## **Extended Data Table 1.** Primers used in Real-Time reverse transcriptase PCR

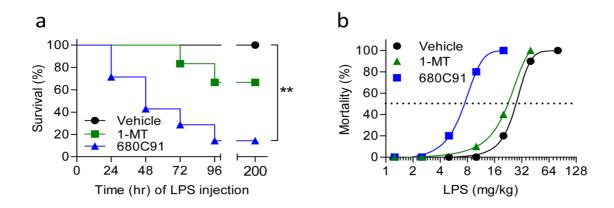
GENE	FORWARD SEQUENCE	REVERSE SEQUENCE	
Gapdh	5'- GCC TTC CGT GTT CCT ACC C -3'	5'- CAG TGG GCC CTC AGA TGC -3'	
Ahr	5'- CCA CTG ACG GAT GAA GAA GGA -3'	5'- ATC TCG TAC AAC ACA GCC TCT C -3'	
Cyp1a1	5'- GAC ACA GTG ATT GGC AGA G -3'	5'- GAA GGT CTC CAG AAT GAA GG -3'	
Ido1	5'- TCT GCC TGT GCT GAT TGA -3'	5'- CTG TAA CCT GTG TCC TCT CA -3'	
Ido2	5'- CTC AGA CTT CCT CAC TTA ATC G -3'	5'- GCT GCT CAC GGT AAC TCT -3'	
Tdo2	5'- GTG AAC GAC GAC TGT CAT ACC G -3'	5'- GCT GGA AAG GGA CCT GGA AT -3'	
Foxp3	5' - CCC AGG AAA GAC AGC AAC CTT TT -3'	5' - TTC TCA CAA CCA GGC CAC TTG - 3'	
Rorc	5' - ACA ACA GCA GCA AGT GAT GG - 3'	5' - CCT GGA TTT ATC CCT GCT GA - 3'	
Tgfb1	5' - CAC AGA GAA GAA CTG CTG TG- 3'	5' - AGG AGC GCA CAA TCA TGT TG- 3'	
Il10	5' - ACC AGC TGG ACA ACA TAC TG- 3'	5' - CGC ATC CTG AGG GTC TTC AG- 3'	

**Extended Data Table 2.** Complete list of datasets used to assess gene expression levels of tyrosine kinases in mouse myeloid dendritic cells.

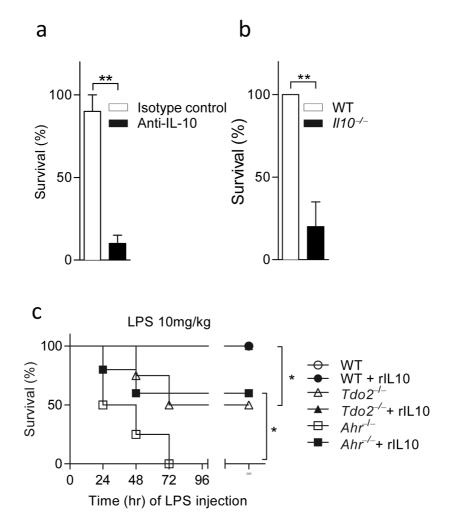
GEO accession	Platform	Total samples in series	# of un- stimulated samples	# of LPS stimulated samples	GEO accession of used samples	Reference
GSE28231	MOE 430 2.0	5	1	1	GSM698691 GSM698692	1
GSE28340	MOE 430 2.0	8	2	2	GSM700775 GSM700776 GSM700777 GSM700778	2
GSE18115	MOE 430 2.0	8	1	2	GSM452757 GSM452759 GSM452761	3
GSE23641	MOE 430 2.0	2	0	1	GSM579979	4
GSE16761	MOE 430 2.0	3	1	1	GSM420166 GSM420165	5
GSE10246	MOE 430 2.0	182	2	0	GSM258647 GSM258648	6
GSE7219	MOE 430 2.0	12	3	0	GSM173580 GSM173581 GSM173582	7

