

Enantiospecific, Nickel-Catalyzed Cross-Couplings of Allylic Pivalates and Arylboroxines

Harathi D. Srinivas, Qi Zhou, Mary P. Watson*

Department of Chemistry and Biochemistry, University of Delaware, Newark, DE 19716

mpwatson@udel.edu

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General Information

Reactions were performed either in a N₂-atmosphere glovebox in oven-dried 1-dram vials with Teflon-lined caps or in oven-dried round-bottomed flasks unless otherwise noted. Flasks were fitted with rubber septa, and reactions were conducted under a positive pressure of N₂. Stainless steel syringes or cannulae were used to transfer air- and moisture-sensitive liquids. Flash chromatography was performed on silica gel 60 (40-63 µm, 60Å) unless otherwise noted. Thin layer chromatography (TLC) was performed on glass plates coated with silica gel 60 with F254 indicator. Commercial reagents were purchased from Sigma Aldrich, Acros, Fisher, Strem, TCI, Combi Blocks, Alfa Aesar, or Cambridge Isotopes Laboratories and used as received with the following exceptions: toluene, CH₂Cl₂, dioxane, MeCN and Et₂O were dried by passing through drying columns.¹ Toluene and MeCN were then degassed by sparging with N₂ and stored over activated 4Å MS in a N₂-atmosphere glovebox. Pivaloyl chloride was purchased from Sigma Aldrich and distilled before use. Boronic acids were purchased from Combi Blocks and were converted to the boroxines according to literature procedure.² Racemic products **2–24** were prepared from the corresponding racemic pivalates using the General Procedure for the cross coupling described below. CDCl₃ was stored over oven-dried potassium carbonate. Proton nuclear magnetic resonance (¹H NMR) spectra and carbon nuclear magnetic resonance (¹³C NMR) spectra were recorded on both 400 MHz and 600 MHz spectrometers. Chemical shifts for protons are reported in parts per million downfield from tetramethylsilane and are referenced to residual protium in the NMR solvent (CHCl₃ = δ 7.26; (CD₃)₂CO = δ 2.07). Chemical shifts for carbon are reported in parts per million downfield from tetramethylsilane and are referenced to the carbon resonances of the solvent (CDCl₃ = δ 77.2; (CD₃)₂CO = δ 28.94). Data are represented as follows: chemical shift, multiplicity (br = broad, s = singlet, d = doublet, t = triplet, q = quartet, p = pentet, m = multiplet, dd = doublet of doublets, h = heptet), coupling constants in Hertz (Hz), integration. Infrared (IR) spectra were obtained using FTIR spectrophotometers with material loaded onto a NaCl plate. The mass spectral data were

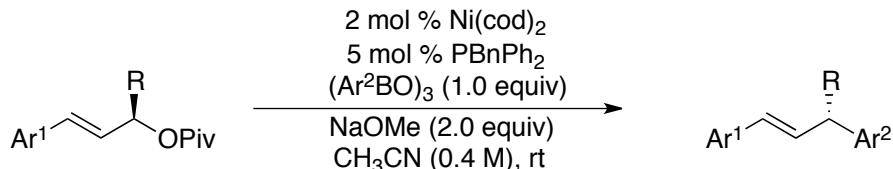
¹ Pangborn, A. B.; Giardello, M. A.; Grubbs, R. H.; Rosen, R. K.; Timmers, F. J. *Organometallics* **1996**, *15*, 1518.

² Xiao, Q.; Tian, L.; Tan, R.; Xia, Y.; Qiu, D.; Zhang, Y.; Wang, J. *Org. Lett.* **2012**, *14*, 4230.

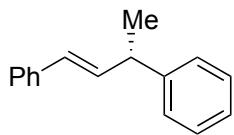
obtained at the University of Delaware mass spectrometry facility. Optical rotations were measured using a 2.5 mL cell with a 0.1 dm path length. Melting points were taken on a Stuart SMP10 instrument.

Stereospecific Cross Coupling of Allylic Pivalates with Boroxines

General Procedure for the Stereospecific, Nickel-Catalyzed Coupling of Allylic Pivalates with Boroxines

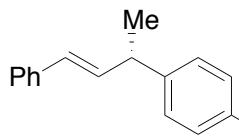


In a N₂-atmosphere glovebox, Ni(cod)₂ (1.7 mg, 0.06 mmol, 2 mol %), benzyl diphenyl phosphine (4.2 mg, 0.015 mmol, 5 mol %) and NaOMe (32.5 mg, 0.6 mmol, 2.0 equiv) were weighed into a 1-dram vial. Allylic pivalate (0.30 mmol, 1.0 equiv) and boroxine (0.30 mmol, 1.0 equiv) were added, followed by acetonitrile (0.75 mL, 0.4 M). The vial was capped with a Teflon-lined cap and removed from the glovebox. The mixture was stirred at room temperature for the time specified below. The reaction mixture was then diluted with Et₂O (1.5 mL) and filtered through a plug of silica gel, which was rinsed with Et₂O (10 mL). The filtrate was concentrated and then purified by silica gel chromatography to give the arylated product.

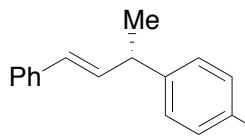


(S,E)-But-1-ene-1,3-diylbenzene (2). Product **2** was prepared via the General Procedure using pivalate **1a** (prepared in 96% ee). The reaction was stirred at room temperature for 4 h. The crude material was purified by silica gel chromatography (100% hexanes) to give compound **2** (run 1: 55.6 mg, 89%; run 2: 54.3 mg, 87%) as colorless oil. The enantiomeric excess was determined to be 95% (run 1: 95% ee; run 2: 95% ee) by chiral HPLC analysis (CHIRALCEL OD-H, 1.0 mL/min, 0.1% *i*-PrOH/hexane $\lambda=254$ nm); $t_R(\text{major})=6.07$ min, $t_R(\text{minor})=6.48$ min. $[\alpha]_D^{24} = -21.1^\circ$ (c 1.42, CHCl₃): ¹H NMR (400 MHz, CDCl₃) δ 7.40 – 7.36 (m, 2H), 7.35 – 7.27 (m, 6H), 7.25 – 7.19 (m, 2H), 6.46 – 6.37 (m, 2H), 3.69 – 3.63 (m, 1H), 1.49 (d, $J = 7.0$ Hz, 3H); ¹³C NMR (101 MHz, (CD₃)₂CO) δ 145.8, 137.6, 135.2, 128.5, 128.43, 128.40, 127.1,

127.0, 126.11, 126.07, 42.6, 20.9. The spectral data for this compound matches that reported in the literature.³



(S)-2-(1-Phenylethyl)naphthalene (3). Compound **3** was prepared via the General Procedure using pivalate **1a** (prepared in 96% ee). The crude material was purified by silica gel chromatography (2% Et₂O/hexanes) to give compound **3** (run 1: 65.9 mg, 92%; run 2: 63.0 mg, 88%) as colorless oil. The enantiomeric excess was determined to be 88% (run 1: 87% ee; run 2: 89% ee) by chiral HPLC analysis (CHIRALCEL OD-H, 0.8 mL/min, 0.1% *i*-PrOH/hexane, λ =254 nm); t_R (major)=14.47 min, t_R (minor)=13.13 min. $[\alpha]_D^{24} = -2.8^\circ$ (c 3.56, CHCl₃): ¹H NMR (400 MHz, CDCl₃) δ 7.41 – 7.35 (m, 2H), 7.34 – 7.26 (m, 2H), 7.25 – 7.16 (m, 3H), 6.93 – 6.84 (m, 2H), 6.45 – 6.34 (m, 2H), 3.81 (s, 3H), 3.66 – 3.55 (m, 1H), 1.46 (d, J = 7.0 Hz, 3H); ¹³C NMR (101 MHz, CDCl₃) δ 158.1, 137.8, 137.7, 135.7, 128.6, 128.39, 128.36, 127.2, 126.3, 114.0, 55.4, 41.9, 21.5. The spectral data for this compound matches that reported in the literature.⁴

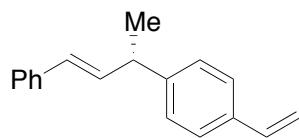


((S,E)-Methyl(4-(4-phenylbut-3-en-2-yl)phenyl)sulfane (4). Compound **4** was prepared via the General Procedure using pivalate **1a** (prepared in 96% ee). The crude material was purified by silica gel chromatography (100% hexanes) to give compound **4** (run 1: 73.2 mg, 96%; run 2: 70.6 mg, 93%) as a white amorphous solid (mp 49–51 °C). The enantiomeric excess was determined to be 93% (run 1: 93% ee; run 2: 92% ee) by chiral HPLC analysis (CHIRALPAK IA, 1.0 mL/min, 100% hexanes, λ =254 nm); t_R (major)=13.90 min, t_R (minor)=17.71 min. $[\alpha]_D^{24} = -8.3^\circ$ (c 1.44, CHCl₃): ¹H NMR (600 MHz, (CD₃)₂CO) δ 7.43 – 7.38 (m, 2H), 7.32 – 7.16 (m, 7H), 6.48 – 6.44 (m, 2H), 3.68 – 3.60 (m, 1H), 2.46 (s, 3H), 1.43 (d, J = 7.0 Hz, 3H); ¹³C NMR (151 MHz,

³ Sanz, R.; Miguel, D.; Martínez, A.; Álvarez-Gutiérrez, J. M.; Rodríguez, F. *Org. Lett.* **2007**, 9, 727.

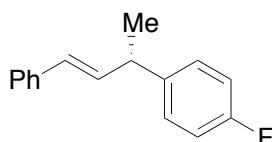
⁴ Malkov, A. V.; Davis, S. L.; Baxendale, I. R.; Mitchell, W. L.; Kocovský, P. *J. Org. Chem.* **1999**, 64, 2751.

CDCl_3) δ 142.8, 137.5, 135.9, 135.0, 128.6, 128.5, 127.9, 127.2, 127.1, 126.2, 42.1, 21.2, 16.3; FTIR (NaCl/thin film) 3023, 2963, 2920, 2869, 1891, 1799, 1598, 1493, 1446, 1010, 964, 746; HRMS (EI+) $[\text{M}]^+$ calculated for $\text{C}_{17}\text{H}_{18}\text{S}$: 254.1129, found: 254.1134.



(*S,E*)-1-(4-Phenylbut-3-en-2-yl)-4-vinylbenzene (5).

Compound **5** was prepared via the General Procedure using pivalate **1a** (prepared in 96% ee). The crude material was purified by silica gel chromatography (100% hexanes) to give compound **5** (run 1: 65.4 mg, 93%; run 2: 64.8 mg, 92%) as yellow oil. The enantiomeric excess was determined to be 91% (run 1: 91% ee; run 2: 91% ee) by chiral HPLC analysis (CHIRALPAK IA, 1.0 mL/min, 100% hexanes, $\lambda=254$ nm); $t_{\text{R}}(\text{major})=7.82$ min, $t_{\text{R}}(\text{minor})=9.24$ min. $[\alpha]_D^{24} = -22.6^\circ$ (c 2.21, CHCl_3): ^1H NMR (400 MHz, CDCl_3) δ 7.43 – 7.35 (m, 4H), 7.33 – 7.17 (m, 5H), 6.73 (dd, $J = 17.6, 10.9$ Hz, 1H), 6.49 – 6.34 (m, 2H), 5.75 (dd, $J = 17.6, 1.0$ Hz, 1H), 5.23 (dd, $J = 10.8, 1.0$ Hz, 1H), 3.66 (qd, $J = 6.9, 5.4$ Hz, 1H), 1.48 (d, $J = 7.0$ Hz, 3H); ^{13}C NMR (101 MHz, CDCl_3) δ 145.5, 137.7, 136.7, 135.8, 135.2, 128.74, 128.67, 127.6, 127.3, 126.5, 126.3, 113.4, 42.5, 21.3; FTIR (NaCl/thin film) 3023, 2964, 2928, 1629, 1509, 1494, 965, 838, 746 cm^{-1} ; HRMS (EI+) $[\text{M}]^+$ calculated for $\text{C}_{18}\text{H}_{18}$: 234.1409, found: 234.1412.



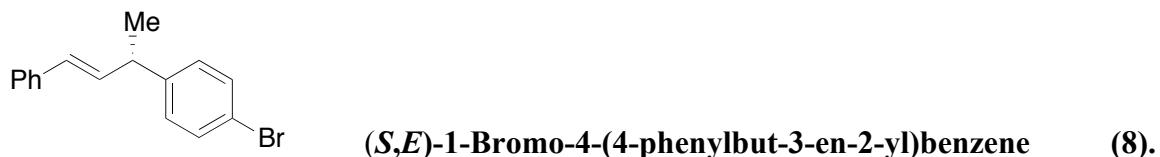
(*S,E*)-1-Fluoro-4-(4-phenylbut-3-en-2-yl)benzene (6).

Compound **6** was prepared via the General Procedure using pivalate **1a** (prepared in 95% ee). The crude material was purified by silica gel chromatography (100% hexanes) to give compound **6** (run 1: 57.6 mg, 87%; run 2: 58.9 mg, 85%) as a yellow oil. The enantiomeric excess was determined to be 93% (run 1: 93% ee; run 2: 93% ee) by chiral HPLC analysis (CHIRALPAK IB, 1.0 mL/min, 100% hexanes, $\lambda=254$ nm); $t_{\text{R}}(\text{major})=7.86$ min, $t_{\text{R}}(\text{minor})=7.41$ min. $[\alpha]_D^{24} = -15.4^\circ$ (c 2.34, CHCl_3): ^1H NMR (400 MHz, CDCl_3) δ 7.41 – 7.18 (m, 7H), 7.07 – 6.97 (m, 2H), 6.46 – 6.31 (m, 2H), 3.69 – 3.60 (m, 1H), 1.47 (d, $J = 7.0$ Hz, 3H); ^{13}C NMR (101 MHz, CDCl_3) δ 161.5 (d, $J_{\text{C}-\text{F}} = 243.8$ Hz), 141.4 (d, $J_{\text{C}-\text{F}} = 3.2$ Hz), 137.5, 135.1, 128.9, 128.8 (d, $J_{\text{C}-\text{F}} = 1.4$ Hz), 128.7,

127.3, 126.3, 115.4 (d, $J_{C-F} = 21.0$ Hz), 42.0, 21.5. The spectral data for this compound matches that reported in the literature.⁵



Compound **7** was prepared via the General Procedure using pivalate **1a** (prepared in 96% ee). The crude material was purified by silica gel chromatography (100% hexanes) to give compound **7** (run 1: 53.7 mg, 74%; run 2: 54.6 mg, 75%) as a yellow oil. The enantiomeric excess was determined to be 94% (run 1: 94% ee; run 2: 94% ee) by chiral HPLC analysis (CHIRALPAK IA, 1.0 mL/min, 100% hexanes, $\lambda = 254$ nm); $t_R(\text{major})=7.38$ min, $t_R(\text{minor})=8.11$ min. $[\alpha]_D^{24} = -13.9^\circ$ (c 3.36 CHCl₃): ¹H NMR (400 MHz, CDCl₃) δ 7.41 – 7.28 (m, 6H), 7.25 – 7.18 (m, 3H), 6.46 – 6.29 (m, 2H), 3.68 – 3.59 (m, 1H), 1.46 (d, $J = 7.0$ Hz, 3H); ¹³C NMR (101 MHz, CDCl₃) δ 144.2, 137.5, 134.7, 132.0, 129.0, 128.9, 128.74, 128.71, 127.4, 126.3, 42.1, 21.3. The spectral data for this compound matches that reported in the literature.⁶

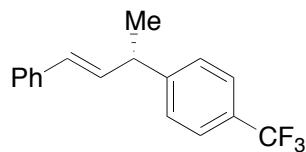


Compound **8** was prepared via the General Procedure using pivalate **1a** (prepared in 96% ee). The crude material was purified by silica gel chromatography (100% hexanes) to give compound **7** (run 1: 53.4 mg, 62%; run 2: 54.2 mg, 63%) as a yellow oil. The enantiomeric excess was determined to be 96% (run 1: 96% ee; run 2: 95% ee) by chiral HPLC analysis (CHIRALPAK IA, 1.0 mL/min, 100% hexanes, $\lambda = 254$ nm); $t_R(\text{major})=8.22$ min, $t_R(\text{minor})=9.44$ min. $[\alpha]_D^{24} = -5.8^\circ$ (c 1.38, CHCl₃): ¹H NMR (400 MHz, CDCl₃) δ 7.47 – 7.42 (m, 2H), 7.38 – 7.34 (m, 2H), 7.30 (dd, $J = 8.5, 6.7$ Hz, 2H), 7.24 – 7.19 (m, 1H), 7.18 – 7.13 (m, 2H), 6.47 – 6.29 (m, 2H), 3.66 – 3.57 (m, 1H), 1.45 (d, $J = 7.0$ Hz, 3H); ¹³C NMR (101 MHz, CDCl₃) δ 144.7, 137.4, 134.6, 131.7, 129.3,

⁵ Mauleón, P.; Carretero, J. C. *Org. Lett.* **2004**, 6, 3195.

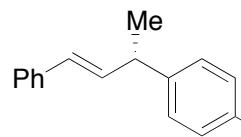
⁶ Li, M.-B.; Wang, Y.; Tian, S.-K. *Angew. Chem. Int. Ed.* **2012**, 51, 2968.

129.1, 128.7, 127.4, 126.3, 120.1, 42.2, 21.3. The spectral data for this compound matches that reported in the literature.⁷



(S,E)-1-(4-Phenylbut-3-en-2-yl)-4-(trifluoromethyl)benzene

(9). Compound **9** was prepared via the General Procedure using pivalate **1a** (prepared in 96% ee). The crude material was purified by silica gel chromatography (100% hexanes) to give compound **9** (run 1: 64.6 mg, 78%; run 2: 63.8 mg, 77%) as a colorless oil. The enantiomeric excess was determined to be 91% (run 1: 90% ee; run 2: 91% ee) by chiral HPLC analysis (CHIRALCEL OD-H, 0.6 mL/min, 0.1% *i*-PrOH/hexane, $\lambda=254$ nm); $t_R(\text{major})=11.14$ min, $t_R(\text{minor})=12.14$ min. $[\alpha]_D^{24} = -9.4^\circ$ (c 2.32, CHCl₃): ¹H NMR (400 MHz, CDCl₃) δ 7.58 (d, *J* = 8.1 Hz, 2H), 7.43 – 7.35 (m, 4H), 7.31 (dd, *J* = 8.5, 6.7 Hz, 2H), 7.25 – 7.19 (m, 1H), 6.47 – 6.32 (m, 2H), 3.76 – 3.68 (m, 1H), 1.49 (d, *J* = 7.0 Hz, 3H); ¹³C NMR (101 MHz, CDCl₃) δ 149.8 (q, *J*_{C-F} = 1.3 Hz), 137.3, 134.2, 129.5, 128.72, 128.69 (q, *J*_{C-F} = 32.2 Hz), 127.8, 127.5, 125.6 (q, *J*_{C-F} = 3.8 Hz), 124.5 (q, *J*_{C-F} = 271.6 Hz), 120.4, 42.6, 21.2. The spectral data for this compound matches that reported in the literature.⁸



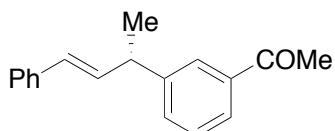
(S,E)-Methyl 4-(4-phenylbut-3-en-2-yl)benzoate (10).

Compound **10** was prepared via the General Procedure using pivalate **1a** (prepared in 96% ee). The crude material was purified by silica gel chromatography (5% Et₂O/hexanes) to give compound **10** (run 1: 75.1 mg, 94%; run 2: 73.4 mg, 92%) as a colorless oil. The enantiomeric excess was determined to be 89% (run 1: 89% ee; run 2: 89% ee) by chiral HPLC analysis (CHIRALPAK IA, 0.8 mL/min, 1% *i*-PrOH/hexane, $\lambda=254$ nm); $t_R(\text{major})=21.99$ min, $t_R(\text{minor})=19.68$ min. $[\alpha]_D^{24} = -30.2^\circ$ (c 2.91, CHCl₃):

⁷ Zhou, T.; Peters, B.; Maldonado, M. F.; Govender, T.; Andersson, P. G. *J. Am. Chem. Soc.* **2012**, 134, 13592.

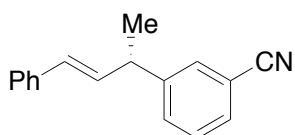
⁸ Zhao, J.; Ye, J.; Zhang, Y. *J. Adv. Synth. Catal.* **2013**, 355, 491.

¹H NMR (400 MHz, CDCl₃) δ 8.07 – 7.97 (m, 2H), 7.43 – 7.28 (m, 6H), 7.25 – 7.18 (m, 1H), 6.50 – 6.30 (m, 2H), 3.92 (s, 3H), 3.75 – 3.67 (m, 1H), 1.49 (d, *J* = 7.0 Hz, 3H); ¹³C NMR (101 MHz, CDCl₃) δ 167.2, 151.1, 137.4, 134.3, 130.0, 129.3, 128.7, 128.3, 127.5, 127.4, 126.3, 52.2, 42.8, 21.2. The spectral data for this compound matches that reported in the literature.⁹



(*S,E*)-1-(3-(4-Phenylbut-3-en-2-yl)phenyl)ethanone (11).

Compound **11** was prepared via the General Procedure using pivalate **1a** (prepared in 95% ee). The crude material was purified by silica gel chromatography (6% Et₂O/hexanes) to give compound **11** (run 1: 65.4 mg, 87%; run 2: 64.2 mg, 85%) as a colorless oil. The enantiomeric excess was determined to be 91% ee (run 1: 91% ee; run 2: 91% ee) by chiral HPLC analysis (CHIRALPAK IB, 1.0 mL/min, 0.5% *i*-PrOH/hexane, λ =254 nm); *t*_R(major)=18.72 min, *t*_R(minor)=21.37 min. [α]_D²⁴ = -26.1° (c 2.64, CHCl₃): ¹H NMR (400 MHz, CDCl₃) δ 7.88 (t, *J* = 1.8 Hz, 1H), 7.81 (dt, *J* = 7.6, 1.5 Hz, 1H), 7.49 (dt, *J* = 7.7, 1.6 Hz, 1H), 7.42 (t, *J* = 7.6 Hz, 1H), 7.39 – 7.34 (m, 2H), 7.33 – 7.27 (m, 2H), 7.24 – 7.18 (m, 1H), 6.57 – 6.22 (m, 2H), 3.76 – 3.68 (m, 1H), 2.62 (s, 3H), 1.50 (d, *J* = 7.0 Hz, 3H); ¹³C NMR (101 MHz, CDCl₃) δ 198.5, 146.4, 137.5, 137.4, 134.6, 132.4, 129.2, 128.9, 128.7, 127.4, 127.1, 126.7, 126.3, 42.7, 26.9, 21.3; FTIR (NaCl/thin film) 3025, 2965, 2927, 1681, 1598, 1448, 1356, 1264, 966, 750, 693 cm⁻¹; HRMS (EI+) [M]⁺ calculated for C₁₈H₁₈O: 250.1358, found: 250.1376.

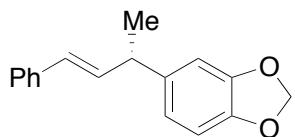


(*S,E*)-3-(4-Phenylbut-3-en-2-yl)benzonitrile (12).

Compound **12** was prepared via the General Procedure using pivalate **1a** (prepared in 96% ee). The crude material was purified by silica gel chromatography (2% Et₂O/hexanes) to give compound **12** (run 1: 60.2 mg, 85%; run 2: 58.4 mg, 83%) as a colorless oil. The enantiomeric excess was determined to be 93% (run 1: 92% ee; run 2: 93% ee) by chiral HPLC analysis (CHIRALPAK IA, 1.0 mL/min, 2.0% *i*-PrOH/hexane, λ =254 nm);

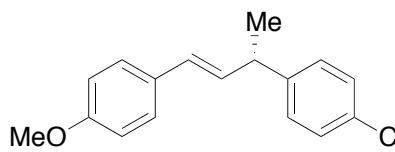
⁹ Liao, L.; Sigman, M. S. *J. Am. Chem. Soc.* **2010**, 132, 10209.

t_R (major)=9.53 min, t_R (minor)=10.52 min. $[\alpha]_D^{24} = -19.5^\circ$ (c 2.40, CHCl₃): ¹H NMR (400 MHz, CDCl₃) δ 7.57 (t, J = 1.7 Hz, 1H), 7.52 (dd, J = 8.2, 1.4 Hz, 2H), 7.46 – 7.28 (m, 5H), 7.26 – 7.20 (m, 1H), 6.48 – 6.40 (m, 1H), 6.31 (dd, J = 15.9, 6.7 Hz, 1H), 3.73 – 3.64 (m, 1H), 1.48 (d, J = 7.0 Hz, 3H); ¹³C NMR (101 MHz, CDCl₃) δ 147.1, 137.1, 133.6, 132.2, 131.1, 130.2, 129.8, 129.4, 128.7, 127.6, 126.3, 119.2, 112.6, 42.3, 21.1; FTIR (NaCl/thin film) 3025, 2967, 2930, 2228, 1590, 1481, 1446, 960, 798, 751 cm⁻¹; HRMS (EI+) [M]⁺ calculated for C₁₇H₁₅N: 233.1204, found: 233.1221.



(S,E)-5-(4-Phenylbut-3-en-2-yl)benzo[d][1,3]dioxole (13).

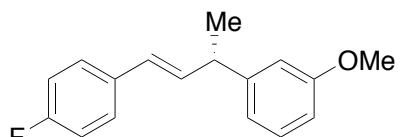
Compound **13** was prepared via the General Procedure using pivalate **1a** (prepared in 96% ee). The crude material was purified by silica gel chromatography (2% Et₂O/hexanes) to give compound **13** (run 1: 67.3 mg, 89%; run 2: 65.1 mg, 86%) as a colorless oil. The enantiomeric excess was determined to be 92% (run 1: 92% ee; run 2: 91% ee) by chiral HPLC analysis (CHIRALPAK IA, 0.6 mL/min, 0.1% *i*-PrOH/hexane, λ =254 nm); t_R (major)=13.70 min, t_R (minor)=14.68 min. $[\alpha]_D^{24} = -7.7^\circ$ (c 2.98, CHCl₃): ¹H NMR (400 MHz, CDCl₃) δ 7.40 – 7.34 (m, 2H), 7.29 (dd, J = 8.5, 6.8 Hz, 2H), 7.24 – 7.16 (m, 1H), 6.84 – 6.70 (m, 3H), 6.44 – 6.30 (m, 2H), 5.93 (s, 2H), 3.61 – 3.52 (m, 1H), 1.43 (d, J = 7.0 Hz, 3H); ¹³C NMR (101 MHz, CDCl₃) δ 147.8, 146.0, 139.8, 137.6, 135.4, 128.6, 128.5, 127.2, 126.3, 120.2, 108.3, 108.0, 101.0, 42.4, 21.5; FTIR (NaCl/thin film) 3024, 2964, 2873, 2228, 1486, 1437, 1245, 1040, 938, 693 cm⁻¹; HRMS (EI+) [M]⁺ calculated for C₁₇H₁₆O₂: 252.1150, found: 252.1156.



(S,E)-1-Chloro-4-(4-(4-methoxyphenyl)but-3-en-2-yl)benzene (14).

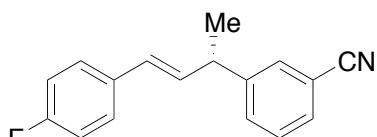
Compound **14** was prepared via the General Procedure using pivalate **1b** (prepared in 82% ee). The crude material was purified by silica gel chromatography (4% Et₂O/hexanes) to give compound **14** (run 1: 68.2 mg, 83%; run 2: 62.5 mg, 82%) as a white solid (mp 58–61 °C). The enantiomeric excess was determined to be 76% ee (run 1: 76% ee; run 2: 75% ee) by chiral HPLC analysis (CHIRALPAK IA, 0.8 mL/min, 0.1%

i-PrOH/hexane, $\lambda=254$ nm); t_R (major)=11.45 min, t_R (minor)=13.71 min. $[\alpha]_D^{24} = -8.3^\circ$ (c 1.07, CHCl_3): ^1H NMR (400 MHz, CDCl_3) δ 7.33 – 7.27 (m, 4H), 7.23 – 7.18 (m, 2H), 6.88 – 6.81 (m, 2H), 6.40 – 6.30 (m, 1H), 6.19 (dd, $J = 15.9, 6.7$ Hz, 1H), 3.80 (s, 3H), 3.66 – 3.55 (m, 1H), 1.44 (d, $J = 7.0$ Hz, 3H); ^{13}C NMR (101 MHz, CDCl_3) δ 159.1, 144.5, 132.6, 131.9, 130.3, 128.9, 128.7, 128.4, 127.4, 114.1, 55.5, 42.1, 21.4; FTIR (NaCl/thin film) 3030, 2963, 2931, 2835, 1607, 1510, 1251, 1174, 1035, 816 cm^{-1} ; HRMS (EI+) $[\text{M}]^+$ calculated for $\text{C}_{17}\text{H}_{17}\text{ClO}$: 272.0968, found: 272.0960.



(*S,E*)-1-(4-(4-Fluorophenyl)but-3-en-2-yl)-3-methoxybenzene (15).

Compound **15** was prepared via the General Procedure using pivalate **1c** (prepared in 98% ee). The crude material was purified by silica gel chromatography (2% $\text{Et}_2\text{O}/\text{hexanes}$) to give compound **15** (run 1: 69.4 mg, 90%; run 2: 68.2 mg, 89%) as a colorless oil. The enantiomeric excess was determined to be 91% (run 1: 91% ee; run 2: 91% ee) by chiral HPLC analysis (CHIRALPAK IB, 0.6 mL/min, 0.1% *i*-PrOH/hexane, $\lambda=254$ nm); t_R (major)=14.19 min, t_R (minor)=17.07 min. $[\alpha]_D^{24} = -18.9^\circ$ (c 2.80, CHCl_3): ^1H NMR (400 MHz, CDCl_3) δ 7.36 – 7.29 (m, 2H), 7.29 – 7.23 (m, 1H), 7.02 – 6.95 (m, 2H), 6.90 – 6.75 (m, 3H), 6.39 (d, $J = 15.9$ Hz, 1H), 6.29 (dd, $J = 15.9, 6.5$ Hz, 1H), 3.82 (s, 3H), 3.65 – 3.56 (m, 1H), 1.47 (d, $J = 7.0$ Hz, 3H); ^{13}C NMR (101 MHz, CDCl_3) δ 162.1 (d, $J_{\text{C}-\text{F}} = 245.9$ Hz), 159.9, 147.4, 134.93 (d, $J_{\text{C}-\text{F}} = 2.3$ Hz), 133.80 (d, $J_{\text{C}-\text{F}} = 3.2$ Hz), 129.64, 127.72 (d, $J_{\text{C}-\text{F}} = 7.8$ Hz), 127.53, 119.83, 115.47 (d, $J_{\text{C}-\text{F}} = 21.5$ Hz), 113.46, 111.34, 55.33, 42.73, 21.30; FTIR (NaCl/thin film) 2964, 2834, 1600, 1508, 1259, 1227, 1157, 779 cm^{-1} ; HRMS (EI+) $[\text{M}]^+$ calculated for $\text{C}_{17}\text{H}_{17}\text{FO}$: 256.1263, found: 256.1252.



(*S,E*)-3-(4-(4-Fluorophenyl)but-3-en-2-yl)benzonitrile (16).

Compound **16** was prepared via the General Procedure using pivalate **1c** (prepared in 98% ee). The crude material was purified by silica gel chromatography (5% $\text{Et}_2\text{O}/\text{hexanes}$) to give compound **16** (run 1: 56.4 mg, 75%; run 2: 55.2 mg, 73%) as a

colorless oil. The enantiomeric excess was determined to be 96% (run 1: 96% ee; run 2: 95% ee) by chiral HPLC analysis (CHIRALPAK IA, 1.0 mL/min, 0.1% *i*-PrOH/hexane, $\lambda=254$ nm); t_R (major)=11.64 min, t_R (minor)=12.68 min. $[\alpha]_D^{24} = -5.0^\circ$ (c 2.80, CHCl₃): ¹H NMR (400 MHz, CDCl₃) δ 7.61 – 7.47 (m, 3H), 7.42 (t, $J = 7.7$ Hz, 1H), 7.36 – 7.29 (m, 2H), 7.03 – 6.96 (m, 2H), 6.38 (dd, $J = 15.9, 1.3$ Hz, 1H), 6.22 (dd, $J = 15.9, 6.8$ Hz, 1H), 3.74 – 3.61 (m, 1H), 1.47 (d, $J = 7.0$ Hz, 3H); ¹³C NMR (101 MHz, CDCl₃) δ 162.3 (d, $J_{C-F} = 246.6$ Hz), 147.0, 133.4 (d, $J_{C-F} = 2.2$ Hz), 133.2 (d, $J_{C-F} = 3.3$ Hz), 132.1, 131.1, 130.2, 129.5, 128.7, 127.8 (d, $J_{C-F} = 8.0$ Hz), 119.2, 115.6 (d, $J_{C-F} = 21.5$ Hz), 112.7, 42.3, 21.1; FTIR (NaCl/thin film) 3038, 2968, 2228, 1600, 1507, 1481, 1227, 1157, 967, 816, 693 cm⁻¹; HRMS (EI+) [M]⁺ calculated for C₁₇H₁₄FN: 251.1110, found: 251.1115.

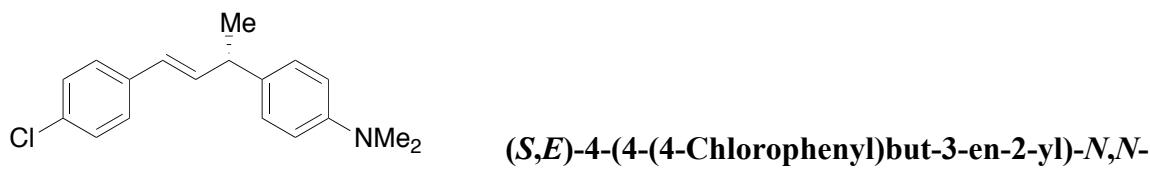
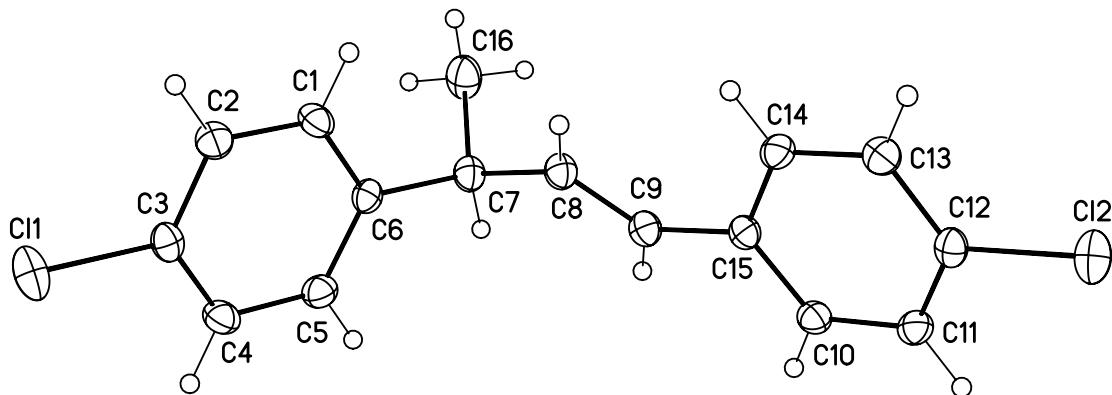


Compound **17** was prepared via the General Procedure using pivalate **1d** (prepared in 96% ee). The crude material was purified by silica gel chromatography (100% hexanes) to give compound **17** (run 1: 72.1 mg, 87%; run 2: 70.8 mg, 85%) as a white solid (mp 69–73 °C). The enantiomeric excess was determined to be 90% (run 1: 90% ee; run 2: 90% ee) by chiral HPLC analysis (CHIRALCEL OD-H, 0.4 mL/min, 100% hexanes, $\lambda=254$ nm); t_R (major)=26.72 min, t_R (minor)=24.66 min. $[\alpha]_D^{24} = -11.7^\circ$ (c 0.76, CHCl₃): ¹H NMR (400 MHz, CDCl₃) δ 7.34 – 7.22 (m, 6H), 7.23 – 7.15 (m, 2H), 6.41 – 6.24 (m, 2H), 3.61 (qd, $J = 7.0, 4.8$ Hz, 1H), 1.44 (d, $J = 7.0$ Hz, 3H); ¹³C NMR (101 MHz, CDCl₃) δ 143.9, 136.0, 135.5, 132.9, 132.2, 128.83, 128.81, 128.79, 127.9, 127.5, 42.1, 21.2. The spectral data for this compound matches that reported in the literature.¹⁰

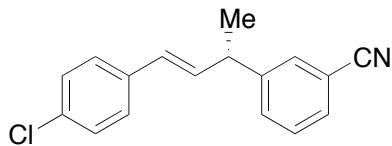
X-ray quality crystals were obtained from an Et₂O/hexanes mixture cooled to –18 °C. The crystal structure demonstrated that the absolute configuration is *S* (Fig S1).

¹⁰ Sanz, R.; Miguel, D.; Martinez, A.; Álvarez-Gutierrez, J. M.; Rodriguez, F. *Org. Lett.* **2007**, 9, 2027.

Figure S1. Molecular diagram of (*S*)-**17** with ellipsoids at 30% probability. H-atoms depicted with arbitrary radii. (CCDC-973168)

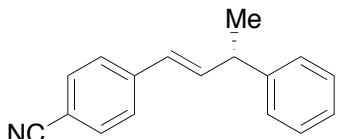


dimethylaniline (18). Compound **18** was prepared via the General Procedure using pivalate **1d** (prepared in 96% ee). The crude material was purified by silica gel chromatography (5% Et₂O/hexanes) to give compound **18** (run 1: 71.8 mg, 84%; run 2: 70.2 mg, 82%) as a colorless oil. The enantiomeric excess was determined to be 89% (run 1: 89% ee; run 2: 88% ee) by chiral HPLC analysis (CHIRALPAK IA, 0.8 mL/min, 0.5% *i*-PrOH/hexane, $\lambda=254$ nm); t_R (major)=10.18 min, t_R (minor)=8.99 min. $[\alpha]_D^{24} = -14.8^\circ$ (c 0.74, CHCl₃): ¹H NMR (400 MHz, CDCl₃) δ 7.32 – 7.20 (m, 4H), 7.19 – 7.10 (m, 2H), 6.78 – 6.69 (m, 2H), 6.40 – 6.29 (m, 2H), 3.56 (qd, $J = 7.0, 4.4$ Hz, 1H), 2.93 (s, 6H), 1.43 (d, $J = 7.0$ Hz, 3H); ¹³C NMR (101 MHz, CDCl₃) δ 149.5, 137.0, 136.4, 133.4, 132.5, 128.7, 128.0, 127.5, 126.8, 113.1, 41.7, 41.0, 21.3; FTIR (NaCl/thin film) 2962, 2926, 2799, 1895, 1646, 1521, 1489 1403, 1090, 1011, 947, 809 cm⁻¹; HRMS (EI+) [M]⁺ calculated for C₁₈H₂₀ClN: 285.1284, found: 285.1260.



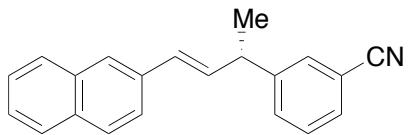
(*S,E*)-3-(4-Chlorophenyl)but-3-en-2-yl benzonitrile

(19). Compound **19** was prepared via the General Procedure using pivalate **1d** (prepared in 96% ee). The crude material was purified by silica gel chromatography (5% Et₂O/hexanes) to give compound **19** (run 1: 69.6 mg, 87%; run 2: 68.3 mg, 85%) as a colorless oil. The enantiomeric excess was determined to be 90% ee (run 1: 90% ee; run 2: 90% ee) by chiral HPLC analysis (CHIRALCEL OD-H, 0.8 mL/min, 1.0% *i*-PrOH/hexane, λ =254 nm); t_R (major)=15.06 min, t_R (minor)=17.46 min. $[\alpha]_D^{24} = -22.1^\circ$ (c 2.76, CHCl₃): ¹H NMR (400 MHz, CDCl₃) δ 7.56 – 7.48 (m, 3H), 7.45 – 7.40 (m, 1H), 7.27 (s, 4H), 6.37 (dd, J = 15.9, 1.1 Hz, 1H), 6.28 (dd, J = 15.9, 6.5 Hz, 1H), 3.71 – 3.63 (m, 1H), 1.47 (d, J = 7.0 Hz, 3H); ¹³C NMR (101 MHz, CDCl₃) δ 146.9, 135.6, 134.3, 133.2, 132.1, 131.1, 130.3, 129.5, 128.9, 128.7, 127.6, 119.2, 112.7, 42.3, 21.0; FTIR (NaCl/thin film) 3028, 2967, 2871, 2229, 1896, 1581, 1490, 1091, 968 cm⁻¹; HRMS (EI+) [M]⁺ calculated for C₁₇H₁₄ClN: 267.0815, found: 267.0807.



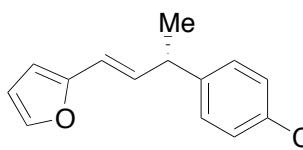
(*S,E*)-4-(3-Phenylbut-1-en-1-yl)benzonitrile (20).

Compound **20** was prepared via the General Procedure using pivalate **1e** (prepared in 80% ee). The crude material was purified by silica gel chromatography (5% Et₂O/hexanes) to give compound **20** (run 1: 60.2 mg, 86%; run 2: 58.6 mg, 84%) as a colorless oil. The enantiomeric excess was determined to be 76% (run 1: 76% ee; run 2: 76% ee) by chiral HPLC analysis (CHIRALPAK IA, 0.8 mL/min, 1.0% *i*-PrOH/hexane, λ =254 nm); t_R (major)=15.73 min, t_R (minor)=14.13 min. $[\alpha]_D^{24} = -51.7^\circ$ (c 2.28, CHCl₃): ¹H NMR (400 MHz, CDCl₃) δ 7.57 (d, J = 8.4 Hz, 2H), 7.42 (d, J = 8.3 Hz, 2H), 7.38 – 7.31 (m, 2H), 7.27 – 7.21 (m, 3H), 6.54 (dd, J = 15.9, 6.7 Hz, 1H), 6.41 (dd, J = 15.8, 1.3 Hz, 1H), 3.75 – 3.63 (m, 1H), 1.49 (d, J = 7.0 Hz, 3H); ¹³C NMR (101 MHz, CDCl₃) δ 144.8, 142.2, 139.6, 132.5, 128.8, 127.4, 127.3, 126.8, 126.7, 119.3, 110.3, 42.9, 21.1; FTIR (NaCl/thin film) 3060, 3027, 2967, 2225, 1645, 1581, 1492, 1299, 1012, 969, 819 cm⁻¹; HRMS (EI+) [M]⁺ calculated for C₁₇H₁₅N: 233.1204, found: 233.1230.



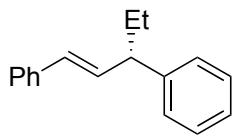
(*S,E*)-3-(4-(Naphthalen-2-yl)but-3-en-2-yl)benzonitrile (21).

Compound **21** was prepared via the General Procedure using pivalate **1f** (prepared in 96% ee). The crude material was purified by silica gel chromatography (6% Et₂O/hexanes) to give compound **21** (run 1: 66.5 mg, 78%; run 2: 64.2 mg, 76%) as a white solid (mp 90–92 °C). The enantiomeric excess was determined to be 91% (run 1: 90% ee; run 2: 91% ee) by chiral HPLC analysis (CHIRALPAK IB, 1.0 mL/min, 1.0% *i*-PrOH/hexane, λ =254 nm); t_R (major)=18.77 min, t_R (minor)=23.33 min. $[\alpha]_D^{24} = -17.8^\circ$ (c 0.56, CHCl₃): ¹H NMR (400 MHz, CDCl₃) δ 7.83 – 7.75 (m, 3H), 7.71 (s, 1H), 7.63 – 7.51 (m, 4H), 7.50 – 7.40 (m, 3H), 6.59 (d, J = 15.9 Hz, 1H), 6.44 (dd, J = 15.9, 6.8 Hz, 1H), 3.78 – 3.69 (m, 1H), 1.52 (d, J = 7.0 Hz, 3H); ¹³C NMR (101 MHz, CDCl₃) δ 147.1, 134.6, 134.1, 133.7, 133.0, 132.2, 131.2, 130.3, 130.0, 129.5, 128.4, 128.1, 127.8, 126.5, 126.2, 126.0, 123.6, 119.2, 112.7, 42.5, 21.2; FTIR (NaCl/thin film) 3057, 2967, 2927, 2228, 1627, 1597, 1481, 1267, 934, 813, 747, 694 cm⁻¹; HRMS (EI+) [M]⁺ calculated for C₂₁H₁₇N: 283.1361, found: 283.1335.

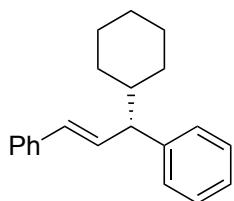


(*S,E*)-2-(3-(4-chlorophenyl)but-1-en-1-yl)furan (22).

Compound **22** was prepared via the General Procedure using pivalate **1g** (prepared in 97% ee). The crude material was purified by silica gel chromatography (100% hexanes) to give compound **22** (56.8 mg, 82%) as a colorless oil. The enantiomeric excess was determined to be 86% by chiral HPLC analysis (CHIRALCEL OD-H, 0.8 mL/min, 100% hexanes, λ =254 nm); t_R (major)=10.25 min, t_R (minor)=11.07 min. $[\alpha]_D^{24} = -48.5^\circ$ (c 1.38, CHCl₃): ¹H NMR (600 MHz, CDCl₃) δ 7.31 – 7.30 (m, 1H), 7.29 – 7.26 (m, 2H), 7.19 – 7.16 (m, 2H), 6.34 (dd, J = 3.3, 1.8 Hz, 1H), 6.30 (dd, J = 15.9, 6.7 Hz, 1H), 6.20 – 6.13 (m, 2H), 3.65 – 3.53 (m, 1H), 1.42 (d, J = 7.0 Hz, 3H); ¹³C NMR (151 MHz, CDCl₃) δ 153.0, 144.0, 141.7, 133.7, 132.1, 128.9, 128.8, 117.9, 111.4, 107.1, 41.9, 21.2; FTIR (NaCl/thin film) 2966, 2928, 2871, 1897, 1491, 1454, 1150, 1094, 1012, 963, 826, 733 cm⁻¹; HRMS (EI+) [M]⁺ calculated for C₁₄H₁₃ClO: 232.0655, found: 232.0667.



(*S,E*)-Pent-1-ene-1,3-diyldibenzene (23). Compound **23** was prepared via the General Procedure using pivalate **1h** (prepared in 90% ee). The crude material was purified by silica gel chromatography (100% hexanes) to give compound **23** (run 1: 52.7 mg, 79%; run 2: 54.7 mg, 82%) as a colorless oil. The enantiomeric excess was determined to be 82% (run 1: 82% ee; run 2: 81% ee) by chiral HPLC analysis (CHIRALCEL OD-H, 0.4 mL/min, 0.1% *i*-PrOH/hexane, $\lambda=254$ nm); $t_R(\text{major})=13.87\text{ min}$, $t_R(\text{minor})=14.76\text{ min}$. $[\alpha]_D^{24} = -15.1^\circ$ (*c* 1.45, CHCl_3): ^1H NMR (600 MHz, CDCl_3) δ 7.37 – 7.33 (m, 2H), 7.33 – 7.30 (m, 2H), 7.29 – 7.25 (m, 4H), 7.24 – 7.18 (m, 2H), 6.41 (d, $J=15.9$ Hz, 1H), 6.35 (dd, $J=15.9, 7.6$ Hz, 1H), 3.32 (q, $J=7.5$ Hz, 1H), 1.91 – 1.78 (m, 2H), 0.93 (t, $J=7.4$ Hz, 3H); ^{13}C NMR (101 MHz, $(\text{CD}_3)_2\text{CO}$) δ 145.7, 138.6, 135.1, 130.2, 129.30, 129.27, 128.4, 127.8, 126.94, 126.90, 52.0, 29.7, 12.5. The spectral data for this compound matches that reported in the literature.¹¹



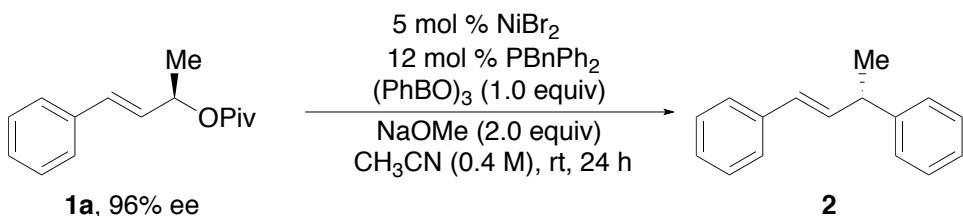
(*R,E*)-(3-Cyclohexylpent-1-en-1-yl)benzene (24). Compound **24** was prepared via the General Procedure using pivalate **1i** (prepared in 87% ee). The crude material was purified by silica gel chromatography (0.5% Et_2O /hexanes) to give compound **24** (run 1: 73.8 mg, 89%; run 2: 76.2 mg, 92%) as a colorless oil. The enantiomeric excess was determined to be 62% (run 1: 62% ee; run 2: 61% ee) by chiral HPLC analysis (CHIRALCEL OD-H, 0.8 mL/min, 100% hexanes, $\lambda=254$ nm); $t_R(\text{major})=8.46\text{ min}$, $t_R(\text{minor})=8.04\text{ min}$. $[\alpha]_D^{24} = -6.8^\circ$ (*c* 1.89, CHCl_3): ^1H NMR (400 MHz, CDCl_3) δ 7.38 – 7.25 (m, 5H), 7.24 – 7.15 (m, 5H), 6.49 – 6.19 (m, 2H), 3.08 (dt, $J=8.8, 4.5$ Hz, 1H), 2.00 – 1.88 (m, 1H), 1.79 – 1.60 (m, 4H), 1.52 – 1.42 (m, 1H), 1.31 – 1.08 (m, 3H), 1.06 – 0.78 (m, 2H); ^{13}C NMR (101 MHz, CDCl_3) δ 144.2, 137.8, 133.4,

¹¹ Sarkar, S. M.; Uozumi, Y.; Yamada, Y. M. A. *Angew. Chem. Int. Ed.* **2011**, 50, 9437.

130.3, 128.59, 128.58, 128.1, 127.1, 126.3, 126.2, 56.9, 42.8, 33.2, 31.7, 31.6, 26.8, 26.5.

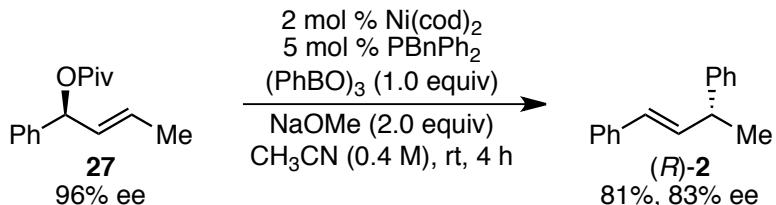
The spectral data for this compound matches that reported in the literature.¹²

Benchtop Experiment: Preparation of (*S,E*)-But-1-ene-1,3-diylbenzene (2)



This experiment, including weighing of reagents, was conducted entirely on the benchtop without the use of a glovebox. NiBr_2 (5.5 mg, 0.025 mmol, 5 mol %), benzyldiphenyl phosphine (16.6 mg, 0.06 mmol, 12 mol %) and NaOMe (54.03 mg, 1.0 mmol, 2.0 equiv) were weighed into a 1-dram vial. Pivalate **1a** (116.2 mg, 0.5 mmol, 1.0 equiv, 96% ee), phenylboroxine (156 mg, 0.5 mmol, 1.0 equiv) and zinc (32.6 mg, 0.5 mmol, 1.0 equiv) were added. Capped with a Teflon-lined cap, the vial was evacuated and backfilled with N_2 three times. Acetonitrile (1.25 mL, 0.4 M) was then added. The mixture was stirred for 24 h at room temperature. The reaction mixture was then diluted with Et_2O (3.0 mL) and filtered through a plug of silica gel, which was rinsed with Et_2O (15 mL). The filtrate was concentrated and purified by silica gel chromatography (100% hexanes) to give compound **2** (54.4 mg, 53%) as a colorless oil. The enantiomeric excess of the product was determined to be 93% by chiral HPLC analysis.

Cross Coupling of Pivalate 27

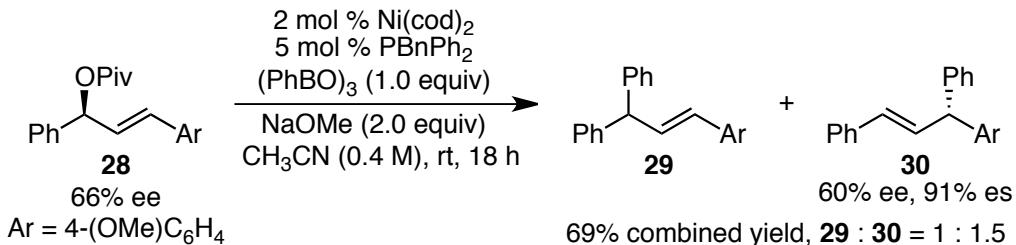


The arylation of **27** was performed according to the General Procedure, but on a 0.2-mmol scale. In a N_2 -atmosphere glovebox, $\text{Ni}(\text{cod})_2$ (1.1 mg, 0.04 mmol, 2 mol %),

¹² Miranda, M. A.; Pérez-Prieto, J.; Font-Sanchis, E.; Kónya, K.; Scaiano, J. C. *J. Org. Chem.* **1997**, 62, 5713.

benzyl diphenyl phosphine (2.8 mg, 0.010 mmol, 5 mol %) and NaOMe (21.7 mg, 0.40 mmol, 2.0 equiv) were weighed into a 1-dram vial. Allylic pivalate **27** (46.5 mg, 96% ee, 0.20 mmol, 1.0 equiv) and phenylboroxine (62.2 mg, 0.20 mmol, 1.0 equiv) were added, followed by acetonitrile (0.5 mL, 0.4 M). The vial was capped with a Teflon-lined cap and removed from the glovebox. The mixture was stirred at room temperature for 4 h. The reaction mixture was then diluted with Et₂O (1.5 mL) and filtered through a plug of silica gel, which was rinsed with Et₂O (10 mL). The filtrate was concentrated and purified by silica gel chromatography (1% Et₂O/hexanes) to afford (*R*)-**2** (33.8 mg, 81%) as a colorless oil. The absolute configuration was determined by comparison to the HPLC trace and optical rotation of (*S*)-**2**. The enantiomeric excess was determined by chiral HPLC analysis (CHIRALCEL OD-H, 1.0 mL/min, 0.1% *i*-PrOH/hexane $\lambda=254$ nm); $t_R(\text{major})=6.59$ min, $t_R(\text{minor})=6.18$ min. $[\alpha]_D^{24}=+12.9^\circ$ (c 1.39, CHCl₃).

Cross Coupling of Pivalate **28**



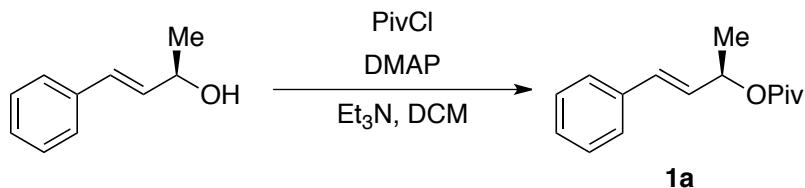
The arylation of **28** was performed according to the General Procedure, but on a 0.2-mmol scale. In a N₂-atmosphere glovebox, Ni(cod)₂ (1.1 mg, 0.04 mmol, 2 mol %), benzyl diphenyl phosphine (2.8 mg, 0.010 mmol, 5 mol %) and NaOMe (21.7 mg, 0.40 mmol, 2.0 equiv) were weighed into a 1-dram vial. Allylic pivalate **28** (64.9 mg, 66% ee, 0.20 mmol, 1.0 equiv) and phenylboroxine (62.2 mg, 0.20 mmol, 1.0 equiv) were added, followed by acetonitrile (0.5 mL, 0.4 M). The vial was capped with a Teflon-lined cap and removed from the glovebox. The mixture was stirred at room temperature for 18 h. The reaction mixture was then diluted with Et₂O (1.5 mL) and filtered through a plug of silica gel, which was rinsed with Et₂O (10 mL). The filtrate was concentrated and purified by silica gel chromatography (5% Et₂O/hexanes) to afford a 1: 1.5 mixture of **29**¹³ and **30**¹⁴ (41.7 mg, 69% combined yield) as colorless oil. The enantiomeric excess of

¹³ Baba, A.; Kajioka, M.; Nishimoto, Y.; Saito, T.; Yasuda, M. *Chem. Comm.*, **2008**, 47, 6396.

30 was determined by chiral HPLC analysis (CHIRALCEL OD-H, 0.4 mL/min, 0.1% *i*-PrOH/hexane λ =254 nm); t_R (major)=43.72 min, t_R (minor)=37.96 min. $[\alpha]_D^{24} = +22.6^\circ$ (c 0.66, CHCl_3). The spectral data of each compound in the mixture matches that reported in the literature.¹³⁻¹⁴

¹⁴ Ji, J.-X.; Li, M.-M.; Wei, W.; Yang, Y.-R.; Yue, H.-L. *Adv. Synth. Catal.*, **2011**, 1 353 , 3139.

General Procedure for the Preparation of Allylic Pivalates: Preparation of (*R*)-4-Phenylbutan-2-yl Pivalate (**1a**)

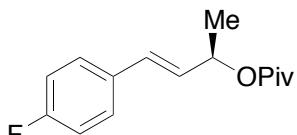


(*R*)-4-Phenyl-3-buten-2-ol (1.80 g, 12.2 mmol, 1.0 equiv, 96% ee) and DMAP (149.5 mg, 1.22 mmol, 0.10 equiv) were dissolved in CH₂Cl₂ (20 mL, 0.61 M). Et₃N (3.5 mL, 24.5 mmol, 2.0 equiv) and pivaloyl chloride (1.80 mL, 14.7 mmol, 1.2 equiv) were then added. The reaction mixture was then stirred for 15 h at room temperature, before H₂O (20 mL) was added. The organic layer was extracted with CH₂Cl₂ (2 x 40 mL). The combined organic layers were washed with aq. KOH (2.0 M, 40 mL), dried (MgSO₄), filtered and concentrated. The resulting residue was purified by silica gel chromatography (5% Et₂O/hexanes) to give compound **1a** (2.20 g, 77%) as a white solid (mp 47–50 °C). The enantiomeric excess was determined to be 96% by chiral HPLC analysis (CHIRALPAK IC, 0.8 mL/min, 1.0% *i*-PrOH/hexane, $\lambda=254$ nm); t_R (major)=6.80 min, t_R (minor)=8.01 min. $[\alpha]_D^{24} = +106.1^\circ$ (c 2.78, CHCl₃): ¹H NMR (400 MHz, CDCl₃) δ 7.40 – 7.35 (m, 2H), 7.34 – 7.28 (m, 2H), 7.25 – 7.21 (m, 1H), 6.59 (d, J = 16.0 Hz, 1H), 6.19 (dd, J = 16.0, 6.5 Hz, 1H), 5.54 – 5.46 (m, 1H), 1.39 (d, J = 6.5 Hz, 3H), 1.21 (s, 9H); ¹³C NMR (151 MHz, CDCl₃) δ 178.0, 136.6, 131.2, 129.3, 128.73, 128.70, 126.7, 70.7, 38.9, 27.3, 20.5; FTIR (NaCl/thin film) 2977, 2932, 1727, 1479, 1279, 1159, 1041, 965, 747, 692 cm⁻¹; HRMS (EI) [M]⁺ calculated for C₁₅H₂₀O₂: 232.1463, found: 232.1479.

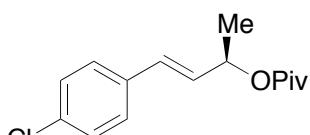


Prepared according to the General Procedure on a 2.48 mmol scale to give **1b** (500 mg, 77%) as a white solid (mp 76–80 °C). The enantiomeric excess was determined to be 82% by chiral HPLC analysis (CHIRALPAK IC, 0.7 mL/min, 2.0% *i*-PrOH/hexane,

$\lambda=254$ nm); t_R (major)=9.23 min, t_R (minor)=10.47 min. $[\alpha]_D^{24} = +116.1^\circ$ (c 1.08, CHCl_3): ^1H NMR (400 MHz, CDCl_3) δ 7.36 – 7.27 (m, 2H), 6.89 – 6.81 (m, 2H), 6.53 (d, $J = 15.9$ Hz, 1H), 6.05 (dd, $J = 15.9, 6.7$ Hz, 1H), 5.52 – 5.43 (m, 1H), 3.81 (s, 3H), 1.37 (d, $J = 6.5$ Hz, 3H), 1.20 (s, 9H); ^{13}C NMR (101 MHz, CDCl_3) δ 178.0, 159.5, 130.8, 129.4, 127.9, 127.1, 114.1, 70.9, 55.5, 38.9, 27.3, 20.6; FTIR (NaCl/thin film) 2830, 2821, 1714, 1573, 1380, 1080, 754 cm^{-1} ; HRMS (EI+) $[\text{M}]^+$ calculated for $\text{C}_{16}\text{H}_{22}\text{O}_3$: 262.1569, found: 262.1569.



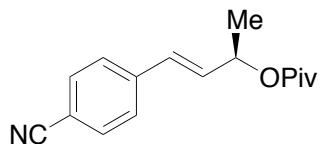
(*R,E*)-4-(4-Fluorophenyl)but-3-en-2-yl pivalate (1c). Prepared according to the General Procedure on a 3.32 mmol scale to give **1c** (660 mg, 79%) as a colorless oil. The enantiomeric excess was determined to be 98% by chiral HPLC analysis (CHIRALPAK IA, 1.0 mL/min, 2.0% *i*-PrOH/hexane, $\lambda=254$ nm); t_R (major)=4.66 min, t_R (minor)=5.49 min. $[\alpha]_D^{24} = +134.2^\circ$ (c 0.92, CHCl_3): ^1H NMR (400 MHz, CDCl_3) δ 7.38 – 7.30 (m, 2H), 7.05 – 6.96 (m, 2H), 6.55 (d, $J = 16.0$ Hz, 1H), 6.10 (dd, $J = 16.0, 6.5$ Hz, 1H), 5.53 – 5.44 (m, 1H), 1.38 (d, $J = 6.5$ Hz, 3H), 1.24 – 1.19 (m, 9H); ^{13}C NMR (101 MHz, CDCl_3) δ 177.9, 162.6 (d, $J = 247.1$ Hz), 132.76 (d, $J = 3.3$ Hz), 130.05, 129.02 (d, $J = 2.3$ Hz), 128.2 (d, $J = 8.0$ Hz), 115.6 (d, $J = 21.6$ Hz), 70.6, 38.9, 27.3, 20.5; FTIR (NaCl/thin film) 2977, 2935, 2873, 1727, 1509, 1231, 1157, 1041 cm^{-1} ; HRMS (EI+) $[\text{M}]^+$ calculated for $\text{C}_{15}\text{H}_{19}\text{FO}_2$: 250.1369, found: 250.1388.



(*R,E*)-4-(4-Chlorophenyl)but-3-en-2-yl pivalate (1d).

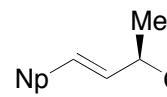
Prepared according to the General Procedure on a 5.40 mmol scale to give **1d** (1.3 g, 92%) as a white solid (mp 72–74 °C). The enantiomeric excess was determined to be 96% by chiral HPLC analysis (CHIRALPAK IA, 0.8 mL/min, 1.0% *i*-PrOH/hexane, $\lambda=254$ nm); t_R (major)=6.09 min, t_R (minor)=7.06 min. $[\alpha]_D^{24} = +150.8^\circ$ (c 0.68, CHCl_3): ^1H NMR (400 MHz, CDCl_3) δ 7.34 – 7.24 (m, 4H), 6.53 (dd, $J = 15.9, 1.2$ Hz, 1H), 6.16 (dd, $J = 16.0, 6.4$ Hz, 1H), 5.53 – 5.44 (m, 1H), 1.38 (d, $J = 6.5$ Hz, 3H), 1.21 (s, 9H); ^{13}C NMR (101 MHz, CDCl_3) δ 177.9, 135.1, 133.6, 130.0, 129.9, 128.9, 127.9, 70.5, 39.0,

27.3, 20.4; FTIR (NaCl/thin film) 2923, 2840, 1824, 1713, 1569, 1417, 1380, 1340, 1080 cm⁻¹; HRMS (EI+) [M]⁺ calculated for C₁₅H₁₉ClO₂: 266.1074, found: 266.1060.

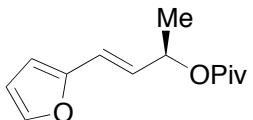


(R,E)-4-(4-Cyanophenyl)but-3-en-2-yl pivalate (1e).

Prepared according to the General Procedure on a 2.49 mmol scale to give **1e** (425 mg, 66%) as a white solid (mp 84–86 °C). The enantiomeric excess was determined to be 80% by chiral HPLC analysis (CHIRALPAK IC, 1.0 mL/min, 2.0% *i*-PrOH/hexane, λ =254 nm); t_R (major)=24.46 min, t_R (minor)=32.11 min. $[\alpha]_D^{24} = +115.0^\circ$ (c 1.00, CHCl₃): ¹H NMR (400 MHz, CDCl₃) δ 7.64 – 7.56 (m, 2H), 7.49 – 7.42 (m, 2H), 6.58 (d, J = 16.0 Hz, 1H), 6.31 (dd, J = 16.0, 6.1 Hz, 1H), 5.55 – 5.47 (m, 1H), 1.40 (d, J = 6.5 Hz, 3H), 1.22 (s, 9H); ¹³C NMR (101 MHz, CDCl₃) δ 177.8, 141.1, 133.3, 132.7, 129.3, 127.2, 119.1, 111.2, 70.1, 39.0, 27.3, 20.3; FTIR (NaCl/thin film) 2978, 2935, 2872, 2227, 1717, 1604, 1480, 1282, 1173, 1039, 968 cm⁻¹; HRMS (EI+) [M]⁺ calculated for C₁₆H₁₉NO₂: 257.1416, found: 257.1408.

