

1 Supporting Online Materials for

2
3 Molecular Network Analysis of Endometriosis Reveals a
4 Novel Role for c-Jun Regulated Macrophage Activation
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16 Materials and Methods

17 Figure S1. Sample size estimation and power calculations

18 Figure S2. Multiplex immunoassay performance

19 Figure S3. Non-negative matrix factorization of randomized and reduced data sets

20 Figure S4. Peritoneal aspirate characteristics across molecular subpopulations

21 Figure S5. Isolation and relative cytokine secretion of adherent peritoneal macrophages.

22 Figure S6. Differential cytokine secretion by peritoneal macrophages

23 Figure S7. Enrichment analysis of ASRM III/IV cytokines

24 Figure S8. Inferred co-expression of ASRM III/IV cytokines

25 Table S1. Pair-wise Wilcoxon rank-sum tests – Treatment Status

26 Table S2. Pair-wise Wilcoxon rank-sum tests – ASRM Staging

27 Table S3. Pair-wise Wilcoxon rank-sum tests – Cycle Phase

28 Table S4. Pair-wise Wilcoxon rank-sum tests – Recurrence Status

29 Table S5. Pair-wise Wilcoxon rank-sum tests – Lesion Distribution

30 Table S6. Pair-wise Wilcoxon rank-sum tests – Primary Indication

31 Table S7. Reported associations between peritoneal cytokines, chemokines, growth
32 factors and pelvic endometriosis.

33 Table S8. Over-represented transcriptional binding sites among macrophage secreted
34 cytokines

35 Table S9. Luminex Targets and Assay Performance Characteristics

36 Table S10. Leukocyte Sub-populations Among Peritoneal Aspirates

37 Table S11. Kinase Inhibitor Treatments
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38 Supplemental Materials and Methods

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40 *Sample Size Calculations*

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42 Sample size estimates were determined using previously reported effect sizes from case-control
43 studies investigating peritoneal fluid cytokine associations with endometriosis. Twenty
44 publications in which parametric statistical findings were reported indicated a broad range of
45 effect sizes achieving significance ($P < 0.05$). Shown in [Fig. S1](#), published effect sizes - reported
46 as Standardized Mean Differences (SMD) following log-normalization - ranged between 0.54-
47 4.32 with a median of 0.83. Statistical power for two-tailed Mann-Whitney U-tests tests of
48 association between cytokine concentrations and clinical variables was calculated using
49 G*Power3. Maintaining a 2:1 case:control allocation ratio, approximately 60 samples are
50 required to obtain 0.8 power for moderate effect sizes corresponding to SMD = 0.8.

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53 *Gene Set Enrichment Analysis*

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55 Gene set enrichment analysis was performed as originally described ([Subramanian 2005](#)) using
56 gene profiles derived from the Immune Response in silico (IRIS) expression compendia and
57 Human Immune Cell Transcriptome (GEO Accession GSE22886 and GSE3982, respectively)
58 ([Abbas 2005](#), [Jeffrey 2006](#)).

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60 For all analyses, 22,283 U133A probe sets were first mapped to 14,339 unique genes and
61 expressed sequence tags by taking the maximum intensity value across all probes. Expression
62 values for all genes were then median-centered across all profiles in the compendia, and
63 enrichment scores for each profile calculated using a weighting exponent of one. Finally,
64 empirical P values for each enrichment score were determined by generating null distributions
65 of scores from 1000 random cytokine signatures of equivalent size selected from the 47
66 detected cytokines or 79 corresponding receptors.

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69 *Pair-wise Enrichment and Reconstruction of Hierarchical Immune Cell Networks*

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71 To quantify the degree to which a set of differentially regulated cytokines contribute to specific
72 routes of intercellular communication, we introduce a pair-wise enrichment statistic that captures
73 the coordinate expression of signature cytokines by secreting cell populations and their cognate
74 receptors by responding cell populations. Specifically, we define the pair-wise enrichment score,
75 ES_{ij} between secreting population i and responding population j as a two-dimensional
76 generalization of the Kolmogorov-Smirnov statistic ([Ni 2012](#)):

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$$ES_{ij} = \max |F(x_i, y_j) - G(x_0, y_0)|$$

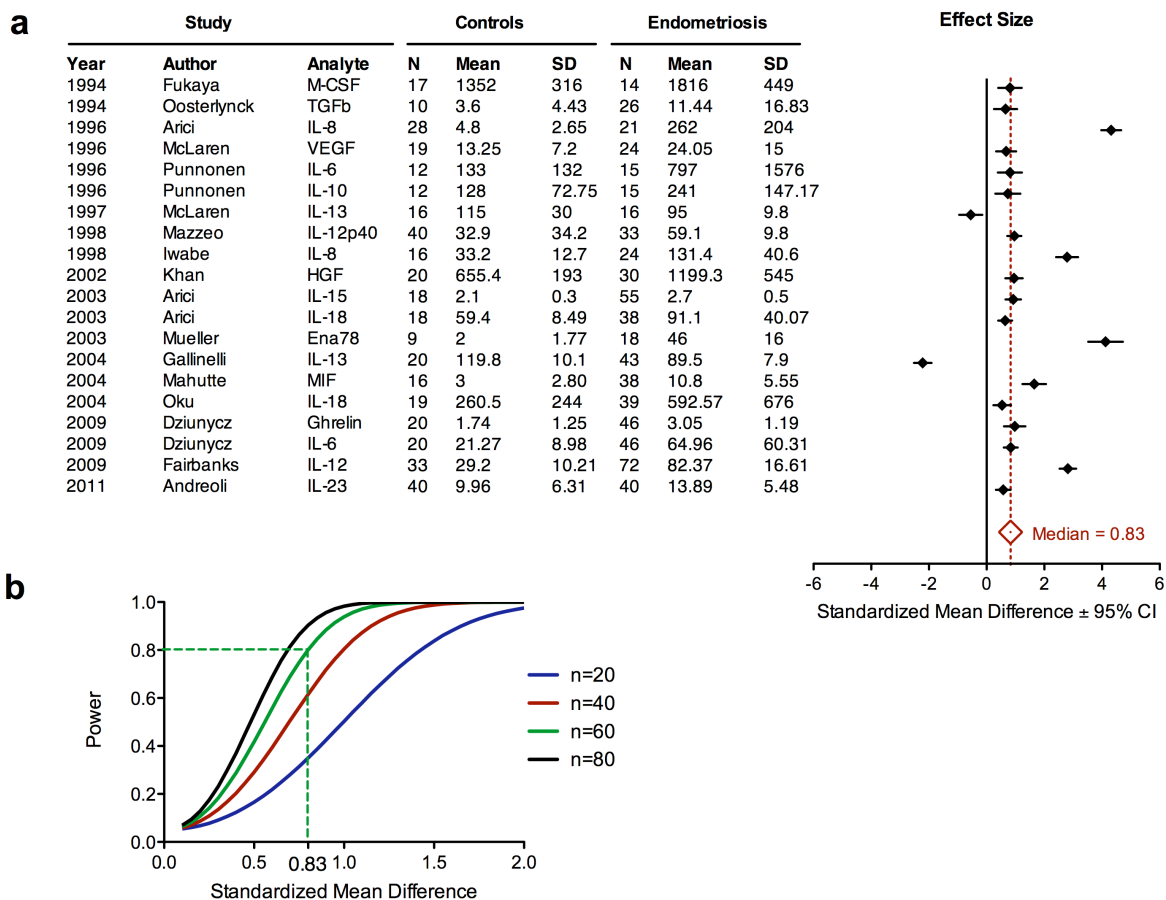
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80 where $F(x_i, y_j)$ is the two-dimensional empirical cumulative distribution function (ECDF) for
81 signature cytokine and receptor expression levels, x_i and y_j , respectively, and $G(x_0, y_0)$ is the
82 reference ECDF for all extracellular gene products and their receptors across all lineages. The
83 reference distribution may be constructed explicitly or approximated; here we use the median
84 cytokine and receptor expression levels of the forty-seven assayed proteins across all cell
85 lineages to approximate $G(x_0, y_0)$.

