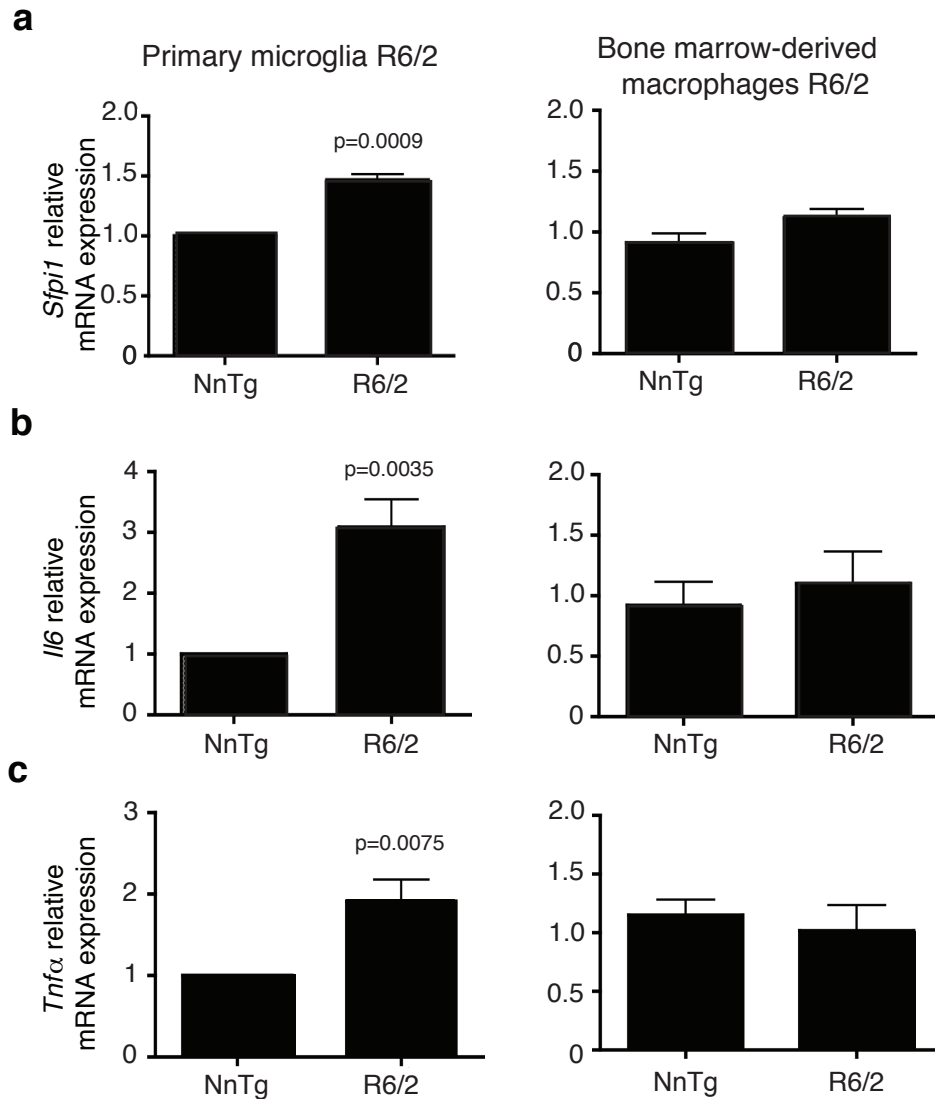
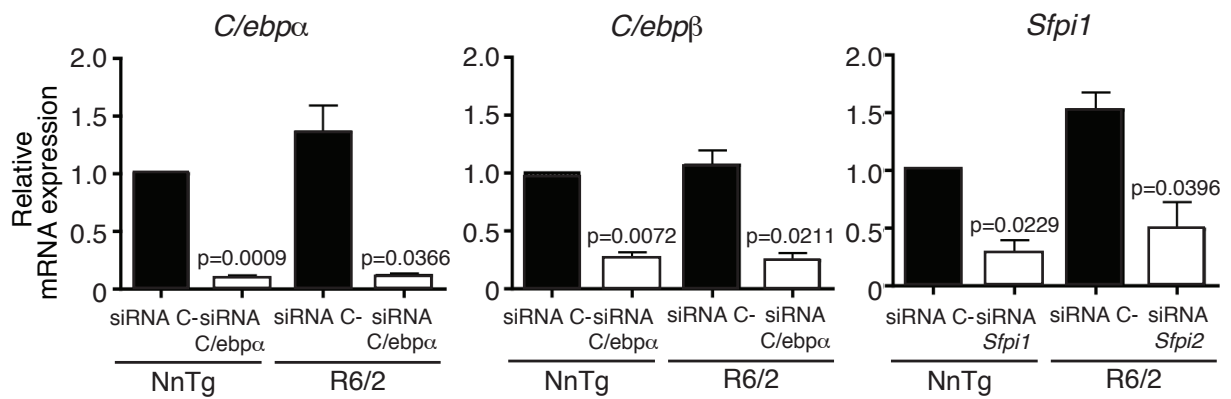