#### References

- 1. Pletinckx, K., *et al.* Similar inflammatory DC maturation signatures induced by TNF or *Trypanosoma brucei* antigens instruct default Th2-cell responses. *Eur. J. Immunol.* **41**, 3479-3494 (2011).
- 2. Sun, Y., et al. Targeting of microRNA-142-3p in dendritic cells regulates endotoxin-induced mortality. *Blood* **117**, 6172-6183 (2011).
- 3. Clavarino, G., *et al.* Protein phosphatase 1 subunit Ppp1r15a/GADD34 regulates cytokine production in polyinosinic:polycytidylic acid-stimulated dendritic cells. *Proc. Natl. Acad. Sci. U S A* **109**, 3006-3011 (2012).
- 4. O'Connell, R.M., *et al.* MicroRNA-155 promotes autoimmune inflammation by enhancing inflammatory T cell development. *Immunity* **33**, 607-619 (2010).
- 5. Koga, K., *et al.* Cyclic adenosine monophosphate suppresses the transcription of proinflammatory cytokines via the phosphorylated c-Fos protein. *Immunity* **30**, 372-383 (2009).
- 6. Lattin, J.E., *et al.* Expression analysis of G Protein-Coupled Receptors in mouse macrophages. *Immunome Res.* **4**, 5 (2008).
- 7. Lind, E.F., *et al.* Dendritic cells require the NF-κB2 pathway for cross-presentation of soluble antigens. *J. Immunol.* **181**, 354-363 (2008).

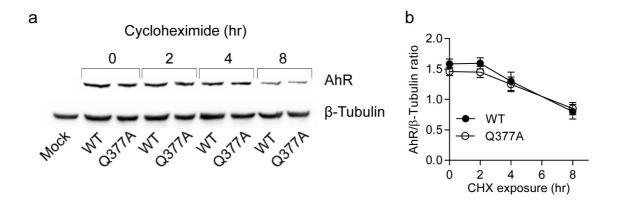


Extended Data Fig. 1. Increased susceptibility to primary LPS challenge in mice treated with a TDO2 inhibitor. (a) Twelve hours before LPS challenge (10 mg/kg), WT mice were treated with vehicle, the IDO1 and IDO2 inhibitor 1-MT (200 mg/kg), or the TDO2 inhibitor 680C91 (10 mg/kg), control groups receiving 1-MT or 680C91 but no LPS. Survival was monitored every 24 h through day 8 of LPS challenge (n = 10 mice per group per experiment, in one out of three). \*\*P < 0.001 (log-rank test). (b) Estimation of LD<sub>50</sub> (mg/kg) in mice treated with 1.25, 2.5, 5, 10, 20, 40, or 80 mg/kg LPS. n = 10 per group per dose. LD<sub>50</sub> values were calculated by curve-fitting ( $r^2 \ge 0.95$ ) in one experiment representative of two.



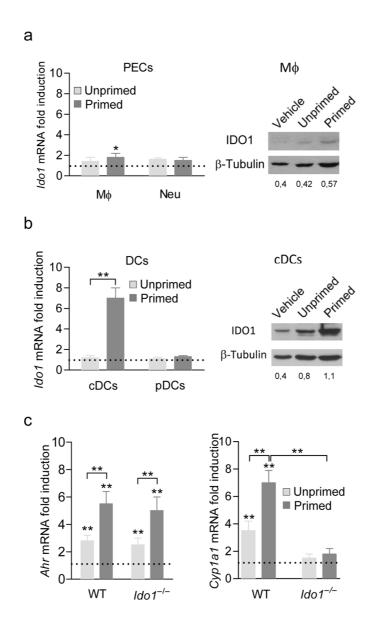
#### Extended Data Fig. 2. Lack of endogenous IL-10 increases susceptibility to