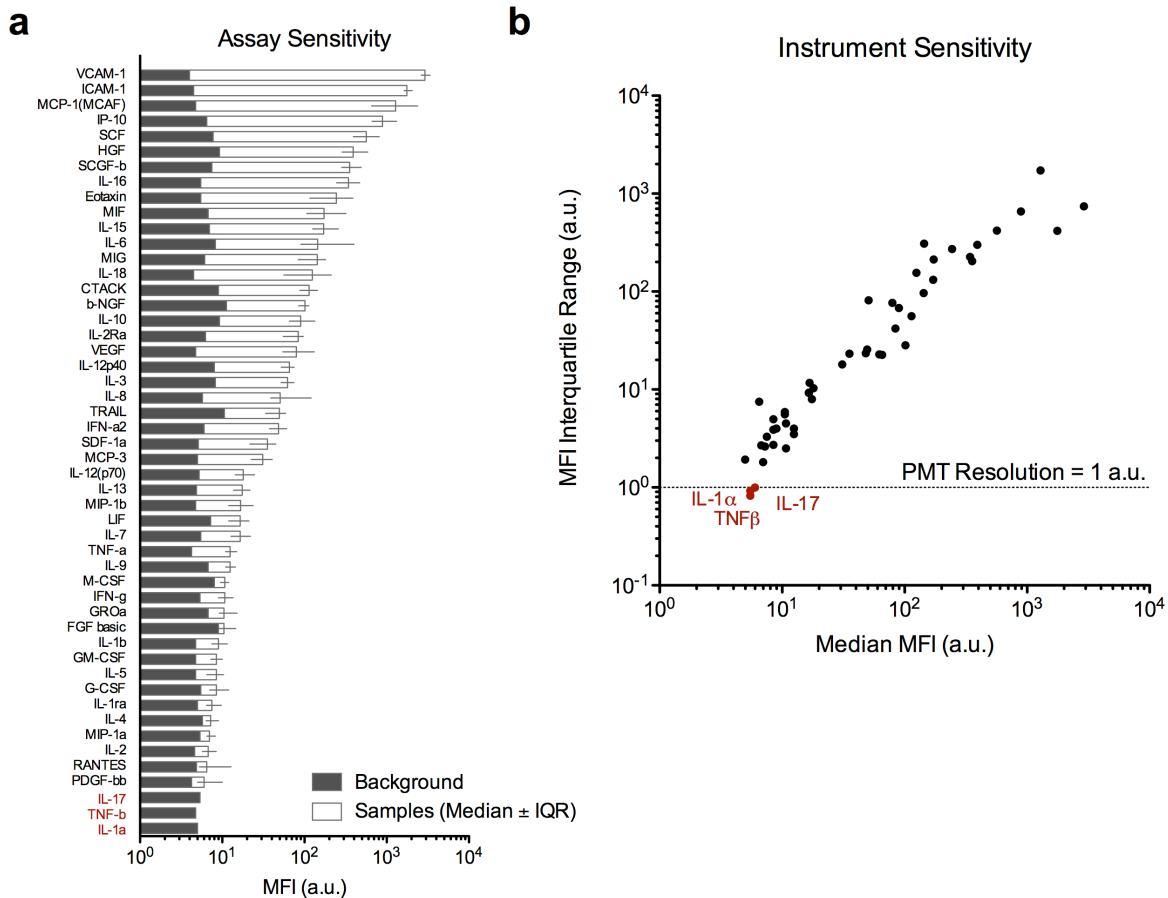
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87 For n distinct cell lineages, the 2D-KS enrichment score ES_{ij} is calculated over all n^2 possible
88 pairwise combinations of secreting and responding populations. Null distributions of ES_{ij} for

89 each n^2 interactions are similarly constructed from randomly generated cytokine/receptor
90 signatures to obtain the corresponding significance levels. Finally, we rearrange the edge-
91 directed network of significant ($P < 0.05$) interactions according to decreasing out-degree to
92 evaluate the hierarchy of cell-cell interactions.
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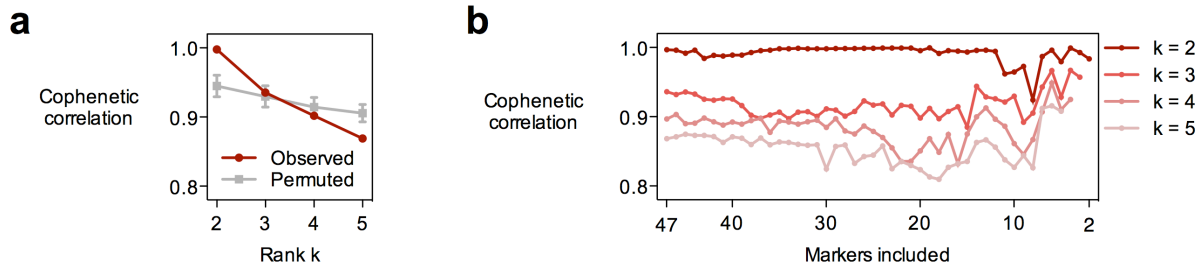
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 94 **Figure S1. Sample size estimation and power calculations.** (a) Published associations
 95 between peritoneal fluid cytokines and endometriosis were evaluated to estimate the range of
 96 anticipated effect sizes. Standardized mean differences \pm 95% confidence intervals are shown
 97 to the right (Range = 0.54-4.32; Median = 0.83). (b) Power curves for two-tailed Mann-Whitney
 98 U-tests determined at the $\alpha=0.05$ significance level for the indicated effect sizes and total
 99 sample size, assuming a 2:1 case:control allocation.
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Figure S2. Multiplex immunoassay performance

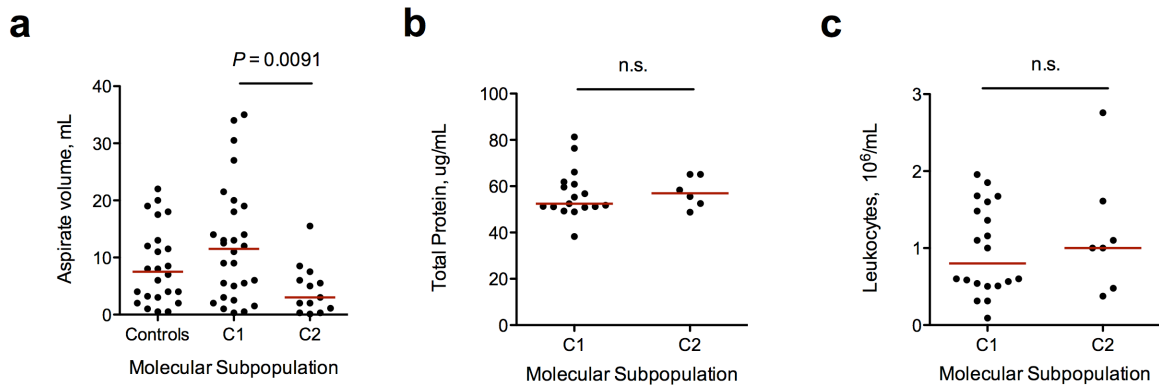
Prior to analysis, raw fluorescent intensities were evaluated for relative variation above background levels (a) and total sample variation above instrument resolution (b). In (a), bars represent median \pm interquartile range. A majority of concentrations for three peritoneal analytes (IL-1 α , TNF β , and IL-17) fall below the lower detection limits for the immunoassay.



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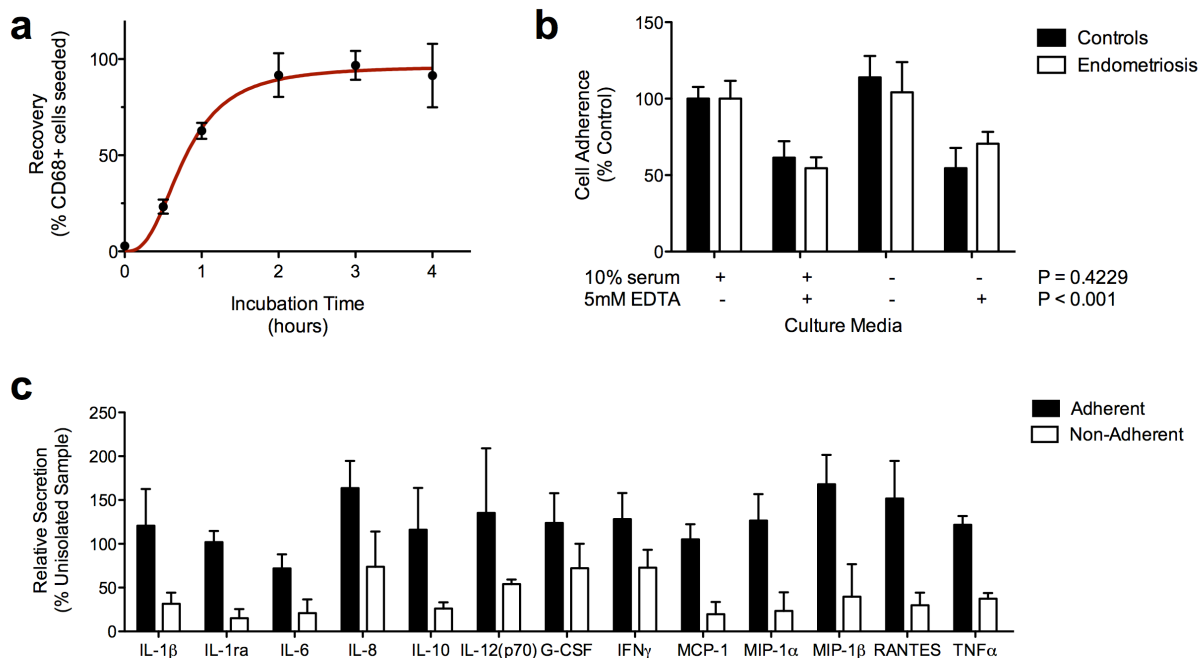
Figure S3. Non-negative matrix factorization of randomized and reduced data sets

(a) Cophenetic correlation of 1000 rank k factorizations in the observed (red) and permuted (shaded) data sets. Error bars indicate 95% confidence intervals. (b) Optimal factorization into two subsets is conserved across reduced data sets comprising markers exhibiting maximal variation.

