Supplementary Figure Legends

Supplementary Figure 1. The loss of Gas1 enhances rDNA silencing and rDNA stability in the presence of Sir2. (A) The absence of Gas1 increases rDNA silencing. Silencing within rDNA was assessed by monitoring the growth of cells (10-fold serial dilutions) plated on SC medium without uracil. SC medium was used as a plating control. (B) The loss of Gas1 promotes transcriptional silencing of the mURA3 reporter gene at the rDNA locus in a Sir2dependent manner. Total RNA was extracted from wild-type (WT), $gas1\Delta$, $sir2\Delta$, and $gas1\Delta$ $sir2\Delta$ cells. Quantitative real-time reverse transcription-PCR analysis was performed to measure the transcript levels of the mURA3 reporter gene inserted inside (RDN1-NTS1::mURA3) or outside the rDNA array (*leu2::mURA3*). Amplification efficiencies were validated and normalized against ACT1. Relative mURA3 transcript levels were calculated as the ratio of the normalized transcript level of the *mURA3* reporter gene inside the NTS1 region to that outside the rDNA array. The values were the mean of three independent experiments and error bar indicates standard deviations. Asterisks indicate P < 0.05, compared with WT cells (two-tailed Student's t-test). (C) The loss of Gas1 represses rDNA recombination in a Sir2-dependent manner. rDNA recombination is represented by the rate of loss of the ADE2 marker gene integrated at the rDNA locus in WT, $gas1\Delta$, $sir2\Delta$, and $gas1\Delta$ $sir2\Delta$ cells. Values represent the average of three independent experiments, and error bars indicate the standard deviation. Mean values of the recombination rates for WT, $gas1\Delta$, $sir2\Delta$, and $gas1\Delta$ sir2 Δ cells are 1.53×10^{-3} , 0.57×10^{-3} , 4.68×10^{-3} , and 4.84×10^{-3} , respectively. Asterisks indicate *P*<0.05, compared with WT cells (two-tailed Student's *t*-test).

Supplementary Figure S2. The protein level of Sir2 is not changed in the absence of Gas1 or Msn2/4. Total protein was extracted from wild-type (WT), $gas1\Delta$, $msn2\Delta$ $msn4\Delta$, and $gas1\Delta$ $msn2\Delta$ $msn4\Delta$ cells, and immunoblotting was performed using an HRP-conjugated anti-mouse IgG antibody for the detection of TAP-tagged protein. Actin was used as a loading control. The relative ratio of Sir2 to actin, normalized against that of WT cells, is shown below each lane. Data are representative of at least three independent experiments.

Supplementary Figure S3. The protein level of Gas1 is not changed in the lack of Gas1 β -1,3-glucanosyltransferase activity. Total protein was extracted from WT and *gas1* Δ cells containing an empty vector and *gas1* Δ cells expressing WT *GAS1* and *gas1*^{*E161Q, E262Q*} on the pRS413 vector, and immunoblotting was performed using a mouse anti-GFP antibody for the detection of GFP-tagged protein. Hexokinase was used as a loading control. The relative ratio of Gas1 to hexokinase, normalized against that of WT cells, is shown below each lane. Data are representative of at least three independent experiments.

Supplementary Figure S4. Congo red treatment promotes Sir2-mediated rDNA silencing. (A) The association of Msn2/4 with the PNC1 promoter region is enhanced under Congo red treatment. The degree of association of Msn2-TAP (left panel) and Msn4-TAP (right panel) with the PNC1 promoter region was measured using a ChIP assay with or without treatment with 100 µg/ml Congo red for 1 h. Cells without Congo red treatment were used as a control. Values represent the average of three independent experiments, and error bars indicate the standard deviation. Asterisks indicate P < 0.05, compared with untreated control cells (twotailed Student's t-test). (B) The protein level of Pnc1 increases under Congo red treatment. Total protein was extracted from cells with or without treatment with 100 µg/ml Congo red for 1 h, and immunoblotting was performed using an HRP-conjugated anti-mouse IgG antibody for the detection of TAP-tagged protein. Actin was used as a loading control. The relative ratio of Pnc1 to actin, normalized against that of untreated control cells, is shown below each lane. Data are representative of at least three independent experiments. (C) The association of Sir2 with rDNA is enhanced under Congo red treatment. The degree of Sir2 binding to four representative regions in the rDNA locus (25S, NTS1, NTS2/18S, and 18S regions) was measured using a ChIP assay with or without treatment with 100 µg/ml Congo red for 1 h. Values represent the average of three independent experiments, and error bars indicate the standard deviation. Asterisks indicate P < 0.05, compared with untreated control cells (two-tailed Student's t-test). (D) Congo red increases rDNA silencing in the presence of Sir2. Silencing within the rDNA region was assessed by monitoring the growth of cells (10fold serial dilutions) plated on SC medium without uracil in the presence or absence of 100 µg/ml Congo red. SC medium was used as a plating control. (E) Congo red promotes transcriptional silencing of the *mURA3* reporter gene at the rDNA locus in a Sir2-dependent manner. Total RNA was extracted from WT and $sir2\Delta$ cells with or without treatment with 100 µg/ml Congo red for 1 h. Quantitative real-time reverse transcription-PCR analysis was performed as in Supplementary Figure 1B. The values were the mean of three independent experiments and error bar indicates standard deviations. Asterisks indicate P < 0.05, compared with untreated WT cells (two-tailed Student's t-test). (F) Congo red suppresses rDNA

