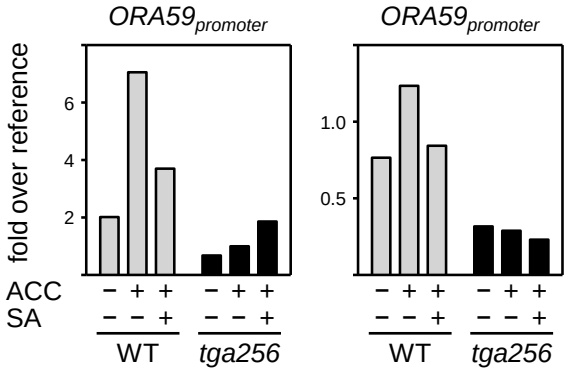
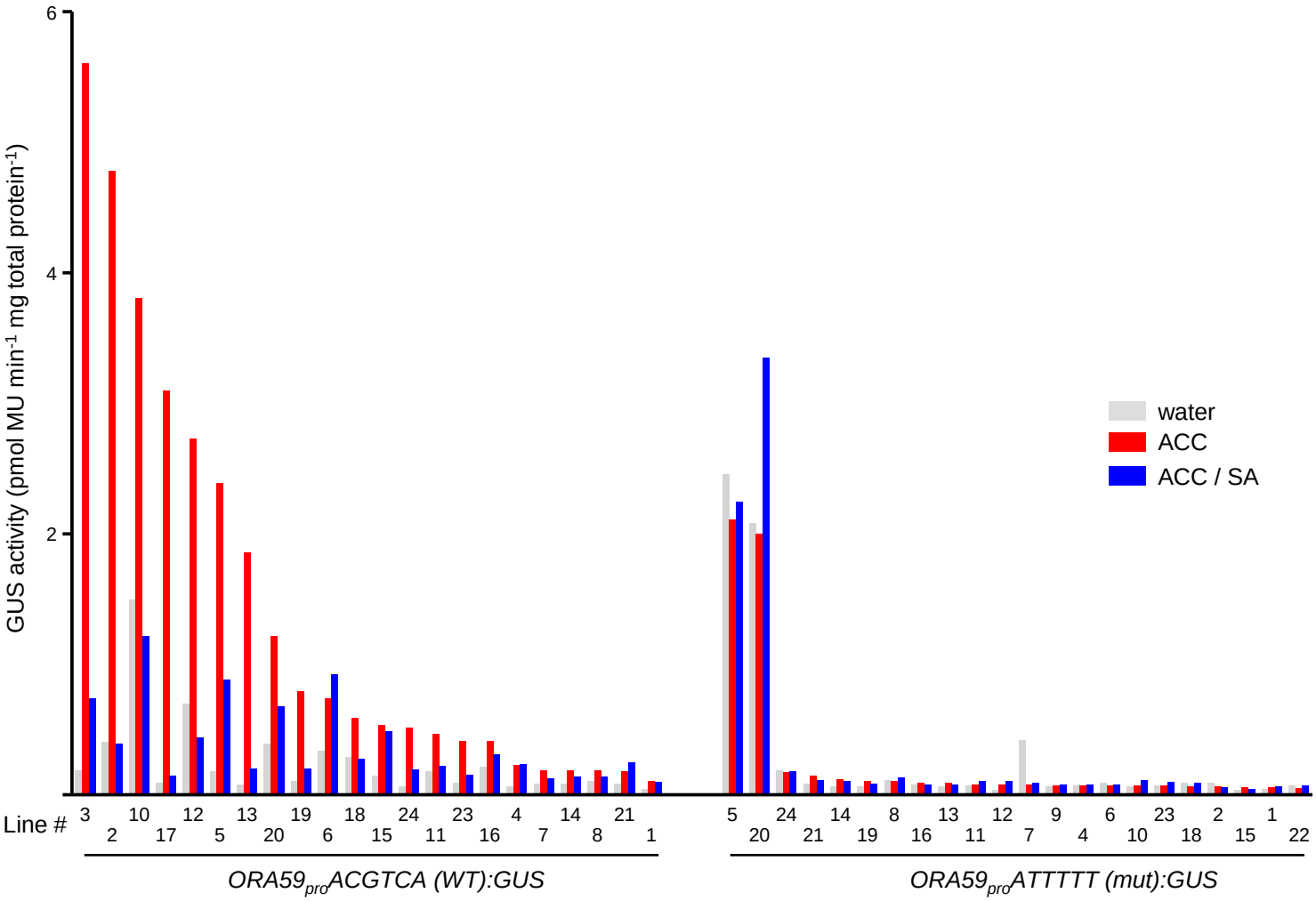