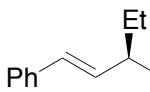


(R,E)-4-(Naphthalen-2-yl)but-3-en-2-yl pivalate (1f). Prepared according to the General Procedure on a 2.36 mmol scale to give **1f** (515 mg, 77%) as a white solid (mp 103–105 °C). The enantiomeric excess was determined to be 96% by chiral HPLC analysis (CHIRALPAK IA, 1.0 mL/min, 1.0% *i*-PrOH/hexane, λ =254 nm); t_R (major)=6.12 min, t_R (minor)=6.92 min. $[\alpha]_D^{24} = +141.9^\circ$ (c 0.82, CHCl₃): ¹H NMR (400 MHz, CDCl₃) δ 7.84 – 7.71 (m, 4H), 7.59 (dd, J = 8.6, 1.8 Hz, 1H), 7.51 – 7.40 (m, 2H), 6.75 (d, J = 15.7 Hz, 1H), 6.32 (dd, J = 16.0, 6.4 Hz, 1H), 5.60 – 5.52 (m, 1H), 1.43 (d, J = 6.5 Hz, 3H), 1.24 (s, 9H); ¹³C NMR (101 MHz, CDCl₃) δ 178.0, 134.1, 133.7, 133.2, 131.2, 129.6, 128.3, 128.1, 127.8, 126.8, 126.4, 126.1, 123.7, 70.7, 39.0, 27.3, 20.5; FTIR (NaCl/thin film) 2974, 1718, 1653, 1368, 1278, 1164, 1035, 972, 743 cm⁻¹; HRMS (EI+) [M]⁺ calculated for C₁₉H₂₂O₂: 282.1620, found: 282.1634.

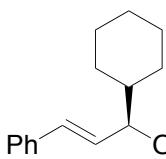


(*R,E*)-4-(Furan-2-yl)but-3-en-2-yl pivalate (1g). Prepared

according to the General Procedure on a 1.66 mmol scale to give **1g** (320 mg, 74%) as a colorless oil. The enantiomeric excess was determined to be 97% by chiral HPLC analysis (CHIRALPAK IA, 0.6 mL/min, 0.1% *i*-PrOH/hexane, $\lambda=254$ nm); t_R (major)=8.98 min, t_R (minor)=9.95 min. $[\alpha]_D^{24} = +124.1^\circ$ (*c* 1.25, CHCl_3): ^1H NMR (400 MHz, CDCl_3) δ 7.37 – 7.32 (m, 1H), 6.44 – 6.34 (m, 2H), 6.25 (d, $J = 3.3$ Hz, 1H), 6.12 (dd, $J = 15.9, 6.4$ Hz, 1H), 5.50 – 5.42 (m, 1H), 1.36 (d, $J = 6.4$ Hz, 3H), 1.21 (s, 9H); ^{13}C NMR (151 MHz, CDCl_3) δ 177.9, 152.3, 142.3, 127.9, 119.5, 111.5, 108.7, 70.3, 39.0, 27.3, 20.4; FTIR (NaCl/thin film) 2976, 2933, 2872, 1727, 1480, 1280, 1161, 1040, 959, 734 cm^{-1} ; HRMS (EI+) [M] $^+$ calculated for $\text{C}_{13}\text{H}_{18}\text{O}_3$: 222.1256, found: 222.1262.



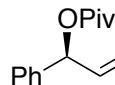
(*R,E*)-1-Phenylpent-1-en-3-yl pivalate (1h). Prepared according to the General Procedure on a 1.50 mmol scale to give **1h** (295 mg, 70%) as a pale yellow oil. The enantiomeric excess was determined to be 90% by chiral HPLC analysis (CHIRALCEL OD-H, 0.8 mL/min, 1.0% *i*-PrOH/hexane, $\lambda=254$ nm); t_R (major)=7.37min, t_R (minor)=5.84min. $[\alpha]_D^{24} = +87.1^\circ$ (*c* 2.41, CHCl_3): ^1H NMR (600 MHz, CDCl_3) δ 7.40 – 7.35 (m, 2H), 7.31 (t, $J = 7.6$ Hz, 2H), 7.27 – 7.21 (m, 1H), 6.59 (d, $J = 15.9$ Hz, 1H), 6.13 (dd, $J = 16.0, 6.9$ Hz, 1H), 5.37 – 5.30 (m, 1H), 1.78 – 1.69 (m, 2H), 1.23 (s, 9H), 0.95 (t, $J = 7.4$ Hz, 3H); ^{13}C NMR (151 MHz, CDCl_3) δ 178.0, 136.7, 132.1, 128.7, 128.0, 127.9, 126.7, 75.5, 39.10, 27.9, 27.4, 9.7. The spectral data for this compound matches that reported in the literature.¹⁵



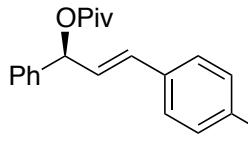
(*R,E*)-1-Cyclohexyl-3-phenylallyl pivalate (1i). Prepared according to the General Procedure on a 1.50 mmol scale to give **1i** (295 mg, 70%) as a white solid

¹⁵ Phipps, R. J.; McMurray, L.; Ritter, S.; Duong, H. A.; Gaunt, M. J. *J. Am. Chem. Soc.* **2012**, 134, 10773.

(mp 69–71 °C). The enantiomeric excess was determined to be 87% by chiral HPLC analysis (CHIRALCEL OD-H, 0.8 mL/min, 1.0% i-PrOH/hexane, $\lambda=254$ nm); t_R (major)=6.89 min, t_R (minor)=5.32 min. $[\alpha]_D^{24} = +29.2^\circ$ (c 0.82, CHCl_3): ^1H NMR (400 MHz, CDCl_3) δ 7.41 – 7.19 (m, 5H), 6.55 (d, $J = 15.9$ Hz, 1H), 6.12 (dd, $J = 15.9, 7.4$ Hz, 1H), 5.23 – 5.14 (m, 1H), 1.85 – 1.59 (m, 6H), 1.22 (s, 10H), 1.22 – 0.99 (m, 4H); ^{13}C NMR (101 MHz, CDCl_3) δ 177.9, 136.8, 132.8, 128.7, 127.9, 127.0, 126.7, 78.4, 42.3, 39.2, 29.07, 28.6, 27.4, 26.6, 26.2, 26.1; FTIR (NaCl/thin film) 3026, 2929, 2854, 1728, 1479, 1280, 1158, 966, 748 cm^{-1} ; HRMS (EI+) [M] $^+$ calculated for $\text{C}_{20}\text{H}_{28}\text{O}_2$: 300.2089, found: 300.2066.



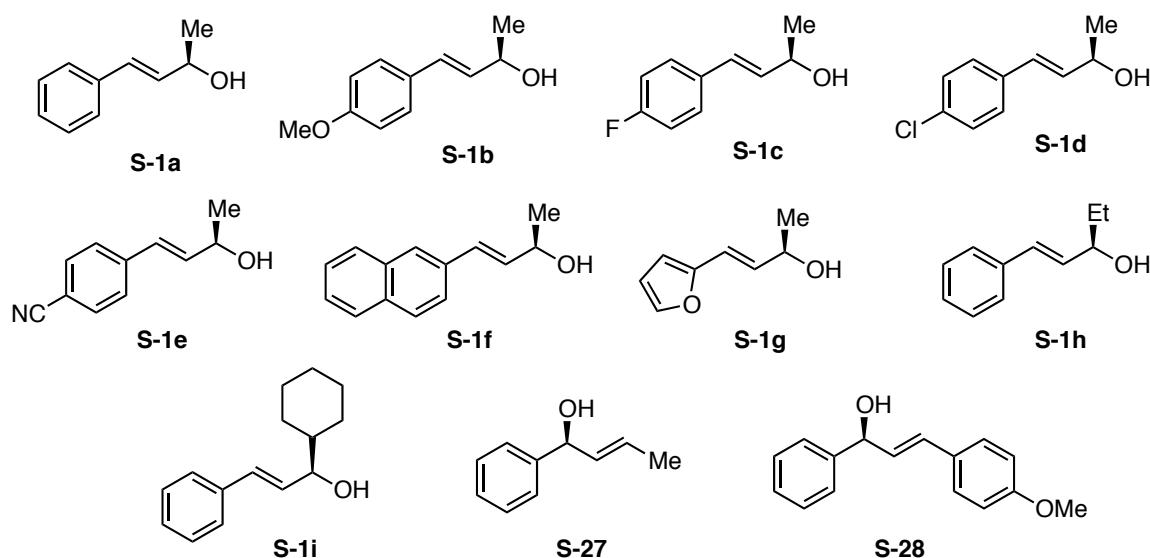
(S,E)-1-Phenylbut-2-en-1-yl pivalate (27). Prepared according to the General Procedure on a 0.6 mmol scale to give **27** (103.5 mg, 78%) as a colorless oil. The enantiomeric excess was assumed to be unchanged from allylic alcohol **S-27** (96% ee). $[\alpha]_D^{24} = -10.2^\circ$ (c 2.15, CHCl_3): ^1H NMR (600 MHz, CDCl_3) δ 7.39 – 7.26 (m, 5H), 6.17 (d, $J = 6.9$ Hz, 1H), 5.80 – 5.70 (m, 1H), 5.67 – 5.60 (m, 1H), 1.71 (d, $J = 6.6$ Hz, 3H), 1.22 (s, 9H); ^{13}C NMR (151 MHz, CDCl_3) δ 177.5, 140.3, 130.0, 129.2, 128.6, 127.8, 126.7, 76.0, 39.0, 27.3, 17.9; FTIR (NaCl/thin film) 2972, 1732, 1480, 1455, 1278, 1149, 963 cm^{-1} ; HRMS (EI+) [M] $^+$ calculated for $\text{C}_{21}\text{H}_{24}\text{O}_3$: 232.1463, found: 232.1470.



(S,E)-3-(4-Methoxyphenyl)-1-phenylallyl pivalate (28). Compound **28** was prepared via the General Procedure a 1.0 mmol scale. The crude material was purified by silica gel chromatography (5–7.5% Et_2O /hexanes, silica gel pre-saturated with Et_3N) to give **28** (167 mg, 54%) as a colorless oil. The enantiomeric excess was determined to be 66% by chiral HPLC analysis (CHIRALPAK IA, 1.0 mL/min, 5% i-PrOH/hexane, $\lambda=210$ nm); t_R (major)=7.21 min, t_R (minor)=6.26 min. $[\alpha]_D^{24} = +4.1^\circ$ (c 4.83, CHCl_3): ^1H NMR (400 MHz, CDCl_3) δ 7.40 – 7.29 (m, 7H), 6.84 (d, $J = 8.8$ Hz, 2H), 6.46 (d, $J = 15.6$ Hz, 1H), 6.37 (d, $J = 7.2$ Hz, 1H), 6.17 (dd, $J = 15.6, 6.8$ Hz, 1H), 3.80 (s, 3H), 1.24 (s, 9H); ^{13}C NMR (101 MHz, CDCl_3) δ 177.4, 159.5, 139.9, 131.9,

129.0, 128.6, 128.0, 127.9, 126.7, 125.6, 113.9, 76.0, 55.3, 38.9, 27.1; FTIR (NaCl/thin film) 2971, 2933, 1732, 1608, 1512, 1277, 1252, 1149, 699 cm^{-1} ; HRMS (LIFDI) [M]⁺ calculated for C₂₁H₂₄O₃: 324.1720, found: 324.1725.

Preparation of Enantioenriched Allylic Alcohols

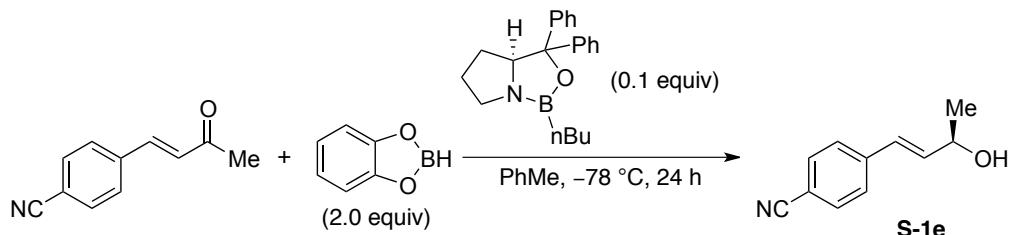


Enantioenriched allylic alcohols (**S-1a** to **S-1i**) were prepared via CBS reduction of the corresponding ketones according to the procedure reported in the literature.¹⁶ These reductions are known to give the stereochemistry shown. A detailed example of the synthesis of **S-1e** obtained by this method is given below. **S-27** and **S-28** were prepared according to a reported procedure.¹⁷

¹⁶ (a) Corey, E. J.; Bakshi, R. K. *Tetrahedron Lett.* **1990**, 31, 611. (b) Corey, E. J.; Helal, C. J. *Tetrahedron Lett.* **1995**, 36, 9153.

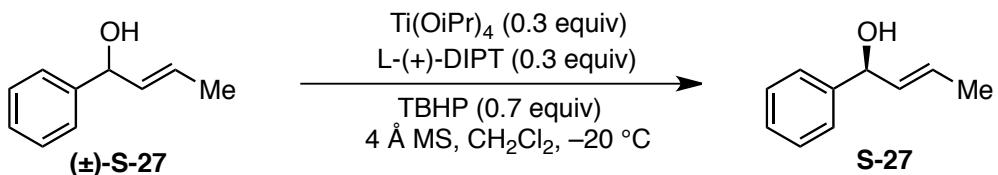
¹⁷ Hidaka, I.; Michiko, S.; Kei, T.; Masatoshi, K.; Kentaro, Y. *Chem. Eur. J.* **2009**, 15, 4663.

**General Procedure for the Preparation of Allylic Alcohols S-1a to S-1i:
Preparation of (R)-4-(4-Cyanophenyl)-3-butene-2-ol (S-1e)**

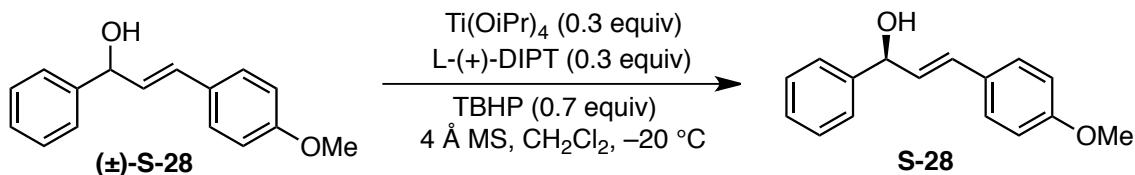


In an oven-dried, 100-mL round-bottomed flask, (E)-4-(4-cyanophenyl)-3-butene-2-one (1.08 g, 5.84 mmol, 1.0 equiv) was dissolved in 10 mL PhMe. Under N₂, (S)-(-)-2-butyl-CBS-oxazaborolidine (0.58 mL, 0.58 mmol, 1.0 M in PhMe, 0.1 equiv) was added. After stirring at room temperature for 15 min, the mixture was cooled to -78 °C, and catecholborane (1.25 mL, 11.68 mmol, 2.0 equiv) was added slowly. The mixture was stirred at -78 °C for additional 24 h. Sat. NaHCO₃ (10 mL) was added, and the crude product was extracted with EtOAc (3 x 20 mL). The combined organic layers were washed with aq. NaOH (1.5 M) until the color of the solution was light yellow, indicating the complete removal of residual catecholborane. The organic layers were then treated with sat. NaCl, dried (MgSO₄), filtered and concentrated. The resulting residue was purified by silica gel chromatography (50% Et₂O/hexanes) to give compound S-1e (920 mg, 92%) as a yellow oil. The enantiomeric excess was determined to be 80% by chiral HPLC analysis (CHIRALPAK IA, 1.0 mL/min, 6.0% i-PrOH/hexane, $\lambda = 254$ nm); $t_R(\text{major})=21.77$ min, $t_R(\text{minor})=19.56$ min. $[\alpha]_D^{24} = -9.2^\circ$ (c 2.39, CHCl₃): ¹H NMR (600 MHz, CDCl₃) δ 7.60 (d, J = 8.2 Hz, 2H), 7.45 (d, J = 8.2 Hz, 2H), 6.60 (d, J = 16.0 Hz, 1H), 6.39 (dd, J = 15.9, 5.8 Hz, 1H), 4.57 – 4.50 (m, 1H), 1.65 (brs, 1H), 1.39 (d, J = 6.4 Hz, 3H); ¹³C NMR (151 MHz, CDCl₃) δ 141.5, 137.6, 132.57, 127.6, 127.1, 119.1, 111.0, 68.6, 23.6; FTIR (NaCl/thin film) 3421(brs), 2973, 2226, 1604, 1504, 1142, 971 cm⁻¹; HRMS (EI) [M]⁺ calculated for C₁₁H₁₁NO: 173.0841, found: 173.0867.

Preparation of Allylic Alcohols S-27 and S-28



In an oven-dried, 100-mL round-bottomed flask, L-(+)-DIPT (0.74 mL, 3.54 mmol, 0.3 equiv) was added to a suspension of (*E*)-1-phenyl-2-buten-1-ol (1.76g, 11.8 mmol, 1.0 equiv), 4 Å MS (0.85 g, crushed before use), and CH₂Cl₂ (47 mL). The suspension was then cooled to -20 °C, and Ti(OⁱPr)₄ (1.06 mL, 3.54 mmol, 0.3 equiv) and TBHP (1.5 mL, 8.25 mmol, 5.5 M in decane, 0.7 equiv) were added. The mixture was stirred for 3 h at -20 °C. FeSO₄·7H₂O (6.5 g) and H₂O (40 mL) were then added, followed by tartaric acid (2.2 g), H₂O (20 mL), and aq. HCl (1.0 M, 30 mL) to dissolve the precipitate. The layers were separated. The organic layer was then washed with sat. NaCl, dried (Na₂SO₄), filtered, and concentrated. The resulting residue was purified by silica gel chromatography (15% Et₂O/hexanes) to give compound **S-27** (240 mg, 27%) as colorless oil. The enantiomeric excess was determined to be 96% by chiral GC analysis (Agilent 19091C-133/HP-225, 50% cyanopropyl, 50% phenylmethyl polysiloxane capillary column, initial temperature of 40 °C, ramp 1.0 °C/min to 180 °C, hold for 10 min); *t*_R(major)=105.67 min, *t*_R(minor)=106.02 min. [α]_D²⁴ = +16.2° (c 3.29, CHCl₃). The absolute configuration was determined to be *S* by comparison of the optical rotation to that reported in the literature.¹⁸

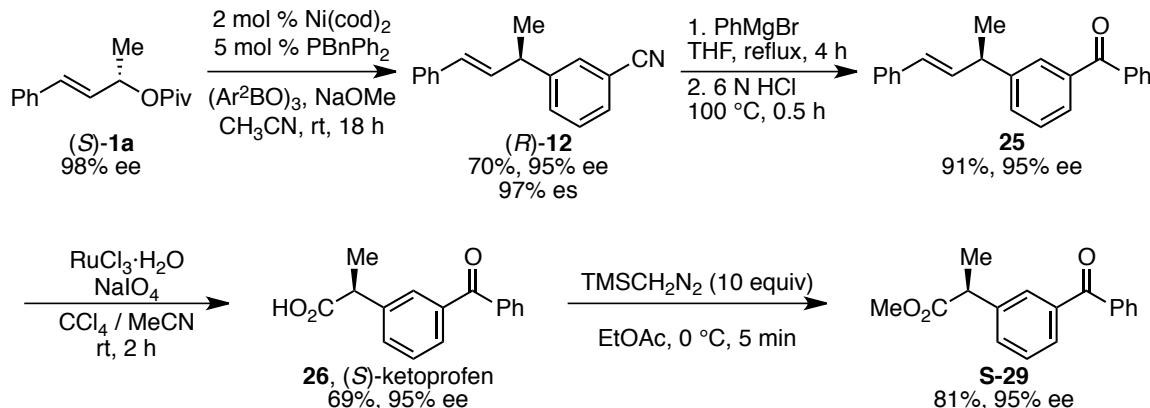


S-28 was prepared on a 8.0-mmol scale from (±)-S-28 according to the procedure described for **S-27** above. **S-28** (662 mg, 70%) was obtained as a sticky yellow oil from the crude residue by silica gel chromatography (33% Et₂O/hexanes). The enantiomeric

¹⁸ Hayashi, T.; Yamamoto, A.; Hagishita, T. *J. Org. Chem.* **1986**, 51, 723.

excess was determined to be 67% by chiral HPLC analysis (CHIRALPAK IA, 1.0 mL/min, 8.0% *i*-PrOH/hexane, $\lambda = 254$ nm); $t_R(\text{major})=20.38$ min, $t_R(\text{minor})=17.82$ min. $[\alpha]_D^{24} = -5.1^\circ$ (*c* 4.26, CHCl₃). The spectral data for this compound matches that reported in the literature.¹⁹

Preparation of (*S*)-Ketoprofen



Compound *(R)*-12 was prepared in 95% ee via the General Procedure using pivalate *(S)*-1a (prepared in 98% ee), as described above.

The conversion of *(R)*-12 to **25** followed a procedure adapted from the literature.²⁰ In a 25-mL, three-neck round-bottomed flask equipped with a reflux condenser, **12** (180 mg, 0.77 mmol, 95% ee, 1.0 equiv) and THF (1.6 mL) were combined. PhMgBr (1.0 M in THF, 3.9 mL, 3.9 mmol, 5.0 equiv) was added slowly at room temperature. Upon completion of addition, the mixture was refluxed for 4 h. After cooling to room temperature, HCl (6 N, 2 mL) was added. The mixture was heated at 100 °C for 30 min. After cooling to room temperature, H₂O (20 mL) was added. The crude product was extracted with Et₂O (3 x 10 mL), dried (MgSO₄), filtered, and concentrated. The resulting material was purified by silica gel chromatography (5% Et₂O/hexanes) to give **25** (219 mg, 91%) as a light yellow oil. The enantiomeric excess was determined to be 95% by chiral HPLC analysis (CHIRALPAK IA, 1.0 mL/min, 1.0% *i*-PrOH/hexane, $\lambda=220$ nm); $t_R(\text{major})=13.43$ min, $t_R(\text{minor})=14.41$ min. $[\alpha]_D^{24} = +35.2^\circ$ (*c* 1.7, CHCl₃):

¹⁹ Xu, W.; Zhou, Y.; Wang, R.; Wu, G.; Chen, P. *Org. Biomol. Chem.* **2012**, 10, 367.

²⁰ Riva, E.; Gagliardi, M.; Martinelli, M.; Passarella, D.; Vigo, D.; Rencurosi, A. *Tetrahedron* **2010**, 66, 3242.

¹H NMR (600 MHz, CDCl₃) δ 7.84 – 7.79 (m, 2H), 7.75 (s, 1H), 7.64 – 7.61 (m, 1H), 7.60 – 7.56 (m, 1H), 7.53 – 7.45 (m, 3H), 7.42 (t, *J* = 7.6 Hz, 1H), 7.38 – 7.34 (m, 2H), 7.32 – 7.27 (m, 2H), 7.23 – 7.18 (m, 1H), 6.48 – 6.34 (m, 2H), 3.76 – 3.69 (m, 1H), 1.50 (d, *J* = 7.0 Hz, 3H); ¹³C NMR (151 MHz, CDCl₃) δ 197.0, 146.2, 138.0, 137.8, 137.5, 134.7, 132.5, 131.6, 130.2, 129.2, 129.0, 128.7, 128.5, 128.39, 128.38, 127.4, 126.3, 42.7, 21.3; FTIR (NaCl/thin film) 3024, 2965, 1658, 1597, 1493, 1447, 1317, 1178, 966 cm⁻¹; HRMS (EI) [M]⁺ calculated for C₂₃H₂₀O: 312.1514, found: 312.1534.

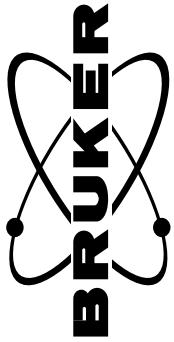
The conversion of **25** to **26** followed a procedure adapted from the literature.²¹ A suspension of RuCl₃·H₂O (7.3 mg, 0.035 mmol, 0.05 equiv) in H₂O (7.3 mL) was added to a solution of **25** (219 mg, 0.7 mmol, 1.0 equiv), NaIO₄ (749 mg, 3.5 mmol, 5.0 equiv), and CCl₄/MeCN (v/v = 1/1, 9.4 mL) at 0 °C. The mixture was then stirred at room temperature for 2 h. Et₂O (30 mL) was added, and the crude product was then extracted with sat. NaHCO₃ (5 x 15 mL). The combined aqueous layers were then cooled to 0 °C, and HCl (6 N) was carefully added until the pH ≤ 1. The crude product was then extracted from aqueous layer with CH₂Cl₂ (5 x 15 mL). The combined organic layers were dried (Na₂SO₄), filtered, and concentrated. The crude material was purified by silica gel chromatography (0–3% MeOH/CH₂Cl₂) to give (*S*)-ketoprofen (**26**) as an off-white solid (124 mg, 69%). The spectral data of **26** matches that reported in the literature.²²

The enantiomeric excess of **26** was determined to be 95% based on the corresponding methyl ester **S-29**, which was prepared via a reported procedure on a 0.17 mmol scale.^{22a} The spectral data of **S-29** matches that reported in the literature.²² The enantiomeric excess of **S-29** was determined to be 95% by chiral HPLC analysis (CHIRALPAK IB, 0.8 mL/min, 0.5% EtOH/hexane, λ=254 nm); *t*_R(major)=39.65 min, *t*_R(minor)=37.39 min. [α]_D²⁴ = +43.5° (c 0.80, CHCl₃). The absolute configuration of **S-29** was determined to be *S* by comparing the optical rotation with reported data.²³

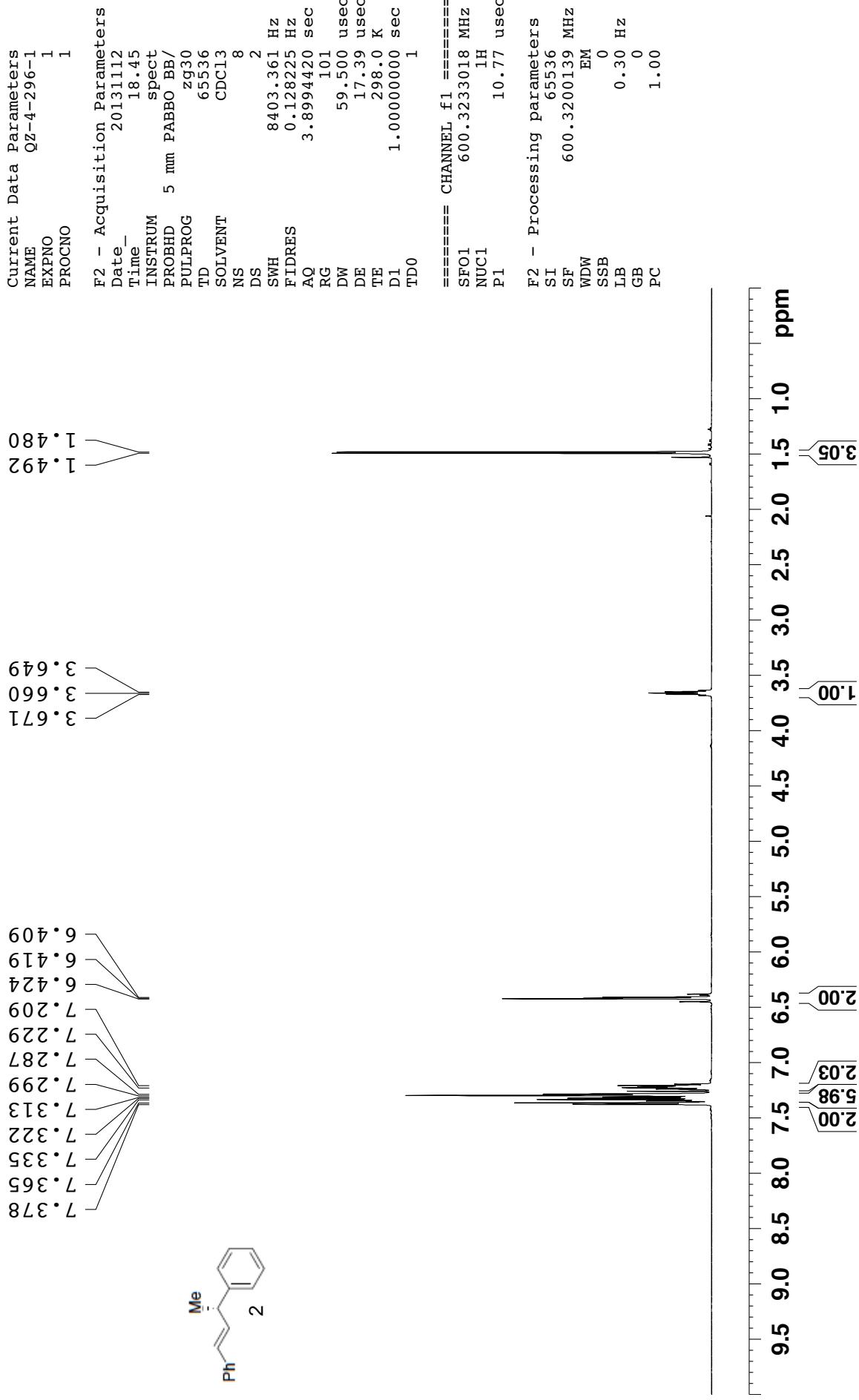
²¹ Norinder, J.; Bogár, K.; Kanupp, L.; Bäckvall, J.-E. *Org. Lett.* **2007**, 9, 5095.

²² (a) Allen, A. E.; MacMillan, D. W. C. *J. Am. Chem. Soc.*, **2011**, 133, 4260; (b) Shiina, I.; Nakata, K.; Onda, Y.-S. *Eur. J. Org. Chem.* **2008**, 5887; (c) Fadel, A. *Synlett* **1992**, 48.

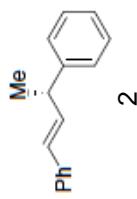
²³ Neumann, H.; Brennführer, A.; Beller, M. *Adv. Synth. Catal.* **2008**, 350, 2437.



compound 2



compound 2



145.83
137.65
135.19
128.48
128.44
128.39
127.15
126.99
126.11
126.07

20.93
42.65

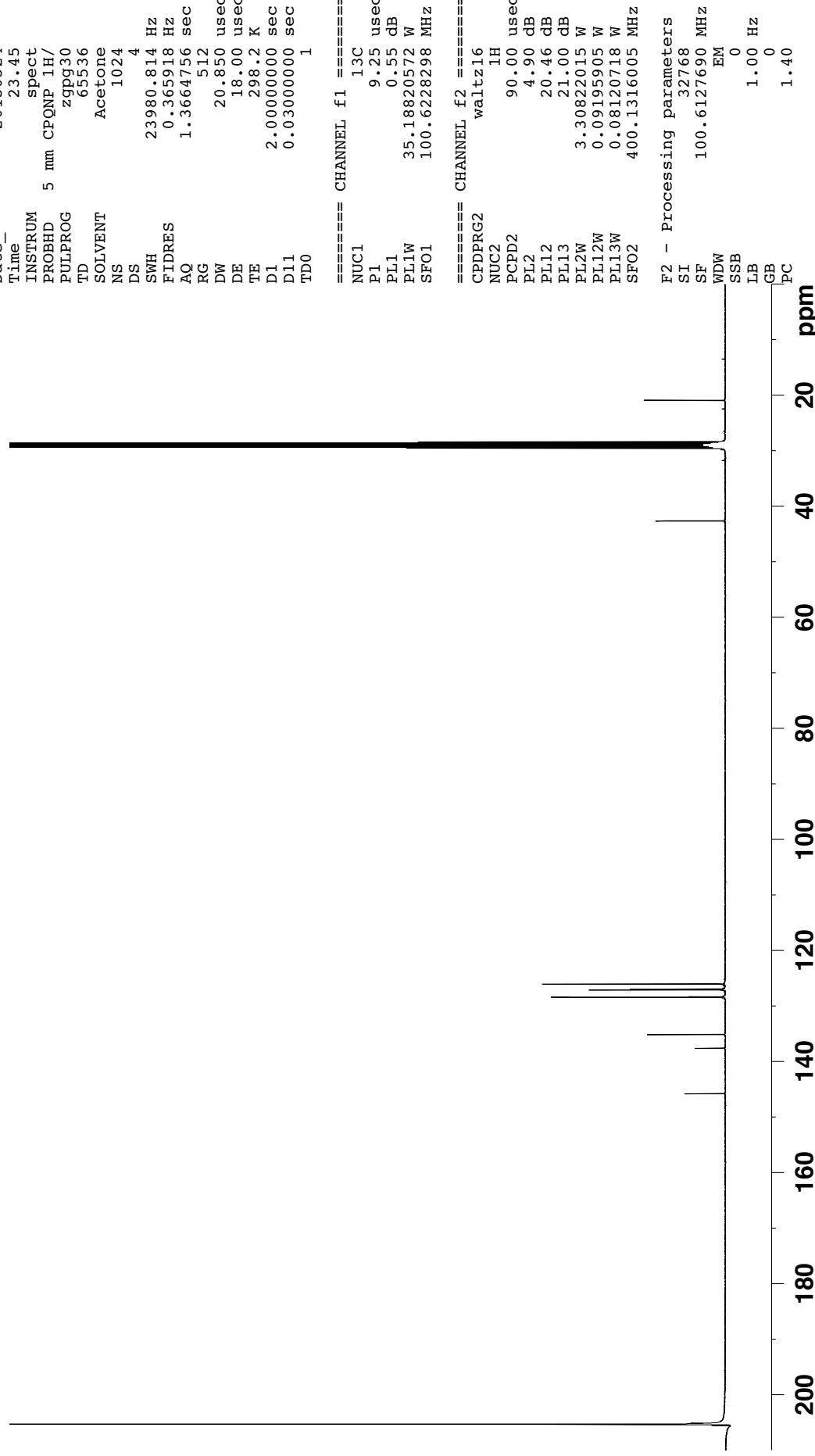
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PROCNO 1

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PULPROG zgpg30
TD 65536
SOLVENT Acetone
NS 1024
DS 4
SWH 23980.814 Hz
FIDRES 0.365918 Hz
AQ 1.3664756 sec
RG 512
DW 20.850 usec
DE 18.00 usec
TE 298.2 K
D1 2.0000000 sec
D11 0.0300000 sec
TD0 1

===== CHANNEL f1 =====
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P1 9.25 usec
PL1 0.55 dB
PL1W 35.18820572 W
SFO1 100.6228298 MHz

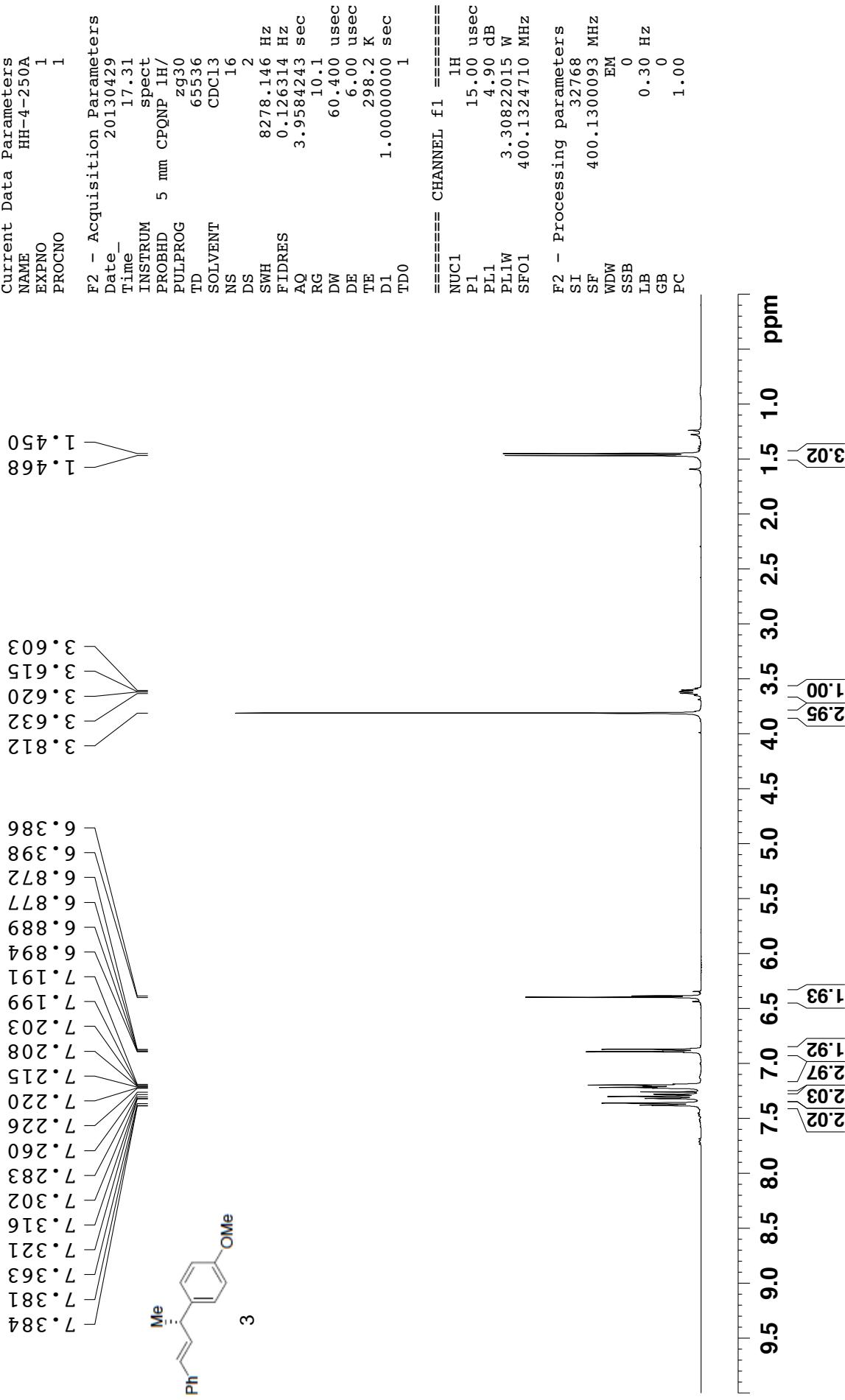
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PL2 4.90 dB
PL12 20.46 dB
PL13 21.00 dB
PL2W 3.30822015 W
PL12W 0.09195905 W
PL13W 0.08120718 W
SFO2 400.1316005 MHz

F2 - Processing parameters
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SF 100.6127690 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40



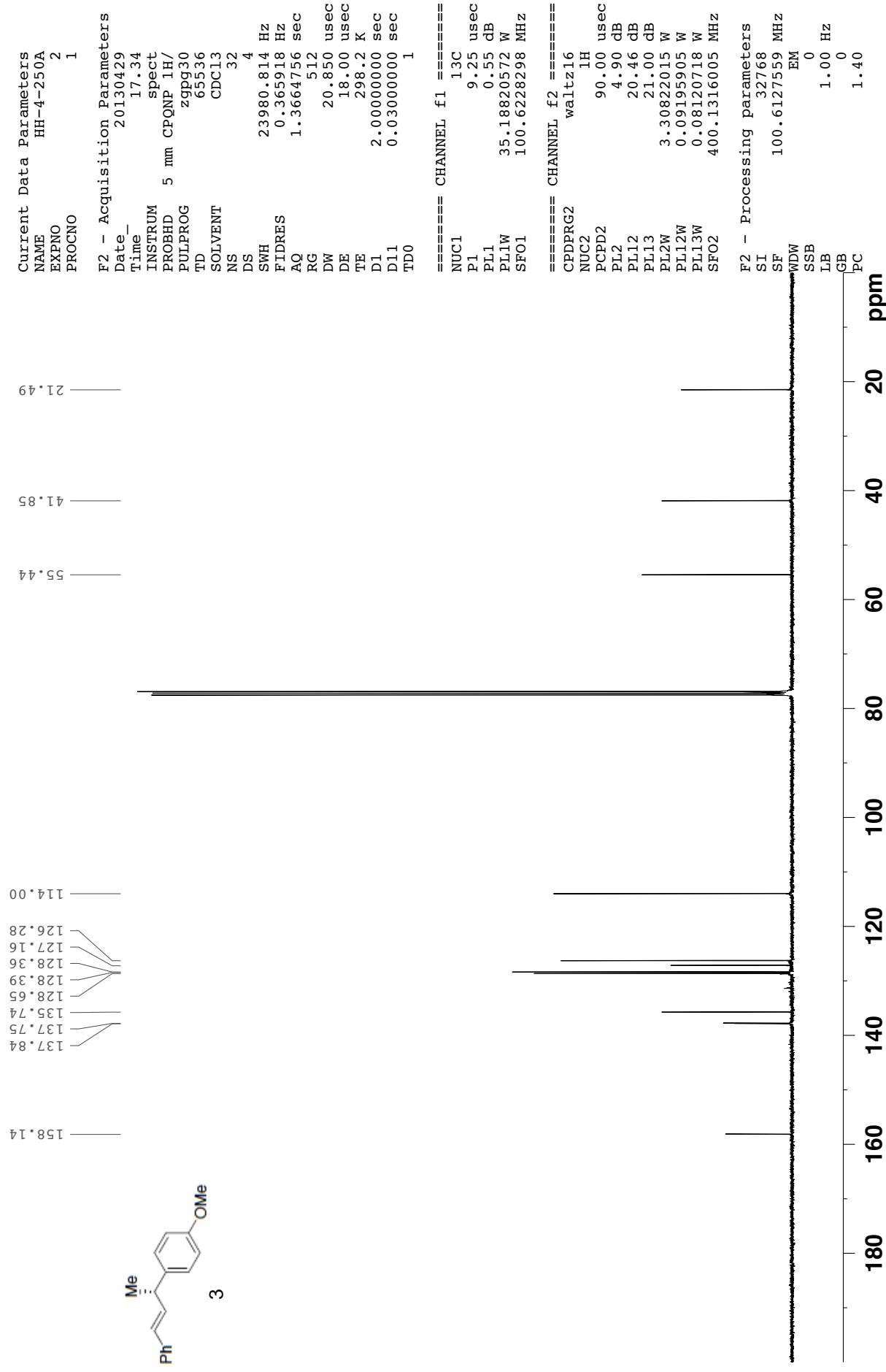
BRUKER

compound 3



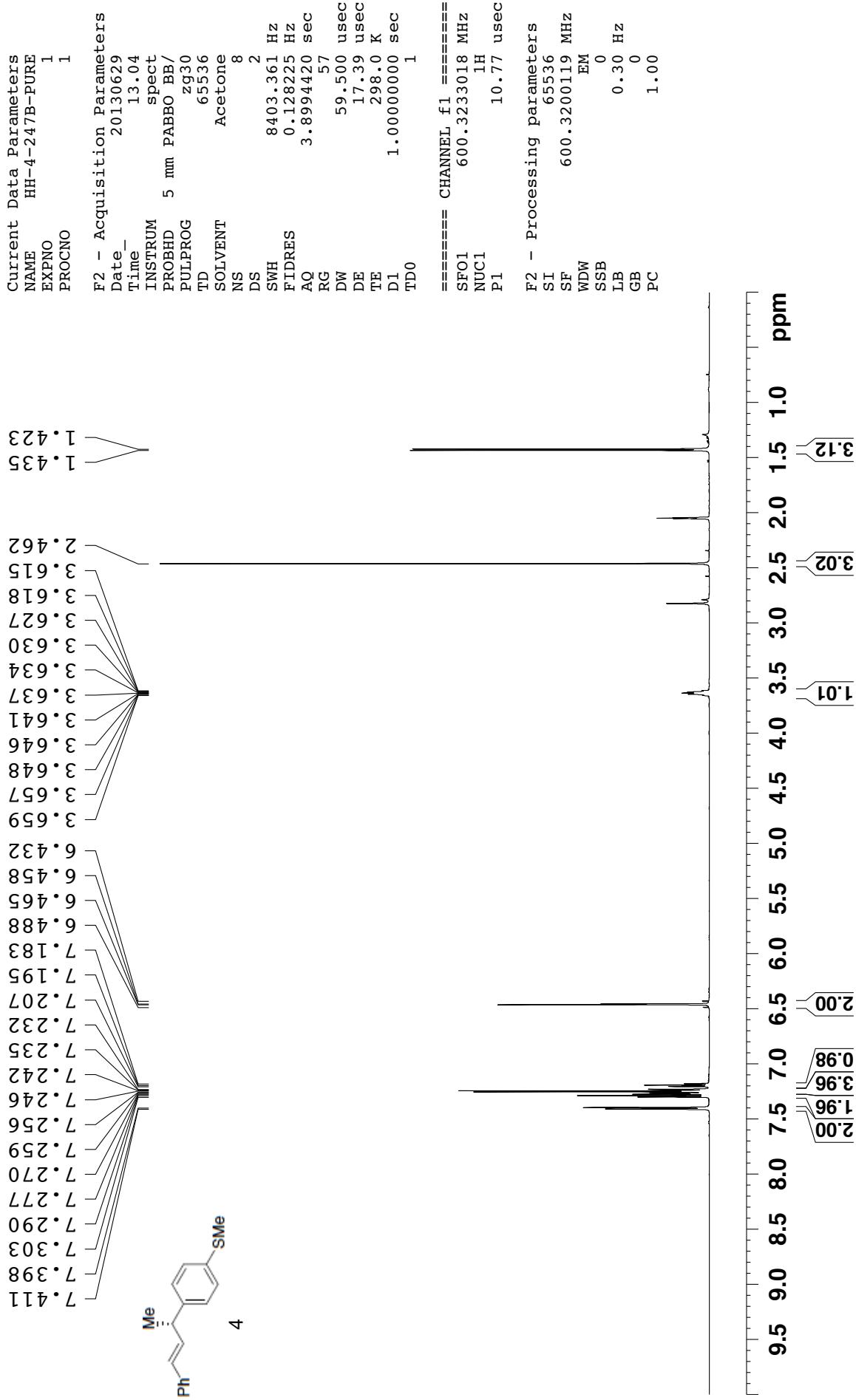


compound 3



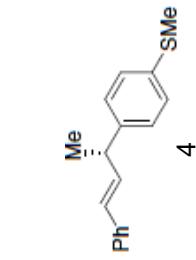
The Bruker logo consists of the word "BRUKER" in a bold, black, sans-serif font. The letters are partially enclosed by two stylized, symmetrical, black, wavy lines that resemble atomic orbits or magnetic field lines.

compound 4





compound 4



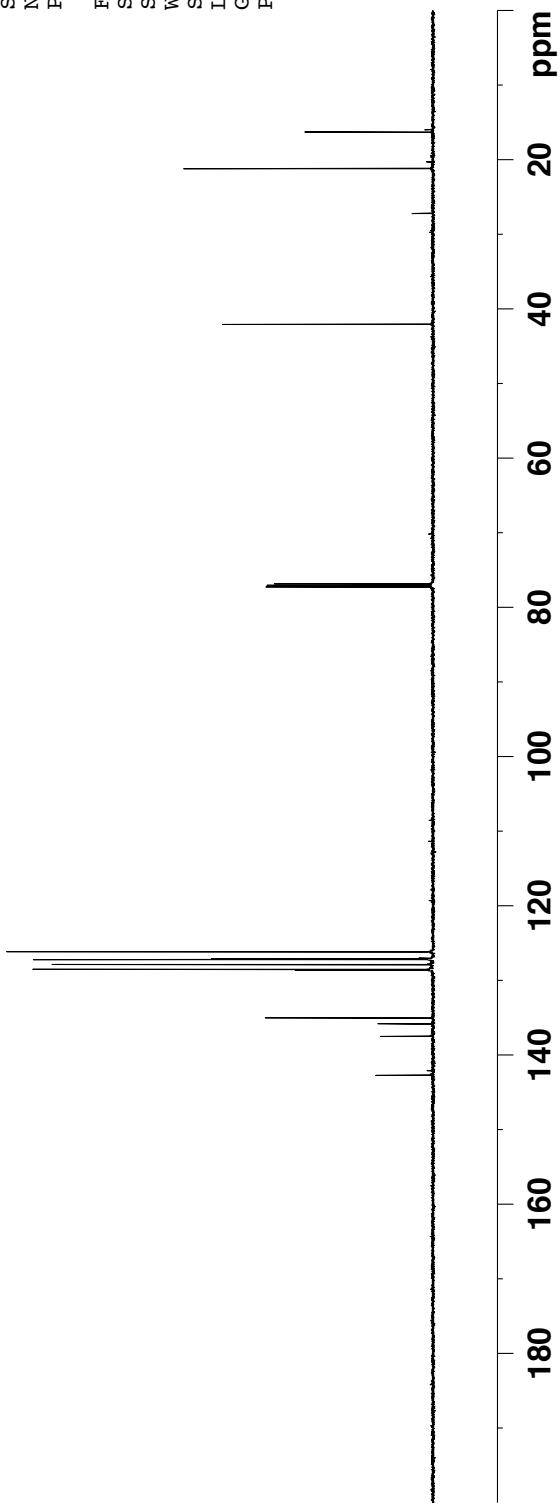
142.76
137.53
135.05
135.087
135.05
128.66
128.54
127.90
127.23
127.13
126.18

Current Data Parameters
 NAME HH-4-247B
 EXPNO 2
 PROCNO 1

F2 - Acquisition Parameters
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 Time 16.39
 INSTRUM spect
 PROBHD 5 mm PABBO BB/
 PULPROG zgpg55
 TD 65536
 SOLVENT CDCl3
 NS 256
 DS 4
 SWH 34722.223 Hz
 FIDRES 0.529819 Hz
 AQ 0.9437684 sec
 RG 2050
 DW 14.400 usec
 DE 19.34 usec
 DE 298.0 K
 TE 1.10000002 sec
 D1 0.03000000 sec
 D11 TD0 1

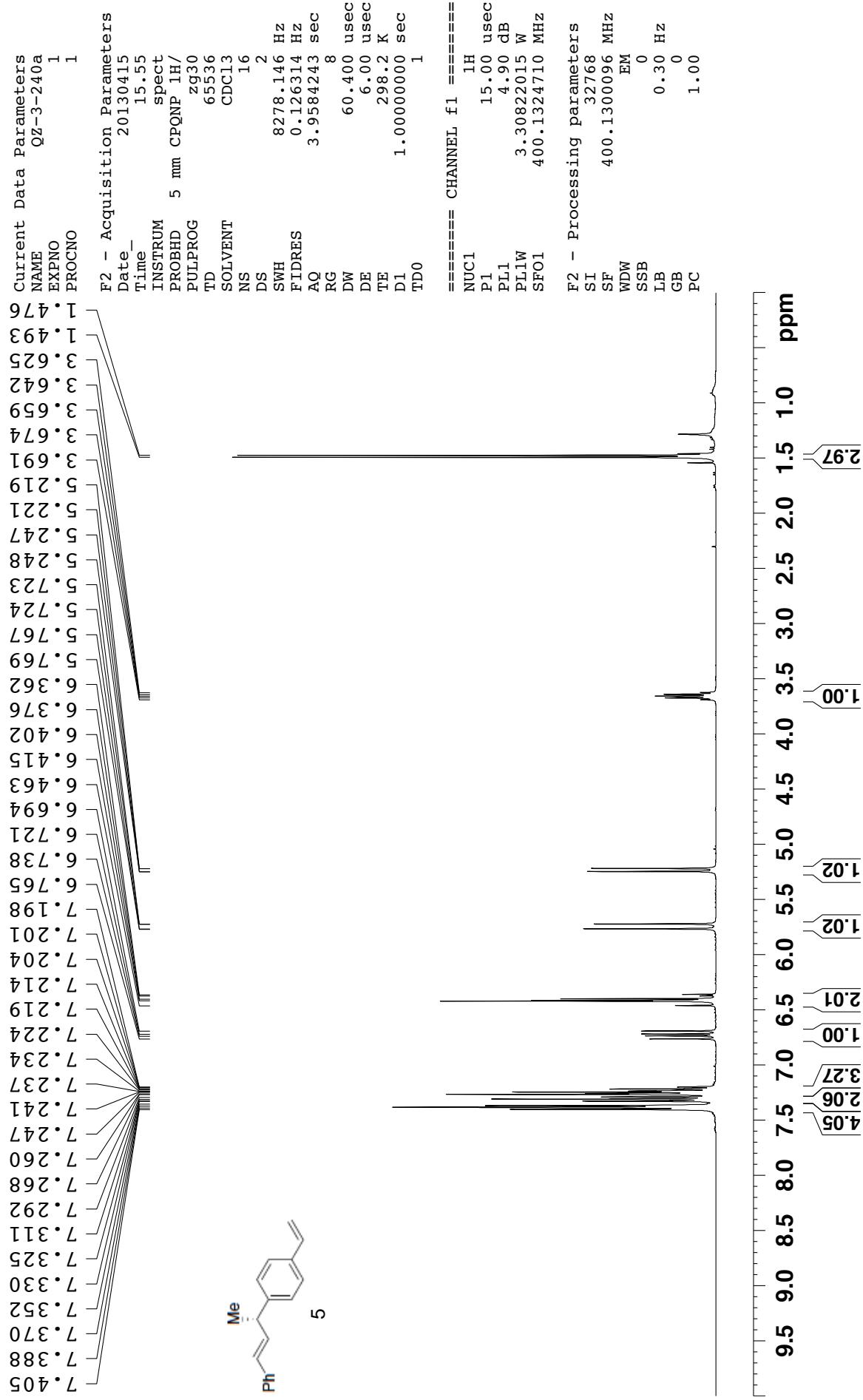
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 P1 10.63 usec

F2 - Processing parameters
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compound 5

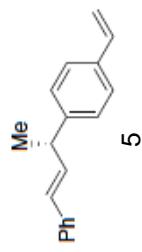
BRUKER



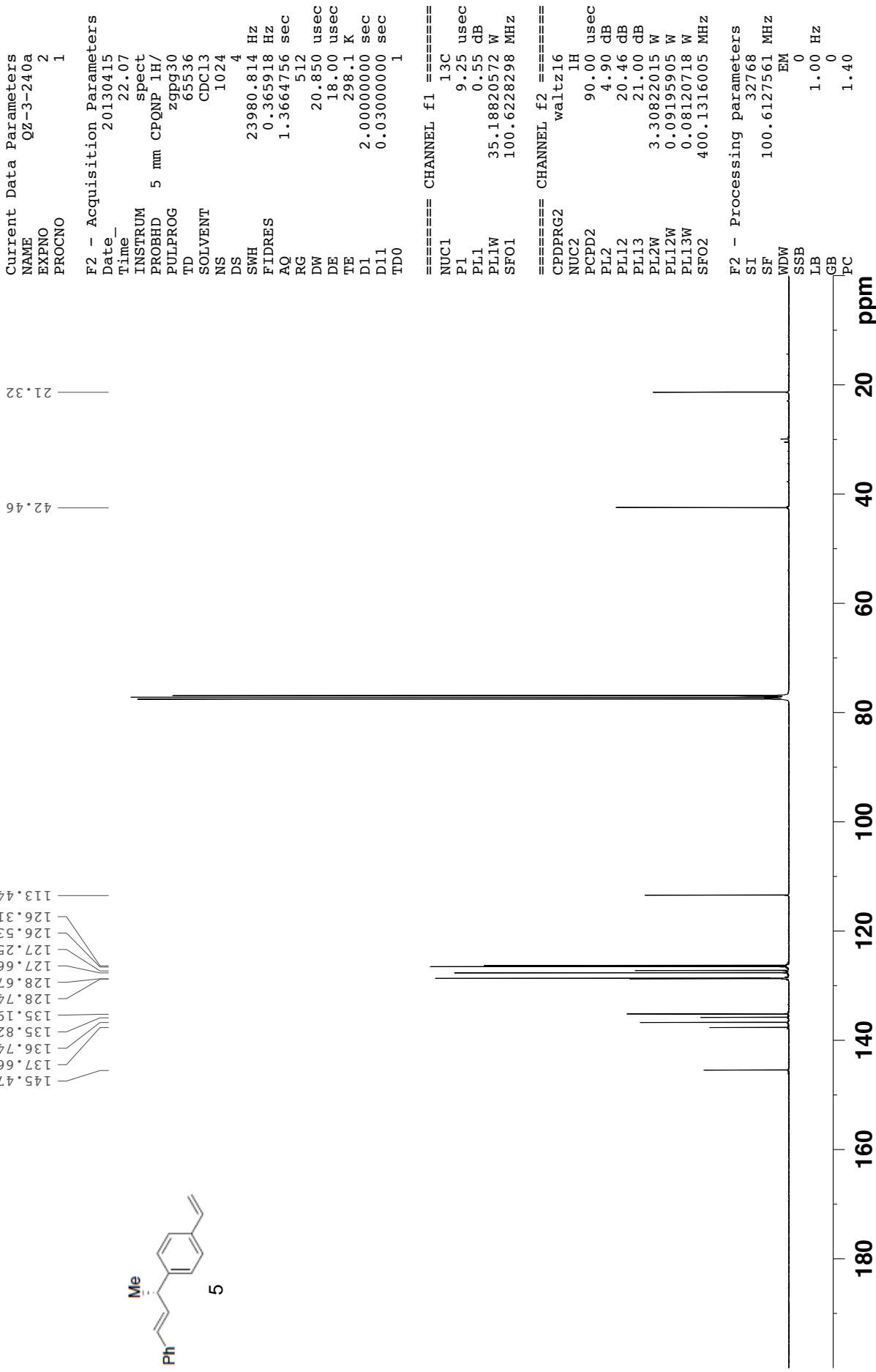


compound 5

145.47
136.74
135.19
135.82
137.66
136.74
135.19
128.67
127.66
126.53
126.31
113.44



21, 32
—
42, 46
—





Compound 6

Current Data Parameters

NAME	HH-4-242
EXPNO	1
PROCNO	1

F2 - Acquisition Parameters

Date	20130413
Time	5.33
INSTRUM	spect
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PULPROG	Zg30
TD	65536
SOLVENT	CDCl ₃
NS	16
DS	2
SWH	8278.146 Hz
FIDRES	0.126314 Hz
AQ	3.9584243 sec
RG	8
DW	60.400 usec
DE	6.00 usec
TE	298.1 K
D1	1.0000000 sec
TDO	1

===== CHANNEL f1 =====

NUC1	1H
P1	15.00 usec
PL1	4.90 dB
PL1W	3.30822015 W
SFO1	400.1324710 MHz

F2 - Processing parameters

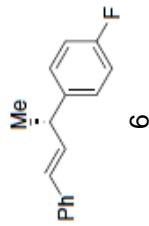
SI	32768
SF	400.1300092 MHz
WDW	EM
SSB	0
LB	0.30 Hz
GB	0
PC	1.00

Chemical structure of Compound 6: A benzene ring with a phenyl group (Ph) at position 1, a methyl group (Me) at position 2, and a fluorine atom (F) at position 4.

Integration values (ppm): 1.00, 2.02, 2.01, 1.99, 2.00, 1.99, 2.01, 9.5, 9.0, 8.5, 8.0, 7.5, 7.0, 6.5, 6.0, 5.5, 5.0, 4.5, 4.0, 3.5, 3.0, 2.5, 2.0, 1.5, 1.0, 3.09 ppm.