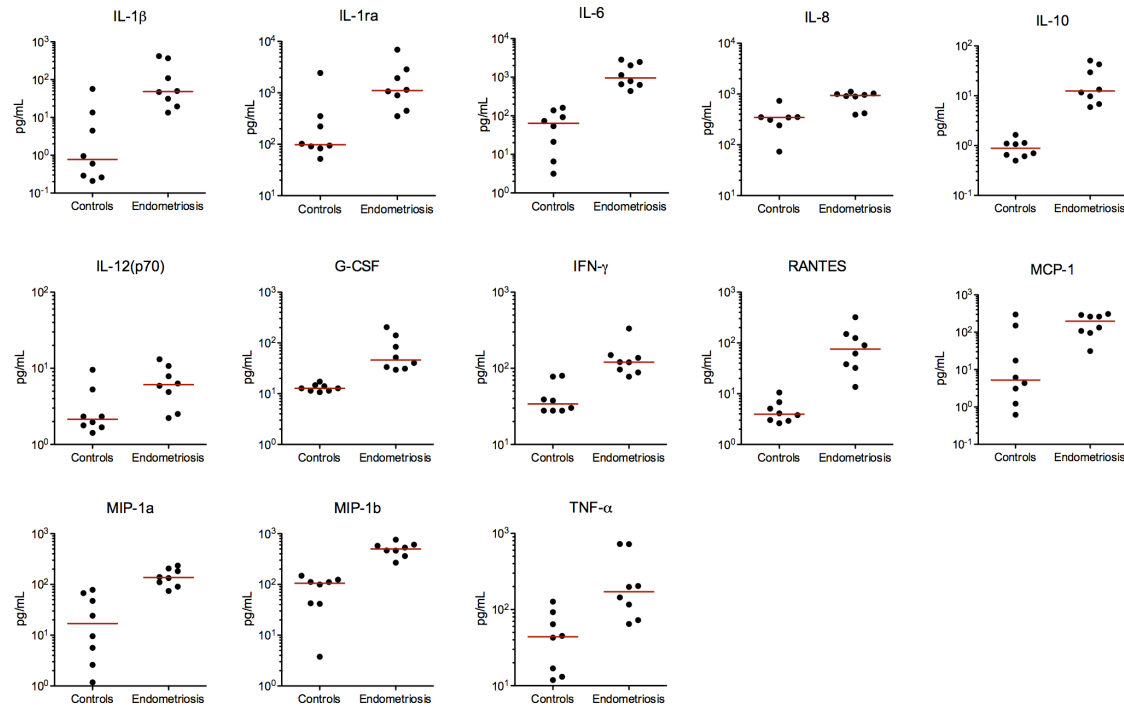


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Figure S4. Peritoneal aspirate characteristics across molecular subpopulations
 Patient subpopulations defined by low (C1) and elevated (C2) peritoneal fluid cytokine concentrations show markedly reduced median aspirate volumes (a), but equivalent total protein and leukocyte levels (b and c).



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 119 **Figure S5. Isolation and relative cytokine secretion of adherent peritoneal macrophages.**
 120 (A) Recovery of adherent CD68+ macrophages as a function of static incubation period with
 121 tissue culture treated polystyrene plates. (B) After two hours of incubation, macrophage
 122 adherence is sensitive to the presence of divalent cations, but not substrate deposition of serum
 123 components among both control and endometriosis samples. P values indicate significance by
 124 two-way ANOVA. (C) Cytokine secretion among adherent and non-adherent peritoneal
 125 leukocytes (relative to unisolated, sample-matched controls). For all panels, data are mean \pm
 126 s.e.m. of three donor samples.

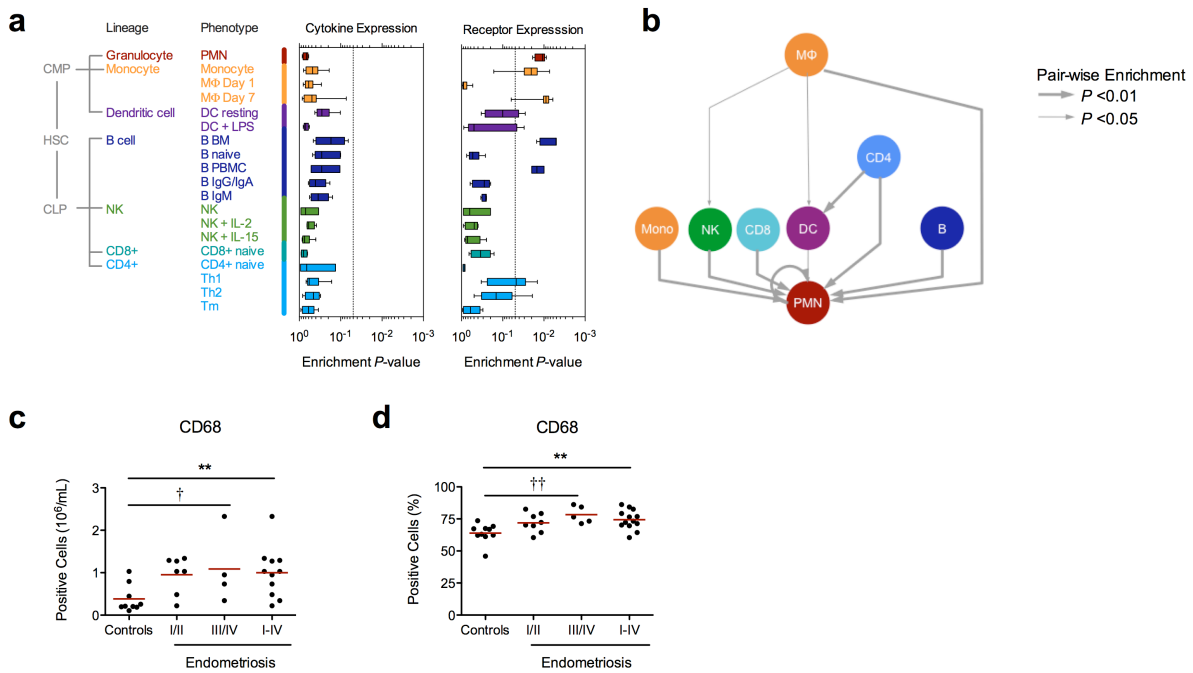
a**b**

Cytokine	Rank-sum	P-value	Fold Change
IL-1β	0.0047		62.5
IL-1ra	0.0086		11.28
IL-6	0.0002		15.1
IL-8	0.0012		2.71
IL-10	0.0002		14.13
IL-12(p70)	0.0312		2.85
G-CSF	0.0009		3.62
IFN-γ	0.0016		3.54
MCP-1	0.0281		37.7
MIP-1α	0.0003		8.09
MIP-1β	0.0002		4.73
RANTES	0.0002		19.2
TNF-α	0.0003		3.90

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Figure S6. Differential cytokine secretion by peritoneal macrophages

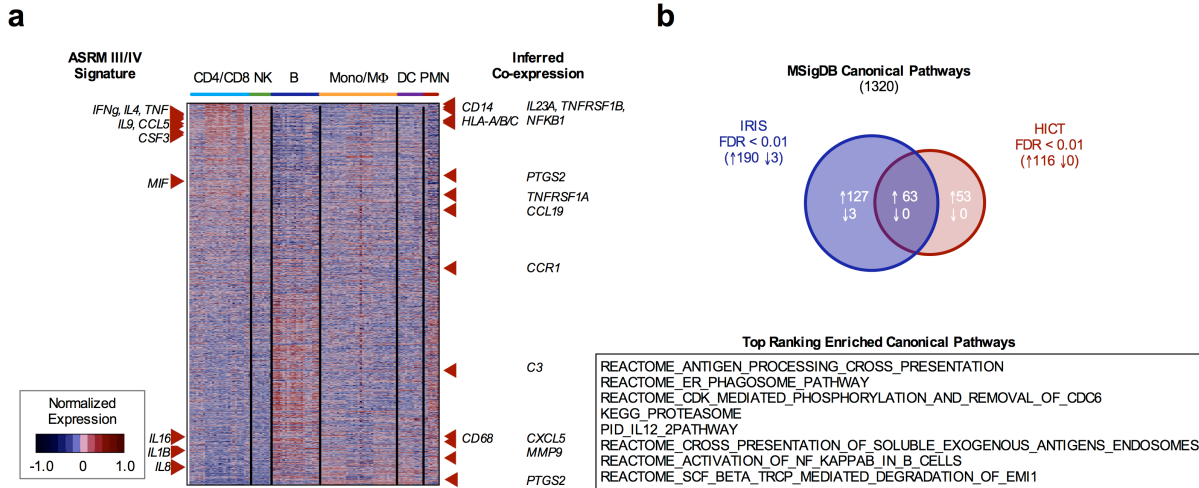
(a) Thirteen of fifty cytokines assayed in media conditioned by isolated peritoneal macrophages demonstrated significant increases (Wilcoxon rank sum test $P < 0.05$ and minimum two-fold change) in endometriosis samples versus controls. (b) Statistical summary for differentially secreted cytokines in (a).



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Figure S7. Enrichment analysis of ASRM III/IV cytokines

(A) Significance of lineage-specific enrichment scores for ASRM III/IV cytokine expression obtained by GSEA among Immune Response *in silico* (IRIS) transcriptional profiles. (B) Hierarchy of enriched intercellular cytokine-receptor interactions. Flow cytometric confirmation of increased (C) absolute and (D) relative CD68+ macrophage abundance among endometriosis patient populations. Increasing CD68+ counts among patient subpopulations grouped by ASRM stage are shown for comparison. $**P < 0.01$ Wilcoxon rank-sum test; $†P < 0.05$, $††P < 0.01$, linear trend test.



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Figure S8. Inferred co-expression of ASRM III/IV cytokines

(A) Transcripts from the IRIS compendia rank-ordered according to median correlation distance to cytokines differentially associated with ASRM stage III/IV endometriosis (left). In contrast to co-expression analysis using the multivariate consensus signature, inferred co-expression of macrophage surface markers and reported disease markers were broad and incoherent (right).