recombination in a Sir2-dependent manner. rDNA recombination is represented by the rate of loss of the *ADE2* marker gene integrated at the rDNA locus in WT and *sir2* Δ cells plated on SC medium with or without 100 µg/ml Congo red. Values represent the average of three independent experiments, and error bars indicate the standard deviation. Mean values of the recombination rates for WT (control), WT (Congo red), *sir2* Δ (control), and *sir2* Δ (Congo red) cells are 1.45×10^{-3} , 0.64×10^{-3} , 4.68×10^{-3} , and 4.21×10^{-3} , respectively. Asterisks indicate *P*<0.05, compared with untreated WT cells (two-tailed Student's *t*-test).

Supplementary Figure S5. Cell wall stress agents, such as calcofluor white, SDS, vanadate, and caffeine, do not affect rDNA silencing. (A) Calcofluor white (CFW) does not promote rDNA silencing. Silencing within the rDNA region was assessed by monitoring the growth of cells (10-fold serial dilutions) plated on SC medium without uracil in the presence or absence of 50 µg/ml CFW. SC medium was used as a plating control. (B) CFW does not promote transcriptional silencing of the mURA3 reporter gene at the rDNA locus. Total RNA was extracted from wild-type and $sir2\Delta$ cells with or without treatment with 50 µg/ml CFW for 1 h. Quantitative real-time reverse transcription-PCR analysis was performed to measure the transcript levels of the mURA3 reporter gene inserted inside (RDN1-NTS1::mURA3) or outside the rDNA array (*leu2::mURA3*). Values represent the average of three independent experiments, and error bars indicate the standard deviation. (C) CFW does not promote rDNA stability. rDNA recombination is represented by the rate of loss of the ADE2 marker gene integrated at the rDNA locus in wild-type and $sir2\Delta$ cells plated on SC medium with or without 50 µg/ml CFW. Values represent the average of three independent experiments, and error bars indicate the standard deviation. Mean values of the recombination rates for WT (control), WT (CFW), $sir2\Delta$ (control), and $sir2\Delta$ (CFW) cells are 1.53×10^{-3} , 1.39×10^{-3} , 4.68×10⁻³, and 4.62×10⁻³, respectively. (D) SDS, vanadate, and caffeine do not promote transcriptional silencing of the *mURA3* reporter gene at the rDNA locus. Total RNA was extracted from cells with or without treatment with 0.01% SDS, 5 mM vanadate, or 12 mM caffeine for 1 h. Quantitative real-time reverse transcription-PCR analysis was performed as described above. Values represent the average of three independent experiments, and error bars indicate the standard deviation.

Supplementary Figure S6. Gas1 paralogs are not involved in rDNA silencing. (A) The absence of Gas1 paralogs does not increase rDNA silencing. The spot assay was performed in