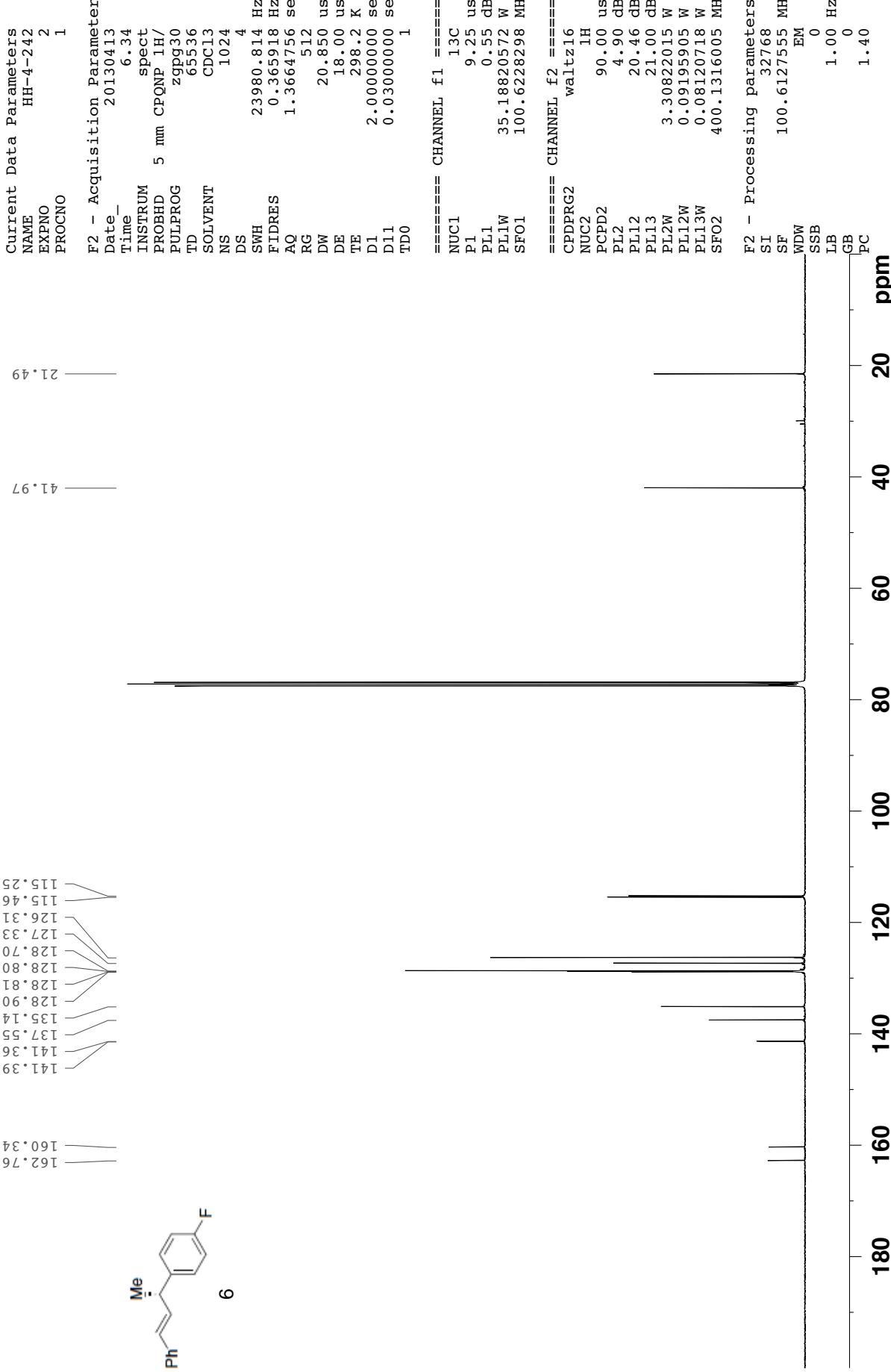
compound 6

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128.80
128.70
127.33
126.31
125.46
115.25



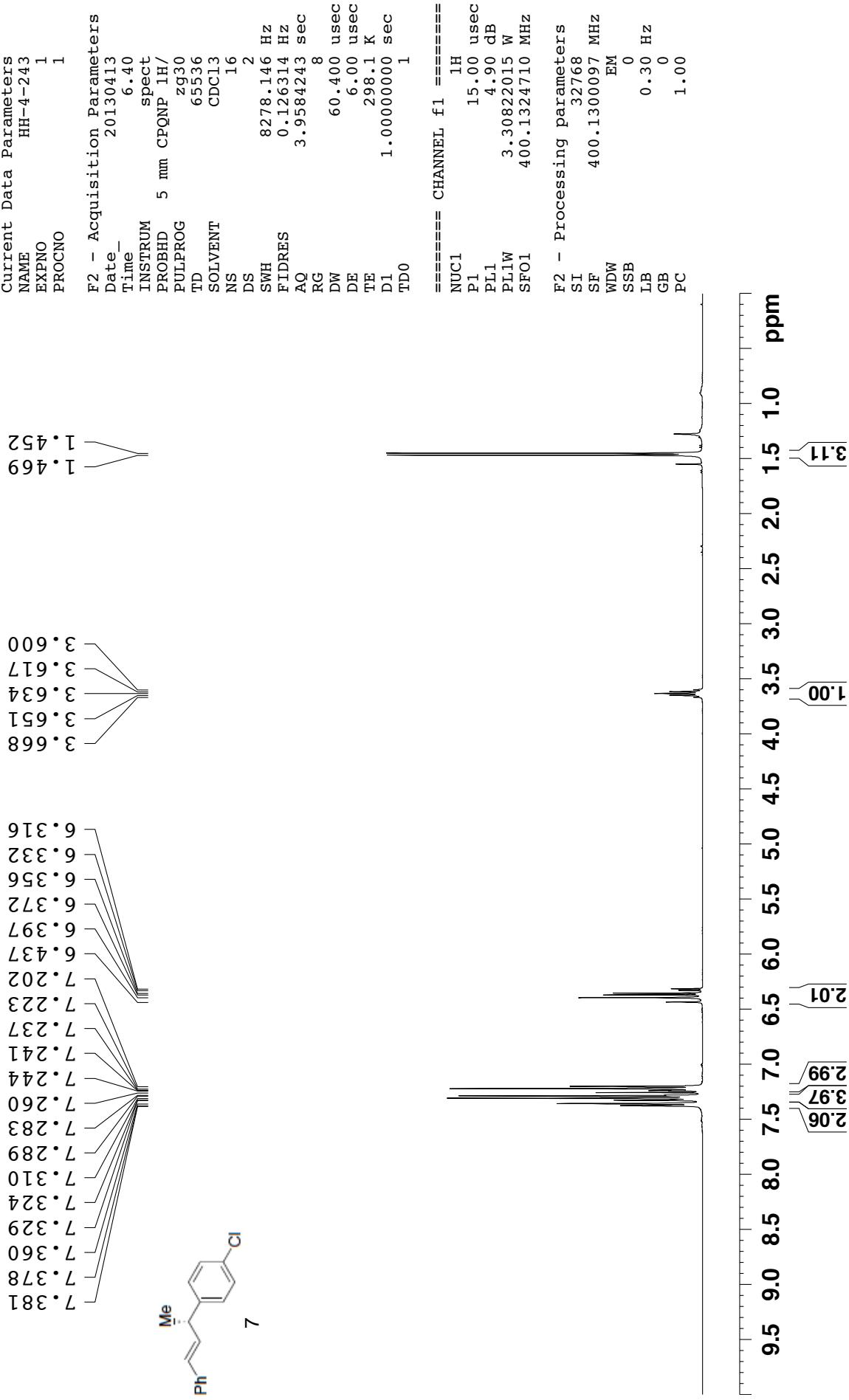
— 21.49 —

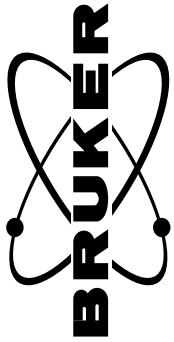
— 41.97 —





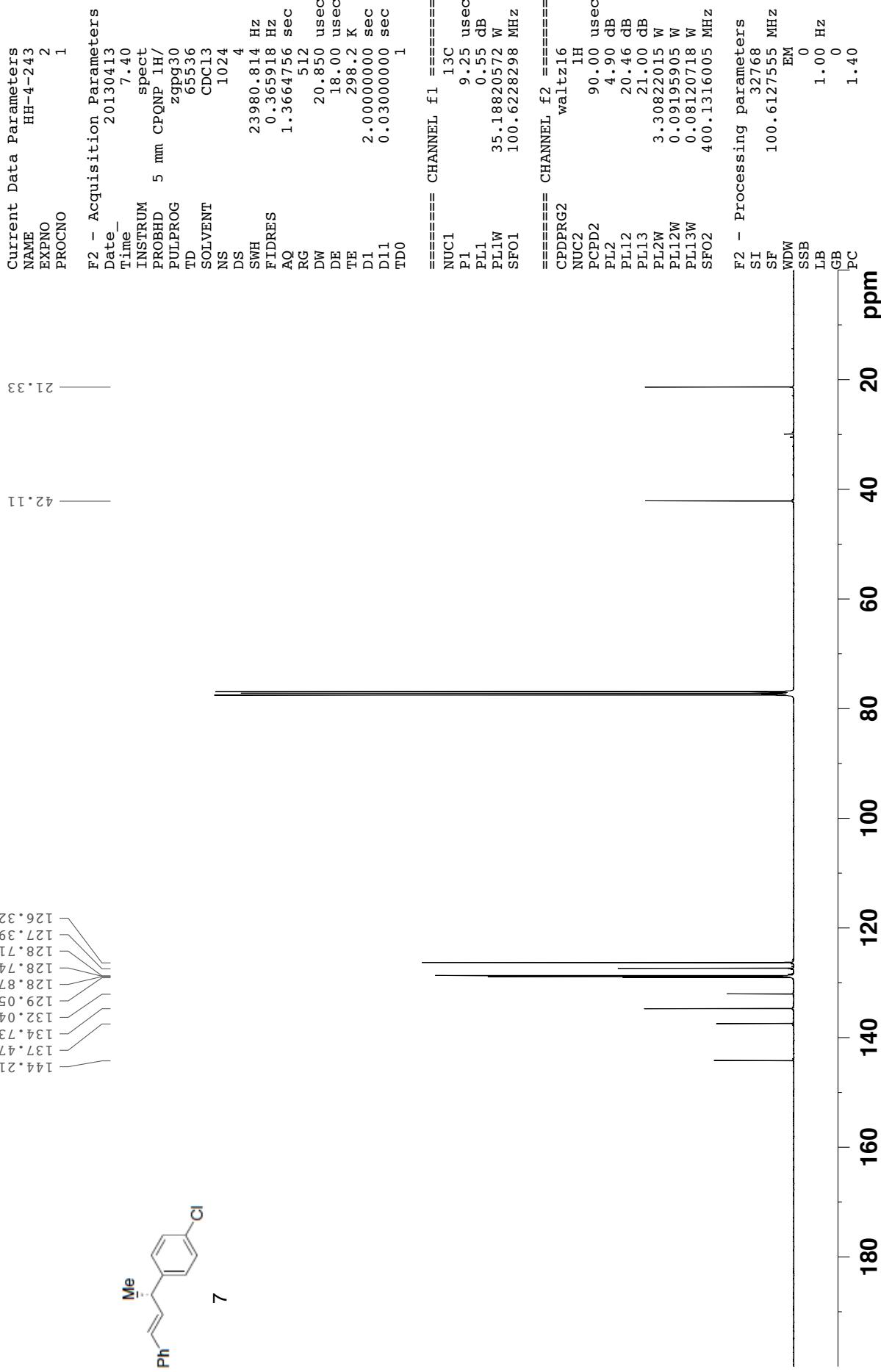
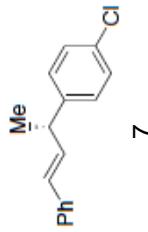
compound 7





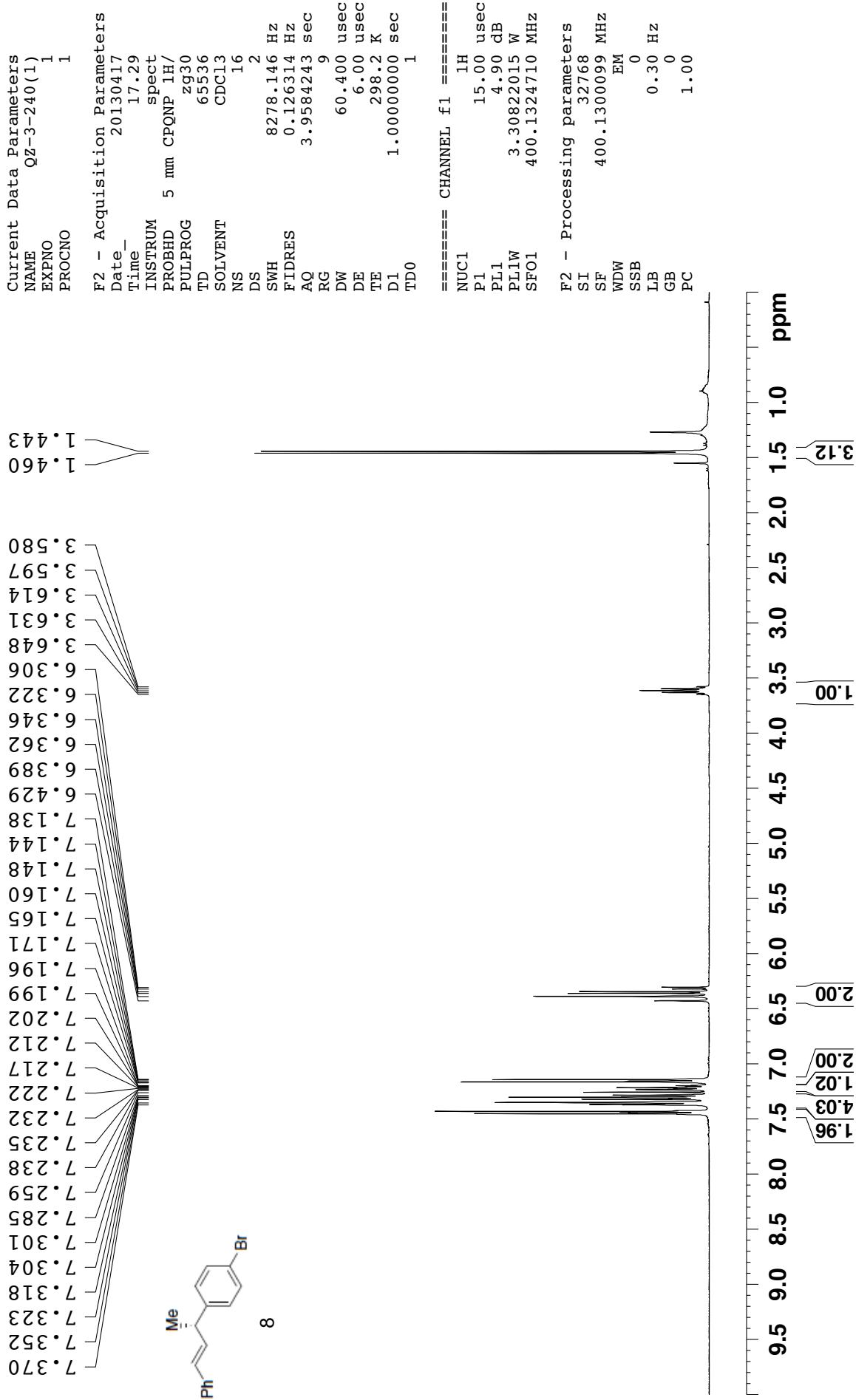
compound 7

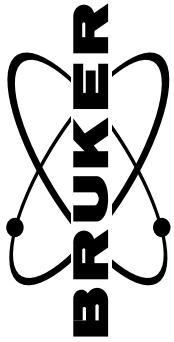
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129.05
128.87
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127.39
126.32





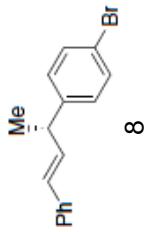
compound 8





compound 8

144.74
137.45
134.63
131.69
129.29
129.10
128.71
127.40
126.32
120.11



21.28
—
42.17
—

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Current Data Parameters
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PROCNO    1

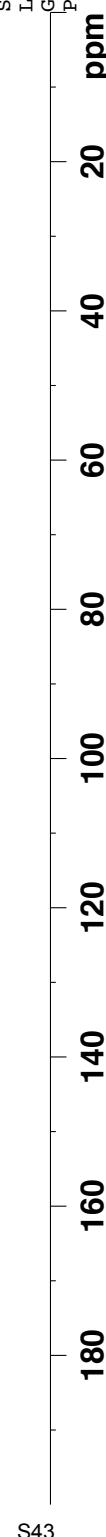
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PULPROG  zgpg30
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NS        1024
DS         4
SWH      23980.814 Hz
FIDRES   0.365918 Hz
AQ        1.3664756 sec
RG        512
DW       20.850 usec
DE        18.00 usec
TE       298.1 K
D1      2.0000000 sec
D11     0.0300000 sec
TD0        1

===== CHANNEL f1 =====
NUC1      13C
P1        9.25 usec
PL1      35.18820572 W
PL1W    100.6228298 MHz
SFO1

===== CHANNEL f2 =====
CPDPRG2
NUC2      1H
PCPD2     90.00 usec
PL2        4.90 dB
PL12     20.46 dB
PL13     21.00 dB
PL2W    3.30822015 W
PL12W   0.09195905 W
PL13W   0.08120718 W
SFO2     400.1316005 MHz

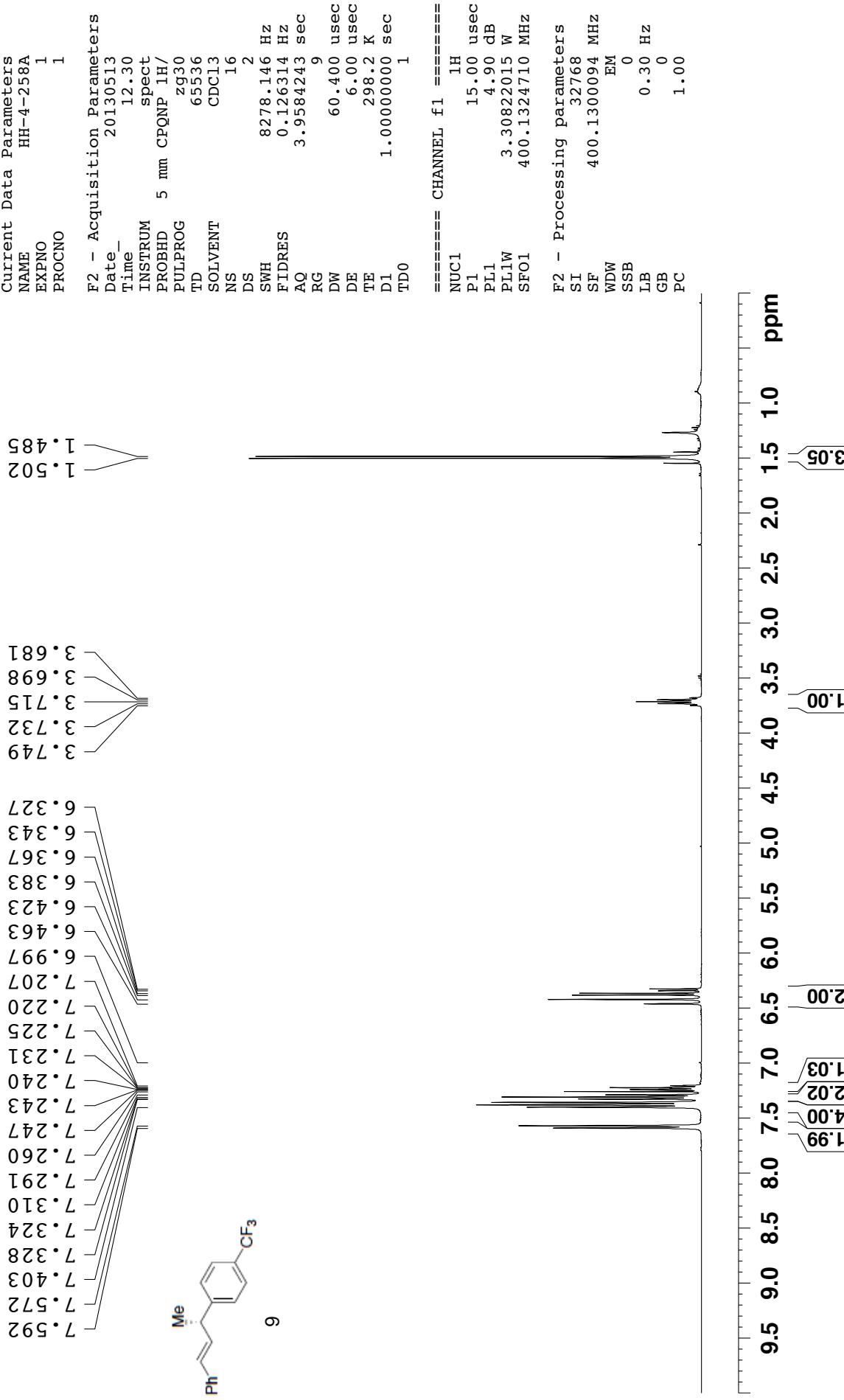
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WDW
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PC      1.40

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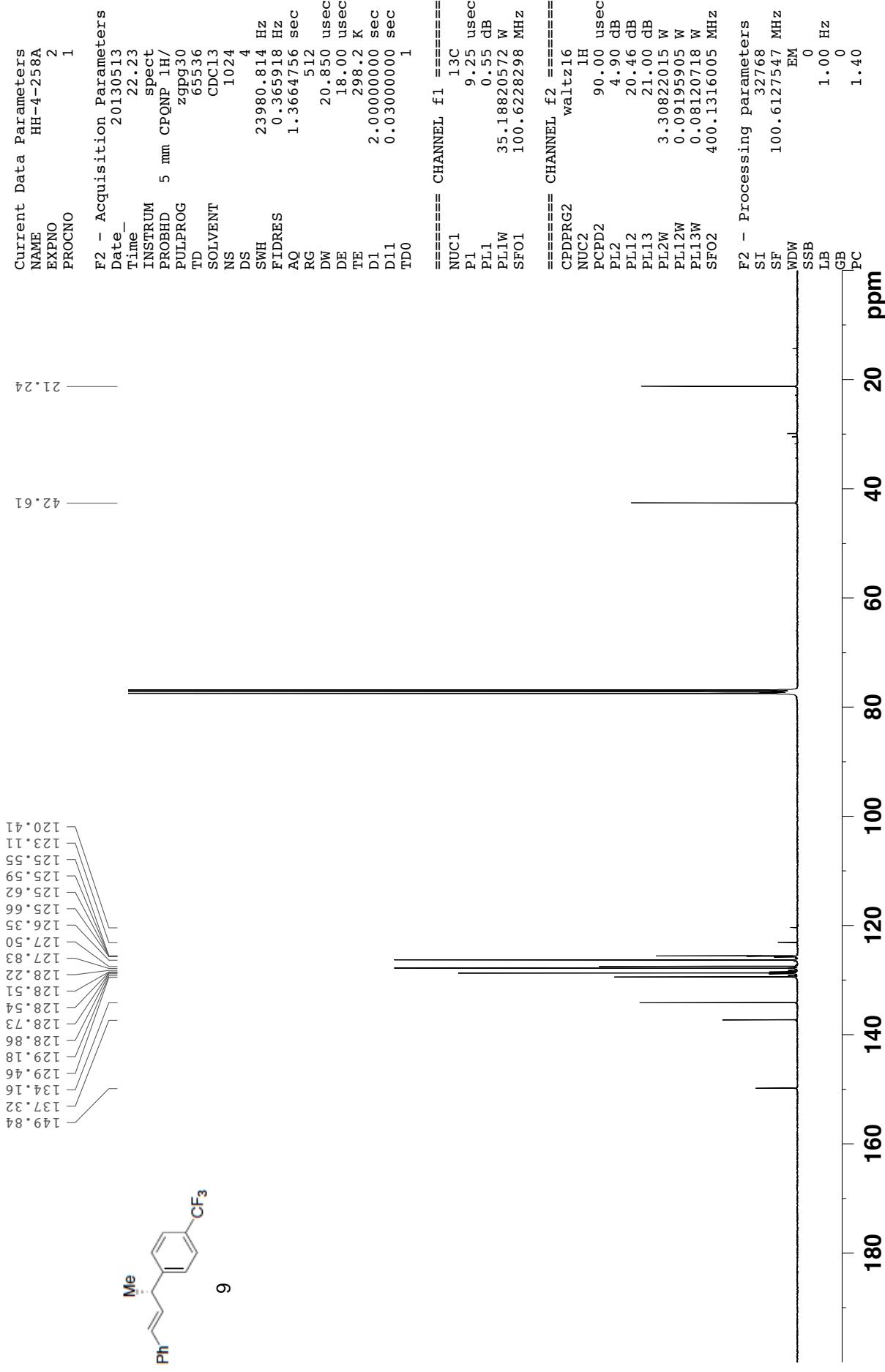
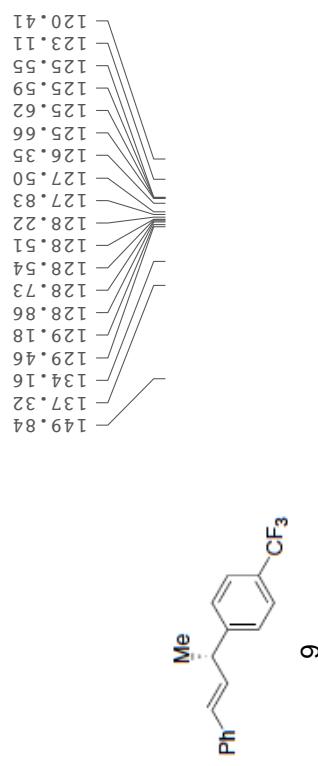
BRUKER

compound 9

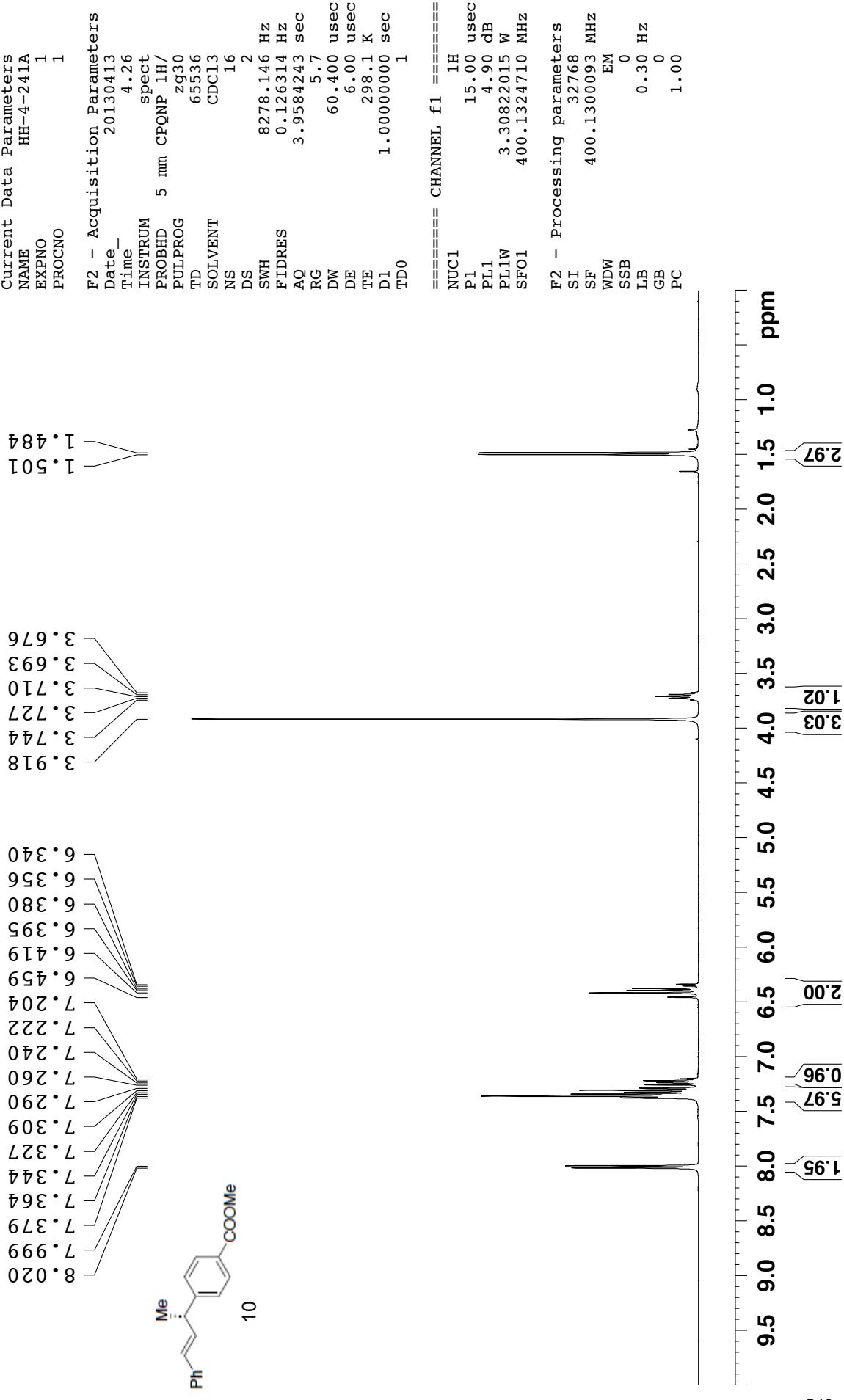




compound 9



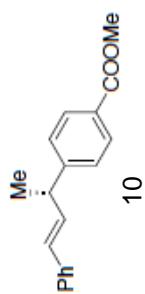
compound 10



compound 10



151.14
167.21
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130.02
129.32
128.69
128.31
127.51
127.41
126.31



52.18
42.75
21.18

```

Current Data Parameters
NAME HH-4-241A
EXPNO 2
PROCNO 1

F2 - Acquisition Parameters
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Time 5.26
INSTRUM spect
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PULPROG zgpg30
TD 65536
SOLVENT CDCl3
NS 1024
DS 4
SWH 23980.814 Hz
FIDRES 0.365918 Hz
AQ 1.3664756 sec
RG 512
DW 20.850 usec
DE 18.00 usec
TE 298.2 K
D1 2.0000000 sec
D11 0.0300000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 13C
P1 9.25 usec
PL1 0.55 dB
PL1W 35.18820572 W
SF01 100.6228298 MHz

===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 90.00 usec
PL2 4.90 dB
PL12 20.46 dB
PL13 21.00 dB
PL2W 3.30822015 W
PL12W 0.09195905 W
PL13W 0.08120718 W
SF02 400.1316005 MHz

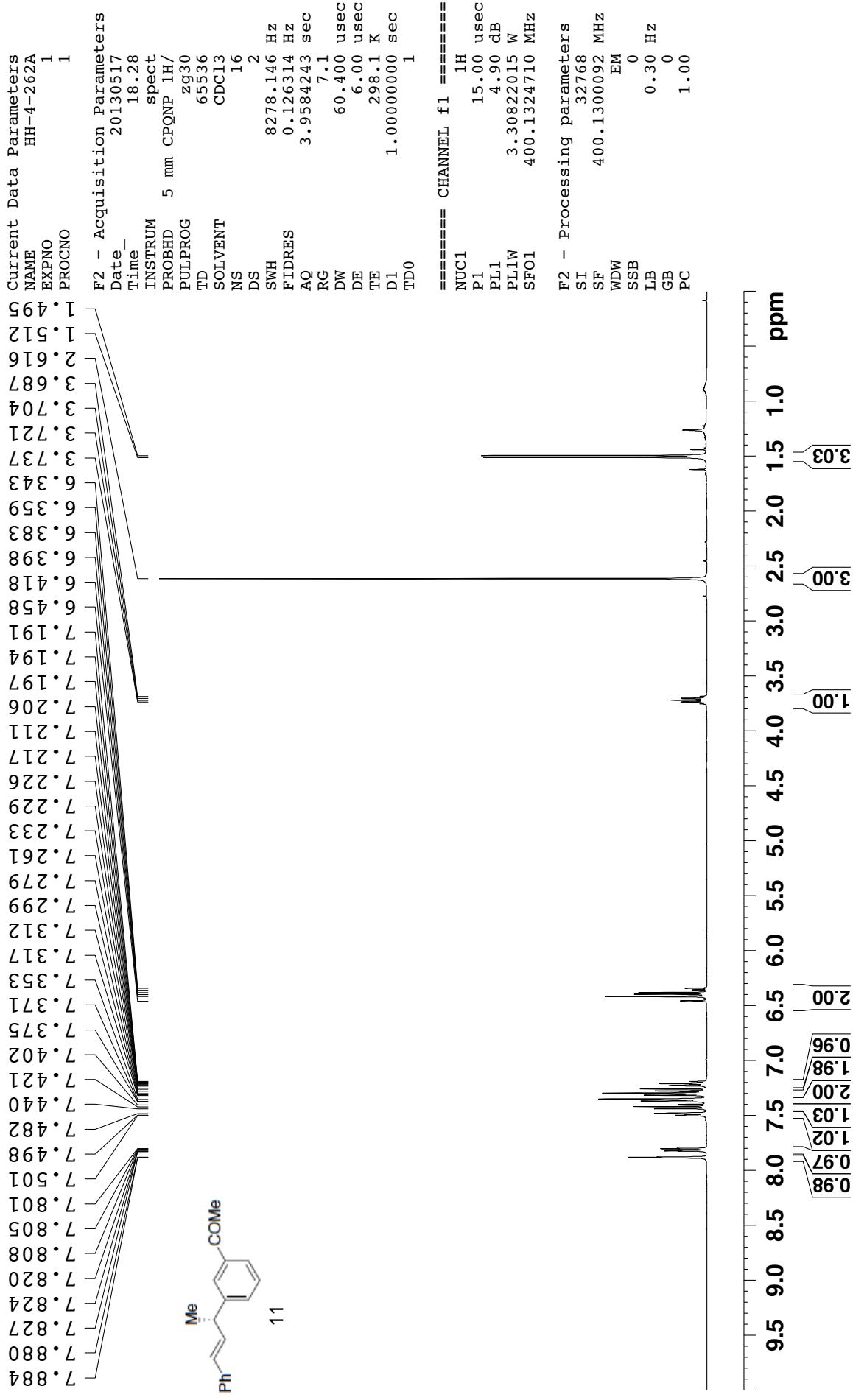
F2 - Processing parameters
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WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

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200 180 160 140 120 100 80 60 40 20 0 ppm

compound 11

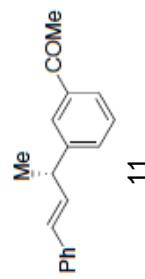
BRUKER



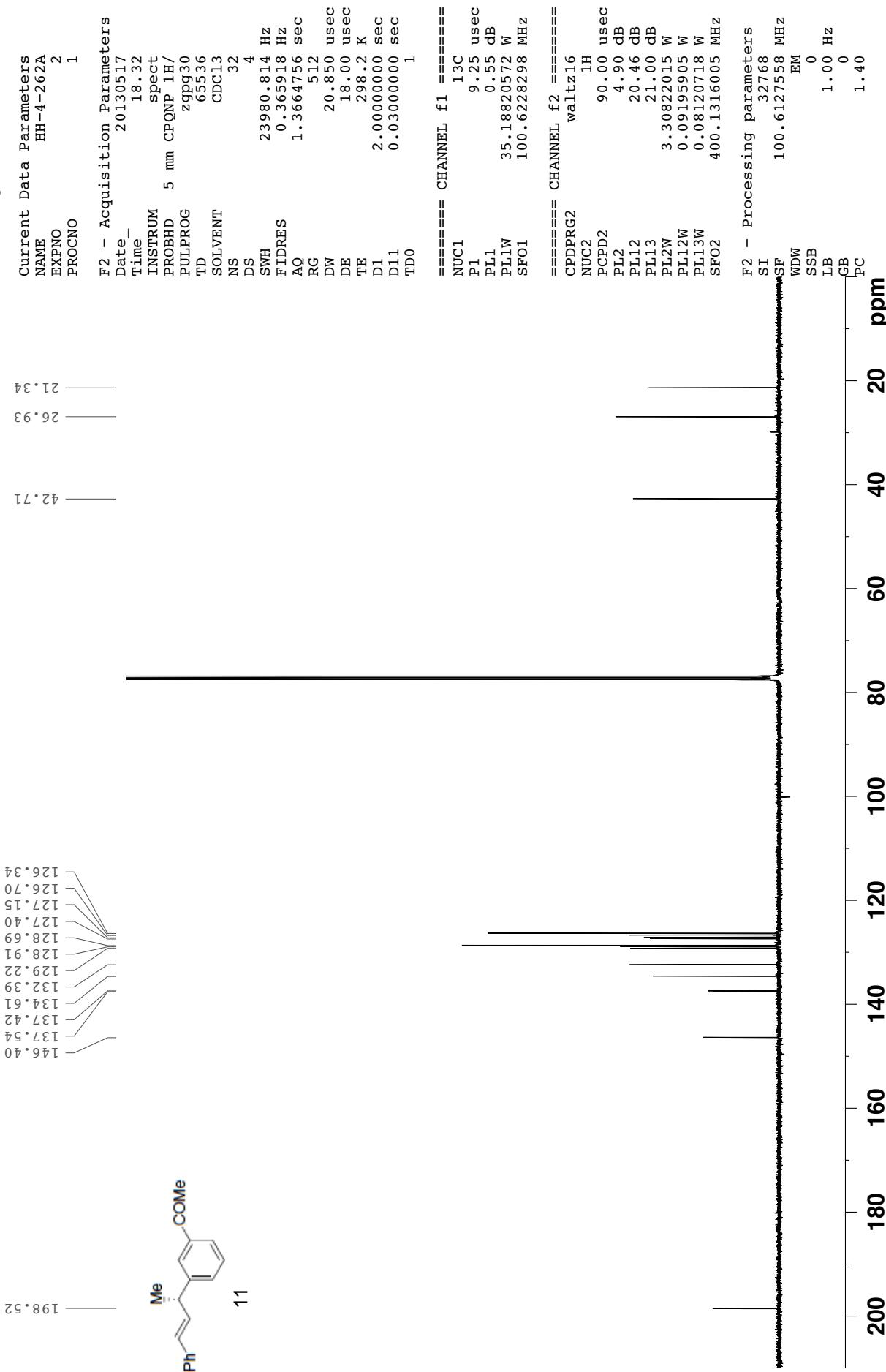


compound 11

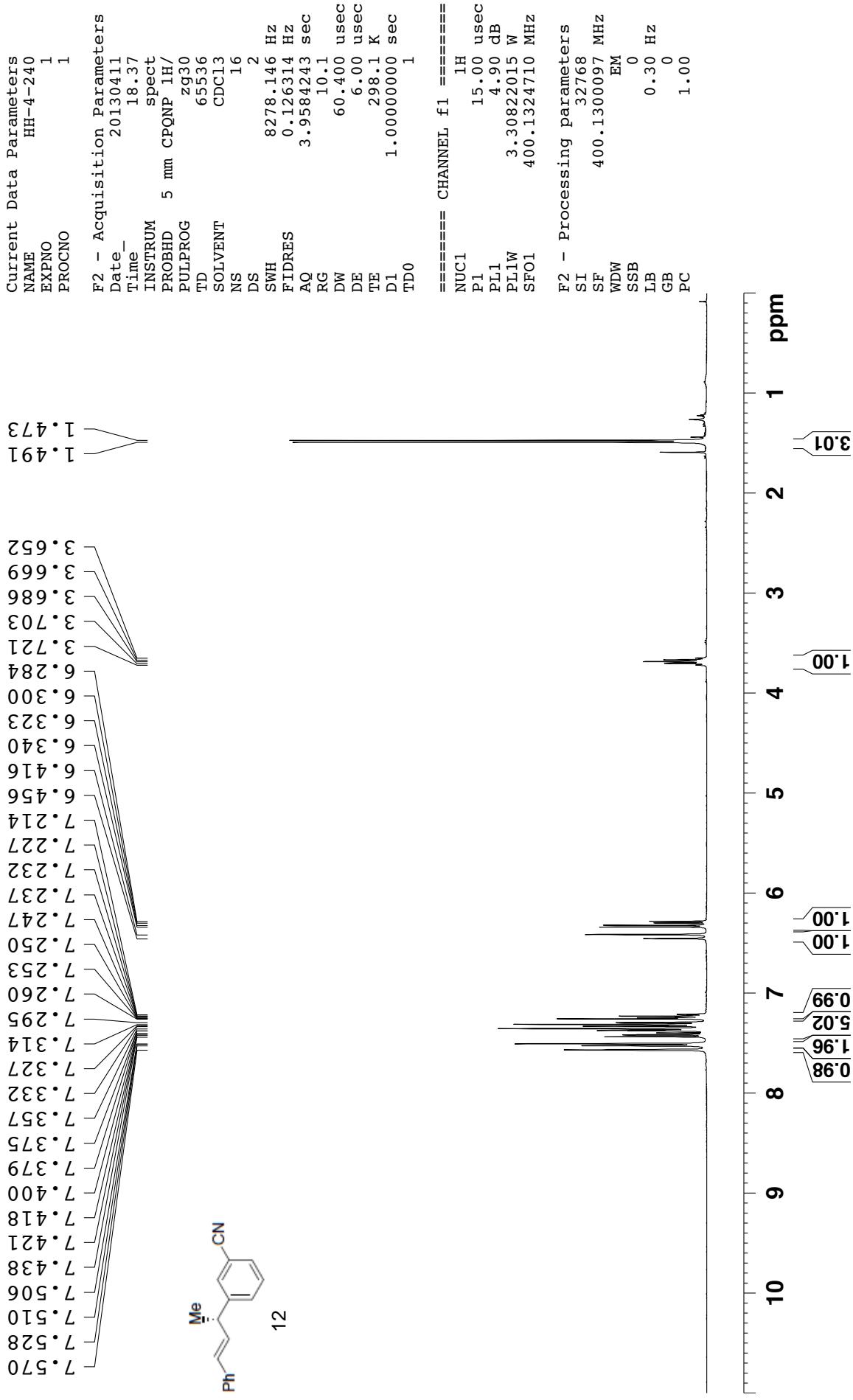
146.40
137.54
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134.61
132.39
129.22
128.91
128.69
127.40
127.15
126.70
126.34

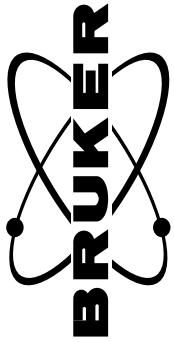


42.71
26.93
21.34



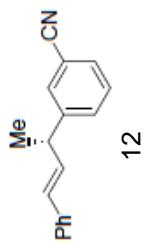
compound 12





compound 12

147.14
137.09
133.60
132.18
131.11
130.20
129.82
129.44
128.75
127.62
126.35
119.20
112.62



21.10
42.32

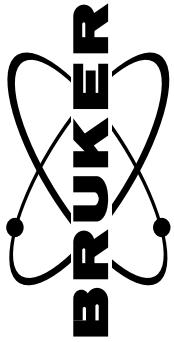
Current Data Parameters
NAME HH-4-240
EXPNO 2
PROCNO 1

F2 - Acquisition Parameters
Date 20130412
Time 4.48
INSTRUM spect
PROBHD 5 mm CPQNP 1H/
PULPROG zpg30
TD 65536
SOLVENT CDCl3
NS 1024
DS 4
SWH 23980.814 Hz
FIDRES 0.365918 Hz
AQ 1.3664756 sec
RG 512
DW 20.850 usec
DE 18.00 usec
TE 298.2 K
D1 2.0000000 sec
D11 0.0300000 sec
TD0 1

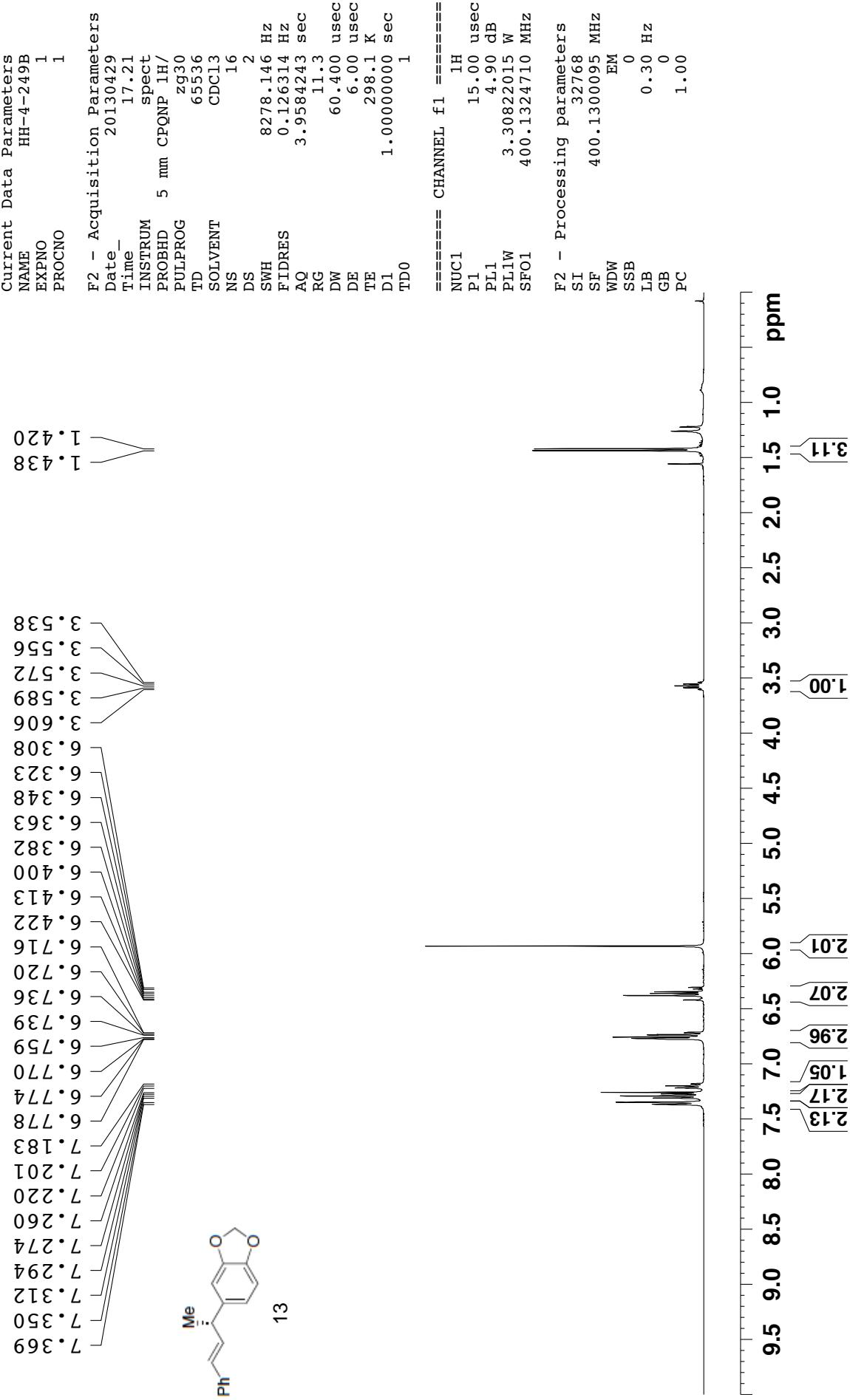
===== CHANNEL f1 =====
NUC1 13C
P1 9.25 usec
PL1 0.55 dB
PL1W 35.18820572 W
SFO1 100.6228298 MHz

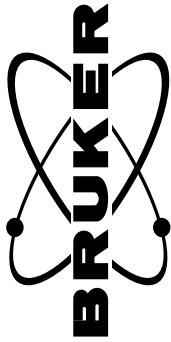
===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 90.00 usec
PL2 4.90 dB
PL12 20.46 dB
PL13 21.00 dB
PL2W 3.30822015 W
PL12W 0.09195905 W
PL13W 0.08120718 W
SFO2 400.1316005 MHz

F2 - Processing parameters
SI 32768
SF 100.6127574 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40



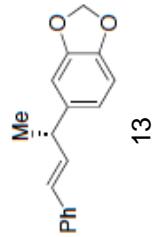
compound 13



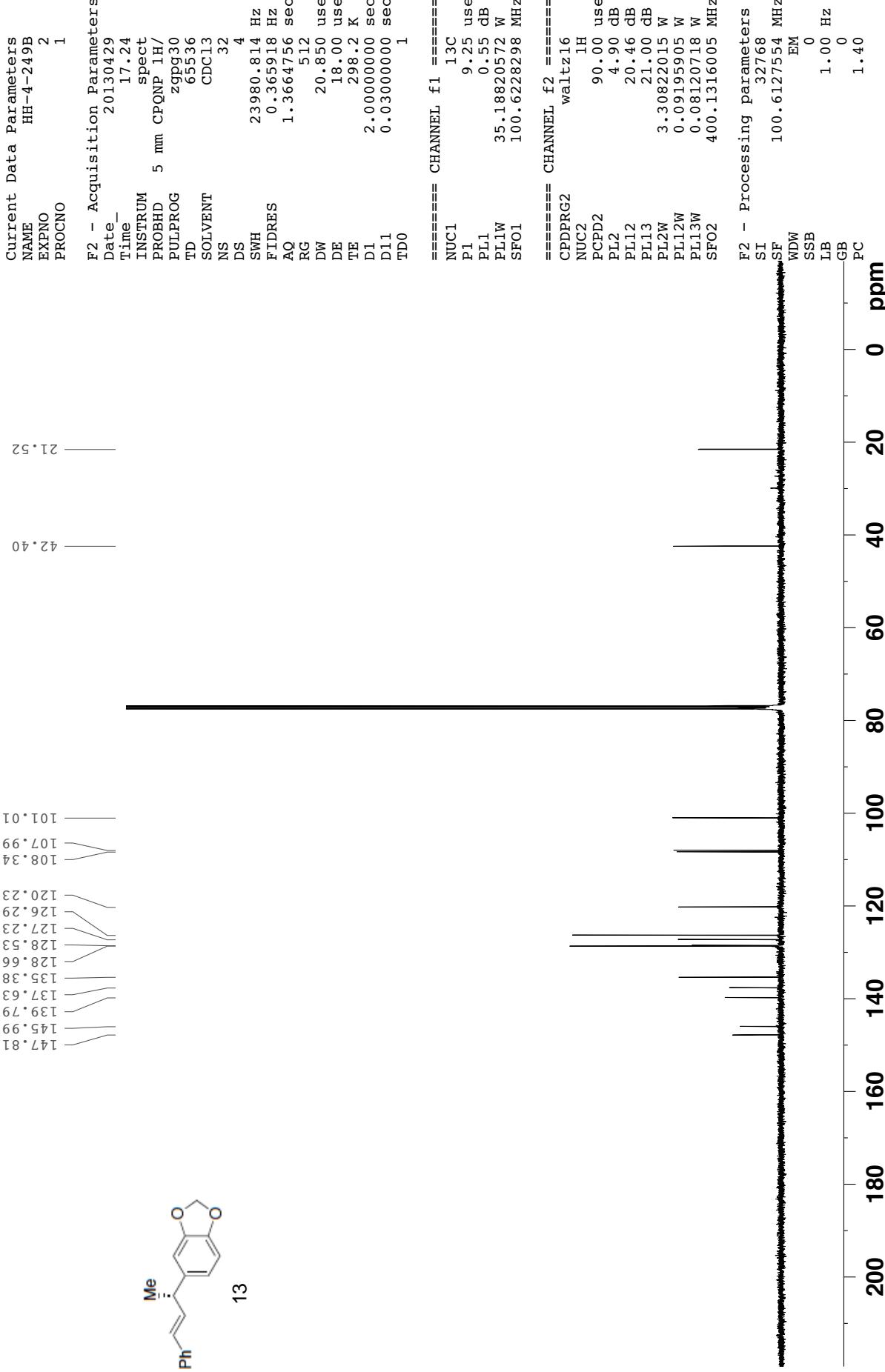


compound 13

147.81
145.99
139.79
137.63
135.38
128.66
128.53
127.23
126.29
120.23
108.34
107.99
101.01

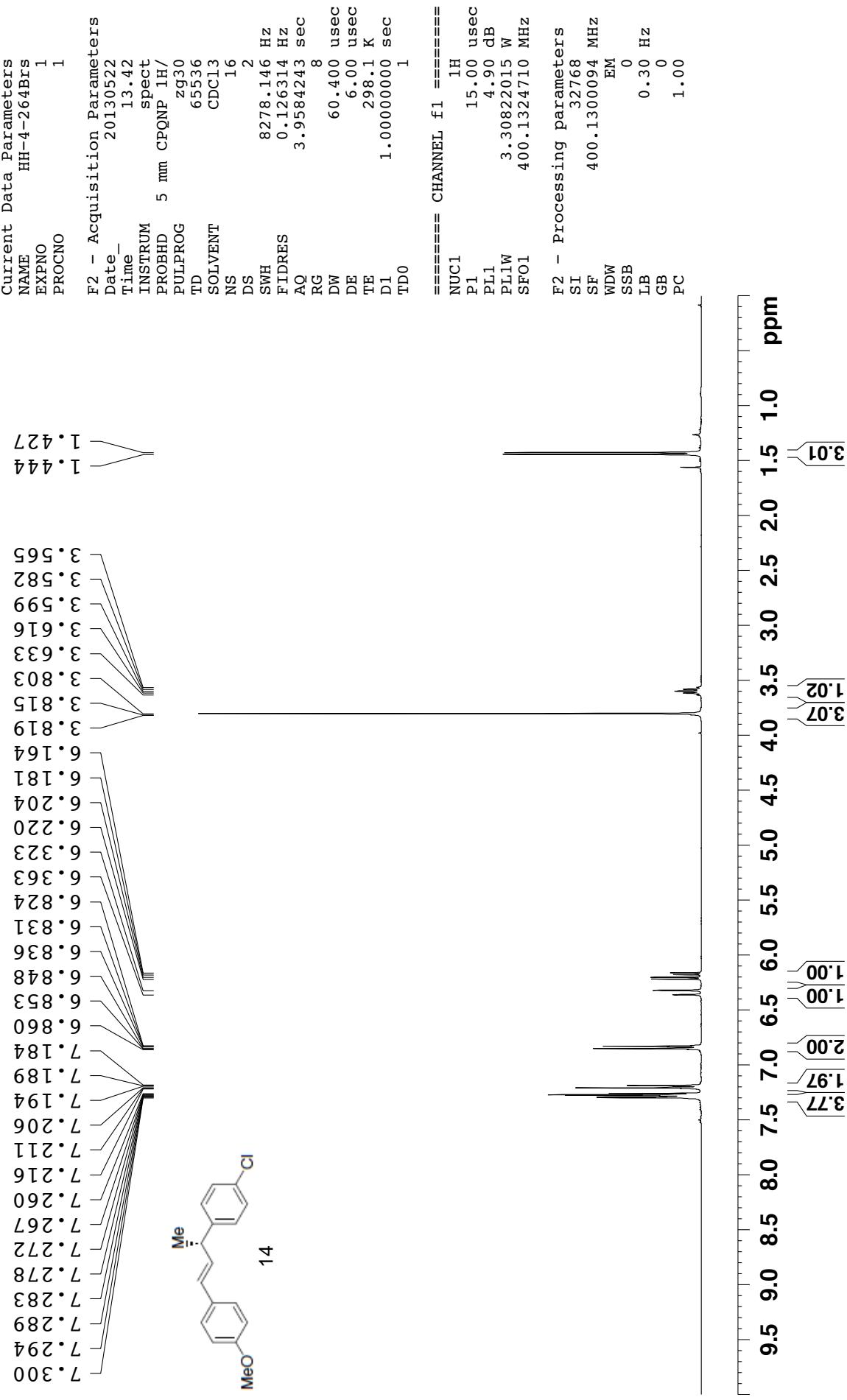


21.52
42.40

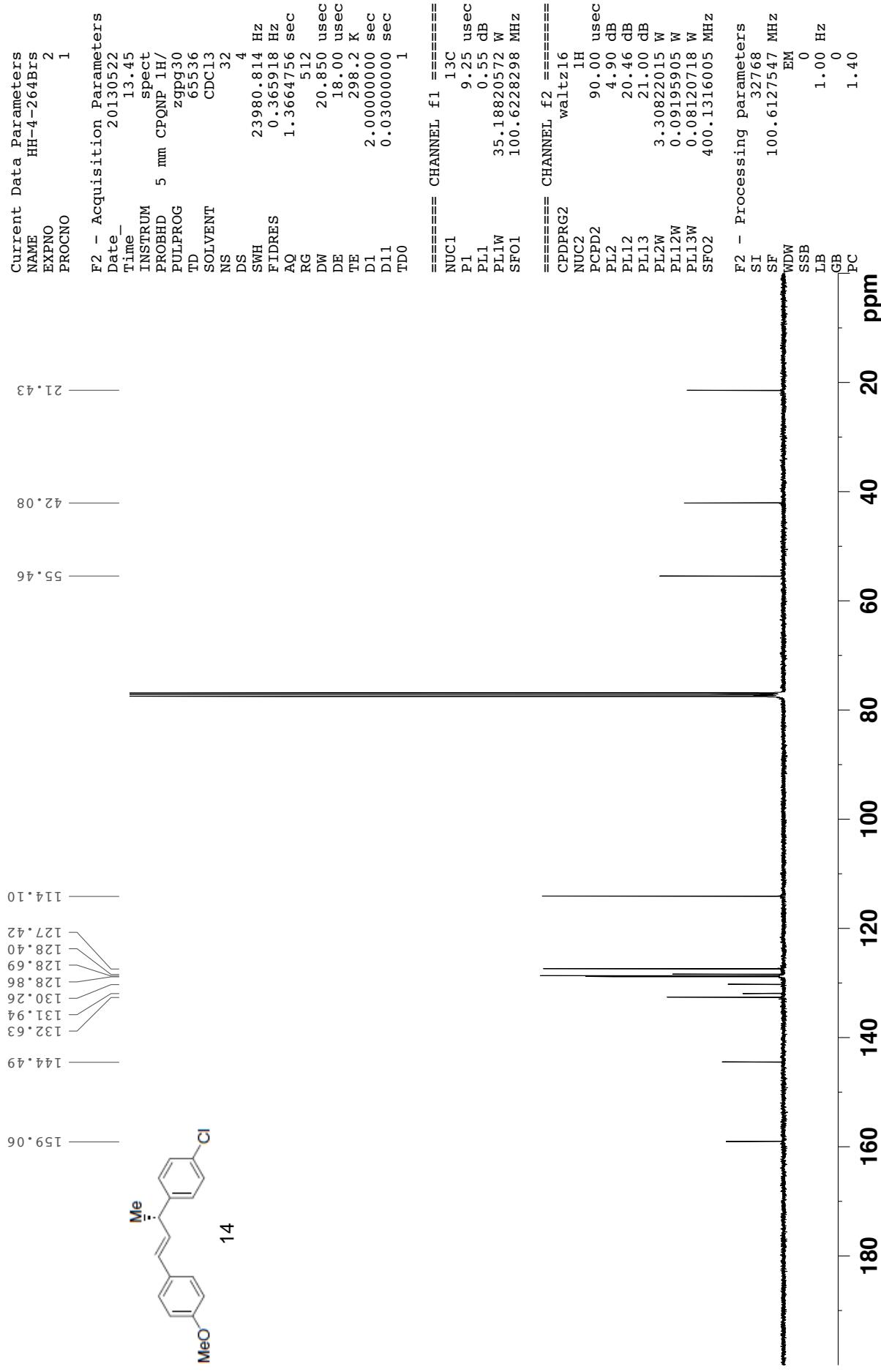


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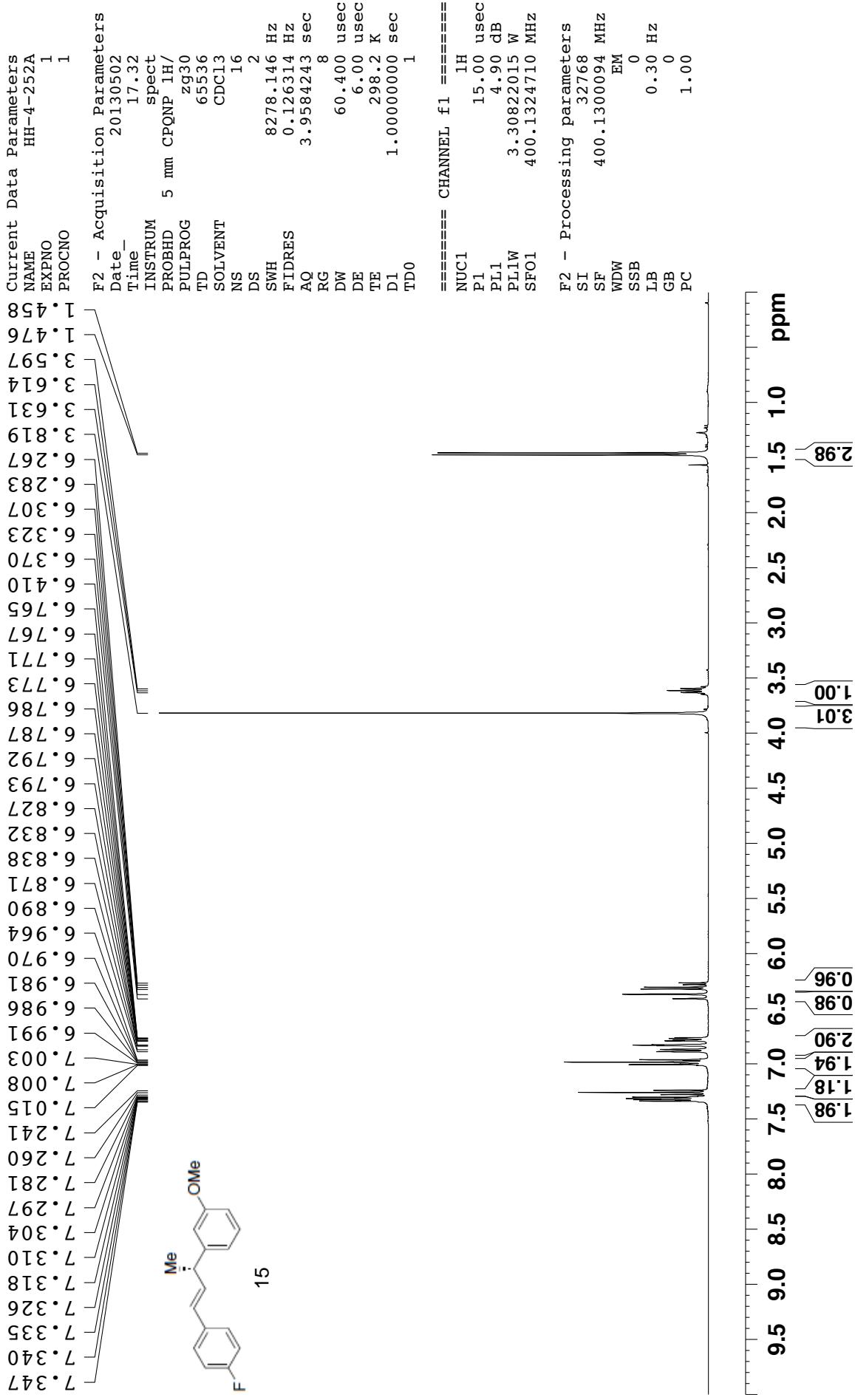
compound 14



compound 14



compound 15



The Bruker logo consists of the word "BRUKER" in a bold, black, sans-serif font. The letters are oriented vertically, with the "B" at the bottom and the "R" at the top. Two thick, black, curved lines resembling orbits or magnetic field lines are positioned above and below the letters, intersecting them.

21.30

42.73

55.34

15

Current Data Parameters

NAME	HH-4-252A
EXPNO	2
PROCNO	1

F2 - Acquisition Parameters

Date	20130502
Time	22.12
INSTRUM	spect
PROBHD	5 mm CPQNP 1H/
PULPROG	zgpg30
TD	65536
SOLVENT	CDC13
NS	1024
DS	4
SWH	23980.814 Hz
FIDRES	0.365918 Hz
AQ	1.3664756 sec
RG	512
DW	20.850 usec
DE	18.00 usec
TE	298.1 K
D1	2.00000000 sec
D11	0.03000000 sec
TD0	1

===== CHANNEL f1 =====

NUC1	13C
P1	9.25 usec
PL1	0.55 dB
PL1W	35.18820572 W
SFO1	100.6228298 MHz

===== CHANNEL f2 =====

CPDPRG2	waltz16
NUC2	1H
PCPD2	90.00 usec
PL2	4.90 dB
PL12	20.46 dB
PL13	21.00 dB
PL2W	3.30822015 W
PL12W	0.09195905 W
PL13W	0.08120718 W
SFO2	400.1316005 MHz

F2 - Processing parameters

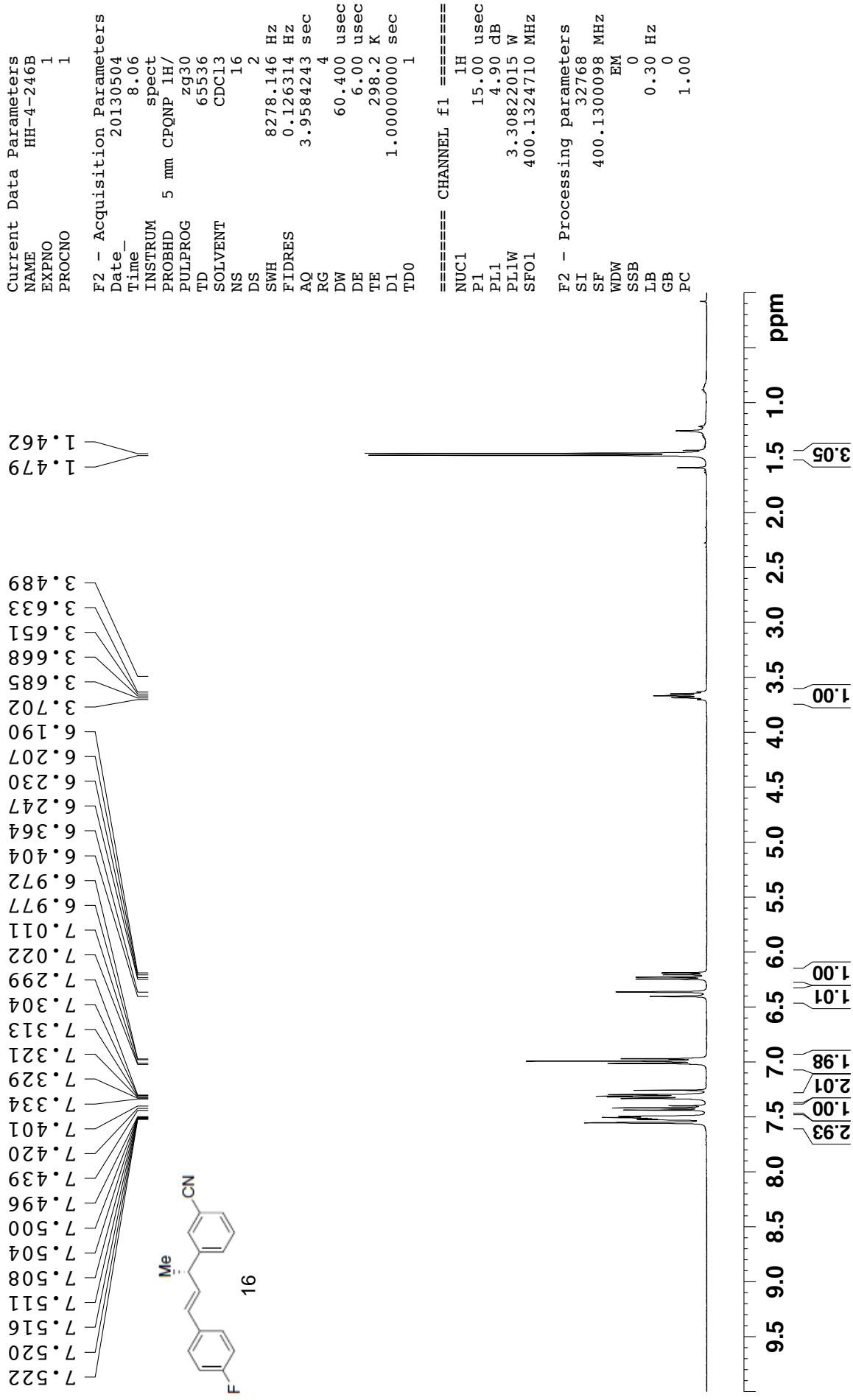
SI	32768
SF	100.6127562 MHz
WDW	EM
SSB	0
LB	1.00 Hz
GB	0
PC	1.40

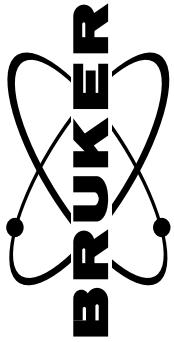
180 160 140 120 100 80 60 40 20 ppm

compound 15

Compound 16

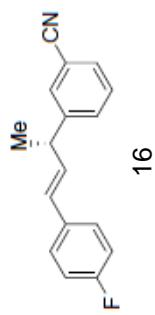
BRUKER



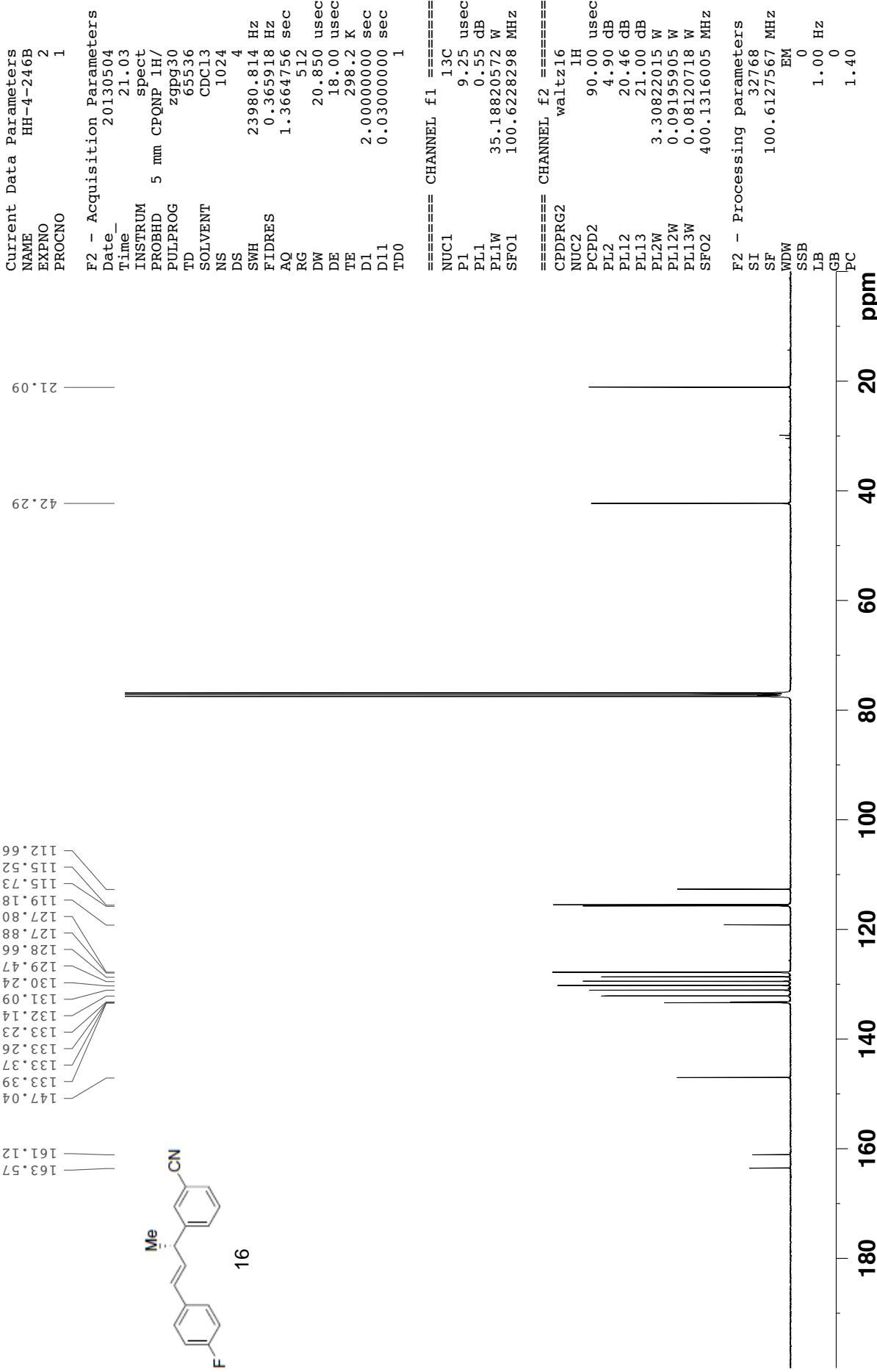


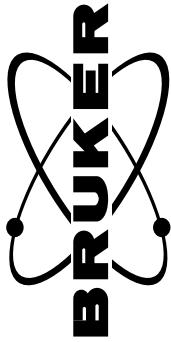
compound 16

147.04
133.39
133.37
133.26
133.23
132.14
131.09
130.24
129.47
128.66
127.88
127.80
119.18
115.73
115.52
112.66

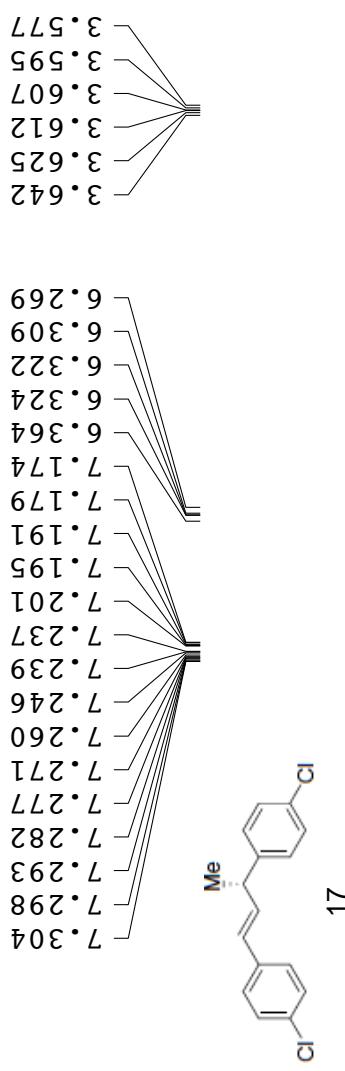


42.29
21.09





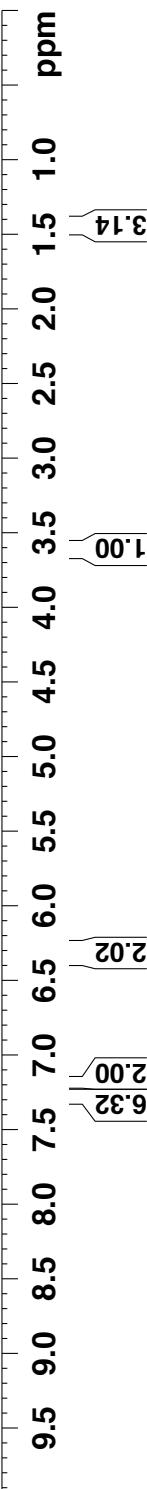
compound 17



1.448
1.431

3.642
3.625
3.612
3.607
3.595
3.577

7.304
7.298
7.293
7.277
7.271
7.260
7.246
7.239
7.201
7.195
7.191
7.179
7.174
6.364
6.324
6.309
6.269



=====
F2 - Acquisition Parameters
 NAME HH-4-257A
 EXPNO 1
 PROCNO 1

Date 20130513
Time 2.34
INSTRUM spect
PROBHD 5 mm CPQNP 1H/
PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 16
DS 2
SWH 8278.146 Hz
FIDRES 0.126314 Hz
AQ 3.9584243 sec
RG 11.3
DW 60.400 usec
DE 6.00 usec
TE 298.1 K
D1 1.0000000 sec
TDO 1

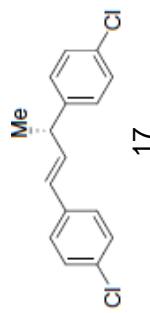
===== CHANNEL f1 =====
NUC1 1H
P1 15.00 usec
PL1 4.90 dB
PL1W 3.30822015 W
SFO1 400.1324710 MHz

=====
F2 - Processing parameters
SI 32768
SF 400.1300105 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

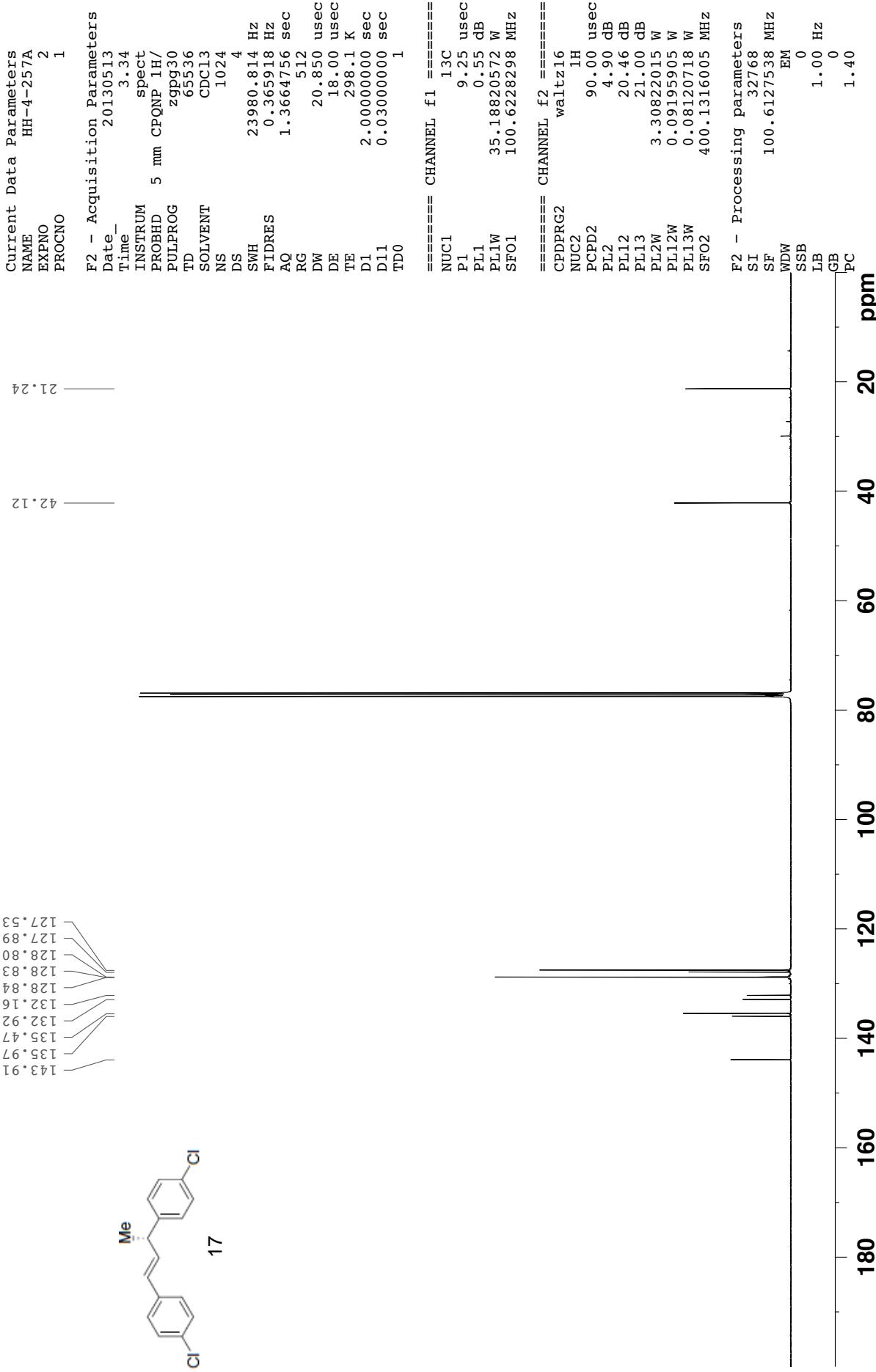


compound 17

143.91
135.97
135.47
132.92
132.16
128.84
128.83
128.80
127.89
127.53

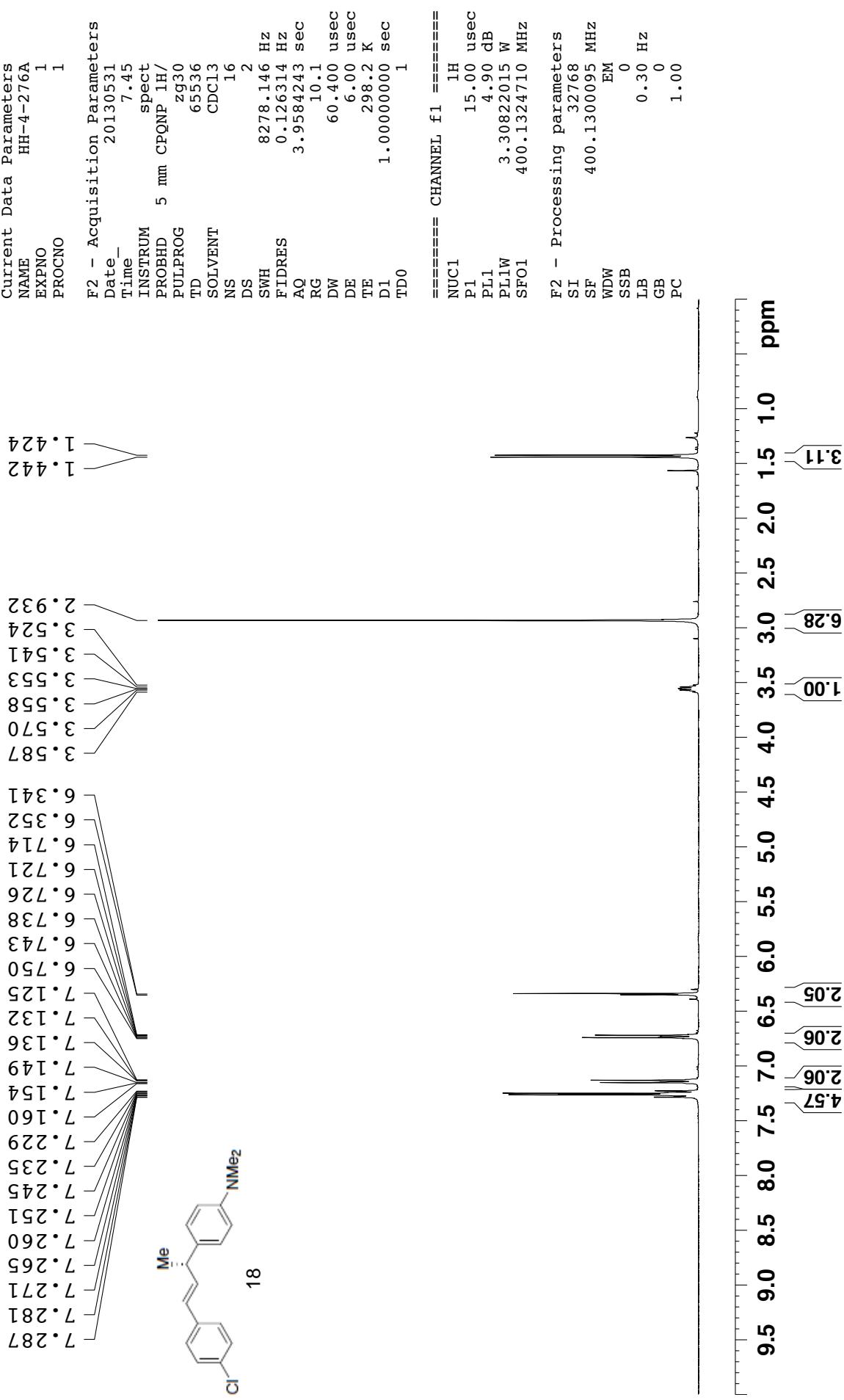


21.24
42.12

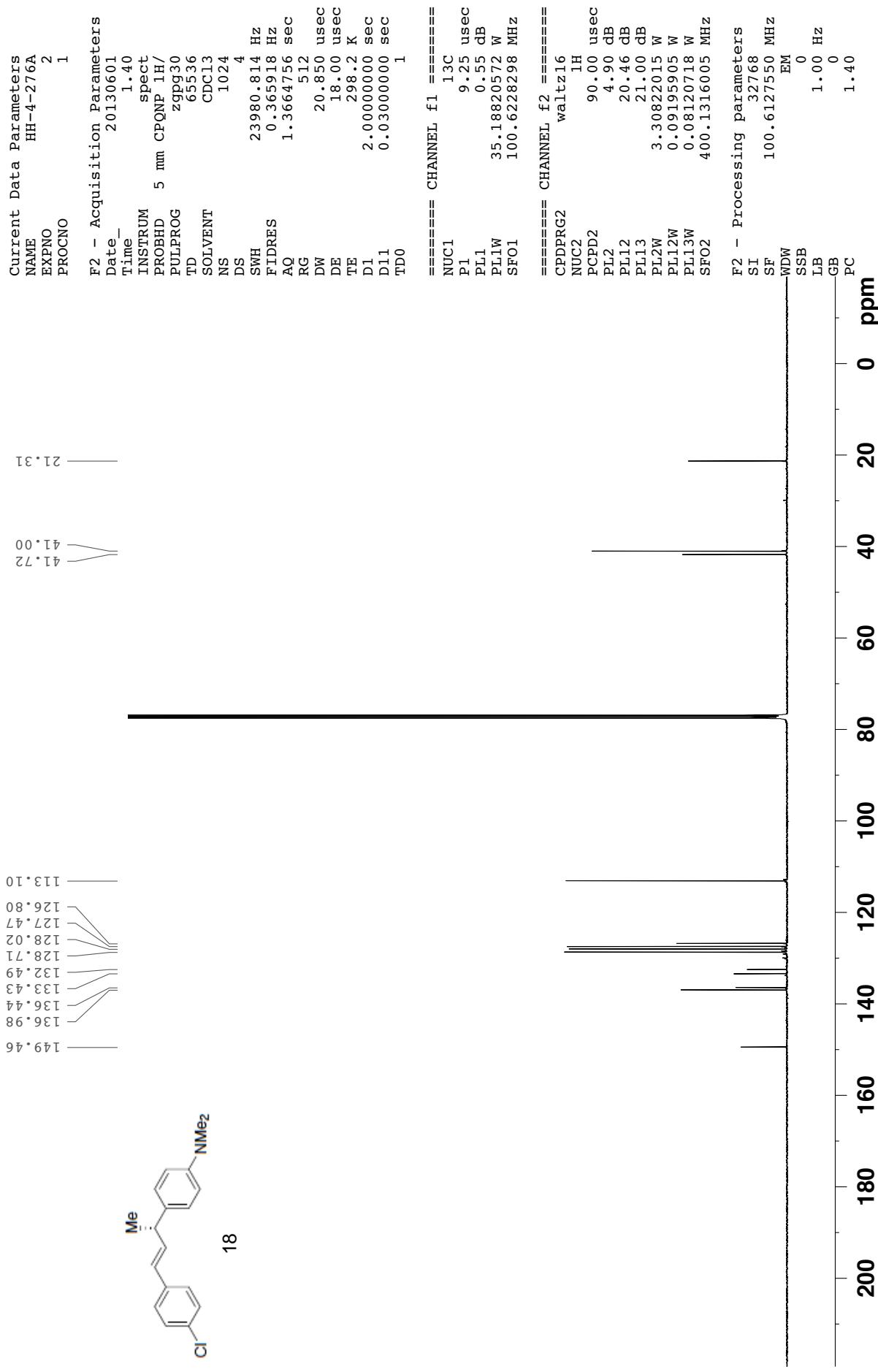




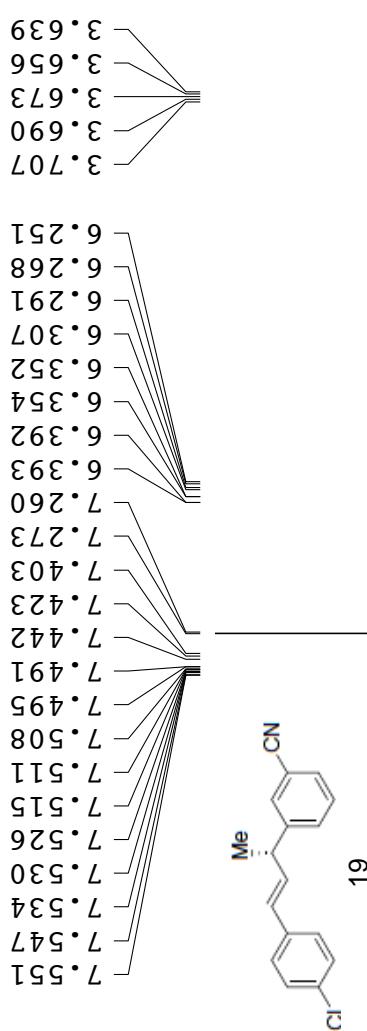
compound 18



compound 18



compound 19



1.464
1.481

3.639
3.656
3.673
3.690
3.707

6.251
6.268
6.291
6.307
6.352
6.392
6.393
6.395
6.396
6.397
6.398
6.399
6.403
6.423
6.442
7.491
7.495
7.508
7.511
7.515
7.526
7.530
7.534
7.541
7.551

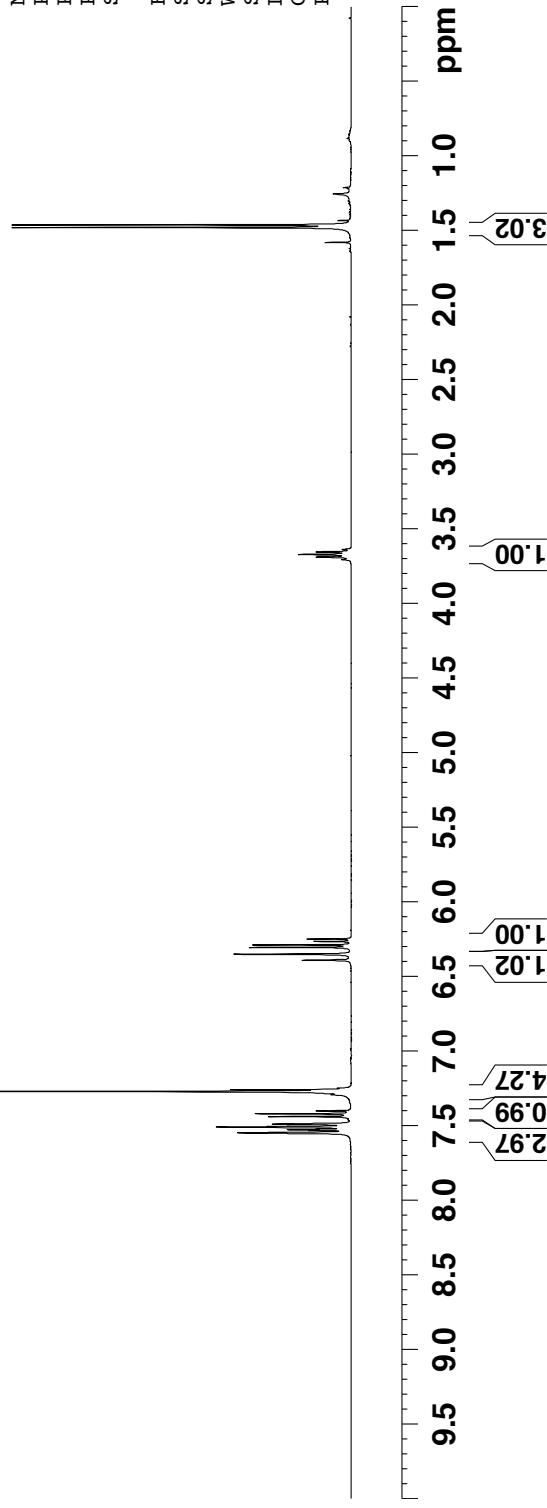
```