(B) Gene set enrichment of IRIS and HICT co-expression profiles derived from the ASRM III/IV signature cytokines likewise demonstrate incoherent enrichment of unrelated canonical pathways.

152 **Table S1.** Pair-wise Wilcoxon rank-sum tests – Treatment Status
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Cytokine	Median Concentration (pg/mL)			Untreated vs. Controls		Treated vs. Controls		Treated vs. Untreated	
	Controls	Untreated	Treated	Unadjusted	BH	Unadjusted	BH	Unadjusted	BH
PDGF-bb	2.50	2.50	2.35	0.5274	0.1148	0.9617	0.9414	0.4870	0.45478
IL-1b	0.23	0.26	0.24	0.0131	0.0307	0.3586	0.8719	0.1906	0.39612
IP-10	667.11	707.89	826.82	0.5908	0.1153	0.8113	0.9006	0.8870	0.59928
IL-1ra	12.27	13.06	11.30	0.3851	0.1081	0.4163	0.8719	0.1309	0.39230
IL-2	0.15	0.15	0.13	0.6834	0.1153	0.6782	0.9006	0.4498	0.44631
IL-4	0.32	0.35	0.33	0.0193	0.0307	0.3069	0.8719	0.2548	0.39612
IL-5	0.52	0.52	0.56	0.7176	0.1162	0.0939	0.8719	0.1301	0.39230
IL-6	10.22	10.97	10.70	0.5039	0.1132	0.6906	0.9006	0.3651	0.44351
IL-7	4.93	5.06	5.19	0.4655	0.1081	0.4445	0.8719	0.8730	0.59928
IL-8	2.72	3.48	3.29	0.0221	0.0307	0.1150	0.8719	0.4190	0.44351
IL-9	2.81	3.27	2.94	0.0068	0.0307	0.6437	0.9006	0.0929	0.39230
IL-10	2.08	2.23	2.32	0.4608	0.1081	0.3988	0.8719	0.8660	0.59928
IL-12(p70)	2.45	2.66	2.79	0.4154	0.1081	0.4353	0.8719	0.8870	0.59928
IL-13	0.64	0.72	0.71	0.2187	0.0881	0.3310	0.8719	0.8800	0.59928
IL-15	1.26	1.32	1.35	0.7470	0.1162	0.6906	0.9006	0.8381	0.59928
IL-17	0.70	0.76	0.68	0.0192	0.0307	0.7236	0.9006	0.1467	0.39230
Eotaxin	70.03	89.44	109.51	0.3606	0.1081	0.1474	0.8719	0.3842	0.44351
FGFb	0.90	0.94	0.90	0.1115	0.0777	0.8483	0.9006	0.1308	0.39230
G-CSF	1.69	2.03	1.76	0.0284	0.0330	0.8234	0.9006	0.1451	0.39230
GM-CSF	0.10	0.10	0.08	0.4457	0.1081	0.8232	0.9006	0.2850	0.41140
IFN-g	23.77	27.51	26.04	0.0576	0.0573	0.3724	0.8719	0.2481	0.39612
MCP-1	44.98	64.62	43.12	0.1465	0.0850	0.8361	0.9006	0.1453	0.39230
MIP-1a	0.85	0.89	0.91	0.1653	0.0881	0.4349	0.8719	0.7351	0.59853
MIP-1b	1.23	1.20	1.25	0.6895	0.1153	0.5450	0.9006	0.3650	0.44351
RANTES	2.34	2.66	2.56	0.1442	0.0850	0.3809	0.8719	0.5940	0.52393
TNF-a	3.80	4.20	3.97	0.0709	0.0617	0.7741	0.9006	0.1451	0.39230
VEGF	15.93	17.27	16.65	0.4288	0.1081	0.4168	0.8719	0.8870	0.59928
CTACK	104.86	94.18	82.78	0.2278	0.0881	0.1388	0.8719	0.4665	0.44889
GROa	3.73	3.73	3.45	0.7997	0.1185	0.3985	0.8719	0.1309	0.39230
ICAM-1	828.72	831.64	827.01	0.3447	0.1081	0.9113	0.9325	0.4039	0.44351
IL-1a	0.13	0.12	0.13	0.7505	0.1162	0.4978	0.9006	0.2620	0.39612
IL-2Ra	49.19	45.35	46.20	0.2248	0.0881	0.6444	0.9006	0.1606	0.39230
IL-3	71.01	69.32	64.54	0.6783	0.1153	0.6558	0.9006	0.3604	0.44351
IL-12p40	216.90	219.76	189.39	0.8718	0.1248	0.2086	0.8719	0.2411	0.39612
IL-16	185.68	188.22	189.44	0.4655	0.1081	0.7989	0.9006	0.6316	0.54202
IL-18	6.08	5.22	4.04	0.8900	0.1248	0.8610	0.9006	0.9575	0.63342
LIF	1.92	1.82	1.98	0.6782	0.1153	0.7866	0.9006	1.0000	0.63507
MCP-3	16.09	17.08	13.39	0.2218	0.0881	0.6907	0.9006	0.1357	0.39230
M-CSF	0.33	0.32	0.33	0.8959	0.1248	1.0000	0.9414	0.9929	0.63507
MIF	75.35	96.61	88.26	0.0825	0.0639	0.3477	0.8719	0.4140	0.44351
MIG	452.94	436.38	469.03	0.6176	0.1153	0.2790	0.8719	0.4451	0.44631
b-NGF	7.32	7.38	6.47	0.6448	0.1153	0.2517	0.8719	0.2010	0.39612
SCF	82.52	71.04	94.45	0.3069	0.1081	0.4937	0.9006	0.1606	0.39230
SCGF	1261.96	1216.32	1118.20	0.3770	0.1081	0.9619	0.9414	0.5578	0.50608
SDF-1a	49.09	51.09	51.82	0.6067	0.1153	1.0000	0.9414	0.7091	0.59258
TNFb	0.32	0.32	0.32	0.4115	0.1081	0.5954	0.9006	0.2393	0.39612
TRAIL	32.60	31.65	23.94	0.6952	0.1153	0.0370	0.8719	0.1887	0.39612
VCAM-1	1156.08	1212.30	1068.50	0.7762	0.1175	0.1151	0.8719	0.1061	0.39230
HGF	331.54	342.44	359.96	0.1942	0.0881	0.3642	0.8719	0.8382	0.59928
IFNa2	17.43	16.94	16.24	0.6783	0.1153	0.2386	0.8719	0.3420	0.44351