wild-type (WT), $gas1\Delta$, $gas2\Delta$, $gas3\Delta$, $gas4\Delta$, $gas5\Delta$, and $bgl2\Delta$ cells. Silencing within rDNA was assessed by monitoring the growth of cells (10-fold serial dilutions) plated on SC medium without uracil. SC medium was used as a plating control. (B) The absence of Gas1 paralogs does not significantly contribute to rDNA stability. rDNA recombination is represented by the rate of loss of the *ADE2* marker gene integrated at the rDNA locus in WT, $gas1\Delta$, $gas2\Delta$, $gas3\Delta$, $gas4\Delta$, $gas5\Delta$, and $bgl2\Delta$ cells. Values represent the average of three independent experiments, and error bars indicate the standard deviation. Mean values of the recombination rates for WT, $gas1\Delta$, $gas2\Delta$, $gas3\Delta$, $gas4\Delta$, $gas5\Delta$, $gas3\Delta$, $gas4\Delta$, $gas5\Delta$, and $bgl2\Delta$ cells are 1.48×10^{-3} , 0.40×10^{-3} , 2.07×10^{-3} , 1.28×10^{-3} , 1.01×10^{-3} , 1.05×10^{-3} , and 1.35×10^{-3} , respectively. Asterisks indicate P < 0.05, compared with WT cells (two-tailed Student's *t*-test).

Supplementary Figure S7. The lack of Gas1 β -1,3-glucanosyltransferase activity and the treatment of Congo red decrease the *in vivo* activity of PKA. (A) The absence of Gas1 β -1,3-glucanosyltransferase activity decreases the *in vivo* activity of PKA. Total protein was extracted from WT and *gas1* Δ cells harboring pRS423-pr^{CUP}-6×MYC-cki1^{2-200(S125/130A)} and containing an empty, WT *GAS1* and *gas1*^{E161Q, E262Q} on the pRS415 vector. Immunoblotting was performed using a mouse anti-Myc antibody. The relative ratio of phosphorylated (Cki1-P) to non-phosphorylated (Cki1) forms of Cki1, normalized against that of WT cells, is shown below each lane. Data are representative of at least three independent experiments. (B) The Congo red treatment decreases the *in vivo* activity of PKA. Total protein was extracted from the cells harboring pRS423-pr^{CUP}-6×MYC-cki1^{2-200(S125/130A)}, and immunoblotting was performed using a mouse anti-Myc antibody. The relative ratio of phosphorylated (Cki1-P) to non-phosphorylated (Cki1) forms of Cki1, normalized against that of untreated cells, is shown below each lane. Data are representative of at least three independent experiments. (B) The congo red treatment decreases the *in vivo* activity of PKA. Total protein was extracted from the cells harboring pRS423-pr^{CUP}-6×MYC-cki1^{2-200(S125/130A)}, and immunoblotting was performed using a mouse anti-Myc antibody. The relative ratio of phosphorylated (Cki1-P) to non-phosphorylated (Cki1) forms of Cki1, normalized against that of untreated cells, is shown below each lane. Data are representative of at least three independent experiments.

Supplementary Figure S8. The lack of PKA-dependent phosphorylation of Msn2 abolishes the effect of $gas1\Delta$ on rDNA silencing in $gas1\Delta$ cells. Silencing within the rDNA region was assessed by monitoring the growth of cells (10-fold serial dilutions) plated on SC medium without uracil. SC medium was used as a plating control. The spot assay was performed with WT and $gas1\Delta msn2\Delta$ cells containing an empty vector and $gas1\Delta msn2\Delta$ cells expressing WT *MSN2* and $msn2^{S582D, S620D, S625D, S633D}$.