=====
F2 - Acquisition Parameters
NAME      HH-4-256B
EXPNO     1
PROCNO    1
Date      20130513
Time      1.28
INSTRUM   spect
PROBHD   5 mm CPQNP 1H/
PULPROG  zg30
TD       65536
SOLVENT   CDCl3
NS        16
DS         2
SWH      8278.146 Hz
FIDRES   0.126314 Hz
AQ        3.9584243 sec
RG        11.3
DW       60.400 usec
DE        6.00 usec
TE        298.2 K
D1      1.0000000 sec
TD0           1

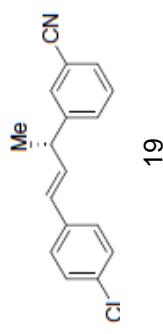
===== CHANNEL f1 =====
NUC1      1H
P1        15.00 usec
PL1      4.90 dB
PL1W
SFO1    400.1324710 MHz

F2 - Processing parameters
SI        32768
SF      400.1300093 MHz
WDW
SSB
LB        0 Hz
GB        0
PC        1.00

```

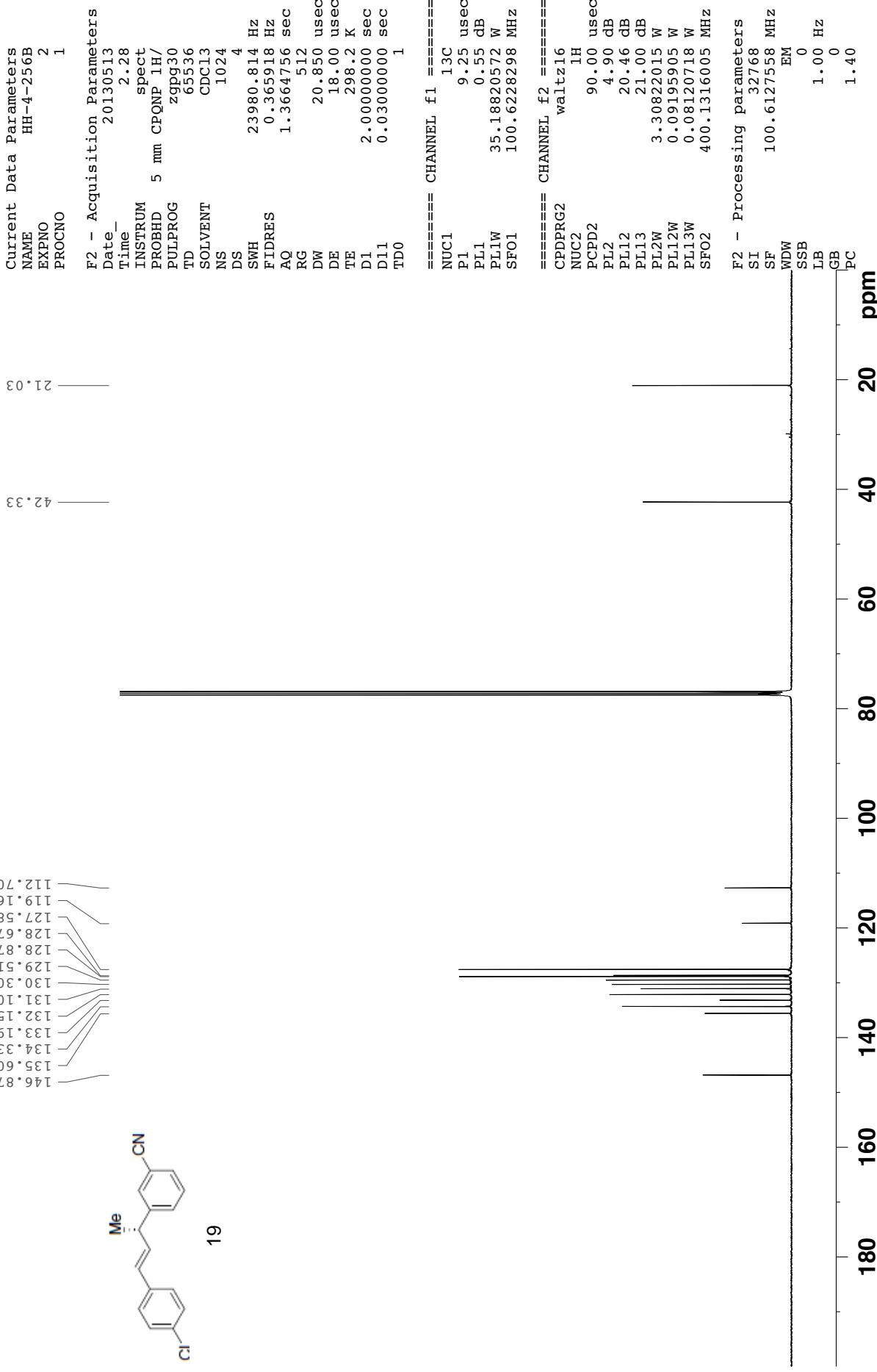


compound 19



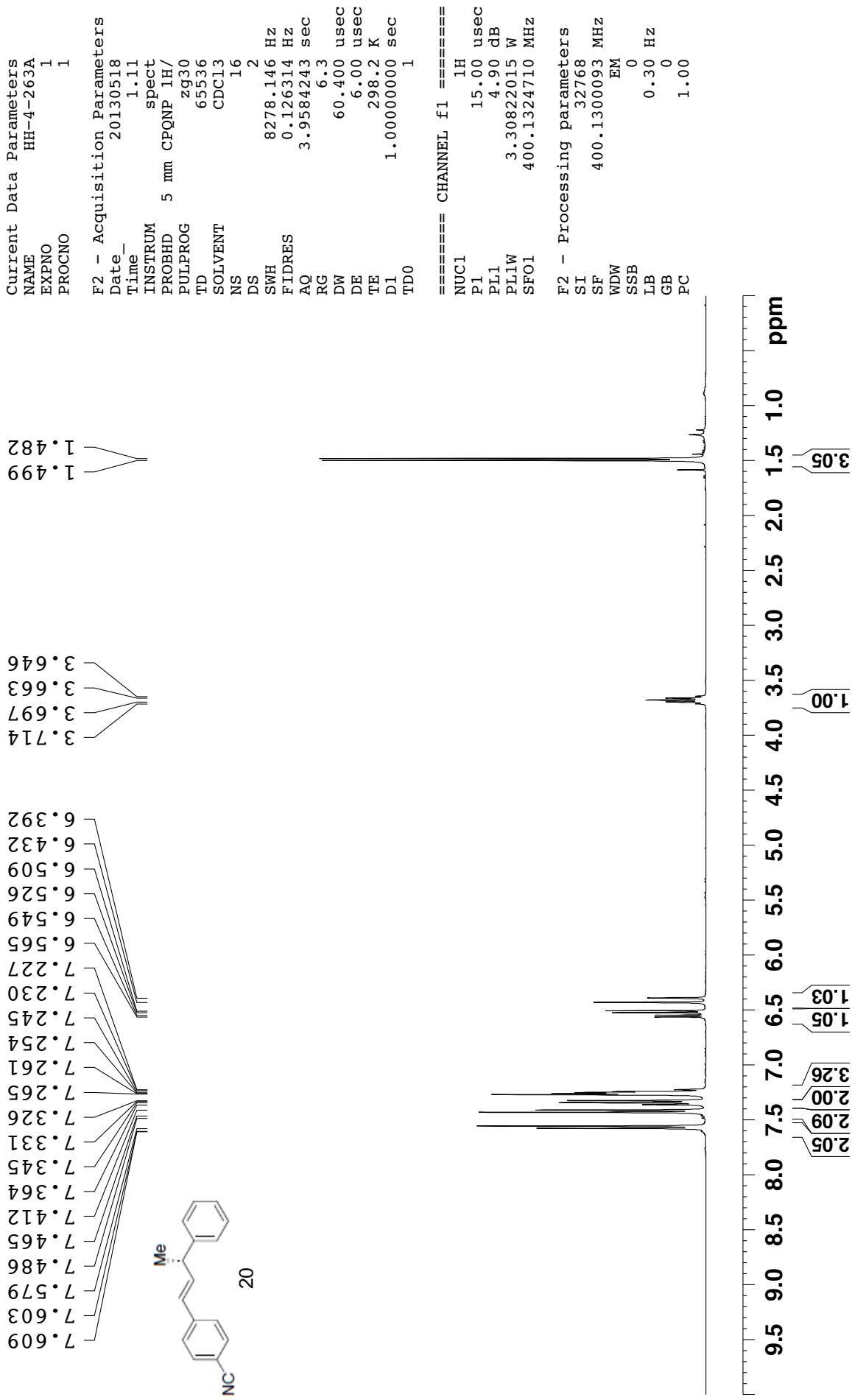
146.87
135.60
134.33
133.19
132.15
131.10
130.30
129.51
128.87
128.67
127.58
127.16
119.16
112.70

21.03
42.33



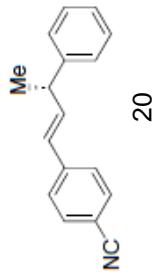
BRUKER

compound 20



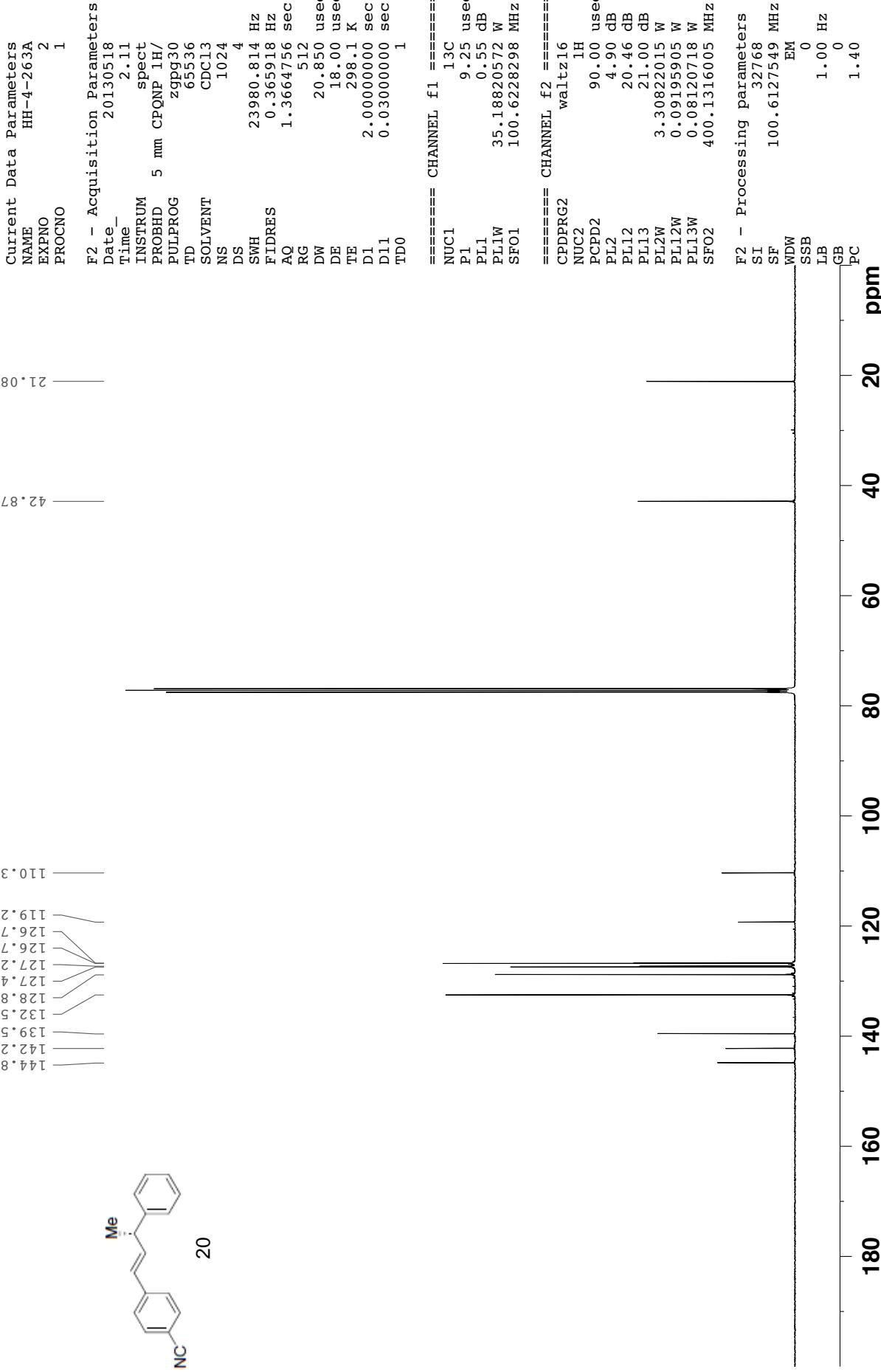


compound 20



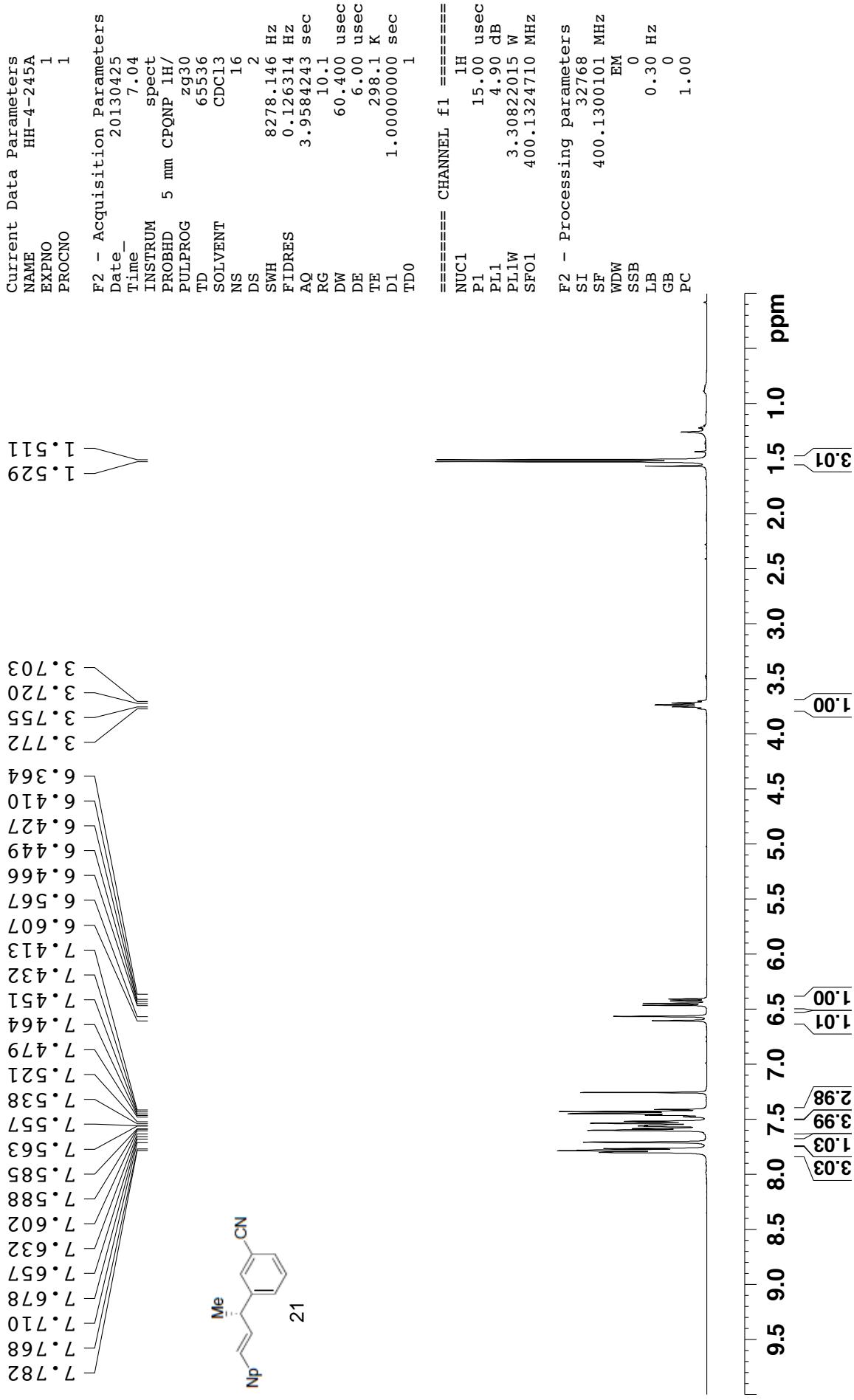
144.84
142.24
139.57
132.51
128.82
127.45
127.29
126.79
126.71
126.29
110.35

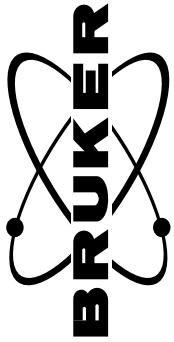
21.08
42.87



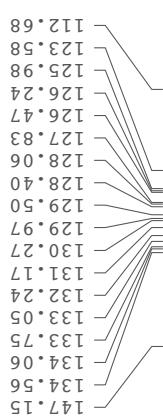
BRUKER

compound 21

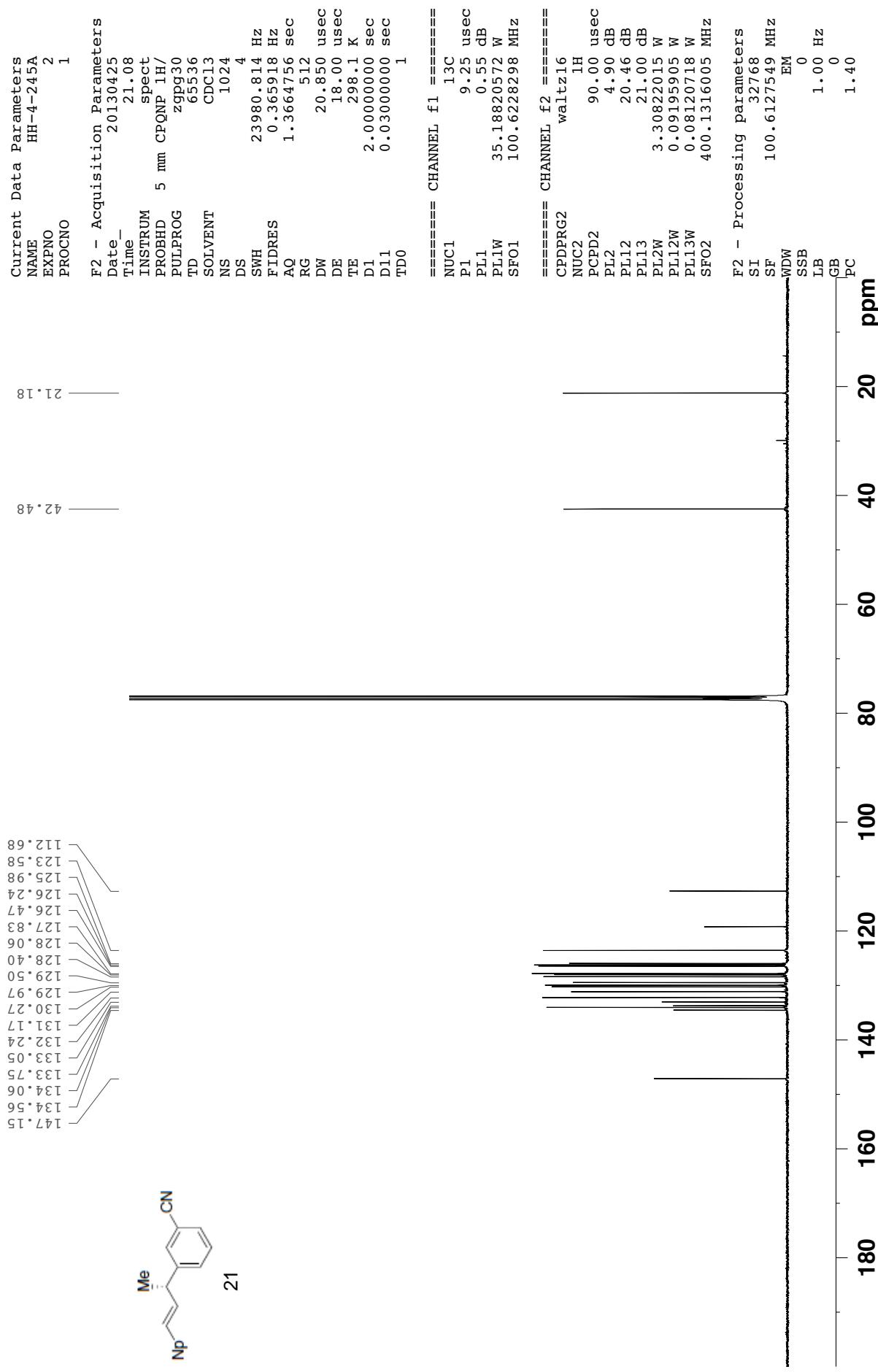




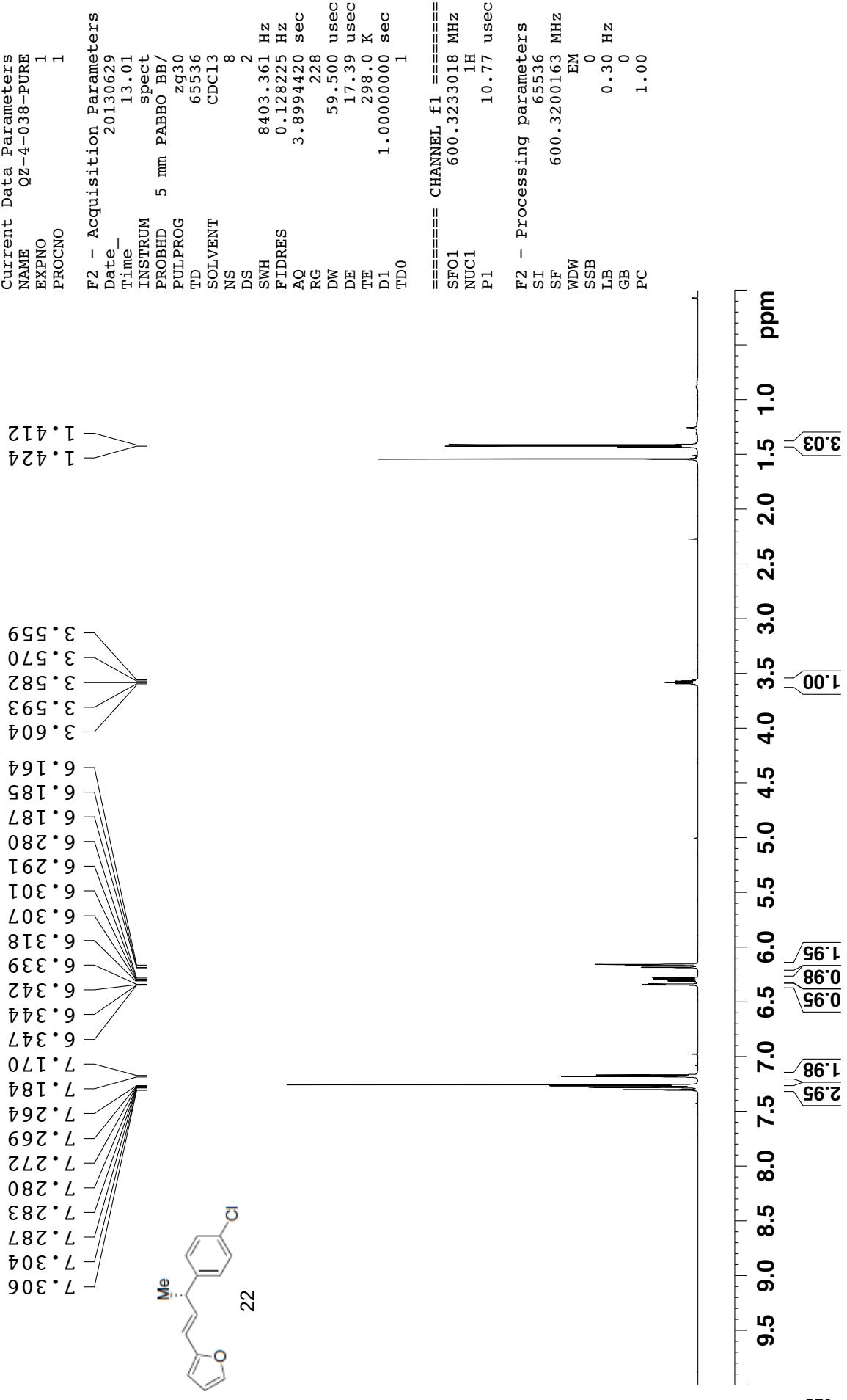
compound 21



21



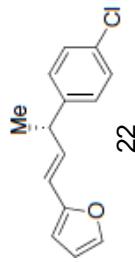
compound 22



compound 22



153.04
143.98
141.72
133.67
132.11
128.88
128.76
117.86
111.37
107.15

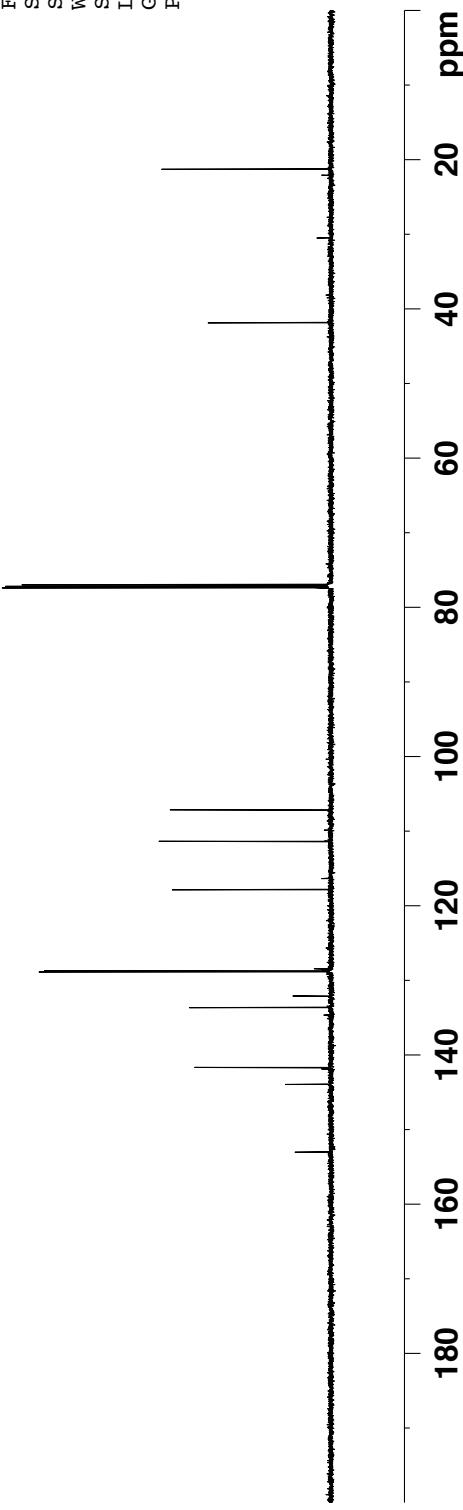


Current Data Parameters
 NAME QZ-4-038
 EXPNO 2
 PROCNO 1

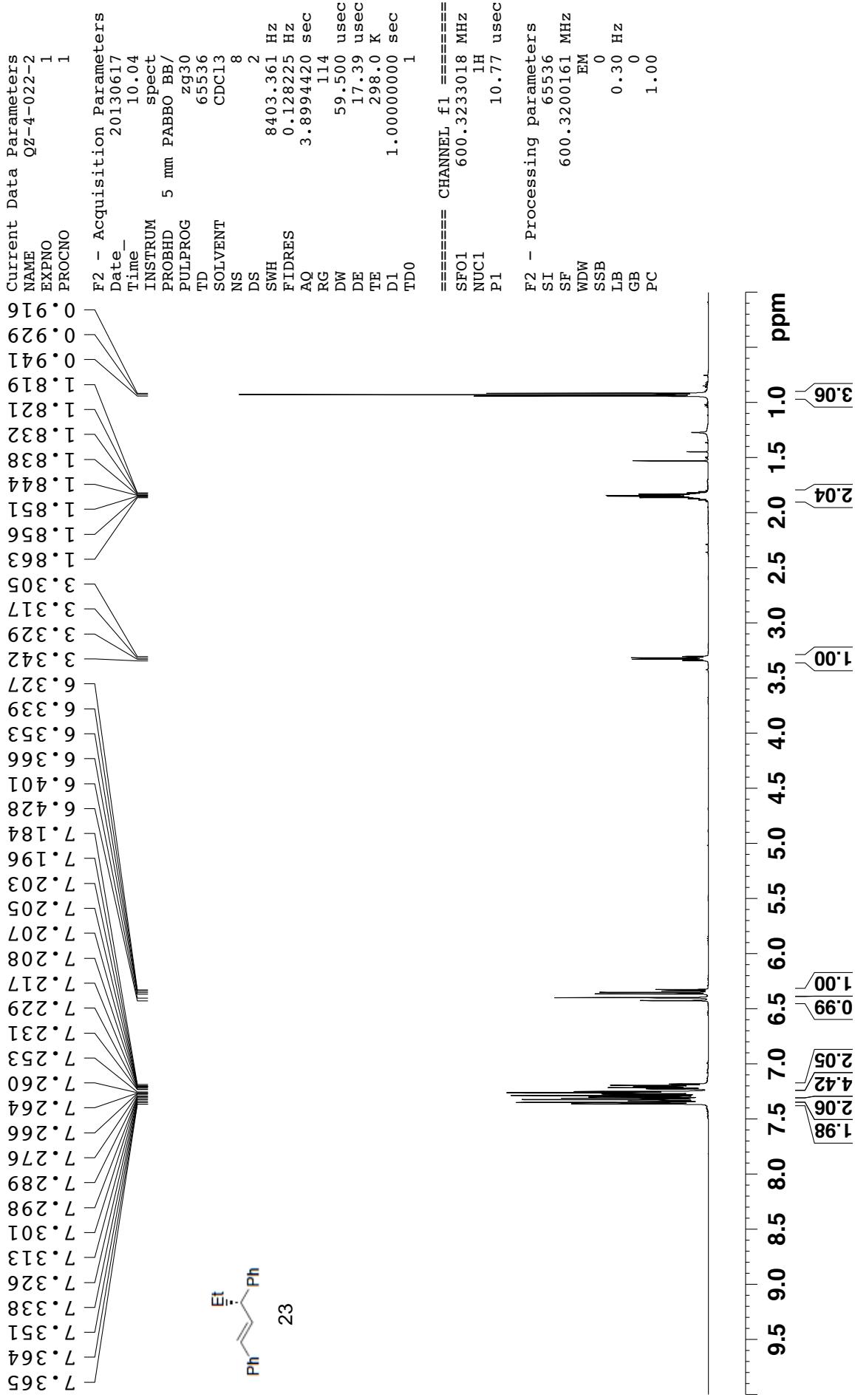
F2 - Acquisition Parameters
 Date 20130628
 Time 16.10
 INSTRUM spect
 PROBHD 5 mm PABBO BB/
 PULPROG zgpg55
 TD 65536
 SOLVENT CDCl3
 NS 256
 DS 4
 SWH 34722.223 Hz
 FIDRES 0.529819 Hz
 AQ 0.9437684 sec
 RG 2050
 DW 14.400 usec
 DE 19.34 usec
 TE 298.0 K
 D1 1.1000002 sec
 D11 0.03000000 sec
 TDO 1

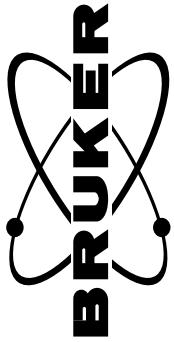
===== CHANNEL f1 ======
 SFO1 150.9656784 MHz
 NUC1 13C
 P1 10.63 usec

F2 - Processing parameters
 SI 32768
 SF 150.9505603 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40



compound 23

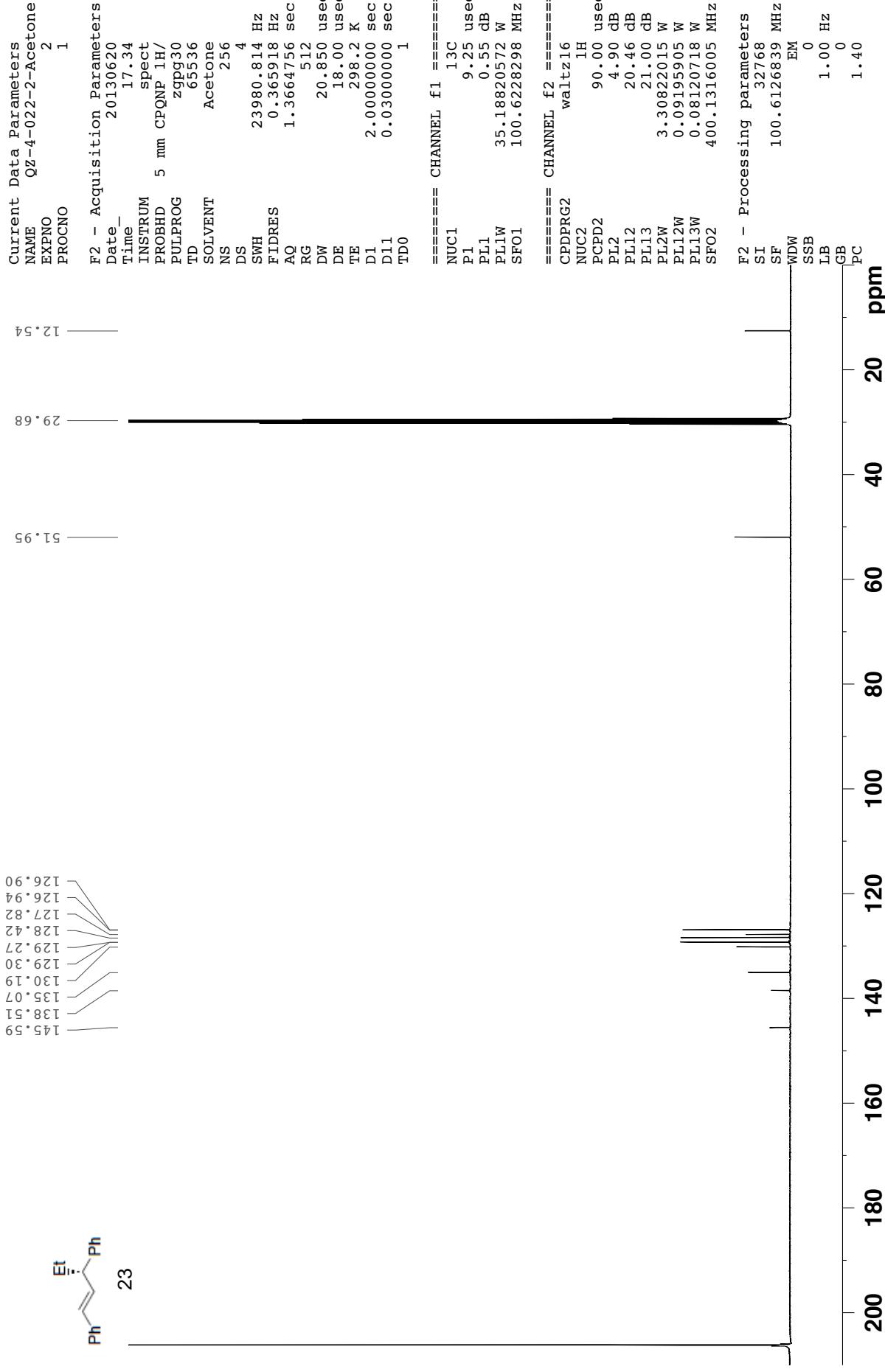




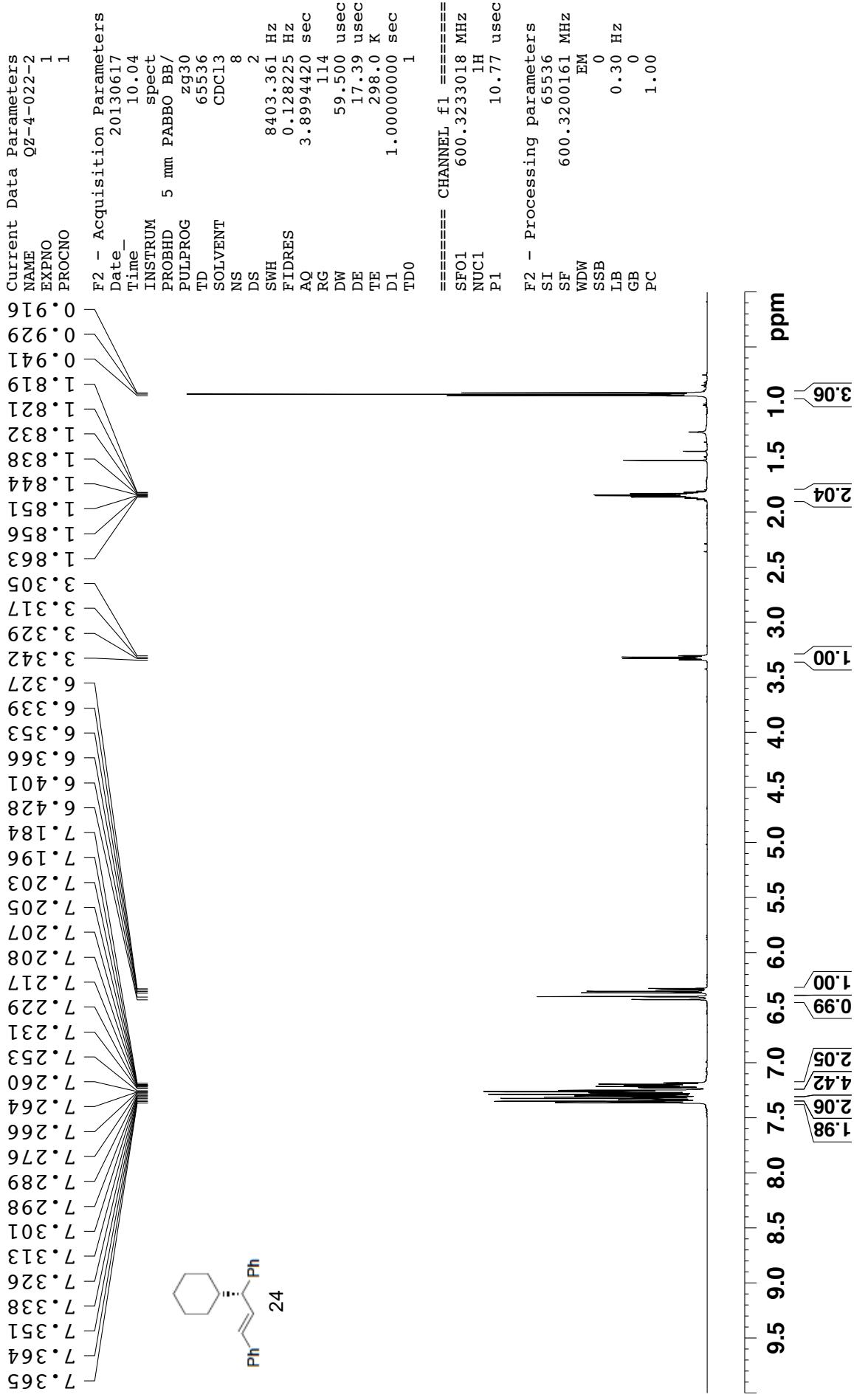
compound 23



145.59
 138.51
 135.07
 130.19
 129.30
 129.27
 128.42
 127.82
 126.94
 126.90

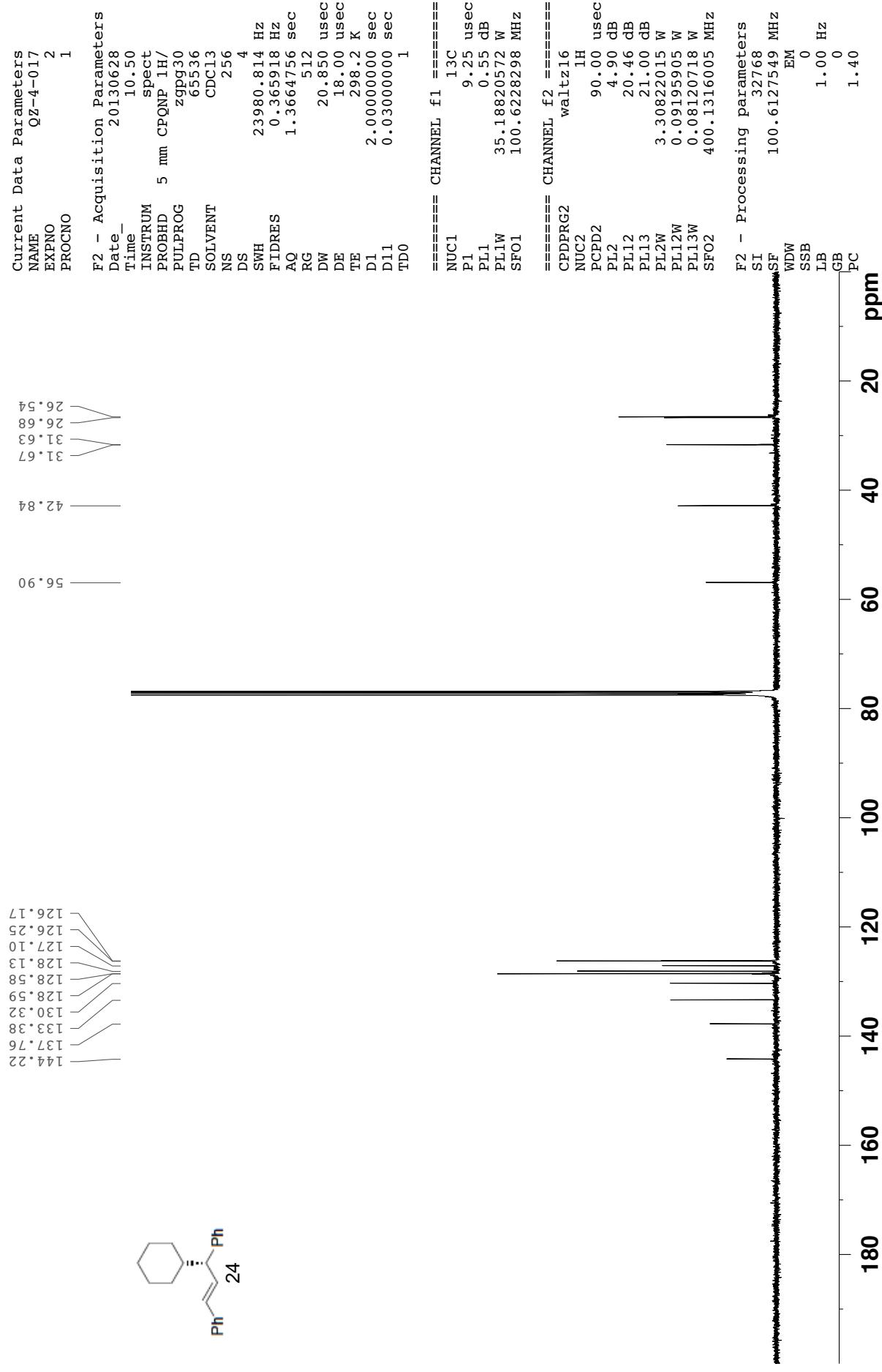


compound 24



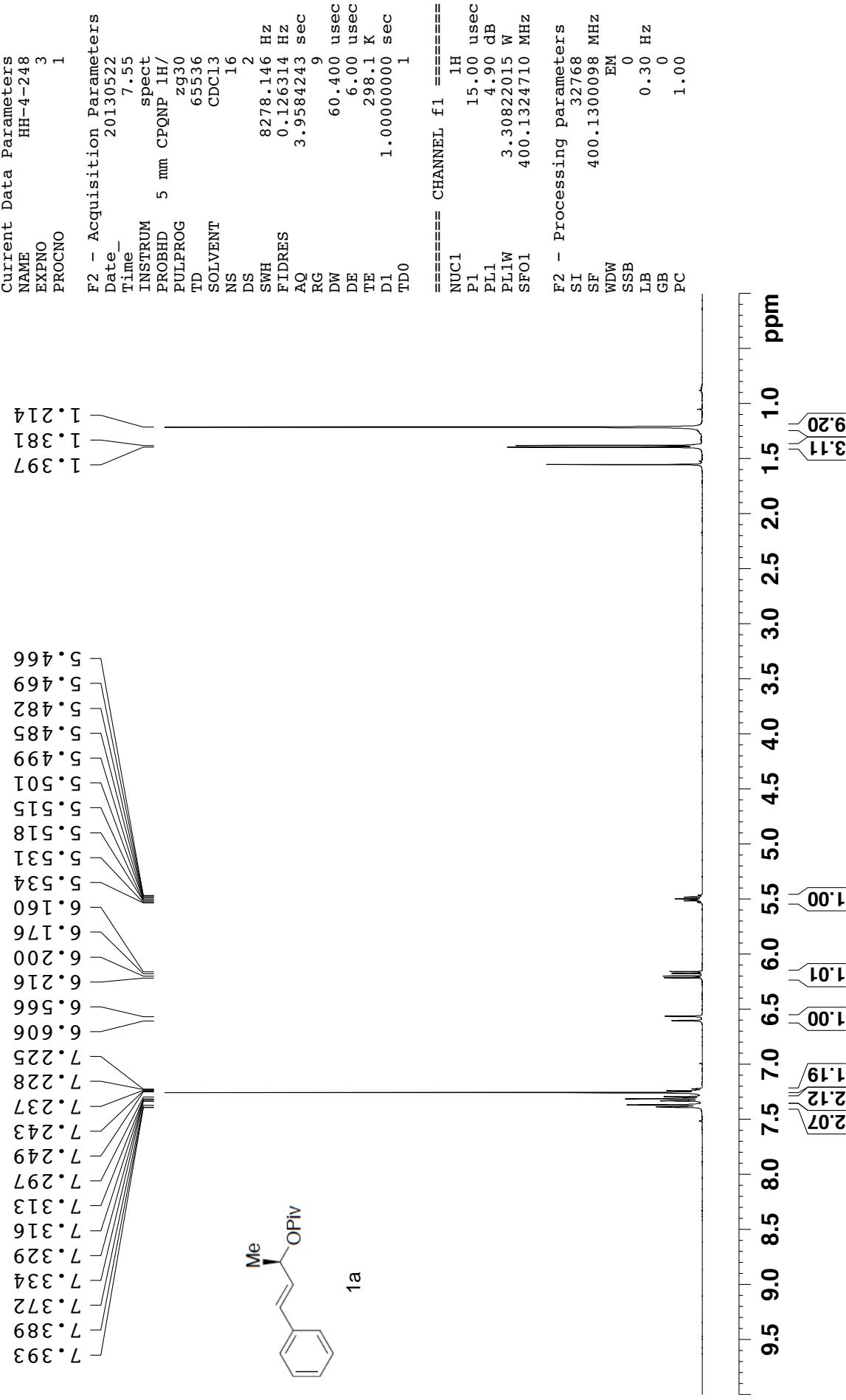


compound 24

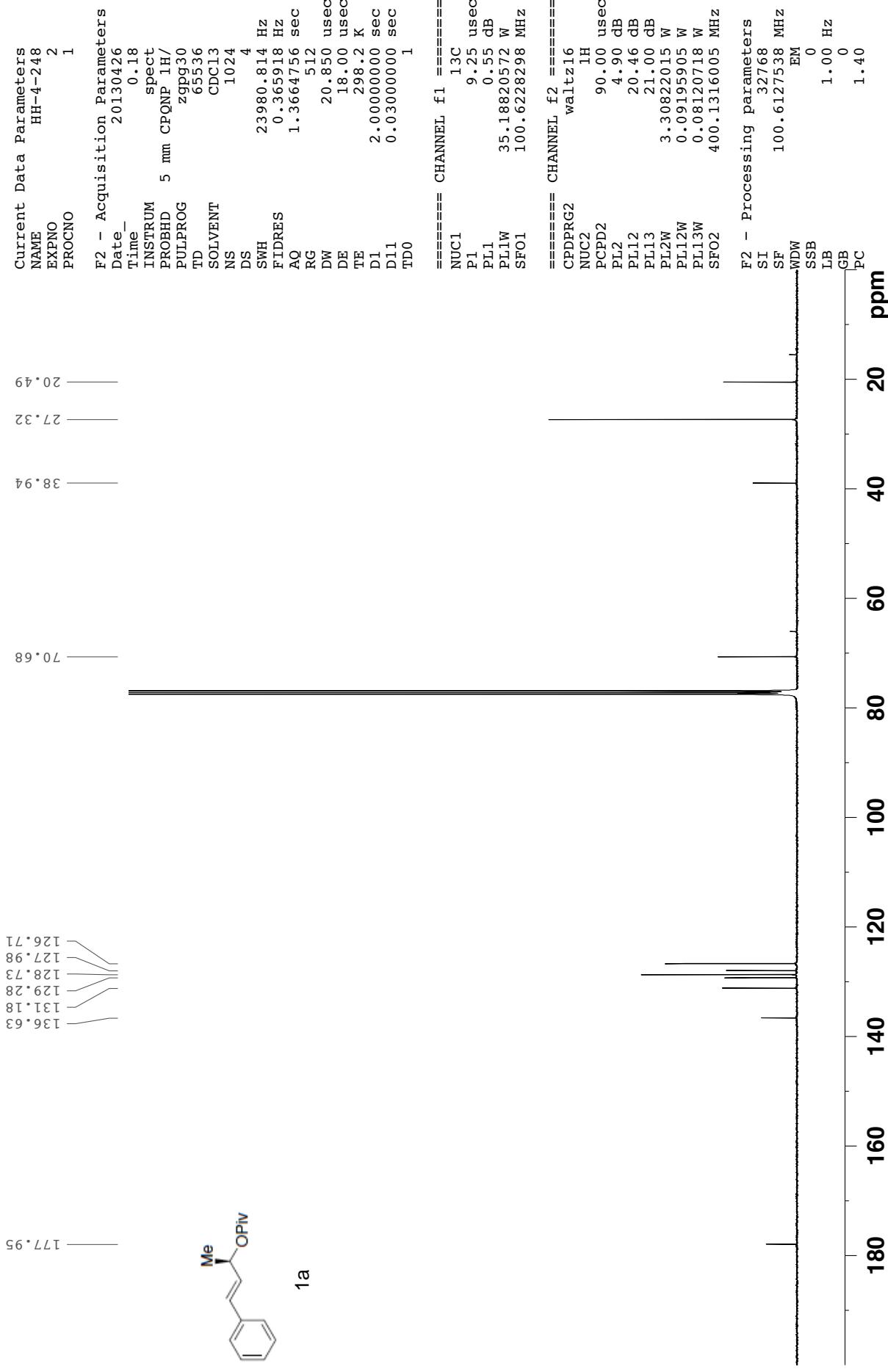




compound 1a

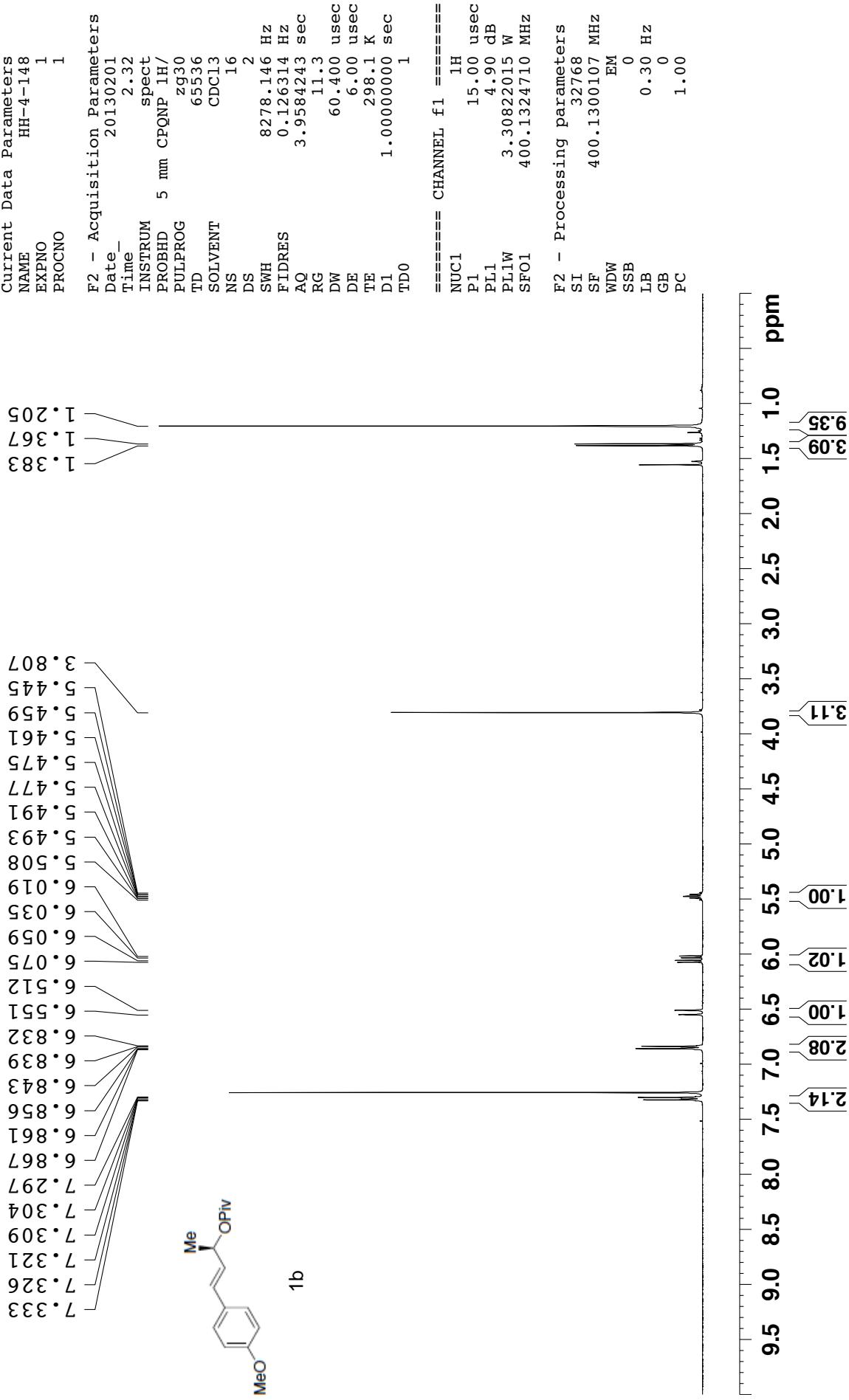


compound 1a



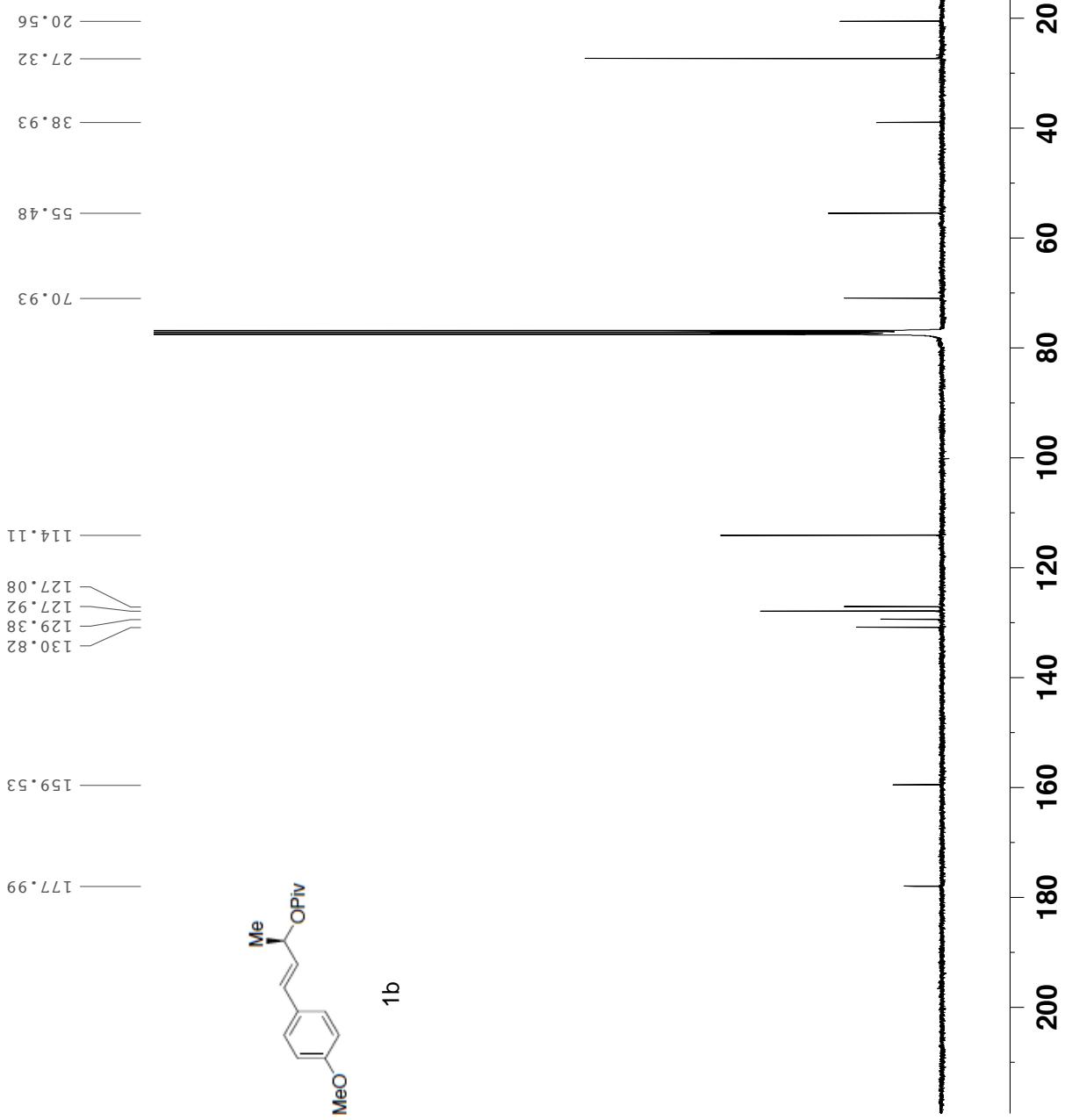
The Bruker logo consists of the word "BRUKER" in a bold, black, sans-serif font, oriented vertically. The letters are partially enclosed by two stylized, intersecting arcs that resemble atomic orbits or magnetic field lines.

Compound 1b



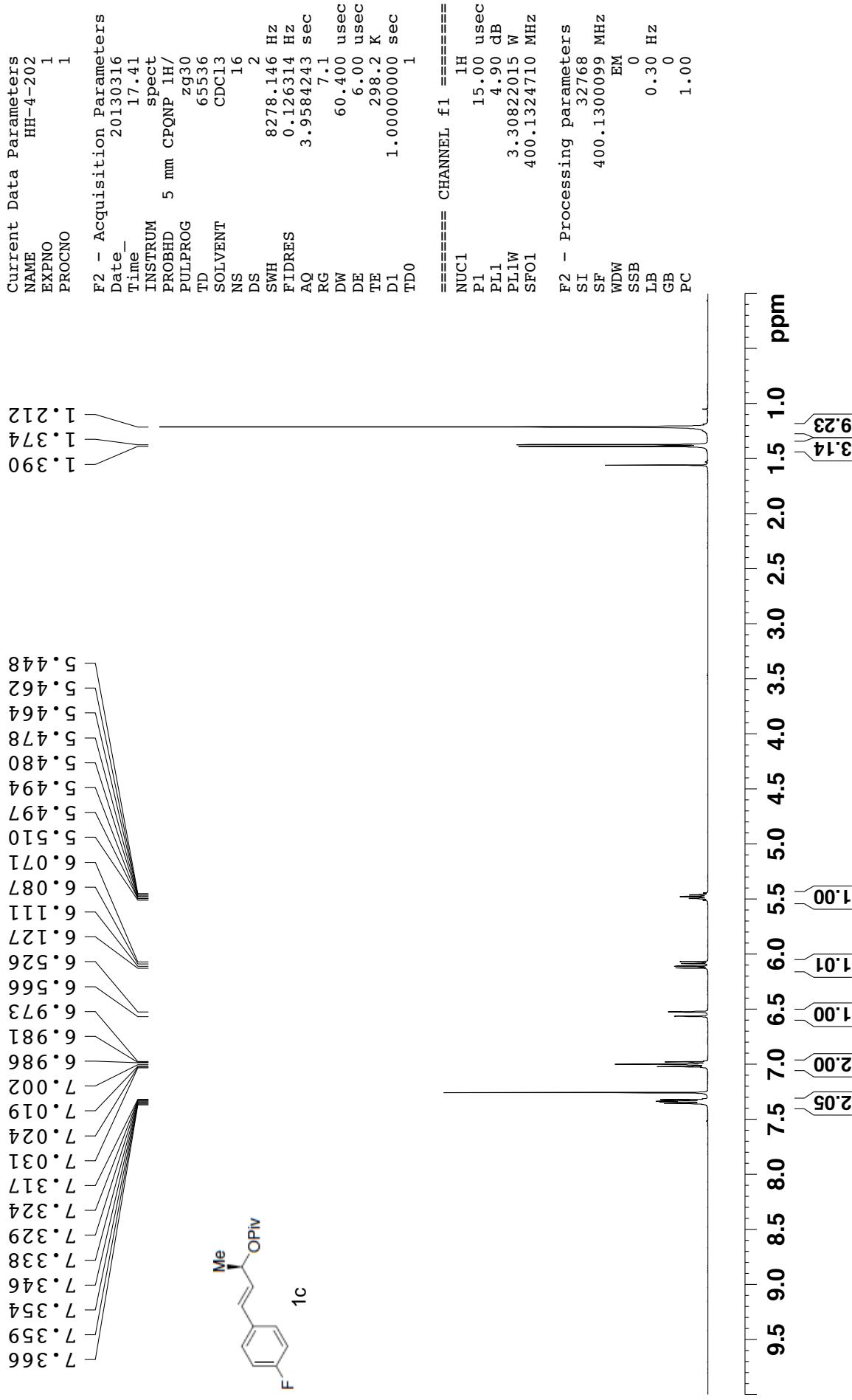


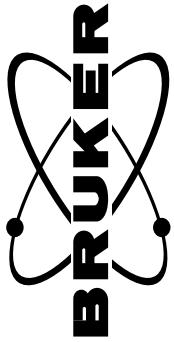
compound 1b



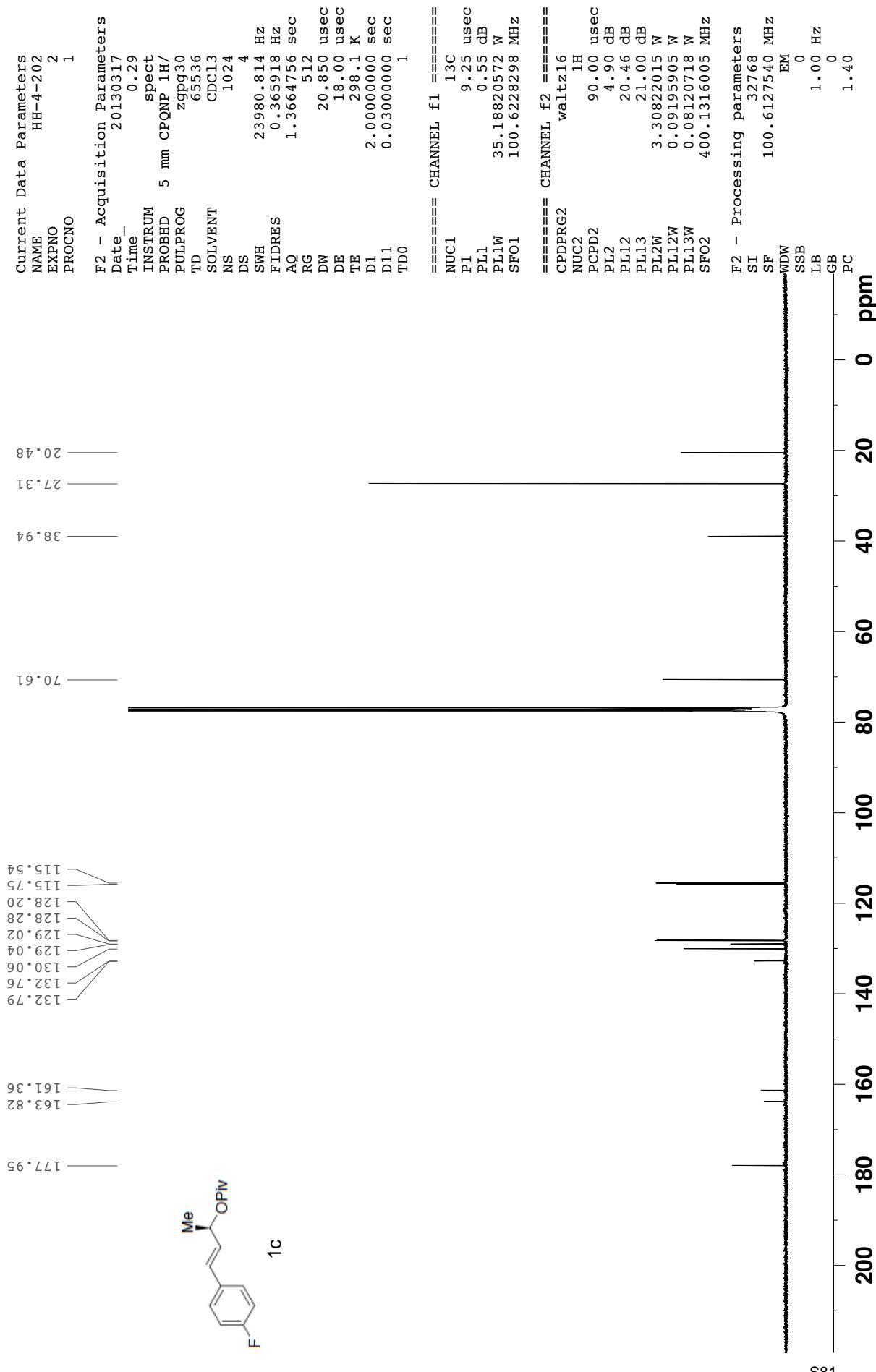
The Bruker logo consists of the word "BRUKER" in a bold, black, sans-serif font, oriented vertically. The letters are partially enclosed by two stylized, intersecting arcs that resemble atomic orbits or magnetic field lines.

compound 1c



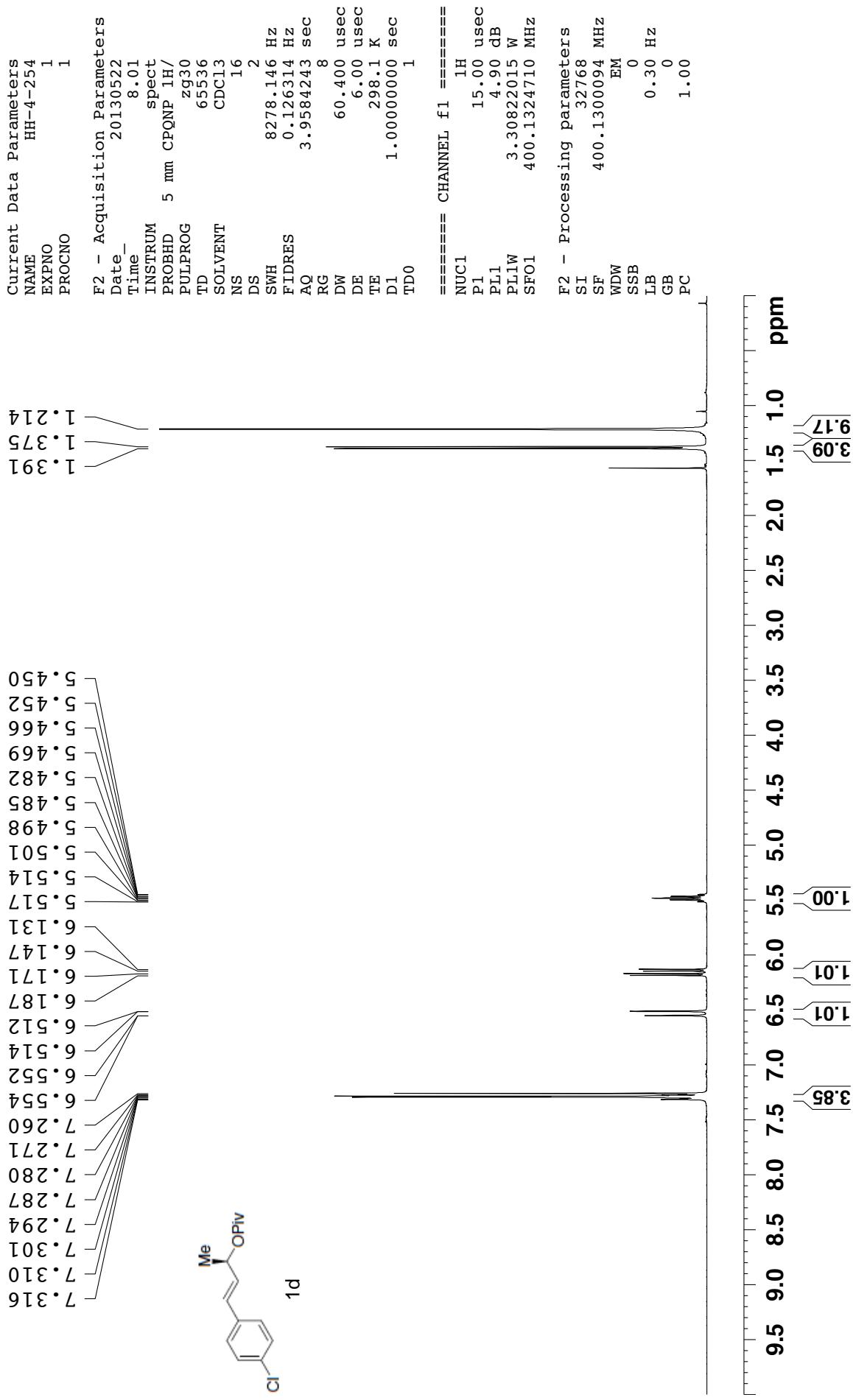


compound 1c



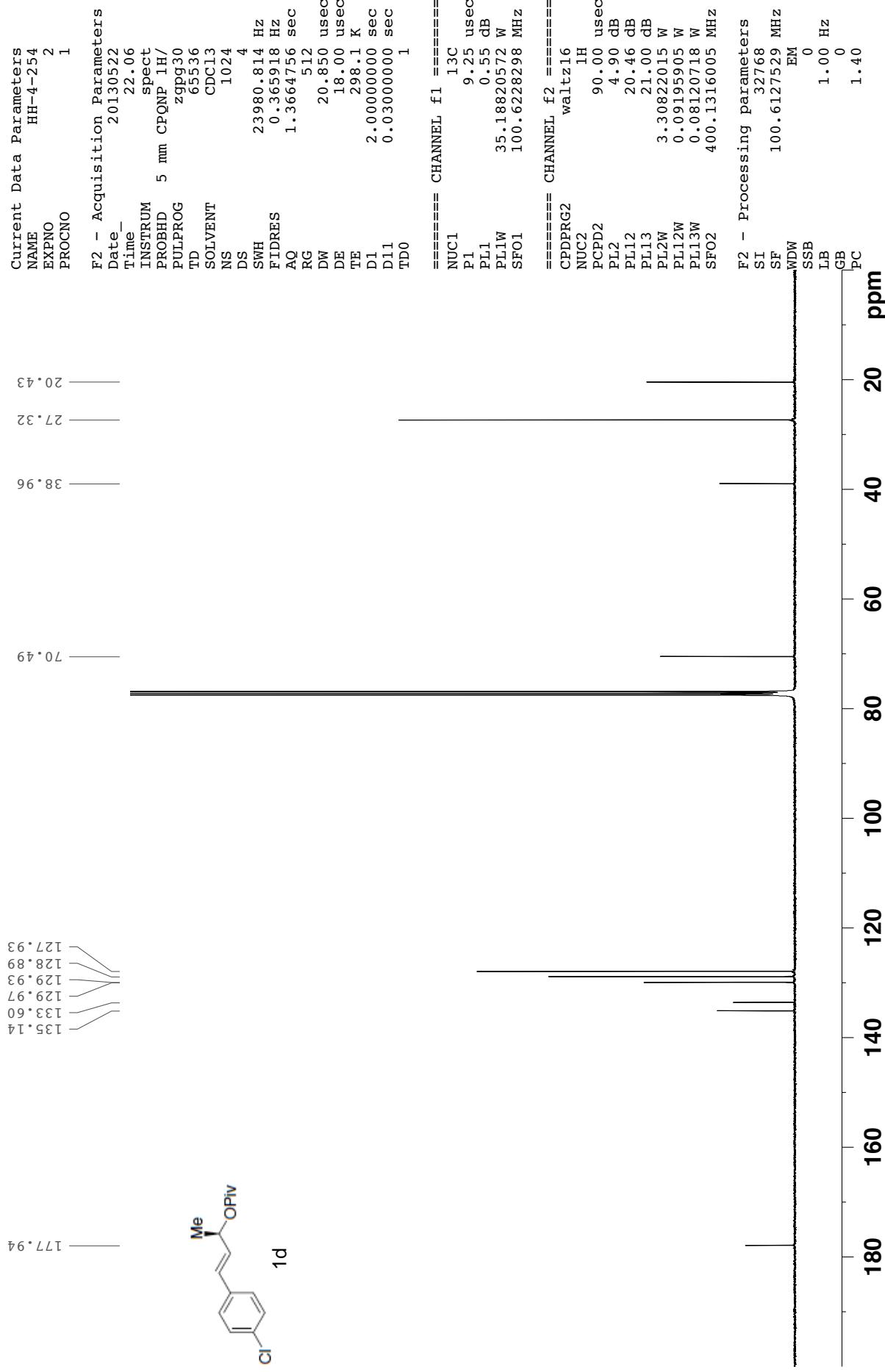


compound 1d



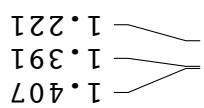
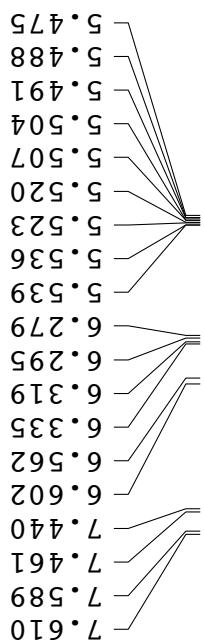
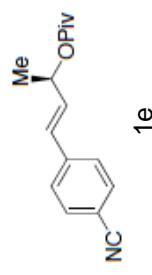


compound 1d



The Bruker logo consists of the word "BRUKER" in a bold, black, sans-serif font, oriented vertically. The letters are partially enclosed by two stylized, intersecting arcs that resemble atomic orbits or magnetic field lines.

Compound 1e