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155 **Table S2.** Pair-wise Wilcoxon rank-sum tests – ASRM Staging
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Cytokine	Median Concentration (pg/mL)			ASRM I/II vs. Controls		ASRM III/IV vs. Controls		ASRM III/IV vs. I/II	
	Controls	Minimal/Mild	Moderate/Severe	Unadjusted	BH	Unadjusted	BH	Unadjusted	BH
PDGF-bb	2.50	2.32	3.03	0.5566	0.9805	0.0550	0.0744	0.0190	0.0531
IL-1b	0.23	0.24	0.30	0.3395	0.9805	0.0004	0.0032	0.0095	0.0478
IP-10	667.11	707.89	707.97	0.6700	0.9805	0.6192	0.2534	0.9476	0.4864
IL-1ra	12.27	12.27	13.86	0.9708	0.9805	0.0870	0.0777	0.0502	0.1147
IL-2	0.15	0.14	0.16	0.9805	0.9805	0.4640	0.2149	0.4776	0.3475
IL-4	0.32	0.34	0.48	0.5575	0.9805	0.0002	0.0032	0.0017	0.0416
IL-5	0.52	0.50	0.55	0.9708	0.9805	0.5290	0.2315	0.4852	0.3475
IL-6	10.22	9.60	28.06	0.6878	0.9805	0.0769	0.0777	0.0345	0.0866
IL-7	4.93	4.98	5.45	0.6005	0.9805	0.4559	0.2149	0.6742	0.4095
IL-8	2.72	3.10	10.56	0.3941	0.9805	0.0009	0.0032	0.0181	0.0531
IL-9	2.81	3.11	3.61	0.0299	0.9805	0.0113	0.0260	0.3712	0.2977
IL-10	2.08	1.95	2.48	0.8170	0.9805	0.0899	0.0777	0.0902	0.1417
IL-12(p70)	2.45	2.45	2.97	0.9224	0.9805	0.0927	0.0777	0.0621	0.1274
IL-13	0.64	0.67	0.73	0.5504	0.9805	0.1073	0.0859	0.1675	0.1832
IL-15	1.26	1.45	1.28	0.4801	0.9805	0.8264	0.3030	0.7826	0.4374
IL-17	0.70	0.76	0.78	0.1098	0.9805	0.0118	0.0260	0.2108	0.2097
Eotaxin	70.03	88.90	92.30	0.5507	0.9805	0.3131	0.1604	0.6085	0.3825
FGFb	0.90	0.94	0.97	0.2778	0.9805	0.0870	0.0777	0.5025	0.3475
G-CSF	1.69	1.86	2.90	0.5180	0.9805	0.0007	0.0032	0.0065	0.0478
GM-CSF	0.10	0.10	0.10	0.7142	0.9805	0.3190	0.1604	0.5617	0.3622
IFN-g	23.77	24.10	32.65	0.8264	0.9805	0.0009	0.0032	0.0086	0.0478
MCP-1	44.98	64.62	86.72	0.2894	0.9805	0.1398	0.1026	0.3647	0.2977
MIP-1a	0.85	0.89	0.92	0.4860	0.9805	0.0737	0.0777	0.2563	0.2242
MIP-1b	1.23	1.09	1.28	0.2838	0.9805	0.6087	0.2534	0.2585	0.2242
RANTES	2.34	2.34	4.81	0.9126	0.9805	0.0034	0.0101	0.0035	0.0445
TNF-a	3.80	4.06	4.59	0.3297	0.9805	0.0242	0.0426	0.2168	0.2097
VEGF	15.93	16.06	17.52	0.7608	0.9805	0.2603	0.1432	0.4306	0.3282
CTACK	104.86	95.20	91.00	0.6879	0.9805	0.0722	0.0777	0.1412	0.1693
GROa	3.73	3.73	4.73	0.7421	0.9805	0.3722	0.1820	0.1481	0.1693
ICAM-1	828.72	806.21	979.12	0.8170	0.9805	0.0365	0.0536	0.0156	0.0531
IL-1a	0.13	0.12	0.12	0.9593	0.9805	0.4962	0.2239	0.2053	0.2097
IL-2Ra	49.19	45.78	39.44	0.5508	0.9805	0.1144	0.0875	0.2531	0.2242
IL-3	71.01	69.32	69.64	0.7149	0.9805	0.7367	0.2882	0.8233	0.4501
IL-12p40	216.90	223.82	216.59	0.8076	0.9805	0.5392	0.2315	0.5113	0.3475
IL-16	185.68	170.73	267.55	0.5187	0.9805	0.0294	0.0470	0.0130	0.0531
IL-18	6.08	5.47	4.11	0.8171	0.9805	1.0000	0.3520	0.9895	0.4977
LIF	1.92	2.28	1.17	0.6968	0.9805	0.2034	0.1279	0.0659	0.1274
MCP-3	16.09	16.36	18.26	0.3937	0.9805	0.1982	0.1279	0.7228	0.4227
M-CSF	0.33	0.32	0.33	0.9319	0.9805	0.8951	0.3215	0.9371	0.4864
MIF	75.35	94.57	210.55	0.4954	0.9805	0.0135	0.0264	0.1062	0.1483
MIG	452.94	432.04	533.47	0.7240	0.9805	0.1649	0.1161	0.0806	0.1352
b-NGF	7.32	7.48	7.20	0.7516	0.9805	0.2033	0.1279	0.1377	0.1693
SCF	82.52	72.73	61.65	0.9127	0.9805	0.0722	0.0777	0.1448	0.1693
SCGF	1261.96	1191.07	1355.14	0.6878	0.9805	0.2364	0.1342	0.3788	0.2977
SDF-1a	49.09	57.54	46.12	0.2785	0.9805	0.7589	0.2884	0.0979	0.1448
TNFb	0.32	0.32	0.32	0.6825	0.9805	0.2926	0.1560	0.5360	0.3547
TRAIL	32.60	31.65	32.38	0.7424	0.9805	0.7367	0.2882	0.7625	0.4358
VCAM-1	1156.08	1061.17	1303.37	0.6006	0.9805	0.2364	0.1342	0.0806	0.1352
HGF	331.54	329.31	376.77	0.2895	0.9805	0.2364	0.1342	0.6839	0.4095
IFNa2	17.43	16.94	17.36	0.6878	0.9805	0.7700	0.2884	0.9476	0.4864

157

158

Table S3. Pair-wise Wilcoxon rank-sum tests – Cycle Phase

Cytokine	Median Concentration (pg/mL)				Controls		Endometriosis	
	Follicular	Luteal	Follicular	Luteal	Unadjusted	BH	Unadjusted	BH
	(-)	(-)	(+)	(+)				
PDGF-bb	2.50	2.35	2.50	2.50	0.5930	1.0000	0.9685	0.8925
IL-1b	0.23	0.22	0.26	0.27	0.3959	0.9166	0.9685	0.8925
IP-10	588.31	903.94	674.08	841.48	0.1965	0.8983	0.0719	0.4128
IL-1ra	12.27	12.27	13.46	12.27	1.0000	1.0000	0.8232	0.8747
IL-2	0.15	0.14	0.12	0.15	0.9393	1.0000	0.0849	0.4211
IL-4	0.33	0.31	0.36	0.35	0.8189	1.0000	0.7721	0.8747
IL-5	0.51	0.52	0.52	0.52	0.7893	1.0000	0.7321	0.8747
IL-6	8.19	18.42	13.15	8.76	0.3047	0.8983	0.1761	0.5351
IL-7	4.87	5.33	4.98	5.07	0.7322	1.0000	0.7625	0.8747
IL-8	2.07	2.83	3.33	3.63	0.3619	0.9048	1.0000	0.8925
IL-9	2.95	2.57	2.95	3.72	0.9696	1.0000	0.0030	0.0660
IL-10	1.67	2.27	2.53	1.93	0.1105	0.8983	0.1485	0.5097
IL-12(p70)	2.76	2.35	2.76	2.53	0.2545	0.8983	0.6268	0.8747
IL-13	0.67	0.62	0.72	0.66	0.5423	1.0000	0.1306	0.5097
IL-15	1.06	1.56	1.58	1.21	0.0333	0.8983	0.0740	0.4128
IL-17	0.68	0.72	0.76	0.79	0.6728	1.0000	0.1431	0.5097
Eotaxin	58.41	82.86	88.90	95.93	0.2871	0.8983	0.7033	0.8747
FGFb	0.91	0.89	0.94	0.93	1.0000	1.0000	0.7825	0.8747
G-CSF	1.74	1.69	2.39	1.90	0.9391	1.0000	0.3244	0.8042
GM-CSF	0.06	0.10	0.09	0.12	0.3410	0.8983	0.0109	0.1623
IFN-g	23.00	25.73	27.51	30.79	0.7322	1.0000	0.6361	0.8747
MCP-1	28.78	51.40	64.47	65.59	0.4033	0.9166	0.4230	0.8505
MIP-1a	0.89	0.84	0.89	0.98	0.4931	1.0000	0.0646	0.4128
MIP-1b	1.14	1.26	1.14	1.36	0.3414	0.8983	0.0247	0.2752
RANTES	1.84	2.34	3.06	2.59	0.2227	0.8983	0.4945	0.8747
TNF-a	3.85	3.74	4.14	4.64	0.8786	1.0000	0.2370	0.6610
VEGF	16.65	14.48	17.43	16.29	0.7038	1.0000	0.6363	0.8747
CTACK	104.66	111.05	90.61	102.75	0.9394	1.0000	0.0979	0.4368
GROa	3.40	3.89	3.73	4.57	0.2386	0.8983	0.1799	0.5351
ICAM-1	825.79	831.64	831.64	833.30	0.5687	1.0000	0.7327	0.8747
IL-1a	0.13	0.13	0.12	0.12	0.4946	1.0000	0.9891	0.8925
IL-2Ra	48.81	50.61	42.32	45.42	0.7040	1.0000	0.9476	0.8925
IL-3	73.62	68.40	68.78	73.85	0.5949	1.0000	0.8029	0.8747
IL-12p40	213.41	218.61	212.14	223.94	1.0000	1.0000	0.4700	0.8738
IL-16	184.78	200.19	184.78	191.96	0.5947	1.0000	0.4384	0.8505
IL-18	8.65	5.44	4.53	5.88	0.1286	0.8983	0.6646	0.8747
LIF	1.47	2.02	1.60	2.33	0.2875	0.8983	0.0551	0.4128
MCP-3	12.86	16.55	18.14	16.62	0.0805	0.8983	0.8851	0.8925
M-CSF	0.32	0.34	0.30	0.38	1.0000	1.0000	0.0021	0.0660
MIF	95.59	60.67	117.40	87.56	0.1106	0.8983	0.2994	0.7860
MIG	278.30	462.03	482.08	416.41	0.3233	0.8983	0.7229	0.8747
b-NGF	7.28	7.32	7.37	7.43	0.9697	1.0000	0.6269	0.8747
SCF	96.38	71.04	69.49	71.31	0.2241	0.8983	0.4306	0.8505
SCGF	1284.13	1239.79	1191.07	1347.98	0.7040	1.0000	0.8233	0.8747
SDF-1a	51.24	46.94	51.09	54.22	1.0000	1.0000	0.4383	0.8505
TNFb	0.32	0.32	0.32	0.32	0.3185	0.8983	0.5097	0.8747
TRAIL	26.24	34.54	32.74	29.80	0.1286	0.8983	0.4383	0.8505
VCAM-1	1101.68	1262.57	1212.30	1172.52	0.1489	0.8983	0.5902	0.8747
HGF	347.02	270.51	370.93	333.56	1.0000	1.0000	0.9686	0.8925
IFNa2	17.16	17.61	16.94	17.42	0.9394	1.0000	0.8955	0.8925