**endotoxemia**. (a) Survival of WT mice exposed to 10 mg/kg LPS in the presence of anti–IL-10 (0.2 mg/mouse daily, for 4 d, commencing 6 h before challenge) or an isotype control. Data are from three independent experiments (mean  $\pm$  s.d.). (b) Survival of WT and  $II10^{-/-}$  mice treated with 10 mg/kg LPS. \*\*P < 0.001 (log-rank test). (c) Survival curves of mice of different genotypes challenged with 10 mg/kg LPS, with or without therapeutic subcutaneous IL-10 at 250 ng/mouse, daily, from challenge (day 0) through day 5. \*P < 0.05 (IL-10 vs. vehicle). The data show that exogenous IL-10 compensates for both the TDO2 and AhR defects at the lower LPS dosage. IL-10 is protective only in TDO2 knockouts when 20 mg/kg LPS is used.

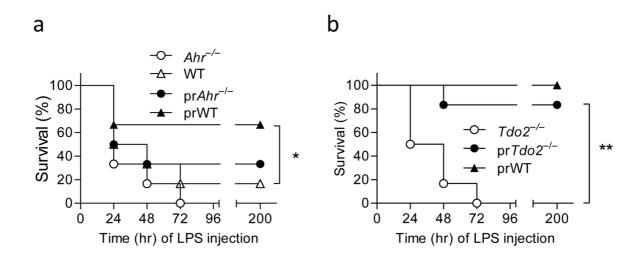


# Extended Data Fig. 3. Mutation of Gln377 to Ala in AhR PAS-B domain does not alter receptor half-life, and apparently results in increased TCDD ligand potency.

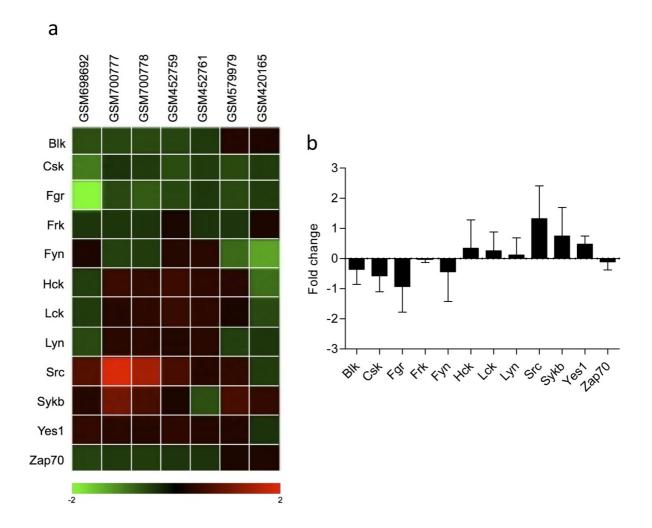
(a) AhR-deficient cDCs were transfected with WT or Q377A AhR. After 24 h, cells were incubated with cycloheximide (CXM) (10  $\mu$ g/ml) and harvested at different times, lysed, and analyzed for AhR expression by immunoblotting, using a specific antibody.  $\beta$ -tubulin was used as a loading control. Data are from one experiment of three. (b) Ratios (means  $\pm$  s.d. of three experiments) of WT or Q377A AhR to  $\beta$ -tubulin in transfected cDCs at different times of CHX treatment . (No differences by Student's t-test.)