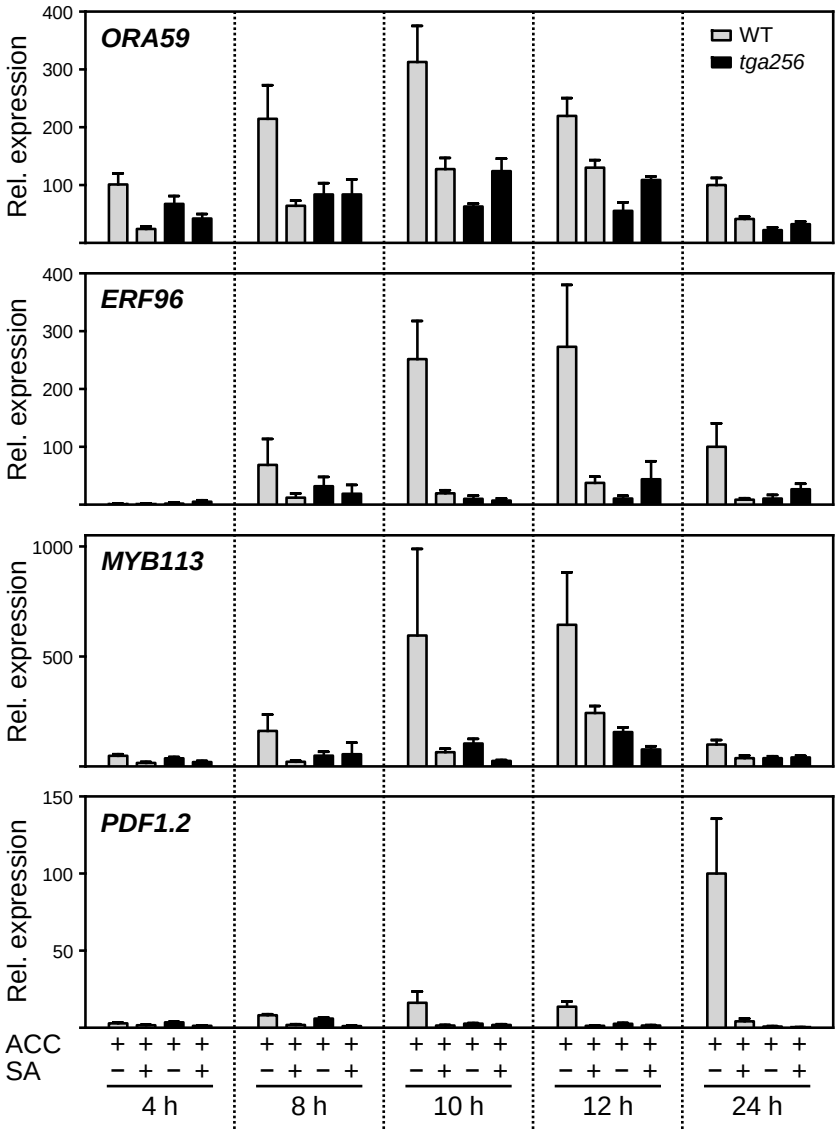
Supplemental Figure S1. Chromatin immunoprecipitation analysis of TGA binding to the *ORA59* promoter

Four-week old soil-grown wild-type (WT) and *tga256* mutant plants were treated with water, 1 mM ACC, or 1 mM ACC/1 mM SA for 24 h. Ten plants were combined per treatment and experiment. Chromatin samples were subjected to immunoprecipitation using the αTGA2,5 antiserum. The co-immunoprecipitated DNA was recovered and *ORA59* promoter fragments were amplified with quantitative PCR using the primer pair indicated in Figure 5. A genomic fragment of *ACTIN8* was used as a reference. Results of two independent experiments are shown.



Supplemental Figure S2. GUS activities of independent transformants encoding the wild-type (WT) and the mutated *ORA59* promoter upstream of the *GUS* reporter gene

Plants (20 to 30 F2 plants per treatment and line) were sprayed with either water, 1 mM ACC or 1 mM ACC/1 mM SA and harvested after 24 h.