160 **Table S4.** Pair-wise Wilcoxon rank-sum tests – Recurrence Status
 161

Cytokine	Median Concentration (pg/mL)		Recurrent Disease vs. Initial Diagnosis	
	Initial Diagnosis	Recurrent Disease	Unadjusted	BH
PDGF-bb	2.5	2.5	0.5778	1.0000
IL-1b	0.26	0.27	0.6741	1.0000
IP-10	674.08	777.26	0.2284	1.0000
IL-1ra	13.46	12.985	0.9137	1.0000
IL-2	0.13	0.155	0.1160	1.0000
IL-4	0.35	0.36	0.5151	1.0000
IL-5	0.5	0.55	0.2221	1.0000
IL-6	14.91	9.86	0.5976	1.0000
IL-7	4.49	5.29	0.5606	1.0000
IL-8	3.76	3.405	0.8817	1.0000
IL-9	3.11	3.58	0.0782	1.0000
IL-10	2.3	2.175	1.0000	1.0000
IL-12(p70)	2.62	2.71	0.9784	1.0000
IL-13	0.72	0.685	0.6068	1.0000
IL-15	1.24	1.435	0.5697	1.0000
IL-17	0.76	0.76	0.8477	1.0000
Eotaxin	89.44	90.645	0.8603	1.0000
FGFb	0.88	0.96	0.0527	1.0000
G-CSF	2.03	2.185	0.9030	1.0000
GM-CSF	0.1	0.1	0.9892	1.0000
IFN-g	27.51	27.955	0.6549	1.0000
MCP-1	43.11	85.98	0.1103	1.0000
MIP-1a	0.89	0.92	0.8704	1.0000
MIP-1b	1.15	1.22	0.1632	1.0000
RANTES	3.83	2.56	0.3433	1.0000
TNF-a	4.2	4.17	0.7970	1.0000
VEGF	16.06	17.435	0.3366	1.0000
CTACK	94.18	92.67	0.7351	1.0000
GROa	4.35	3.645	0.2969	1.0000
ICAM-1	831.64	845.67	0.7556	1.0000
IL-1a	0.12	0.12	0.4742	1.0000
IL-2Ra	45.35	43.605	0.9892	1.0000
IL-3	79.96	67.81	0.1133	1.0000
IL-12p40	223.82	213.41	0.3717	1.0000
IL-16	179.21	205.625	0.4901	1.0000
IL-18	5.1	5.38	0.2973	1.0000
LIF	1.99	1.795	0.8710	1.0000
MCP-3	18.37	16.985	0.7452	1.0000
M-CSF	0.32	0.33	0.8495	1.0000
MIF	117.4	92.38	0.6552	1.0000
MIG	435.18	458.115	0.8817	1.0000
b-NGF	7.38	7.38	1.0000	1.0000
SCF	71.04	71.09	0.6650	1.0000
SCGF	1216.32	1236.435	0.6650	1.0000
SDF-1a	51.09	51.605	0.7970	1.0000
TNFb	0.35	0.32	0.2858	1.0000
TRAIL	32.01	31.65	0.6165	1.0000
VCAM-1	1237.57	1123.315	0.8180	1.0000
HGF	297.14	375.135	0.1553	1.0000
IFNa2	18.98	16.73	0.1593	1.0000

162 **Table S5.** Pair-wise Wilcoxon rank-sum tests – Lesion Distribution

163

Cytokine	Median Concentration (pg/mL)				Peritoneal vs. Controls		Ovarian vs. Controls		Deep vs. Controls	
	Controls	Peritoneal	Ovarian	Deep	Unadjusted	BH	Unadjusted	BH	Unadjusted	BH
PDGF-bb	2.43	2.50	1.86	2.31	0.8165	1.0000	0.0343	0.8201	0.7596	0.9786
IL-1b	0.22	0.23	0.21	0.25	0.5203	1.0000	0.3056	0.9257	0.0956	0.4316
IP-10	706.02	736.40	838.86	786.78	1.0000	1.0000	0.5823	0.9751	0.2128	0.6130
IL-1ra	12.63	13.00	11.87	15.06	0.9805	1.0000	0.5970	0.9751	0.1145	0.4316
IL-2	0.14	0.14	0.14	0.13	0.9414	1.0000	0.9297	1.0000	0.7792	0.9786
IL-4	0.31	0.34	0.32	0.31	0.7503	1.0000	0.8945	1.0000	0.7580	0.9786
IL-5	0.51	0.52	0.47	0.53	0.7507	1.0000	0.9649	1.0000	0.8185	0.9786
IL-6	9.72	9.60	6.43	8.98	0.9127	1.0000	0.0989	0.9257	1.0000	1.0000
IL-7	5.15	5.14	5.28	6.37	0.9320	1.0000	0.8430	1.0000	0.2524	0.6130
IL-8	2.25	2.46	2.07	3.36	0.6966	1.0000	0.1724	0.9257	0.0793	0.4316
IL-9	2.81	2.91	3.11	3.21	0.6000	1.0000	0.2079	0.9257	0.0705	0.4316
IL-10	1.83	1.88	1.59	2.65	0.9514	1.0000	0.1525	0.9257	0.8587	0.9786
IL-12(p70)	2.41	2.47	2.29	2.47	0.7421	1.0000	0.3906	0.9257	0.9797	1.0000
IL-13	0.63	0.64	0.64	0.64	0.8167	1.0000	1.0000	1.0000	0.5083	0.8301
IL-15	1.01	0.96	0.98	1.84	0.8169	1.0000	0.3669	0.9257	0.0118	0.2897
IL-17	0.70	0.76	0.71	0.76	0.6836	1.0000	0.9294	1.0000	0.0845	0.4316
Eotaxin	46.98	47.26	49.15	105.09	0.9708	1.0000	0.5822	0.9751	0.0394	0.4316
FGFb	0.91	0.94	0.87	1.05	0.3729	1.0000	0.5375	0.9751	0.2627	0.6130
G-CSF	1.69	1.81	1.68	2.11	0.4632	1.0000	0.7914	1.0000	0.0190	0.3102
GM-CSF	0.12	0.12	0.10	0.11	0.6335	1.0000	0.5955	0.9751	0.5568	0.8553
IFN-g	23.93	24.10	24.10	22.74	0.6786	1.0000	0.9649	1.0000	0.9391	1.0000
MCP-1	41.01	40.81	50.72	81.55	0.9127	1.0000	0.8776	1.0000	0.0446	0.4316
MIP-1a	0.87	0.89	0.79	0.97	0.6516	1.0000	0.3649	0.9257	0.3868	0.7285
MIP-1b	1.20	1.21	1.05	1.10	0.9030	1.0000	0.3328	0.9257	0.1544	0.5042
RANTES	2.34	2.34	2.10	2.83	0.6688	1.0000	0.1786	0.9257	0.3590	0.7285
TNF-a	3.79	3.85	3.99	4.03	0.7140	1.0000	0.4142	0.9257	0.7404	0.9786
VEGF	14.25	15.20	10.64	13.73	0.7513	1.0000	0.2711	0.9257	0.5586	0.8553
CTACK	107.48	108.09	116.27	75.14	0.9320	1.0000	0.4156	0.9257	0.0984	0.4316
GROa	3.88	3.90	3.28	3.91	0.8357	1.0000	0.3219	0.9257	0.8189	0.9786
ICAM-1	837.80	831.64	889.34	826.38	0.9127	1.0000	0.7084	1.0000	0.7410	0.9786
IL-2Ra	49.19	45.49	44.02	57.23	0.4574	1.0000	0.5234	0.9751	0.2525	0.6130
IL-3	69.20	67.01	65.16	68.29	0.8934	1.0000	0.9824	1.0000	0.9190	1.0000
IL-12p40	216.89	215.19	215.35	214.53	0.9223	1.0000	0.8430	1.0000	1.0000	1.0000
IL-16	185.67	188.80	137.29	206.36	0.6173	1.0000	0.0209	0.8201	0.3467	0.7285
IL-18	4.77	4.52	3.31	5.32	0.9320	1.0000	0.4156	0.9257	0.4305	0.7534
LIF	2.40	2.38	2.54	3.75	1.0000	1.0000	0.6920	1.0000	0.2855	0.6358
MCP-3	13.39	13.41	12.72	12.73	0.7057	1.0000	0.7917	1.0000	0.9594	1.0000
M-CSF	0.33	0.33	0.31	0.38	1.0000	1.0000	0.4540	0.9551	0.1088	0.4316
MIF	70.06	90.30	59.13	98.48	0.7057	1.0000	0.3670	0.9257	0.1472	0.5042
MIG	452.94	451.86	433.61	569.86	0.8742	1.0000	0.4156	0.9257	0.2525	0.6130
b-NGF	7.52	7.48	7.40	8.42	0.9903	1.0000	0.7917	1.0000	0.5933	0.8810
SCF	76.00	79.66	62.09	86.57	0.6522	1.0000	0.3011	0.9257	0.8190	0.9786
SCGF	1261.77	1284.13	1061.32	1430.55	0.8169	1.0000	0.4678	0.9551	0.4014	0.7285
SDF-1a	51.90	57.23	52.74	65.75	0.6260	1.0000	0.8950	1.0000	0.1092	0.4316
TNFb	0.33	0.34	0.34	0.33	0.8153	1.0000	0.7905	1.0000	0.8574	0.9786
TRAIL	36.52	38.69	27.00	43.35	0.7607	1.0000	0.0502	0.8201	0.4014	0.7285
VCAM-1	1081.96	1110.15	1065.03	834.71	0.9127	1.0000	0.7414	1.0000	0.1947	0.5963
HGF	324.22	327.65	246.95	536.20	0.6522	1.0000	0.1657	0.9257	0.0029	0.1436
IFNa2	17.43	17.61	16.70	16.10	0.9223	1.0000	0.9124	1.0000	0.4609	0.7787