Supplementary Data

Supplementary Table S1. Yeast strains used in this study

Strain	Genotype	Source
BY4741	MATa his $3\Delta 1 \ leu 2\Delta 0 \ met 15\Delta 0 \ ura 3\Delta 0$	Open Biosystems
OMY2798	MATa ade2-1 ura3-1 trp1-1 leu2-3,112 his3-11 can1-100 leu2::mURA3	[6]
DMY2804	MATa ade2-1 ura3-1 trp1-1 leu2-3,112 his3-11 can1-100 RDN1-NTS1::mURA3	[6]
HY0245	MAT a ade2-1 ura3-1 trp1-1 leu2-3,112 his3-11 can1-100 leu2::mURA3 sir2∆::TRP1	This study
HY0291	MATa ade2-1 ura3-1 trp1-1 leu2-3,112 his3-11 can1-100 RDN1-NTS1::mURA3 sir2∆::TRP1	This study
IY1164	MATa ade2-1 ura3-1 trp1-1 leu2-3,112 his3-11 can1-100 leu2::mURA3 gas1Δ::TRP1	This study
IY1165	MATa ade2-1 ura3-1 trp1-1 leu2-3,112 his3-11 can1-100 RDN1-NTS1::mURA3 gas1A::TRP1	This study
IY1167	MAT a ade2-1 ura3-1 trp1-1 leu2-3,112 his3-11 can1-100 leu2::mURA3 gas1∆::TRP1 sir2∆::HIS3	This study
IY1168	MATa ade2-1 ura3-1 trp1-1 leu2-3,112 his3-11 can1-100 RDN1-NTS1::mURA3 gas1A::TRP1 sir2A::HIS3	This study
OMY3010	MATa ade2-1 ura3-1 trp1-1 leu2-3,112 his3-11 can1-100 RAD5 ⁺ with RDN1::ADE2	[6]
IY1185	MATa ade2-1 ura3-1 trp1-1 leu2-3,112 his3-11 can1-100 RAD5 ⁺ with RDN1::ADE2 gas1A::TRP1	This study
IY0236	MATa ade2-1 ura3-1 trp1-1 leu2-3,112 his3-11 can1-100 RAD5 ⁺ with RDN1::ADE2 sir2 Δ ::TRP1	This study
IY1448	MATa ade2-1 ura3-1 trp1-1 leu2-3,112 his3-11 can1-100 RAD5 ⁺ with RDN1::ADE2 gas1A::LEU2 sir2A::TRP1	This study
Y1170	MATa his $3\Delta 1$ leu $2\Delta 0$ met $15\Delta 0$ ura $3\Delta 0$ MSN2-GFP-HIS $3MX6$	This study
Y1171	MAT a his3Δ1 leu2Δ0 met15Δ0 ura3Δ0 MSN2-GFP-HIS3MX6 gas1Δ::LEU2	This study

HY1172	MAT \mathbf{a} his3 $\Delta 1$ leu2 $\Delta 0$ met15 $\Delta 0$ ura3 $\Delta 0$ MSN2-TAP-HIS3MX6	This study
HY1173	MATa his $3\Delta 1$ leu $2\Delta 0$ met $15\Delta 0$ ura $3\Delta 0$ MSN2-TAP-HIS $3MX6$ gas 1Δ ::URA 3	This study
HY1174	MATa his $3\Delta 1$ leu $2\Delta 0$ met $15\Delta 0$ ura $3\Delta 0$ PNC1-GFP-HIS $3MX6$	This study
HY1175	MATa his $3\Delta 1$ leu $2\Delta 0$ met $15\Delta 0$ ura $3\Delta 0$ PNC1-GFP-HIS $3MX6$ gas 1Δ ::URA 3	This study
HY1176	MATa his $3\Delta 1$ leu $2\Delta 0$ met $15\Delta 0$ ura 3Δ PNC1-GFP-HIS $3MX6$ msn 2Δ ::KanMX4 msn 4Δ ::LEU2	This study
HY1177	MATa his3Δ1 leu2Δ0 met15Δ0 ura3Δ PNC1-GFP-HIS3MX6 msn2Δ::KanMX4 msn4Δ::LEU2 gas1Δ::URA3	This study
HY1178	MATa his $3\Delta 1$ leu $2\Delta 0$ met $15\Delta 0$ ura $3\Delta 0$ SIR2-TAP-HIS $3MX6$	This study
HY1179	MATa his $3\Delta 1$ leu $2\Delta 0$ met $15\Delta 0$ ura $3\Delta 0$ SIR2-TAP-HIS $3MX6$ gas 1Δ ::URA 3	This study
HY1180	$MATa$ his $3\Delta 1$ leu $2\Delta 0$ met $15\Delta 0$ ura 3Δ SIR2-TAP-HIS $3MX6$ msn 2Δ ::Kan $MX4$ msn 4Δ ::LEU 2	This study
HY1181	MATa his $3\Delta 1$ leu $2\Delta 0$ met $15\Delta 0$ ura 3Δ SIR2-TAP-HIS $3MX6$ msn 2Δ ::KanMX4 msn 4Δ ::LEU2 gas 1Δ ::URA 3	This study
HY1186	MATa ade2-1 ura3-1 trp1-1 leu2-3,112 his3-11 can1-100 RAD5 ⁺ with RDN1::ADE2 msn2∆::URA3 msn4∆::LEU2	This study
HY1187	MATa ade2-1 ura3-1 trp1-1 leu2-3,112 his3-11 can1-100 RAD5 ⁺ with RDN1::ADE2 msn2A::URA3 msn4A::LEU2 gas1A::TRP1	This study
HY1320	$MATa$ his3 $\Delta 1$ leu2 $\Delta 0$ met15 $\Delta 0$ ura3 $\Delta 0$ MSN2-GFP-HIS3MX6 [pRS415]	This study
HY1323	$MATa$ his $3\Delta 1$ leu $2\Delta 0$ met $15\Delta 0$ ura $3\Delta 0$ MSN2-GFP-HIS $3MX6$ gas 1Δ ::URA 3 [pRS415]	This study
HY1324	$MATa$ his $3\Delta 1$ leu $2\Delta 0$ met $15\Delta 0$ ura $3\Delta 0$ MSN2-GFP-HIS $3MX6$ gas 1Δ ::URA 3 [pRS415 GAS1-TAP]	This study
HY1325	MATa his $3\Delta 1$ leu $2\Delta 0$ met $15\Delta 0$ ura $3\Delta 0$ MSN2-GFP-HIS $3MX6$ gas 1Δ ::URA 3 [pRS415 gas $1^{E161Q, E262Q}$ -TAP]	This study
HY1332	$MATa$ his $3\Delta 1$ leu $2\Delta 0$ met $15\Delta 0$ ura $3\Delta 0$ MSN2-TAP-HIS $3MX6$ [pRS415]	This study
HY1335	$MATa$ his $3\Delta 1$ leu $2\Delta 0$ met $15\Delta 0$ ura $3\Delta 0$ MSN2-TAP-HIS $3MX6$ gas 1Δ ::URA 3 [pRS415]	This study
HY1336	MATa his3Δ1 leu2Δ0 met15Δ0 ura3Δ0 MSN2-TAP-HIS3MX6 gas1Δ::URA3 [pRS415 GAS1-GFP]	This study