Mutant Huntingtin promotes autonomous microglia activation via myeloid lineage-determining factors

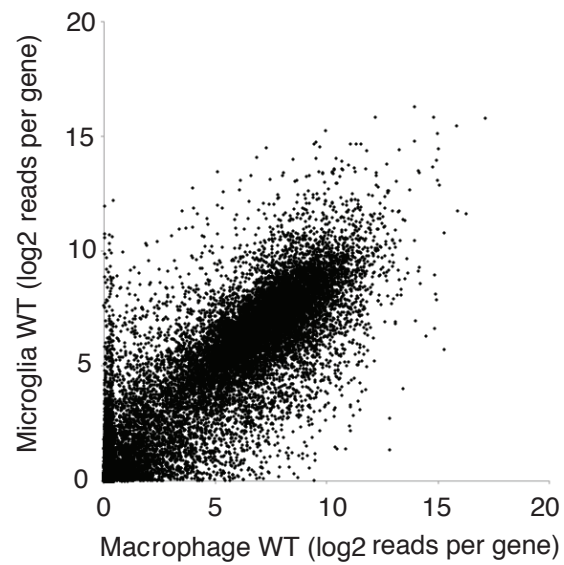
Crotti A., Benner C., Kerman B., Gosselin D., Lagier-Tourenne C., Zuccato C., Cattaneo E., Gage F.H., Cleveland D.W., Glass C.K.