164

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165 **Table S6.** Pair-wise Wilcoxon rank-sum tests – Primary Indication
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Cytokine	Median Concentration (pg/mL)		Infertility vs. Pain	
	Pain Only	Infertility +/- Pain	Unadjusted	BH
PDGF-bb	2.5	2.13	0.4126	0.8618
IL-1b	0.26	0.285	0.4466	0.8618
IP-10	707.89	689.47	0.5569	0.8618
IL-1ra	13.06	13.455	0.5188	0.8618
IL-2	0.14	0.165	0.6566	0.8872
IL-4	0.35	0.445	0.2334	0.8192
IL-5	0.55	0.48	0.9199	0.9738
IL-6	10.97	12.645	0.5007	0.8618
IL-7	5.06	5.145	0.4916	0.8618
IL-8	3.79	3.1	0.7965	0.9738
IL-9	3.11	3.35	0.2174	0.8192
IL-10	2.12	2.62	0.6160	0.8801
IL-12(p70)	2.62	2.96	0.5097	0.8618
IL-13	0.72	0.695	1.0000	1.0000
IL-15	1.42	0.985	0.0855	0.8192
IL-17	0.76	0.76	0.8731	0.9738
Eotaxin	102.89	67.645	0.1184	0.8192
FGFb	0.94	0.935	0.9428	0.9738
G-CSF	2.34	1.82	0.5860	0.8618
GM-CSF	0.12	0.09	0.0615	0.8192
IFN-g	27.51	28.54	0.5567	0.8618
MCP-1	51.41	92.495	0.1253	0.8192
MIP-1a	0.89	0.94	0.7955	0.9738
MIP-1b	1.2	1.12	0.4737	0.8618
RANTES	2.66	2.81	0.8075	0.9738
TNF-a	4.2	4.01	0.5006	0.8618
VEGF	15.36	21.44	0.2126	0.8192
CTACK	92.91	100.1	0.6989	0.9196
GROa	4.35	3.2	0.1599	0.8192
ICAM-1	818.72	950.84	0.2233	0.8192
IL-1a	0.12	0.115	0.0249	0.8192
IL-2Ra	45.49	39.33	0.3821	0.8618
IL-3	69.32	72.485	0.8974	0.9738
IL-12p40	219.76	216.585	0.4737	0.8618
IL-16	198.74	167.4	0.2126	0.8192
IL-18	4.89	6.485	0.4307	0.8618
LIF	1.99	1.605	0.2458	0.8192
MCP-3	17.64	16.49	0.4915	0.8618
M-CSF	0.32	0.31	0.2015	0.8192
MIF	114.75	81.015	0.6364	0.8838
MIG	482.08	336.64	0.3517	0.8618
b-NGF	7.38	7.32	0.9543	0.9738
SCF	71.48	54.61	0.1253	0.8192
SCGF	1202.35	1366.225	0.9087	0.9738
SDF-1a	57.54	48.245	0.1922	0.8192
TNFb	0.32	0.32	0.8487	0.9738
TRAIL	31.65	31.83	0.8298	0.9738
VCAM-1	1091.14	1274.385	0.2956	0.8618
HGF	342.44	412.09	0.5763	0.8618
IFNa2	16.86	18.37	0.2826	0.8618

167
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Table S7. Reported associations between peritoneal cytokines, chemokines, growth factors and pelvic endometriosis.

Interleukins	Endometriosis vs. Controls			
	Increased	No Difference	Decreased	Transient
IL-1		Koyama 1993		
IL-1 α	Kondera-Anasz 2005	Dziunycz 2009		
IL-1 β	Fakih 1987	Keenan 1989		
	Taketani 1992	Kalu 2007		
	Ho 1996	Bedaiwy 2002		
	Sukhikh 2004	Oku 2004		
	Mier-Cabrerra 2010	Milewski 2008		
	Michaud 2011			
	Sikora 2012			
IL-1Ra	Kondera-Anasz 2005	Ho 1996	Zhang 2007	
		Bersinger 2012	Mier-Cabrerra 2010	
IL-1 sRII			Kondera-Anasz 2005	
sIL1RAcP			Michaud 2011	
IL-2		Punnonen 1996	Hsu 1997	
		Oku 2004		
		Podgaec 2007		
		Hassa 2009		
		Mier-Cabrerra 2010		
IL-2Ra				
IL-3				
IL-4	Hsu 1997	Punnonen 1996		
		Oku 2004		
		Podgaec 2007		
		Hassa 2009		
		Mier-Cabrerra 2010		
IL-5	Koyama 1993	Punnonen 1996		
IL-6	Koyama 1993	Keenan 1989		
	Rier 1995	Oku 2004		
	Ho 1996			
	Punnonen 1996			
	Harada 1997			
	Bedaiwy 2002			
	Khan 2002			
	Kalu 2007			
	Milewski 2008			
	Dziunycz 2009			
	Mier-Cabrerra 2010			
	Velasco 2010			
	Drosdzol-Cop 2012			
	Bersinger 2012			
IL-6sR			Rier 1995	
IL-7				
IL-9				
IL-10	Punnonen 1996	Rana 1996		Ho 1997
	Tabibzadeh 2003	Hsu 1997		
	Kondera-Anasz 2004	Oku 2004		
	Podgaec 2007	Hassa 2009		
	Mier-Cabrerra 2010	Andreoli 2011		
		Bersinger 2012		
		Gazvani 2001		
IL-11		Zeyneloglu 1998		Ho 1997
IL-12(p70)	Mazzeo 1998	Gazvani 2001		
	Gallinelli 2004	Mier-Cabrerra 2010		
	Fairbanks 2009	Andreoli 2011		
		Bersinger 2012		
IL-12(p40)		Mazzeo 1998		
IL-13		Bedaiwy 2002	McLaren 1997	
		Mier-Cabrerra 2010	Gallinelli 2004	
		Velasco 2010		
IL-14				
IL-15		Bersinger 2012	Mier-Cabrerra 2010	Arici 2003
IL-16	Koga 2005			

IL-17		Velasco 2010 Andreoli 2011		Zhang 2005
IL-18	Oku 2004 Bersinger 2012	Fairbanks 2009 Glitz 2009	Zhang 2004 Sikora 2012	Arici 2003
IL-19				
IL-20				
IL-21				
IL-22				
IL-23	Andreoli 2011			
IL-24				
IL-25				
IL-26				
IL-27				
IL-28				
IL-29				
IL-30				
IL-31				
IL-32				
IL-33	Santulli 2012			
IL-35				
IL-36				
IL-37				
IL-38				
MIF	Kats 2002 Mahutta 2004			
IFN α 2		Sukhikh 2004		
IFN γ	Podgaec 2007	Keenan 1989 Khorram 1993 Oku 2004 Sukhikh 2004 Milewski 2008 Hassa 2009	Ho 1996 Hsu 1997 Wu 1998 Mier-Cabrera 2010	
TNF Superfamily				
TNF α	Rana 1996 Ho 1996 Overton 1996 Harada 1997 Bedaiwy 2002 Sukhikh 2004 Mier-Cabrera 2010 Drosdzol-Cop 2012	Keenan 1989 Calhaz-Jorge 2000 Oku 2004 Kalu 2007 Podgaec 2007 Milewski 2008 Dziunycz 2009		Pizzo 2002
TNFRSF1A/B/sTNFR TNFRSF11B/OPG	Koga 2000 Harada 2004 Bersinger 2006			
TNF β TRAIL sFas sFasL ICAM-1				Harada 2004
VCAM-1		Kalu 2007 Kalu 2007 Somigliana 1996 Daniel 2000 Bersinger 2012 Daniel 2000 Bersinger 2012		
TGF β	Oosterlynck 1994 Kupker 1998 Pizzo 2002			
Hematopoietic				
LIF G-CSF GM-CSF		Bersinger 2012 Punnonen 1996 Oku 2004		
CSF1/M-CSF	Fukaya 1994 Budrys 2012		Weinberg 1991	
SCGF SCF				Osuga 2000
Tissue Remodeling				
bFGF PDGF-bb VEGF	Bourlev 2006 McLaren 1996 Kupker 1998	Bersinger 2012 Overton 1996 Kalu 2007 Dziunycz 2009		

171 **Table S8.** Over-representation of transcriptional binding sites among macrophage secreted
 172 cytokines.
 173