HY1337	$MATa$ his $3\Delta 1$ leu $2\Delta 0$ met $15\Delta 0$ ura $3\Delta 0$ MSN2-TAP-HIS $3MX6$ gas 1Δ :: URA 3 [pRS415 gas $1^{E161Q, E262Q}$ -GFP]	This study	
HY1402	$MATa$ his $3\Delta 1$ leu $2\Delta 0$ met $15\Delta 0$ ura $3\Delta 0$ PNC1-TAP-HIS $3MX6$ [pRS415]	This study	
HY1403	$MATa$ his $3\Delta 1$ leu $2\Delta 0$ met $15\Delta 0$ ura $3\Delta 0$ PNC1-TAP-HIS $3MX6$ gas 1Δ ::URA 3 [pRS415]	This study	
HY1404	$MATa$ his $3\Delta 1$ leu $2\Delta 0$ met $15\Delta 0$ ura $3\Delta 0$ PNC1-TAP-HIS $3MX6$ gas 1Δ ::URA 3 [pRS415 GAS1-GFP]	This study	
HY1405	$MATa$ his $3\Delta 1$ leu $2\Delta 0$ met $15\Delta 0$ ura $3\Delta 0$ PNC1-TAP-HIS $3MX6$ gas 1Δ :: URA 3 [pRS415 gas $1^{E161Q, E262Q}$ -GFP]	This study	
HY1338	$MATa$ his $3\Delta 1$ leu $2\Delta 0$ met $15\Delta 0$ ura $3\Delta 0$ SIR2-TAP-HIS $3MX6$ [pRS415]	This study	
HY1341	$MATa$ his $3\Delta 1$ leu $2\Delta 0$ met $15\Delta 0$ ura $3\Delta 0$ SIR2-TAP-HIS $3MX6$ gas 1Δ ::URA3 [pRS415]	This study	
HY1342	$MATa$ his $3\Delta 1$ leu $2\Delta 0$ met $15\Delta 0$ ura $3\Delta 0$ SIR2-TAP-HIS $3MX6$ gas 1Δ ::URA3 [pRS415 GAS1-GFP]	This study	
HY1343	MATa his $3\Delta 1 \ leu 2\Delta 0 \ met 15\Delta 0 \ ura 3\Delta 0 \ SIR2-TAP-HIS3MX6 \ gas 1\Delta:: URA3 \ [pRS415 \ gas 1^{E161Q, E262Q}-GFP]$	This study	
HY1350	MATa ade2-1 ura3-1 trp1-1 leu2-3,112 his3-11 can1-100 leu2::mURA3 [pRS413]	This study	
HY1353	MATa ade2-1 ura3-1 trp1-1 leu2-3,112 his3-11 can1-100 RDN1-NTS1::mURA3 [pRS413]	This study	
HY1356	MATa ade2-1 ura3-1 trp1-1 leu2-3,112 his3-11 can1-100 leu2::mURA3 gas1A::TRP1 [pRS413]	This study	
HY1357	MATa ade2-1 ura3-1 trp1-1 leu2-3,112 his3-11 can1-100 leu2::mURA3 gas1Δ::TRP1 [pRS413 GAS1-GFP]	This study	
HY1358	MATa ade2-1 ura3-1 trp1-1 leu2-3,112 his3-11 can1-100 leu2::mURA3 gas1Δ::TRP1 [pRS413 gas1 ^{E161Q, E262Q} - GFP]	This study	
HY1359	MATa ade2-1 ura3-1 trp1-1 leu2-3,112 his3-11 can1-100 RDN1-NTS1::mURA3 gas1A::TRP1 [pRS413]	This study	
HY1360	MATa ade2-1 ura3-1 trp1-1 leu2-3,112 his3-11 can1-100 RDN1-NTS1::mURA3 gas1A::TRP1 [pRS413 GAS1- GFP]	This study	
HY1361	MATa ade2-1 ura3-1 trp1-1 leu2-3,112 his3-11 can1-100 RDN1-NTS1::mURA3 gas1Δ::TRP1 [pRS413 gas1 ^{E161Q} , ^{E262Q} -GFP]	This study	
HY1344	MATa ade2-1 ura3-1 trp1-1 leu2-3,112 his3-11 can1-100 RAD5 ⁺ with RDN1::ADE2 [pRS415]	This study	
HY1347	MATa ade2-1 ura3-1 trp1-1 leu2-3,112 his3-11 can1-100 RAD5 ⁺ with RDN1::ADE2 gas1∆::URA3 [pRS415]	This study	