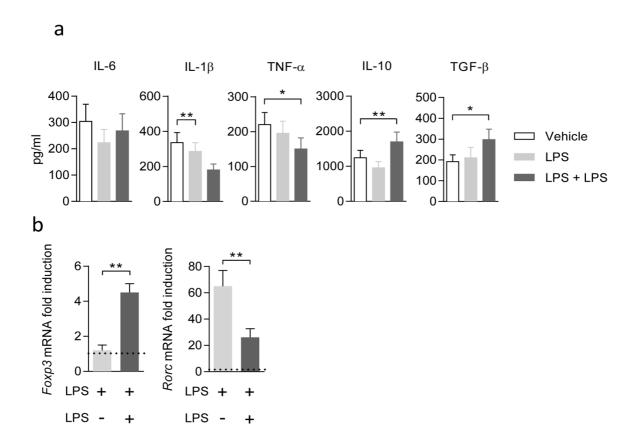
Extended Data Fig. 4. LPS tolerance potentiates IDO1 expression and AhR activation in splenic cDCs. (a) Real-time PCR analysis of *Ido1* mRNA expression and immunoblot analysis of IDO1 protein in peritoneal exudate macrophages (M $\Phi$ ) and neutrophils (Neu) (a), as well as in splenic conventional DCs (cDCs) or plasmacytoid DCs (pDCs) (b). Cells were harvested and purified at 24 (a) or 72 (b) h of LPS rechallenge. For comparison, samples were included from mice on first exposure to 40 mg/kg LPS (unprimed), as opposed to tolerized mice (primed). Data of *Ido1* mRNA fold induction are presented as in **Fig. 2a** (mean  $\pm$  s.d. of three experiments; \*P < 0.05 and \*\*P < 0.001, Shapiro test). Immunoblotting data are from one experiment of three. (c) Real-time PCR analysis of *Ahr* and *Cyp1a1* transcript expressions in cDCs from the same mice as in b. \*\*P < 0.001, Shapiro test.



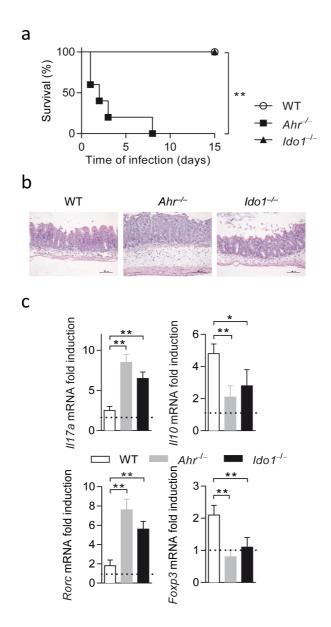
**Extended Data Fig. 5. Absolute requirement for functional AhR, but not TDO2, in LPS tolerance manifestations.** (a) Survival curves of WT and LPS-primed (0.5 mg/kg, day 0) WT (prWT) and AhR-deficient (pr $Ahr^{-}/^{-}$ ) mice after a second challenge (on day +7) with 40 mg/kg LPS. Survival was monitored every 24 h through day 8 of LPS challenge. n = 8-10 mice per group per experiment. One experiment of three. \*P < 0.05, log-rank test. (b) Survival curves of WT and LPS-primed (10 mg/kg, day 0) WT (prWT) and TDO2-deficient (pr $Tdo2^{-}/^{-}$ ) mice after a second challenge (on day +7) with 40 mg/kg LPS. Survival was monitored every 24 h through day 8 of LPS challenge. n = 8-10 mice per group per experiment. One experiment of three. \*\*P < 0.001, log-rank test.



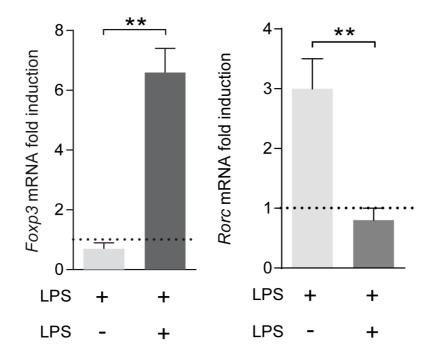
Extended Data Fig. 6. Bioinformatic data from myeloid cDCs data sets, as follows, (a) expression changes of tyrosine kinases in LPS-primed myeloid DCs as compared to untreated counterparts, and (b)  $\log_2$  fold changes, depicted as mean values and standard errors.