```

Current Data Parameters          FF2 - Acquisition Parameters
NAME      HH-4-227               Date_    20130522
EXPNO     1                      Time_   5.43
PROCNO    1

INSTRUM  PROBHD                mm      CPQNPH
         PULPROG              Zg30
         TD                   65536
         SOLVENT              CDC13
         NS                  16
DS        2
SWH      8278.146 Hz
        0.126314 Hz
        3.9584243 sec
FIDRES
AQ
AQ
RG      9
DW      60.400 usec
DE      6.00  usec
TE      298.1 K
TD1    1.00000000 sec
TDDO

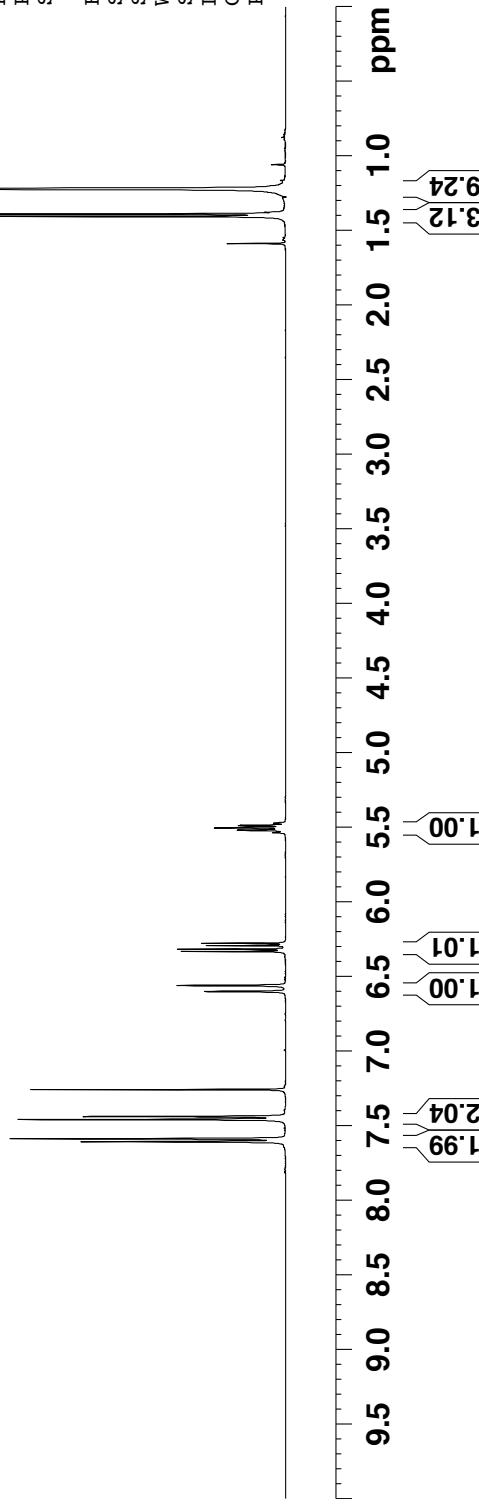
```

```

===== CHANNEL f1 =====
NUC1          1H
P1           15.00 usec
PL1          4.90 dB
PPL1W        3.30822015 W
SFO1         400.1324710 MHz