HY1348	MATa ade2-1 ura3-1 trp1-1 leu2-3,112 his3-11 can1-100 RAD5 ⁺ with RDN1::ADE2 gas1∆::URA3 [pRS415 GAS1- GFP]	This study
HY1349	MATa ade2-1 ura3-1 trp1-1 leu2-3,112 his3-11 can1-100 RAD5 ⁺ with RDN1::ADE2 gas1A::URA3 [pRS415 gas1 E161Q, E262Q-GFP]	This study
HY1202	MATa his $3\Delta 1 \ leu 2\Delta 0 \ met 15\Delta 0 \ ura 3\Delta 0 \ TPK1-GFP-HIS3MX6$	This study
HY1203	MATa his $3\Delta 1 \ leu 2\Delta 0 \ met 15\Delta 0 \ ura 3\Delta 0 \ TPK1-GFP-HIS3MX6 \ gas 1\Delta::URA3$	This study
HY1398	$MATa$ his $3\Delta 1$ leu $2\Delta 0$ met $15\Delta 0$ ura $3\Delta 0$ TPK1-GFP-HIS $3MX6$ [pRS415]	This study
HY1399	$MATa$ his $3\Delta 1$ leu $2\Delta 0$ met $15\Delta 0$ ura $3\Delta 0$ TPK1-GFP-HIS $3MX6$ gas 1Δ ::URA3 [pRS415]	This study
HY1400	$MATa$ his $3\Delta 1$ leu $2\Delta 0$ met $15\Delta 0$ ura $3\Delta 0$ TPK1-GFP-HIS $3MX6$ gas 1Δ ::URA3 [pRS415 GAS1-TAP]	This study
HY1401	$MATa$ his $3\Delta 1$ leu $2\Delta 0$ met $15\Delta 0$ ura $3\Delta 0$ TPK1-GFP-HIS3MX6 gas 1Δ ::URA3 [pRS415 gas $1^{E161Q, E262Q}$ -TAP]	This study
HY1497	MATa his $3\Delta 1 \ leu 2\Delta 0 \ met 15\Delta 0 \ ura 3\Delta 0 \ HIS3MX6:: P_{RPL7B}$ -HA-BCY1	This study
HY1498	MATa his $3\Delta 1 \ leu 2\Delta 0 \ met 15\Delta 0 \ ura 3\Delta 0 \ HIS3MX6:: P_{RPL7B}$ -HA-BCY1 gas 1Δ :: URA3	This study
HY1499	MATa his $3\Delta 1 \ leu 2\Delta 0 \ met 15\Delta 0 \ ura 3\Delta 0 \ HIS3MX6:: P_{RPL7B}$ -HA-BCY1 mpk 1Δ ::LEU2	This study
HY1500	MATa his $3\Delta 1 \ leu 2\Delta 0 \ met 15\Delta 0 \ ura 3\Delta 0 \ HIS3MX6:: P_{RPL7B}$ -HA-BCY1 mpk 1Δ :: LEU2 gas 1Δ :: URA3	This study
HY1390	MATa ade2-1 ura3-1 trp1-1 leu2-3,112 his3-11 can1-100 leu2::mURA3 gas2∆::TRP1	This study
HY1391	MATa ade2-1 ura3-1 trp1-1 leu2-3,112 his3-11 can1-100 RDN1-NTS1::mURA3 gas2A::TRP1	This study
HY1392	MATa ade2-1 ura3-1 trp1-1 leu2-3,112 his3-11 can1-100 leu2::mURA3 gas3∆::TRP1	This study
HY1393	MATa ade2-1 ura3-1 trp1-1 leu2-3,112 his3-11 can1-100 RDN1-NTS1::mURA3 gas3Δ::TRP1	This study
HY1394	MATa ade2-1 ura3-1 trp1-1 leu2-3,112 his3-11 can1-100 leu2::mURA3 gas4∆::TRP1	This study
HY1395	MATa ade2-1 ura3-1 trp1-1 leu2-3,112 his3-11 can1-100 RDN1-NTS1::mURA3 gas4Δ::TRP1	This study
HY1396	MATa ade2-1 ura3-1 trp1-1 leu2-3,112 his3-11 can1-100 leu2::mURA3 gas5∆::TRP1	This study