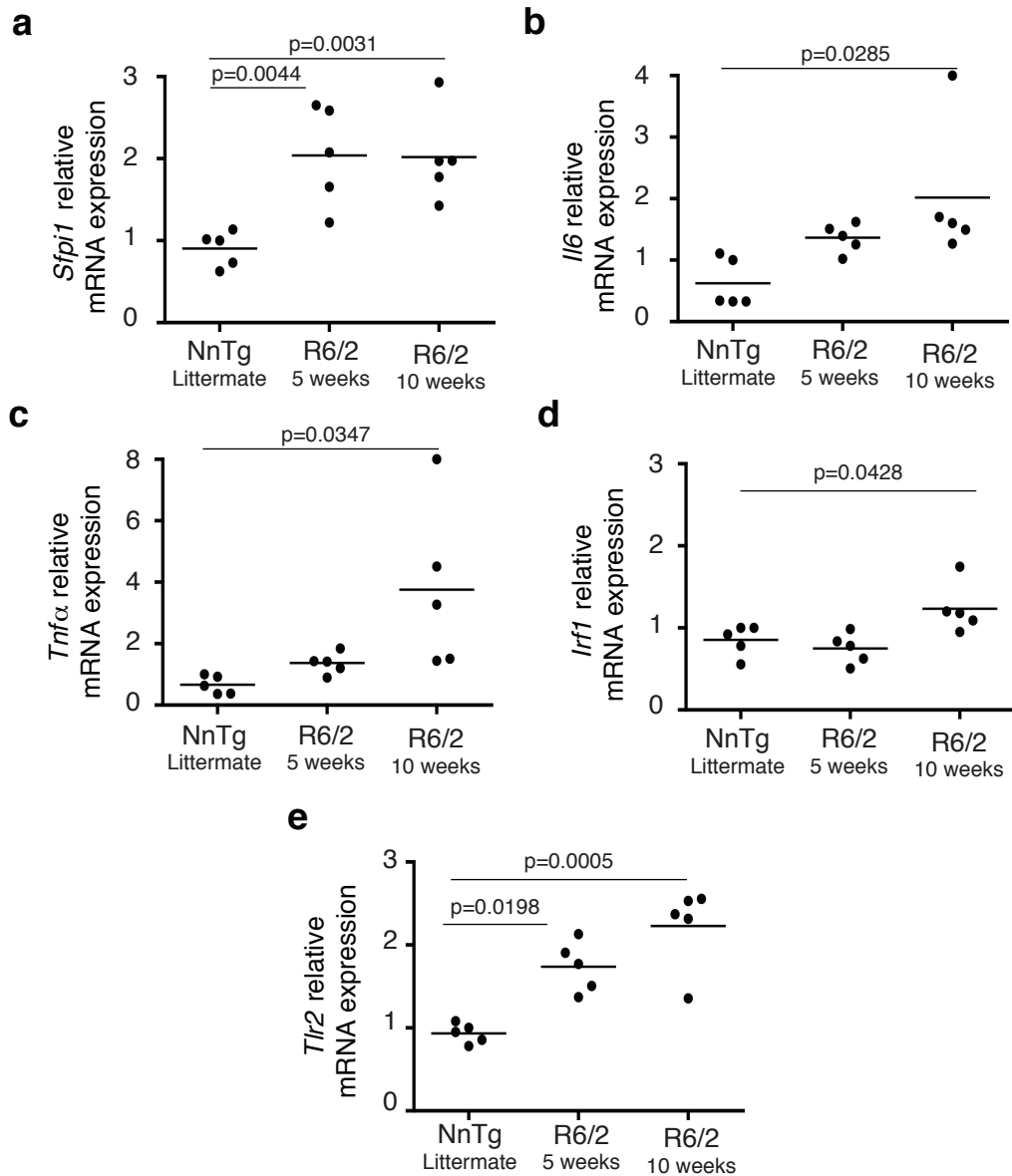
Supplementary Figure 1: PU.1 and PU.1-C/EBPs target genes are up-regulated in primary microglia but not in BMDM from R6/2 mice. qRT-PCR analysis for *Sfp1* (a), *Il6* (b) and *Tnfα* (c) mRNAs expression in primary microglia (mean±sd, n= 6 biological replicates, two-tailed paired student't test) and bone marrow derived macrophages (mean±sd, n= 5 biological replicates, two-tailed paired student't test) purified from non-transgenic littermates and R6/2 mice.