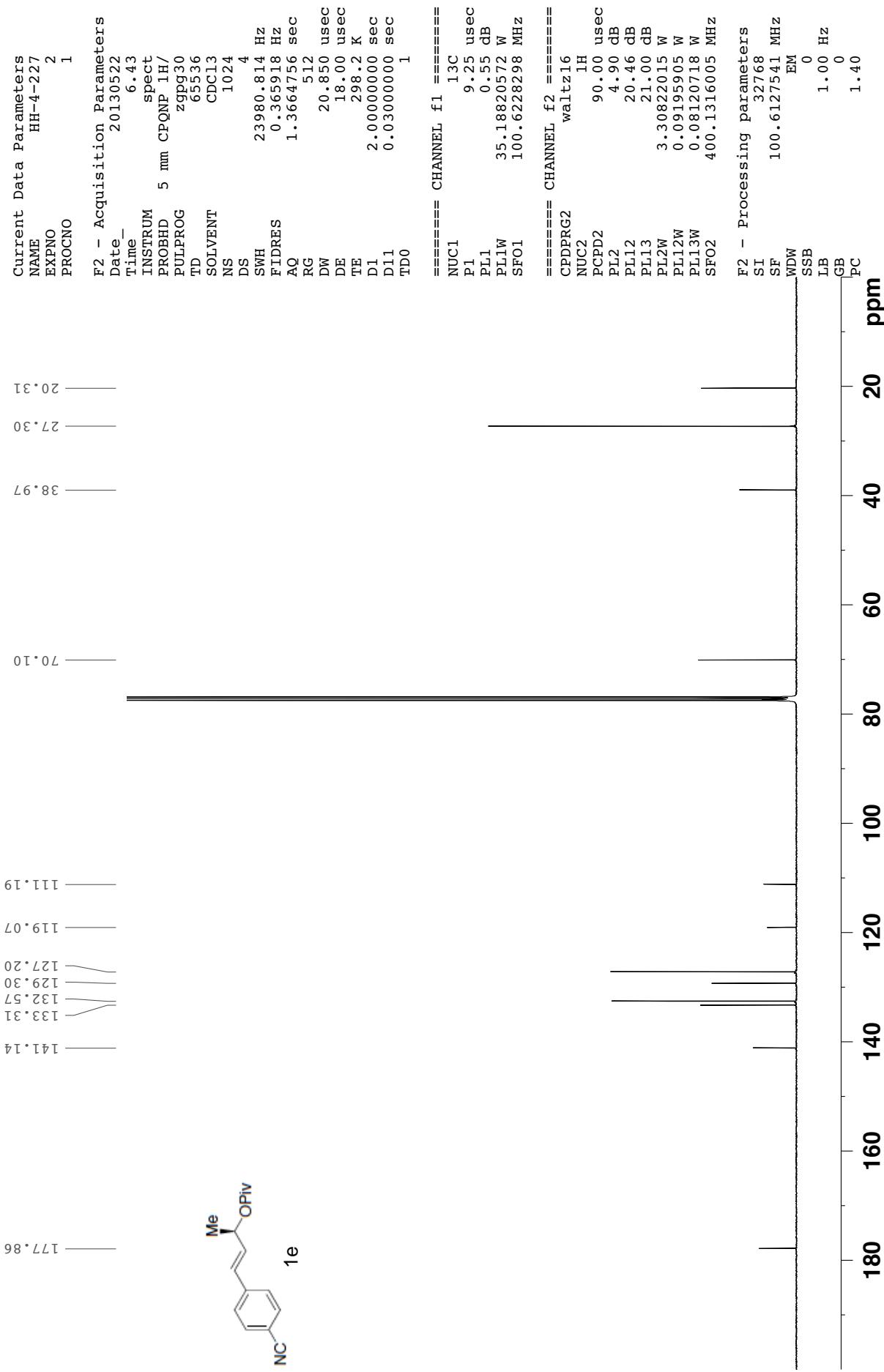
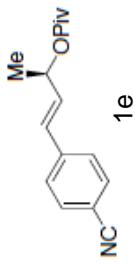
F2 - Processing parameters
SI           32768
SF          400.1300094 MHz
WDW
SSB          0
LB           0.30 Hz
GB
PC

```



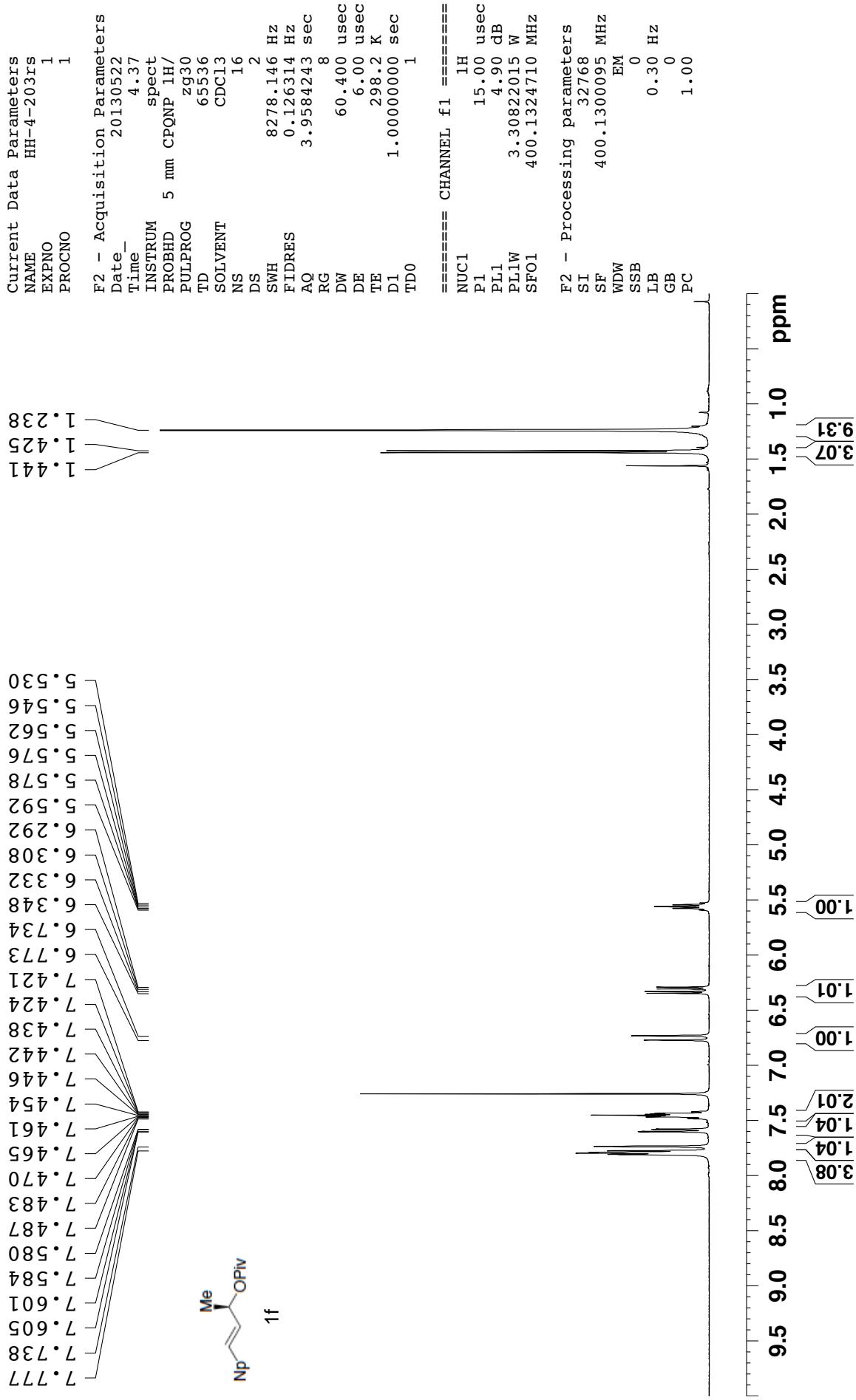
BRUKER

Compound 1e



BRUKER

Compound 1f



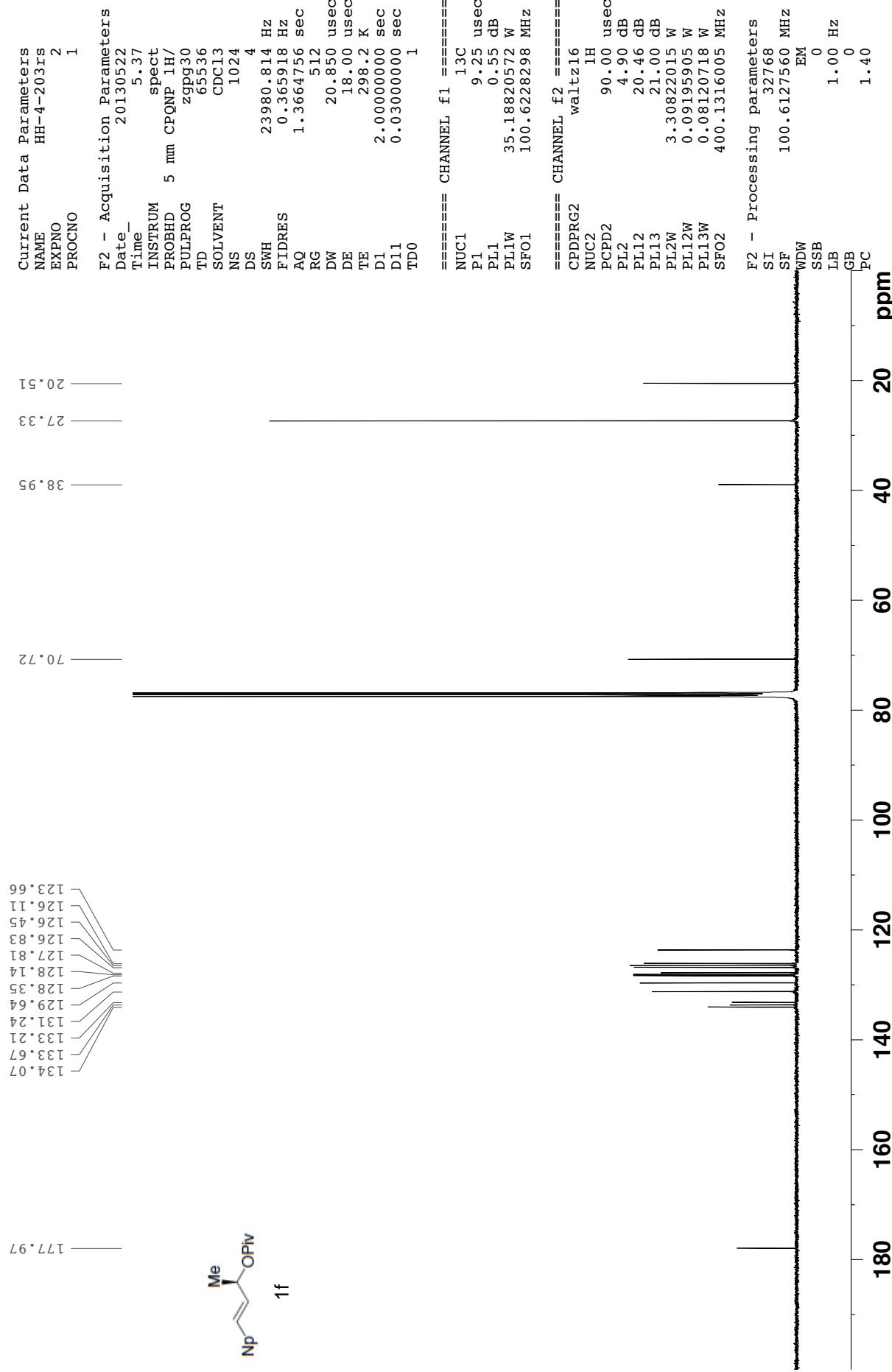
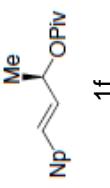


compound 1f

134.07
133.67
133.21
131.24
129.64
128.35
128.14
127.81
126.83
126.45
126.11
123.66

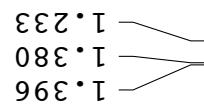
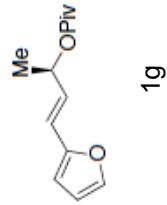
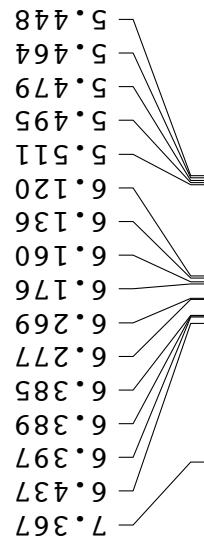
38.95
27.33
20.51

70.72





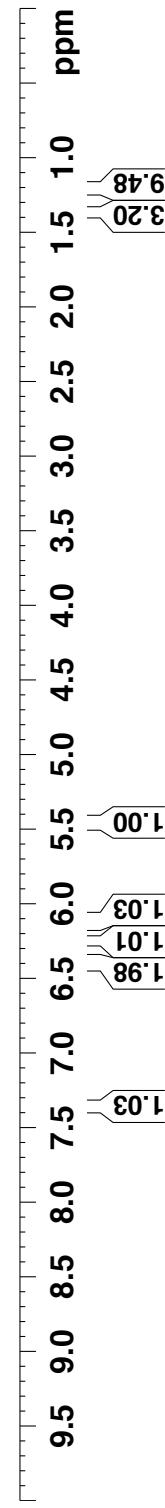
compound 1g



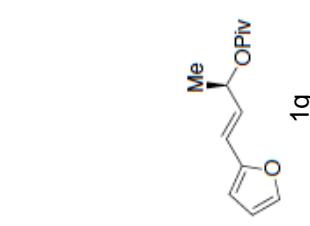
Current Data Parameters
NAME QZ-3-242-1-RAC
EXPNO 1
PROCNO 1

F2 - Acquisition Parameters
Date 20130422
Time 13.33
INSTRUM spect
PROBHD 5 mm CPQNP 1H/
PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 16
DS 2
SWH 8278.146 Hz
FIDRES 0.126314 Hz
AQ 3.9584243 sec
RG 5.7
DW 60.400 usec
DE 6.00 usec
TE 298.2 K
D1 1.0000000 sec
TDO 1

===== CHANNEL f1 ======
NUC1 1H
P1 15.00 usec
PL1 4.90 dB
PL1W 3.30822015 W
SFO1 400.1324710 MHz
F2 - Processing parameters
SI 32768
SF 400.1300095 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00



compound 1g



177.92
152.29
142.27
127.96
119.48
111.51
108.70

70.31
38.95
27.33
20.45

Current Data Parameters
NAME QZ-3-242-1
EXPNO 2
PROCNO 1

F2 - Acquisition Parameters
Date 20130620
Time 15.33
INSTRUM spect
PROBHD 5 mm PABBO BB/
PULPROG zgpg55
TD 65536
SOLVENT CDCl3
NS 256
DS 4
SWH 34722.223 Hz
FIDRES 0.529819 Hz
AQ 0.9437684 sec
RG 2050
DW 14.400 usec
DE 19.34 usec
TE 298.0 K
D1 1.1000002 sec
D11 0.03000000 sec
TDO 1

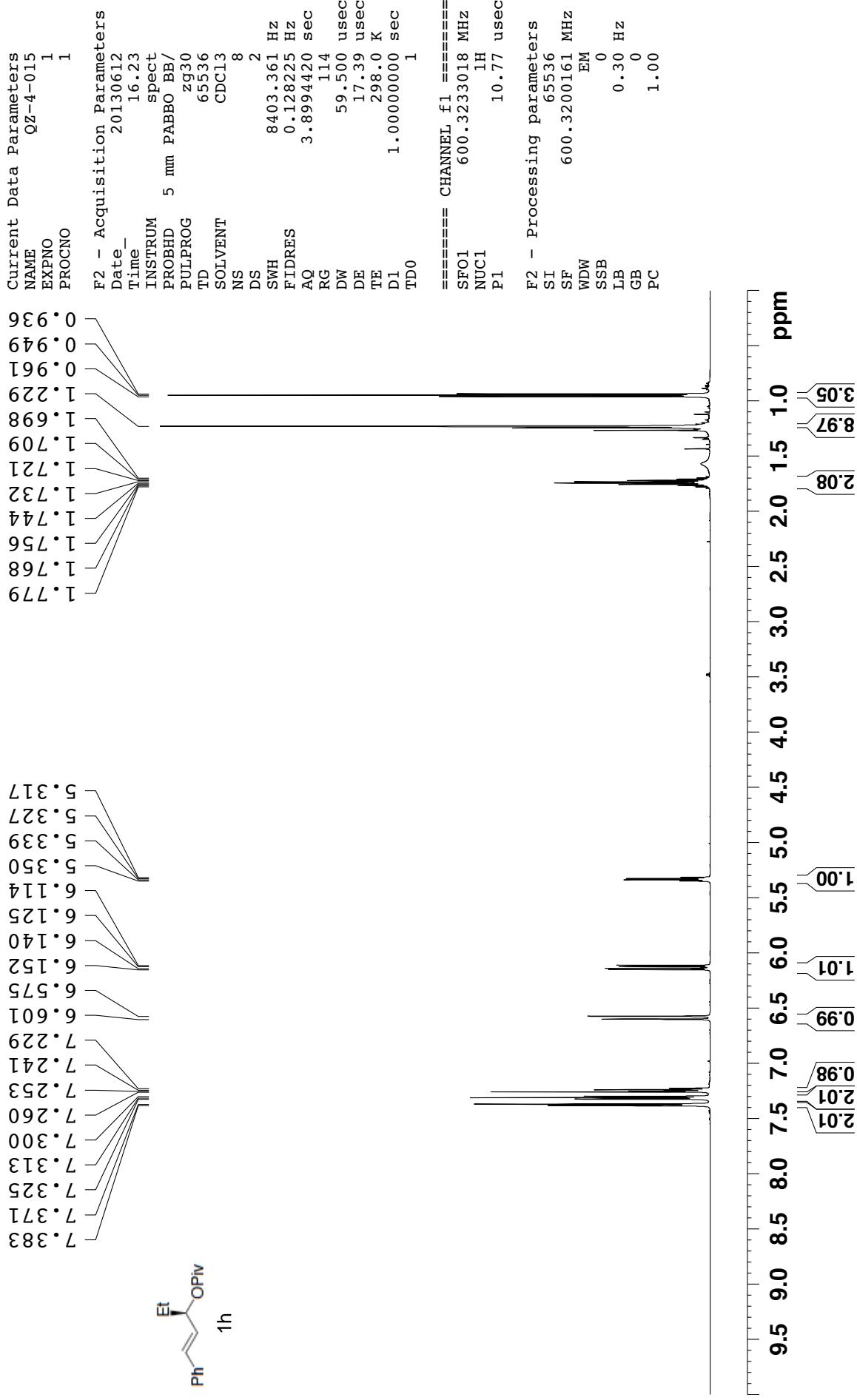
===== CHANNEL f1 =====
SFO1 150.9656784 MHz
NUC1 13C
P1 10.63 usec

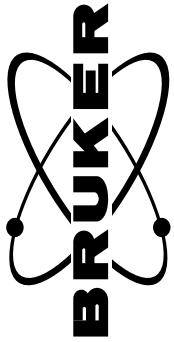
F2 - Processing parameters
SI 32768
SF 150.9505582 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

200 180 160 140 120 100 80 60 40 20 0 ppm

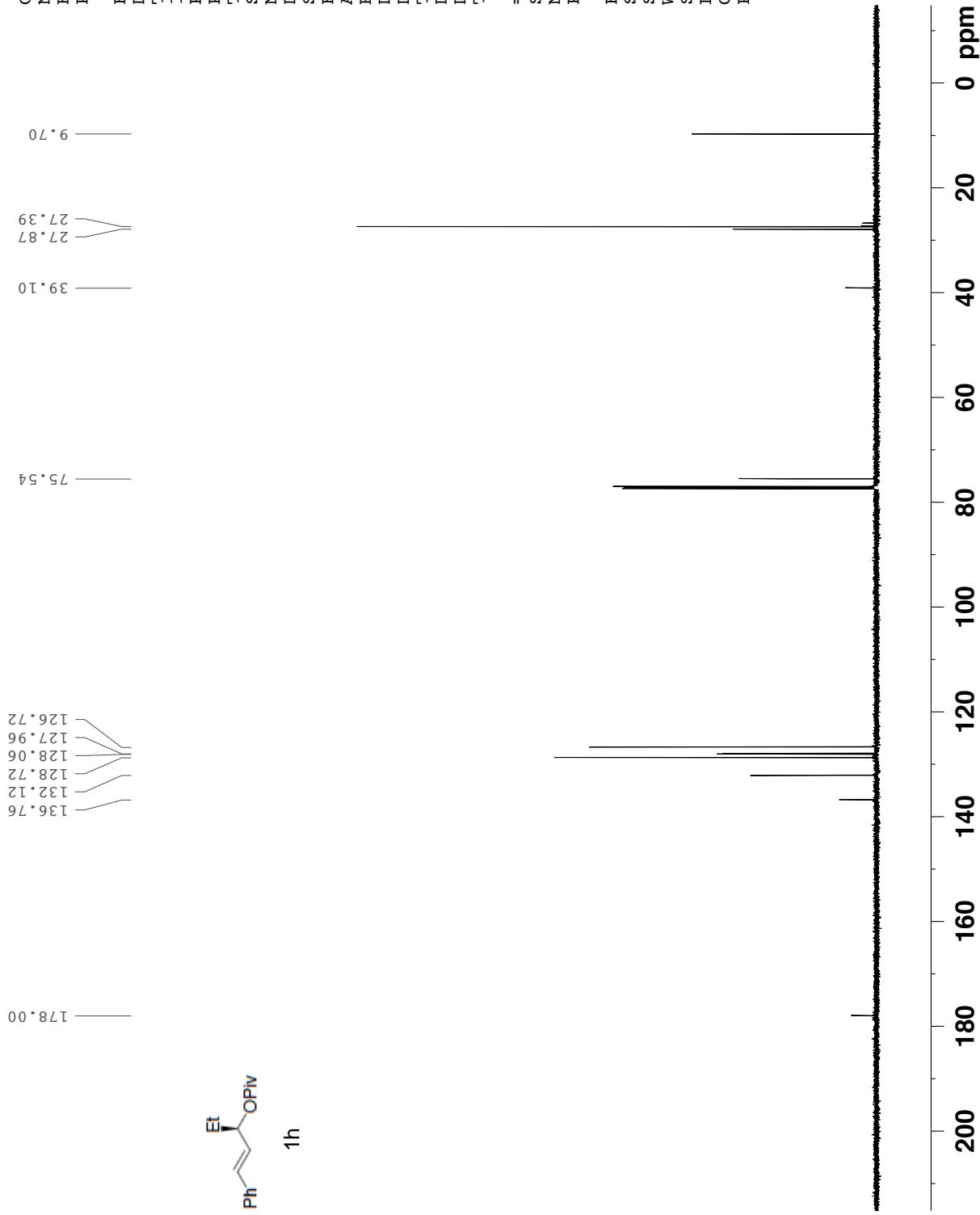


compound 1h



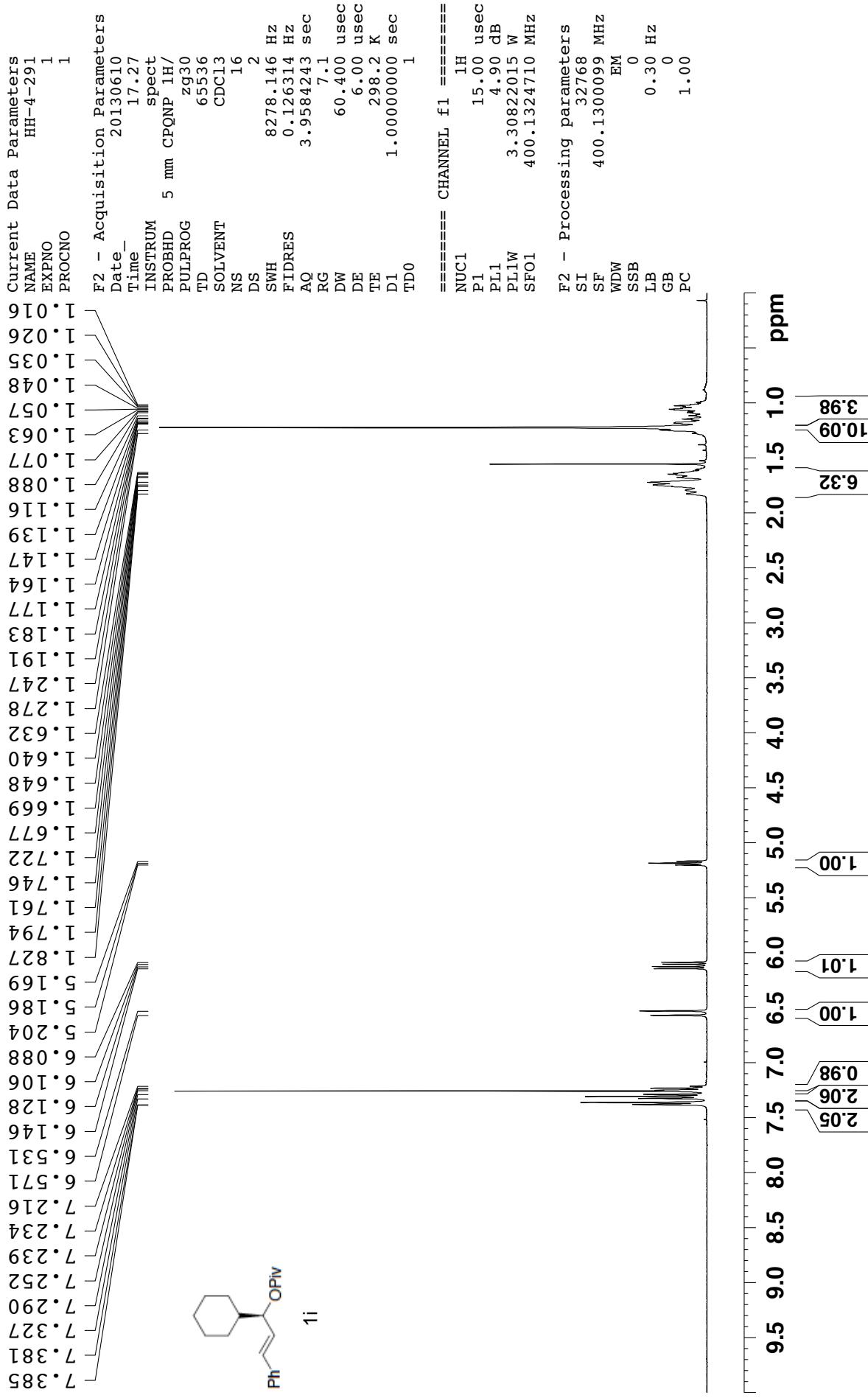


compound 1h



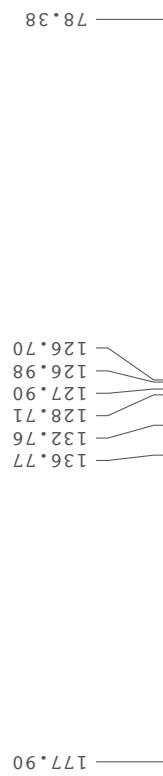
BRUKER

Compound 1*i*





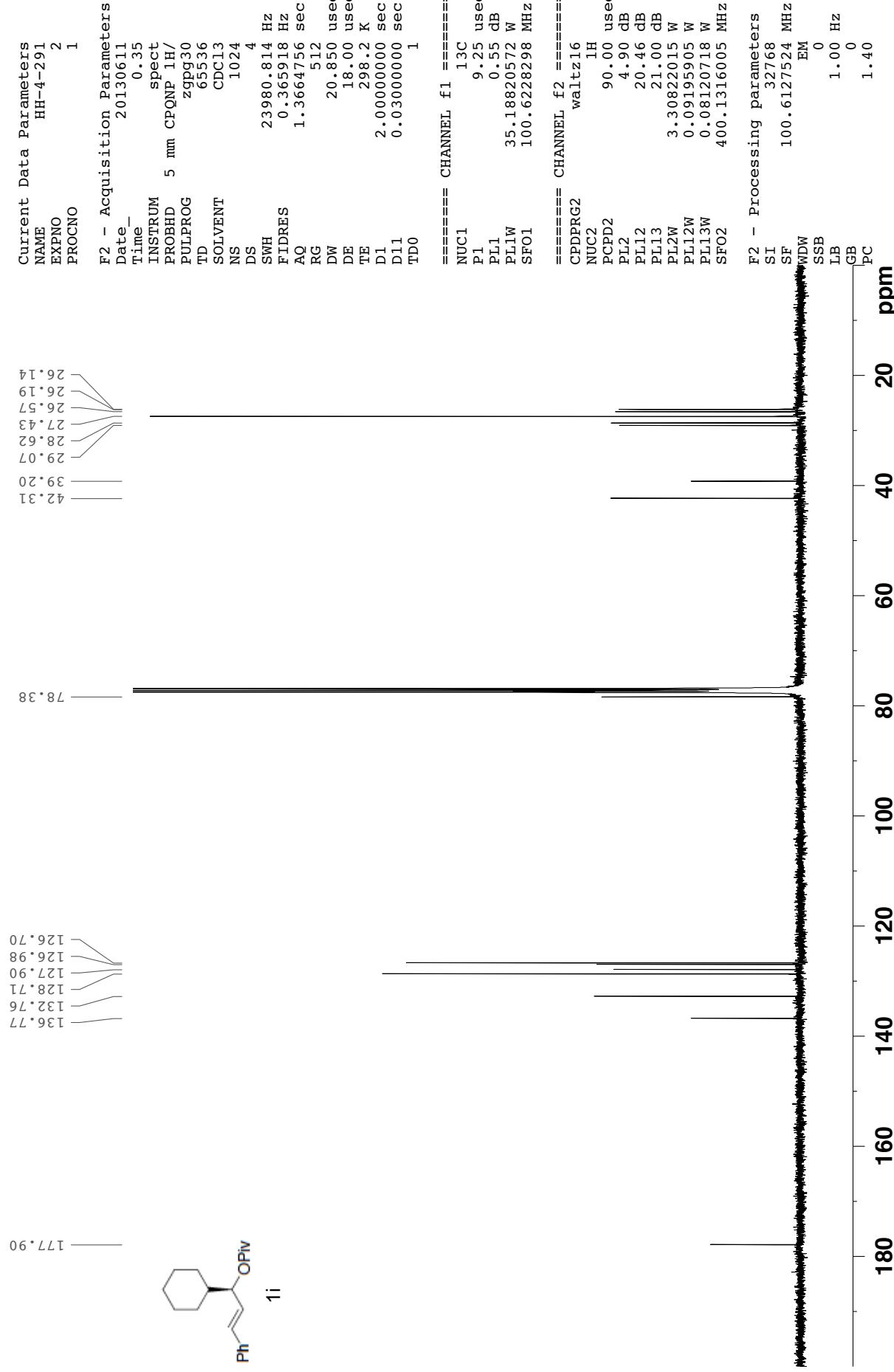
compound 1i



136, 77
132, 76
128, 71
127, 90
126, 98
126, 70

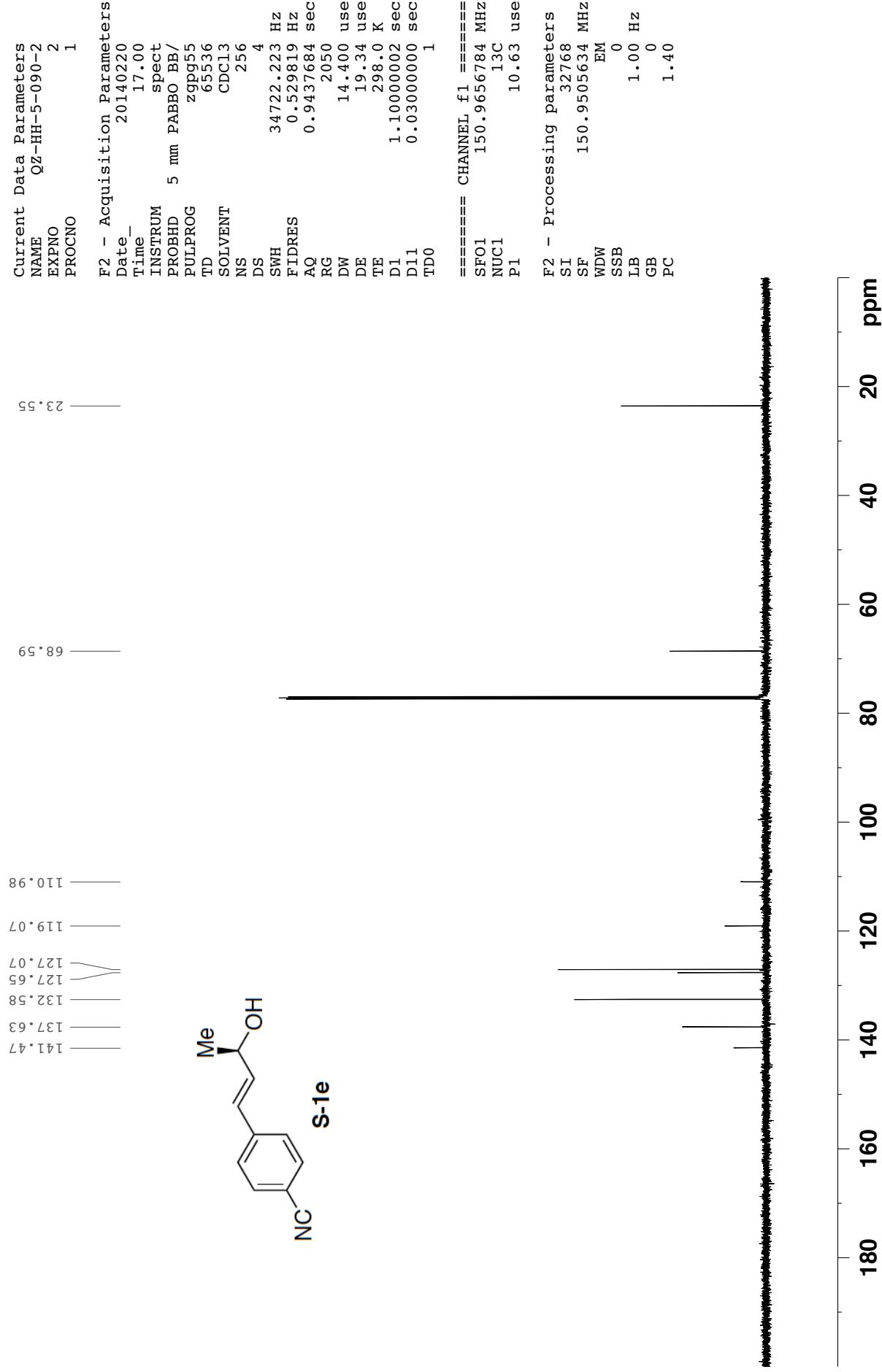
42, 31
39, 20
29, 07
28, 62
27, 43
26, 57
26, 19
26, 14

78, 38



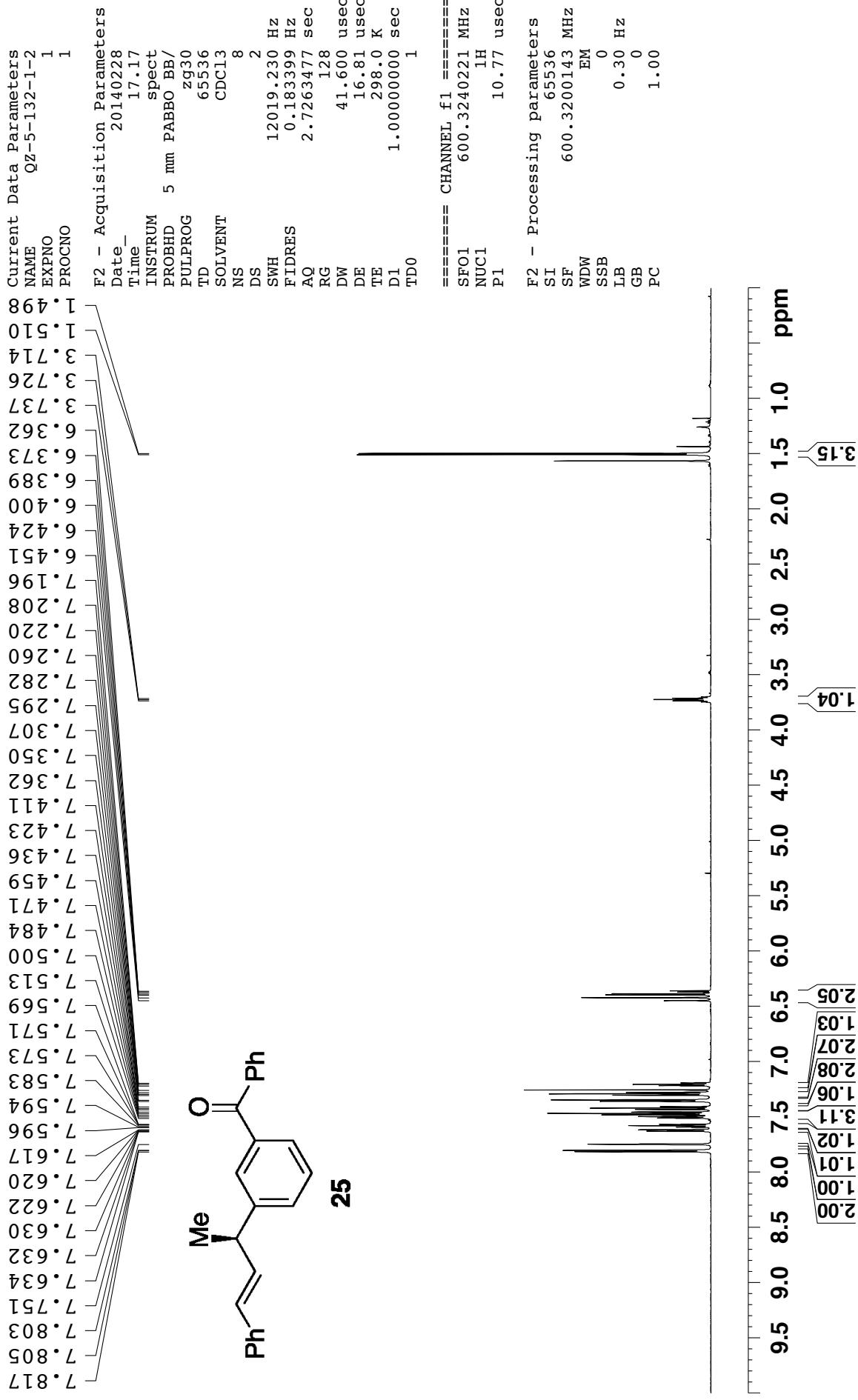
compound S-1e 1H NMR

BRUKER

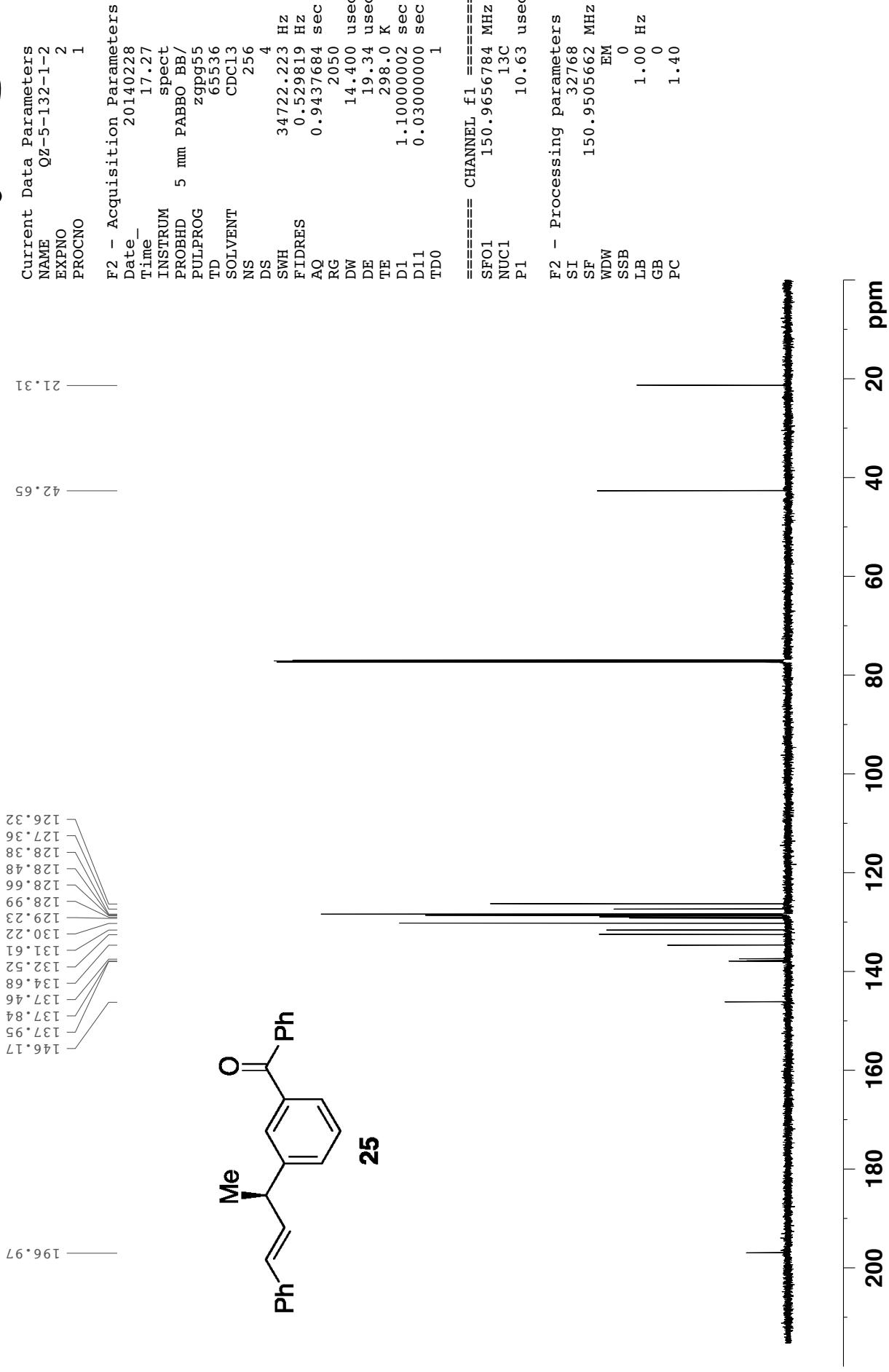




compound 25 1HNMR

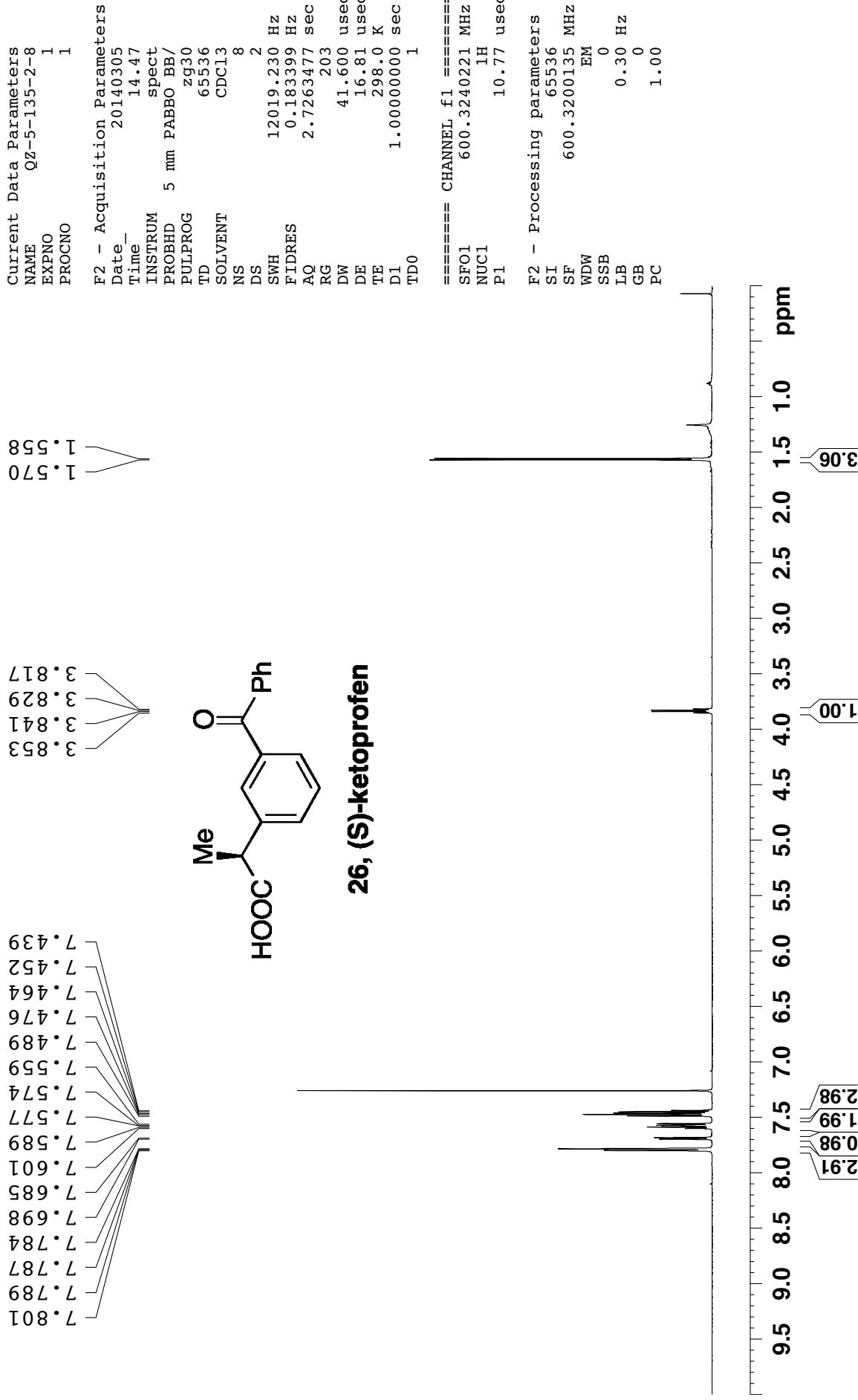
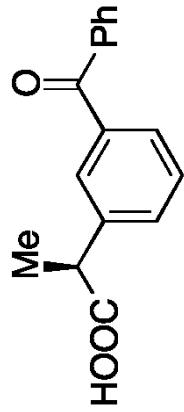
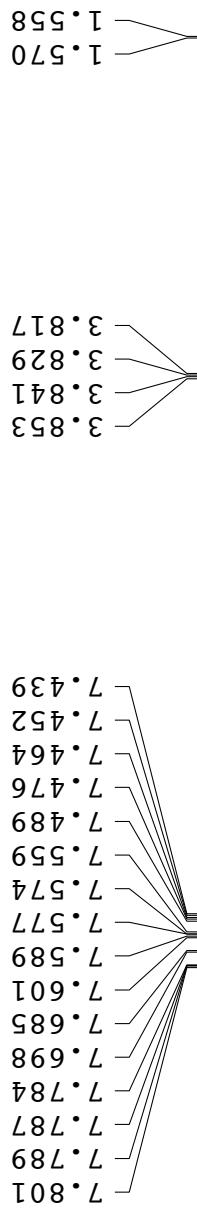


compound 25 13CNMR



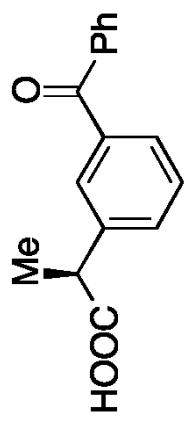


Compound 26 1H NMR (*S*)-ketoprofen



compound 26 13CNMR (*S*)-ketoprofen

140.24
138.12
137.58
132.68
131.76
130.23
129.43
128.76
128.46

**26, (S)-ketoprofen**

18.30

45.20

179.21

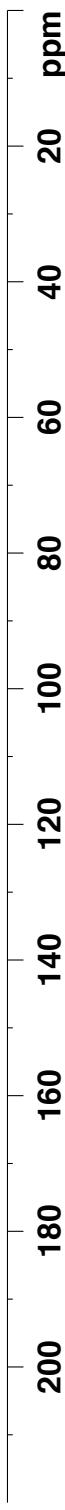
196.56

Current Data Parameters
NAME QZ-5-135-2-8
EXPNO 2
PROCNO 1

F2 - Acquisition Parameters
Date 20140305
Time 15.00
INSTRUM spect
PROBHD 5 mm PABBO BB/
PULPROG zgpg55
TD 65536
SOLVENT CDCl3
NS 256
DS 4
SWH 34722.223 Hz
FIDRES 0.529819 Hz
AQ 0.9437684 sec
RG 2050
DW 14.400 usec
DE 19.34 usec
TE 298.0 K
D1 1.10000002 sec
D11 0.03000000 sec
TD0 1

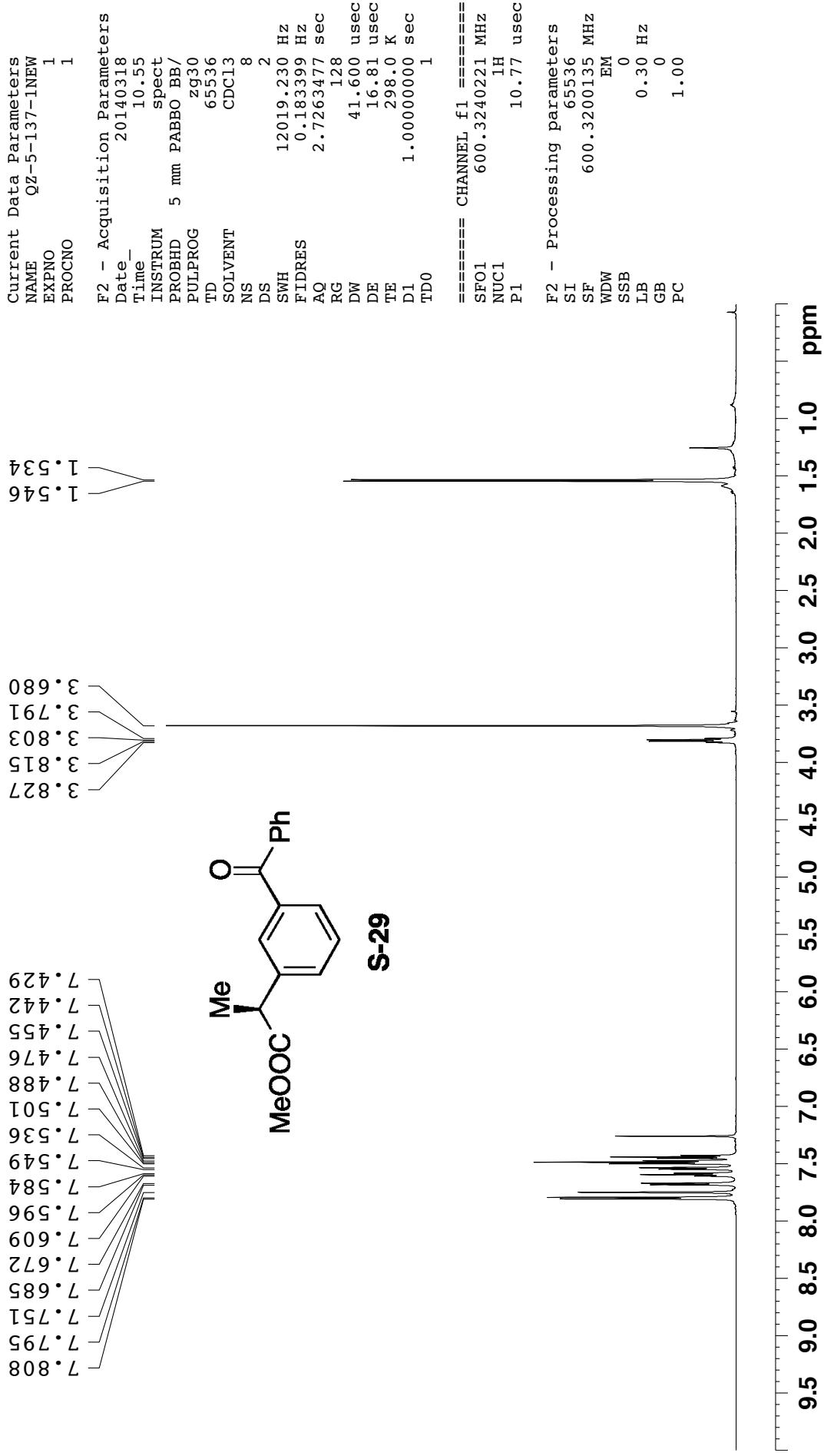
===== CHANNEL f1 =====
SFO1 150.9656784 MHz
NUC1 13C
P1 10.63 usec

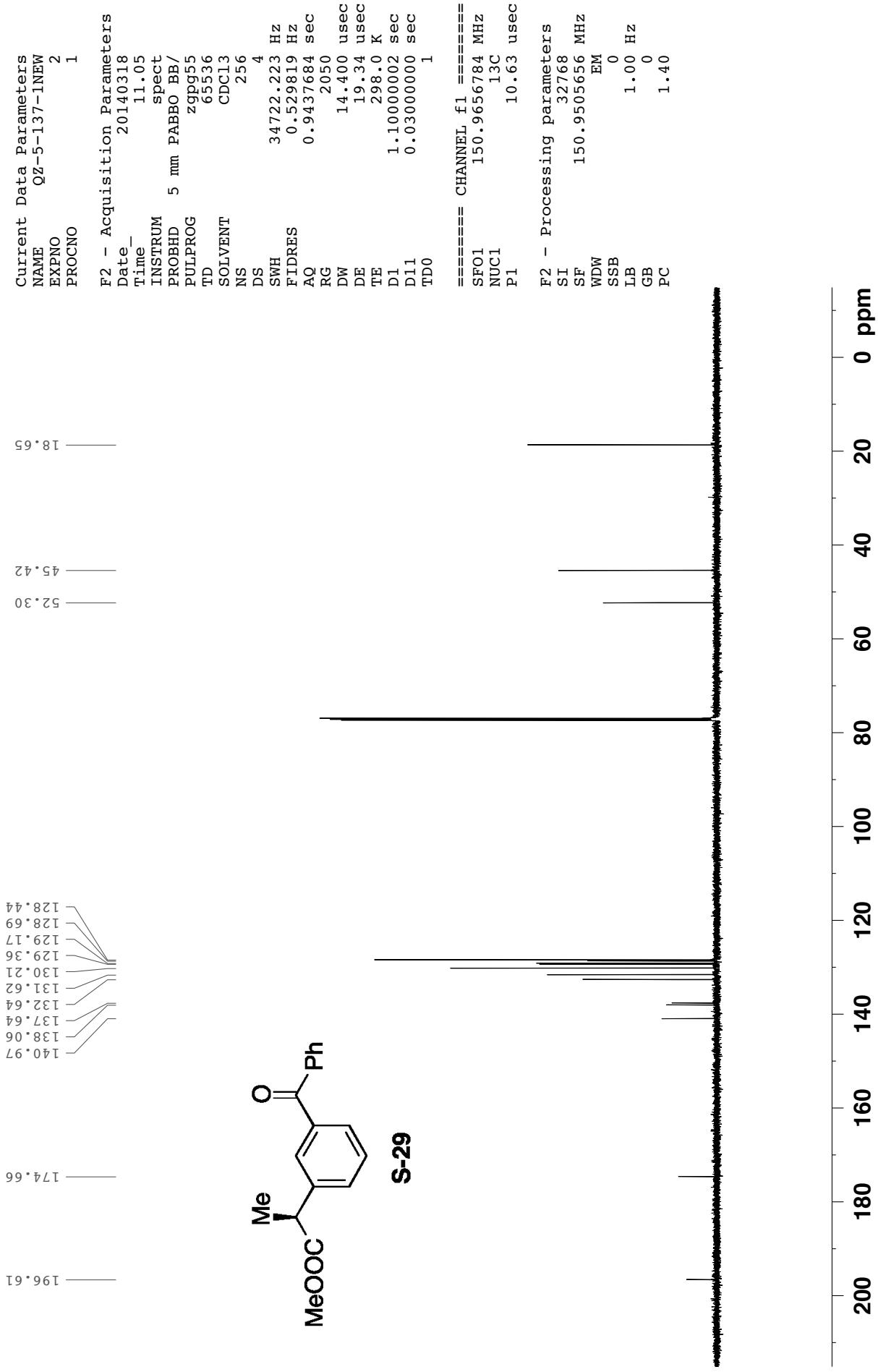
F2 - Processing parameters
SI 32768
SF 150.9505633 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40





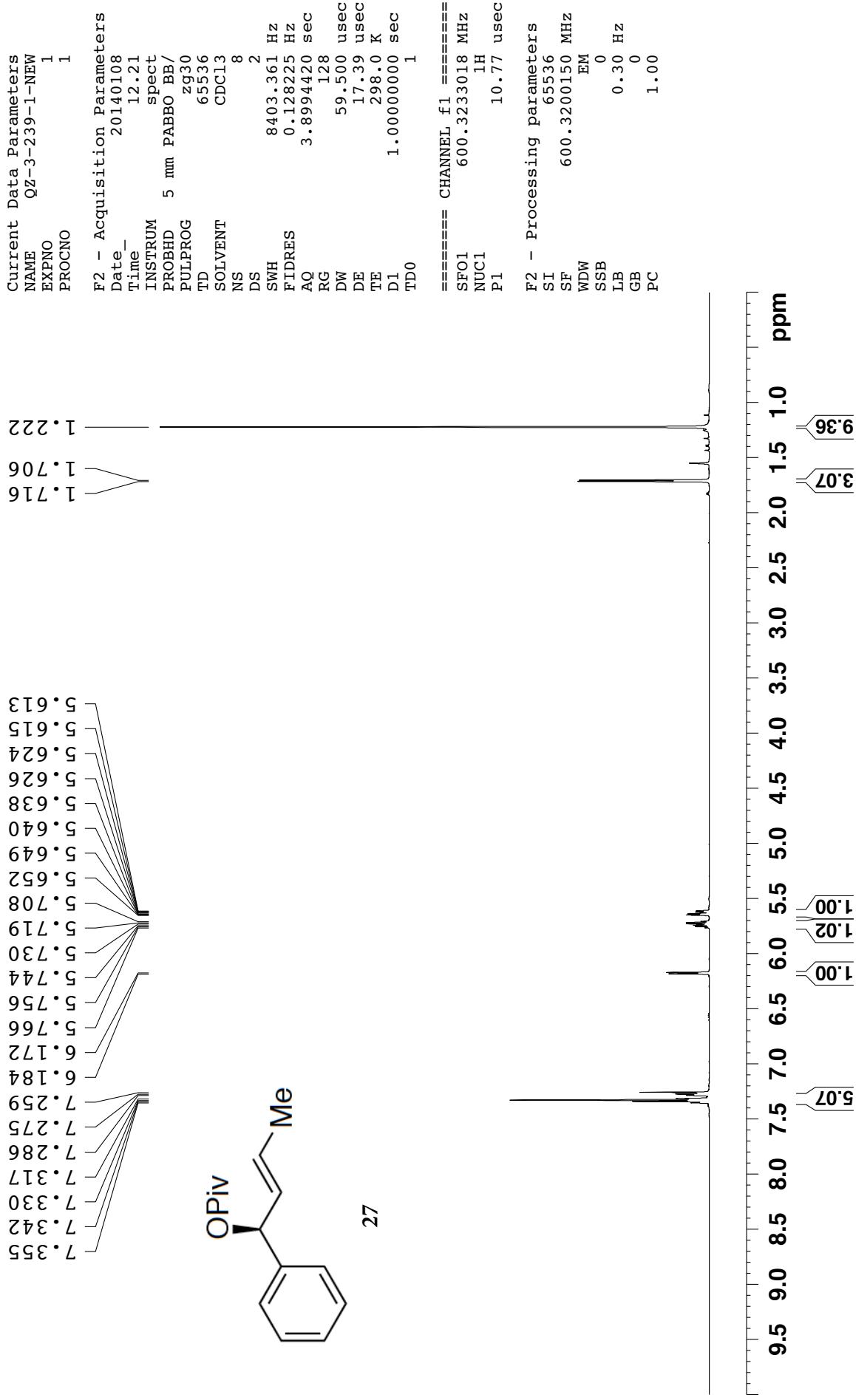
compound S-29 1H NMR



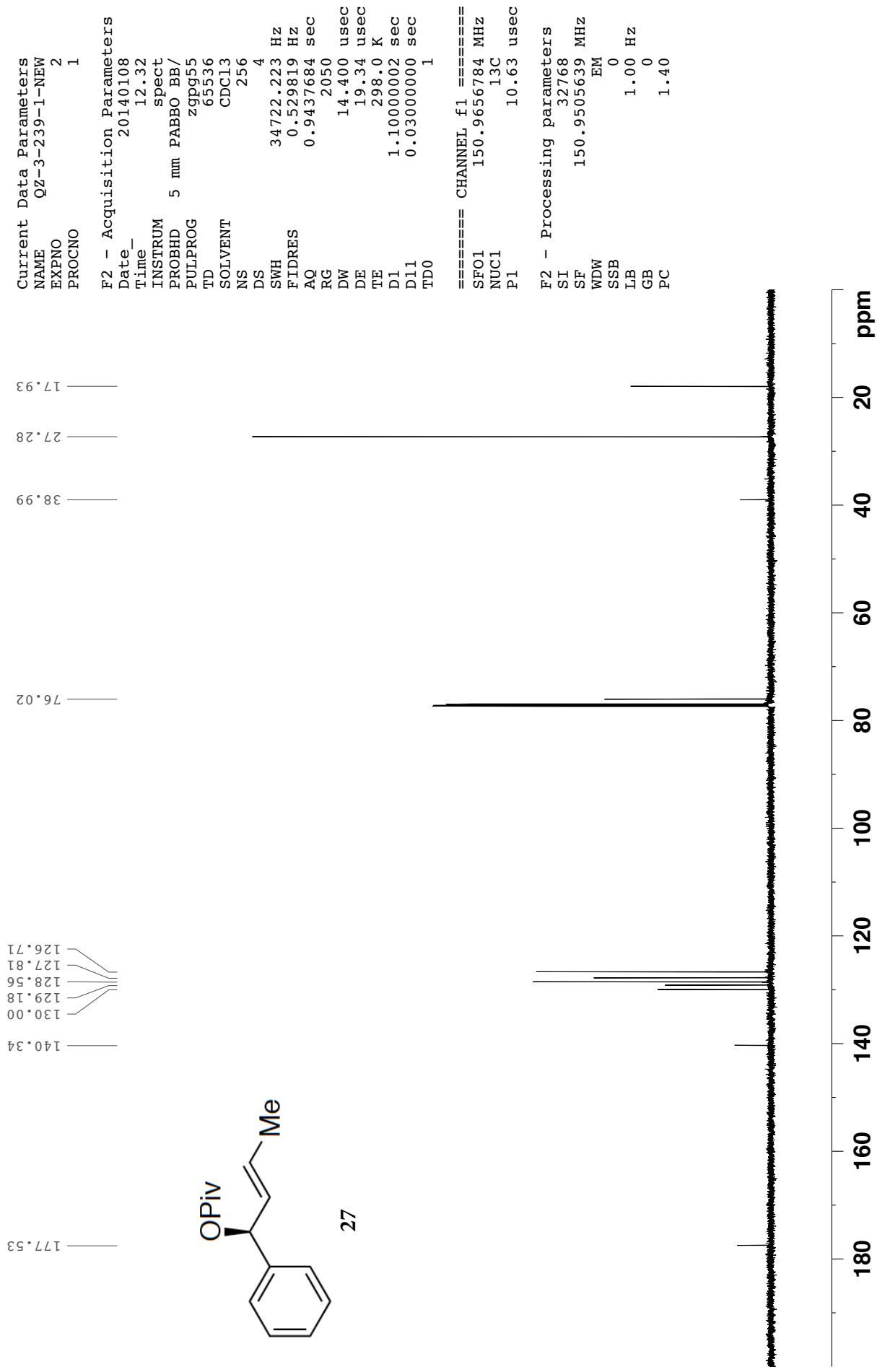


BRUKER

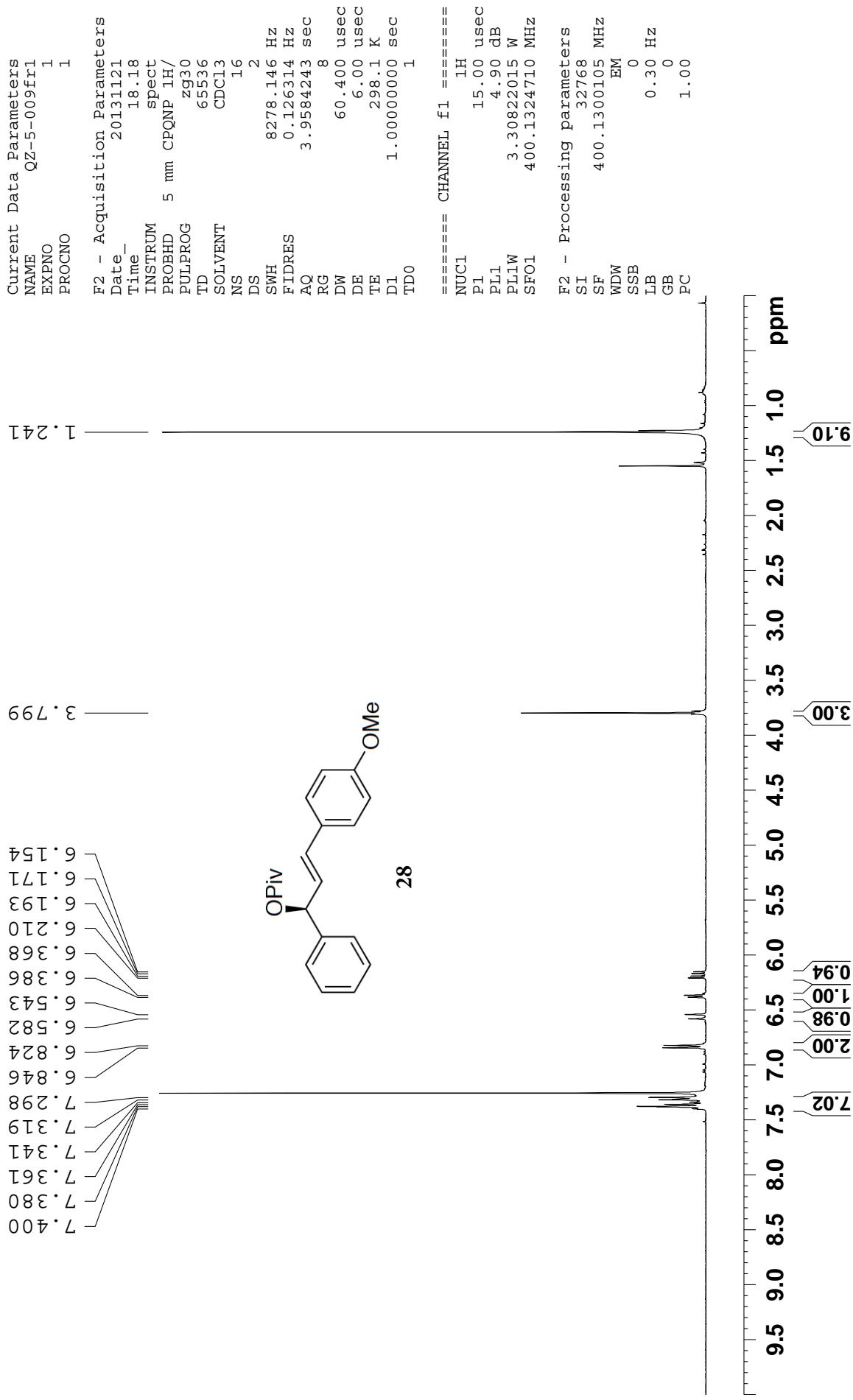
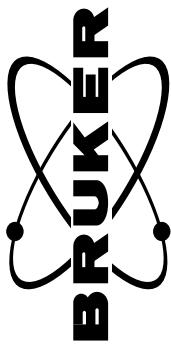
compound 27 1H NMR

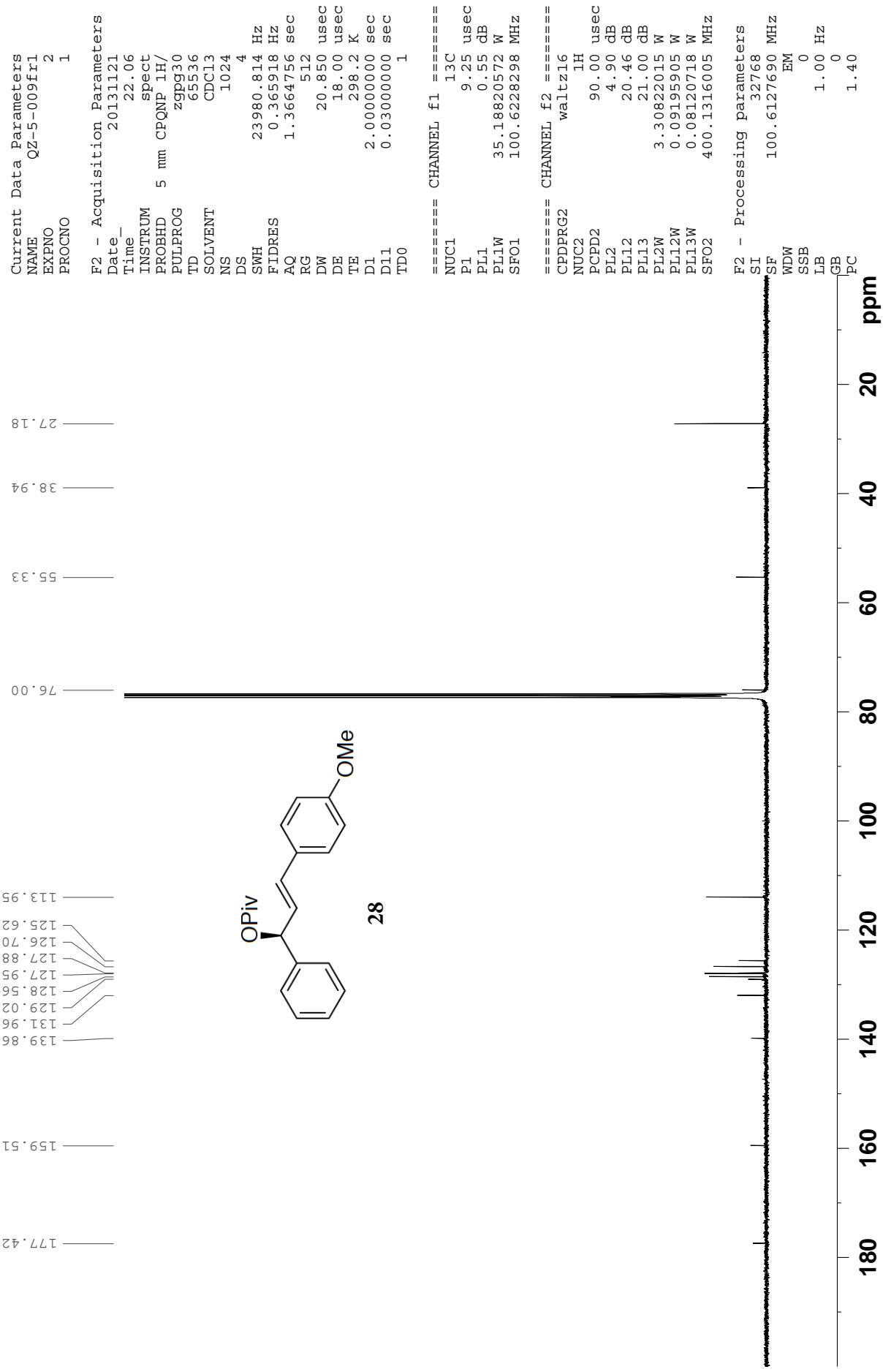


compound 27 13CNMR

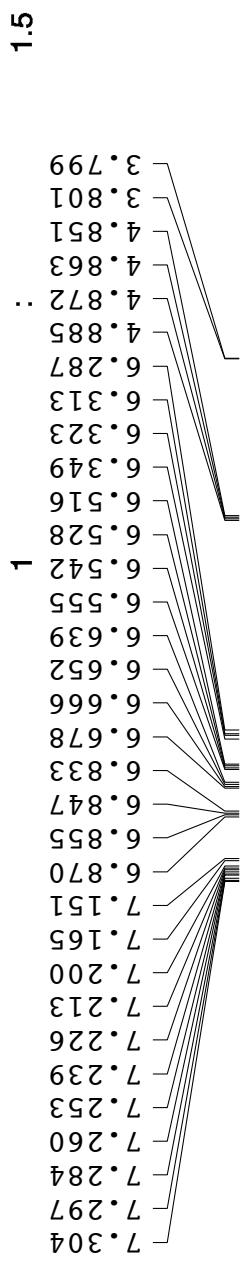


compound 28 1H NMR





29 and enantioenriched 30



29 and enantioenriched 30

1

30

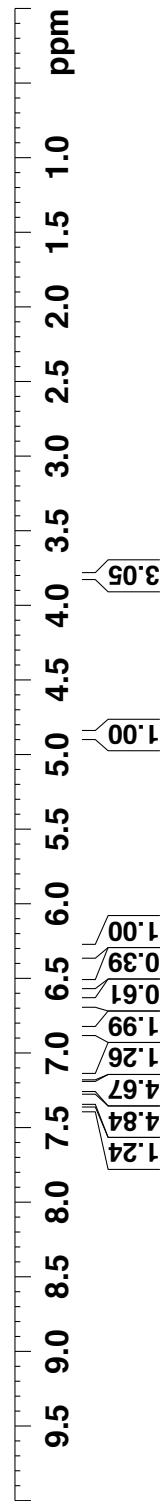
BRUKER

Current Data Parameters
NAME QZ-5-123-1
EXPNO 1
PROCNO 1

F2 - Acquisition Parameters
Date 20140214
Time 13.20
INSTRUM spect
PROBHD 5 mm PABBO BB/
PULPROG zg30
TD 65536
SOLVENT CDCl₃
NS 8
DS 2
SWH 8403.361 Hz
FIDRES 0.128225 Hz
AQ 3.8994420 sec
RG 144
DW 59.500 usec
DE 17.39 usec
TE 298.0 K
D1 1.0000000 sec
TDO 1

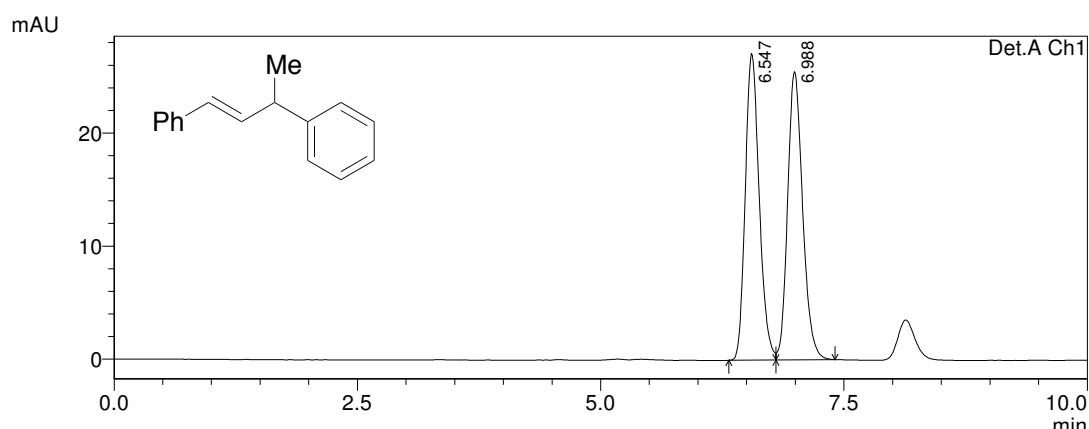
===== CHANNEL f1 ======
SFO1 600.3233018 MHz
NUC1 1H
P1 10.77 usec

F2 - Processing parameters
SI 65536
SF 600.3200141 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00



3.05
1.00
0.61
0.67
1.26
1.99
4.84
1.24
0.39
0.61
1.99
4.67
1.00

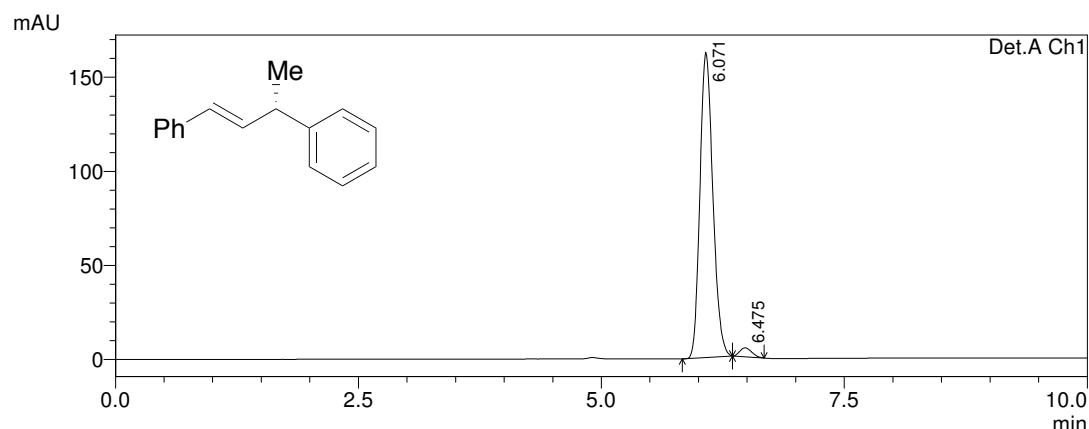
Racemic 2



Detector A Ch1 254nm

Peak#	Ret. Time	Area	Height	Area %	Height %
1	6.547	270789	27126	49.766	51.556
2	6.988	273337	25488	50.234	48.444
Total		544126	52614	100.000	100.000

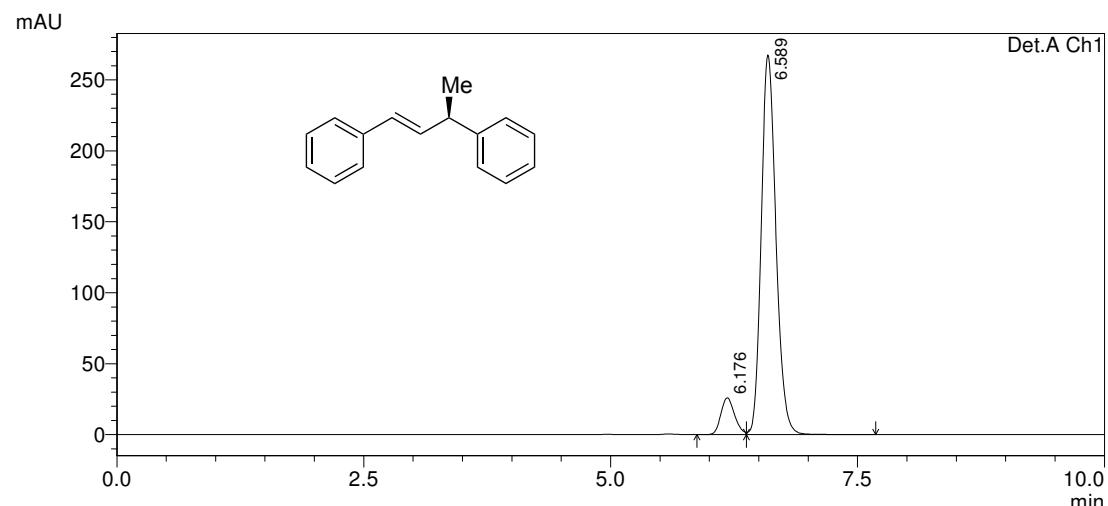
Enantioenriched 2, ee = 95%



Detector A Ch1 254nm

Peak#	Ret. Time	Area	Height	Area %	Height %
1	6.071	1497700	162221	97.383	97.181
2	6.475	40241	4706	2.617	2.819
Total		1537941	166928	100.000	100.000

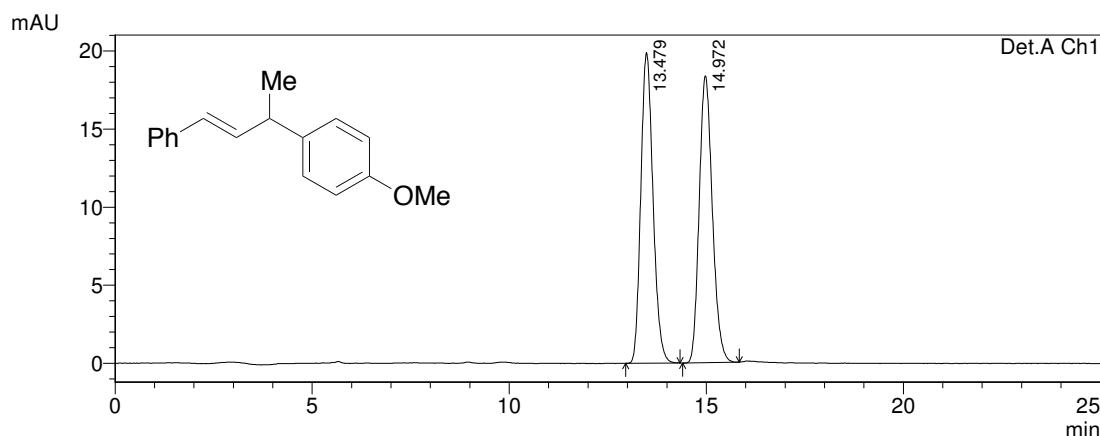
Enantioenriched (*R*)-**2**, prepared from **27**, as shown in Scheme 5, ee = 83%



Detector A Ch1 254nm

Peak#	Ret. Time	Area	Height	Area %	Height %
1	6.176	249802	25887	8.275	8.818
2	6.589	2768940	267665	91.725	91.182
Total		3018741	293552	100.000	100.000

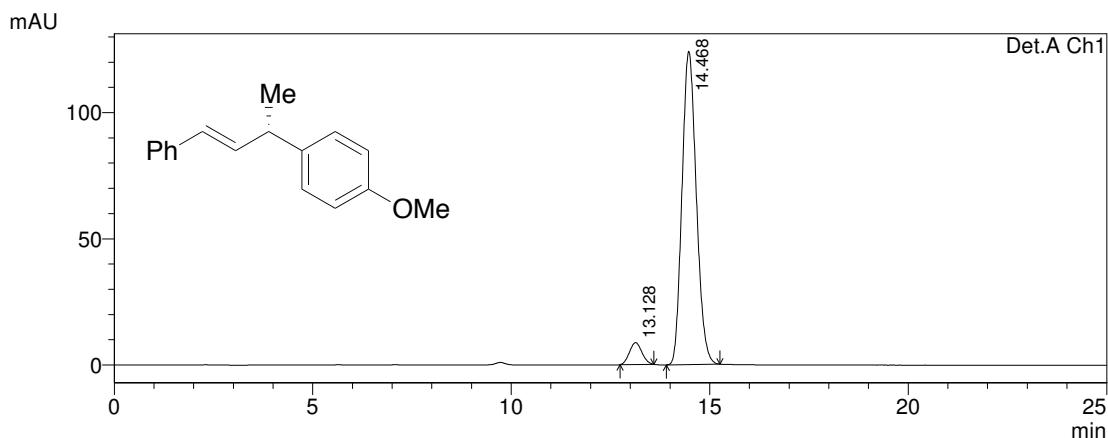
Racemic 3



Detector A Ch1 254nm

Peak#	Ret. Time	Area	Height	Area %	Height %
1	13.479	427846	19884	50.127	51.998
2	14.972	425680	18356	49.873	48.002
Total		853526	38240	100.000	100.000

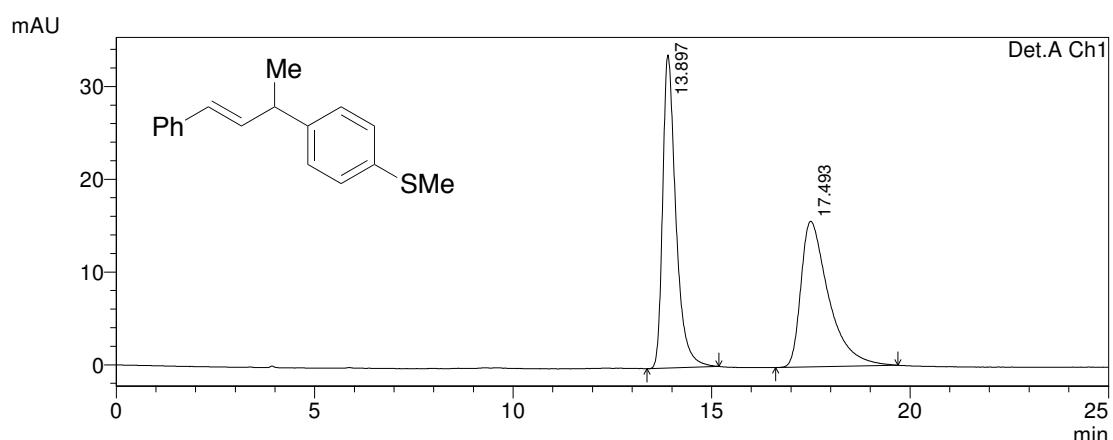
Enantioenriched 3, ee = 89%



Detector A Ch1 254nm

Peak#	Ret. Time	Area	Height	Area %	Height %
1	13.128	189418	8679	5.739	6.535
2	14.468	3111331	124131	94.261	93.465
Total		3300749	132811	100.000	100.000

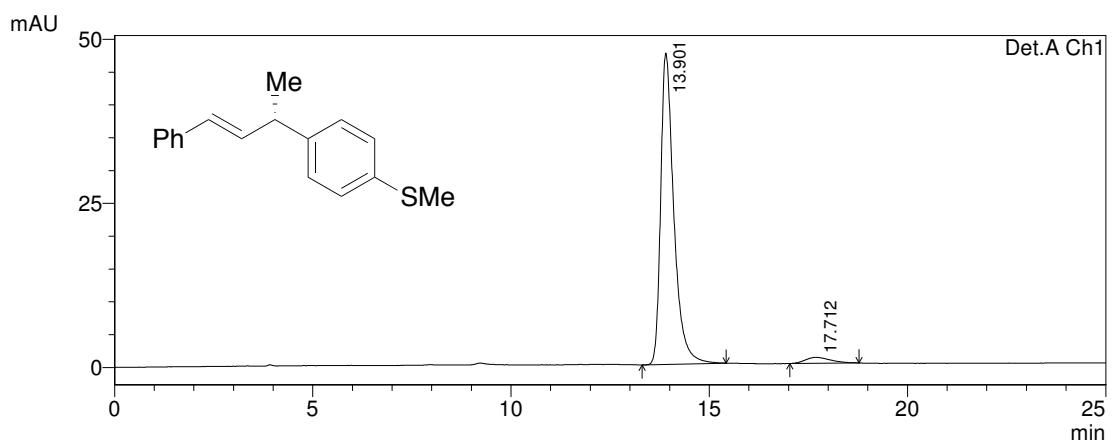
Racemic **4**



Detector A Ch1 254nm

Peak#	Ret. Time	Area	Height	Area %	Height %
1	13.897	780775	33745	50.587	68.263
2	17.493	762670	15688	49.413	31.737
Total		1543445	49433	100.000	100.000

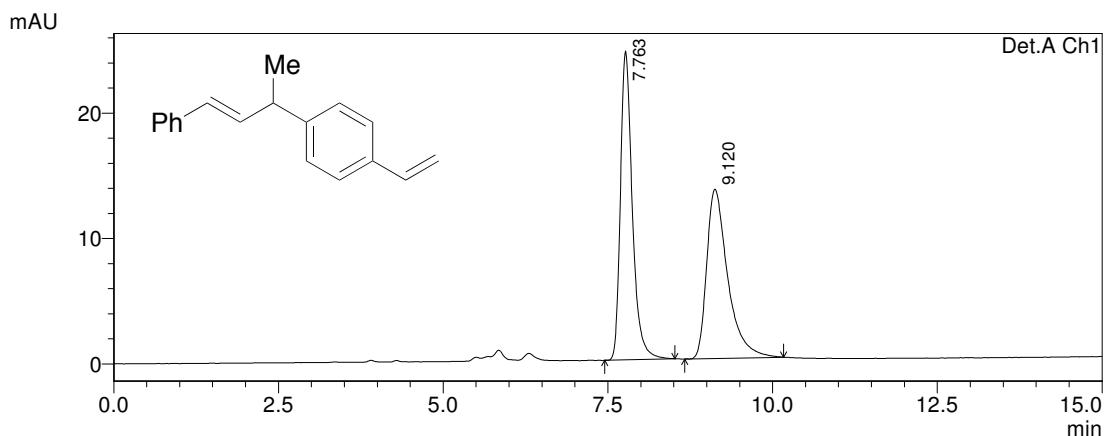
Enantioenriched **4**, ee = 93%



Detector A Ch1 254nm

Peak#	Ret. Time	Area	Height	Area %	Height %
1	13.901	1093788	47430	96.408	98.119
2	17.712	40748	909	3.592	1.881
Total		1134537	48339	100.000	100.000

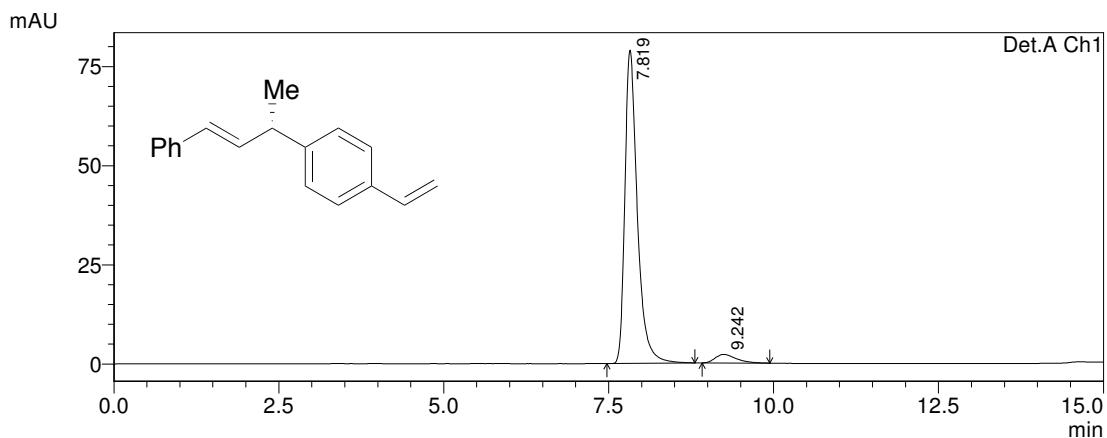
Racemic 5



Detector A Ch1 254nm

Peak#	Ret. Time	Area	Height	Area %	Height %
1	7.763	309122	24632	50.797	64.592
2	9.120	299424	13503	49.203	35.408
Total		608545	38135	100.000	100.000

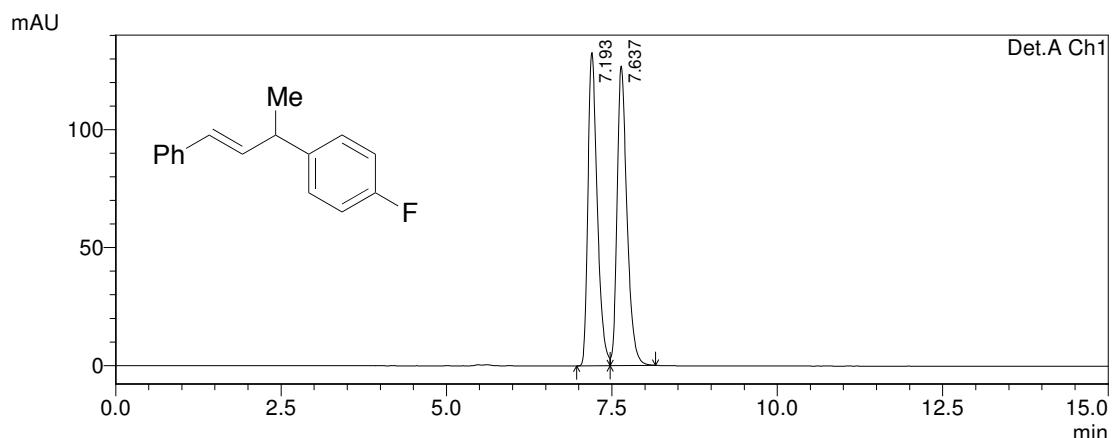
Enantioenriched 5, ee = 91%



Detector A Ch1 254nm

Peak#	Ret. Time	Area	Height	Area %	Height %
1	7.819	1036677	78979	95.516	97.324
2	9.242	48666	2171	4.484	2.676
Total		1085343	81150	100.000	100.000

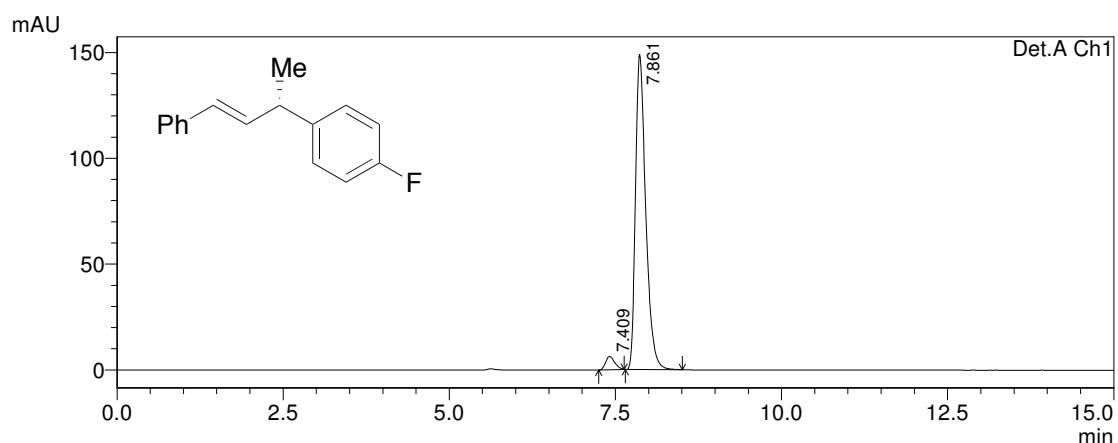
Racemic **6**



Detector A Ch1 254nm

Peak#	Ret. Time	Area	Height	Area %	Height %
1	7.193	1303841	132756	49.668	51.098
2	7.637	1321281	127053	50.332	48.902
Total		2625122	259810	100.000	100.000

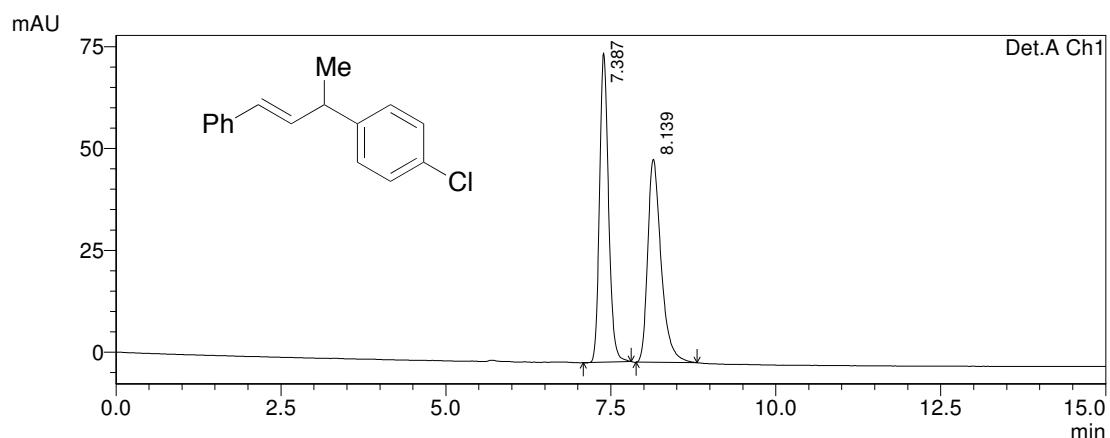
Enantioenriched **6**, ee = 93%



Detector A Ch1 254nm

Peak#	Ret. Time	Area	Height	Area %	Height %
1	7.409	59690	6261	3.607	4.036
2	7.861	1595262	148864	96.393	95.964
Total		1654951	155125	100.000	100.000

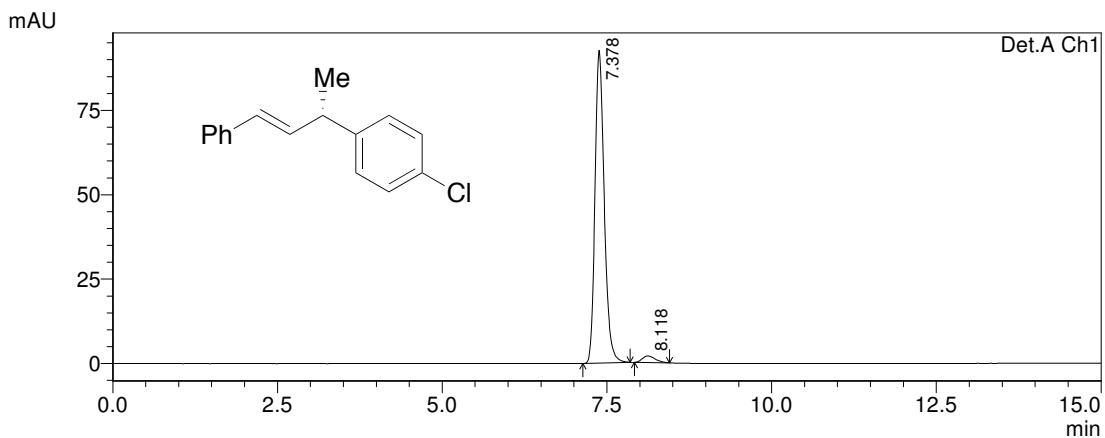
Racemic 7



Detector A Ch1 254nm

Peak#	Ret. Time	Area	Height	Area %	Height %
1	7.387	719087	75904	50.501	60.370
2	8.139	704825	49826	49.499	39.630
Total		1423912	125730	100.000	100.000

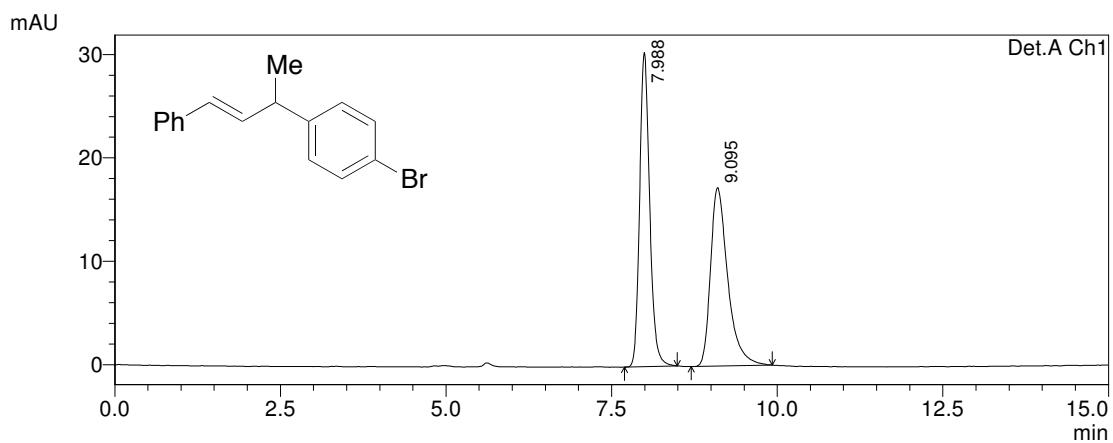
Enantioenriched 7, ee = 94%



Detector A Ch1 254nm

Peak#	Ret. Time	Area	Height	Area %	Height %
1	7.378	904758	92637	97.094	97.924
2	8.118	27077	1964	2.906	2.076
Total		931836	94601	100.000	100.000

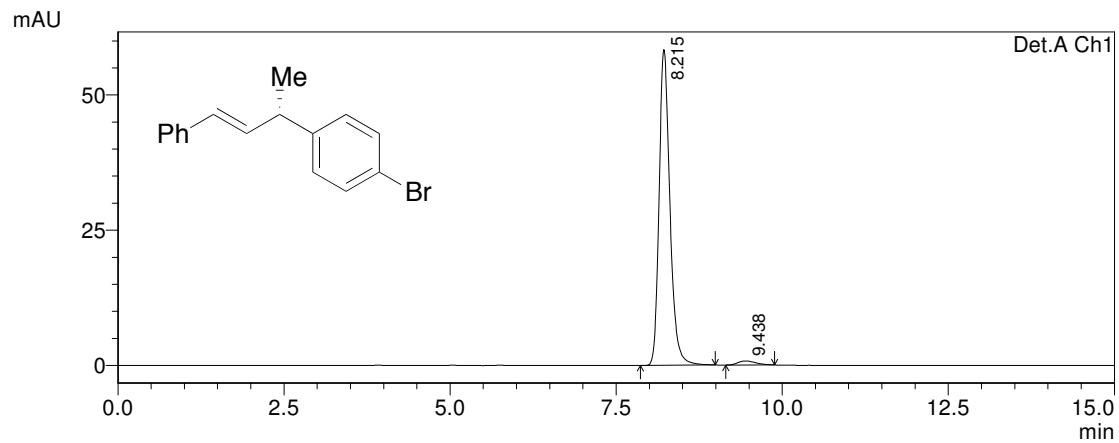
Racemic 8



Detector A Ch1 254nm

Peak#	Ret. Time	Area	Height	Area %	Height %
1	7.988	320442	30359	50.720	63.778
2	9.095	311345	17242	49.280	36.222
Total		631787	47601	100.000	100.000

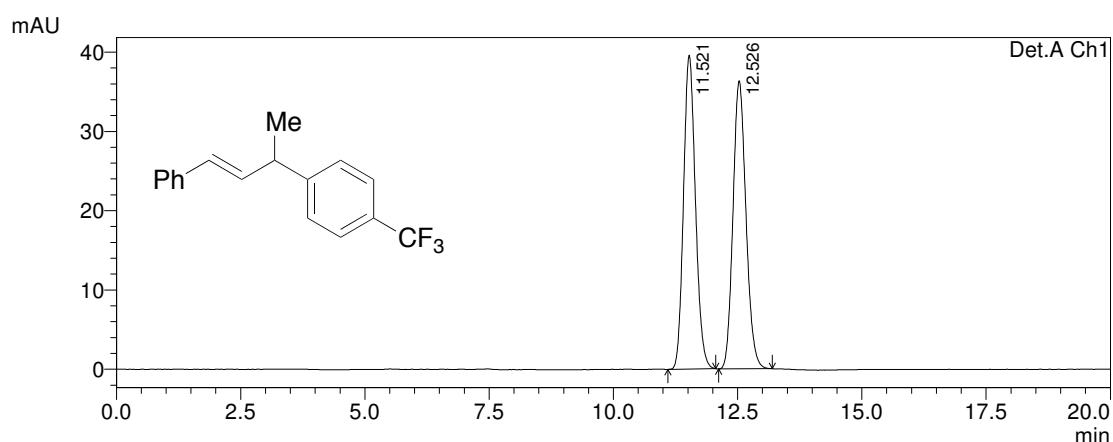
Enantioenriched 8, ee = 96%



Detector A Ch1 254nm

Peak#	Ret. Time	Area	Height	Area %	Height %
1	8.215	661539	58358	97.938	98.732
2	9.438	13930	749	2.062	1.268
Total		675469	59108	100.000	100.000

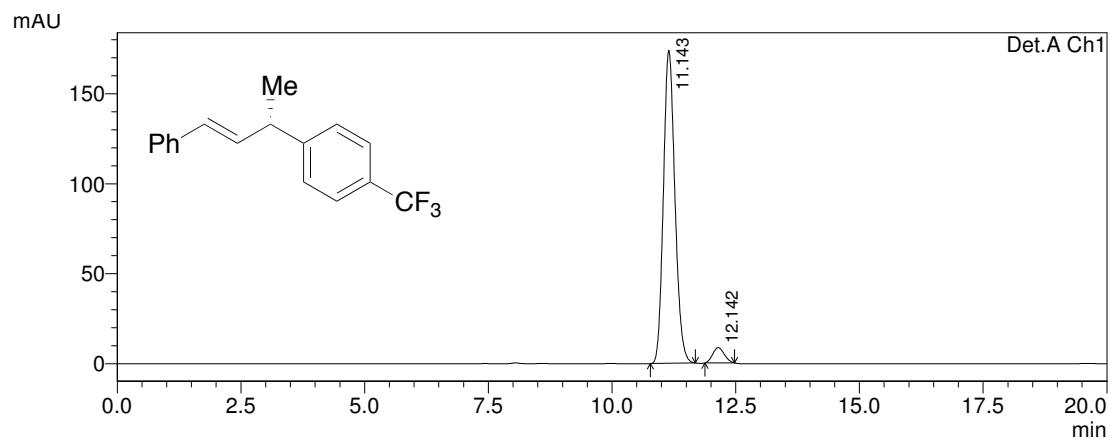
Racemic **9**



Detector A Ch1 254nm

Peak#	Ret. Time	Area	Height	Area %	Height %
1	11.521	660610	39588	50.006	52.136
2	12.526	660455	36345	49.994	47.864
Total		1321065	75933	100.000	100.000

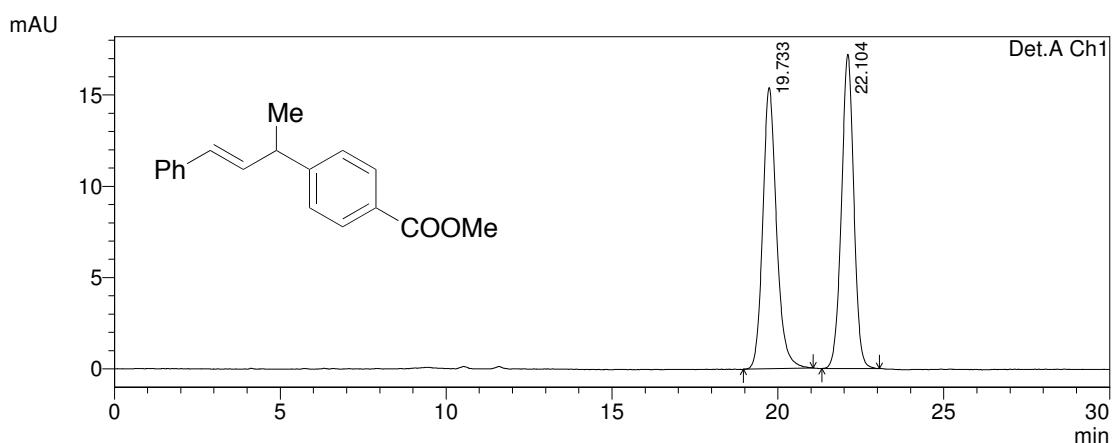
Enantioenriched **9**, ee = 91%



Detector A Ch1 254nm

Peak#	Ret. Time	Area	Height	Area %	Height %
1	11.143	2817309	173908	95.290	95.325
2	12.142	139267	8530	4.710	4.675
Total		2956576	182438	100.000	100.000

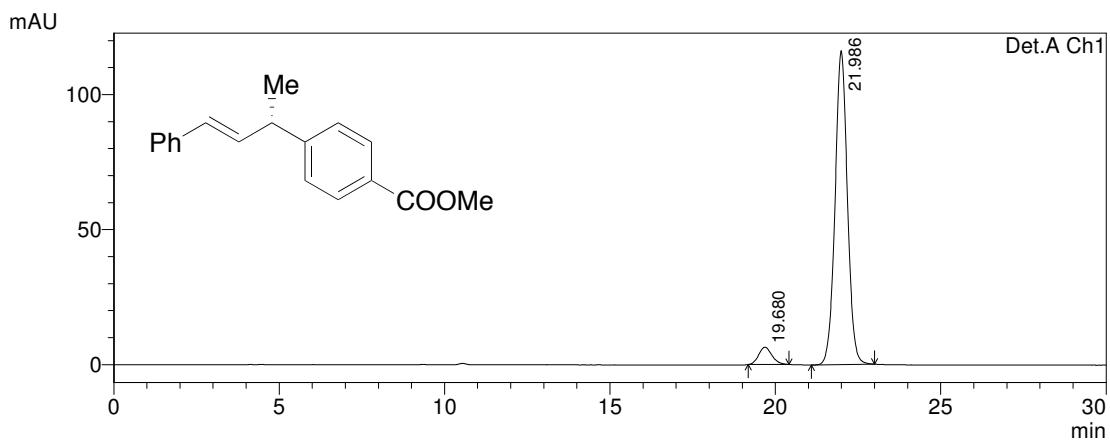
Racemic **10**



Detector A Ch1 254nm

Peak#	Ret. Time	Area	Height	Area %	Height %
1	19.733	444003	15399	49.901	47.223
2	22.104	445768	17211	50.099	52.777
Total		889771	32610	100.000	100.000

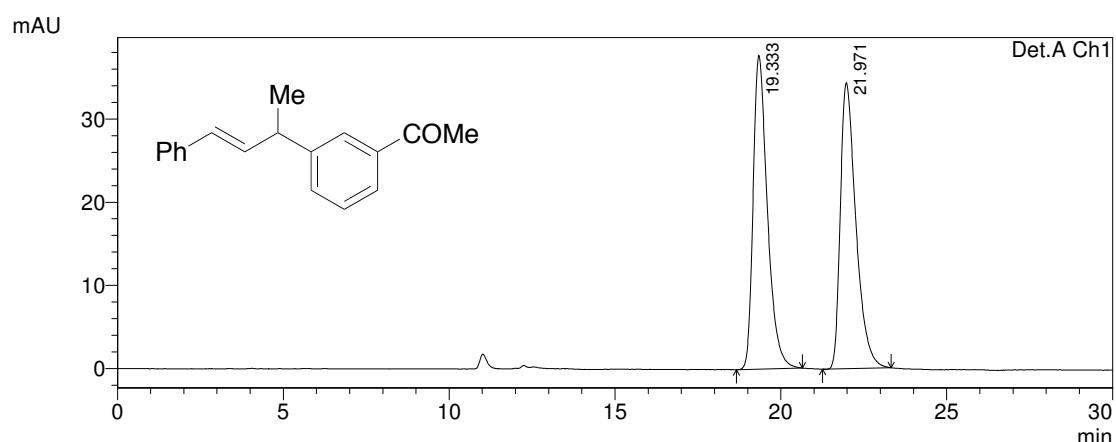
Enantioenriched **10**, ee = 89%



Detector A Ch1 254nm

Peak#	Ret. Time	Area	Height	Area %	Height %
1	19.680	178540	6440	5.592	5.249
2	21.986	3014453	116234	94.408	94.751
Total		3192993	122673	100.000	100.000

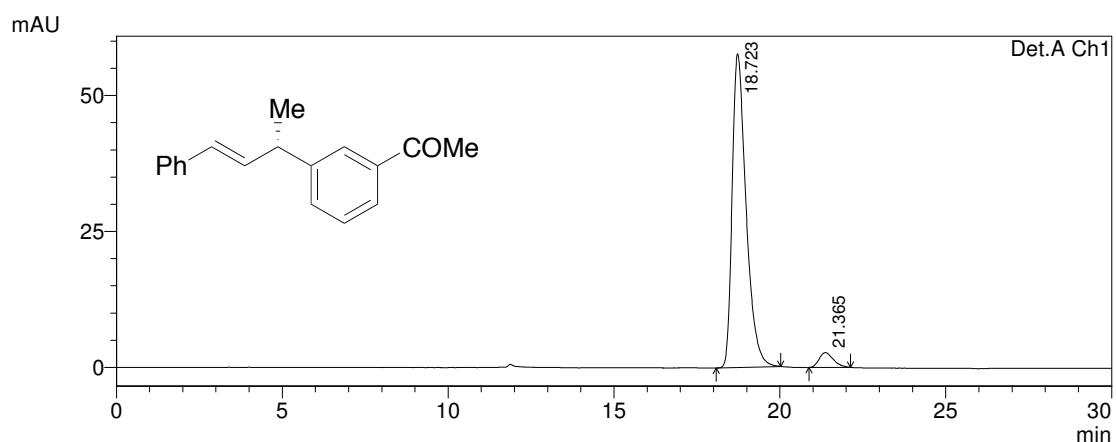
Racemic **11**



Detector A Ch1 254nm

Peak#	Ret. Time	Area	Height	Area %	Height %
1	19.333	1126904	37748	50.126	52.323
2	21.971	1121253	34396	49.874	47.677
Total		2248157	72144	100.000	100.000

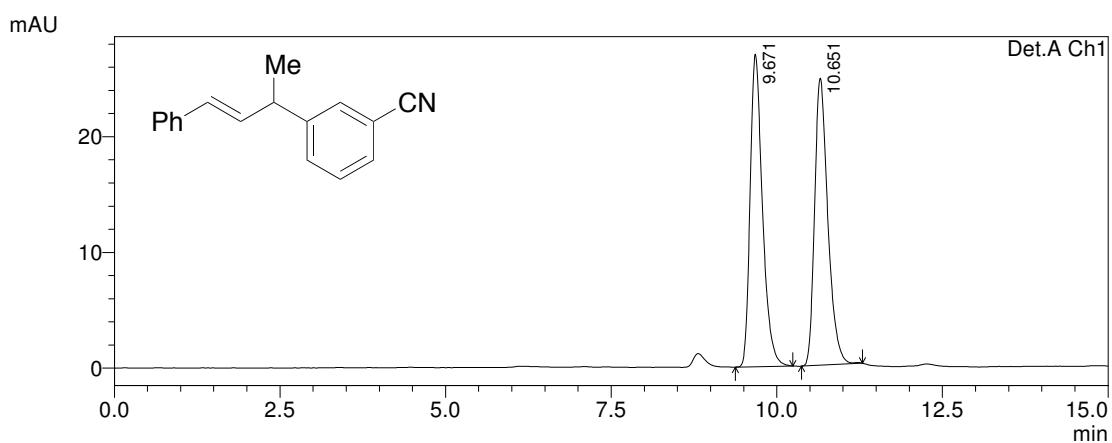
Enantioenriched **11**, ee = 91%



Detector A Ch1 254nm

Peak#	Ret. Time	Area	Height	Area %	Height %
1	18.723	1683869	57676	95.364	95.458
2	21.365	81861	2744	4.636	4.542
Total		1765730	60420	100.000	100.000

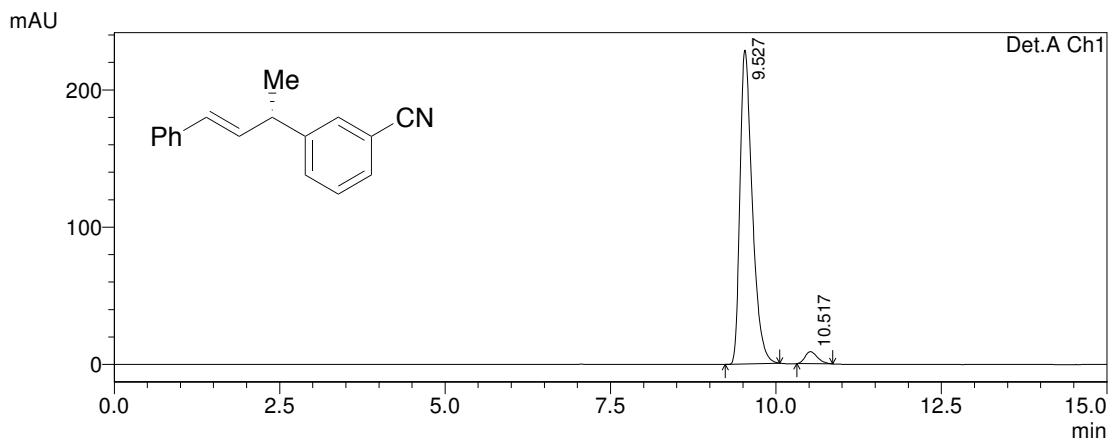
Racemic **12**



Detector A Ch1 254nm

Peak#	Ret. Time	Area	Height	Area %	Height %
1	9.671	349082	27005	50.386	52.117
2	10.651	343730	24810	49.614	47.883
Total		692812	51815	100.000	100.000

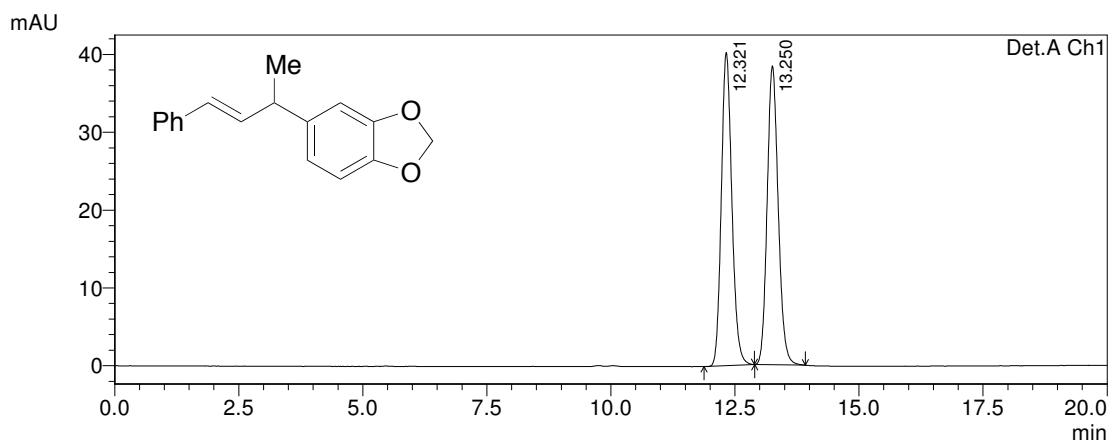
Enantioenriched **12**, ee = 93%



Detector A Ch1 254nm

Peak#	Ret. Time	Area	Height	Area %	Height %
1	9.527	2955263	228575	96.263	96.292
2	10.517	114736	8801	3.737	3.708
Total		3069999	237376	100.000	100.000

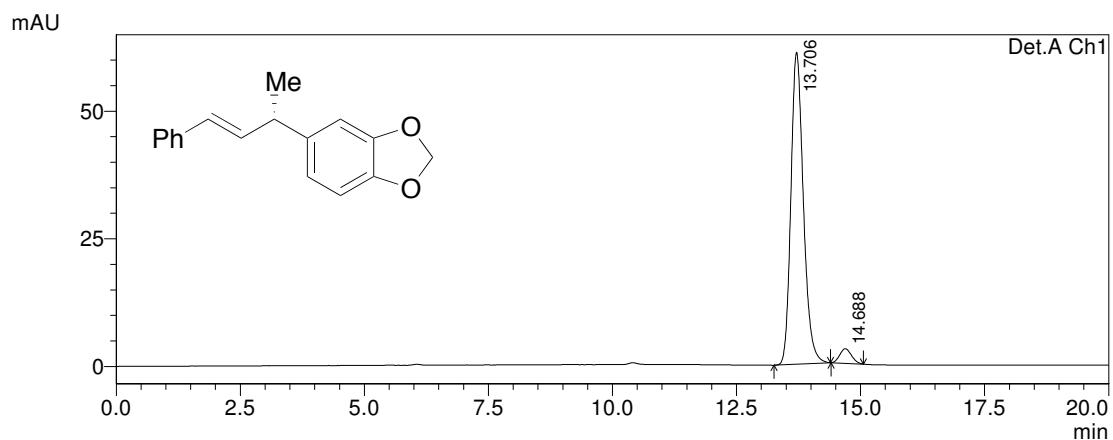
Racemic **13**



Detector A Ch1 254nm

Peak#	Ret. Time	Area	Height	Area %	Height %
1	12.321	594232	40220	49.983	51.167
2	13.250	594630	38386	50.017	48.833
Total		1188862	78605	100.000	100.000

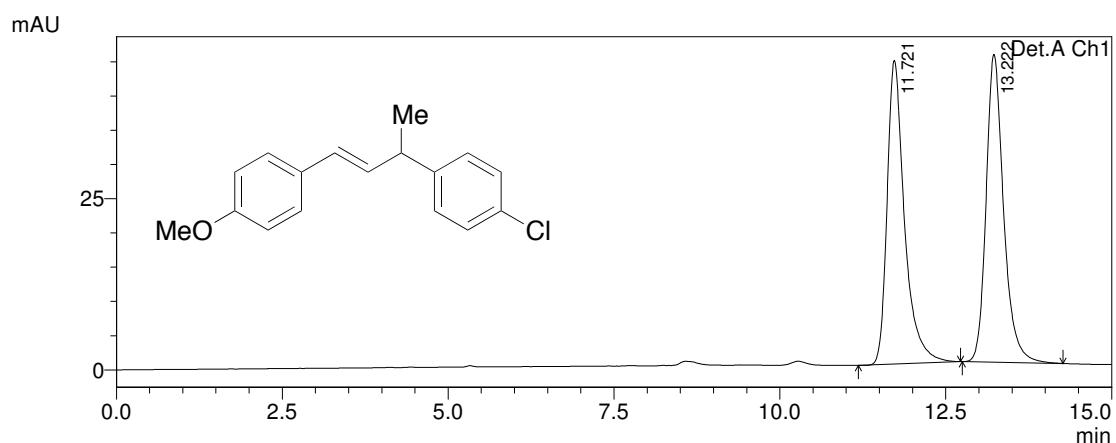
Enantioenriched **13**, ee = 92%



Detector A Ch1 254nm

Peak#	Ret. Time	Area	Height	Area %	Height %
1	13.706	1067949	61099	95.807	95.549
2	14.688	46740	2846	4.193	4.451
