGAC	reverse
T2R40	T2R11 26U19	CGGATAAAGGCATGTCCAG	forward
	T2R11 40U22	TCCAGATTTAAAATCGTCTTCA	forward
	T2R11 893L18	TCAGGCCAGGATTGCCCA	reverse
	T2R11 917L18	ACTGCAGCCGCTTCCAGG	reverse
T2R42	T2R22 31U21	GTACTGTCAATAGCAGAATTC	forward
	T2R22_66U20	GGGAAATGTGTTCATTGGAC	forward
	T2R22 867L21	TTCAAGATTGTCTGTCTTAGC	reverse
	T2R22 927L22	CTATCTGTAAATCTGTAACAGA	reverse
<i>T1R3</i>	T1R3-re2-F	ACCAGGACAGCCCCTTGGT	forward
	T1R3-re2-R	GGGGCATGAAGGAGATCCAG	reverse
<i>Calhm1</i> (exon 1)	CALHM1e1p2-F	GCAGCGGTGAGGTGGGAGG	forward
	CALHM1e1p2-R	CCCCTCACCTGGGAGATGCAG	reverse
<i>Calhm1</i> (exon 2)	CALHM1e2p3-F	CTCTCCCATGCAGGCACTG	forward
· · /	CALHM1e2p3-R	GGCCACAGCTCACACTTTGC	reverse

Table S2. P	rimers used	in th	nis study.
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* Each forward primer can pair with each reverse primer.

Table S3. *T2R*s of the four bats with available genome sequences. Intact genes are characterized by complete and intact ORFs, partial genes contain incomplete and intact ORFs due to incomplete genome sequences, and pseudogenes are characterized by disrupted ORFs due to nonsense or frame-shifting mutations.

Species	Gene number				Percent of
	Intact	Partial	Pseudogene	Total	pseudogenes
Myotis davidii	25	4	10	39	25.6%
Myotis lucifugus	27	0	7	34	20.6%
Pteropus alecto	13	0	13	26	50.0%
Pteropus vampyrus	14	0	10	24	41.7%

Table S4. Likelihood values and parameter estimates for likelihood ratio tests of selective pressures on bat *T2R*s and *Calhm1*. The assumption of each model was given in table 1.