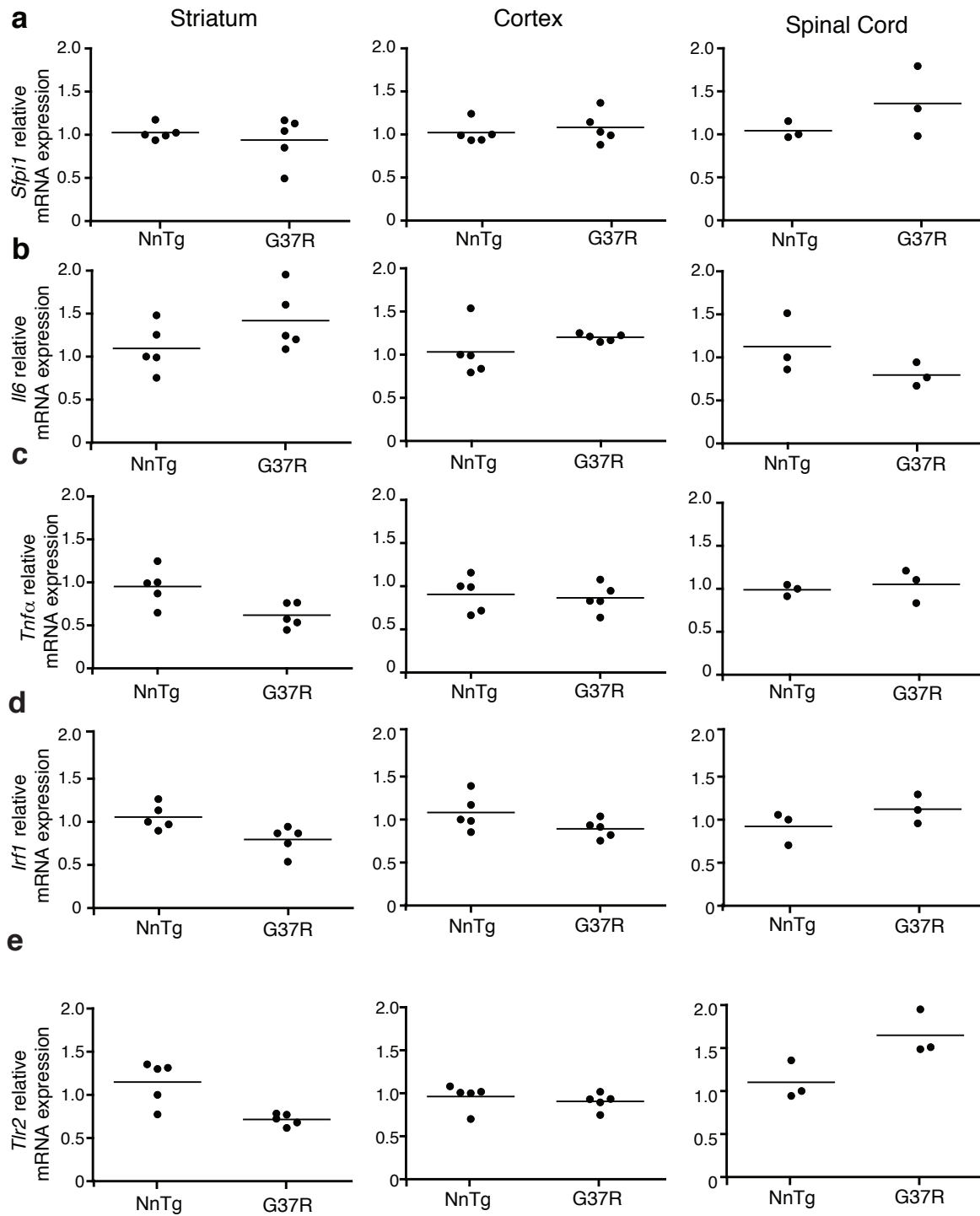
Supplementary Figure 2: Effect of siRNA knockdown of PU.1, C/EBP α and C/EBP β . Efficiency of siRNA knockdown in primary microglia derived from R6/2 mice and nontransgenic littermates as determined by qRT-PCR (mean \pm sd, n= 3 biological replicates, p values determined by two-tailed paired student t-test).