Total		1114689	63945	100.000	100.000

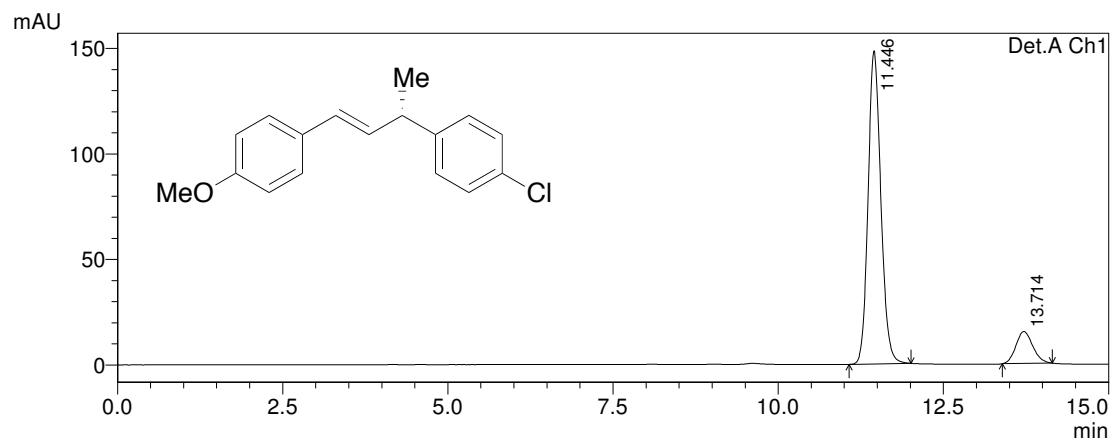
Racemic **14**



Detector A Ch1 254nm

Peak#	Ret. Time	Area	Height	Area %	Height %
1	11.721	807561	44373	49.506	49.663
2	13.222	823662	44976	50.494	50.337
Total		1631223	89349	100.000	100.000

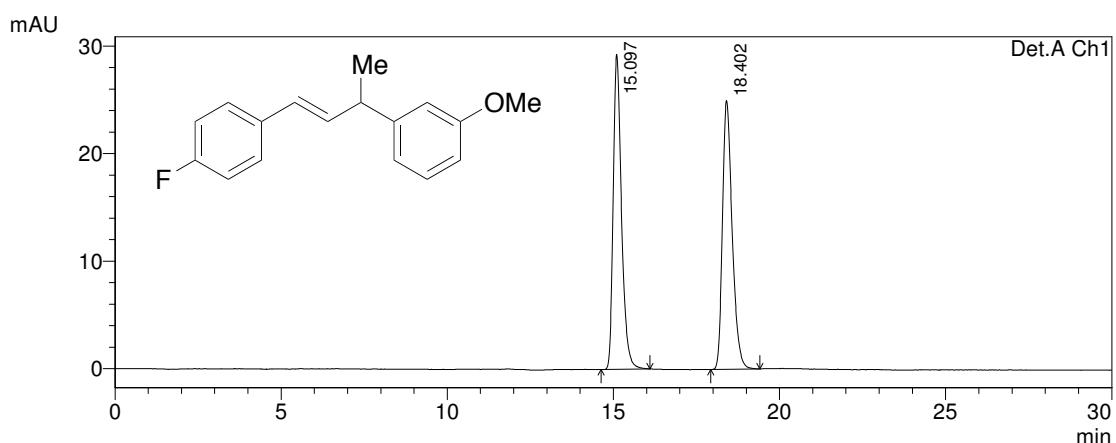
Enantioenriched **14**, ee = 76%



Detector A Ch1 254nm

Peak#	Ret. Time	Area	Height	Area %	Height %
1	11.446	1995853	148409	88.115	90.733
2	13.714	269209	15157	11.885	9.267
Total		2265062	163566	100.000	100.000

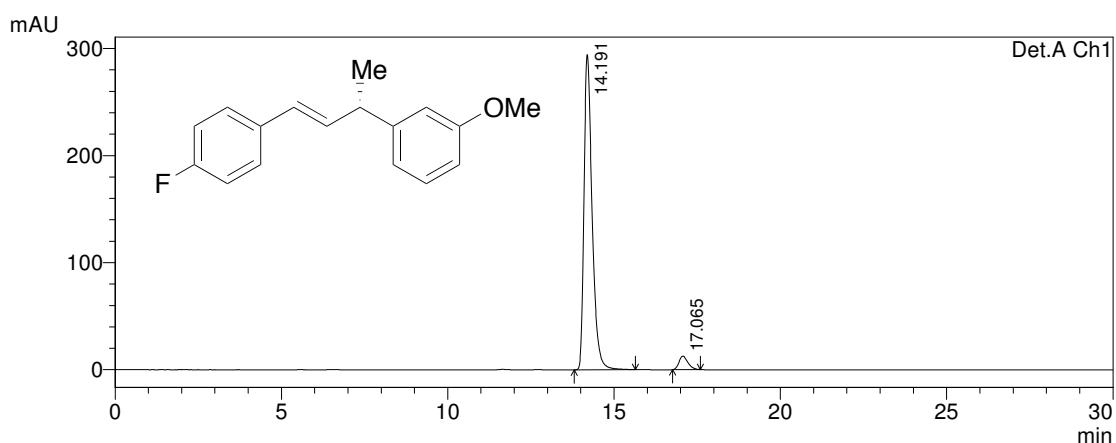
Racemic **15**



Detector A Ch1 254nm

Peak#	Ret. Time	Area	Height	Area %	Height %
1	15.097	508175	29289	49.940	53.947
2	18.402	509391	25003	50.060	46.053
Total		1017566	54292	100.000	100.000

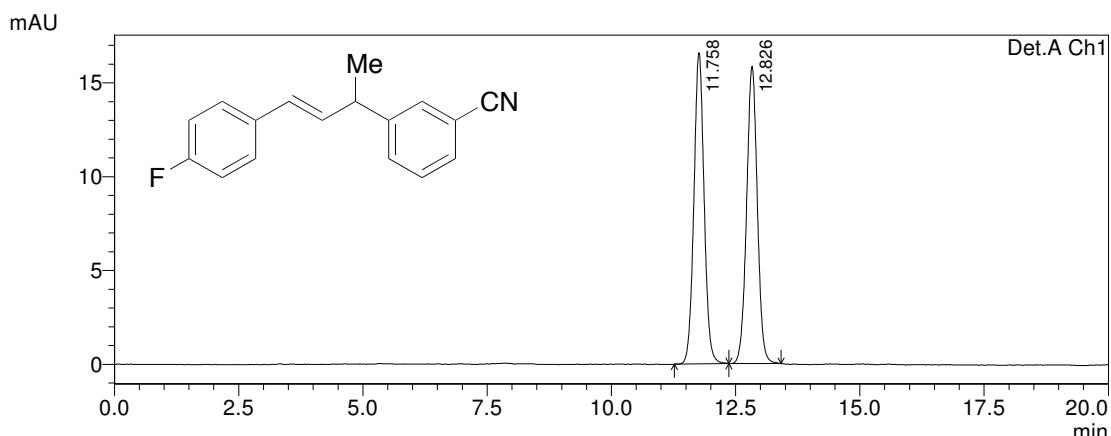
Enantioenriched **15**, ee = 91%



Detector A Ch1 254nm

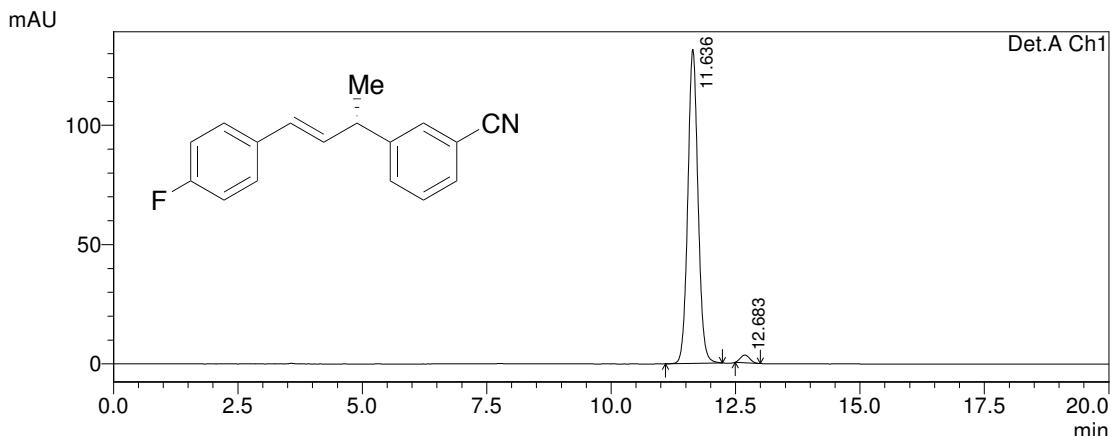
Peak#	Ret. Time	Area	Height	Area %	Height %
1	14.191	5000569	294197	95.503	95.887
2	17.065	235491	12621	4.497	4.113
Total		5236060	306818	100.000	100.000

Racemic **16**



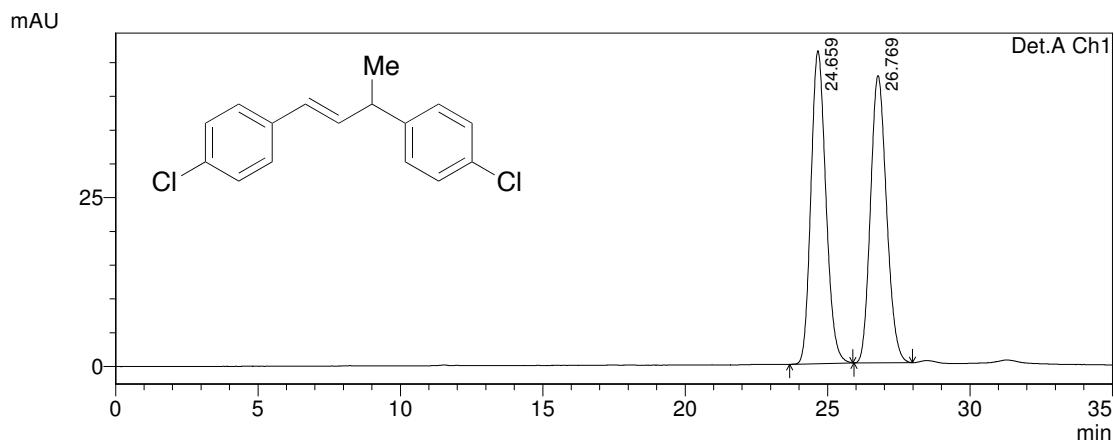
Detector A Ch1 254nm					
Peak#	Ret. Time	Area	Height	Area %	Height %
1	11.758	237836	16587	49.983	51.145
2	12.826	237998	15845	50.017	48.855
Total		475834	32432	100.000	100.000

Enantioenriched **16**, ee = 96%



Detector A Ch1 254nm					
Peak#	Ret. Time	Area	Height	Area %	Height %
1	11.636	1878029	131646	97.804	97.592
2	12.683	42169	3248	2.196	2.408
Total		1920198	134894	100.000	100.000

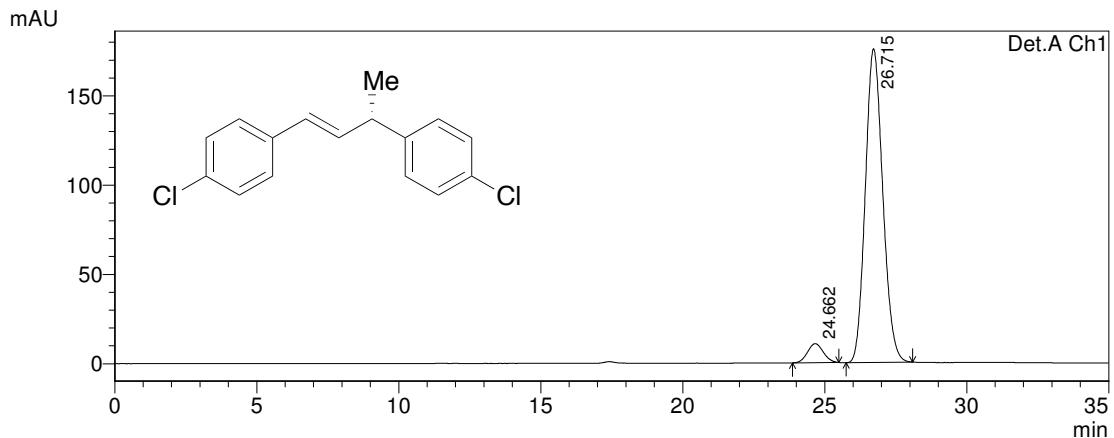
Racemic **17**



Detector A Ch1 254nm

Peak#	Ret. Time	Area	Height	Area %	Height %
1	24.659	1689765	46329	50.349	52.130
2	26.769	1666345	42543	49.651	47.870
Total		3356110	88872	100.000	100.000

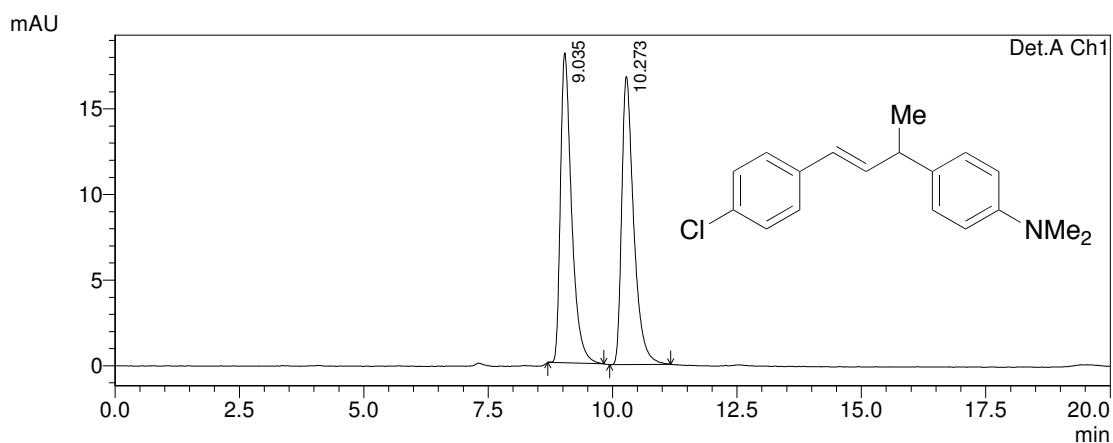
Enantioenriched **17**, ee = 90%



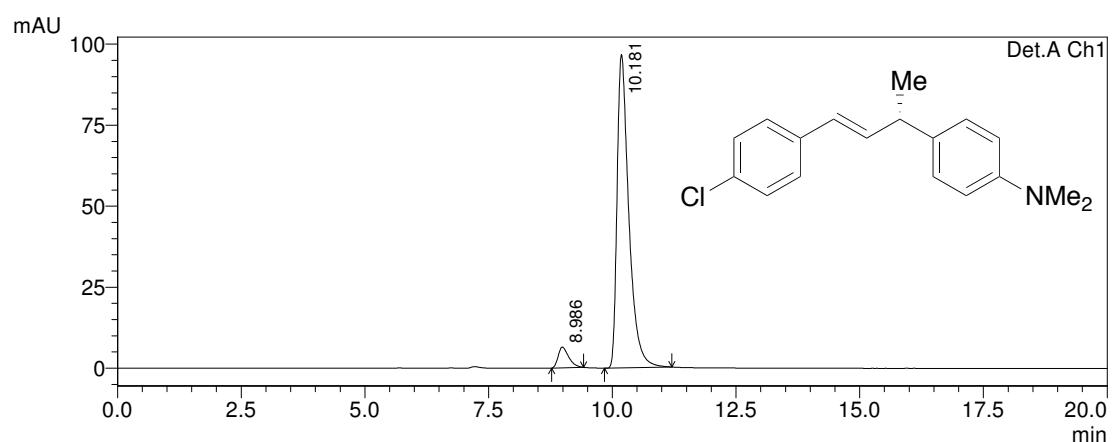
Detector A Ch1 254nm

Peak#	Ret. Time	Area	Height	Area %	Height %
1	24.662	411573	10667	5.191	5.724
2	26.715	7516797	175693	94.809	94.276
Total		7928370	186360	100.000	100.000

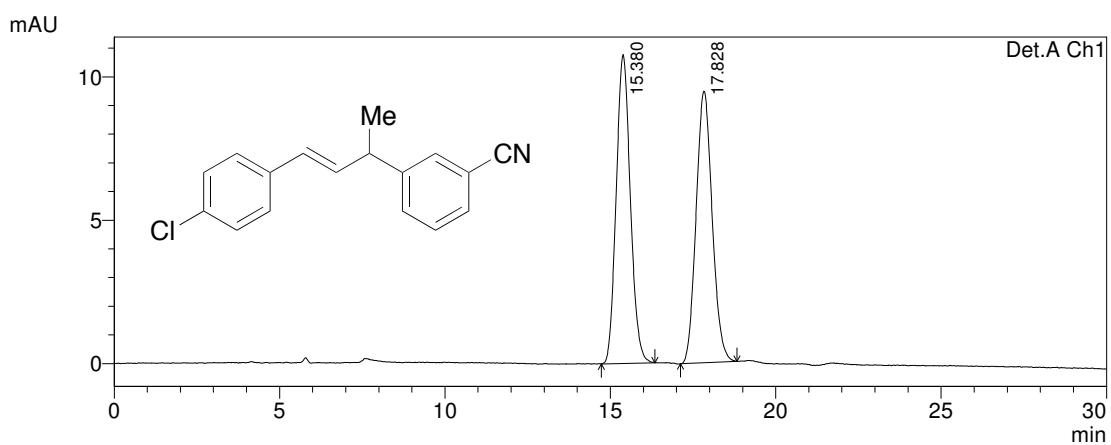
Racemic **18**



Enantioenriched **18**, ee = 89%



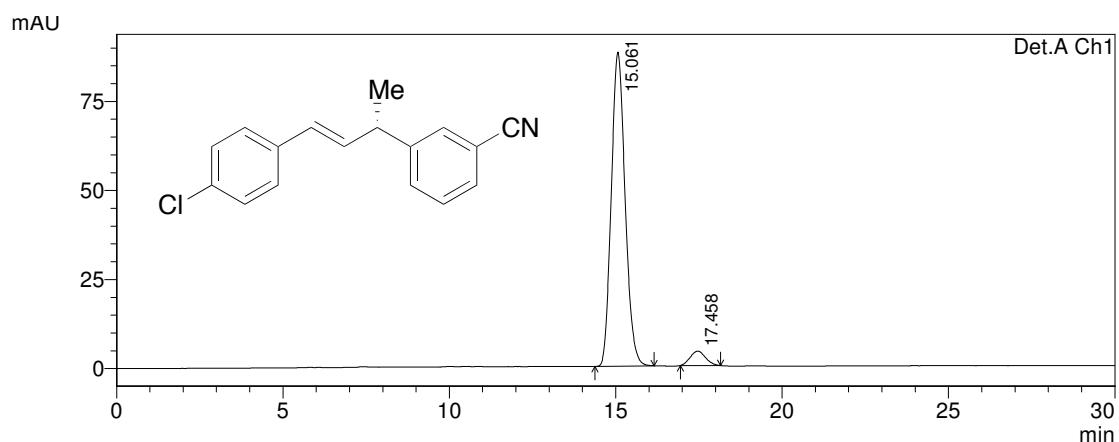
Racemic **19**



Detector A Ch1 254nm

Peak#	Ret. Time	Area	Height	Area %	Height %
1	15.380	320415	10768	50.063	53.201
2	17.828	319614	9472	49.937	46.799
Total		640029	20239	100.000	100.000

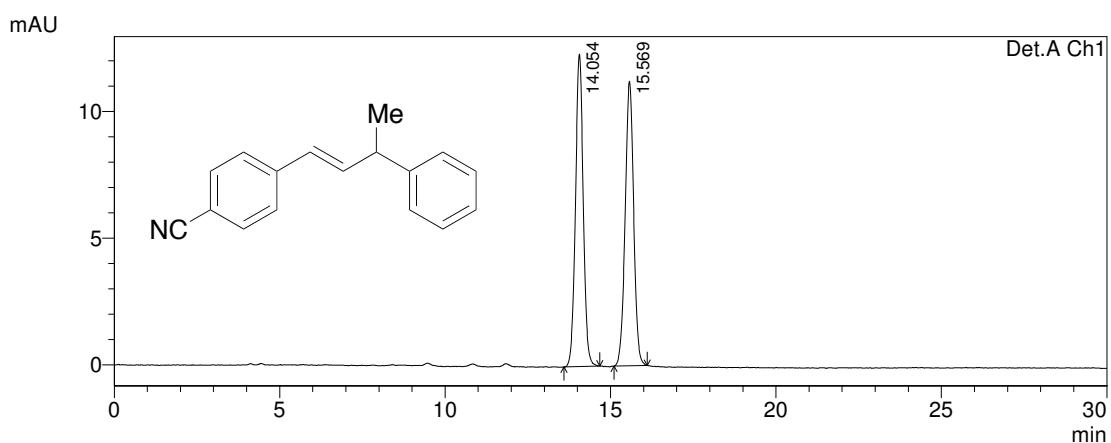
Enantioenriched **19**, ee = 90%



Detector A Ch1 254nm

Peak#	Ret. Time	Area	Height	Area %	Height %
1	15.061	2546825	88146	95.154	95.602
2	17.458	129704	4055	4.846	4.398
Total		2676529	92201	100.000	100.000

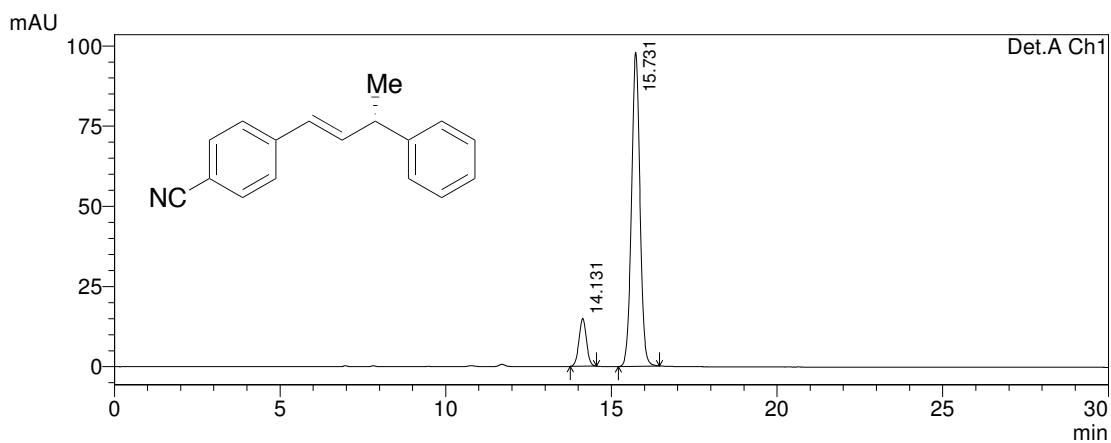
Racemic **20**



Detector A Ch1 254nm

Peak#	Ret. Time	Area	Height	Area %	Height %
1	14.054	200511	12325	50.027	52.327
2	15.569	200293	11229	49.973	47.673
Total		400804	23554	100.000	100.000

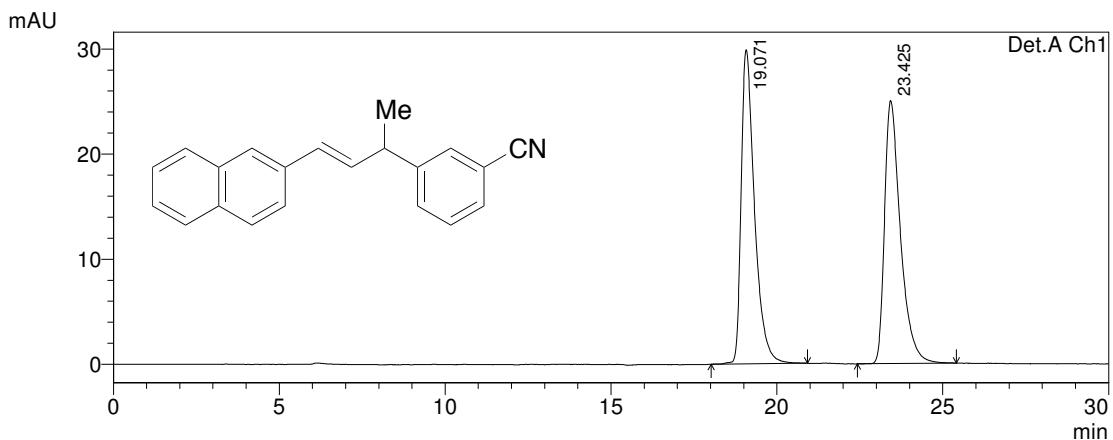
Enantioenriched **20**, ee = 76%



Detector A Ch1 254nm

Peak#	Ret. Time	Area	Height	Area %	Height %
1	14.131	235932	14981	11.961	13.275
2	15.731	1736595	97870	88.039	86.725
Total		1972527	112851	100.000	100.000

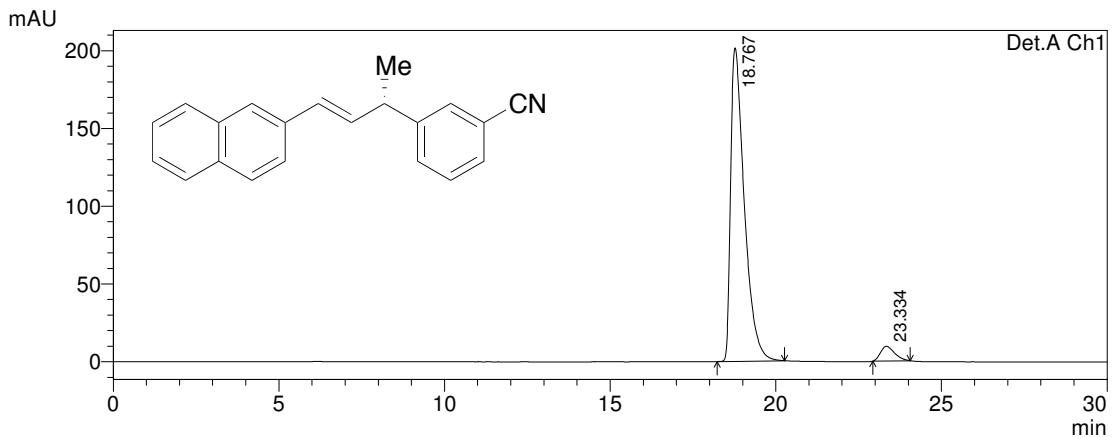
Racemic **21**



Detector A Ch1 254nm

Peak#	Ret. Time	Area	Height	Area %	Height %
1	19.071	835245	29879	50.109	54.427
2	23.425	831604	25018	49.891	45.573
Total		1666849	54897	100.000	100.000

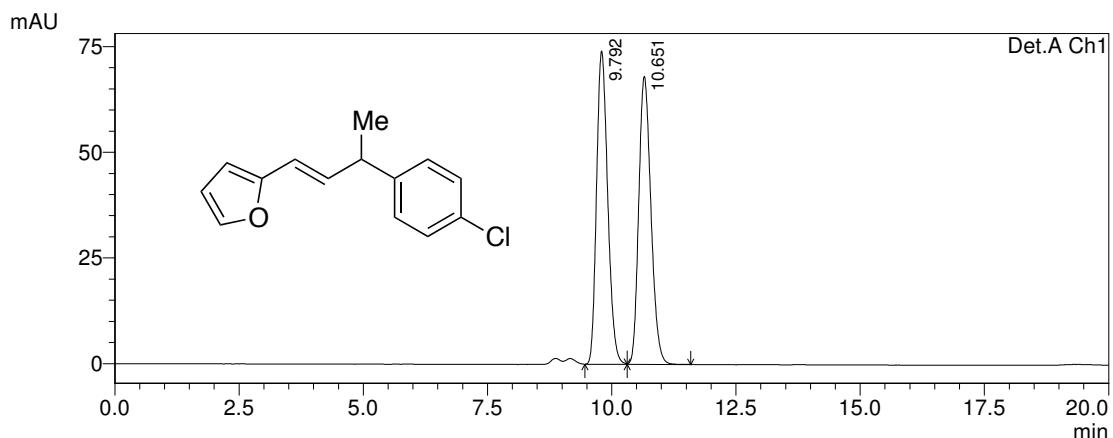
Enantioenriched **21**, ee = 91%



Detector A Ch1 254nm

Peak#	Ret. Time	Area	Height	Area %	Height %
1	18.767	5778302	201515	95.422	95.541
2	23.334	277193	9405	4.578	4.459
Total		6055496	210920	100.000	100.000

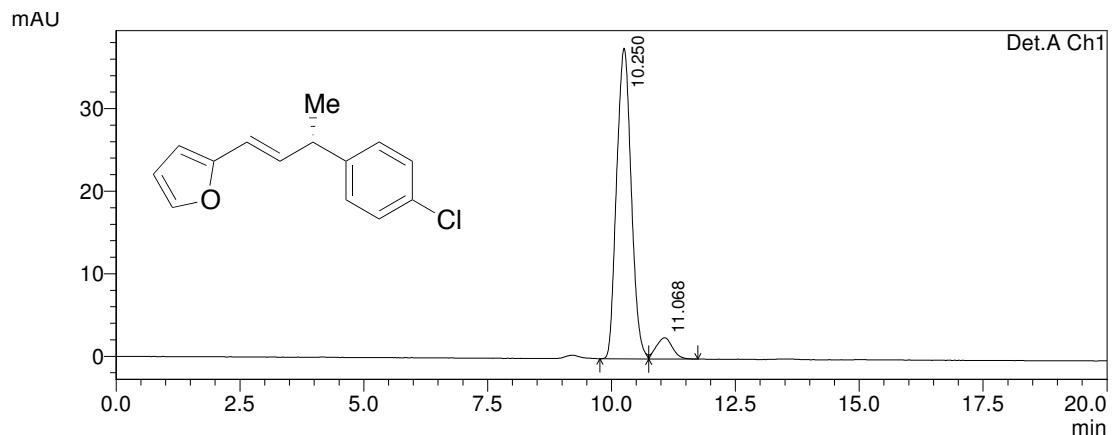
Racemic **22**



Detector A Ch1 254nm

Peak#	Ret. Time	Area	Height	Area %	Height %
1	9.792	1156522	74059	49.899	52.103
2	10.651	1161221	68081	50.101	47.897
Total		2317742	142140	100.000	100.000

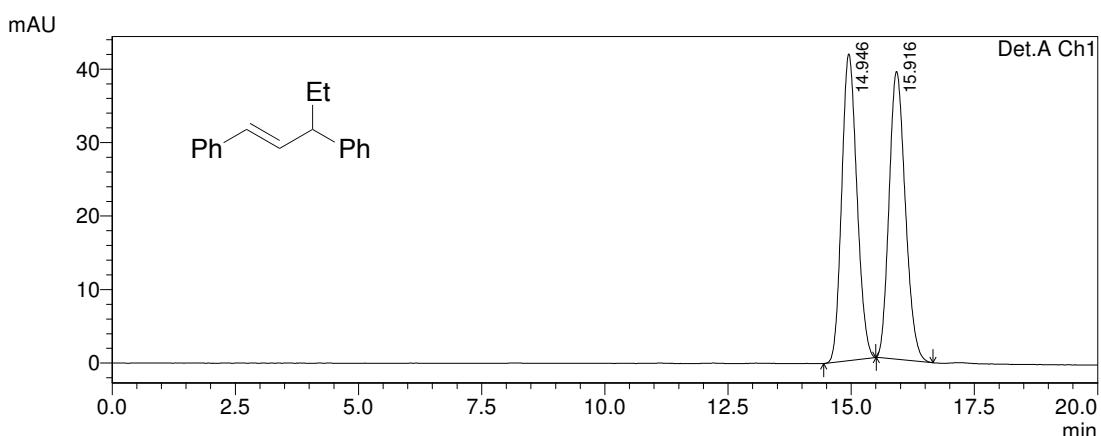
Enantioenriched **22**, ee = 86%



Detector A Ch1 254nm

Peak#	Ret. Time	Area	Height	Area %	Height %
1	10.250	771297	37625	92.947	93.597
2	11.068	58527	2574	7.053	6.403
Total		829824	40198	100.000	100.000

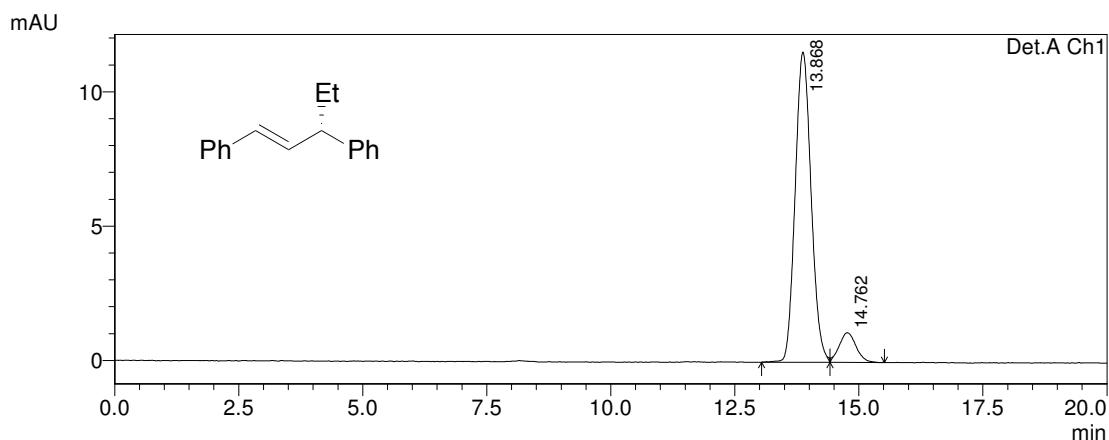
Racemic 23



Detector A Ch1 254nm

Peak#	Ret. Time	Area	Height	Area %	Height %
1	14.946	895104	41713	49.972	51.580
2	15.916	896105	39157	50.028	48.420
Total		1791209	80870	100.000	100.000

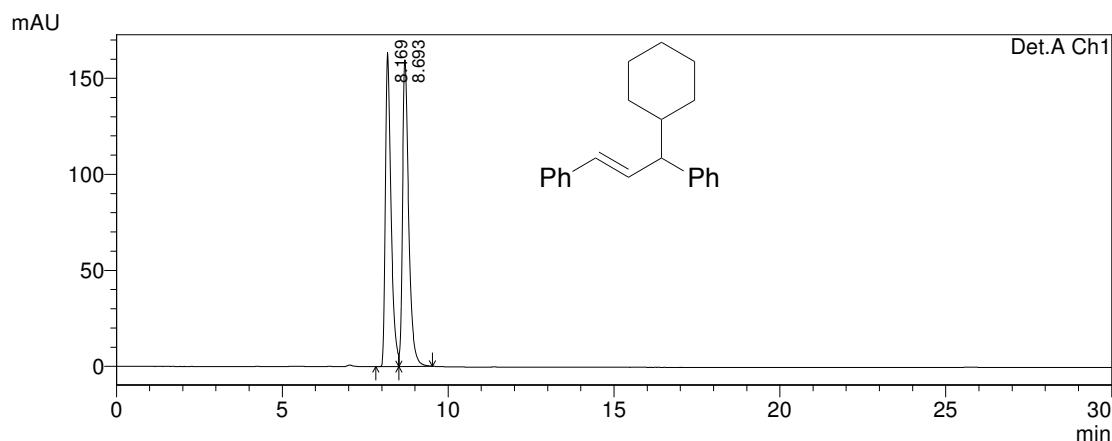
Enantioenriched 23, ee = 82%



Detector A Ch1 254nm

Peak#	Ret. Time	Area	Height	Area %	Height %
1	13.868	258330	11546	90.794	91.272
2	14.762	26194	1104	9.206	8.728
Total		284524	12650	100.000	100.000

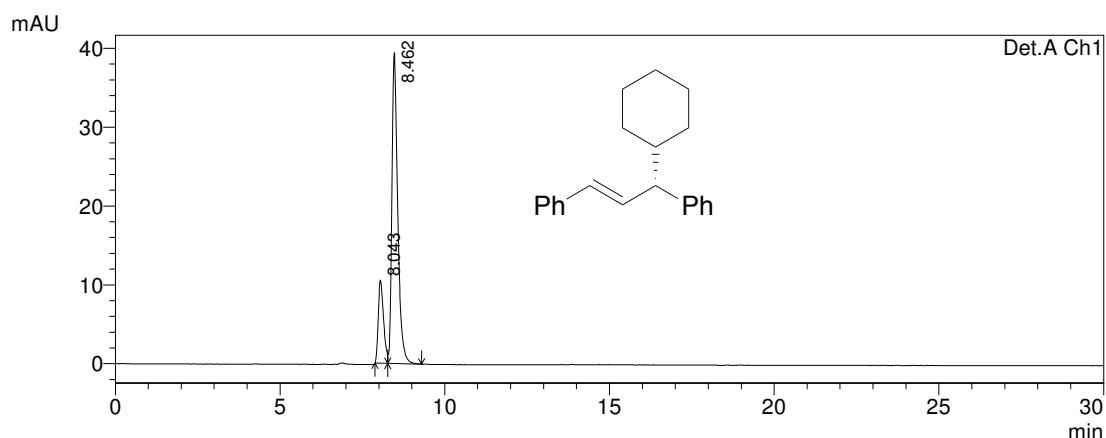
Racemic 24



Detector A Ch1 254nm

Peak#	Ret. Time	Area	Height	Area %	Height %
1	8.169	1940959	163447	48.814	50.644
2	8.693	2035273	159293	51.186	49.356
Total		3976232	322740	100.000	100.000

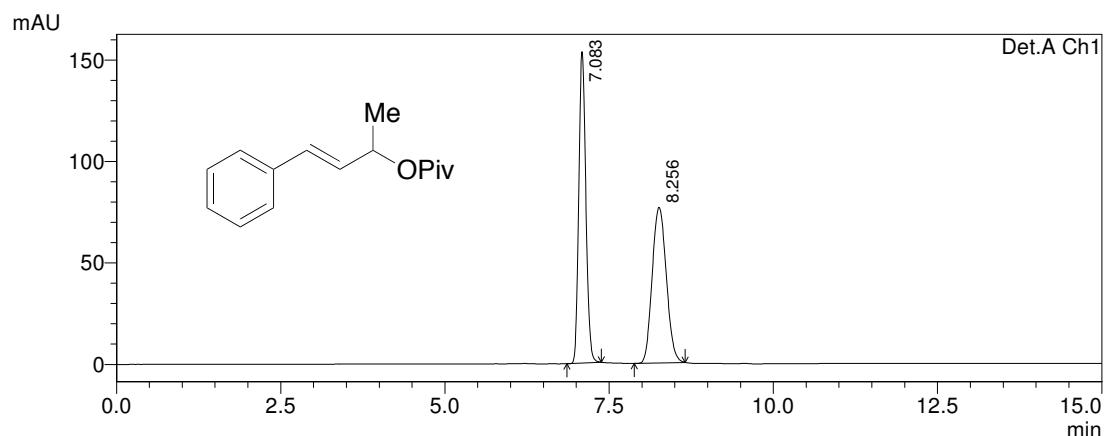
Enantioenriched 24, ee = 61%



Detector A Ch1 254nm

Peak#	Ret. Time	Area	Height	Area %	Height %
1	8.043	116720	10518	19.377	21.073
2	8.462	485645	39394	80.623	78.927
Total		602365	49912	100.000	100.000

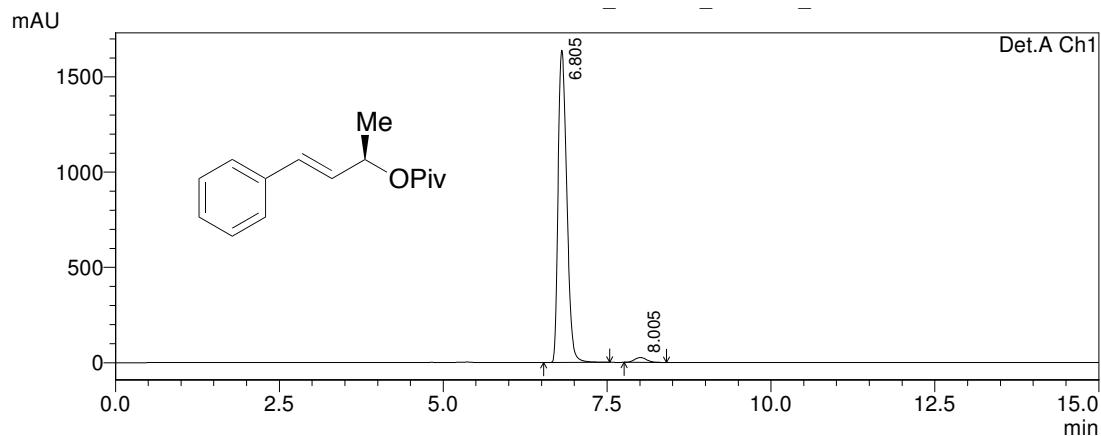
Racemic **1a**



Detector A Ch1 254nm

Peak#	Ret. Time	Area	Height	Area %	Height %
1	7.083	1148930	153378	50.011	66.649
2	8.256	1148419	76750	49.989	33.351
Total		2297349	230128	100.000	100.000

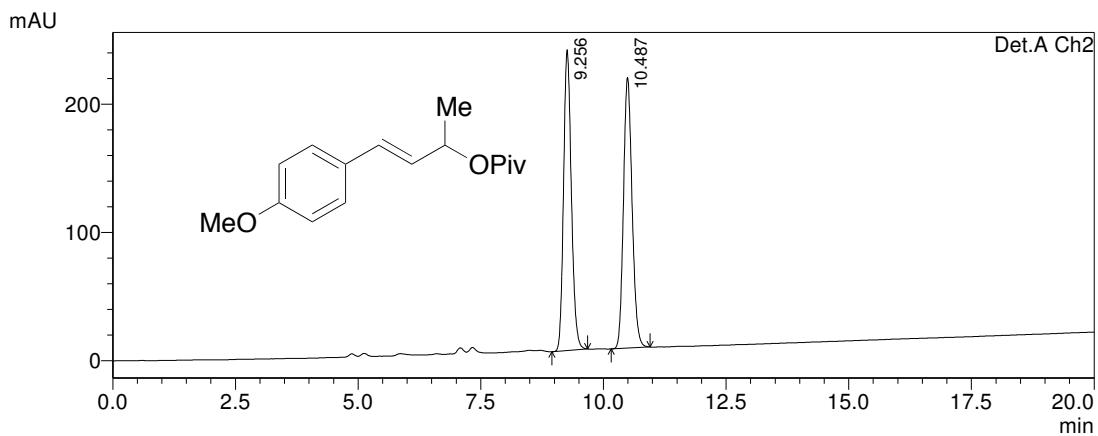
Enantioenriched **1a**, ee = 96%



Detector A Ch1 254nm

Peak#	Ret. Time	Area	Height	Area %	Height %
1	6.805	15085485	1638394	97.867	98.461
2	8.005	328742	25601	2.133	1.539
Total		15414227	1663995	100.000	100.000

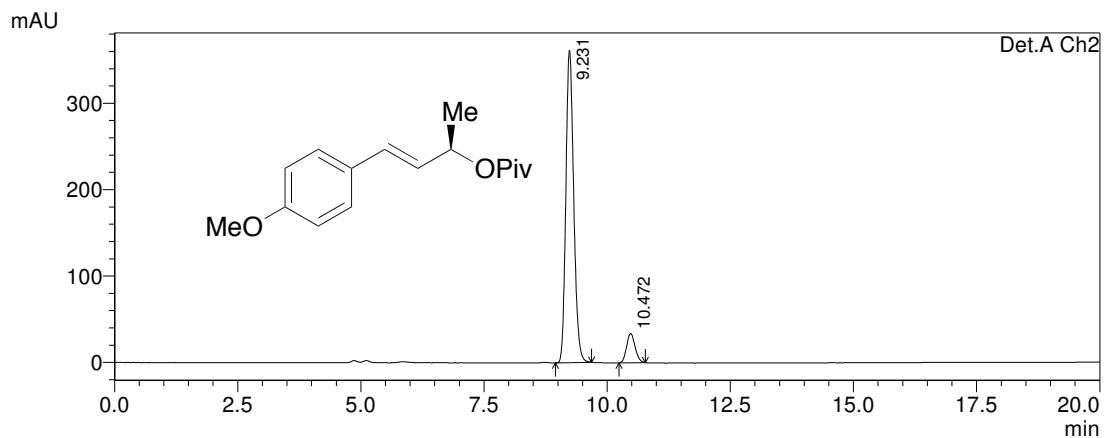
Racemic **1b**



Detector A Ch2 210nm

Peak#	Ret. Time	Area	Height	Area %	Height %
1	9.256	2587105	234367	50.020	52.612
2	10.487	2585032	211095	49.980	47.388
Total		5172137	445462	100.000	100.000

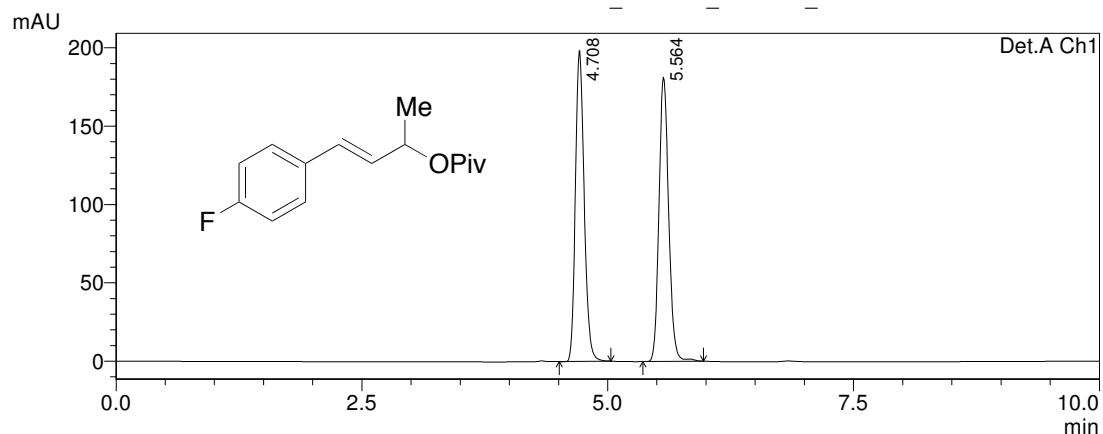
Enantioenriched **1b**, ee = 82%



Detector A Ch2 210nm

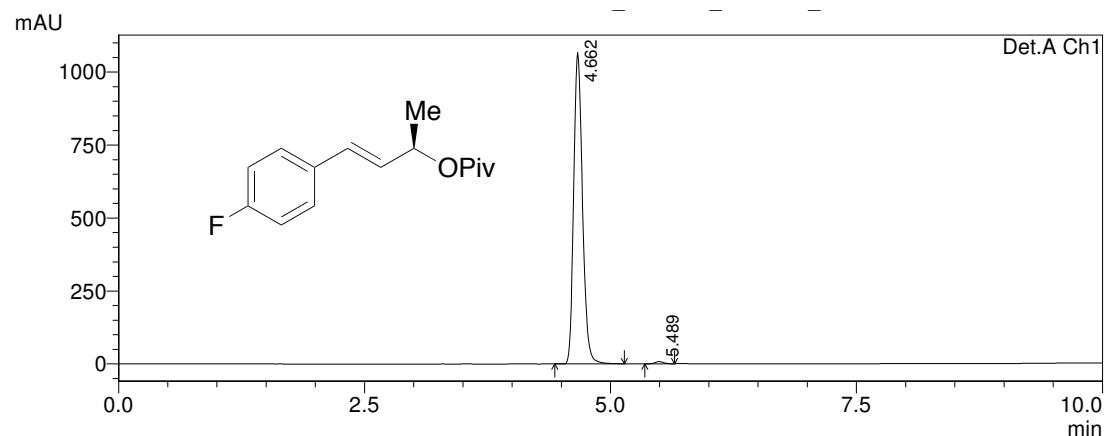
Peak#	Ret. Time	Area	Height	Area %	Height %
1	9.231	4008739	361390	90.806	91.434
2	10.472	405887	33855	9.194	8.566
Total		4414626	395245	100.000	100.000

Racemic **1c**



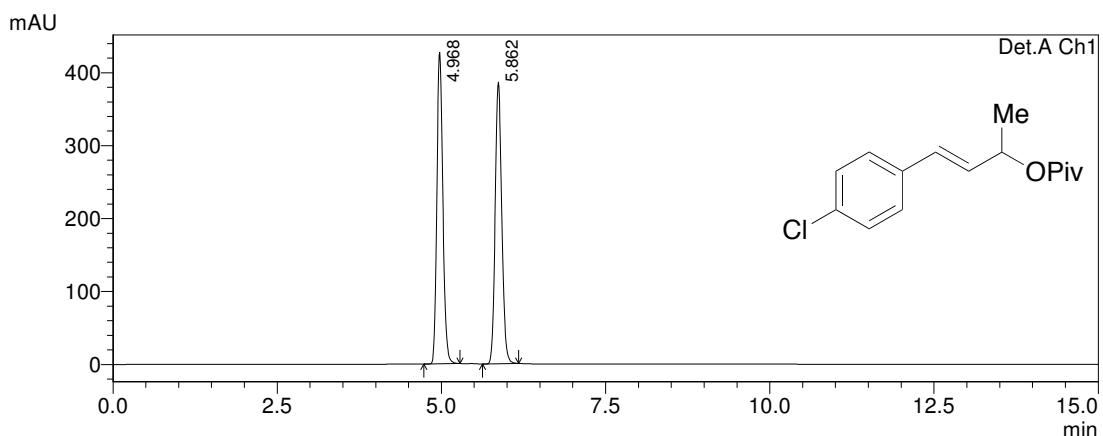
Detector A Ch1 254nm					
Peak#	Ret. Time	Area	Height	Area %	Height %
1	4.708	1233223	198303	49.818	52.220
2	5.564	1242245	181443	50.182	47.780
Total		2475468	379746	100.000	100.000

Enantioenriched **1c**, ee = 98%



Detector A Ch1 254nm					
Peak#	Ret. Time	Area	Height	Area %	Height %
1	4.662	6650232	1066854	99.298	99.295
2	5.489	47028	7574	0.702	0.705
Total		6697260	1074428	100.000	100.000

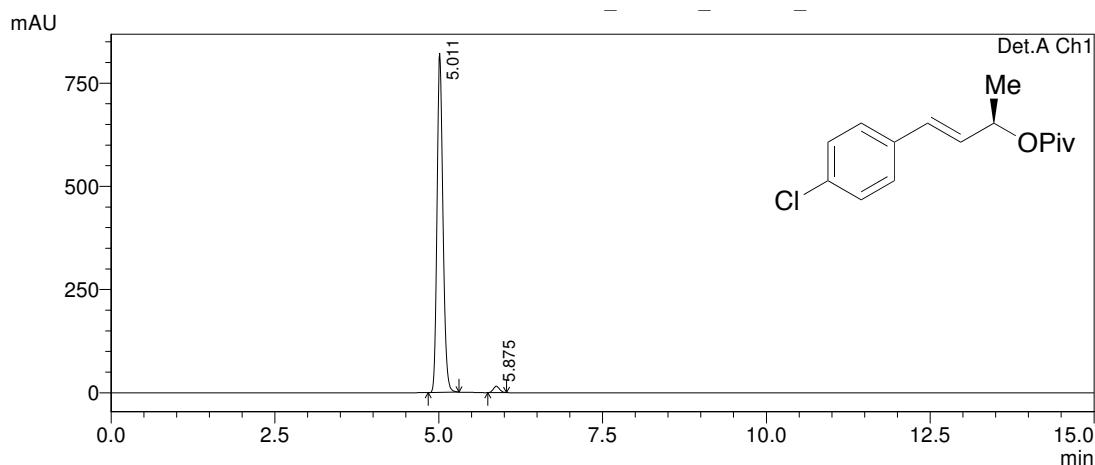
Racemic **1d**



Detector A Ch1 254nm

Peak#	Ret. Time	Area	Height	Area %	Height %
1	4.968	2698912	427008	50.039	52.519
2	5.862	2694758	386051	49.961	47.481
Total		5393670	813059	100.000	100.000

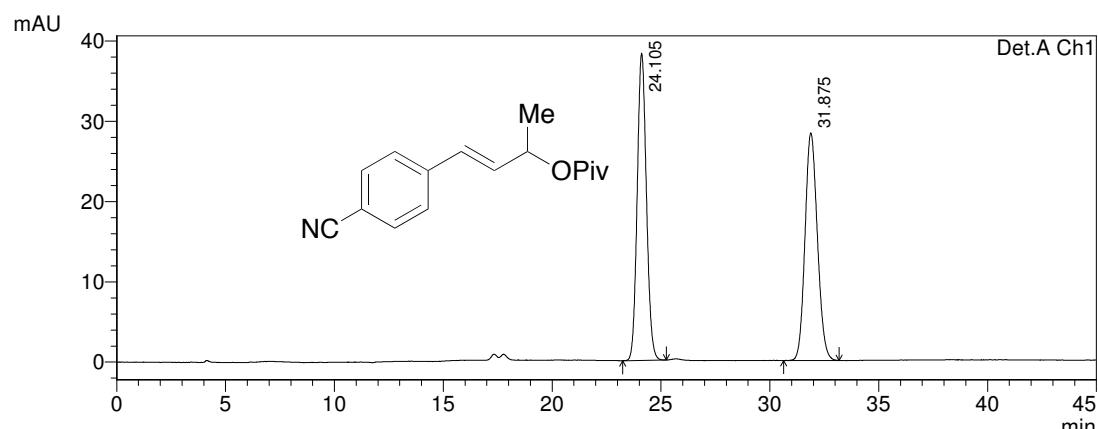
Enantioenriched **1d**, ee = 96%



Detector A Ch1 254nm

Peak#	Ret. Time	Area	Height	Area %	Height %
1	5.011	5220879	821541	98.026	98.146
2	5.875	105126	15518	1.974	1.854
Total		5326005	837059	100.000	100.000

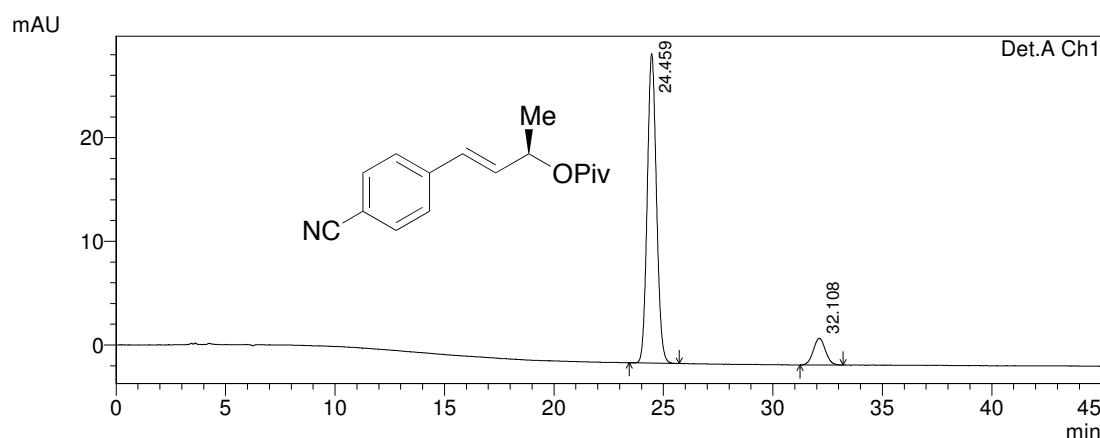
Racemic **1e**



Detector A Ch1 254nm

Peak#	Ret. Time	Area	Height	Area %	Height %
1	24.105	1125881	38286	50.089	57.434
2	31.875	1121870	28374	49.911	42.566
Total		2247751	66661	100.000	100.000

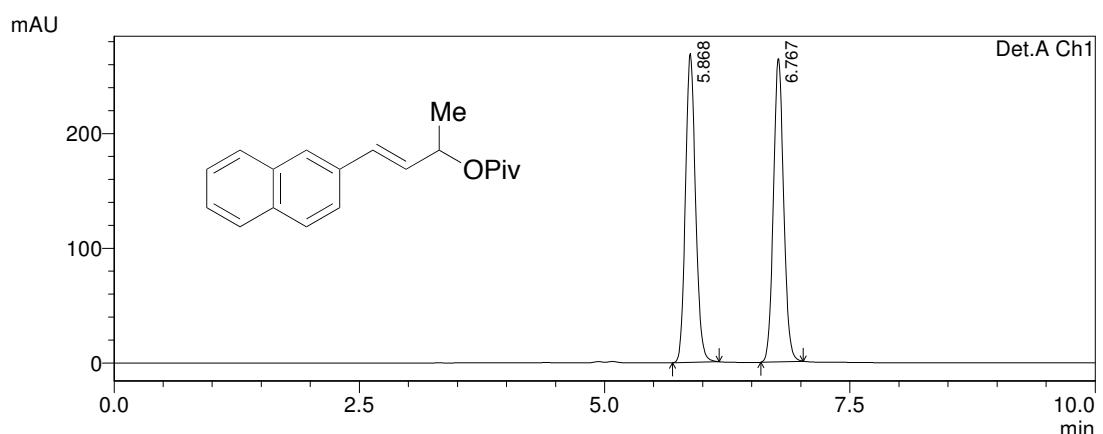
Enantioenriched **1e**, ee = 80%



Detector A Ch1 254nm

Peak#	Ret. Time	Area	Height	Area %	Height %
1	24.459	874057	29841	89.821	92.069
2	32.108	99056	2571	10.179	7.931
Total		973112	32411	100.000	100.000

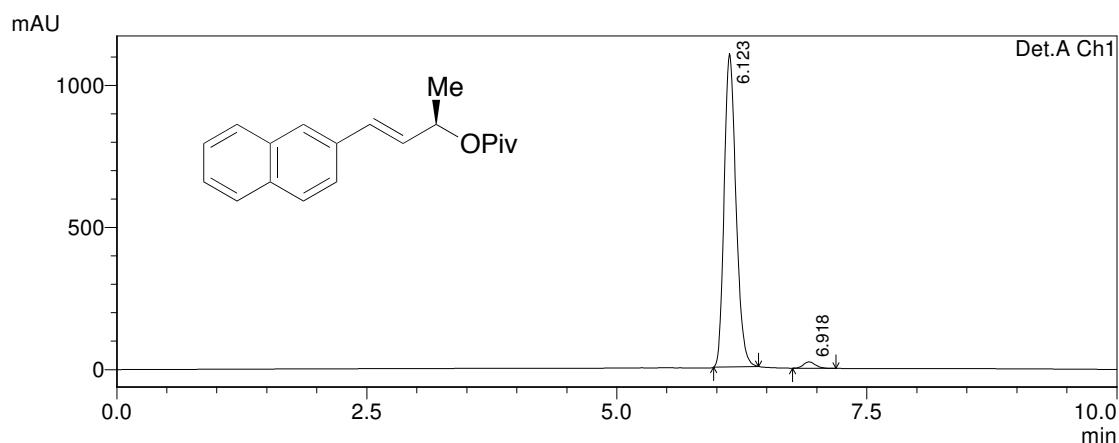
Racemic **1f**



Detector A Ch1 254nm

Peak#	Ret. Time	Area	Height	Area %	Height %
1	5.868	1927376	269194	50.115	50.459
2	6.767	1918496	264295	49.885	49.541
Total		3845872	533490	100.000	100.000

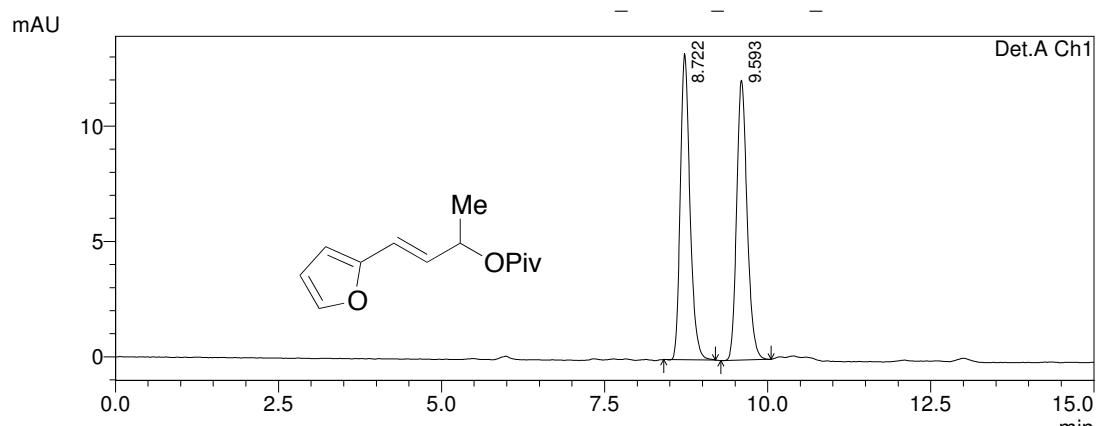
Enantioenriched **1f**, ee = 96%



Detector A Ch1 254nm

Peak#	Ret. Time	Area	Height	Area %	Height %
1	6.123	9044130	1102126	97.972	98.006
2	6.918	187203	22422	2.028	1.994
Total		9231333	1124548	100.000	100.000

Racemic **1g**

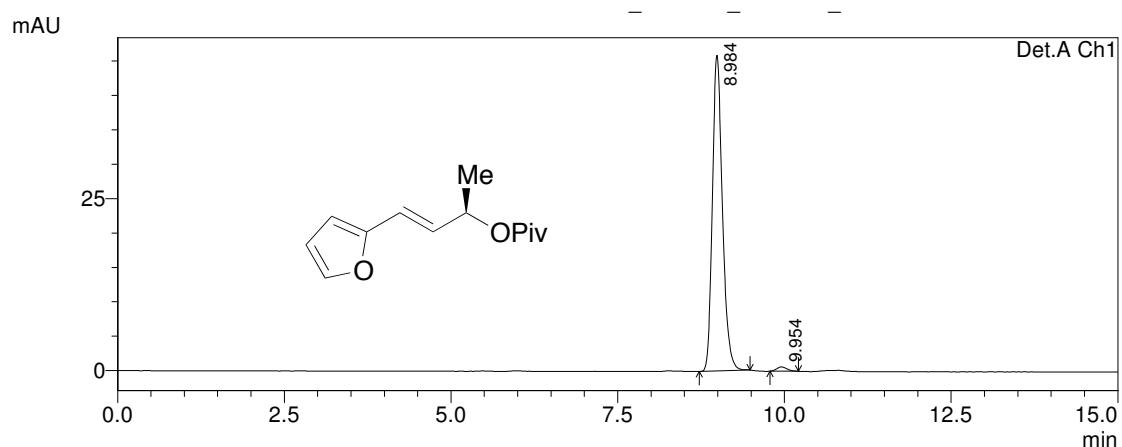


0

Detector A Ch1 254nm

Peak#	Ret. Time	Area	Height	Area %	Height %
1	8.722	136201	13271	50.103	52.242
2	9.593	135642	12132	49.897	47.758
Total		271843	25403	100.000	100.000

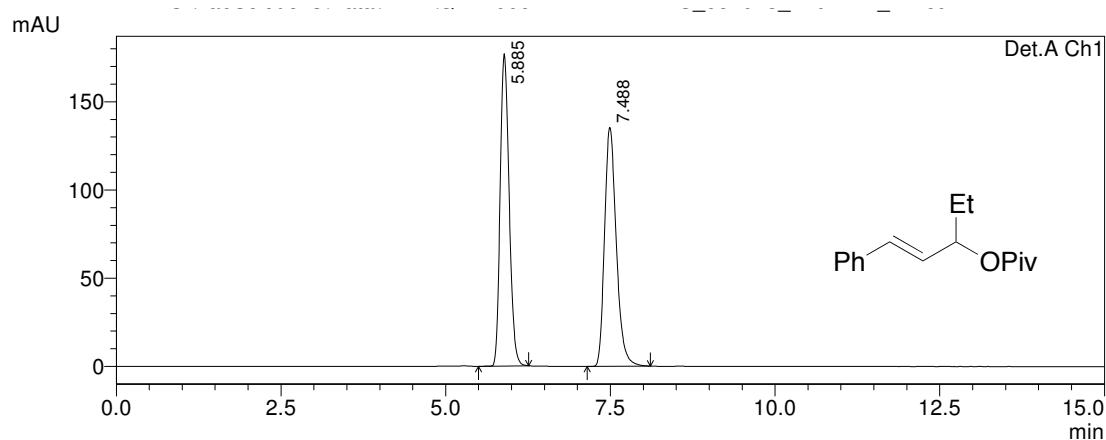
Enantioenriched **1g**, ee = 97%



Detector A Ch1 254nm

Peak#	Ret. Time	Area	Height	Area %	Height %
1	8.984	480703	45863	98.617	98.634
2	9.954	6741	635	1.383	1.366
Total		487444	46498	100.000	100.000

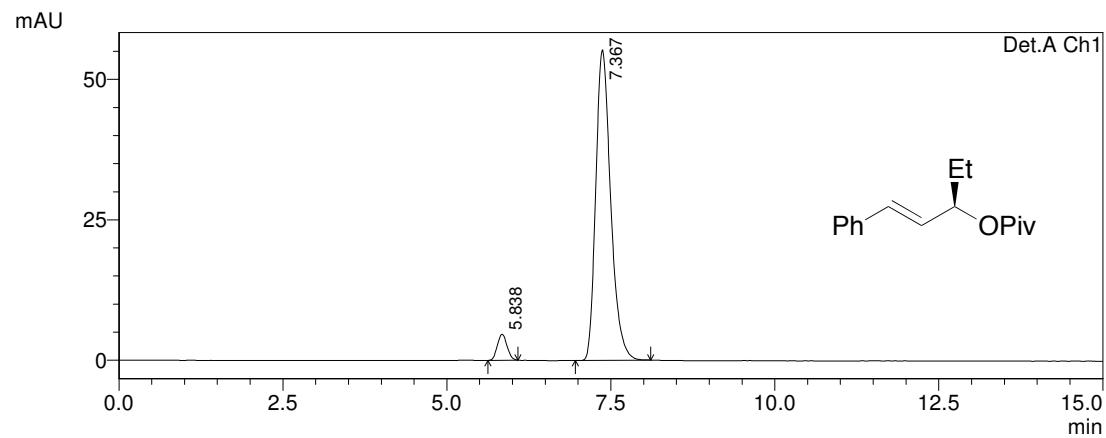
Racemic **1h**



Detector A Ch1 254nm

Peak#	Ret. Time	Area	Height	Area %	Height %
1	5.885	1677000	176994	50.017	56.641
2	7.488	1675869	135488	49.983	43.359
Total		3352870	312481	100.000	100.000

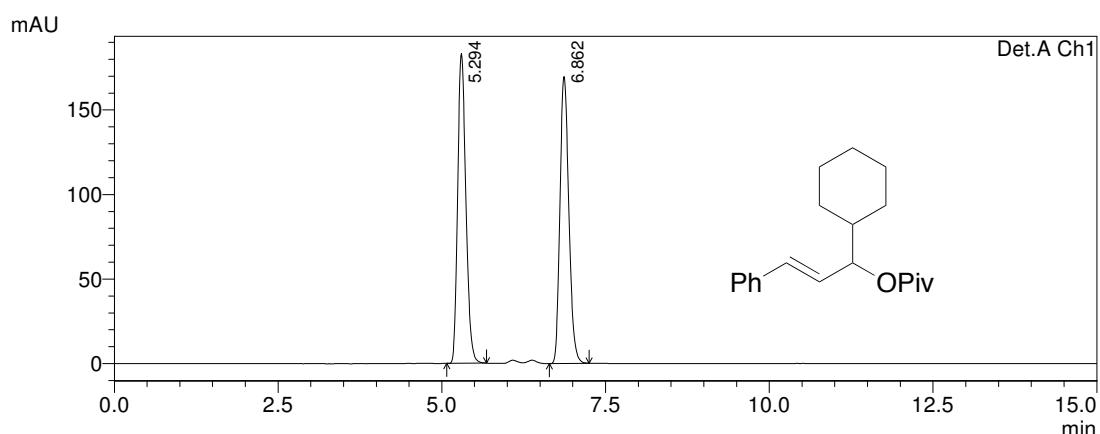
Enantioenriched **1h**, ee = 90%



Detector A Ch1 254nm

Peak#	Ret. Time	Area	Height	Area %	Height %
1	5.838	47604	4634	5.158	7.735
2	7.367	875250	55270	94.842	92.265
Total		922854	59903	100.000	100.000

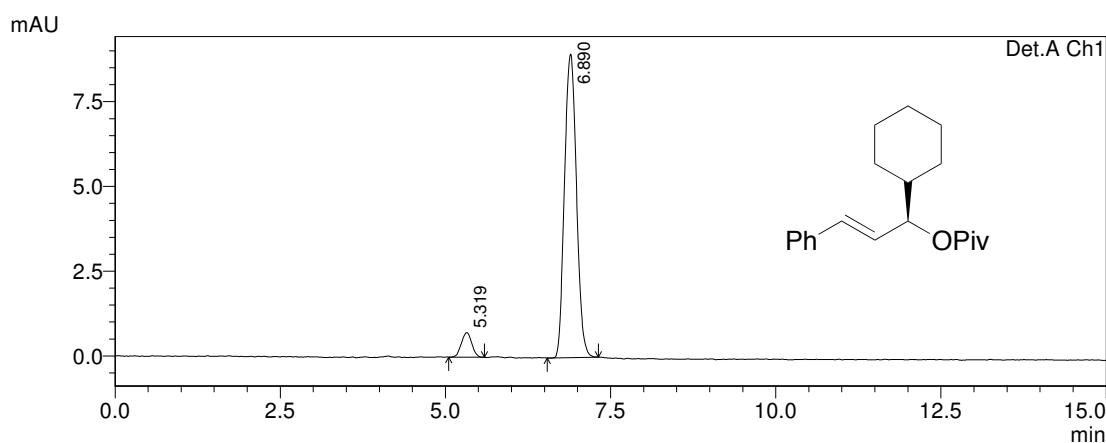
Racemic **1i**



Detector A Ch1 254nm

Peak#	Ret. Time	Area	Height	Area %	Height %
1	5.294	1600071	183097	50.004	51.903
2	6.862	1599846	169669	49.996	48.097
Total		3199916	352766	100.000	100.000

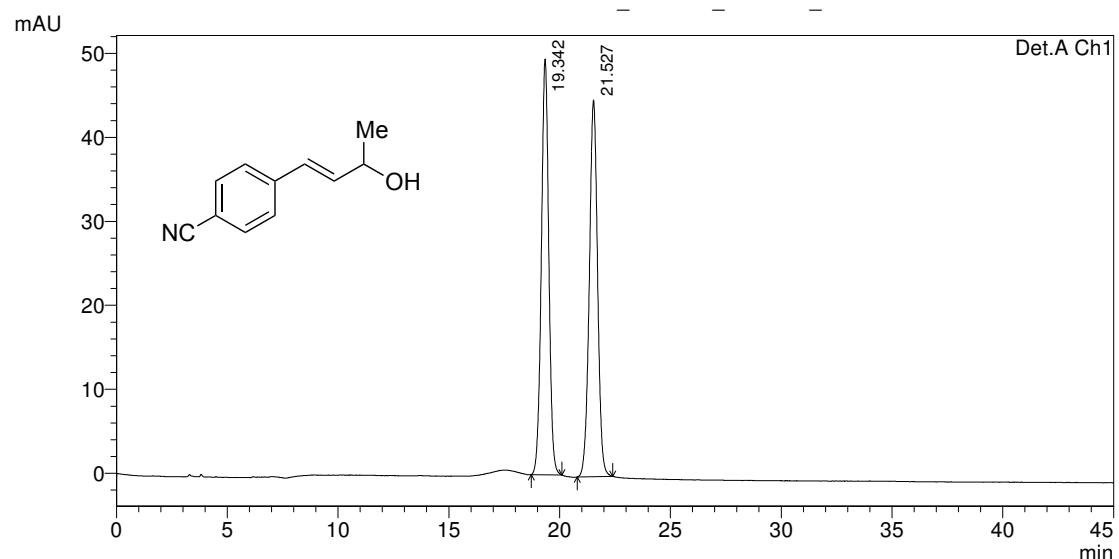
Enantioenriched **1i**, ee = 87%



Detector A Ch1 254nm

Peak#	Ret. Time	Area	Height	Area %	Height %
1	5.319	7957	726	6.585	7.505
2	6.890	112885	8953	93.415	92.495
Total		120842	9680	100.000	100.000

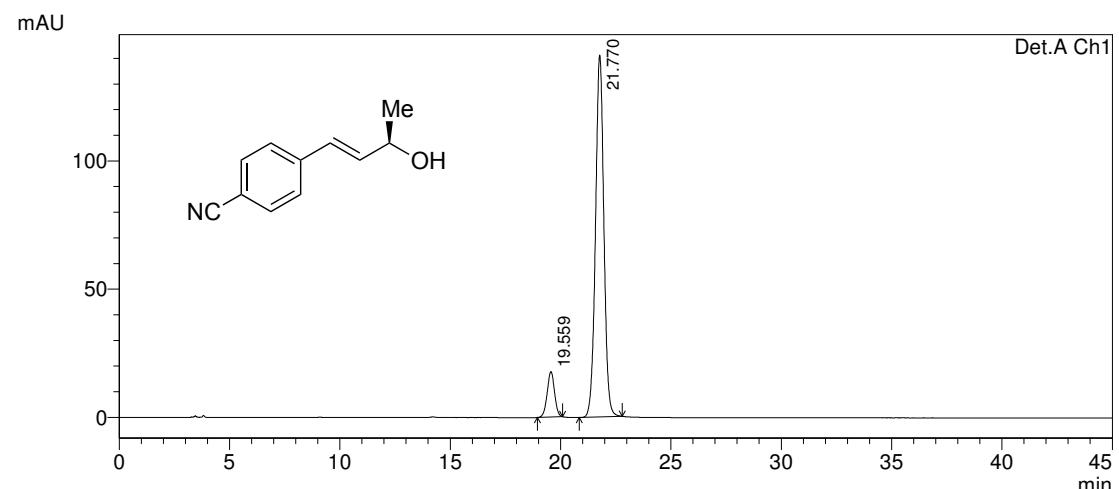
Racemic S-1e



Detector A Ch1 254nm

Peak#	Ret. Time	Area	Height	Area %	Height %
1	19.342	1155111	49477	49.791	52.459
2	21.527	1164813	44838	50.209	47.541
Total		2319925	94314	100.000	100.000

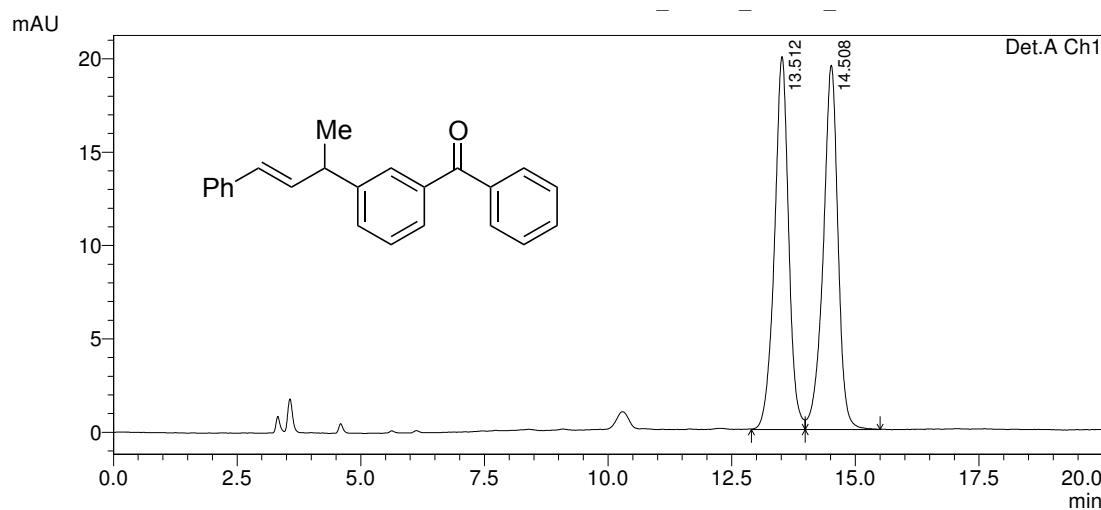
Enantioenriched S-1e, 80% ee



Detector A Ch1 254nm

Peak#	Ret. Time	Area	Height	Area %	Height %
1	19.559	411243	17736	9.981	11.160
2	21.770	3708859	141186	90.019	88.840
Total		4120101	158922	100.000	100.000

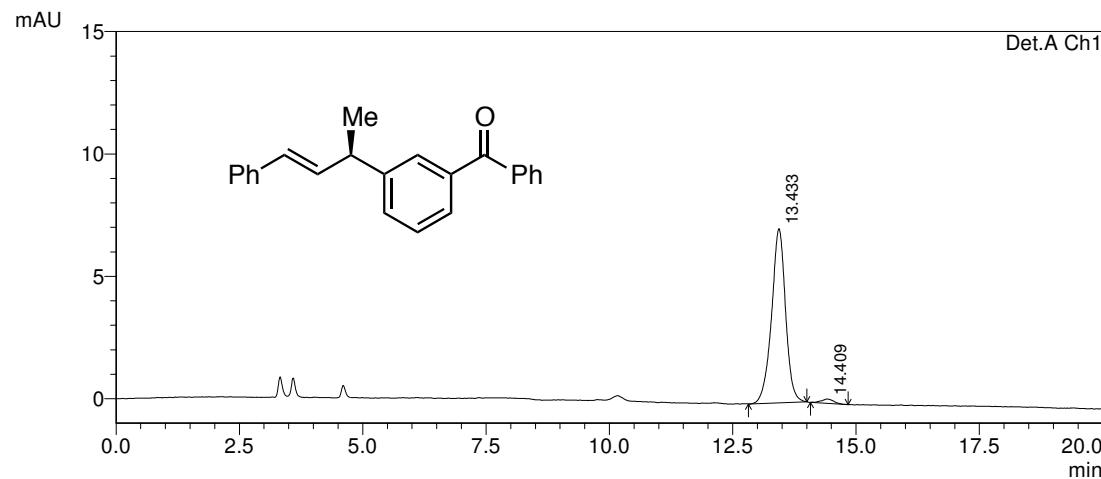
Racemic **25**



Detector A Ch1 220nm

Peak#	Ret. Time	Area	Height	Area %	Height %
1	13.512	399033	19975	49.585	50.590
2	14.508	405720	19509	50.415	49.410
Total		804753	39484	100.000	100.000

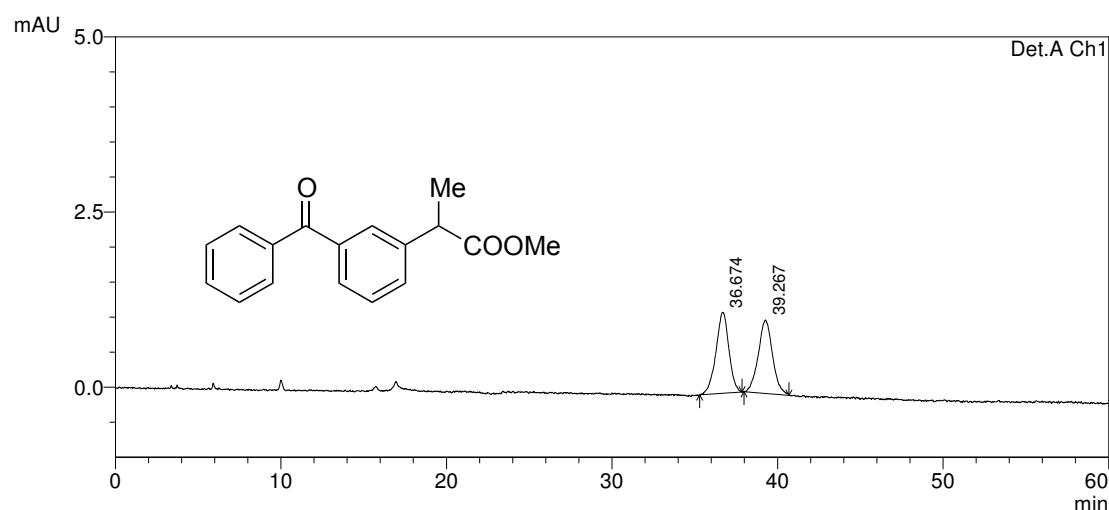
Enantioenriched **25**, 95% ee



Detector A Ch1 220nm

Peak#	Ret. Time	Area	Height	Area %	Height %
1	13.433	140135	7107	97.873	97.615
2	14.409	3045	174	2.127	2.385
Total		143180	7281	100.000	100.000

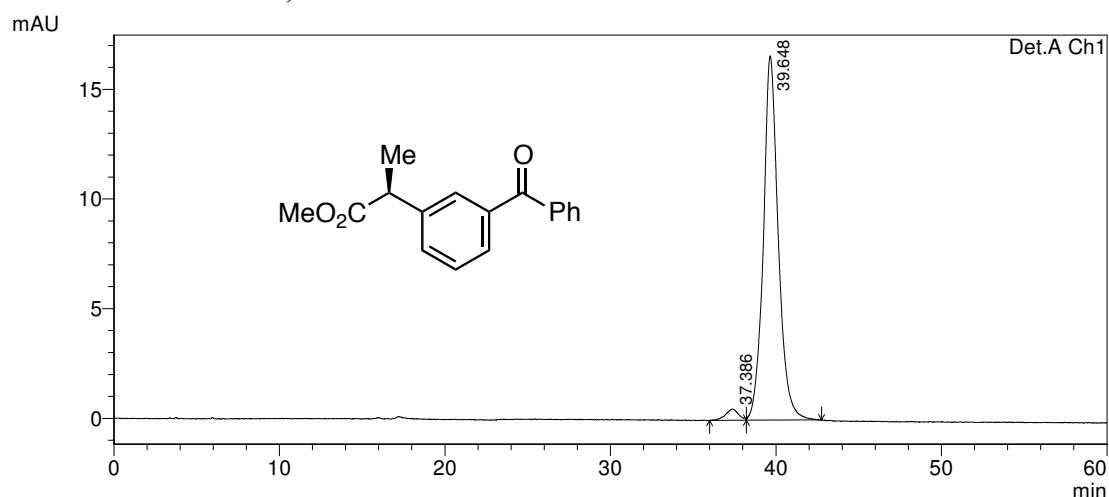
Racemic S-29



Detector A Ch1 254nm

Peak#	Ret. Time	Area	Height	Area %	Height %
1	36.674	62988	1156	50.223	52.472
2	39.267	62430	1047	49.777	47.528
Total		125418	2203	100.000	100.000