	Model A	Model B	Model C	Model D	Model E	Model F
T2R1	ln <i>L</i> =-3427.132844	ln <i>L</i> =-3428.833944	ln <i>L</i> =-3527.065761	ln <i>L</i> =-3526.200984	ln <i>L</i> =-3627.802949	ln <i>L</i> =-3627.174896
	ω=0.8276	ω=1	ω=0.8263	$\omega_1=0.8430, \omega_2=0.3047$	ω ₁ =0.8730, ω ₂ =0.7150	ω_1 =0.8695, ω_2 =1.0069, ω_3 =0.3313
T2R3	ln <i>L</i> =-4263.749638	ln <i>L</i> =-4275.088529	ln <i>L</i> =-4400.220476	ln <i>L</i> =-4399.407653	ln <i>L</i> =-4053.415269	ln <i>L</i> =-4052.104084
	ω=0.6585	ω=1	ω=0.6697	ω_1 =0.6594, ω_2 =1.8957	ω ₁ =0.7100, ω ₂ =0.7281	ω_1 =0.6941, ω_2 =0.8084, ω_3 =3.5745
T2R4	ln L=-3209.936509	ln <i>L</i> =-3225.293118	ln <i>L</i> =-2621.778461	ln <i>L</i> =-2621.229081	ln <i>L</i> =-2825.868116	ln <i>L</i> =-2825.543076
	ω=0.572	ω=1	ω=0.5854	ω_1 =0.6006, ω_2 =0.2878	ω ₁ =0.6178, ω ₂ =0.7636	ω_1 =0.6300, ω_2 =0.8823, ω_3 =0.3936
T2R5	ln L=-3428.39021	ln <i>L</i> =-3442.627536	ln <i>L</i> =-3427.53808	ln <i>L</i> =-3427.312371	ln <i>L</i> =-3430.459055	ln <i>L</i> =-3430.459055
	ω=0.5908	ω=1	ω=0.5837	ω_1 =0.2643, ω_2 =0.3578	ω ₁ =0.2726, ω ₂ =0.3941	ω_1 =0.6036, ω_2 =1.0264, ω_3 =0.2059
T2R7	ln <i>L</i> =-1905.509419	ln <i>L</i> =-1910.159618	ln <i>L</i> =-1917.232499	ln <i>L</i> =-1917.232491	ln <i>L</i> =-2069.501724	ln <i>L</i> =-2069.08614
	ω=0.64228	ω=1	ω=0.6421	ω_1 =0.6421, ω_2 =239.4661	ω ₁ =0.6435, ω ₂ =0.2534	ω_1 =0.617, ω_2 =0.1052, ω_3 =1.3055
T2R38	ln <i>L</i> =-2815.328795	ln L=-2867.976306	ln <i>L</i> =-2705.549881	ln <i>L</i> =-2705.545709	ln <i>L</i> =-3174.197658	ln <i>L</i> =-3174.677814
	ω=0.3296	ω=1	ω=0.3566	$\omega_1 = 0.3571, \omega_2 = 0.3311$	ω ₁ =0.3450, ω ₂ =0.4829	ω_1 =0.3460, ω_2 =0.4928, ω_3 =0.5434
T2R39	ln <i>L</i> =-3071.946349	ln <i>L</i> =-3082.233326	ln <i>L</i> =-2274.85814	ln <i>L</i> =-2274.589168	ln <i>L</i> =-2643.864733	ln <i>L</i> =-2643.851076
	ω=0.6112	ω=1	ω=0.4388	ω_1 =0.4282, ω_2 =0.6227	ω ₁ =0.5368, ω ₂ =0.7020	ω_1 =0.5374, ω_2 =0.7233, ω_3 =0.6518
T2R40	ln <i>L</i> =-1933.761815	ln <i>L</i> =-1981.834222	ln <i>L</i> =-1976.861014	ln <i>L</i> =-1976.848337	ln <i>L</i> =-2279.690887	ln <i>L</i> =-2278.46542
	ω=0.2643	ω=1	ω=0.2654	ω_1 =0.2643, ω_2 =0.3578	ω ₁ =0.2726, ω ₂ =0.3941	ω_1 =0.2639, ω_2 =0.5168, ω_3 =0.4964
T2R42	ln <i>L</i> =-3238.165213	ln <i>L</i> =-3238.214621	ln <i>L</i> =-3213.889067	ln <i>L</i> =-3213.834829	ln <i>L</i> =-4086.161606	ln <i>L</i> =-4086.275863
	ω=0.9668	ω=1	ω=0.8962	ω ₁ =0.9100, ω ₂ =0.8327	ω_1 =0.9078, ω_2 =0.7592	$\omega_1=0.8587, \omega_2=0.5558, \omega_3=1.1973$
Calhm1	ln <i>L</i> =-3205.467111	ln <i>L</i> =-3555.917574	ln <i>L</i> =-3255.222199	ln <i>L</i> =-3255.222033	ln <i>L</i> =-3517.697681	ln <i>L</i> =-3516.483621
	ω=0.0565	ω=1	ω=0.0557	ω_1 =0.0557, ω_2 =0.0001	ω ₁ =0.0558, ω ₂ =0.0683	ω ₁ =0.0576, ω ₂ =0.0390, ω ₃ =223.3136

Figure legends:

Figure S1. Phylogenetic positions of bats revealed by *Cytb* genes, which were newly sequenced or obtained from the GenBank (indicated with an asterisk). The tree was constructed with the Bayesian method, and numbers at the nodes are the Bayesian posterior probabilities as percentages.

Figure S2. Evolutionary relationships of all intact *T2Rs* from the four bats with available genome sequences. A total of 211 codons were used to reconstruct a maximum-likelihood tree with the Bayesian method. The tree was rooted with the mouse *V1Rd8* and *V1Re9*, and numbers at the nodes are the Bayesian posterior probabilities as percentages. Species include *Pteropus vampyrus* (Ptva), *P. alecto* (Ptal), *Myotis lucifugus* (Mylu), and *M. davidii* (Myda).

Figure S3. Phylogenetic trees of each T2R gene in all bats using the Bayesian method. The Bayesian trees were rooted with the orthologous genes in humans, and numbers at the nodes are the Bayesian posterior probabilities as percentages.

Figure S4. An alignment of *T1R3* encoding the shared subunit of sweet and umami tastes in two vampire bats (*D. rotundus* and *D. ecaudata*), one megabat (*P. vampyrus*), and dog (*C. familiaris*). *D. rotundus* was sequenced previously [ref. 10 in the main text] while *D. ecaudata* was sequenced in this study (GenBank Accession no. KJ557282). Dashes indicate alignment gaps and question marks represent unamplified nucleotides, regions corresponding to transmembrane domains are boxed.

Figure S1







0.1

Figure S3 continued

- Artibeus cinereus

Diaemus youngi

– Diphylla ecaudata

- Carollia perspicillata

Desmodus rotundus



T2R40



C	familiaris	
ι.	jumilluris	

P. vampyrus

D. rotundus

D. ecaudata

360

480

600

720

840

960

969 AATGAGTGA ????????? ??????????