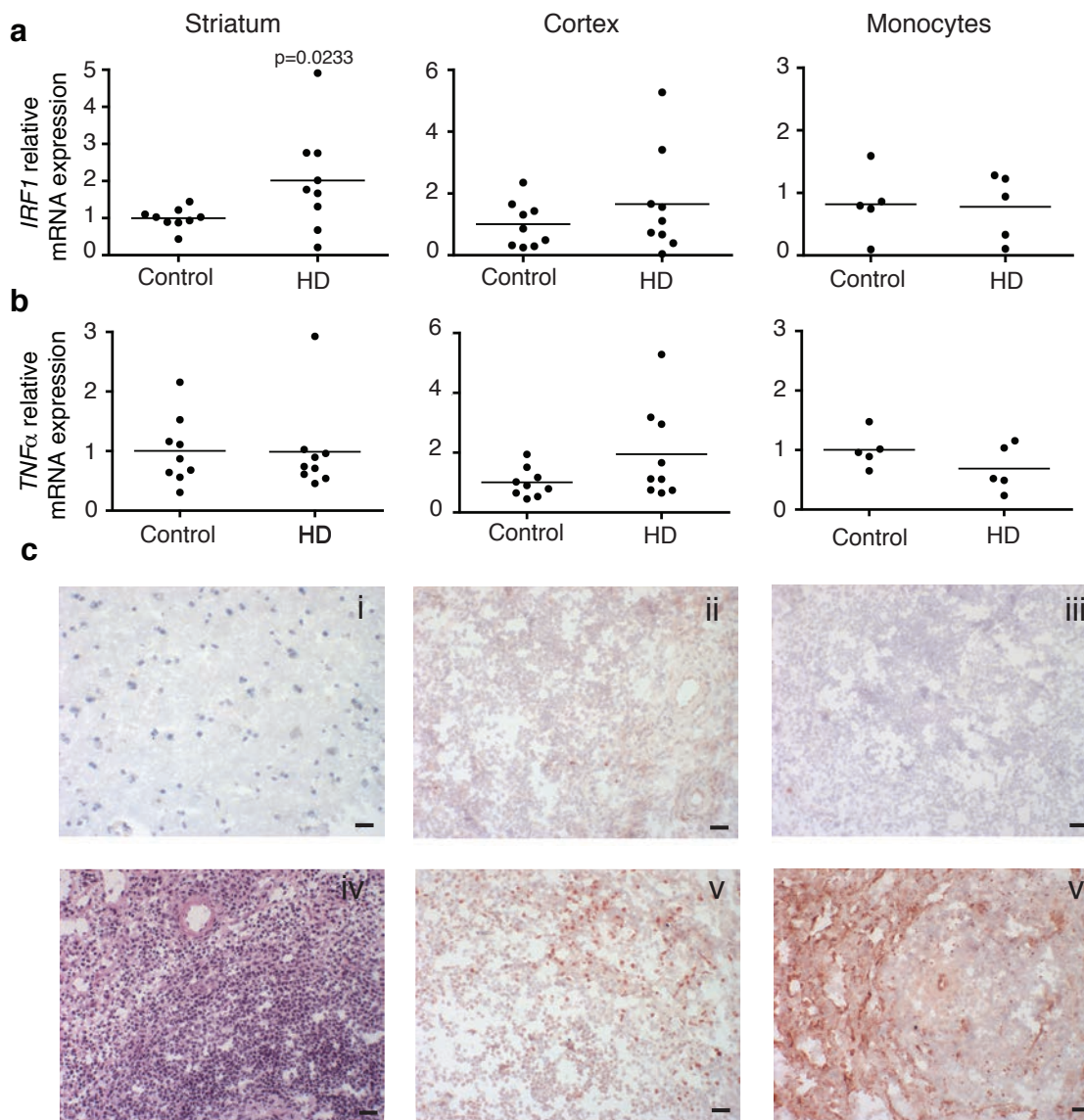
Supplementary Figure 3: Differential gene expression between Microglia WT and Macrophages WT. Scatter Plot representing the differential gene expression observed in microglia from wild-type *Hdh*^{7/7} versus BMDM from wild-type *Hdh*^{7/7} mice.