HY1397	MATa ade2-1 ura3-1 trp1-1 leu2-3,112 his3-11 can1-100 RDN1-NTS1::mURA3 gas5Δ::TRP1	This study
HY1388	MATa ade2-1 ura3-1 trp1-1 leu2-3,112 his3-11 can1-100 leu2::mURA3 bgl2A::TRP1	This study
HY1389	MATa ade2-1 ura3-1 trp1-1 leu2-3,112 his3-11 can1-100 RDN1-NTS1::mURA3 bgl2Δ::TRP1	This study
HY1444	MATa ade2-1 ura3-1 trp1-1 leu2-3,112 his3-11 can1-100 RAD5 ⁺ with RDN1::ADE2 gas2A::TRP1	This study
HY1445	MATa ade2-1 ura3-1 trp1-1 leu2-3,112 his3-11 can1-100 RAD5 ⁺ with RDN1::ADE2 gas3A::TRP1	This study
HY1446	MATa ade2-1 ura3-1 trp1-1 leu2-3,112 his3-11 can1-100 RAD5 ⁺ with RDN1::ADE2 gas4A::TRP1	This study
HY1447	MATa ade2-1 ura3-1 trp1-1 leu2-3,112 his3-11 can1-100 RAD5 ⁺ with RDN1::ADE2 gas5A::TRP1	This study
HY1443	MATa ade2-1 ura3-1 trp1-1 leu2-3,112 his3-11 can1-100 RAD5 ⁺ with RDN1::ADE2 bgl2Δ::TRP1	This study

Supplementary	y Table S2. Oligonucleotide	primers used for ChIP assa	vs in this study

Locus	Forward Primer	Reverse Primer
rDNA-25S	CGACTAACCCACGTCCAACT	CCGAATGAACTAGCCCTGAA
rDNA-NTS1	TCCCCACTGTTCACTGTTCA	AGGGCTTTCACAAAGCTTCC
rDNA-NTS2/18S	AAGATGCCCACGATGAGACT	GGGAGGTACTTCATGCGAAA
rDNA-18S	CCAGAACGTCTAAGGGCATC	CTCACCAGGTCCAGACACAA
CUP1	TGAAGGTCATGAGTGCCAAT	TTCGTTTCATTTCCCAGAGCA
PNC1 promoter	GATCAAGGTGGCACACAGGG	ATACATAGTGGGCCAAACGG
ACTI	TGACTGACTACTTGATGAAG	ACAGAAGGATGGAACAAAGC