Transcription Factor	All Motifs	Consensus Sequence(s)	Co-expression Profile P-Value		Co-expression Profile FDR			
			IRIS	HICT	IRIS	HICT		
C/EBPbeta	<0.0001	V\$CEBPB_01	0.0000	0.0000	0.1330	0.0645		
		V\$CEBPB_02	0.0022	0.0000	0.1448	0.0392		
		V\$CEBP_Q2_01	0.0129	0.1626	0.2820	0.4249		
		V\$CEBP_Q3	0.2298	0.3982	0.6020	0.7252		
		V\$CEBPB_Q6	NA	NA	NA	NA		
RelA-p65-isoform1	0.0026	V\$NFKAPPAB65_01	0.0021	0.0000	0.0749	0.0078		
		V\$NFKAPPAB_01	0.0000	0.0000	0.0371	0.0015		
		V\$NFKB_Q6_01	0.0000	0.0000	0.0456	0.0089		
		V\$RELA_Q6	NA	NA	NA	NA		
CREB1	0.0047	V\$CREB_01	0.9017	0.8568	0.9857	1.0000		
		V\$CREB_Q2	0.6960	0.1106	0.9826	0.3907		
		V\$CREB_Q4	0.7767	0.8489	1.0000	1.0000		
		V\$CREB_Q2	0.0895	0.7080	0.4423	1.0000		
		V\$TAXCREB_01	0.8693	0.1147	0.9997	0.3401		
		V\$TAXCREB_Q2	0.4063	0.2286	0.7197	0.3837		
		V\$CREB_Q2_01	0.6716	0.7511	0.9015	0.9338		
		V\$CREB_Q4_01	0.2041	0.3401	0.5608	0.6951		
		V\$CREB_Q3	0.2981	0.1549	0.7330	0.7430		
		V\$CREBATF_Q6	NA	NA	NA	NA		
		V\$CREB1_01	NA	NA	NA	NA		
		V\$CREB1_Q6	NA	NA	NA	NA		
		NF-kappaB1-p50:RelA-p65	0.0095	V\$P50RELAP65_Q5_01	NA	NA	NA	NA
ATF-2	0.0186	V\$CREBP1_01	0.6869	0.9660	0.9999	1.0000		
		V\$CREBP1CJUN_01	0.7810	0.6883	NA	NA		
		V\$CREBP1_Q2	0.8366	0.4664	0.9642	0.7473		
		V\$CREB_Q3	0.2981	0.1549	NA	NA		
		V\$CREBATF_Q6	NA	NA	NA	NA		
NF-kappaB1-p50	0.0198	V\$ATF2_Q5	NA	NA	NA	NA		
		V\$NFKAPPAB50_01	NA	NA	NA	NA		
		V\$NFKAPPAB_01	0.0000	0.0000	0.0371	0.0015		
		V\$NFKB_Q6	0.0000	0.0000	0.0300	0.0064		
		V\$NFKB_C	0.0022	0.0000	0.0982	0.0128		
c-Fos:c-Jun	0.0274	V\$NFKB_Q6_01	0.0000	0.0000	0.0456	0.0089		
		V\$P50_Q6	NA	NA	NA	NA		
		V\$CFOSCJUN_Q5	NA	NA	NA	NA		
		c-Ets-1	0.0286	V\$CETS1P54_01	0.2140	0.7336	0.6859	0.9423
		V\$CETS1P54_02	NA	NA	NA	NA		
AP-1	0.0417	V\$ETS1_B	0.0712	0.0319	0.4129	0.3434		
		V\$ETS_B	NA	NA	NA	NA		
		V\$ETS_Q6	NA	NA	NA	NA		
		V\$ETS_Q4	0.0000	0.0000	0.0035	0.0131		
		V\$CETS1P54_03	NA	NA	NA	NA		
		V\$CETS1_Q6	NA	NA	NA	NA		
		V\$CETS1_02	NA	NA	NA	NA		
		V\$ETS1_02	NA	NA	NA	NA		
		V\$AP1FJ_Q2	0.0044	0.0000	0.1437	0.1090		
		V\$AP1_Q2	0.0713	0.0050	0.3912	0.1115		
STAT1	0.0444	V\$AP1_Q6	0.0000	0.0041	0.0835	0.1088		
		V\$AP1_Q4	0.0000	0.0000	0.1027	0.1030		
		V\$AP1_C	0.0000	0.4241	0.1214	0.7488		
		V\$AP1_01	0.0000	0.0000	0.0159	0.0796		
		V\$AP1_Q2_01	0.0109	0.0807	0.2366	0.3499		
		V\$AP1_Q6_01	0.0066	0.0449	0.1453	0.2619		
		V\$AP1_Q4_01	0.0000	0.0000	0.0488	0.0730		
		V\$AP1_Q6_02	NA	NA	NA	NA		
		V\$STAT1_01	0.3418	0.4359	0.7391	1.0000		
		V\$STAT_01	0.0146	0.0000	0.1433	0.0385		
STAT1	0.0444	V\$STAT1_02	0.5580	0.1652	0.8404	0.4480		
		V\$STAT1_03	0.8896	0.7739	0.9931	1.0000		
		V\$STAT_Q6	0.6152	0.8502	0.8555	0.9888		
		V\$STAT1_05	NA	NA	NA	NA		
		V\$STAT1_Q6	NA	NA	NA	NA		

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 175 NA – Not Annotated in MSigDB v4.0
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176 **Table S9.** Luminex Targets and Assay Performance Characteristics

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Panel	Analyte	HUGO Gene Symbol	xMAP Region	LoD, pg/mL	Intra-Assay CV, %	Inter-Assay CV, %
Group I - 27-plex	IL-1b	IL1B	39	0.02	0.7	2.5
	IL-1ra	IL1RA	25	1.28	0.9	5.7
	IL-2	IL2	38	0.34	0.8	3.6
	IL-4	IL4	52	0.02	2	3.5
	IL-5	IL5	33	0.03	1.7	4.5
	IL-6	IL6	19	0.13	0.7	3.2
	IL-7	IL7	74	0.07	1.4	3.2
	IL-8	IL8	54	0.06	2.4	2.9
	IL-9	IL9	77	0.83	1	4.3
	IL-10	IL10	56	0.12	0.4	6.4
	IL-12 (p70)	IL12A,B	75	3.10	2.2	1.8
	IL-13	IL13	51	0.05	1.2	4.4
	IL-15	IL15	73	0.62	0.4	4.5
	IL-17	IL17	76	0.59	0.8	7.5
	Eotaxin	CCL11	43	0.52	1.2	8
	bFGF	FGFB	44	0.47	0.7	6.1
	G-CSF	CSF3	57	0.21	1.5	3.7
	GM-CSF	CSF2	34	0.65	0.4	3.4
	IFNg	IFNG	21	0.52	1.3	2.9
	IP-10	CXCL10	48	0.63	1.1	3.8
	MCP-1	CCL2	53	0.23	0.5	4
	MIP-1a	CCL3	55	0.22	0.6	5.3
	MIP-1b	CCL4	18	0.21	0.5	2.5
	PDGF	PDGF	47	0.45	2	3.8
	RANTES	CCL5	37	0.46	1	6.5
	TNFa	TNF	36	1.55	1.4	3.6
	VEGF	VEGF	45	0.47	1.1	4.1
Group II - 21-plex	IFNa2	IFNA2	20	0.93	1.7	9
	IL-1a	IL1A	63	0.13	1.1	3.3
	IL-2ra	IL2RA	13	0.70	1.1	11.1
	IL-3	IL3	64	1.64	0.9	6.4
	IL-12 (p40)	IL12B	28	0.40	1.4	7.6
	IL-16	IL16	27	0.51	1.3	6.8
	IL-18	IL18	42	0.09	6	8.6
	CTACK	CCL27	72	1.10	2.5	6.1
	GRO-a	CXCL1	61	1.87	1	5.6
	HGF	HGF	62	0.74	0.9	6
	LIF	LIF	29	0.54	0.9	2.9
	MCP-3	CCL7	26	0.31	1.5	10.9
	M-CSF	CSF1	67	0.53	1	4.8
	MIF	MIF	35	1.38	4.6	7.9
	MIG	CXCL9	14	1.50	1.3	9.7
	b-NGF	NGFB	46	0.21	0.9	5
	SCF	SCF	65	0.38	1.3	3.8
	SCGF-b	CLEC11A	78	5.72	1.5	5.8
	SDF-1a	CXCL12	22	1.19	1.3	12.3
	TNFb	TNFB	30	0.51	0.4	3.8
TRAIL	TNFSF10	66	0.79	1.5	6.3	
ICAM-1 singleplex	ICAM-1	ICAM1	12	0.95	1.3	5.4
VCAM-1 singleplex	VCAM-1	VCAM1	15	0.70	1.8	9.4

178 **Table S10.** Leukocyte Sub-populations Among Peritoneal Aspirates
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Population	Surface Staining	Peripheral Blood ¹	Total	Peritoneal Aspirate	
				Adherent	Non-adherent
Leukocytes	CD45+	85%	98	98	98
Granulocytes	CD14+ CD68- CD10+	59 ± 20	<1	<1	<1
Monocytes/Macrophages	CD14+ CD68+ CD10-	6 ± 2	64 ± 7 74 ± 8**	96 ± 7	41 ± 6
Lymphocytes	CD14- CD68- CD10-	34 ± 8	20 ± 6	<5	60 ± 13

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 181 Values are mean ± s.d. percent composition of three donors.
 182 ¹Peripheral blood values are reference ranges for healthy adults (Reichert et al. 1991, Bain
 183 1996)
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Table S11. Kinase Inhibitor Treatments

Inhibitor	Primary Target(s)	IC50 (nM)	Experimental Dose (uM)
LY294002	PI3K β	310	10
	PI3K α	730	
	PI3K δ	1,060	
	PI3K γ	6,600	
PD98059	MEK1	4,000	50
	MEK2	50,000	
SB203580	p38 α /MAPK14	50	10
	p38 β /MAPK11	500	
SP600125	JNK1	40	10
	JNK2	40	
	JNK3	90	
BMS-345541	IKK2	300	10
	IKK1	4,000	
SU6656	SRC	280	10
	Fyn	170	
	Yes	20	
	Lyn	130	
JAK Inhibitor I	JAK1	15 (murine)	10
	JAK2	1	
	JAK3	5 (Ki50)	
	Tyk2	1	
Gö6983	PKC α	7	10
	PKC β	7	
	PKC γ	6	
	PKC δ	10	
	PKC ζ	60	

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