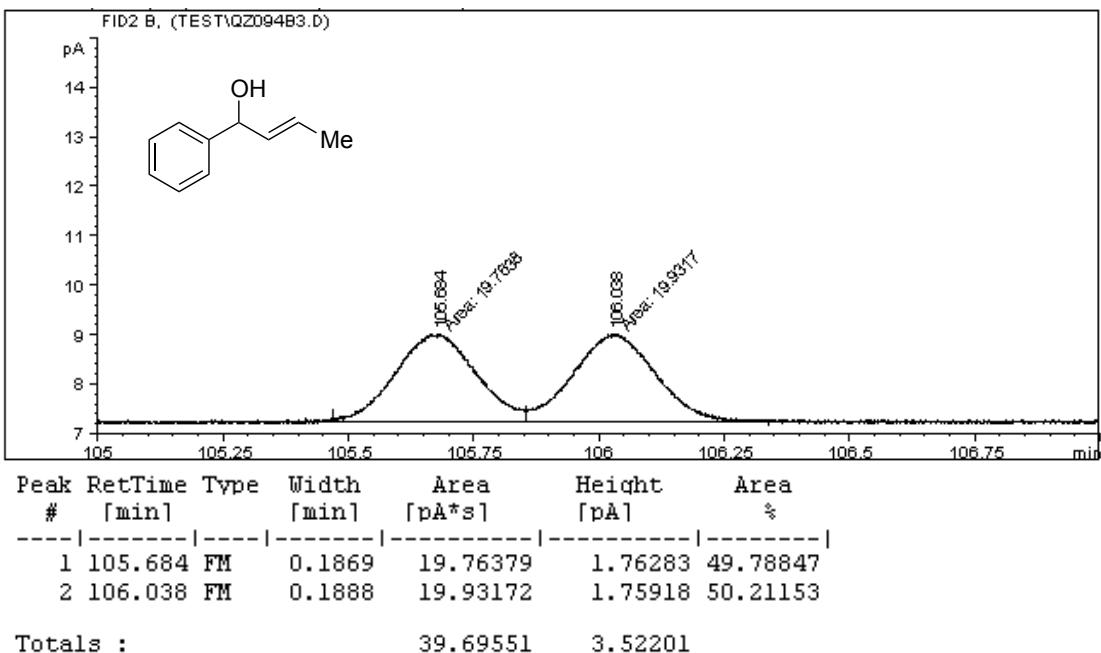
Enantioenriched S-29, 95% ee



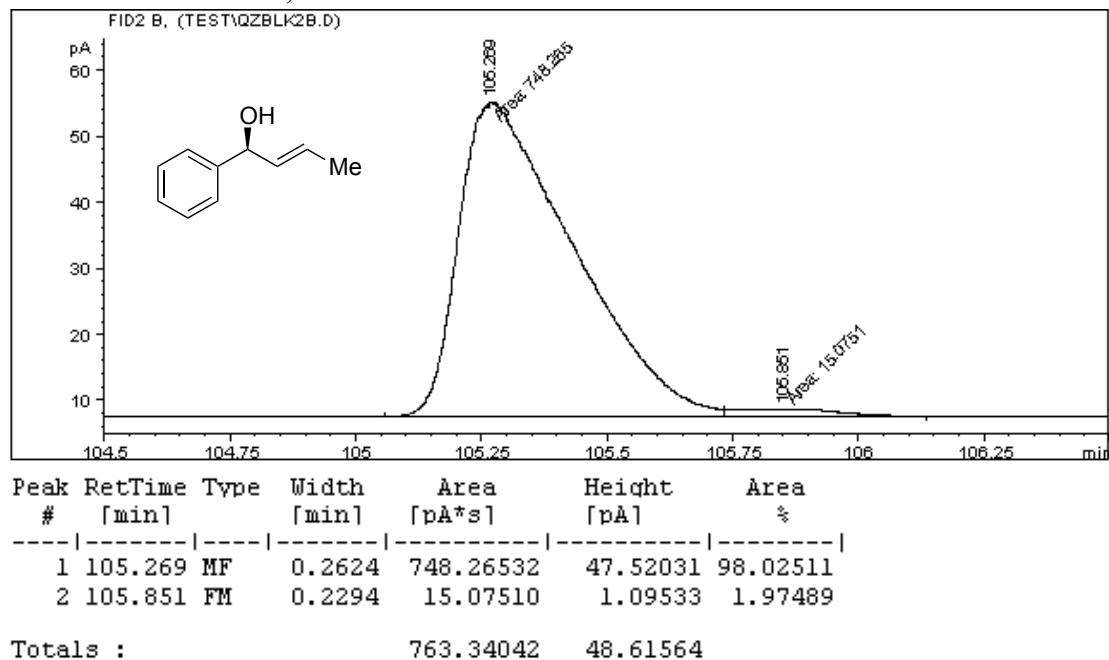
Detector A Ch1 254nm

Peak#	Ret. Time	Area	Height	Area %	Height %
1	37.386	27762	513	2.592	2.995
2	39.648	1043427	16616	97.408	97.005
Total		1071188	17129	100.000	100.000

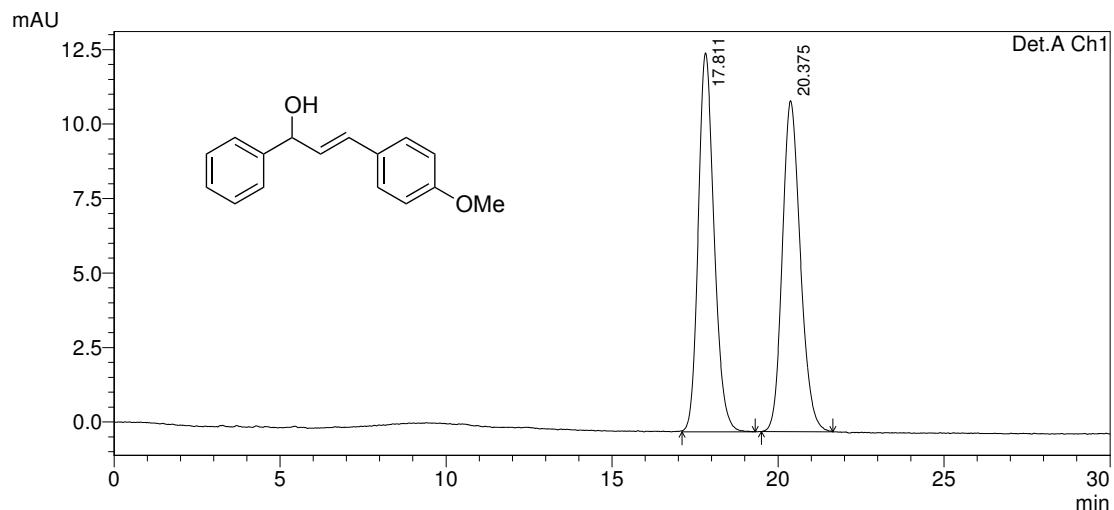
Racemic S-27



Enantioenriched S-27, 96% ee



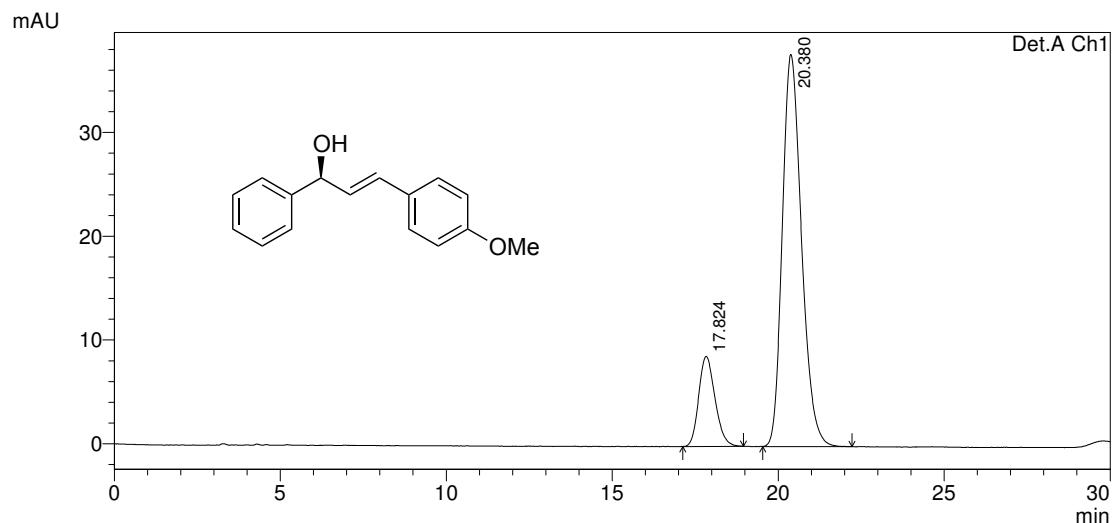
Racemic S-28



Detector A Ch1 254nm

Peak#	Ret. Time	Area	Height	Area %	Height %
1	17.811	421028	12719	50.067	53.386
2	20.375	419895	11106	49.933	46.614
Total		840923	23825	100.000	100.000

Enantioenriched S-28, 67% ee

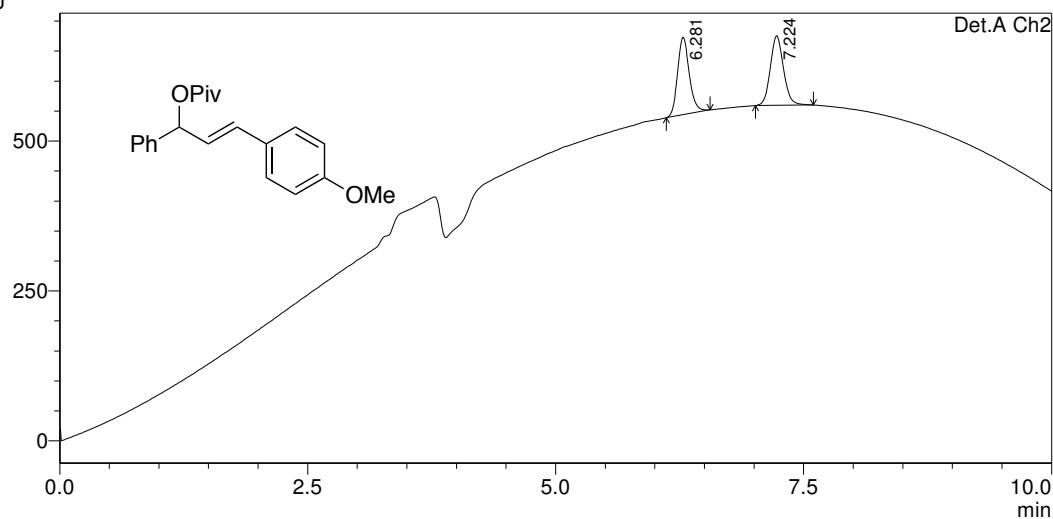


Detector A Ch1 254nm

Peak#	Ret. Time	Area	Height	Area %	Height %
1	17.824	297238	8687	16.342	18.689
2	20.380	1521674	37796	83.658	81.311
Total		1818913	46483	100.000	100.000

Racemic **28**

mAU

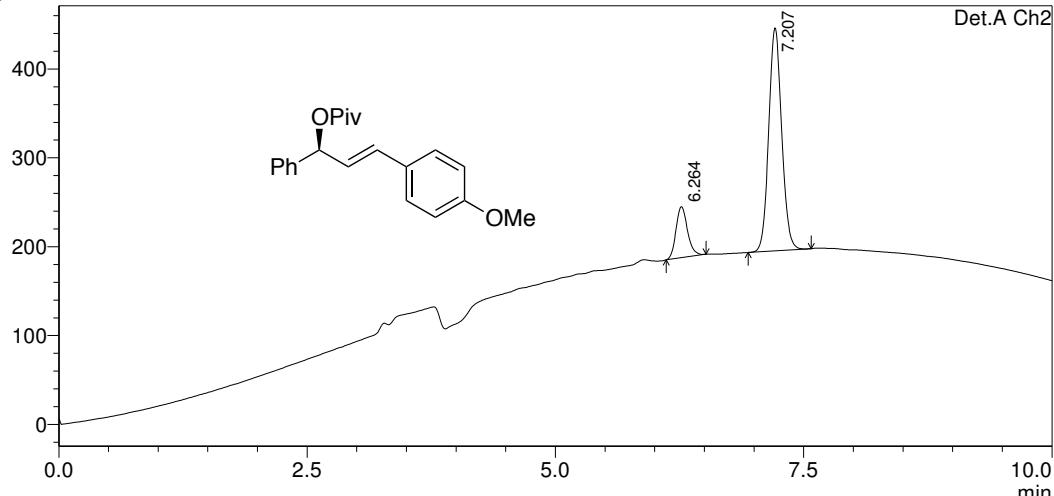


Detector A Ch2 210nm

Peak#	Ret. Time	Area	Height	Area %	Height %
1	6.281	1057077	129088	49.605	52.686
2	7.224	1073915	115925	50.395	47.314
Total		2130992	245013	100.000	100.000

Enantioenriched **28**, 66% ee

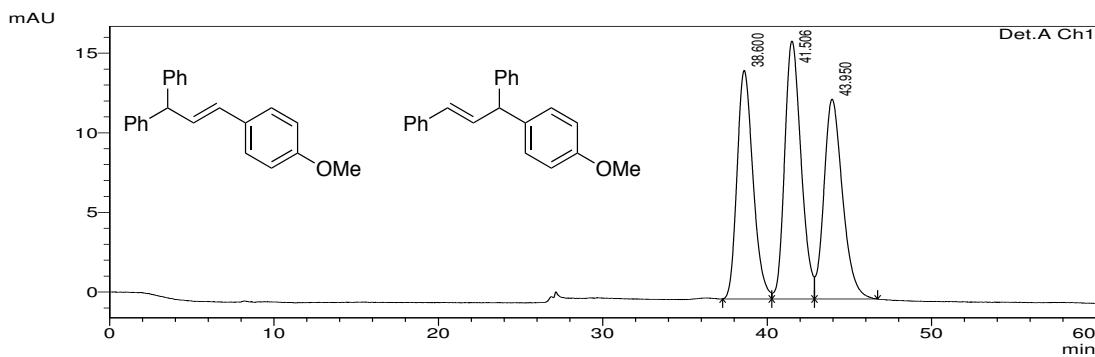
mAU



Detector A Ch2 210nm

Peak#	Ret. Time	Area	Height	Area %	Height %
1	6.264	463024	57592	16.886	18.670
2	7.207	2279005	250877	83.114	81.330
Total		2742028	308469	100.000	100.000

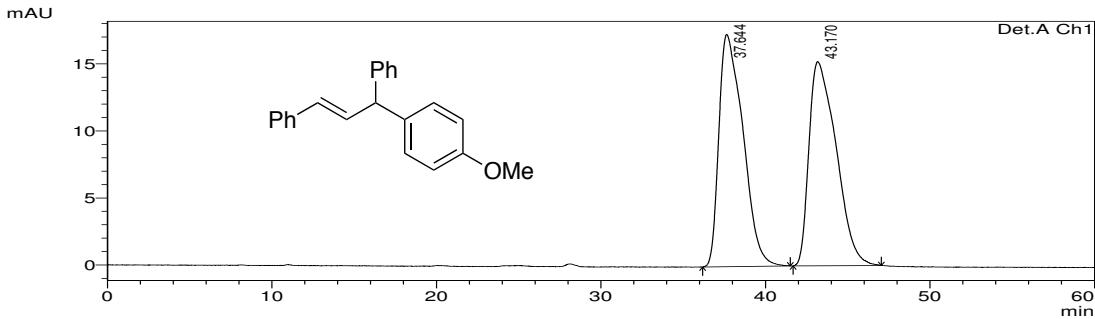
Mixture of **29** and Racemic **30**



Detector A Ch1 254nm

Peak#	Ret. Time	Area	Height	Area %	Height %
1	38.600	968147	14354	31.460	33.306
2	41.506	1121651	16195	36.449	37.578
3	43.950	987549	12548	32.091	29.116
Total		3077347	43098	100.000	100.000

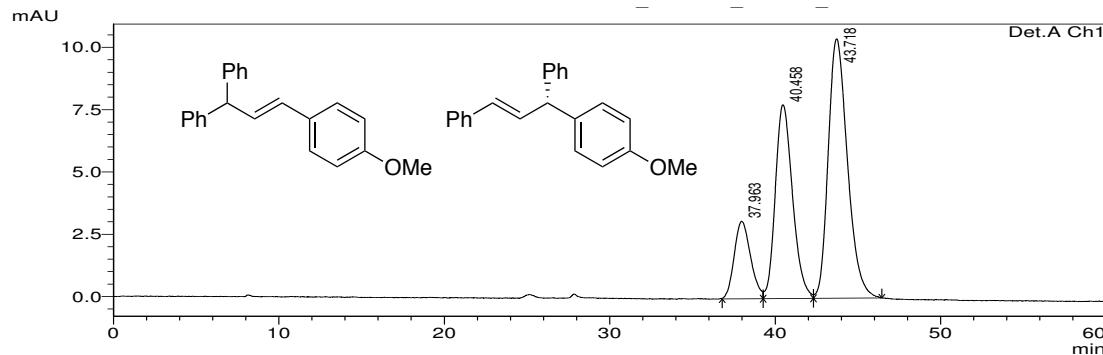
Racemic **30**



Detector A Ch1 254nm

Peak#	Ret. Time	Area	Height	Area %	Height %
1	37.644	1763496	17331	50.233	53.226
2	43.170	1747151	15230	49.767	46.774
Total		3510647	32562	100.000	100.000

Mixture of **29** and Enantioenriched **30** (60% ee)



Detector A Ch1 254nm

Peak#	Ret. Time	Area	Height	Area %	Height %
1	37.963	213435	3115	13.183	14.627
2	40.458	563166	7775	34.784	36.504
3	43.718	842453	10408	52.034	48.869
Total		1619054	21299	100.000	100.000

Crystal Structure Report for (S)-17 (CCDC-973168)

A specimen of C₁₆H₁₄Cl₂, approximate dimensions 0.270 mm x 0.400 mm x 0.440 mm, was used for the X-ray crystallographic analysis. The X-ray intensity data were measured.

The total exposure time was 9.95 hours. The frames were integrated with the Bruker SAINT software package using a narrow-frame algorithm. The integration of the data using an orthorhombic unit cell yielded a total of 20983 reflections to a maximum θ angle of 73.55° (0.80 Å resolution), of which 2806 were independent (average redundancy 7.478, completeness = 98.9%, R_{int} = 3.25%, R_{sig} = 2.06%) and 2788 (99.36%) were greater than 2σ(F²). The final cell constants of a = 5.6849(8) Å, b = 7.6858(10) Å, c = 32.425(4) Å, volume = 1416.7(3) Å³, are based upon the refinement of the XYZ-centroids of 9863 reflections above 20 σ(I) with 11.83° < 2θ < 147.1°. Data were corrected for absorption effects using the multi-scan method (SADABS). The ratio of minimum to maximum apparent transmission was 0.724. The calculated minimum and maximum transmission coefficients (based on crystal size) are 0.2748 and 0.4188.

The structure was solved and refined using the Bruker SHELXTL Software Package, using the space group P 21 21 21, with Z = 4 for the formula unit, C₁₆H₁₄Cl₂. The final anisotropic full-matrix least-squares refinement on F² with 164 variables converged at R1 = 2.88%, for the observed data and wR2 = 8.08% for all data. The goodness-of-fit was 1.025. The largest peak in the final difference electron density synthesis was 0.276 e⁻/Å³ and the largest hole was -0.220 e⁻/Å³ with an RMS deviation of 0.036 e⁻/Å³. On the basis of the final model, the calculated density was 1.299 g/cm³ and F(000), 576 e⁻.

Table 1. Sample and crystal data for (S)-17.

Identification code	mary015
Chemical formula	C ₁₆ H ₁₄ Cl ₂
Formula weight	277.17
Temperature	200(2) K
Wavelength	1.54178 Å
Crystal size	0.270 x 0.400 x 0.440 mm
Crystal system	orthorhombic
Space group	P 21 21 21
Unit cell dimensions	a = 5.6849(8) Å α = 90° b = 7.6858(10) Å β = 90° c = 32.425(4) Å γ = 90°
Volume	1416.7(3) Å ³
Z	4
Density (calculated)	1.299 g/cm ³
Absorption coefficient	3.932 mm ⁻¹
F(000)	576

Table 2. Data collection and structure refinement for (S)-17.

Theta range for data collection	2.73 to 73.55°
Index ranges	-7<=h<=7, -9<=k<=9, -32<=l<=39
Reflections collected	20983
Independent reflections	2806 [R(int) = 0.0325]
Coverage of independent reflections	98.9%
Absorption correction	multi-scan
Max. and min. transmission	0.4188 and 0.2748
Structure solution technique	direct methods
Structure solution program	SHELXS-97 (Sheldrick, 2008)
Refinement method	Full-matrix least-squares on F ²
Refinement program	SHELXL-97 (Sheldrick, 2008)
Function minimized	$\Sigma w(F_o^2 - F_c^2)^2$
Data / restraints / parameters	2806 / 0 / 164
Goodness-of-fit on F²	1.025
Δ/σ_{\max}	0.001
Final R indices	2788 data; I>2σ(I) R1 = 0.0288, wR2 = 0.0806 all data R1 = 0.0290, wR2 = 0.0808
Weighting scheme	w=1/[σ ² (F _o ²)+(0.0478P) ² +0.3632P] where P=(F _o ² +2F _c ²)/3
Absolute structure parameter	0.0(0)
Largest diff. peak and hole	0.276 and -0.220 eÅ ⁻³
R.M.S. deviation from mean	0.036 eÅ ⁻³

Table 3. Atomic coordinates and equivalent isotropic atomic displacement parameters (Å²) for (S)-17.

U(eq) is defined as one third of the trace of the orthogonalized U_{ij} tensor.

	x/a	y/b	z/c	U(eq)
C1	0.4469(3)	0.1683(2)	0.21565(5)	0.0349(4)
C2	0.4286(3)	0.1699(2)	0.25857(5)	0.0356(4)
C3	0.2386(3)	0.25589(19)	0.27638(5)	0.0325(3)
C4	0.0703(3)	0.3383(2)	0.25260(6)	0.0374(4)
C5	0.0940(3)	0.3351(2)	0.20982(5)	0.0356(4)
C6	0.2809(3)	0.25149(19)	0.19077(5)	0.0295(3)
C7	0.3019(3)	0.2518(2)	0.14363(5)	0.0341(3)
C8	0.2461(3)	0.0744(2)	0.12590(5)	0.0339(4)

x/a	y/b	z/c	U(eq)
C9 0.0853(3)	0.0478(2)	0.09675(5)	0.0321(3)
C10 0.8084(3)	0.8693(2)	0.05562(5)	0.0325(3)
C11 0.7382(3)	0.7147(2)	0.03709(5)	0.0340(3)
C12 0.8825(3)	0.5704(2)	0.03977(5)	0.0314(3)
C13 0.0957(3)	0.5769(2)	0.06084(5)	0.0347(4)
C14 0.1620(3)	0.7316(2)	0.07961(5)	0.0331(3)
C15 0.0207(3)	0.8804(2)	0.07753(4)	0.0293(3)
C16 0.5463(4)	0.3127(3)	0.12941(6)	0.0493(5)
C11 0.21302(11)	0.26192(6)	0.330252(13)	0.05065(14)
C12 0.79771(10)	0.37802(6)	0.015223(13)	0.04754(14)

Table 4. Bond lengths (Å) for (S)-17.

C1-C2	1.396(2)	C1-C6	1.396(2)
C1-H1	0.95	C2-C3	1.391(2)
C2-H2	0.95	C3-C4	1.383(2)
C3-C11	1.7535(16)	C4-C5	1.394(3)
C4-H4	0.95	C5-C6	1.387(2)
C5-H5	0.95	C6-C7	1.533(2)
C7-C8	1.513(2)	C7-C16	1.537(3)
C7-H7	1.0	C8-C9	1.331(2)
C8-H8	0.95	C9-C15	1.476(2)
C9-H9	0.95	C10-C11	1.390(2)
C10-C15	1.403(2)	C10-H10	0.95
C11-C12	1.382(2)	C11-H11	0.95
C12-C13	1.392(2)	C12-C12	1.7470(16)
C13-C14	1.388(2)	C13-H13	0.95
C14-C15	1.400(2)	C14-H14	0.95
C16-H16A	0.98	C16-H16B	0.98
C16-H16C	0.98		

Table 5. Bond angles (°) for (S)-17.

C2-C1-C6	121.47(16)	C2-C1-H1	119.3
C6-C1-H1	119.3	C3-C2-C1	118.40(16)
C3-C2-H2	120.8	C1-C2-H2	120.8
C4-C3-C2	121.56(15)	C4-C3-C11	119.07(13)
C2-C3-C11	119.37(13)	C3-C4-C5	118.69(16)
C3-C4-H4	120.7	C5-C4-H4	120.7

C6-C5-C4	121.69(16)	C6-C5-H5	119.2
C4-C5-H5	119.2	C5-C6-C1	118.18(15)
C5-C6-C7	120.21(14)	C1-C6-C7	121.60(14)
C8-C7-C6	111.17(13)	C8-C7-C16	110.51(15)
C6-C7-C16	111.72(14)	C8-C7-H7	107.8
C6-C7-H7	107.8	C16-C7-H7	107.8
C9-C8-C7	123.52(15)	C9-C8-H8	118.2
C7-C8-H8	118.2	C8-C9-C15	127.21(16)
C8-C9-H9	116.4	C15-C9-H9	116.4
C11-C10-C15	121.17(15)	C11-C10-H10	119.4
C15-C10-H10	119.4	C12-C11-C10	119.25(15)
C12-C11-H11	120.4	C10-C11-H11	120.4
C11-C12-C13	121.22(15)	C11-C12-Cl2	119.12(13)
C13-C12-Cl2	119.64(13)	C14-C13-C12	118.87(15)
C14-C13-H13	120.6	C12-C13-H13	120.6
C13-C14-C15	121.53(14)	C13-C14-H14	119.2
C15-C14-H14	119.2	C14-C15-C10	117.94(15)
C14-C15-C9	123.30(14)	C10-C15-C9	118.76(15)
C7-C16-H16A	109.5	C7-C16-H16B	109.5
H16A-C16-H16B	109.5	C7-C16-H16C	109.5
H16A-C16-H16C	109.5	H16B-C16-H16C	109.5

Table 6. Torsion angles (°) for (S)-17.

C6-C1-C2-C3	0.5(3)	C1-C2-C3-C4	0.0(3)
C1-C2-C3-Cl1	-179.32(14)	C2-C3-C4-C5	-0.3(3)
Cl1-C3-C4-C5	179.03(14)	C3-C4-C5-C6	0.1(3)
C4-C5-C6-C1	0.4(3)	C4-C5-C6-C7	-179.60(16)
C2-C1-C6-C5	-0.7(3)	C2-C1-C6-C7	179.30(16)
C5-C6-C7-C8	-107.64(17)	C1-C6-C7-C8	72.4(2)
C5-C6-C7-C16	128.40(18)	C1-C6-C7-C16	-51.6(2)
C6-C7-C8-C9	125.84(17)	C16-C7-C8-C9	-109.5(2)
C7-C8-C9-C15	179.54(16)	C15-C10-C11-C12	-1.1(2)
C10-C11-C12-C13	0.4(2)	C10-C11-C12-Cl2	-178.22(12)
C11-C12-C13-C14	0.4(2)	Cl2-C12-C13-C14	179.00(12)
C12-C13-C14-C15	-0.5(2)	C13-C14-C15-C10	-0.2(2)
C13-C14-C15-C9	-179.49(15)	C11-C10-C15-C14	1.1(2)
C11-C10-C15-C9	-179.66(15)	C8-C9-C15-C14	-17.4(3)

Table 7. Anisotropic atomic displacement parameters (\AA^2) for (S)-17.

The anisotropic atomic displacement factor exponent takes the form: $-2\pi^2 [h^2 a^{*2} U_{11} + \dots + 2 h k a^* b^* U_{12}]$

	U_{11}	U_{22}	U_{33}	U_{23}	U_{13}	U_{12}
C1	0.0362(8)	0.0336(8)	0.0350(8)	-0.0031(7)	0.0045(7)	0.0081(7)
C2	0.0397(9)	0.0326(9)	0.0346(8)	0.0025(7)	-0.0018(7)	0.0068(7)
C3	0.0408(8)	0.0257(7)	0.0309(8)	-0.0027(6)	0.0024(6)	-0.0039(7)
C4	0.0343(9)	0.0362(9)	0.0416(9)	-0.0057(7)	0.0050(7)	0.0053(7)
C5	0.0314(8)	0.0356(8)	0.0398(9)	-0.0008(7)	-0.0042(7)	0.0034(7)
C6	0.0323(7)	0.0252(7)	0.0309(7)	-0.0027(6)	-0.0032(6)	-0.0029(7)
C7	0.0397(8)	0.0326(8)	0.0301(8)	-0.0011(6)	-0.0027(6)	0.0026(8)
C8	0.0388(9)	0.0335(8)	0.0295(7)	-0.0006(6)	-0.0013(6)	0.0017(7)
C9	0.0370(8)	0.0318(8)	0.0274(7)	0.0020(6)	0.0005(6)	0.0007(7)
C10	0.0315(8)	0.0368(8)	0.0293(7)	0.0003(6)	-0.0016(6)	0.0065(7)
C11	0.0302(8)	0.0443(9)	0.0274(7)	0.0011(6)	-0.0014(6)	-0.0006(7)
C12	0.0382(9)	0.0322(8)	0.0237(7)	0.0011(6)	0.0016(6)	-0.0054(7)
C13	0.0373(9)	0.0369(9)	0.0299(8)	0.0035(7)	-0.0016(7)	0.0051(7)
C14	0.0305(8)	0.0412(8)	0.0276(7)	0.0004(6)	-0.0038(6)	0.0014(7)
C15	0.0317(8)	0.0349(8)	0.0213(7)	0.0017(6)	0.0025(6)	-0.0014(7)
C16	0.0550(12)	0.0561(12)	0.0367(9)	-0.0018(9)	0.0067(9)	-0.0162(10)
C11	0.0776(3)	0.0446(2)	0.0298(2)	-0.00252(16)	0.0078(2)	0.0012(2)
C12	0.0649(3)	0.0365(2)	0.0412(2)	-0.00480(16)	-0.0075(2)	-0.0067(2)

Table 8. Hydrogen atomic coordinates and isotropic atomic displacement

parameters (\AA^2) for (S)-17.

	x/a	y/b	z/c	U(eq)
H1	0.5751	0.1094	0.2031	0.042
H2	0.5431	0.1137	0.2752	0.043
H4	-0.0589	0.3960	0.2652	0.045
H5	-0.0208	0.3917	0.1933	0.043
H7	0.1834	0.3358	0.1326	0.041
H8	0.3303	-0.0233	0.1361	0.041
H9	0.0031	0.1476	0.0872	0.038
H10	-0.2892	-0.0307	0.0534	0.039
H11	-0.4073	-0.2917	0.0228	0.041
H13	0.1941	-0.5228	0.0623	0.042
H14	0.3068	-0.2634	0.0942	0.04
H16A	0.6657	0.2302	0.1390	0.074
H16B	0.5790	0.4281	0.1409	0.074
H16C	0.5499	0.3187	0.0992	0.074