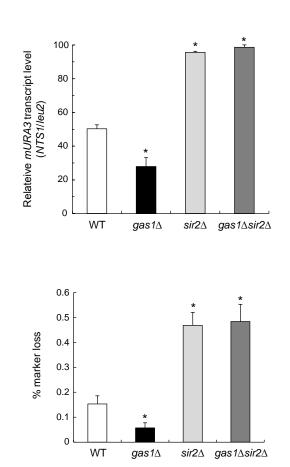


SC

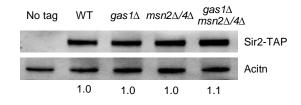
SC-Ura

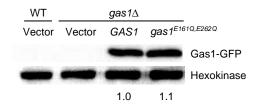
Cells		
••••		leu2::mURA3 NTS1::mURA3
• • • • •		NTS1::mURA3
•••		leu2::mURA3 sir2∆
6 6 6 ÷	• • • • •	leu2::mURA3 NTS1::mURA3
• • • • • •	• • • • •	leu2::mURA3 gas1∆
🕑 🌒 🌒 🌮 🕗	© >	leu2::mURA3 NTS1::mURA3
S 🖲 🕲 🍕 🕻 👘		leu2::mURA3
🌒 🚳 🏘 👌 📩 👘	🔴 🌰 🏶 🥐 🕚 🔵	leu2::mURA3 NTS1::mURA3

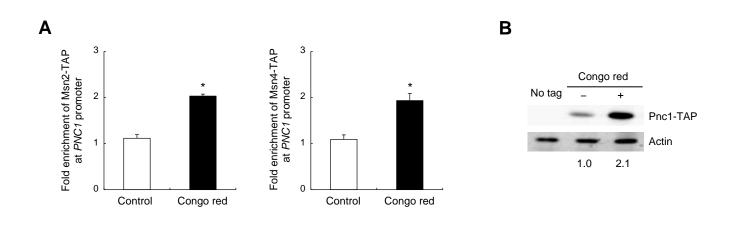
В



С

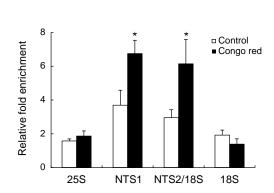




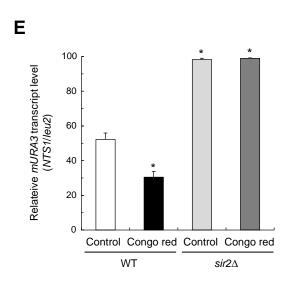


D





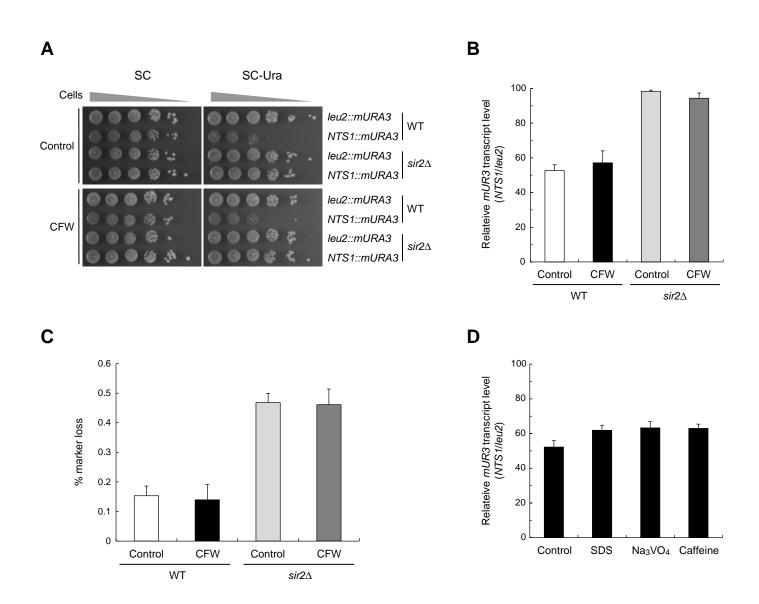
	SC	SC-Ura	
Cells	6		
			leu2::mURA3
Quarteral	48000		NTS1::mURA3
Control			leu2::mURA3 sir2∆
			NTS1::mURA3
	***		leu2::mURA3
Congo	🕒 🔍 🏶 😒 😒 🕓	0 0 0	leu2::mURA3 NTS1::mURA3
red		•••	leu2::mURA3
	•••		NTS1::mURA3

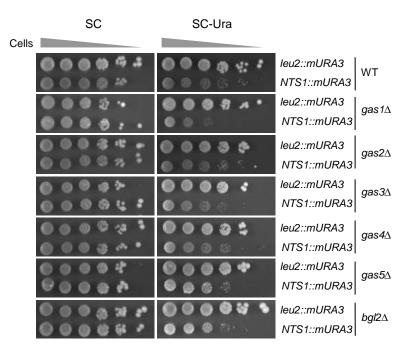


0.6 0.5 0.4 0.3 0.2 0.1 0 Control Congo red WT sir2Δ

F

Г







Α