Extended Data Fig. 7. LPS tolerance modulates cytokine production and *Foxp3* and *Rorc* transcription in *S.* Typhimurium infection. (a) IL-6, IL-1 $\beta$ , TNF- $\alpha$ , IL-10, and TGF- $\beta$  were measured in cecum cell supernatants from LPS-tolerant mice infected with *S. enterica* Typhimurium. Data are from three independent experiments (means  $\pm$  s.d.). \*P < 0.05 and \*\*P < 0.001 (Student's *t*-test). (b) RT-PCR expression of *Foxp3* and *Rorc* transcripts in mesenteric lymph node cells from LPS-tolerant, *Salmonella*-infected mice. Data (mean  $\pm$  s.d. of three experiments) are presented as normalized transcript expression in the samples relative to normalized transcript expression in control cultures (that is, cells from vehicle-treated mice, in which fold change = 1; dotted line). \*P < 0.05; \*\*P < 0.001 (Shapiro test).

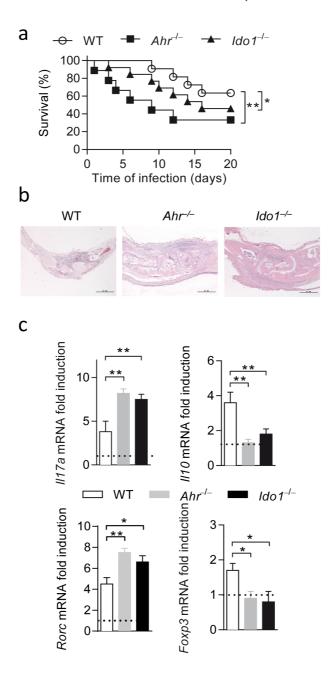


Extended Data Fig. 8. *Ahr*-/- and *Ido*-/- mice are more susceptible than WT mice to *S.* Typhimurium infection. (a) Naïve mice of different genotypes were challenged intragastrically with *S.* Typhimurium. Mortality data were recorded (\*\*P < 0.001, WT vs. all other genotypes; log-rank test) and (b) H&E staining of mouse ceca was performed at 7 days of infection. Scale bars, 50  $\mu$ m. One of three experiments. (c) Transcript expressions of *Il17a*, *Rorc*, *Il10*, and *Foxp3* were quantified in mesenteric lymph node cells. Data (mean  $\pm$  s.d. of three experiments) are presented as normalized transcript expression in the samples relative to normalized transcript expression in cells from uninfected donors, in which fold change = 1. \*\*P < 0.001 (Shapiro test).



### Extended Data Fig. 9. LPS tolerance modulates Foxp3 and Rorc transcription in

**GBS infection**. RT-PCR expression of *Foxp3* and *Rorc* transcripts in joint-draining lymph node cells from LPS-tolerant, GBS-infected mice. Data (mean  $\pm$  s.d. of three experiments) are presented as normalized transcript expression in the samples relative to normalized transcript expression in control cells (that is, cells from vehicle-treated mice, in which fold change = 1; dotted line). \*\*P < 0.001 (Shapiro test).



Extended Data Fig. 10. *Ahr*<sup>-/-</sup> and *Ido*<sup>-/-</sup> mice are more susceptible than WT mice to GBS immunopathology. (a) Naïve mice of different genotypes were infected with GBS ( $1 \times 10^7$  CFU). Mortality data were recorded (\*P < 0.05 and \*\*P < 0.001, WT vs. all other genotypes; log-rank test). (b) H&E staining of joints was performed at 10 days of infection. Scale bars,  $100 \mu m$ . One of three experiments. (c) Transcript expressions of *Il17a*, *Rorc*, *Il10*, and *Foxp3* were quantified in joint-draining lymph nodes. Data (mean  $\pm$  s.d. of three experiments) are presented as normalized transcript expression in the samples relative to normalized transcript expression in cells from uninfected donors, in which fold change = 1.\*\*P < 0.001 (Shapiro test).