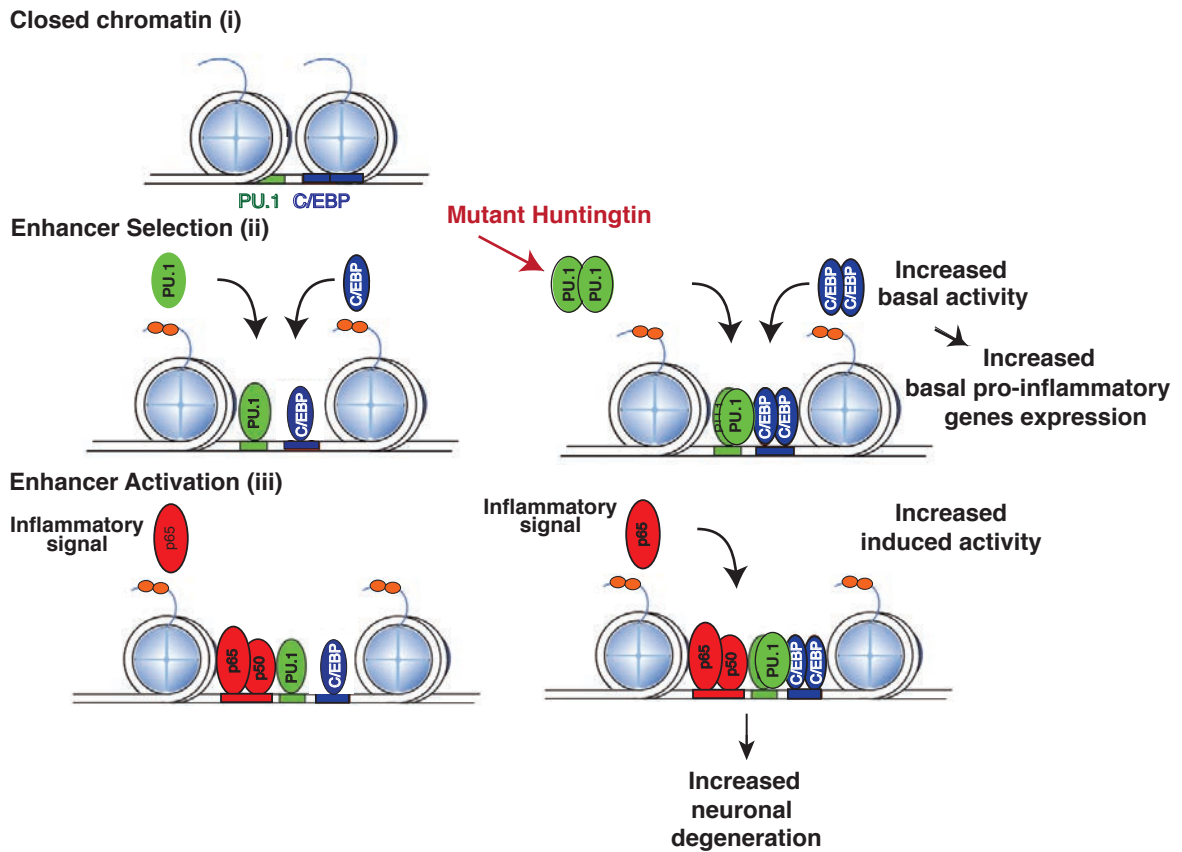
Supplementary Figure 4: PU.1 and PU.1-C/EBPs target genes are up-regulated in the striatum from R6/2 mice. qRT-PCR analysis for *Sfp1* (a), *Il6* (b), *Tnfα* (c), *Irf1* (d) and *Tlr2* (e) mRNAs expression in striatum from nontransgenic littermates, pre-symptomatic (5 weeks-old) and symptomatic (10 weeks-old) R6/2 mice. Each dot is representative of one mouse (unpaired student's test).



Supplementary Figure 5: PU.1 and PU.1-C/EBPs target genes are not differentially expressed in SOD1^{G37R} mouse model of ALS. qRT-PCR analysis for *Sfp1* (a), *Il6* (b), *Tnfα* (c), *Irf1* (d) and *Tlr2* (e) mRNAs expression in striatum, cortex and spinal cord from nontransgenic littermates and SOD1^{G37R} mice (8-12 months old). Each dot is representative of one mouse (unpaired student's test).



Supplementary Figure 6: Inflammation *in vivo* in HD individuals. qRT-PCR analysis for *IRF1* (a) and *TNFα* (b) mRNAs expression in striatum (first column, n= 9 individual per group), cortex (second column, n= 9 individual per group) and monocytes (third column, n= 5 individual per group) from controls and HD individuals. Each dot is representative of one individual. All p values were determined by unpaired student t-test. (c) IHC controls: brain section in presence of rabbit IgG (negative control) (i); spleen section in presence of rabbit IgG (negative control) (ii); spleen section in presence of diluting buffer (BSA, 1% bovine serum albumin in PBS phosphate buffered saline) (iii); H&E staining on spleen section (iv); PU.1 IHC staining on spleen section (positive control) (v); Von Willebrand factor IHC staining on spleen section (positive control for endothelial cells/blood vessels) (vi). Scale bar: 100μm.