Supplemental Figure S3. Time-dependent expression analysis of selected transcription factors and *PDF1.2* in ACC- and ACC/SA-treated wild-type and *tga256* plants

Four-week old soil-grown wild-type (WT) and *tga256* were sprayed with 1 mM ACC or 1 mM ACC/1 mM SA and harvested after 4, 8, 10, 12, and 24 hours. The relative transcript levels of the indicated transcription factors and *PDF1.2* were determined by quantitative RT-PCR analysis. The relative expression values in wild-type plants after 24 h after ACC treatment were set to 100%. The mean values (±SE) of four to five biological replicates are shown.

Supplemental Table S1: Primers for Genotyping, Cloning and ChIP Analyses

Abbr.	Description	Sequence
P1	ora59 GK-061A12_LP	ATGCTGCAGACGGTAACAAAC
P2	ora59 GK-061A12_RP	CTTCAAGCTCATGGAGCTCAC
P3	TGA25 fwd.	GTCAATCCGGTTTCATATTCTCCTC
P4	TGA25 rev.	CCGCATAAACAATAAACCAAGAGAG
P5	tga25 rev.	GAGCGACAACTCCTTTCAACTCATC
P6	TGA6 fwd.	AAGCGGATAGGTGATCAGTG
P7	tga6 fwd.	TTCTCACTTTGTGATTTGCCTTTGG
P8	TGA6 rev.	TGGGCAATCTTGCTATGATTTCAAG
P9	ORA59-Pro. fwd.	GGGGACAAGTTTGTACAAAAAAGCAGGCTCCG GATTGGTTGCAGGTTACGATG
P10	ORA59-Pro. rev.	GGGGACCACTTTGTACAAGAAAGCTGGGTCGC ATTTTCGATCTTTTTTTTTCTTCTTG
P11	pDONR fwd.	ACGACGTTGTAAAACGACGGCCAG
P12	ORA59 TGACG overlap rev.	GGACAAGACCAGGTTGAGTGTAAAAAATACGG CGGCGTATTCCCGAC
P13	ORA59 TGACG overlap fwd.	GTATTTTTTACACTCAACCTGGTCTTGTCC
P14	pDONR rev.	GTAACATCAGAGATTTTGAGACAC
P15	ACTIN8 ChIP fwd.	GGTTTTCCCCAGTGTTGTTG
P16	ACTIN8 ChIP rev.	CTCCATGTCATCCCAGTTGC
P17	ORA59 ChIP fwd.	CGAGAGAGTATATGAAGAGGCCAA
P18	ORA59 ChIP rev.	GGACAAGACCAGGTTGAGTG
P19	ERF96 ChIP fwd.	TCTTTCATTATATAGAGGTGTCGAGGG
P20	ERF96 ChIP fwd.	GCGTGTTTGTATGTGTTTGTGTATAGG

Supplemental Table S2: Primers for Quantitative RT-PCR Analyses

Description	Sequence / Order No.
UBQ5 RT fwd.	GACGCTTCATCTCGTCC
UBQ5 RT rev.	GTAAACGTAGGTGAGTCCA
PDF1.2 RT fwd.	CTTGTTCTCTTTGCTGCTTTC
PDF1.2 RT rev.	CATGTTTGGCTCCTTCAAG
ORA59	QuantiTect Primer Assay QT00852054 (Qiagen, Hilden, Germany)
ERF96	QuantiTect Primer Assay QT00737828 (Qiagen, Hilden, Germany)
MYB113	QuantiTect Primer Assay QT00887684 (Qiagen, Hilden, Germany)
ERF1	QuantiTect Primer Assay QT00777777 (Qiagen, Hilden, Germany)
ATERF14	QuantiTect Primer Assay QT00868315 (Qiagen, Hilden, Germany)
ATERF15	QuantiTect Primer Assay QT00760648 (Qiagen, Hilden, Germany)
ERF9	QuantiTect Primer Assay QT00738395 (Qiagen, Hilden, Germany)
ANAC032	QuantiTect Primer Assay QT00743561 (Qiagen, Hilden, Germany)

Supplemental Table S3: Accession Numbers

Gene	Accession Number
UBQ5	*At3g62250*
PDF1.2	*At5g44420*
ORA59	*At1g06160*
ERF96	*At5g43410*
MYB113	*At1g66370*
ERF1	*At3g23240*
ATERF14	*At1g04370*
ATERF15	*At2g31230*
ERF9	*At5g44210*
ACTIN8	*At1g49240*
TGA2	*At5g06950*
TGA5	*At5g06960*
TGA6	*At3g12250*
CTR1	*At5g03730*