Supplementary Figure 7: A model for mechanisms by which mutant Huntingtin influence the selection and activation of microglia enhancers. Left: PU.1 and C/EBPs function in a collaborative manner to select microglia enhancers from inactive chromatin in basal conditions. Pro-inflammatory signals that activate transcription factors such as the p65 component of NF κ B lead to inflammatory response. Right: mutant Huntingtin expression enhances this process by increasing PU.1 expression and PU.1-C/EBPs promoter binding, leading to increased enhancer activity under basal conditions that results in increased basal pro-inflammatory and neurotoxic genes expression. This phenomenon increases the sensitivity to pro-inflammatory signals. In fact, under conditions of sterile inflammation mutant Huntingtin-expressing microglia appears to be more efficient in inducing neuronal death.

Figure 1e

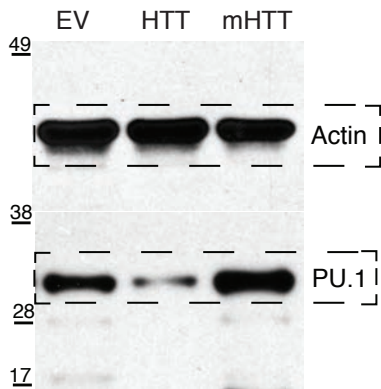


Figure 3c

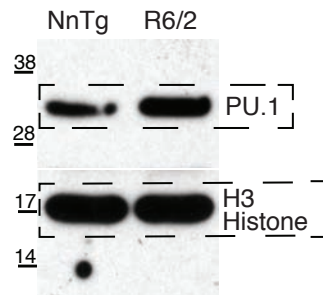


Figure 4a

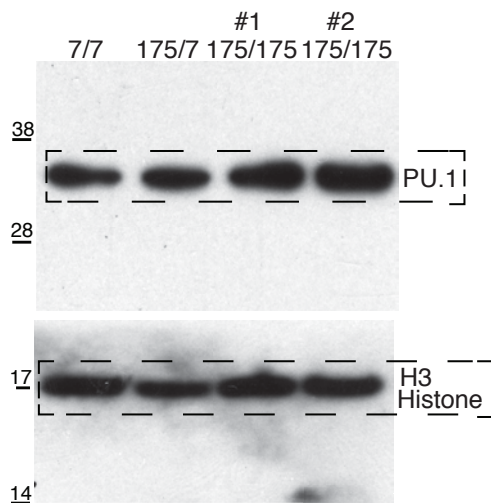
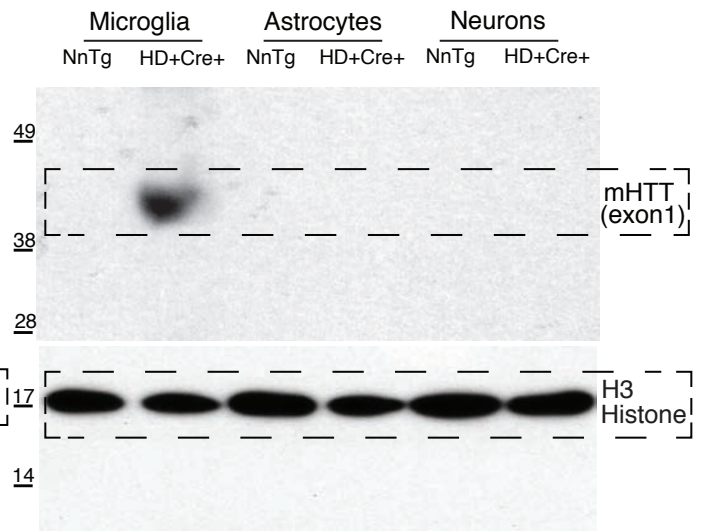


Figure 7b



Supplementary Figure 8: Full-length pictures of the blots presented in the main figures. To examine proteins of interest on the same samples, blots were cut first and then probed with indicated antibodies.